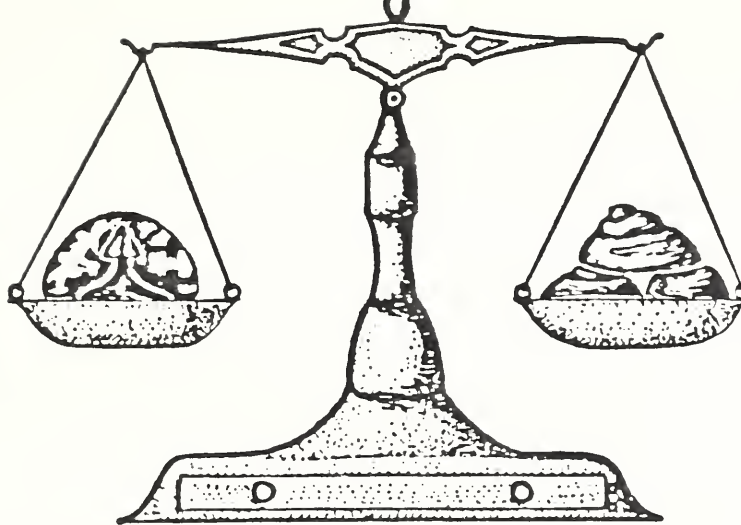


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THE SCALE



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International Advisory Council
for Biosystematic Services in Entomology

by Ke Chung Kim
Chair, International Advisory Council
for Biosystematic Services in Entomology

The International Advisory Council for Biosystematic Services in Entomology (IACBSE) (initially called "...Committee...") was formally established at the XVIIth International Congress of Entomology, held August 23, 1984, in Hamburg, Federal Republic of Germany, by Resolution No. 1 upon the recommendation of the Permanent Council of the International Congress of Entomology. The Resolution emphasized that (a) the number of available insect taxonomists is not sufficient to meet demands for their expertise, (b) the training and employment of biosystematists necessary for the supply of biosystematic services need to be promoted, (c) all potential users communicate in the planning stages of their projects with potential providers of biosystematic services, and (d) national and international organizations make specific budgetary allocations for the support of the biosystematic component in their projects.

The Organizing Committee (K. M. Harris, K. C. Kim as Chairman, L. Knutson, I. M. Smith) was formed by the final Discussion Session at the Congress Symposium (SI.2) "Biosystematic Services in Entomology," to organize the first International Advisory Committee for Biosystematic Services in Entomology. The mandate of the Organizing Committee was to develop initial guidelines for the structure and operation of the Committee, to establish a well balanced membership for the inaugural Committee, and to propose a number of projects to be pursued by IACBSE during its initial term leading up to the XVIII International Congress of Entomology, to be held in Vancouver, British Columbia, Canada, in 1988. The Organizing Committee developed the guidelines and identified prospective members for IACBSE on 29 March 1985, at the National Museum of Natural History, Smithsonian Institution, Washington, D.C., U.S.A.

Working Groups (WG) are project-oriented groups with specific mission. Each WG is organized as a new project and is developed and approved by the EC. The Chair of each WG is appointed by the EC Chair with the concurrence of the Executive Committee. The membership of each WG is appointed by its Chair, in consultation with EC.

MEMBERSHIP

1. The IACBSE consists of 20 members; the Chair as Representative-at-Large, and 19 Representatives from 11 regions (six biogeographic zones).

2. Program Regions and Representation

PALEARCTIC ZONE

Region 1, Eastern Europe and USSR - 2 members

Region 2, Western Europe - 2 members

Region 3, Far East - 2 members

AFRICA/NEAR EAST ZONE

Region 4, Near East - 2 members

Region 5, East Africa - 1 member

Region 6, West Africa - 1 member

ORIENTAL ZONE

Region 7, Southeast Asia - 2 members

AUSTRALIAN-OCEANIA ZONE

Region 8, Australia, New Zealand, and Oceania - 1 member

NEARCTIC ZONE

Region 9, North America (incl. Mexico) - 3 members

NEOTROPICAL ZONE

Region 10, Central America - 1 member

Region 11, South America - 2 members

3. Inaugural Membership

PALEARCTIC ZONE

Region 1, Eastern Europe and USSR

Eastern Europe: L. Papp (Hungarian Natural History Museum,
Budapest, HUNGARY)

USSR: V. F. Zaitsev (Zoological Institute, Academy of Sciences,
Leningrad, USSR)

Region 2, Western Europe

England: K. M. Harris (Commonwealth Institute of Entomology,
British Museum (Natural History), London, UNITED KINGDOM)

Continental Europe: R. zur Strassen (Forschungsinstitut
Senckenberg, Frankfurt am Main, FEDERAL
REPUBLIC OF GERMANY)

Region 3, Far East

Japan: K. Hirashima (Faculty of Agriculture, Kyushu University,
Hakozaki, JAPAN)

Peoples Republic of China: Wu Yan-ru (Institute of Zoology,
Academia Sinica, Beijing, CHINA)

AFRICA/NEAR EAST ZONE

Region 4, Near East

Israel: D. Rosen (Department of Entomology, Hebrew University of
Jerusalem, Rehovot, ISRAEL)

Egypt: M. A. Zaher (Acarology Division, Agricultural Zoology
Department, Faculty of Agriculture, Cairo University,
Giza, EGYPT)

Region 5, Eastern Africa

Kenya: J. M. Ritchie (Entomology Section, National Museums of Kenya, Nairobi, KENYA)

Region 6, Western Africa

Nigeria: A. Akingbohunbe (Department of Plant Science, University of Ife, Ile-Ife, NIGERIA)

ORIENTAL ZONE

Region 7, Southeast Asia

India: T. N. Anathakrishna (Entomology Research Institute, Loyola College, Madras, INDIA)

Philippines: Nelson Esquerra (Visayas State College of Agriculture, Leyte, PHILIPPINES)

AUSTRALIAN-OCEANIA ZONE

Region 8, Australia, New Zealand, and Oceania

New Zealand: J. C. Watt (Entomology Division, DSIR, Private Bag, Auckland, New Zealand)

NEARCTIC ZONE

Region 9, North America

Canada: I. M. Smith (Biosystematics Research Institute, Research Branch, Agriculture Canada, Ottawa, Ontario, CANADA)

Mexico: P. Reyes-Castillo (Institute de Ecologia, A. C., MEXICO, D.F.)

USA: L. Knutson (BARC-WEST, Agricultural Research Service, U.S. Department of Agriculture, Beltsville, MD 20705, USA)

NEOTROPICAL ZONE

Region 10, Central America

Costa Rica: L. Fernando Jiron (Universidad de Costa Rica, Ciudad Universitaria, Rodrigo Facio, San Jose, COSTA RICA)

Region 11, South America

Brazil: N. Papavero (Museu de Zoologia da Universidade de Sao Paulo, Sao Paulo, BRAZIL)

Southern S.A.: R. Gonzalez (Faculty of Agronomy, University of Chila, Santiago, CHILE)

REPRESENTATIVE-AT-LARGE

IACBSE Chair: K. C. Kim (The Frost Entomological Museum, Department of Entomology, The Pennsylvania State University, University Park, PA 16802, USA)

INAUGURAL PROJECTS

The following projects are being organized as Working Groups:

- A. Faunal Assessment of Insects and Arachnids in the World: Current knowledge, research needs, etc.

Convenor: I. M. Smith

- B. International Directory of Systematic Entomology: Services, specialists, research needs, etc.

Convenor: L. Knutson

- C. Regional Biosystematic Service Centers: Service network, models, coordination, workshops, etc.

Convenor: K. M. Harris

- D. IACBSE Newsletter

Convenor: I. M. Smith

- E. Heliothis

Convenor: L. Knutson

- F. Tephritidae

Convenor: K. C. Kim

Any inquiry concerning IACBSE may be made to Prof. K. C. Kim, IACBSE, The Frost Entomological Museum, Department of Entomology, The Pennsylvania State University, University Park, PA 16802, U.S.A.

NECROLOGY

From the Starkville Daily News, Starkville, Mississippi, September 8, 1981:

Mary Hoke Lobdell

"Starkville friends will regret to learn of the recent death of Mary Gladys Hoke Lobdell, of Pisgah Forest, North Carolina, a former resident of Starkville. Mrs. Lobdell, 86, was the widow of Richard Nugent Lobell.

"Mrs. Lobdell was educated at Mississippi University for Women (then MSCW) and the University of Illinois receiving the bachelor's degree in entomology. She became the first woman faculty member at Mississippi State University (then Mississippi A&M College).

"She taught at Brevard (North Carolina) from 1942 until 1964. During her career she discovered and described two genera and 24 species of scale insects. She also contributed to 'Naturalist's Guide to the Americas.'

"Survivors include a son, David Lobdell of Pisgah Forest, a sister and two granddaughters."

NEARCTIC ENTOMOLOGY: CONTINENTAL COOPERATION
ESA, ESC AND ESO MEETING
29 November - 3 December 1982
Toronto, Canada

TITLE: THE MISSISSIPPI ENTOMOLOGICAL MUSEUM SCALE INSECT COLLECTION

AUTHOR: R. G. Baer, Dept. of Entomology, Drawer EM, MSU, Mississippi State,
MS 39762

ABSTRACT: The scale insect collection was started in 1930 by the late Mary Gladys Hoke Lobdell. Presently the collection of 6,000 slides includes 10 families, 42 genera and 150 species of scale insects and mealybugs found in Mississippi. Currently, information on species, hosts and localities for the purposes of retrieval is being computerized.

Holdings of the Mississippi Entomological Museum

Foreign Scale Insects on Slides
(mainly from A. Balachowsky)

<u>Lot No.</u>	<u>Species</u>	<u>Host</u>
1	<i>Aonida tlaiae</i> Balach.	<i>Tamarix articulata</i>
2	<i>Asterolecanium ilicicola</i> Targ.	<i>Quercus coccifera</i>
3	<i>Aspidiotus abietis</i> Schrank.	<i>Pinus silvestris</i>
4	<i>Ceroplastes rusci</i> (L.)	<i>Ficus carica</i>
5	<i>Ceroplastes sinensis</i> DelGue	<i>Schinus molle</i>
6	<i>Ceroplastes sinensis</i> DelGue	<i>Citrus sinensis</i>
7	<i>Chionaspis etrusca</i> Leon	<i>Tamarix</i> sp.
8	<i>Chionaspis longiloba</i>	
9	<i>Chionaspis nereii</i> Newst.	<i>Nerium oleander</i>
10	<i>Chionaspis salicis</i>	<i>Sophora japonica</i>
11	<i>Chionaspis salicis</i>	<i>Salix incana</i>
12	<i>Chionaspis striata</i> Newst.	<i>Thuja orientalis</i>
13	<i>Cryptoparlatoria lencaspis</i> Lind.	<i>Cryptomeria japonica</i>
14	<i>Diaspis calyptroides</i> Casta	<i>Opuntia tomentosa</i>
15	<i>Diaspis visci</i> Schr.	<i>Cypressus sempervirens</i>
16	<i>Eriococcus bezzii</i> Leon.	
17	<i>Eriococcus ericae</i> Sign.	<i>Erica tetralix</i>
18	<i>Eriococcus inermis</i> Green	grass
19	<i>Eriococcus insignis</i> Newst.	grass
20	<i>Eriococcus nukii</i> Kuw.	bamboo
21	<i>Eriococcus spurius</i> Modeer	<i>Ulmus campestris</i>
22	<i>Hemiberlesia minima</i> Leon	<i>Quercus coccifera</i>
23	<i>Lepidosaphes ampelodesma</i> Newst.	<i>Amelodesmas tenax</i>
24	<i>Lepidosaphes conchiformis</i> (Gwel.)	<i>Ficus carica</i>
25	<i>Lepidosaphes conchiformis</i> (Gwel.)	elm
26	<i>Lepidosaphes ficifolii</i> (Berl.)	<i>Ficus carica</i>
27	<i>Lepidosaphes hawaiiensis</i> Mask.	<i>Albizzia</i> sp.
28	<i>Lepidosaphes japonica</i>	<i>Pinus silvestris</i>
29	<i>Lepidosaphes lasianthi</i> (Green)	<i>Croton</i>
30	<i>Lepidosaphes sexspira</i> Hoke	<i>Citrus grandis</i>
31	<i>Leucaspis candida pini</i> (Targ.)	<i>Pinus silvestris</i>
32	<i>Leucaspis riccae</i> Targ.	<i>Ephedra campylopoda</i>
33	<i>Leucaspis signoretti</i> (Targ.)	<i>Pinus pinea</i>
34	<i>Leucaspis vayssieri</i> Balachw.	<i>Rhus pentaphylla</i>
35	<i>Lichtensia ephedra</i> Newst.	<i>Coronilla pentophylla</i>
36	<i>Macrocerococcus superbus</i> Leon.	
37	<i>Orthezia insignis</i> Dougl.	<i>Clitoria</i>
38	<i>Parlatoria blanchardi</i> Targ.	<i>Phoenix dactylifera</i>
39	<i>Parlatoria ziziphi</i> Lucas	<i>Citrus nobilis</i>
40	<i>Pinnaspis minor strachani</i> Cooley	<i>Croton</i>
41	<i>Pseudococcus nipae</i> (Mask.)	<i>Hulitzia augusta</i>
42	<i>Pseudoparlatoria ostreata</i> Ckll.	<i>Cestrum nocturnum</i>
43	<i>Trabutina elastica</i> Marchal	<i>Tamarix gallica</i>

Holdings of the Mississippi Entomological Museum

A. Balachowsky Collection
(Dry Material)

<u>Lot No.</u>	<u>Species</u>	<u>Host</u>
16	<i>Aspidiotus britannicus</i> Newst.	<i>Ruscus aculeatus</i>
17	<i>Parlatoria oleae</i>	<i>Prinus</i> sp.
18	<i>Leucaspis pusilla</i> Loew	<i>Pinus halepensis</i>
19	<i>Fiorinia hirsuta</i> Marchal	<i>Nebbelium lougana</i>
20	<i>Fiorinia fioriniae</i> Targ.	<i>Taurus nobilis</i>
21	<i>Chionaspis ceratoniae</i> Marchal	<i>Ceratonia siligua</i>

Chermotheca Italica
Fascicolo IV, Portici 1908
(Dry Material)

<u>Lot No.</u>	<u>Species</u>	<u>Host</u>
76	<i>Asterolecanium hederæ</i> Licht.	<i>Hedera</i>
77	<i>Margarodes mediterraneus</i> Silv.	<i>Nel terreno</i>
78	<i>Filippia oleae</i> (Costa)	<i>Oleae</i>
79	<i>Lecanodiaspis sardoa</i> Targ.	<i>Cistus</i> sp.
80	<i>Eulecanium coryli</i> (Linn.)	<i>Corylus</i>
81	<i>Trabutina elastica</i> March.	<i>Tamarix</i>
82	<i>Leucaspis riccae</i> Targ.	<i>Oleae</i>
83	<i>Eriococcus bezzi</i> Leon.	<i>Rhododendron</i>
84	<i>Ceroplastes sinensis</i> Del Guercio	<i>Citrus</i>
85	<i>Ceroplastes nerii</i> Newst.	<i>Nerium</i>
86	<i>Asterolecanium aureum</i> (Bdv.)	<i>Anturium</i>
87	<i>Asterolecanium thesii</i> (Dougl.)	<i>Pittosporum</i>
88	<i>Chionaspis etrusca</i> Leon.	<i>Tamarix</i>
89	<i>Chionaspis salicis</i> Linn.	<i>Sophora</i>
90	<i>Chionaspis salicis</i> Linn.	<i>Alnus</i>
91	<i>Diaspis pentagona</i> Targ.	<i>Ailanthus</i>
92	<i>Aspidiotus abietis</i> (Schr.)	<i>Pinus</i>
93	<i>Aspidiotus hederæ</i> Vallot	<i>Acacia</i> sp.
94	<i>Chrysomphalus dictyospermi pinnulifera</i> (Mask.)	<i>Agrumi</i>
95	<i>Chrysomphalus dictyospermi pinnulifera</i> (Mask.)	<i>Phoenix</i>
96	<i>Chrysomphalus degeneratus</i> Leon.	<i>Camellia</i>
97	<i>Aonidiella aurantii</i> (Mask.)	<i>Agrumi.</i>
98	<i>Aonidiella taxus</i> Leon.	<i>Taxus</i>
99	<i>Targionia nigra</i> Sign.	<i>Quercus</i> sp.
100	<i>Mytilaspis conchiformis</i> (Gmel.) Sign.	<i>Olmo.</i>

Chermotheca Italica
Fascicolo V, Portici 1909
(Dry Material)

<u>Lot No.</u>	<u>Species</u>	<u>Host</u>
101	Eriopeltis festucae (Fonsc.)	Festuca
102	Aspidiotus pyri Licht.	Pero
103	Asterolecanium variolosum (Ratz.)	Quercus
104	Kermes baciformis Leon.	Quercus
105	Lepidosaphes newsteadi (Sulc)	Pinus
106	Macrocerococcus superbus Leon.	Graminacee
107	Asterolecanium variolosum var. minor Leon.	Quercus
108	Orthezia martelli Leon.	Graminacee
109	Lepidosaphes ficifolii (Berl.)	Ficus
110	Asterolecanium algeriense Newst.	Templetonia
111	Sphaerolecanium emerici (Planch.)	Quercus
112	Lepidosaphes ficifolii var. ulmicola Leon.	Olmo
113	Kermes vermilio Planch.	Quercus
114	Eulecanium Cecconi Leon.	Menispermum
115	Diaspis boisduvalii Sign.	Pandanus
116	Guerinia serratulae (Fab.)	Leguminose
117	Gossyparia spuria (Modeer)	Olmo
118	Lecanium corni var. robiniarum (Dougl.)	Robinia
119	Pulvinaria vitis (Linn.)	Vite
120	Kermes roboris (Fourc.)	Quercus
121	Leucaspis lowi Colvee.	Pinus
122	Targionia vitis var. Suberi Leon.	Quercus
123	Targionia vitis var. arbutus Leon.	Arbutus
124	Aspidiotus ostraiformis Curtis	Oleae
125	Aonidiella aurantii (Mask.)	Diospyrus

Holdings of the Mississippi Entomological Museum

Coccoidea of Japan (Dry material)
S. I. Kuwana

<u>Lot No.</u>	<u>Species</u>	<u>Host</u>
1.	<i>Aspidiotus ficus</i> Ashm.	<i>Euonymus japonica</i>
2.	<i>Chionaspis wistariae</i>	<i>Frannhia floribunda</i>
3.	<i>Pulvinaria amantii</i> Ckll.	Orange
4.	<i>Chionaspis aspidstrae</i> Sign.	<i>Sepidistra elatior</i>
5.	<i>Mytilaspis craweii</i> Ckll.	<i>Pasamia cuspidata</i>
6.	<i>Chiponaspis euonymi</i> Comet.	<i>Euonymus japonica</i>
7.	<i>Chionaspis aucubae</i> Cooley	<i>Aucuba japonica</i>
8.	<i>Aspidiotus aurantii</i> Mask.	<i>Quercus serrata</i>
9.	<i>Diaspis pentagona</i> Targ.	<i>Morus alba</i>
10.	<i>Aspidiotus perniciosus</i> Comst.	Pear
11.	<i>Lecaniodiaspis quercus</i> Ckll.	<i>Pasania cuspidata</i>
12.	<i>Aspidiotus duplex</i> Ckll.	Orange
13.	<i>Ceroplastes rubens</i> Mask.	Orange
14.	<i>Aspidiotus cryptomeriae</i> Kwd.	<i>Crytomeria japonica</i>
15a.	<i>Cryptoparlatoria leucaspis</i> Lindg.	<i>Crytomeria japonica</i>
15b.	<i>Aspidiotus zonatus</i> Frauenf.	<i>Quercus robur</i>
16.	<i>Antonina Crawii</i> Ckll.	Bamboo
17.	<i>Diaspis rosae</i> Bouche	Rose
18.	<i>Lecanium kunoensis</i> Kwd.	Prunus
19.	<i>Eriococcus onukii</i> Kwd.	Bamboo
20.	<i>Chionaspis citri</i> Comst.	Orange
122b.	<i>Asterolecanium variolosum</i>	<i>Quercus robur</i>
133.	<i>Asterolecanium fimbriatum</i> (Fonsc.) Ckll.	<i>Vinicia minor</i>
134.	<i>Antonina purpurea</i> Sign.	<i>Agropyrum intermedium</i>
135.	<i>Aspidotus bavaricus</i> Ldgr.	<i>Erica arborea</i>
136.	<i>Aspidiotus britannicus</i> Newst.	<i>Rhamnus alaternus</i>
137.	<i>Aspidiotus hederæ</i> (Vall.) Sign.	<i>Rosmarinus officinalis</i>
138.	<i>Aspidiotus hederæ</i> (Vall.) Sign.	<i>Thymelaea hirsuta</i>
139.	<i>Chrysomphalus dictyospermi</i> (Morg.) Leon.	<i>Buxus sempervirens</i>
140.	<i>Aulacaspis pentagona</i> (Targ.) Newst.	<i>Clematis vitalba</i>
141.	<i>Chionaspis salicis</i> (L.) Sign.	<i>Sorbus aria</i>
142.	<i>Lepidosaphes conchiformis</i> (Gmel.) Ldgr.	<i>Cistus monspeliensis</i>
143.	<i>Leucaspis signoreti</i> (Targ.) Sign.	<i>Pinus pinea</i>
144.	<i>Ceroplastes sinensis</i> Del Guercio	<i>Schinus molle</i>
145.	<i>Aspidiotus canariensis</i> Ldgr.	<i>Centaurea</i>
146.	<i>Aspidiotus lenticularis</i> Ldgr.	<i>Olea europaea</i>
147.	<i>Aspidiotus hederæ</i> (Vall.) Sign.	<i>Lavatera maritima</i>
148.	<i>Aspidiotus hederæ</i> (Vall.) Sign.	<i>Teucrium chamaedrys</i>
149.	<i>Chrysomphalus dictyospermi</i> (Morg.) Leon.	<i>Citrus aurantium</i>
150.	<i>Targionia vitis</i> (Sign.) Leon.	<i>Quercus ilex</i>

<u>Lot No.</u>	<u>Species</u>	<u>Host</u>
150b.	Targionia vitis (Sign.) Leon.	Quercus ilex
151.	Aulacaspis pentagona (Targ.) Newst.	Rhamnus alaternus
152.	Chionaspis salicis (L.) Sign.	Vaccinium vitis idaea
153.	Epidiaspis subterranea	Agropyrum intermedium
154.	Lepidosaphes ulmi (L.) Fern.	Pistacia lentiscus
155.	Lecanium oleae (Bern.) Walk.	Erica arborea
156.	Pulvinaria ericae Low	Erica carnea
157.	Lecaniodiaspis sardoa Targ.	Cistus monspeliensis
158.	Polilinia pollinii (Costa) Ckll.	Olea europaea
159.	Aspidiotus hederæ (Vall.) Sign.	Atriplex halimus
160.	Aspidiotus hederæ (Vall.) Sign.	Stachys officinalis
161.	Aspidiotus labiatarum March.	Globularia cordifolia
162.	Aspidiotus minimus (Leon.) Ckll.	Quercus ilex
163.	Aulacaspis pentagona (Targ.) Newst.	Sarothamnus scoparius
164.	Chionaspis salicis (L.) Sign.	Salix aurita
165.	Diaspis visci (Schr.) Low	Juniperus phoenicea
166.	Lepidosaphes ulmi (L.) Fern.	Spartium junceum
167.	Aclerda subterranea Sign.	Agropyrum intermedium
168.	Ceroplastes sinensis Del Guercio	Erica arborea
169.	Asterolecanium variolosum (Ratz.) Ckll.	Quercus lanuginosa
170.	Ceroputo superbus (Leon.) Ldgr.	Silene italica
171.	Aspidiotus hederæ (Vall.) Sign.	Calycotome spinosa
172.	Aspidiotus hederæ (Vall.) Sign.	Euphorbia characias
173.	Aspidiotus jaapi. Ldgr. sp. nov.	Genista pilosa
174.	Aspidiotus lenticularis Ldgr.	Populus tremula
175.	Chrysomphalus dictyospermi (Morg.) Leon.	Hedera helix
176.	Targionia nigra Sign.	Cineraria maritima
177.	Aulacaspis pentagona (Targ.) Newst.	Morus alba
178.	Lepidosaphes ulmi (L.) Fern	Sarothamnus scoparius
179.	Lecanium hesperidum (L.) Burm.	Pittosporum tobira
180.	Lecanium oleae (Bern.) Walk.	Myrtus italica
181.	Asterolecanium fimbriatum (Fonsc.) Ckll.	Pittosporum tobira
182.	Eriococcus ericae Sign.	Erica tetralix
183.	Pseudococcus calluneti Ldgr.	Calluna vulgaris
184.	Pseudococcus hibernicus (Newst.) Fern.	Calamagrostis arenaia
185.	Aspidiotus abietus (Schr.) Low	Pinus silvestris
186.	Aspidiotus hederæ (Vall.) Sign.	Bupleurum fruticosum
187.	Aspidiotus hederæ (Vall.) Sign.	Cytisus triflorus
188.	Aspidiotus hederæ (Vall.) Sign.	Silene italica
189.	Aulacaspis pentagona (Targ.) Newst.	Cytisus nigricans
190.	Parlatoria blanchardi (Targ.) Leon	Phoenix dactylifera
191.	Ceroplastes rusci (L.) Sign.	Cistus monspeliensis
192.	Lecanium corni Bouche, Marchal.	Ribes rubrum
193.	Fonscolombea fraxni (Kalt.) Ckll.	Fraxinus excelsior
194.	Pseudococcus vovai Nas.	Juniperas oxycedrus

<u>Lot No.</u>	<u>Species</u>	<u>Host</u>
195.	Aspidiotus cecconii (Leon.) Sand. (A. Trabuti March.)	Ephedra major var. Villarsii
196.	Aspidiotus hederæ (Vall.) Sign.	Ceratonia siliqua
197.	Aspidiotus hederæ (Vall.) Sign.	Daphne gnidium
198.	Aspidiotus hederæ (Vall.) Sign.	Osyris alba
199.	Targionia donacis Ldgr. sp. n.	Arundo donax
200.	Aulacaspis pentagona (Targ.) Newst.	Ribes rubrum
201.	Lepidosaphes conchifmis	Ficus carica
202.	Leucaspis pusilla Low	Pinus pinaster
203.	Leucaspis riccai Targ.	Ephedra campylopoda
204.	Parlatoria oleæ (Colv.) Ldgr.	Olea europæa
205.	Aspidiotus britannicus Newst.	Lonicera implexa
206.	Aspidiotus hederæ (Vall.) Sign.	Coronilla emeroides
207.	Aspidiotus hederæ (Vall.) Sign.	Ligustrum japonicum
208.	Aspidiotus hederæ (Vall.) Sign.	Rubia peregrina
209.	Aspidiotu hedericola Ldgr. sp. n.	Hedera helix
210.	Aulacaspis pentagona (Targ.) Newst.	Pelargonium zonale
211.	Diaspis visci (Schr.) Low.	Juniperus oxycedri
212.	Epidiaspis betulæ (Barensprung) Ldgr. (E. Lepere: (Sign.) Ldgr.)	Prunus domestica
213.	Lepidosaphes newsteadi (Sulc.) Fern. (L. juniperi Ldgr.)	Juniperus oxycedrus
214.	Leucaspis Riccai Targ.	Olea europæa
215.	Parlatoria oleæ (Colv.) Ldgr.	Pirus amygdaliformis
216.	Ceroplastes rusci (L.) Sign.	Fixus carica
217.	Asterolecanium fimbriatum (Fonsc.) Ckll.	Phagnalon annoticum
218.	Lecaniodiaspis sardoa Targ.	Cistus cfeticus
219.	Phenacoccus aceris (Sign.) Ckll.	Betula verrucosa
220.	Aspidiotus hederæ (Vall.) Sign.	Euphorbia wulfenii
221.	Aspidiotus hederæ (Vall.) Sign.	Heliotropium peruvianum
222.	Aspidiotus hederæ (Vall.) Sign.	Smilax aspera
223.	Aspidiotus lenticularis Ldgr.	Pistacia lentiscus
224.	Aspidiotus rapax Comst.	Euonymus japonicus
225.	Chionaspis salicis (L.) Sign.	Fraxinus excelsior
226.	Epidiaspis subterranea (Sign.) Ldgr.	Agropyrum repens
227.	Parlatoria oleæ (Colv.) Ldgr.	Pistacia lentiscus
228.	Filippia oleæ (Costa) Sign.	Olea europæa
229.	Pseudococcus adonidum (L.) Westw.	Acrostichum aureum
230.	Aspidiotus hederæ (Vall.) Sign.	Robinia pseudacacia
231.	Chrysomphalus dictyospermi (Morg.) Leon.	Phoenix dactylifera
232.	Aulacaspis pentagona. (Targ.) Newst.	Sedum reflexum
233.	Chionaspis salicis (L.) Sign.	Salix repens
234.	Diaspis Boisduvali Sign.	Caraguata sp.
235.	Diaspis visci (Schr.) Low.	Juniperus oxycedrus
236.	Lepidosaphes newsteadi (Sulc.) Fern.	Pinus silvestris
237.	Leucaspis candida (Targ.) Sign.	Pinus silvestris
238.	Filippia oleæ (Costa.) Sign.	Hedera helix
239.	Lecanium corni Bouche, Marchal.	Prunus persica
240.	Luzulaspis luzulæ (Duf.) Fern.	Carex remota
241.	Eriococcus spurius (Mod.) Ldgr.	Ulmus campestris

<u>Lot No.</u>	<u>Species</u>	<u>Host</u>
242.	Phenacoccus aceris (Sign.) Ckll.	Quercus robur
243.	Aspidiotus hederæ (Vall.) Sign.	Dianthus caesius
244.	Aspidiotus hederæ (Vall.) Sign.	Lonicera implexa
245.	Aspidiotus ostreiformis Curt.	Salix caprea
246.	Chrysomphalus dictyospermi (Morg.) Leon.	Pandanus veitchi
247.	Chionaspis salicis (L.) Sign.	Salix pentandra
248.	Lepidosaphes ulmi (L.) Fern.	Cornus sanguinea
249.	Leucaspis lowi Colv.	Pinus silvestris
250.	Lecanium ciliatum Dougl. (L. Douglasi Sulc).	Populus tremula
251.	Physokermes fuscus (Gmel.)	Quercus robur
252.	Luzulaspis luzulae (Duf.) Fern.	Carex silvatica
253.	Aspidiotus bornmulleri (Rubs.) Ldgr.	Globularia salicina
254.	Aspidiotus hederæ (Vallot) Sign.	Phoenix dactylifer
255.	Aspidiotus hederæ (Vallot) Sign.	Ruta bracteosa
256.	Aspidiotus lauretorum Ldgr.	Laurus canariensis
257.	Cryptaspidotus aonidioides Ldgr.	Laurus canariensis
258.	Chionaspis salicis (L.) Sign.	Salix incana
259.	Pinnaspis pandani (Comst.) Ckll.	Anthurium magnificum
260.	Lecanium corni Bouche, Marchal.	Alnus alnobetula
261.	Lecanium hemisphaericum Targ.	Zygopetalum sp. and Lycaste skinneri
262.	Physokermes coryli (L.) Ldgr.	Alnus glutinosa
263.	Pulvinaria betulæ (L.) Sign.	Salix repens
264.	Orthezia urticae (L.) Amy. et Serv.	Stellaria holostea
265.	Aspidiotus hederæ (Vall.) Sign.	Yucca clotfolia
266.	Chrysomphalus dictyospermi (Morg.) Leon.	Alpinia vittata
267.	Diaspis boisduvali Sign.	Pandanus ulitis
268.	Diaspis zamiae Morg.	Cycas revoluta
269.	Pinnaspis pandant (Comst.) Ckll. and Aspidiotus cyanophylli Sign.	Phytelephas macrocarpa
270.	Parlatorea zizyphi (Luc.) Sign. and einige P. perganda Comst.	Citrus aurantium
271.	Xyloccus loti Ldgr. Sp. n.	Lotus cytisoides L.
272.	Lecanium hemisphaericum Targ.	Acroctichum aureum
273.	Lecanium hemisphaericum Targ.	Cycas revoluta
274.	Lecanium hemisphaericum Targ.	Polyposium quercifolium
275.	Lecanium hesperidum (L.) Burm. var. clienum (Dougl.) Newst.	Philodendron amazonicum
276.	Orthezia urticae (L.) Am. et Serv.	Betonica officinalts

NOTE ON THE FIFTH INTERNATIONAL SYMPOSIUM OF
SCALE INSECT STUDIES (ISSIS-V)
PORTICI, ITALY, June 24-28, 1986

Michael Kosztarab
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The program started with an **Awards Ceremony** for the three recipients of the Filippo Silvestri Research Competition Award. The **Plenary Session** of ISSIS-V was opened with greetings by Antonio Tranfaglia who introduced Kosztarab as chair of the Plenary Session.

Kosztarab asked the participants to pay tribute to the two coccidologists who died since the last meeting in 1983, **Alfred S. Balachowsky** of France and Professor **Fong-Ge Chen** of China. The participants stood in silence during and after the names were read. Director Gennaro Viggiani made the joint Plenary Session presentation with Tranfaglia.

An **Introductory Business Meeting** followed where a "Resolution and Site Selection Committee for ISSIS-VI" was chosen which included D. J. Williams (as chair), Y. Ben-Dov, M. Kosztarab, F. Kozar, D. Matile-Ferrero, and A. Tranfaglia. The committee was entrusted to study the two written invitations received from Israel and Poland proposing sites for the 1990 meeting and to make recommendations.

The **presentations** were grouped into five major Sessions: I-Phylogeny and Taxonomy, II-Morphology, III-Faunistics, IV-Scale Insects of Economic Importance and their Natural Enemies, and V-Poster Presentations. The individual Sessions were cochaired by: I - E. Tremblay and J. M. Cox; II - C. C. C. Tao and D. J. Williams; IIIa - D. Matile-Ferrero and F. Kozar; IIIb - I. Foldi and S. Paloukis; IVa - D. Rosen and V. Yashnosh; IVb - P. J. Gullan and E. Podsiadlo. **Abstracts** of 44 papers were printed in a program booklet.

Besides the five poster presentations, 35 papers were read. With a total of 40 presentations, and 61 participants from 16 countries, **this was the largest international coccidological conference to date.** When comparing topics with the earlier four meetings, we find at ISSIS-V a definite increase in the number of presentations on parasites of coccoids, also, for the first time, talks on coccoid paleontology, sex pheromones, integrated control, plant quarantine and on literature cataloging.

The **Round Table Discussions** were concerned with the following suggestions from participants for future research and topics at meetings: the effect of plant hosts on scale parasites, plant-scale interactions, scale cover formation, dermal glands, color of substrate and body color in coccoids, population dynamics and ecology, behavior, chromosome studies, and plant quarantine. Some of the participants reported on their **projects in progress** and I filled in gaps for those not present, and reported on the progress made with project commitments listed as priorities in the Summary of ISSIS-IV.

It was concluded that although good progress was made during the past three years, much work remains in the systematics, biology, ecology and zoogeography of Coccoidea. Workers were encouraged to fill existing gaps in our knowledge of this group. Those colleagues who volunteered during the Budapest meeting to prepare family catalogues, or to publish inventories of the collections under their direction, and to compile terminologies for standardization of certain families, should conclude their work. A number of participants were able to report in Portici on the progress already made toward achieving some of these goals.

The Second Business Meeting was chaired by D. J. Williams. Copies of the two letters of invitation (from Ben-Dov and Koteja) were provided for consideration to the participants the day before. Williams gave a summary of the merits of each proposed site: Bet-Dagan/Rehovot, Israel, and Cracow, Poland. The voting favored Cracow, Poland, for 1990. It was the consensus to consider Israel for the 1994 symposium, if the Israeli colleagues still wanted to host it. Because Koteja was not able to be present, I summarized the suggestions received from participants for the 1990 meeting, and discussed these with E. Podsiadlo, and forwarded the suggestions for consideration to Chairman Koteja and his new Local Organization Committee.

The Final Business Meeting was cochaired by Tranfaglia and me. I expressed the heartfelt thanks of the symposium participants to Director Viggiani, also to professors Tranfaglia and Tremblay for organizing this meeting for our scientific community.

Williams read a resolution expressing thanks to the organizers of the symposium. The resolution was unanimously adopted by the participants of the symposium. Director Viggiani expressed his thanks to the participants, and the Fifth International Symposium of Scale Insect Studies was closed by Professor Tranfaglia.

Please plan now to attend ISSIS-VI in Cracow, Poland, during 1990. Dr. Koteja extended a cordial invitation to each of us in a recent letter. ISSIS-VI will be held in an attractive setting at the 100-year-old Academy of Agriculture, probably during the first week in September. By then the tourists will have gone, but the Academy will not yet have opened for classes, so participants may be able to stay in inexpensive dormitories and/or in nearby hotels, and have meals in student cafeterias, to reduce cost. The close by Tatra and Pieniny mountains should provide excellent possibilities for excursions and collecting trips. Those who need more information and want to share ideas about ISSIS-VI with Dr. Jan Koteja (Institute of Applied Zoology, Academy of Agriculture al. Mickiewicz 24, PL-30-059, Cracow, Poland) should write to him directly.

Note: A more detailed summary of ISSIS-V will be printed as part of the Symposium Proceedings.

THE COCCOIDEA SLIDE COLLECTION AT VIRGINIA POLYTECHNIC INSTITUTE
AND STATE UNIVERSITY, BLACKSBURG

Part II - Families Eriococcidae through Stictococcidae

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Part I - Families Aclerdidae through Diaspididae was published in December 1985 in volume XI, No. 1 of 'The Scale,' by M. Kosztarab and M. H. Rhoades. This part concludes the present inventory of our slide collection. For the benefit of those who do not have access to the first publication we have repeated the general information on the collection, our loan policies, and on the abbreviations used.

The collection was begun in 1962 with M. Kosztarab's appointment as curator of the insect collections in this department and with the donation of his private collection. The Coccoidea collection at the present includes approximately 54,000 specimens mounted on about 21,000 slides, including 561 types and 483 voucher specimens. The voucher specimens resulted from research completed in our coccidology laboratory. The specimens, with few exceptions, are permanently mounted in Canada balsam. A large part of the slides resulted from research work completed with M. Kosztarab by his former graduate students: R. G. Baer, R. C. Brachman, S. W. Bullington, A. D'Ascoli, A. B. Hamon, J. O. Howell, P. L. Lambdin, M. L. Williams, P. A. Willoughby, and S. P. Yang. Slides from the work of D. K. Pollet, a former PhD candidate in this department, and S. Afifi, a postdoctoral fellow who worked with M. Kosztarab, are also in our collection. Several colleagues who retired from this department donated their slide collections to us, namely J. M. Amos, M. L. Bobb, J. McD. Grayson, C. H. Hill, and W. S. Hough.

A significant part of the paratype material along with other valuable slides was donated by R. F. Wilkey. Many other people, too numerous to list, sent us donations of slides throughout the past 24 years.

The Coccoidea collection is of world-wide scope and as such its significance for researchers was recognized by the National Science Foundation providing a grant (BSR-8317213) for its improvement and greater utilization, along with other parts of the university's insect collection.

The original spelling of generic and specific names found on slides has been retained, except for correcting a few obvious misspellings.

Requests for loans of Coccoidea material should be sent to M. Kosztarab. Loans are made for a period of six months, unless requested and approved otherwise. Holotypes, and part of the paratypes resulting from our work and/or from specimens loaned from this collection, are, as a rule, donated to the U.S. National Museum of Natural History.

Our slide-mounted specimens are often complemented by dry or alcohol-preserved material in our collection. Exchanges of new material for our duplicate slides are encouraged.

TYPE ABBREVIATIONS: C=Cotype, N=Neotype, P=Paratype or Paralectotype,
T=Type material, TT=Topotype

STAGE ABBREVIATIONS: The number of specimens and their stages are given in
parenthesis for type slides. f=adult female, m=adult male,
n=nymph, p=pupa

Genus	Species	Author	No. of Types	No. of Slides
ERIOCOCCIDAE				
Acalyptococcus	eugeniae	Lambdin & Kosztarab	7(20f)P	
Acanthococcus	aceris	Signoret		2
Acanthococcus	desertus	Matesova		1
Acanthococcus	greeni	(Newstead)		1
Acanthococcus	munroi	Boratynski		4
Acanthococcus	roboris	(Goux)		1
Capulinia	jaboticabae	(Ihering)		1
Cylindrococcus	casuarinae	Maskell		1
Eriococcus	adenostomae	Ehrhorn		1
Eriococcus	araucariae	(Maskell)		1
Eriococcus	arenosus	Cockerell		1
Eriococcus	azaleae	Comstock		9
Eriococcus	borealis	Cockerell		1
Eriococcus	carolinae	Williams		3
Eriococcus	coccineus	Cockerell		5
Eriococcus	coriaceus	Maskell		1
Eriococcus	cryptus	(Cockerell)		1
Eriococcus	diaboli	Ferris		1
Eriococcus	dubius	Cockerell		1
Eriococcus	eriogoni	Ehrhorn		1
Eriococcus	euphorbiae	Ferris		2
Eriococcus	eurythrix	Miller&Gonzalez	1(2f)P	
Eriococcus	galapagoensis	Morrison		1
Eriococcus	gerbergi	McDaniel		1
Eriococcus	gillettei	Tinsley		2
Eriococcus	kemptoni	Parrott		1
Eriococcus	larrae	Parrott&Cockerell)		1
Eriococcus	missouri	Hollinger		1
Eriococcus	pittospori	Ferris		1
Eriococcus	quercus	(Comstock)		19
Eriococcus	texanus	(King)		2
Eriococcus	undetermined			5
Gossyparia	spuria	(Modeer)		11
Greenisca	brachypodii	Borchsenius&Danzig		1
Greenisca	glyceriae	(Green)		5
Greenisca	gouxii	(Balachowsky)		3
Greenisca	rubra	Matesova		1
Hypericicoccus	hyperici	(Ferris)		2

Madarococcus	viridulus	Hoy		1
Opisthocelis	undetermined			1
Oregmopyga	erigoni	Miller		1
Oregmopyga	johnsoni	Miller		1
Ovaticoccus	agavium	(Douglas)		1
Rhizococcus	cingulatus	(Kiritchenko)		1
Rhizococcus	cynodontis	(Kiritchenko)		5
Rhizococcus	inermis	(Green)		1
Rhizococcus	insignis	(Newstead)		3
Rhizococcus	pseudinsignis	Green		3
Eriococcidae	undetermined			9
HALIMOCOCCIDAE				
Halimococcus	thebaicae	Hall		1
KERMESIDAE				
Allokermes	branigani	(King)	2(2f)TT; 3(16n)TT	23
Allokermes	cueroensis	(Cockerell)	2(2f)TT	54
Allokermes	essigi	(King)	1(1f)P; 1(1f)TT	58
Allokermes	ferrisi	Bullington&Kosztarab		15
Allokermes	galliformis	(Riley)	23(52f)TT; 1(6n)P	727
Allokermes	gillettei	(Cockerell)		37
Allokermes	grandis	(Cockerell)	2(2f)P; 1(2n)P	
Allokermes	kingi	(Cockerell)	1(1f)TT	510
Allokermes	kosztarabi	(Baer)	1(1f)TT; 2(9n)P	23
Allokermes	nivalis	(King&Cockerell)	1(3f)TT	
Allokermes	rattani	(Ehrhorn)		17
Fulbrightia	gallicola	Ferris	2(2n)T	
Kermes	bacciformis	Leonardi		5
Kermes	cockerelli	Ehrhorn	4(22n)C	18
Kermes	concinulus	Cockerell	1(1n)T	23
Kermes	corticalis	Nassonov		1
Kermes	nudus	Bullington&Kosztarab	17(26f)P	
Kermes	pinus	Baer&Kosztarab	4(15n)P	
Kermes	quercus	(Linnaeus)		5
Kermes	rimarum	Ferris	15(31n)P; 3(14n)TT	30
Kermes	roboris	Fourcroy		6
Kermes	shastensis	Ehrhorn	6(35n)TT; 2(2f)TT	11
Kermes	sylvestris	(Cockerell&King)	6(6f)TT	22
Kermes	vermilio	Planchon		9
Kermes	undetermined			97
Nanokermes	folium	Bullington&Kosztarab	119(266f)P; 2(5n)P	2
Nanokermes	iselini	(Baer&Kosztarab)	13(25f)P; 4(17n)P	8
Nanokermes	pubescens	(Bogue)	3(16n)P; 2(7n)TT	89
Olliffiella	cristicola	Cockerell		7
Olliffiella	secunda	Ferris	4(10n)P	9
Physeriococcus	cellulosus	Borchsenius	3(3f)P; 1(9n)P	

KERRIDAE

Austrotachardia	acaciae	(Maskell)		1
Austrotachardia	melaleucaae	(Maskell)		1
Laccifer	lacca	(Kerr)		1
Tachardiella	mexicana	(Comstock)		1
Tachardiella	undetermined			1
Tachardina	affluens	(Brain)		1
Kerridae	undetermined			6

LECANODIASPIDIDAE

Anomalococcus	cremastogastris	Green		8
Anomalococcus	indicus	Ayyar Ramakrishna		3
Anomalococcus	undetermined			1
Celaticoccus	cantentulatus	(Froggatt)	2(3f)P; 5(84n)P	
Lecanodiaspis	acaciae	(Maskell)		7
Lecanodiaspis	africana	Newstead		13
Lecanodiaspis	albilineata	Williams&Kosztarab	4(7f)P; 5(17n)C	
Lecanodiaspis	anomala	(Green)	1(1f)T; 4(14n)T	2
Lecanodiaspis	baculifera	Leonardi		5
Lecanodiaspis	brabei	Brain		16
Lecanodiaspis	convexa	Froggatt	3(3f)T; 1(5n)T	5
Lecanodiaspis	cremastogastris	Takagi	3(4f)TT	4
Lecanodiaspis	dealbatae	Lambdin	1(1f)P	
Lecanodiaspis	dendrobii	(Douglas)		12
Lecanodiaspis	dilatata	Froggatt		2
Lecanodiaspis	distincta	Howell&Kosztarab	2(3f)P	5
Lecanodiaspis	elongata	Ferris	2(2f)T	
Lecanodiaspis	eremocitri	Howell&Kosztarab	4(4f)P	1
Lecanodiaspis	erica	Hodgson		2
Lecanodiaspis	erratica	DeLotto		1
Lecanodiaspis	esperancei	Howell	1(1f)P	
Lecanodiaspis	eucalypti	(Maskell)		7
Lecanodiaspis	frenchi	Froggatt		8
Lecanodiaspis	hodgsoni	Howell&Kosztarab	2(2f)T	
Lecanodiaspis	ingae	Howell&Kosztarab	4(5f)P; 1(4n)P	
Lecanodiaspis	magna	Brain	6(9f)T	
Lecanodiaspis	malaboda	Green		3
Lecanodiaspis	melaleucaae	(Fuller)	2(3f)P; 2(6n)T	
Lecanodiaspis	microcribraria	Froggatt	4(7f)T; 4(8n)T	
Lecanodiaspis	mimosae	(Maskell)		7
Lecanodiaspis	mimusopis	Green	1(1n)T	
Lecanodiaspis	murphyi	Lambdin	2(7f)P	1
Lecanodiaspis	parthenii	Howell&Kosztarab	9(15f)P; 3(10n)P	
Lecanodiaspis	prosopidis	(Maskell)		144
Lecanodiaspis	prosopidis complex			453
Lecanodiaspis	quercus	Cockerell		12
Lecanodiaspis	rufescens	(Cockerell)		68
Lecanodiaspis	rugosa	Hempel	4(7f)T; 3(14n)T	3
Lecanodiaspis	russellae	Howell&Kosztarab	9(12f)P; 3(12n)T	
Lecanodiaspis	sardoa	Targioni-Tozzetti		4
Lecanodiaspis	subterranea	Kosztarab&Howell	1(1f)P; 1(4n)P	

Lecanodiaspis	tarsalis	Newstead		8
Lecanodiaspis	thamnosmae	Ferris	3(5f)T; 1(2n)T	7
Lecanodiaspis	varioseta	Howell&Kosztarab	4(4f)P; 2(4f)T	4
Lecanodiaspis	yuccae	Townsend	2(2f)T	20
Lecanodiaspis	zygophylli	Hodgson	2(2f)P	
Psoraleococcus	morrisoni	(Takahashi)	1(2f)P	
Psoraleococcus	multicribratus	Lambdin&Kosztarab	4(4f)P; 1(1n)P	
Psoraleococcus	multiporti	(Morrison)		15
Stictacanthus	azadirachtae	(Green)		2
Virgulicoccus	americanus	(Leonardi)		3
MARGARODIDAE				
Crypticerya	rosae	(Riley&Howard)		1
Crypticerya	undetermined			1
Drosicha	undetermined			1
Eurhizococcus	undetermined			1
Icerya	littoralis	Cockerell		1
Icerya	purchasi	Maskell		3
Icerya	rileyi	Cockerell		1
Icerya	seychellarum	Westwood		1
Icerya	undetermined			3
Kuwania	quercus	(Kuwana)		1
Llaveia	bourari	(Signoret)		1
Margarodes	formicarum	Guilding		1
Margarodes	meridionalis	Morrison		1
Matsucoccus	acalyptus	Herbert		1
Matsucoccus	bisetosus	Morrison		1
Matsucoccus	gallicolus	Morrison		4
Matsucoccus	matsumurae	(Kuwana)		2
Matsucoccus	paucicicatrices	Morrison		1
Neostengelia	texana	Morrison		13
Pityococcus	ferris	McKenzie		1
Porphyrophora	hamelii	(Brandt)		3
Steatococcus	undetermined			1
Stomacoccus	platani	Ferris		1
Tessarobelus	guerinii	Montrouzier		1
Xylococcinae	undeter. (nymphs)			4
Xylococculus	alni	(Florence)		1
Xylococculus	betulae	Pergande		12

Xylococcus	macrocarpae	Coleman		2
Xylococcus	filiferus	Löw		1
Margarodidae	undetermined			2
ORTHEZIIDAE				
Arctorthezia	cataphracta	(Olafsen)		5
Arctorthezia	pseudoccidentalis	Morrison		2
Newsteadia	americana	Morrison	1(4f)P	
Newsteadia	floccosa	(deGeer)		6
Newsteadia	minima	Morrison		1
Nipponorthezia	obscura	Morrison	1(1f)T	1
Orthezia	annae	Cockerell		2
Orthezia	artemisiae	Cockerell		2
Orthezia	insignis	Browne		2
Orthezia	lasiorum	Cockerell		1
Orthezia	newcomeri	Morrison		1
Orthezia	nigrocincta	Cockerell		1
Orthezia	praelonga	Douglas		4
Orthezia	solidaginis	Sanders		1
Orthezia	tillandsiae	Morrison		1
Orthezia	urticae	(Linneaus)		4
Ortheziola	vej dovskyi	Šulc		1
Ortheziidae	undetermined			5
PHOENICOCOCCIDAE				
Phoenicococcus	marlatti	Cockerell		2
Phoenicococcidae	undetermined			2
PSEUDOCOCCIDAE				
Allococcus	vovae	(Nassonov)		2
Amonostherium	lichtensioides	(Cockerell)		11
Anisococcus	adenostomae	(Ferris)		1
Anisococcus	ephedrae	(Coquillett)		1
Anisococcus	oregonensis	Ferris		1
Anisococcus	quercus	(Ehrhorn)		1
Antonina	australis	Green		3
Antonina	boutelouae	Parrott		18
Antonina	crawii	Cockerell		29
Antonina	graminis	(Maskell)		84
=Antonina	indica	Green		
Antonina	maritima	Green		2
Antonina	natalensis	Brain		11
Antonina	pretiosa	Ferris		40
Antonina	purpurea	Signoret		15
Antonina	transvaalensis	Brain		1
Antonina	vera	Borchsenius		1
Antonina	zonata	Green		37

Antonina	undetermined			129
Antoninella	inaudita	Kiritchenko		4
Antoninoides	parrotti	(Cockerell)		23
Atrococcus	achilleae	(Kiritchenko)		1
Atrococcus	bejbienkoi	Kozár&Danzig		3
Atrococcus	cracens	Williams		2
Atrococcus	paludinus	(Green)		3
Cataenococcus	guatemalensis	Ferris		1
Cataenococcus	ingrandi	Balachowsky		1
Cataenococcus	olivaceus	(Cockerell)		1
Centroccoccus	insolitus	(Green)		1
Chaetococcus	bambusae	(Maskell)		27
Chaetococcus	phragmitis	(Marchal)		9
Chaetococcus	sulci	(Green)		8
Chaetococcus	turanicus	Borchsenius		1
Chaetococcus	undetermined			3
Chnaurococcus	danzigae	Kozár&Kosztarab	2(2f)P	
Chnaurococcus	subterraneus	(Newstead)		2
Chnaurococcus	trifolii	(Forbes)		1
Chnaurococcus	villosus	(Ehrhorn)		1
Chorizococcus	abortivus	McKenzie	1(1f)P	
Chorizococcus	abroniae	McKenzie		1
Chorizococcus	brevicruris	McKenzie		1
Chorizococcus	californicus	McKenzie		1
Chorizococcus	coniculus	Miller&McKenzie	1(1f)P	
Chorizococcus	fistulosus	McKenzie	1(1f)P	
Chorizococcus	lounsburyi	(Brain)		4
Chorizococcus	polyporus	McKenzie		1
Chorizococcus	psoraleae	McKenzie	1(1f)P	
Chorizococcus	rostellum	(Hoke)		11
Chorizococcus	variabilis	McKenzie	1(1f)P	
Chorizococcus	viktorina	Kozár		1
Chorizococcus	wilkeyi	McKenzie	1(1f)P	
Claviccoccus	tribulus	Ferris		2
Coccura	comari	Künow		1
Crisicoccus	azaleae	(Tinsley)		3
Crisicoccus	matsumotoi	(Shiraiwa)		2
Crisicoccus	pini	(Kuwana)		2
Cryptoripersia	salina	(Ehrhorn)		1
Cucullococcus	vaccinii	Ferris		2
Discococcus	graminis	(Ferris)		1
Discococcus	hopi	Ferris		1
Discococcus	spectabilis	McKenzie		1

Distichlicoccus	californicus	(Ehrhorn)		1
Distichlicoccus	dasychloae	Ferris		1
Distichlicoccus	salinus	(Cockerell)		1
Dysmicoccus	alazon	Williams		1
Dysmicoccus	boninsis	(Kuwana)		1
Dysmicoccus	brevipes	(Cockerell)		4
Dysmicoccus	diodium	(McConnell)		1
Dysmicoccus	neobrevipes	Beardsley		1
Dysmicoccus	obesus	(Lobdell)		1
Dysmicoccus	pinicolus	McKenzie		1
Dysmicoccus	polymeris	Miller & McKenzie	1(1f)P	
Dysmicoccus	ryani	(Coquillett)		1
Dysmicoccus	timberlakei	(Cockerell)		1
Dysmicoccus	walkeri	(Newstead)		2
Dysmicoccus	wistariae	(Green)		1
Dysmicoccus	undetermined			6
Ehrhornia	cupressi	(Ehrhorn)		8
Epicoccus	acaciae	(Maskell)		1
Erium	globosum	Maskell		1
Euripersia	tomlini	(Newstead)		3
Eurycoccus	jessica	(Hollinger)		1
Ferrisia	virgata	(Cockerell)		23
Geococcus	coffea	Green		2
Heliococcus	bohemicus	Sulc		2
Heliococcus	deserticola	Miller	1(1f)P	
Heliococcus	insignis	(Lobdell)		1
Heliococcus	stachyos	(Ehrhorn)		2
Heliococcus	sulcii	Goux		1
Heterococcus	nudus	Green		5
Heterococcus	tuttlei	Miller & McKenzie	1(1f)P	
Heterococcus	raui	Miller	1(1f)P	
Humococcus	atriplicis	Ferris		1
Humococcus	caritus	McKenzie	1(1f)P	
Humococcus	hilariae	(Ferris)		1
Humococcus	inornatus	McKenzie		1
Humococcus	porterae	(Cockerell)		1
Hypogeococcus	othnius	Miller & McKenzie		1
Idiococcus	bambusae	Takahashi & Kanda		1
Kuwanina	parva	(Maskell)		2
Lachnodius	eucalypti	(Maskell)		1

Laminococcus	giffardi	(Ehrhorn)		3
Leptococcus	metroxyli	Reyne		1
Longicoccus	psammophilus	(Koteja)	1(1f)P	
Macrocerococcus	superbus	Leonardi		3
Metadenopus	festucae	Šulc		3
Mirococcopsis	elongatus	Borchsenius		2
Mirococcopsis	nagyi	Kozár	1(2f)P	
Mirococcopsis	stipae	Borchsenius		3
Misericoccus	arenarius	(Deane&Steinweiden)		1
Nipaecoccus	aurilanatus	(Maskell)		1
Nipaecoccus	nipae	(Maskell)		5
Nipaecoccus	viridis	(Newstead)		1
Oracella	acuta	(Lobdell)		7
Palmicultor	palmarum	(Ehrhorn)		1
Paludicoccus	distichlium	(Kuwana)		2
Paracoccus	burnerae	(Brain)		1
Paradoxococcus	macdanieli	McKenzie		1
Paroudablis	evelinae	Tereznikova		1
Paroudablis	interruptus	(Green)		1
Paroudablis	piceae	(Löw)		1
Pedronia	cibotii	Beardsley		1
Pedronia	strobilanthis	Green		1
Pedronia	tenuispina	Green		1
Peliococcus	balteatus	(Green)		2
Peliococcus	serratus	(Ferris)		6
Phenacoccus	aceris	Signoret		9
Phenacoccus	celtisifoliae	Hollinger		1
Phenacoccus	colemani	Ehrhorn		2
Phenacoccus	dearnessi	King		2
Phenacoccus	dicoriae	McKenzie		1
Phenacoccus	eremicus	Ferris		1
Phenacoccus	eriongoni	Ferris		1
Phenacoccus	eschscholtziae	McKenzie		1
Phenacoccus	flaveolus	(Cockerell)		1
Phenacoccus	giganteus	McKenzie	1(1f)P	
Phenacoccus	gossypii	Townsend & Cockerell		8
Phenacoccus	graminosus	McKenzie		1
Phenacoccus	harbisoni	Peterson	1(1f)P	
Phenacoccus	helianthi	(Cockerell)		1
Phenacoccus	hordei	(Lindeman)		4
Phenacoccus	infernalis	McKenzie		1

Phenacoccus	lotearum	McKenzie		1
Phenacoccus	manihoti	Matile-Ferrero		1
Phenacoccus	milleri	McKenzie	1(1f)P	
Phenacoccus	minimus	Tinsley		1
Phenacoccus	nonarius	McKenzie		2
Phenacoccus	phenacoccoides	(Kiritchenko)		1
Phenacoccus	rubivorus	(Cockerell)		19
Phenacoccus	solani	Ferris		14
Phenacoccus	solenopsis	Tinsley		13
Phenacoccus	undetermined			3
Planococcoides	ireneus	DeLotto		2
Planococcoides	robustus	Ezzat & McConnell		2
Planococcus	citri	(Risso)		61
Planococcus	kenyae	(LePelley)		1
Planococcus	kraunhiae	(Kuwana)		1
Planococcus	lilacinus	(Cockerell)		2
Plotococcus	eugeniae	Miller&Denno		1
Poecilococcus	longilobus	Brookes	1(1f)P	
Porococcus	tintorius	Cockerell		1
Pseudantonina	bambusae	Green		2
Pseudantonina	poae	(Maskell)		1
Pseudococcus	aberrans	McKenzie		1
Pseudococcus	affinis	Maskell		17
Pseudococcus	beardsleyi	Miller & McKenzie	1(1f)P	
Pseudococcus	brevipes	Cockerell		5
Pseudococcus	calceolariae	(Maskell)		4
Pseudococcus	citriculus	Green		2
Pseudococcus	comstocki	Kuwana		32
Pseudococcus	dentatus	Lobdell		1
Pseudococcus	diversus	McKenzie		1
Pseudococcus	elisae	Borchsenius		2
Pseudococcus	floriger	Ferris		1
Pseudococcus	importatus	McKenzie		4
Pseudococcus	landoi	(Balachowsky)		2
Pseudococcus	longispinus	(Targioni-Tozzetti)		4
Pseudococcus	maritimus	(Ehrhorn)		22
Pseudococcus	microcirculus	McKenzie	1(1f)P	
Pseudococcus	montanus	Ehrhorn		2
Pseudococcus	palmarum	Ehrhorn		1
Pseudococcus	pipturicolus	Beardsley		1
Pseudococcus	sorghiiellus	(Forbes)		4
Pseudococcus	sparsus	McKenzie		2
Pseudococcus	straussiae	Ehrhorn		1
Pseudococcus	undetermined			15
Radicoccus	kelloggi	(Ehrhorn)		2
Rastrococcus	undetermined			1
Rhizoecus	albidus	Goux		1

Rhizoecus	americanus	(Hambleton)		2
Rhizoecus	bicirculus	McKenzie		1
Rhizoecus	bituberculatus	McKenzie	1(1f)P	
Rhizoecus	cacticans	Hambleton		2
Rhizoecus	californicus	Ferris		1
Rhizoecus	campestris	Hambleton		1
Rhizoecus	falcifer	(Künckel d'Herculeis)		2
Rhizoecus	floridanus	Hambleton	1(2f)P	
Rhizoecus	gracilis	McKenzie		1
Rhizoecus	graminis	(Hambleton)		1
Rhizoecus	kondonis	Kuwana		1
Rhizoecus	leucosomus	(Cockerell)		1
Rhizoecus	pritchardi	McKenzie		1
Rhizoecus	sonomae	McKenzie		1
Rhizoecus	undetermined			2
Ritsemia	pupifera	Lichtenstein		1
Saccharicoccus	penium	Williams		3
Saccharicoccus	sacchari	(Cockerell)		7
Scaptococcus	californicus	McKenzie	1(1f)P	
Scaptococcus	milleri	McKenzie		1
Serrolecanium	sasae	(Siraiwa)		11
Serrolecanium	tobai	(Kuwana)		2
Sphaerococcus	casuarinae	Maskell		3
Sphaerococcus	floccosus ms. sp.	Green		3
Sphaerococcus	undetermined			2
Spilococcus	andersoni	(Coleman)		1
Spilococcus	atriplicis	Cockerell		1
Spilococcus	cactearum	McKenzie	1(1f)P	
Spilococcus	ceanothi	McKenzie		1
Spilococcus	erionomi	(Ehrhorn)		2
Spilococcus	gutierreziae	(Cockerell)		1
Spilococcus	implicatus	Ferris		1
Spilococcus	juniperi	(Ehrhorn)		2
Spilococcus	keiferi	McKenzie		1
Spilococcus	larreae	Ferris		3
Spilococcus	parkeri	McKenzie	1(1f)P	
Spilococcus	pressus	Ferris		1
Spilococcus	prosopidis	(Cockerell)		1
Spilococcus	proteae	Hall		1
Spilococcus	sequoiae	(Coleman)		1
Spinococcus	calluneti	(Lindinger)		1
Spinococcus	cacteorum	McKenzie		1
Spinococcus	marrubii	(Kiritchenko)		3
Syrmococcus	spirapunctus	(Lobdell)		1
Tridiscus	distichlii	Ferris		1
Tridiscus	multiorbis	Ferris		1
Trionymus	aberrans	(Goux)		4

Trionymus	americana	(Cockerell)		2
Trionymus	caricis	McConnell		1
Trionymus	clandestinus	McConnell		4
Trionymus	diminutus	(Leonardi)		4
Trionymus	dolus	Ferris		1
Trionymus	haancheni	McKenzie	1(1f)P	
Trionymus	idahoensis	Miller & McKenzie	1(2f)P	
Trionymus	magnus	(Cockerell&Cockerell)		
Trionymus	mocus	Ferris	1(1f)P	
Trionymus	multivorus	(Kiritchenko)		3
Trionymus	newsteadi	Green		2
Trionymus	perrisii	(Signoret)		2
Trionymus	quadricirculus	McKenzie		1
Trionymus	smithii	(Essig)		2
Trionymus	trifolii	Forbes		6
Trionymus	thulensis	Green		2
Trionymus	utahensis	(Cockerell)		2
Trionymus	undetermined			5
Vryburgia	distincta	(DeLotto)		2
Pseudococcidae	undetermined			104
PUTOIDAE				
Puto	acirculus	McKenzie	1(1f)P	4
Puto	albicans	McKenzie	4(4f)T	
Puto	arctostaphyli	Ferris		1
Puto	atriplicis	McKenzie		1
Puto	bryanthi	Ferris		1
Puto	cupressi	(Coleman)		1
Puto	decorosus	McKenzie		1
Puto	echinatus	McKenzie		1
Puto	mexicanus	Cockerell		1
Puto	pacificus	McKenzie		1
Puto	pilosellae	(Šulc)		4
Puto	simmondsiae	McKenzie		1
Puto	ulter	Ferris		3
Puto	yuccae	Coquillett		3
STICTOCOCCIDAE				
Stictococcus	diversiseta	Silvestri		1
Stictococcus	sjostadti	Cockerell		1
Stictococcus	undetermined			1

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I wish again to express my sincere appreciation to Mrs. Helen Proctor for preparing the first draft of this list of publications.

The following publications are those received since the last "Scale" was sent out in December 1985. The list also includes references gathered from a "current awareness" computerized literature search provided by the U.S. Department of Agriculture. If you are aware of papers published in 1984-1985 that are not included in this issue or the previous one, please let me know. Please continue to send reprints of your papers for assurance of inclusion in the "Scale".

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