

LIBRARY OF WELLESLEY COLLEGE



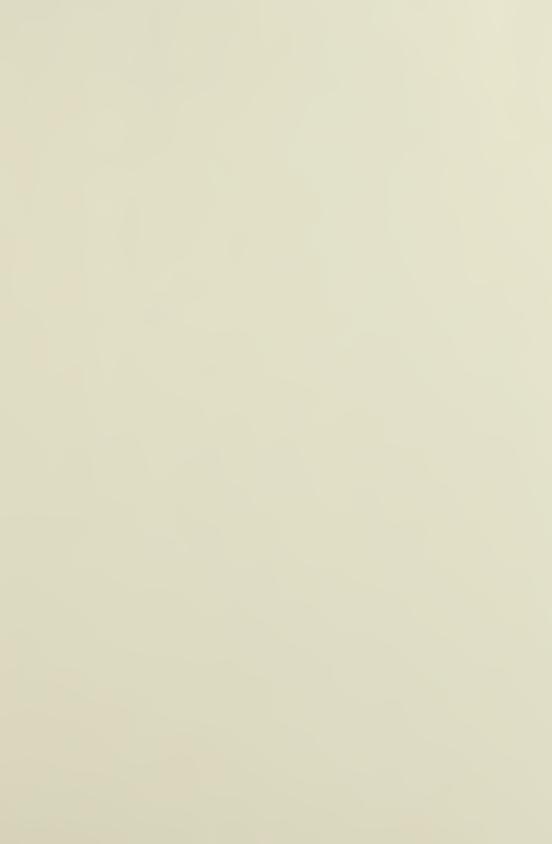
Preservation photocopied with funds from the Barbara Lubin Goldsmith Library Preservation Fund



Digitized by the Internet Archive in 2014











THE RESERVE OF

- WHI

Hibrary of

Millestry Lines Cullege.

Purchased from

The Horstord Kund.













Wild Barley or Squirrel Tail (Hordeum jubatum). Ia. Agr. Exp. Sta.

A Manual of Poisonous Plants

Chiefly of Eastern North America, with Brief Notes on Economic and Medicinal Plants, and Numerous Illustrations

L. H. PAMMEL, Ph. D.

Professor of Botany, Iowa State College Agriculture and Mechanic Arts



THE TORCH PRESS CEDAR RAPIDS, IOWA 1911 COPYRIGHT BY
L. H. PAMMEL
1911

SCIENCE LIBRARY

9 QK

100

U6P32

1911a



FOREWORD

12

During the last decade, there has been much interest manifested in regard to plants injurious to live stock. Numerous contributions have been made along this line, notably by Dr. Chesnut, formerly of the United States Department of Agriculture, Drs. True and Wilcox and their co-workers also of Washington, Dr. Schaffner of the Ohio State University, Dr. Jones of the Vermont Agricultural Experiment Station, Dr. Kennedy of the Nevada Experiment Station, Dr. Nelson of the Wyoming Station, Dr. Nelson of Washington, and Drs. Peters and Bessey of Nebraska. Other station botanists have also contributed to the same line of work. Much of the literature is scattered, however, hence an effort has been made to bring together in the following pages the results obtained.

Much information on this line of investigation has also been obtained from such works as Millspaugh's Medicinal Plants of North America, Dr. Johnson's Manual of the Medical Botany of North America, Lloyd's Drugs and Medicines of North America, Winslow's Veterinary Materia Medica and Therapeutics, Sayre's Organic Materia Medica and Pharmacognosy, Flückiger and Hanbury's Pharmacographia, Greenish's Materia Medica, Ellingwood's Materia Medica, Therapeutics and Pharmacognosy, Pereira's Materia Medica, Luerssen's and Czapek's publications and many others which give details in regard to the effects of poisonous plants.

Many persons may object to the great number of plants which are here regarded as poisonous or described as such in this work. I have placed the broadest interpretation on the subject and have, therefore, included all plants that are injurious although many of these are not known to produce poisons, some even being most useful economic plants and yet injurious to some people.

It has been thought best to arrange the manual so as to consider the plants in the same order as that given in Engler and Prantl's Die Pflanzenfamilien.

The Schizomycetes were contributed by my colleague, Dr. R. E. Buchanan, who has also favored me in many other ways.

The parts concerning the blue-green algae and algae, taking up the higher algae and their relation to the water slimes are given in their sequence under the Schizophyceae and Euphyceae.

The Eumycetes or true fungi are considered chiefly from the pathogenic standpoint; while other fungi are referred to and briefly considered under their respective groups. In regard to the higher fungi, such as the toadstools, much valuable information may be obtained from the works of Dr. Farlow, Prof. Peck and Prof. Atkinson.

The so-called Blastomycetic fungi have been arranged under the group of the imperfectly known forms. There are also brief characterizations of other groups of the cryptogams such as lichens, mosses, ferns and their allies. The flowering plants or Spermatophyta are described more fully than the previous groups, and under the various orders and subdivisions we have also added notes on economic and medicinal plants. Those who desire fuller information in regard to other American species of the different orders described should consult the latest editions of Gray's or Britton's Manuals, in which the descriptions are full and complete. Of course, one cannot expect to add much to the excellent descriptions given in these treatises.

I have freely made use of published literature in the systematic portions as well as in the more technical matter pertaining to poisons, and I wish to acknowledge my assistance from these sources.

In order that the species named may be more readily recognized, a large number have been figured.

In addition to the descriptive part of the work there has been added a chapter on the active principles of plants, by my colleague, Prof. A. A. Bennett. I am also greatly indebted to Miss Harriette S. Kellogg who has carefully read the manuscript and assisted me in other ways besides preparing the bibliography. To Dr. Trelease of the Missouri Botanical Garden, I am indebted for the use of a number of books on the subject. Miss Charlotte M. King, Miss Ada Hayden, Mr. W. S. Dudgeon, and my daughter, Lois, have made drawings especially for this work. Other illustrations are taken from special works, due credit being given in each case.

I am indebted for the use of cuts to the following persons: Dr. C. F. Curtiss of the Iowa Agricultural Experiment Station, Dr. B. D. Halsted of the New Jersey Agricultural Experiment Station, Profs. S. B. Green and Washburn of the Minnesota Station and Prof. A. D. Selby of the Ohio Station, and to the United States Department of Agriculture. Some have been reproduced from Baillon's Dict., and from Bentham's Handbook of British Plants. I have endeavored in each case to give credit for the drawing or cut.

I am indebted to Drs. R. R. Dykstra and C. H. Stange, Profs. L. G. Michael, C. V. Gregory and A. A. Bennett for proof reading and to Dr. W. H. Stuhr for some matter in Part I.

The work does not pretend to be complete; we hope, however, that it may prove useful to the Veterinarian, Physician and Layman.

Ames, Iowa, June 1, 1909.

TABLE OF CONTENTS

PART I

| CHAPTER I. Poisons and Statistics on Poisons | 1 |
|---|----|
| Ancient Use of Poisons, 2; The Rise of Chemistry and Poisons, 3; Ratze- burg on Poisonous Plants, 4; Statistics of Poisoning, 5; Statutes on Poisoning and Actions of Poisons on Different Animals, 6. | |
| CHAPTER II. BACTERIAI, POISONS | 8 |
| Impure Water the Source of Disease, 8; Bacterial Poisons, 10; Botulism, 11; Maidismus or Pellagra, 11. | |
| CHAPTER III. DERMATITIS | 12 |
| Dermatomycosis, 12; Dermatophytes of Domestic Animals, 13; Favus, 17. | |
| CHAPTER IV. FORAGE POISONING, ERGOTISM, AND ASPERGILLOSIS | 20 |
| Forage Poisoning, 20; Poisoning from Silage, 24; Ergotism, 28; Aspergillosis, 29. | |
| CHAPTER V. Peisoning from Fungi | 31 |
| Toadstools — Amanita muscaria, 31; Amanita phalloides, 31. | |
| CHAPTER VI. POISONING FROM OTHER PLANTS. EQUISETOSIS, LOCOISM, | |
| AND LUPINOSIS | 37 |
| Equisetosis, 37; Locoism, 37; Lupinosis, 40. | |
| CHAPTER VII. DELPHINOSIS, LATHYRISMUS (LATHYRISM), ACONITISM, | |
| VERATRISM, UMBELLIFERAE, CONIUM, CICUTA | 44 |
| Delphinosis, 44; Lathyrismus, 45; Aconitism, 46; Veratrism, 46; Poisoning from Umbelliferae, 49; Cowbane, 49; The Poison Hemlock, 51. | |
| CHAPTER VIII. FISH AND ARROW POISONS, HYDROCYANIC POISONING- | |
| TOXALBUMINS — BLACK LOCUST, CASTOR OIL, AND | |
| Jequirity | 52 |
| Fish and Arrow Poisons, 52; Hydrocyanic Poisoning, 53; Poisoning from Toxalbumins, Black Locust, Ricinus and Abrus, 55. | |
| CHAPTER IX. POISONING FROM OPIUM, SOLANACEAE AND PLANTS THAT | |
| CONTAIN SAPONINS | 59 |
| Opium, 59; Poisoning from Solanaccae; Jimson Weed; Hyoscyamin, 60; Poisoning from Plants that Contain Saponin; Bouncing Betty; Corncockle, 62. | |
| CHAPTER X. POISONING FROM FLOWERS, POISONING FROM HONEY, ME- | |
| CHANICAL INJURIES | 64 |
| Poisoning from Flowers, 64; Poisoning from Honey, 64; Mechanical Injuries; Wild Barley; Cheat; Needle Grass; Greasewood; Crimson Clover, 66. | |
| CHAPTER XI. CLASSIFICATION OF POISONS, SYMPTOMS, AND ANTIDOTES | 72 |
| Poisons which Cause Coarse Anatomical Changes of the Organs, 72; Classification of Poisons and Poisonous Symptoms According to Bernard H. Smith — Narcotics, 73; Deliriants, 73; Inebriants, 74; Convulsives, 74; Depressants, 74; Asthenics, 75; Purgatives, 75; Abortives, 76; Irritants with Nervous Symptoms, 76; Simple Irritants, 76. Table of Symptoms Observed After the Administration of Poisons, Adapted from Cattell After the Work of Kobert, 77; Treatment for Poisoning, 79. | |
| CHAPTER XII. THE PRODUCTION OF POISON IN PLANTS | 82 |
| The Use and Actions of Poisons on Plants, 82; Distribution of Poisonous Substances in Plants, 82; Light, 83; Heat, 83; Seasons, 83; Climatic, 85; Culture, 87; Related Species and Toxic Substances, 88; Distribution of Chemical Substances, 88; Saponins, 89; Hydrocyanic Acid, 89; Cumarin, 90; Cytisin, 90. | |
| | |

| CHAPTER XIII. ALGAE IN WATER SUPPLIES | 91 |
|---|-----|
| How Growth of Algae in Water May Be Prevented, 94; The Use of Algicites, 94. | |
| CHAPTER XIV. A CATALOGUE OF THE MORE IMPORTANT POISONOUS PLANTS | |
| of the United States and Canada | 96 |
| Literature, 96; Euthallophyta, Schizophyta, Schizomycetes. Bacteria, 97; Euphyceae — Algae, 98; Eumycetes — Fungi; Phycomycetes. Black Molds and Downy Mildews, 98; Smuts, 98; Rusts, 99; Toadstools, 99; Ascomycetes, 100; Fungi Imperfecti, 100; Embryophyta, Pteridophyta, 100; Embryophyta siphonogama; Flowering Plants; Gynnospermae, Conifers and Allics, 101; Angiospermae, 102; Monocotyledoncae, 102; Grass Family, 102; Lily Family, 103; Dicotyledoncae, 105; Nettle Family, 106; Pink Family, 107; Crowfoot Family, 108; Poppy Family, 112; Rose Family, 115; Pulse Family, 117; Spurge Family, 121; Poison Ivy Family, 122; Carrot Family, 126; Heath Family, 127; Nightshade Family, 131; Composite Family, 137. | |
| CHAPTER XV. CHEMISTRY OF ALKALOIDS, GLUCOSIDES, ETC | 143 |
| Composition and General Properties of Alkaloids, 143; Occurrence of the Alkaloids, 144; Classification of the Alkaloids, 145; On Alkaloids, 147; Glucosides, 148; Gluco-Alkaloids, 149; Saponins, 149; Picrotoxin, Cicutoxin and Toxins, 149; Other Vegetable Poisons, 149; Amins, 149; Organic Acids, 150. | |

TABLE OF CONTENTS

PART II

| KEY FOR PLANT KINGDOM | | | | | | | | 153–158 |
|--------------------------|------|-------|------|-----|----|--|--|---------|
| MYXOTHALLOPHYTA | | | | | | | | 158-160 |
| EUTHALLOPHYTA | | | | | | | | 160 |
| Schizophyta . | | | | | | | | 160 |
| SCHIZOMYCETES | | | | | | | | 161–184 |
| Schizophyceae | | | | | | | | 184–188 |
| FLAGELLATAE | | | | | | | | 188 |
| EUPHYCEAE | | | | | | | | 188-194 |
| PERIDINALES . | | | | | | | | 188 |
| Bacillariales | | | | | | | | 188 |
| Conjugatae . | | | | | | | | 189 |
| CHLOROPHYCEAE | | | | | | | | 190 |
| CHARALES . | | | | | | | | 193 |
| Рилеорнуселе | | | | | | | | 194 |
| DICTYOTALES . | | | | | | | | 194 |
| Rhodophyceae | | | | | | | | 194 |
| Eumycetes | | | | | | | | 195-307 |
| PHYCOMYCETES | | | | | | | | 195-209 |
| Zygomycetes | | | | | | | | 195-204 |
| Oomycetes | ٠. | | | | | | | 204-209 |
| Basidiomycetes | | | | | | | | 209 |
| Hemibasidii | | | | | | | | 210-220 |
| Eubasidii | | | | | | | | 220 |
| ASCOMYCETES . | | | | | | | | 247-281 |
| Fungi Imperfect | I | | | | | | | 281-306 |
| Lichens | | | | | | | | 307 |
| EMBRYOPHYTA ZOIDIOGAMA | | | | | | | | 308-325 |
| Bryophyta . | | | | | | | | 308-312 |
| PTERIDOPHYTA | | | | | | | | 312-325 |
| FILICALES | | | | | | | | 313-322 |
| Equisetales | | | | | | | | 322-325 |
| EMBRYOPHYTA SIPIIONOGAMA | of S | SPERM | ATOI | HYT | ١. | | | 325-802 |
| GYMNOSPERMAE | | | | | | | | 325-332 |
| CONIFERAE | | | | | | | | 327 |
| Anciospermae | | | | | | | | 332 |
| Monocotyledo | ONE | AE | | | | | | 332-395 |
| Pandana | | | | | | | | 332 |
| HELOBIAE | | | | | | | | 332 |
| Glumiflo | RAE | | | | | | | 336-369 |
| | | | | | | | | |

| | GRAMINEA | | | | • | | • | | • | 336–367 |
|---------------|--|-------|---|---|---|---|---|---|---|---------|
| | Cyperacea | Æ | | | | | | | | 367-369 |
| | Principes | | | | | | | | | 369-370 |
| | Spathiflorae | | | | | | | | | 370-372 |
| | FARINOSAE | | | | | | | | | 372-374 |
| | Liliiflorae | | | | | | | | | 374-390 |
| | Scitamineae | | | | | | | | | 390-392 |
| | Microspermae | | | | | | | | | 392-395 |
| | DICOTYLEDONEAE | | | | | | | | | 395-802 |
| | Archichlamy | DEAE | | | | | | | | 395-664 |
| | VERTICILLA | ATAE | | | | | | | | 395 |
| | Piperales | | | | | | | | | 396 |
| | SALICALES | | | | | | | | | 396-397 |
| | MYRICALES | 3 | | | | | | | | 397-399 |
| | BALANOPSI | DALES | 5 | | | | | | | 399 |
| | Leitneriai | | | | | | | | | 400 |
| | JUGLANDAL | ES | | | | | | | | 400-402 |
| | | | | | | | | | | 402-404 |
| | URTICALES | | | | | | | | | 404-415 |
| | PROTEALES | | | | | | | | | 415 |
| | Santalali | | | | | | | | | 415 |
| | Aristoloci | | S | | | | | | | 416 |
| | Polygonal | | _ | | | | | | | 417-423 |
| | CENTROSPE | | • | i | | | | | | 423-444 |
| | RANALES | | | • | · | • | • | | • | 414-479 |
| | RHOEADALI | | • | • | • | • | | | • | 479-496 |
| | SARRACENI | | • | | | • | | | • | 497-500 |
| | and the same of th | | • | • | | : | | • | • | 500-574 |
| | GERANIALE | | • | • | | • | | • | • | 574-604 |
| | Sapindale | | • | • | • | • | • | • | • | 604-620 |
| | RHAMNALI | | • | • | | | • | | • | 620-621 |
| | MALVALES | | • | • | | | • | | • | 621–627 |
| | Parietales | | • | • | | • | • | | • | 627-634 |
| | Opuntiale | | • | • | • | • | • | • | • | 634-637 |
| | Myrtiflor. | | • | • | | • | | | • | 637–645 |
| | Umbellali | | • | • | • | • | • | • | • | 645–664 |
| | METACHLAMYD | | • | • | • | • | • | • | | 664-802 |
| | ERICALES | • | • | • | | • | • | | • | 664-675 |
| | Primulale | | • | • | | • | • | | • | 675-679 |
| | EBENALES | | | • | | • | • | | • | 679-683 |
| | Contortae | | • | • | • | • | • | • | • | 683-697 |
| | Tubiflora | | • | • | • | • | • | • | • | 698–739 |
| | Plantagin | | | • | • | • | • | • | • | 739–740 |
| | _ ' | | | | • | • | • | • | • | 740–748 |
| | Rubiales Campanul | | | • | • | • | • | • | • | 748–802 |
| Camarograpica | Poisonous Plants | | | • | • | • | • | • | • | S03 |
| | of Poisonous Plants | | | | • | • | • | | • | 809-918 |
| | of Poisonous Plant | _ | | | • | • | • | • | • | 919 |
| INDEX | | • | • | • | • | • | | • | • | 919 |

LIST OF PLATES

PART I

| Wild Barley | or Sq | uirre | l Ta | il | | | | | | F_{r} | ontisp | iece |
|---------------|--------|-------|------|----|----|---------|----|--|---|----------|--------|------|
| Anemonella a | | | | | | | | | | opposite | page | 37 |
| Poison Ivy | | | | | | | | | | 44 | 11 | 123 |
| Mountain La | | | | | | | | | | 44 | 44 | 128 |
| Jimson Weed | | | | | | | | | | 44 | " | 133 |
| J | · | Ť | | | | | | | | | | |
| | | | | | D. | T) (11) | II | | | | | |
| | • | | | | FA | KT | 11 | | | | | |
| Honey Colore | ed Arı | nilla | ria | | | | | | | 44 | " | 235 |
| Amanita sp. | | | | | | | | | | 44 | 44 | 236 |
| Deadly Aman | | | | | | | | | | " | 66 | 239 |
| A. Yucca. B | | | | | | | | | | " | " | 375 |
| Death Camas | | | | | | | | | | 16 | " | 378 |
| Wild Indian | | | | | | | | | | ** | 44 | 381 |
| Pennsylvania | | | _ | | | | | | · | ** | ш | 421 |
| Wild Cherry | | | | | | | | | · | 66 | ** | 515 |
| Lupine in flo | | | | | | | | | | 44 | 66 | 546 |
| Great Basin 1 | | | | | | | | | | 4.6 | ** | 548 |
| Common Mil | | | | | | | | | | ee | ** | 697 |
| A. Rhododer | | | | | | | | | | | | 097 |
| Rhododei | | | • | - | | | | | | 46 | " | 667 |



ERRATA

On page 4, line 1, for "von Pragg," substitute "van Praag."

On page 26, line 40 should read, "the several investigators were not working," etc.

On page 29, line 14, for "tubercle," substitute "tuberculosis."

On page 89, line 22, for "Muscari," substitute "Muscaria."

On page 102, line 2, for "Swedish juniper," substitute "Savin."

On page 103, line 14, for "G. gussonianum," substitute "H. gussonianum."

On page 127, line 8, for "Cornus paniculata L'Her," substitute "Cornus Amomum, Mill, Kinnikinnik."

From page 189 to page 243, the heading "EUTHALLEPHYTA" should read "EUTHALLOPHYTA."

On page 201 omit Mucor ramosus, Lindt 1, and description following.

On page 243, lines 29, 33, for "Hand," substitute "Hard."

On page 344, line 13, for "Pamorusa," substitute "Palmarosa."

On page 395, line 31, for "oxogenous," substitute "exogenous."

On page 397, insert "MYRICALES" above Myricaceae, Sweet Gale Family.

On page 450, line 17, for "Dunstan and Anderson," substitute "Dunstan and Henry."

On page 522, Fig. 281 should be inverted.

On page 572, line 4, for "Convincin," substitute "Convicin." On page 877, No. 186, for "Cooke, N. C.," substitute "Cooke, M. C."

On page 884, No. 329 should read "Gerard, A. W..... Wanika, a new," etc.

On page 901, No. 715, for "1901," substitute "191."

Typographical errors in scientific names occurring in the text are corrected in the index.



Part I

A General Treatise on Poisonous Plants



CHAPTER I

POISONS AND STATISTICS ON POISONS

A poison has been defined as "Any substance that, when taken into the system acts in a noxious manner by means not mechanical, tending to cause death or serious detriment to health."

Kobert and other physicians define a poison as "A non-organized body, either organic or inorganic, which under certain conditions, affects temporarily or permanently one or more organs of the body, when in a state of health or in a healthy condition." Such poisons may develop in the body or may come from without. Some substances act injuriously in a mechanical way, that is, they may set up disturbances by irritating some parts of the body. Other substances, while poisonous to one who is ill, may be entirely harmless to persons or animals in a state of health.

Kobert also defines poisons from a pharmacological standpoint as "All pharmacological agents which, in a given case, do not act beneficially but injuriously."

Toxicology is the science of poisons, the word being derived from the ancient word "tox," meaning bow, or arrow, probably from the ancient use of the arrow to kill.

In tracing the application of the word "tox;" "arrow," to its later application, poison, Blyth says: "Perchance the savage found that weapons soiled with the blood of former victims made wounds fatal; from this observation, the next step naturally would be that of experiment,—the arrow or spear would be steeped in all manner of offensive pastes, and smeared with the vegetable juices of those plants which were deemed noxious; and as the effects were mysterious they would be ascribed to the supernatural powers, and covered with a veil of superstition."

The different tribes of Indians in South America have from early days been skilful in preparing arrow poisons, the majority of which contain strychnin in some form. The following plants have, at various times, furnished poisons for arrow tips, not only in South America, but also in other countries: Strychnos toxifera (Strychnine), perhaps the most generally used of any; Antiaris toxicaria, an arrow poison of Java, Borneo, and North Africa; various Leguminosae, as Erythrophloeum in Angola, Sierra Leone, and Seychelles, a different species being used in each place; Pithecolobium, Afzelia, and Derris elliptica of Borneo; of the Menispermaceae, two species of Abuta are used.

Perhaps in this connection, it would not be out of place to mention several fish-poisons, many of which are also legumes. Of this order are Albidizzia, Afzelia, Bauhinia, Enterolobium, Leucaena, Milletia, Piscidia, Acacia, Abrus precatorius, Clitoria, Mundelia, Derris, Lonchocarpus, and Tephrosia. In preparing the last named, the leaves are crushed and mixed with quicklime before using. Among the Menispermaceae are the Indian Berry, Fish-berry or Levant Nut (Anamirta paniculata) of the East Indies which contains picrotoxin; and

Pachygone ovata, used also by the Malays to poison crocodiles. Of the Rhamnaceae, Kraemer mentions Tapura, Gouania, and Zizyphus. Among the Tiliaceae, Kraemer, also mentions species of Grewia which are used as fish-poisons. The same authority adds species of Barringtonia of the order Lecythidiaceae and Laportea stimulans of the Urticaceae to the above list.

ANCIENT USE OF POISONS

In his work on "Poisons: their Effects and Detection," Blyth writes an excellent account of their history from which a few of the following more important data have been taken for the present work.

Their early history is involved in myth. Hecate was said to have been the discoverer of poisonous herbs and her knowledge passed in turn to Medea. The Egyptian kings, Menes and Attalus Philometer, not only had a knowledge of plants but the latter was also familiar with the uses of such plants as hyoscyamus, aconite, conium, and others of similar character. He experimented with poisons and compounded medicines. The Egyptians knew prussic acid, which was extracted from the peach and by means of which those who revealed religious secrets were put to death. The ancient Romans also must have been familiar with this poison, since a Roman knight once took poison and fell dead immediately at the feet of Samolus. The ancient Greeks knew about poisons and it was not considered a dishonorable thing to commit suicide.

Nicander of Colophon (204-138 B. C.) wrote two treatises on poisons, in one of which he described the effect of snake venom; in the other, henbane, aconite, conium, and fungi, were discussed. As antidotes for poisoning from any of these substances, he recommended such remedies as lukewarm oil, in order to excite vomiting.

Dioscorides (40-90 A. D.) divided poisons into (1) Animal poisons, as cantharides, poisonous snakes, the blood of an ox (probably putrid); (2) Poisons from plants, as opium, hyoscyamus, conium, aconite (the latter coming from Akron in Heraclea), and colchicum; (3) Mineral poisons like arsenic and mercury (cinnabar).

Pliny mentions that the Gauls dipped their arrows in a preparation of veratrum.

Toffana of Naples sold under the name of Acquetta di Napoli a solution of arsenious acid, by which, it is said, 605 persons were poisoned, among them the popes, Pius III and Clement XIV.

Poisoning was much practiced in India for the purpose of revenge, robbery and suicide, every little quarrel being liable to end in assassination of one of the parties. Such poisons as arsenic, aconite, opium, and extracts derived from plants of the Solanum family, were also used in India to destroy cattle. It is said that gipsies used *Phycomyces nitens*, having knowledge of its properties from the same country. The spores of the fungus were administered in warm water and death, accompanied with all the symptoms of tuberculosis, followed in a few weeks. The Hebrews seem to have been familiar with certain poisons, as arsenic, aconite and, possibly, ergot.

The deaths of Socrates, Demosthenes, Hannibal, and Cleopatra, were due to the administration of poisons.

In the early part of the Christian era, there were many professional poisoners and their business flourished, kings, emperors, popes, and members of the nobility being among their victims. There were two great criminal schools

in Venice between the tenth and seventeenth centuries, the government secretly recognizing the operations of these criminals and paying a sum of money for the execution of prisoners of note. However, these efforts were not often successful. J. Baptisa Porta, in the sixteenth century, wrote under the title of "Natural Magic," a work devoted partly to cookery and partly to poisons and how to use them.

The early methods of detecting the presence of poisons were crude; the surroundings were always noted; the suspected poison was generally administered to an animal and, if it died, poison was sure to be diagnosed without further investigation, since the early church forbade postmortem examination. Later, however, doctors were permitted to dissect and thus become familiar with pathological changes.

THE RISE OF CHEMISTRY AND POISONS

At the close of the eighteenth and the beginning of the nineteenth centuries, chemistry had advanced sufficiently to test for arsenic and the more important mineral poisons. Scheele discovered prussic acid; other chemists, as Berthollet, Lavoisier, and Stahl, added to the chemical knowledge of poisons The father of modern toxicology, however, was Bonaventura Orfila, whose work was published in 1814. Derosne discovered the alkaloids of narcotin and morphin in 1818. Pelletier and Cavantou discovered strychnin in 1818. Giesecke discovered coniine in 1827, and Geiger and Hesse separated atropin and hyoscyamin in 1833.

The modern aspect of the subject began with treatises on poisons by such workers as Vogel 1 and Richard Mead,2 and writings on the subject of chemistry through the works of Stahl, Scheele, Berthollet, Priestly and Lavoisier. Botanists, too, at this time began to be greatly interested in a study of poisonous plants. Thus we have the work of Bulliard and the work of Gmelin.4 The work of Bulliard discusses a large number of poisonous plants with excellent illustrations, and Gmelin treats quite fully of the then known poisonous plants of Europe.

The works of Gmelin, Bulliard and Plenck 5 on Toxicology, and Buchner's Toxicology were frequently quoted by the older writers. The greatest of the older writers, however, was Orfila 7 whose great work on toxicology became the recognized authority on toxicology. This work was first published in 1814, and passed through many editions. Orfila conducted actual experiments with different plants. This work of Orfila was also translated into different languages,8 Orfila was preceded by Fodere. About that period other toxicologies were published in France and Germany, such as those of Sobernheim and Simon,10

¹ The Usefulness of Natural Philosophy. 1654.

² Mechanical Theory of Poisons.

² Mechanical Theory of Poisons.
8 Historie des plantes veneneuses et suspectes de la France. Paris 1784. Folio X, 177
p., 72 tab. col.—Ed. II: Paris 1798.
4 Abhandlung von den giftigen Gewächsen. Ulm. 1775. Allegemeine Geschichte der
Pflanzengifte. Nurnberg 1777.
5 Toxicolgia, seu doctrina de venenis et antidotis. Viennae, Graeffer. 1785, 338 p.
6 Toxicologie. Nurnberg. 1827.
7 Traite des poisons, ou Toxicologie generale. Paris, 1813. Ed. III, ib. 1826. Orfila
and Bonaventura. Traite des Poisons ou Toxicologie Generale. Paris, 1814. (Ed. 5) 1852.
8 Toxicologie. Seemann & Karls. 2 Vols. Berlin 1829-1831.
9 Nedec leg. Ed. 2.

¹⁰ Handbuch der praktischen Toxicologie. Berlin, 1838. Toxicologie. Nurnberg, 1827, 2nd Ed.

Taylor, Hoffman, Th. Husemann & A. Husemann, van Praag, and Opwyrda Rebuteau,8 Selmi,9 Böhm and von Boeck,10 Dragendorff,11 Falck,12 and more recent works by Joshua Nunn,18 Smith,14 Hutyra and Marck,15 to say nothing of the recent contributions occurring in the American Veterinary Review, the Journal of the Chemical Society, American Journal of Pharmacy, besides treatises in many chemical and pharmaceutical journals.

The modern work of Blyth, though a somewhat exhaustive treatise on the subject of poisons, is not comprehensive so far as a large number of the poisonour plants are concerned. Many popular treatises on the subject of poisonous plants have appeared in nearly every European language both in ancient and modern times, but perhaps no one has contributed more to the subject of poisons than Kobert, who has published several treatises, and one of his works "Practical Toxicology for Physicians and Students" was translated into English by Dr. Friedburg. Such men as Dr. M. Greshoff of Haarlem, published a number of extended treatises on the subject "Poisons, especially Hydrocyanic Acids and Saponins, in Plants." His monograph on fish poisons and subsequent monographs two and three, really survey most of the poisonous plants of the world. Nor should we omit to mention the many treatises by Prof. Power of Wellcome laboratory and his students who have investigated a large number of poisonous plants or the work of Prof. Maiden of New South Wales, or of Cornevin of France.

RATZEBURG ON POISONOUS PLANTS.

Between 1834 and 1838 there appeared the first part of the work of Brandt and Ratzeburg on Phanerogamous poisonous plants of Germany, and in the year 1838 in the same work the poisonous Cryptogams by Phoebus. This like other works of the time contained numerous fine colored plates. This work pertaining to the flowering plants, lists the following plants of Germany as poisonous.

Darnel (Lolium temulentum) Juniperus Sabina, Yew (Taxus baccata) Arum maculatum, Colchicum autumnale, Fritillaria imperialis, Narcissus pseudonar, sissus, Paris quadrifolia, Veratrum album, Aconite (Aconitum Anthora, A Lycoctonum, A. Cammarum, A. altigaleatum, A. variable) Anemone nemorosa, Caltha palustris, Helleborus niger, H. foetidus, H. viridis, Pulsatilla vulgaris, Crowfoot (Ranunculus sceleratus, R. acris, R. alpestris, R. repens, R. bulbosus, R. flammula, R. Thora), Papaver somniferum, Euphorbia palustris, E. Cyparissias, Rhus Toxicodendron, Acthusa Cynapium, Cicuta virosa, Conium maculatum, Coronilla varia, Oenanthe fistulosa, Sium latifolium, Ledum palustre, Cyclamen curopaeum, Daphne Mezereum, Nerium, Oleander, Cynanchum, Vincetoxicum, Atropa Belladonna, Datura Stramonium, Hyoscyamus niger, Mandragora vernalis, Scopolina atropoides, Solanum nigrum, Digitalis purpurea, Gratiola officina-

⁴ Principles and Practice of Medical Jurisprudence. 3 vols. London, 1873.

⁶ Hoffmann, Lebrbuch der gerichtlichen Medicin. 5th ed. Wien, 1890-91.
6 Husemann & Husemann, Handbuch der Toxicologie. Berlin, 1862.
7 Leerboek voor practische giftleer. In Swee Theilen. Utrecht, 1871.
8 Elemens de Toxicologie et de Medecine Legale, appliquees a 'l'Empoisonnement. Paris, 1873. 2nd ed. by Ed. Bourgoing, Paris, 1888.
9 Studi di Tossicologio Chimica. Bologna, 1871.
10 Böhm and von Beeck. Handbuch der Letosicetionen. (Pd. 15 of the Communication)

¹⁰ Böhm and von Boeck. Handbuch der Intoxicationen. (Bd. 15 of the German edition

¹⁰ Bohm and von Boeck.

Anadouch der Intokrationen.

Of Ziemssen's Cyclopaedia).

11 Die gerichtlichchemische Ermittelung von Giften in Nahrungsmitteln, Luftgemischen, Spelseresten, Korpertheilen, etc. St. Petersburg, 1868. 3rd ed. Gottingen, 1888.

12 Die Klinischtigen Intoxicationen (Handbuch der spec. Pathologie u. Therapie red. von R. Virchow, Bd. 2.) Erlangen, 1854.

13 Veterinary Toxicology. Wm. R. Jenkins & Co., N. Y., 1901, 1907.

14 A Manual of Veterinary Hygiene. 5th ed. Wm. R. Jenkins & Co., N. Y., 1035 pages.

15 Specielle Pathologie and Therapie der Hausthiere.

lis, and Lactuca virosa. Many poisonous fungi are enumerated in the second part.

It is interesting here to note their classifications of poisons. Sobernheim elassifies the poisons into, A. vegetable poisons, B. nervous poisons, C. blood poisons. Orfila elasifies poisons into four classes, (1) irritant, acrid, eorrosive, (2) nareotic, (3) narcotic acrid, (4) septic poisons. In 1834 Brandt and Ratzeburg classified poisons as to their origin into (1) mineral, (2) plant, (3) animal. Brandt and Ratzeberg in their treatise on plant poisons make three divisions (1) nareotic (stupefying), (2) aerid (inflammatory), (3) narcotic (inflammatory). They use the elassification of Buehner which is as follows:- (1) narcotic, a HCN Prunus, b, volatile narcotic, Lolium, c, narcotic alkaloidal, Poppy; (2) acrid narcotic, Cicuta, Conium, Ruta, Digitalis; (3) irritant narcotic, a, Aconite, Oleander, Rhus, Smartweed, b, more volatile, hot acrid, Dirca, Pepper; (4) aerid, a, drastic resins, Bryonia, Hypericum, Melia, b, drastic coloring matter, Abrus, Spartium, Pokeweed, c, emetic alkaloids, Iris, Colchicum, Nareissus, d, unknown poisons, Agaricus, Boletus, Phallus, Lycoperdon. Fodere divided poisons into septic, narcotic, narcoacrid, acrid, irritant and astringent.

STATISTICS OF POISONING.

The use of poisons for criminal purposes, although not nearly so extensive at the present time as during the middle ages, still plays an important part in criminal law. The following statistics afford some indication of the use of poisons for suicidal and homicidal purposes.

According to the last census of the United States, the number of persons reported as poisoned was as follows:

| | By Active Poisons | By Gas 1 |
|-------|----------------------|----------|
| 1902. | 1374 | 950 |
| 1903 | 1551 | 1715 |
| 1904 | | 2167 |
| 1905 | | 1306 |
| 1906 | 1734 | 1276 |

Intentional cases of poisoning in live stock are not nearly so frequent as are those in the human family, although there are many cases of the former on record. Poisoning of live stock is generally accidental, caused by consuming plants that are poisonous. Large losses occur annually in this way. In 1900 Prof. Chesnut and Dr. Wilcox investigated the conditions in Montana relative to this subject and published the following statistics resulting from their studies. They state that probably not more than one fourth of the actual cases occurring came under their observation.

¹ These numbers include deaths during, or as a result of conflagrations. The annual average was 1412 from active poisons and 1365 from gas. From 1900-1904, the number of deaths by poison averaged 4.5 per 100,000.

| | Poisoning | cases | among | cattle, | horses, | and | sheep | in | Montana | observed | dur- |
|-----|------------|-------|-------|---------|---------|-----|-------|----|---------|----------|------|
| ing | the season | of 1 | 900: | | | | | | | | |

| | SHE | EP | CATT | LLE | HORSES | |
|-----------------------------------|--------------------|-------------------|----------|----------|----------|------|
| | Poisoned | Died | Poisoned | Died | Poisoned | Died |
| Zygadenus venenosus | 3030 40 3000 | 636 15 1900 | 6 2 100 | 2 2 56 | 4 4 | 3 3 |
| Cicuta occidentalis Loco weeds | 105 3550 | 80 700 | 36 3 | 56 30 | 150 | 3 |
| Total | 9725 | 3331 | 147 | 90 | 154 | 6 |

Nearly every veterinarian has frequent calls to attend cases of poisoning from obscure causes. These can often be traced to plants that occur in the pasture or feed lot.

Accidental cases of poisoning from such wild plants as jimson weed and others are frequent in the United States, several cases occurring annually from cowbane in Iowa. Statistics in regard to such cases are, however, difficult to obtain.

H. W. Cattell, as senior coroner physician in Philadelphia, performed 799 postmortems, in 155 of which, death was due to poisoning. The poisons used were listed as follows: aconite, 1; ammonia, 1; arsenic, 5; carbolic acid, 10; chloroform, 1; creosote, 1; cyanide of potassium, 1; hydrocyanic acid, 2; illuminating gas, 12; lead, 1; oil of merbane, 1; opium, 11; oxalic acid, 1; phosphorus, 1; silver nitrate, 1; stramonium, 1; strychnin, 2; sulphuric acid, 1.

In his work on poisons, Blyth states that the deaths from poisons in England and Wales during the ten years ending December, 1903, were 11,035. Deaths from laudanum were 1,505; cocaine, 12; atropin, 96; prussic acid and oil of almonds, 328; potassic cyanide, 207; strychnin and nux vomica, 244; aconite, 45; alcohol, 87; petroleum, 23; belladonna, 95; cocain, 12.

STATUTES ON POISONING AND ACTION OF POISON ON DIFFERENT ANIMALS.

The statutes do not as a general rule define poisons, but in most of the codes the sale of certain poisonous substances is regulated by law. The following extract from the Iowa code illutrates this: Sale regulated of substances under Schedule A. Arsenic and its preparations, corrosive sublimate, white precipitate, red precipitate, biniodide of mercury, cyanide of potassium, hydrocyanic acid, strychnia and other poisonous vegetable alkaloids and their salts, essential oil of bitter almonds, opium and its preparations, except paregoric and other preparations of opium containing less than two grains to the ounce. Schedule B. Aconite, belladonna, colchicum, conium, nux vomica, henbane, savin, ergot, cotton root, cantharides, creosote, digitalis, and the pharmaceutical preparations, croton oil, chloroform, chloral hydrate, sulphate of zinc, mineral acids, carbolic acid, and oxalic acid.

Not all poisons act in the same way, some acting more quickly than others. Quality and quantity are prime factors in the results obtained. As an illustration of this fact, we may mention ricin which is obtained from the cas-

tor oil bean, one gram of which, if properly diluted, is estimated as sufficient to cause the deaths of 1,500,000 guinea-pigs. The characteristics of the animal affected by the poison is also an important factor in the result. For instance, a fatal dose of strychnin in case of ruminants, when given by mouth is varying; when given hypodermically, it is a little larger than for horses; the minimum fatal dose for a horse being 1½ to 3 grains when given hypodermically, and 3/5 grains (or ½ ounce of nux vomica) when given by mouth, but as much as 2 grains is permissible.

The snail is said to be capable of withstanding more strychnin than an adult man. The minimum dose for man is $\frac{1}{2}$ a grain, while $\frac{4}{7}$ grains constitute a lethal quantity.

Cardiac poisons produce no action upon insects. The rabbit can stand more morphin than a man. Kobert says: "Amygdalin does not affect dogs, but it kills rabbits. The hedgehog takes, with apparent enjoyment, a dose of cantharides that would kill several persons under excruciating pains. The bite of the most venomous snake does not harm him; he can even accommodate no inconsiderable quantity of hydrocyanic acid. Whereas the frog is extraordinarily susceptible to the digitalis poisons, they have no effect on the toad." "Poisons act more powerfully when absorbed from the subcutaneous connective tissue than when administered internally, with the following exceptions: The neutral crotonolglycerid which is found in large quantities in the fresh seeds of Croton Tiglium, but which is often lacking in commercial croton oil, is inactive when introduced under the skin. It possesses, however, terrific action when taken into the stomach. Myronic acid of mustard as an alkaline salt has no effect when it is injected under the skin; it has, on the other hand, a strong action when taken per os by herbivora; the same is true of amygdalin.

"In all three of the foregoing cases, the apparent exception to the rule is explained by the fact that the substance, in itself not poisonous, is split up in the intestinal tract, giving off, amongst others, a toxic substance. In the instance first mentioned, crotonilic acid is the poison thus freed; in the second, ethereal mustard oil; and in the third, hydrocyanic acid. Some substances, such as salts of manganese, iron, tungsten, have no poisonous action when introduced into the intestinal tract, because under these conditions only very minute quantities are absorbed; others are rendered inert because they are excreted almost as quickly as they are taken up, curare being an example; and yet others, such as snake poison, spider poison, quillaic acid, sapotoxin, ergotinic acid, are converted into non-poisonous substances within the intestines."

Persons may become accustomed gradually to the use of poisons. Thus individuals who consume opium or its alkaloids may take large doses without apparent injury, although children are particularly susceptible. The former statement is equally true of those who daily use such poisons as hashish, nicotin, caffein, cocain, alcohol, or morphin. They must have the drug in order to keep up their condition. Many people exhibit idiosyncrasies with reference to food substances or drugs. Some people cannot inhale the odor of morphin, turpentine, or tobacco without becoming ill. Others are uncomfortably affected if the flowers of the common bird cherry or the haw are left in the room. Others become sick when in the presence of the flowers of the tuberose. Coming in contact with the castor-oil plants sometimes causes illness.

CHAPTER II

BACTERIAL POISONS

Impure Water

In all ages great stress has been laid upon the value of the Source

a pure water supply. In ancient times, wherever there were of Disease.

great centers of population, a large amount of labor as well as of money was employed to furnish water. Of this the Claudian aqueduct, built in Rome in the year 50 A. D., is an illustration. Prof. W. P. Mason says: "Not only was a generous daily per capita allowance sought for, but we note in the centuries gone by unmistakable evidences of a keen appreciation of the dangers lurking in a polluted supply; and upon this point many of the ignorant consumers of our day and generation would be benefited did they consult the wisdom of the past."

Of the value placed by the ancients upon the quality of water, Prof. Mason also says: "In ancient times, the valleys of the Euphrates and Tigris, now almost a desert, were densely populated. Four thousand years ago the rulers of Assyria had converted those sterile plains and valleys into gardens of extreme productiveness by the construction of immense artificial lakes for the conservation of the flood-waters of the rivers, and as great distributing canals for irrigation. One of these canals, supplied by the Tigris, was over 400 miles long and from 200-400 feet broad, with sufficient depth for the navigation of the vessels of that time." "In India, tanks, reservoirs, and irrigating canals were constructed many centuries before the Christian era, and a great part of that country was kept in the highest state of cultivation. Some of the tanks or artificial lakes covered many square miles, and were often fifty feet in depth.

A great deal of interest has been manifested recently in all parts of the United States concerning water supplies. This has not been confined to the cities but the interest is manifested in the villages and rural districts as well. We are now demanding more than ever before, not only that a good wholesome supply of water be provided to the citizens of a city or village, but also that as good a supply be furnished the farmer. That such diseases as typhoid and cholera are water borne can not be doubted. Many others, as anthrax, hog cholera, and tuberculosis may also be conveyed by water. Animal parasites are also water borne. In addition to these, there are some poorly defined intestinal disorders that are caused by poor water.

That typhoid fever is quite as prevalent in the country as in the city admits no denial. A record of the cases of typhoid occurring during the fall and winter in any of our rural communities shows that the disease is as widely spread in the country as in the city.

A certain class of animal diseases is produced not by the invasion of microorganisms, but is caused by the water supply being contaminated by the decomposing products of animals. The water may, for example, be highly charged with colon bacilli or other bacteria that produce poisonous products.

Several years ago Dr. Stalker traced an epidemic of horses, cattle, and pigs, all of which had been affected with similar symptoms, the animals uniformly dying after an illness of about two days. The disease was not contagious; the farm buildings were fairly comfortable and clean, and the trouble was, evidently, not due to the food consumed by the animals on the farm. Most of them, however, had been in the habit of drinking from a small creek which ran through the premises. The stream was supplied by a series of springs, and in ordinary seasons flowed for a portion of its course over a gravelly bed. This season the rainfall was light, and it so reduced the supply of water that it ceased to flow. Investigation made on these premises and on the adjoining farms indicated that dead animals were thrown down the steep bluffs into the bed of the stream. During the summer, chickens which had died from cholera and hogs dead from hog cholera had been dumped into the creek. In addition, the creek received the drainage from manure heaps. This was the kind of water that these animals had had to drink. Stock which did not have access to the creek but were watered from a well escaped the disease, while stock on other farms having access to the creek water suffered from the disease.

Dr. Lewis and Mr. Nicholson, in Bulletin 66 of the Oklahoma Agricultural Station, refer to certain troubles of live stock due to faecal contamination. In many cases the pond from which stock is watered is situated where plowed debris is carried into it by heavy rains, partially or completely filling it up, while the stock tramping down the banks soon complete the process. Stock standing in the pond also foul the water with excrement, and in hot weather, when the water is low, such a pond certainly can not afford a very satisfactory water supply. During the winter and spring months, when the rainfall is abundant, this condition is not so noticeable since the water is being continually changed by fresh water running in.

One of the dangers that follow allowing stock of all kinds to stand in a pond is that when the water is at a low stage, and foul, as it becomes in summer seasons, the cattle will not drink a quantity of the hot, foul, surface water, sufficient to prevent certain derangements of the digestive system such as impaction, "dry murrain," and other conditions that are usually ascribed to dry feed, but which are, in a large measure, brought about by insufficient water.

A type of injury resulting from the use of polluted water is illustrated in volume 19, page 74, of the "Journal of Comparative Pathology and Therapeutics." This record is in the form of evidence given in a case in which the plaintiff is the tenant of a farm on which is kept a dairy herd of from 30-35 cows. In 1903, there was no complaint but in 1904 the cows were put to grass in the middle of May and their condition became unsatisfactory at the end of July. Early in September, one of the cows aborted, six others lost their calves between that date and the 7th of October. On the 19th of November, another cow aborted; seventeen of the remaining ones carried their calves the full term and four were barren. The cows drank water from a small lake about one and a half acres in extent, which the town council of Maybole, who were the defendants in the action, used for the deposit of rubbish from the town. About the 18th of October, the cows were removed to another pasture with different water supply, and only one cow slipped her calf. (This occurred Nov. 19). The expert testimony was very conflicting. The plaintiff and expert

testimony held that septic poisoning resulted from the use of this water and thus caused abortion, Prof. Williams maintaining that water holding a large amount of vegetable matter is dangerous to pregnant cows, while the defendants held that this would not be a sufficient cause for the action. Judgment was rendered for the plaintiff.

Bacterial poisons are produced by two classes of bac-Bacterial Poisons. teria: the first includes such as are parasitic or pathogenic; the second, those which form poisonous products by the breaking down of dead animal or plant tissues. An illustration of the first class is seen in the products resulting from the tetanus bacillus and diptheria bacillus which produce an extra-cellular toxin. Another type is the toxin known as endo-toxin. In the extra-cellular form, toxin exudes through the bacterial cell-wall and is found in the body; while in the endo-toxin form, the toxin remains wholly, or in part, in the cell during the life of the organism and is liberated only on the death of the bacteria.

There are on record numerous cases of poisoning as a result of eating certain foods of animal origin, and the same statement may be made in regard to foods of plant origin. Such foods as meats, fish, cheese, and milk, sometimes become injurious because of the products of bacterial growth which they contain. These products are classified as either ptomains or toxins; a third class, the leucomains, result from the breaking down of tissues of the living animal body, being proteid bodies which have been broken down by enzymes, secreted by the cells of the body. These leucomains produce auto-intoxication. Ptomains are soluble, basic substances formed by the action of bacteria on protein material. Dr. Holland illustrates the action as follows: "The aminoacids, ornithin and lysin, constituents of pure protein, subjected to bacterial action, split off CO2, and change to putrescin and cadaverin." Some of these products, as methylamin, are harmless, while others are active poisons. The ptomains are strongly basic, combining with acids to form salts. They are precipitated with chlorides of mercury and are of various kinds. Some are free from oxygen, while others contain that element; some, as typhotoxin, tetanin, pyocyanin, are unclassified; several are injurious in foods; some are produced in fresh oysters and mussels.

Ptomain poisoning. Symptoms: Gastro-enteritis is the most prominent symptom, with depression and nervous disturbances. In most cases, there are, also, marked thirst, salivation, nausea, and vomiting, diarrhoea, cramps in the legs, great prostration, feeble pulse, dilated pupils, delirium, paralysis, and collapse. The postmortem examination generally, but not always, shows inflammation of the stomach and bowels. Cholin, in large doses, nervin, diamin, amanitin, muscarin, all act as poisons; nervin is much more powerful than cholin, the symptoms being those accompanying obstruction of the bowels, together with nausea, pain, and depression; the diamins are all actively poisonous, dilated pupils, convulsions, diarrhoea, and paralysis being prominent symptoms. Muscarin, found in Fly Agaric and certain putrid products, is a much more powerful poison than cholin or nervin and produces vomiting, griping pains in the stomach and intestines, slow pulse, arrested action of the heart, contraction of the pupils and fatal collapse.

Toxins. These are poisonous bases produced by living bacteria or by saprophytic bacteria in the animal body and in higher plants.

Holland arranges the food toxins in two classes: (1) The poisonous

products of specific bacteria, not putrefactive, growing in meat after slaughter, (2) Products of specific bacteria infecting the tissues of food-animals before slaughter. To these should be added: (3) Toxins of higher plants (phytotoxins), in which class would be placed ricin, a product of the castor-oil plant, abrin of the Jequirity plant, crotin of the croton, and robin from the black locust.

This belongs to the first group and is caused by the Bacillus Botulism. botulinus, which contaminates ham, sausage, and fish, and is so poisonous that it frequently is the cause of death in the person using the affected meat or fish. Rabbits, guinea pigs, and cats are very sensitive to the poison and die when given the fluid culture, rabbits succumbing in 36-48 hours after injection of 0.0003-0.001 c. c. Proteus vulgaris, growing in pork and beef sausage produces a similar poisoning.

Symptoms. "Epigastric discomfort, belching nausea, vomiting, gripes, diarrhoea followed by constipation;" nervous symptoms appear in a few days; these are dilated pupils, paralysis of the tongue and pharynx, loss of voice; death may follow delirium and coma, or, after some time, recovery may take place.

Poisoning from this cause manifests itself within a half hour occasionally, generally within twenty-four hours, although it may be delayed a week.

The toxins of the second class come from pathogenic bacteria and are due to their growth in living animals. The meat and milk from animals that have had septicaemia or pyaemia may cause such symptoms as headache, vomiting, profuse diarrhoea, gripes, chills and fever, the usual symptoms of these diseases.

In some of the European countries, and occasionally in the

Maidismus or
Pellagra. United States, there occurs a disease known as Maidismus or
Pellagra. It is common in Northern Italy and Bessarabia and
appears at times in France, Portugal, Spain, Roumania, and
Wallachia.

Etiology. The disease results from the exclusive use of bread made from spoiled maize or Indian corn. It is caused by an unknown toxic substance, partly basic in its nature, called pellagrocein. When maize is kept in a moist place, this poison is likely to develop, probably through the presence of organ-

isms, possibly bacteria or moulds.

Symptoms. The disease produces debility, reddening and swelling of the skin, an antipathy to food, profuse diarrhoea, delirium and raving mania, ataxic walk, spasms and contractions, debility of muscles, and paraplegia. Suicide is not infrequent. In chronic cases, the central nervous system, the skin and digestive tract become the seat of this disease.

Postmortem. The intestines show multiple small ulcers and extensive catarrh. The posterior column and motor area of the lateral tract of the spinal cord show a diseased condition.

Treatment. Change of diet. Use wheat or rye bread or if maize is used it should be thoroughly dried.

A large percentage of cases prove fatal.

CHAPTER III

DERMATITIS

Skin diseases are produced by a variety of causes, some resulting from pathogenic organisms and others through the ingestion of food. On account

of these differences, we have two classes sometimes given: parasitic skin diseases and urticarial diseases. Under the second class are placed such eruptions as those produced by buckwheat and smartweed, known technically as fagopyrismus and rhus venenata (dermatitis), and urticaria, the latter being produced by a large number of plants, especially the nettles. Or there may be internal causes due to innervation of vasomotors. Of the parasitic skin diseases known under the general name of dermatomycoses, we have several types, the co-called Tinea tonsurans and the Favus isms, the former occurring in cattle, dogs, horses, sheep, swine, and poultry.

This term is derived from two Greek words Dermatomycosis. meaning skin and fungus. The classification of the fungi concerned is not at all satisfactory; at present, however, they are generally included in the groups known as Fungi Imperfecti,



The Fungi Imperfecti include a cause of dermatitis or Rhus poisoning (U. S. large group of fungi whose life his-

tory has not been worked out completely. The fungi of this class are formgenera, such as the favus fungus. The so-called Achorion and Trichophyton of various authors represent such form genera, the Oidium albicans being another type; of these, some, perhaps, never produce any other kind of spore than the one commonly seen.

Many of these genera undoubtedly belong to the Ascomycetes, in which the spores are produced in little sacs called asci, the spores being known as ascospores. A kind of ringworm of the dog (Eidamella spinosa) belongs to this group, also the forms that produce aspergillosis, one type of which occurs in the ear.

The Mucoraceae produce an unsegmented mycelium and septa only where the reproductive bodies are formed; the spores usually occur in sporangia, or occasionally small spores may be found in the mycelium; zygospores which result from fertilization also occur in some species. Several species produce surface lesions.

According to Neumann, the dermatophytes of domestic animals belong tosix distinct genera of fungi as follows: Trichophyton, Eidamella, Microsporon, Achorion, Lophophyton, and Oospora. These genera are not, however, all accepted by botanists.

Trichophyton was established by Malmsten in 1848 and is characterized by having a mycelium consisting of simple or dichotomously branched filaments and producing spores from 4-9 μ long. Sabouraud, however, divides the genus into several species depending on the position of the fungus with regard to the invaded hair. The T. endothrix lives inside the hair; the T. endo-ecto-thrix develops outside of the hair, forming a sheath around it, and the T. endo-ecto-thrix develops both inside and outside the hair. This classification is scarcely tenable from either clinical or cultural characters. We have placed this genus with Sporotrichum.

The Eidamella spinosa described by Matruchot and Dassonville in 1901, has a much branched mycelium, 1.5 μ in diameter, divided into short segments and splitting into somewhat squarish oval bodies; it is found on the dog.

The Microsporon discovered by Gruby in 1843, has a branched mycelium, the latter branches bearing conidia from 2-3 μ in diameter. This fungus has also been placed with Sporotrichum.

The Lophophyton, described by Matruchot and Dassonville in 1899, produces a mycelium with some tortuous filaments, others short curved, with thick curved walls; no spores produced; it occurs on fowls and is also referred to Sporotrichum.

The Achorion was described by Remak in 1833. The filaments of the mycelium are from 2-3 μ in diameter, flexuose or straight, variously branched; finally breaking up into spores. This fungus has been placed with the genus Oospora.

Oospora was described by Wallroth in 1833. Its mycelial threads are 2-3 μ in diameter, arranged in irregular chains.

The best expert account of dermatomycosis so far as it affects lower animals will be found in the treatise by Neumann¹ who includes also an excellent bibliography on the subject. Hutyra and Marek have a German text which devotes considerable space to the subject. The work by Hyde and Montgomery treats the subject from a human standpoint. The work by Plaut is also an exhaustive treatise.

Etiology. The cause has been ascribed to various fungi which will be described later in the present work. The predisposing causes are uncleanliness, weakness of animals (those that are worn out may offer a favorable medium for the attacks of the fungus). In the case of cows, Fleming observes that the disease is common in the winter when the stables are dirty, and disappears in the spring when the animals are turned out to

^{1 &}quot;A Treatise on the Parasites and Parasitic Diseases of the Domesticated Animals."

Ranguern. 807, 365 f. 1907. New York.

pasture, the new condition being opposed to contagion. In the case of the cat, early age seems to be an essential condition. In the case of the rabbit, a similar condition seems to hold true. In the dog, early age is not an especially predisposing influence but inoculation is successful in young dogs only, according to Horand, so far as trichophyton is concerned. This latter statement does not hold true for favus.

Contagion. In ringworm of the horse, infection may occur from horse Mégnin states that in one locality 200 horses became infected in this way, a saddle from an infected horse having carried the disease to other animals. In each case, the disease occurred on the left side of the back. It has also been transmitted from an ox to a horse. The tinea tonsurans has been transmitted from the horse to calves, and the tinca of the horse from horse to dogs, sheep, and pigs, and even to man. Neumann says: "The infection of man is exceptional when the frequency of tinea tonsurans in the horse is considered, as there is scarcely a regiment in which it is not always on some young horse," Grooming is the usual way in which the infection is carried and rubbing facilitates inoculation. The ease with which infection occurs on man depends on the character of the fungus, some forms adapting themselves to the conditions present more readily than others. In the case of the bovines, the contagion may be direct. The virus may be preserved a long time in parts of stables where calves were affected with the tinea tonsurans. The infection spreads less readily to sheep and pigs but may be transmitted from bovines to man. In the case of the dog, it is transmitted from dog to dog, from rats and mice to the dog, and, occasionally, from dogs to man. In cats, favus is largely transmitted from mice and it is certain that this form can be transmitted to man. In general, it may be said that the transmission of favus from the rat or mouse is frequently brought about through the domestic cat. The tinea of the fowl is transmitted by contact with a diseased fowl. The favus fungus of the fowl 1 cannot be inoculated on the rat or dog but when inoculated on man, it produces lesions similar to favus. Man may be inoculated very easily by handling a fowl on which large erythematous patches occur, Similar patches have occurred in man when inoculation from a fowl was very probable.

Symptoms. Two forms of skin dermatomycosis in the horse have been (1) called microsporosis, and (2) trichophytosis. important symptoms of the first as given by Neumann are: "It appears in patches which are more especially seated on the upper part of the body — on the shoulders, back, loins, croup, sides, and flanks. These patches may, however, be met with on any part of the body, though they are rare on the lower parts of the legs. What are first noticed are the circular patches, the diameter of which is generally about that of a shilling; they are distinguished from the healthy skin by the dullness and erectness of the hairs covering them. Some time before the circular patches appear, a very small tuft of hairs probably from half a dozen to a dozen - may be seen slightly, but markedly, raised in the form of a fine pencil, and feeling as if they had a somewhat hard base, or were matted together at the bottom, when the finger is passed over them. These tufts may be several in number, and are usually best seen in hindquarters at the very commencement of the disease, or in the vicinity of the patches, of which they are the initial symptom. The hairs fall off in a

¹ Lophophyton gallinae, Mégnin - Trichophyton Mégnini, Blanchard. - Sporotrichum.

few days, and this is often the first symptom that attracts attention. The epidermis of the patch falls off at the same time as the hairs; it appears to be softened, and the surface of the skin has then a dark-grey tint and is slightly moist, which might be attributed to the rupture of vescicles, though their presence has never been demonstrated. It cannot, therefore, be said, as Raillet remarks, that the disease presents itself in the form of herpes, as what is so called in human pathology includes a phase marked by the appearance of vesicles. The humidity of the patch is ephemeral. Its surface generally soon dries, and is covered with epidermic scales of varying thickness, which are agglutinated into flat crusts that are shed and renewed incessantly. These crusts have-more frequently than in the ox-a shinning appearance and a grey or yellowish color like flax. At the same time, the lesion progresses by peripheral extension until it attains the diameter of a five-shilling piece or more, and on each zone invaded, successive symptoms are observed. Pruritis is nearly absent in Microsporosis, and is scarcely even shown to exist, except by movements indicating satisfaction on the part of the animal when the patches are gently scratched.

- "2. Trichophytosis (Sporotrichum). A.—Trichophyton flavum.—The lesions consist of large patches, at least 8 to 10 cm. broad, of a more or less regular form, greyish, and quite smooth. The hairs, raised and matted at their base by a greyish-yellow crust, fall away very rapidly with the crust. The naked surface is not prominent, and shows no trace of suppurative folliculitis (Bodin).
- "B. Trichophyton equinum occurs usually in numerous patches, some isolated, scattered over the croup and shoulders, and attaining at the most 3 cm. in diameter. At first they can be detected only by touch, but later they become visible by the flattening of the hair. The least traction or slight friction carries off a scaly crust which brings away the diseased hairs. The skin then appears smooth, moist, pinkish, or light grey. Very soon the patch becomes dry, scaly, or powdery, and at its base a slaty grey. The lesions spread by the falling out of the marginal hairs (Matruchot and Dassonville).
- "C. Trichophyton verrucosum, var. equi, occurs in numerous patches which average 5-6 cm. in diameter, and are localized on the shoulders, neck, withers, flanks, in fact, everywhere where the harness touches. By their confluence, these patches may produce large, irregular lesions. At first the hairs are raised, not broken, and are matted together at the base by a soft grey crust. This crust falls off in less than a week, carrying away the hairs, and having an absolutely bald, grey surface covered more or less with dry, greyish scales, and without follicular suppuration. In young horses, however, the shedding of the crust leaves a vesicle or pustule, then the surface of the patch is slightly raised, red, and indurated (Bodin).
- "D. Trichophyton verrucosum, var. asini. The lesions are the same as in the preceding Trichophytosis of the Horse, but are generally confined to the neck, head, and ears (Bodin).
- "E. Trichophyton mentagrophytes.— This Trichophytosis occurs usually on the nostrils or head. It forms patches up to 5 or 6 cm. in diameter, which may be mistaken for pustules of horse-pox undergoing regression. Over the whole of these patches the hairs are matted together at their base by a soft, brownish crust of unequal thickness. Slight traction on the hair brings away the crust, exposing a bare, slightly raised surface, which is red, inflamed, and pitted with

small depressions, grey at their base. These result from the opening of the pustules, of which some may be found at the margin of the patch. The hairs are not broken, but shed, and the condition is, in fact a suppurating folliculitis."

The symptoms observed in the bovine, as described by Neumann, are as follows: "The commencement is manifested by a slightly salient ring, on the surface of which the hairs are erect. An active proliferation of the epidermis causes the rapid formation of scales more or less adherent to each other, and crusts of 2 mm. to 7 mm. thick—hence; the name dartre crouteuse was given to the affection by the older (French) veterinarians. According to Gerlach, the crusts are thicker on dark skins, on which they have a greyish-white, fibrous appearance, resembling the amianthus (porrigo asbestinea) on white skins, which are usually finer, the crust is thinner and a little yellow in color."

Dingnosis. The diagnosis based on clinical symptoms should be verified by microscopical examinations. It is best to take material from the younger and deeper parts of the crust which may be moistened with water, or a better examination can be made if it is boiled with a 40 per cent solution of potash after which the particles can be dissected and the fungus threads and spores made out. The different forms cannot readily be distinguished except by cultural methods. The organism grows readily on nutrient media that are neutral or with slightly alkaline reaction. Sabouraud recommended the following:

| Pure glycerine, glucose, lactose, or maltose | 4 | grammes |
|--|------|---------|
| Granulated petone | 1 | gramme |
| Distilled water1 | .00 | grammes |
| Gelose | 1.50 | grammes |

Solid media like potato, agar, and peptonized bouillon are favorable media while liquid media are less favorable. Growth may occur at 15°C, the optimum being 30°C.

Prognosis. The duration of the disease depends on circumstances, it gradually diminishes and may disappear without medical aid. The average length of time of the disease is 40-50 days. Cleanliness has much to do with its disappearance. It lasts longer in thick coated animals than in those with thin coats.

Treatment. Cleanliness and sanitary surroundings, disinfection of stables, careful and regular grooming (all articles used in this process having been thoroughly disinfected, especially when they have previously been used on a diseased animal), avoiding any substance that causes irritation. The following preparations have been used with success: Mercury 1-500; carbolized glycerine; alcohol; an ointment composed of 1 part of carbolic acid, hard and soft soaps, each 20 parts. Fourie and La Calve recommend pure carbolic acid, tincture of iodine, and chloral hydrate in equal parts. The applications should be made once or twice a day or every other day depending upon the irritant properties of the preparations used.

For bovines, the remedies named above will prove efficacious; for the dog, application of an ointment prepared from 1-5 per cent of nitrate of silver, is satisfactory.

Dr. Stuhr has contributed the following account of favus in ani-Favus mals: "Favus is a contagious, vegetable-parasitic disease of the skin, characterized by the formation of more or less circular, cup-shaped crusts, varying in size from very small up to that of a dime. It has been observed in almost every species of domestic animals. The disease is quite rare among horses and cattle although dogs and cats are frequent sufferers. The domestic fowl and pigeon are also susceptible.

Of the laboratory animals, mice, rats, rabbits, and guinea pigs, harbor the

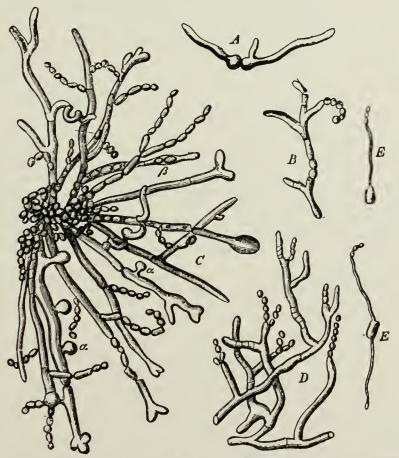


Fig. 2. Favus and Herpes Fungus. A. Spores germinating, grown in gelatine. B. Hypha breaking into segments. C. Formation of spores in chains (a) formation of buds (b) chlamydospores. D. Herpes, threads of the mycelium and formation of spores. E. Oidium lactis spore with germ tubes. After Grawitz.

disease. Young age, thin skin, and debility predispose. Favus is communicable from lower animals to man and vice versa. Man frequently contracts the disease from cats, the latter becoming infected from eating mice and rats.

Etiology. Favus is caused by a vegetable parasite, (Oospora porriginis) which invades the cutaneous structures, especially the epidermal portion. The Achorion Schoenleinii was first discovered by Schoenlein in 1839,

although Remak was the first to demonstrate its pathogenic character by direct inoculation. It consists of mycelium and spores, existing in such profusion that it is readily detected. Skin abrasions are an important accessory cause.

Symptoms. The disease is characterized by dry scabs, brownish or yellowish, gray or silver white on the surface, and white or sulphur yellow in their deep layer. These scabs have a circular form, with a diameter not to exceed that of a dime, and a thickness varying up to one-fifth of an inch. They occasion atrophy of the hair and a slight depression of the skin. These scabs are usually found on the forehead, cheeks, ears, face, abdomen, external side of the hind legs and in the neighborhood of the claws. At first the scabs are perforated by hairs which soon fall out. Later the skin exfoliates under the scab and leaves a pit. In the horse the scabs may become confluent and form bands as wide as the finger. Itching is observed in the dog. In most cases the progress of the disease is quite rapid although the prognosis is favorable unless the disease has become too far advanced.

Lesions. According to Robinson the parasite first obtains a lodgment in the funnel-shaped depression in the epidermis, through which the hair shaft emerges upon the surface. It grows luxuriantly in the upper part of the hair-sac and insinuates itself on all sides between the superficial layers of the epidermis. When it reaches a short distance on all sides of the liair follicle it breaks up the looser layers and appears on the surface producing the characteristic cup-shaped bodies. It also invades the hair shaft itself, penetrates between the cellular layers of the root sheath and by its mechanical pressure upon the papillae interferes with the nutrition of the hair and causes it to fall out. If the pressure is sufficient to cause atrophy of the papilla, a new growth does not occur.

In the skin the parasite usually confines itself to the upper corneous cells and does not extend to the living tissues. In cases where the surface is covered by irregular confluent masses of the parasites, the entire upper layer of the epidermis will be found infiltrated with the achorion.

The corium itself is usually in a state of chronic inflammation, and suppuration, which may be quite abundant, often occurs under crusts. Even in the absence of pus, the pressure of the parasite causes atrophy of the skin, and at last pit-like depressions or more extensive reddened scars are left. The disease ends with the destruction of the glandular structures of the skin.

Treatment. This is purely local except when debility complicates the disease, in which instance tonics should be administered to build up the system. As for the local treatment, its aim is to destroy the parasite and relieve the cutaneous irritation. The dry scabs may be softened and removed by thoroughly washing with soft soap and water. The removal of the hairs, by extraction, from the affected part has been attended with good success since, in so doing, many of the parasites are disposed of. A liniment composed of liquid tar and green soap two parts and alcohol one part will prove beneficial since it is antiparasiticide, disinfectant, dessicating, emollient and cleansing.

Mercuric chlorid in one to two per cent aqueous solution tincture of iodine, sulphur iodid ointment, red iodid of mercury ointment 1-8, sulfur ointment, etc., are all useful applications. It is advisable to clip the hair from unaffected parts adjacent to the diseased foci, so that any spread of the malady may be immediately detected. Whatever the treatment, it is a good plan to wash daily with soft

soap and water before applying it. Cleanliness is extremely essential." Hyde and Montgomery state that the parasiticides are corrosive sublimate in the strength of 1-4 grains (0.066-0.266) to the ounce; formalin (1-4 per cent); sodium hyposulfite in saturated solution; spirit of green soap.

CHAPTER IV

FORAGE POISONING, ERGOTISM, AND ASPERGILLOSIS

We have several excellent illustrations of how other Forage Poisoning external known parasitic organisms may produce disease. Catarrhal stomatitis, for instance, may be produced by the ingestion of fodder which has become infected with any one of several fungi belonging to distinct orders. Among these are the rust of clover, bacteria, mildew of grass, and the rape-destroying fungus, Polydesmus exitiosus; even the common grass rust and other rusts upon grasses as well as the bunts and smuts are known to produce this form of disease. Among higher plants, such products as the pungent spices of pepper and of the roots of horseradish and radish are treated at length in such pathologies as the Friedburger and Frohner Veterinary Pathology.

Serious diseases of the stomach are caused not only by pathogenic germs but also by the ingestion of various foods. Many foods, such as unclean, or damaged fodder, poor water, musty hay, mouldy corn, decomposing potatocs, are responsible for gastro-intestinal catarrh; many fodders, also, contain irritant substances. There are several forms of gastro enteritis. Among forms of the third class (including those caused by ingestion of lower organisms such as fungi or poisonous substances) we may mention botulism, fish poisoning, injuries produced by mould fungi, smuts, rusts, and, finally, the so-called toxic gastro-enteritis produced by numerous poisons. These have sometimes been classed as irritant poisons and narcotic irritant poisons. The vegetable poisons under this head are numerous and have been treated under the different plants. Some pathologists, however, mention especially lupinosis of sheep and equisetosis.

The terms applied to this disease are Cryptogamic Poisoning, Forage Poisoning, Enzootic Cerebritis, Epizootic Cerebro-Spinal Mcningitis, Leuco-Encaphalitis, etc.

Characterization. So-called forage poisoning among horses and mules is a non-communicable disease, which undoubtedly belongs to a group of cryptogamic poisonings. Horses seem to be slightly more susceptible than mules, although it usually terminates fatally in both species.

The disease is characterized by symptoms which are referable to a disturbance in the central nervous system, and by lesions which, if present, are also found there. The course of the disease may be very acute, or it may be greatly lengthened, depending upon the suddenness of the onset. The mortality is very high and but few well developed cases ever recover. Suckling foals do not contract the disease.

History. This disease has prevailed quite generally throughout the Eastern and Central parts of the United States for many years, but until recently has not attracted any considerable attention. During the past few years, however, it has occurred with unusual frequency in the Central West, and, because of the extensive losses directly attributable to it, has

become of great economic importance. In various parts of Iowa, for instance, individual stock-owners have lost several thousand dollars from its ravages. In the different localities the disease has been known by various names, such as "grass staggers," "choking distemper," and "putrid sore throat," and because it apparently presents some of the distinguishing characters of a specific infectious disease, has been frequently recognized as "infectious cerebro-spinal meningitis."

A noteworthy fact however is, that thus far no evidence has been discovered: which would indicate that the disease is transmissable from animal to animal, or that it is even inoculable.

On the other hand, an outstanding feature in every outbreak is, that the affected animals have had access to unwholesome food, either while at pasture or in the stable.

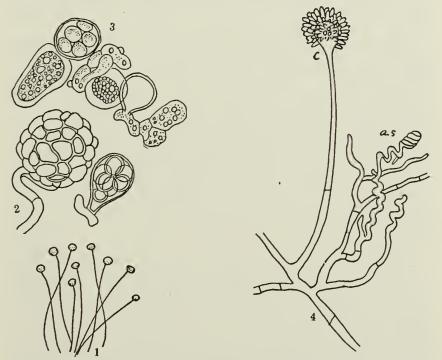


Fig. 3. Common Aspergillus on mouldy corn. 1. General appearance, showing long conidiophore and sterigmata on end. 2. Perithecium with one of its asci and ascospores. 3. Contents from an unripe perithecium. 4. A small part of the mycelium with conidiophore c and spore bearing sterigmata; young ascogonium as. 2, 3, 4 after DeBary.

Geographical distribution. The disease has been reported from nearly every part of the United States. It never becomes epizootic, but is usually confined to isolated localities.

While forage poisoning is not necessarily peculiar to low, poorly drained districts, it is at least most frequently observed in those places where conditions are most favorable for the development of cryptogamic growth.

Etiology. The disease seems especially likely to appear when horses or mules are fed on grain or fodder which has become overgrown with

moulds, or when at pasture, they have had access to grass which, for various reasons, has become fermented or mouldy. Various micro-organisms have been found to be associated with the disease, but as yet none have been proven to possess any etiological significance. Cultural and histological studies have all proved negative. Dr. Moore has in one instance succeeded in obtaining a pure culture of the colon bacillus from the brain.

Symptoms. Depending upon the severity of the attack, the three inay manifest itself in any one of three forms, namely; acute, subacute and abortive. It is possible to observe all of these forms in a single outbreak, as the suddenness of the onset is apparently regulated by the amount of the poison laden food which the animal has ingested.

The acute type is characterized by the abruptness of its appearance, and the grave general disturbances which immediately manifest themselves. There is sometimes violent trembling and twitching of the muscles over the entire body, but most commonly the acute form is ushered in by stupor.

There is manifested a weak, staggering gait and the pharynx is either partially or completely paralyzed. The tongue may also be partially paralyzed and protrude from the mouth, and saliva falls in strings from the lips. The pupil is dilated and the conjunctiva is, as a rule, highly congested. The pulse is variable and may be very rapid and hard, or scarcely perceptible; the respiration is hurried and jerky. The temperature may be slightly elevated, but is most frequently subnormal. Intestines and bladder are paralyzed.

In this form there may be slight muscular rigidity affecting the muscles of the back, neck and jaws, although in many cases this symptom never manifests itself. There is no rigidity of the ocular muscles. The animal soon becomes so weak that he is no longer able to support himself and falls. Delirium may manifest itself, in which the patient may perform a series of movements as if trotting, or become so violent as to do himself serious injury, but most often coma and complete paralysis supervene and death results in from four hours to two days from the commencement of the attack.

The subacute form is much the same as the preceding, except that it developes more slowly and the symptoms are not so violent. It is first noticed by a slowness in mastication and a difficulty in swallowing. A further indication of approaching paralysis is seen in the frequent knuckling and the loss of control over the tail. The temperature is subnormal and the pulse and respiration are but slightly altered. The bowels and bladder are inactive and it is seldom that voidance of urine and faeces occurs voluntarily. There is but slight rigidity of the muscles if indeed there is any, and no evidence of pain is apparent. These symptoms may last two or three days, when gradual improvement takes place, or the paralysis becomes more complete, the general weakness more marked, paroxyms of delirium develop, with inability to stand, breathing becomes more labored, coma comes on and death results apparently without a struggle. This form lasts from six days to two weeks.

In the abortive form there are no well marked constitutional symptoms. The appetite may be somewhat lessened, the ability to swallow slightly impaired, and the animal's movements a little uncertain, but no very noticeable symptoms appear to attract the attention. Improvement usually takes place on the third or fourth day, and recovery is the usual result.

Lesions. As a rule, post-mortem examination reveals no naked eye changes in the tissues of animals dead of forage poisoning. There

may be congestion of the brain and cord with extensive effusion into the ventricles and subarachnoid spaces. Few small hemorrhages and parenchymatous degenerations within the various organs have been mentioned. MacCallum and Buckley have found in the brains of horses dying of this disease, areas of softening "in the frontal region on each side, anterior to the motor region of the cortex." This lesion was practically confined to the white matter immediately under the cortex. In the affected areas there was "complete destruction of the brain substance, in which the anatomical structures are disintegrated and largely replaced by a colloid-like material. The neighboring blood vessels were acutely inflamed, with cellular infiltration of leucocytes and red corpuscles into the perivascular spaces and tissues. In a later outbreak these writers failed to find the brain lesion, but did observe the vascular changes above described.

McCarthy and Ravenel, in a study of fifteen animals found certain lesions in the upper gastro intestinal tract and in the central nervous system. These were:

- (1) In the intervertebral and Gasserian ganglia, where a pericapsular, small round cell accumulation was present. The cells were all of the same type, the nucleus and protoplasm being about the size of a red corpuscle. There was no evidence that these cells were the result of proliferation of the original layer of capsular cells.
- (2) Cortical lesions. These consisted of congestion of the cerebellar and cerebral cortex. There were also capillary hemorrhages. The meninges were normal.
- (3) Changes in the choroid plexus.—In three cases the choroid plexus was changed into a triangular tumor-like mass, of a yellowish red color and of a firm consistency. The increase in size was found to be due to a proliferation of the elastic tissue surrounding the vessels.
- (4) Changes in the nerves.—There was a distinct degeneration of the nerves supplying the larynx and neck. This was present in the nerve up to the ganglion, but was not found in the posterior roots. Other slight changes were detected.

Moore failed to find any gross lesions in the nervous system and other organs in the cases examined by him. In one case the brain, spinal cord, and organs were studied histologically with negative results.

Differential diagnosis. A very important point in the recognition of foragepoisoning is the history which has been referred to previously. It must be distinguished from inflammations of the brain and meninges, and from rabies.

Treatment. In the acute cases this is seldom successful, although quick-acting stimulants to arouse the patient may be tried. In the subacute cases a purge should always be given to rid the intestines of the poison. Strychnin in large doses, to overcome the extreme depression of the nerve centres, and atropin to support a failing circulation may be administered hypodermically at frequent intervals with benefit.

In the very mild cases, all that is necessary is to empty the bowels with a purge.

It is of the utmost importance, in all cases, with the return of the appetite, to supply only such food and water concerning the wholesomeness of which there can be no question.

Prevention. Since it seems to be quite generally accepted that this disease is brought about by the ingestion of mould-contaminated food

the prophylaxis is apparent. Whenever the disease makes its appearance either in a stable or a pasture, the animals should be immediately removed from further exposure by changing the food supply. The food should come, preferably, from a clean, new source and the water should not be contaminated by surface drainage. It is also well to thoroughly disinfect the mangers and feed-boxes, and render inocuous the soiled litter.

There is no known means of artificial protection, and the disease will recur if the animals are again allowed access to spoiled food. (Stuhr).

Poisoning from Silage

History. During the winter of 1908-1909, several cases of poisoning from spoiled silage were reported to Dr. Stange of the Iowa State College. Other cases have no doubt been encountered. In every instance, as in the case reported by Dr. Beaumont, below, moulds occurred in the silage. Dr. Beaumont says: "I am sending you under separate cover by mail a specimen of corn silage upon which you will notice is growing some form of mould which in my opinion is accountable for a very peculiar disease, existing among a herd of young horses and mules belonging to a farmer living here."

Dr. R. E. Buchanan found these moulds occurring in spoiled silage to be a species of *Monascus*. Other moulds, *Mucor*, *Penicillium glaucum*, and *Verticillium* were also present; but there was a preponderance of *Monascus*.

Symptoms. "The first animal, a three year old filly, was taken sick about April 1st, showing symptoms as follows: Gaunt, depressed, stiffness of gait. When lying was unable to rise, but when assisted to rise would stand and show inclination to eat but was unable to masticate and swallow food. Temp. 103.5 F. Pulse 86, Respiration 36; friction sounds distinctly heard at each heart beat. A whistling sound was emitted during expiration and there was also a suppressed painful cough. Animal died in about five days.

"A two-year-old mule and one two-year-old filly were attacked with disease. The mule is improving and will recover but the two-year-old filly shows exactly the same symptoms as Case No. 1, aside from being especially stiff and lame in one fore shoulder, and I think will die within two days."

Treatment. The treatment as followed by Dr. Beaumont is described in detail in his paper before the Missouri Valley Veterinary Association, June 16-17, 1909. Briefly, the method was as follows:

Tincture Strophanthus in two-dram doses, every two hours (given as a cardiac stimulant, the heart action being very weak). I quart of raw linseed oil given in two doses, six hours apart (as general laxative). Potasii Nitras in half to one ounce doses, dissolved in water and given as a drench, every three hours (alterative diuretic, and respiratory stimulant). After the first twenty-four hours the Tr. Strophanthus was discontinued and he began giving Iron Quinine and Strychnin tonic in one-ounce doses three times daily. This was continued with the Potasii Nitras until the animal showed marked improvement when both remedies were discontinued and he prescribed Fowler's Solution (Liquor Potasii Arsenitis) in half-ounce doses three times daily during convalescing stage of the disease which lasted about ten days or two weeks.

Dr. C. H. Stange has contributed the following on forage poisoning and especially with reference to silage:

"Numerous cases have been reported of an affection of the central nervous system, the symptoms being in general quite similar but different and varying

causes are assigned. Dr. Francis reports that in the fall of '03, spring of '04, four to five thousand horses and mules died with a nervous disorder characterized by structural changes in the brain which cause incoordination, delirium, coma and usually death. He concludes that the disease is not caused by moulds but is the result of animals having free access to a labor diet when kept in idleness. He was unable to find the germ described by Wilson and Brimball.

"Professor Harrison of the Ontario Agricultural College reported several cases and as a result of his investigations he concluded that the disease was due to a coccus insolated from the meningeal fluids. Pearson studied an outbreak in seven horses, five of which died. The outbreak occurred soon after opening a new silo, the ensilage from which was mouldy. The symptoms observed were very similar to those observed by Professor Harrison and he emphasized the paralysis of the pharynx and great muscular weakness. He concluded as a result of feeding experiments that the so-called cerebro-spinal meningitis was a forage poisoning. Dr. Dow of Connecticut describes two cases which were attributed to watering from a tub containing a mouldy slime. Dr. Ferguson of Texas describes three cases of forage poisoning due to smutted corn. There was vertigo, coma, low temperature, pulse in later stages rapid and irregular. In 1901 Dr. Hickman investigated an outbreak among horses in North Carolina in which a large number of horses died. In 1906 another outbreak occurred at the same place (Hyde Co.) in which about forty horses and mules died in about three weeks. The cause in these cases seemed to be moulds on vegetation. On the whole the country is low and swampy. The pathological changes of Epizootic Leuco-Encaphalitis were described by McCallum and Buckley in 1902. Muller of Germany reported an outbreak among horses, cattle and sheep due to mouldy straw. (Berliner-Tierärztliche Wochenschrift). Drs. McCarrol and McMullen describe an outbreak of cryptogamic poisoning in horses due to feeding mouldy beet tops. Dr. Lockhart describes several cases in Canada. The prominent symptom seemed to be the inability to swallow.

"Two outbreaks have come under our observation during the past year. The first consisted of eleven head of horses, two horses were being fed for market, the others were fed in the same manner during the nights and turned out during the day. The first animal affected was one being fed for market. It ate part of its feed in the morning but in a few hours showed symptoms of ptyalism, depression and paresis of the hind quarters. By noon the animal was down, unable to rise and struggling some, and died that night. The next animal to show symptoms was its mate. The symptoms shown in this case were similar to acute cases of the so-called cerebro-spinal meningitis, coming on with trembling and weakness causing the animal to stagger. An early symptom in all cases coming under our observation is the ptyalism due to inability to swallow. (Dyspagia). As a result the saliva collects in the mouth and hangs from it in strings. Muscles of different regions of the body are liable to contract. The breathing is rapid and in some cases may be of the Cheyne-Stokes variety. The temperature in this case was sub-normal. In some of the more chronic cases and when the animal has been down for some time with considerable struggling the temperature was somewhat elevated. The pulse was variable, being about normal in some cases and rapid and almost imperceptible in others. The animal became quite violent at times and finally died living but a few hours longer than the first animal. The other seven animals showed a more chronic course, showing inability to swallow, slow, weak pulse, difficult, noisy respiration, weakness and paralysis, spasm of muscles of head, neck and back, death taking place in from two to six days. The other two animals showed a mild type of the disease as slight loss of control, some exophthalmia, loss of appetite and thirst and loss of condition. These animals were placed on potassium iodid and nux vomica and recovered.

"This outbreak was attributed to mouldy silage, which was being fed to the horses, but in order to be more certain 150 pounds of silage were shipped to the college and fed, first to one horse which died in two days from an acute form of the disease. Another horse was fed but would not eat the silage so well, consequently did not die quite so soon, living for several days. In both cases the symptoms resembled those seen in the original outbreak. Post mortem revealed no changes except a few petechia along the small intestine, a few infarcts in the kidney and slight softening of the brain. This however was not very marked, probably due to the fact that they were acute cases. Microscopic examination revealed the presence of mould in the mucosa and submucosa of the intestine, also mycelial threads growing between and around the renal tubules.

"The other outbreak consisted of four horses, three of which died of an acute form of the disease, the symptoms being similar to those already described. The fourth being of a more chronic nature was placed on potassium iodid and nux vomica and recovered. In this outbreak the hay was found to contain a fine mould and was cut from an old pond which had been plowed up and seeded. The water had overflown this, however, and stood for some time. The symptoms and post mortems were similar to those described in the first outbreak, with the exception that no histological examinations were made.

"A form of cerebro spinal meningitis is quite common in Germany. It has also been described in Australia, Great Britain and Russia. It may be that these outbreaks are due to other causes than those already described. Sidamgrotzky and Schlegel found a form of coccus in the sub-arachnoid fluid, but it was necessary to make sub-dural injections of cultures of this organism to cause meningo encephalitis. Johne found a diplococcus in the cerebro-spinal fluid of affected horses.

"Ostertag found a diplococcus similar to the one found by Johne in the cerebro-spinal fluid in the so-called Borna's disease. They were pathogenic for horses and sub dural injections produced symptoms and death similar to cases of Borna's disease. Hutyra and Marek call attention to the fact that bacteriological investigations have not been followed by the same result but possibly the several investigators were working with the same organism. Nevertheless it remains to be shown whether all cases of cerebro-spinal meningitis are due to the same cause and resemble Borna's disease. On the other hand it is possible that epizootic cerebro-spinal meningitis of domestic animals has no specific cause.

"It is apparent that mouldy food and water has caused several outbreaks in this country. Natural infection in European outbreaks is also supposed to take place through infected food and drinking water. The disease is not transmitted from one animal to another. Mohler calls attention to the very interesting work of Schlegel and the Berliner Tierärztliche Wochenschrift who



ERGOT ON VARIOUS GRASSES

Fig. 4-1. Manna Grass (Glyceria nervata). 2. Blue Grass (Poa). 3. Spikelet of Bottle Grass (Asprella Hystrix). 4. Reed Canary Grass (Phalaris Arundinacea). 5. Wild Rye (Elymus robustus). 6. Koeleria cristata. 7. Wheat Grass (Agroppron Smithii). 8. Red Top (Agrostis alba). 9. Blue Joint (Calamagrostis canadensis). 10. Timothy (Phleum pratense).

associates with the affection an organism which he termed Streptococcus melanogenes. Mohler states, however, that whether the disease is of microbian origin or an intoxication has not yet been definitely established."

Ergotism is a disease of bovines caused by the ingestion of Ergotism considerable quantities of food contaminated by ergot. Equines are apparently less susceptible than bovines, although the horses have been known to suffer severely from the disease. Ergotism in man is not an uncommon occurrence, and in nearly every instance it has resulted from eating bread made of ergotized grain. The disease makes its appearance among cattle chiefly in the winter and spring seasons and has at times been the cause of serious losses throughout the central and western states.

Ergot is the sclerotium of a parasitic fungus, Claviceps purpurea, which infests many species of native and cultivated grasses, and appears on some of our grains, especially rye. The sclerotium represents a stage in the life history of the fungus, which is intermediate between that of the mycelium or spawn, and that of the spore-bearing thallus. It flourishes particularly well on rich soil and in warm, damp seasons. The chemistry of ergot is not exactly known, although Kobert succeeded in separating three bodies; namely, ergotinic acid, cornutin, and sphacelinic acid.

Ergotinic acid is a protoplasmic poison, and when injected intravenously produces inflammation of serous and mucous membranes, disintegration of red blood cells, and wide-spread ecchymoses; cornutin excites the central nervous system and causes general convulsions; and sphacelinic acid induces gangrene.

Symptoms. Ergotism manifests itself among animals chiefly in the chronic form, since, as a rule, the poison is acquired in small amounts and accumulation takes place slowly. Two distinct types of the disease are recognized, namely: spasmodic and gangrenous. Symptoms referable to the digestive tract, such as nausea, vomiting, colic, diarrhoea or constipation appear in both forms. Pregnant animals very frequently abort.

In the spasmodic type of the disease, symptoms due to over stimulation of the central nervous system, appear. These are tonic contraction of the flexor tendons of the limbs, anaesthesia of the extremities, muscular trembling, general tetanic spasm, convulsions and delirium. Death usually occurs from secondary causes.

Gangrenous ergotism is attributed to prolonged constriction of the arterioles, and more directly perhaps to degenerative changes in the vessel walls, and the consequent formation of hyaline thrombi.

It is characterized by coldness and anaesthesia of the extremities, followed ultimately by dry gangrene of these parts. The effects of this dry gangrene are often very serious and amount to sloughing of the feet, tips of the ears, tip of the tail, shedding of the hair, teeth, etc. Death takes place from exhaustion.

Lesions. With the exception of the gangrene which may vary greatly in severity, there are no lesions of especial significance. Degenerative changes in the sensory area of the cord and in the vessel walls have been observed in animals slowly poisond with ergot.

Treatment. The first essential in the treatment of ergotism is to remove the cause. In well established cases treatment does not as a rule prove satisfactory.

Tannic acid is the chemical antidote, and should be given to neutralize the unabsorbed portion of the poison. Chloral is the physiological antidote. In

addition to giving the antidote, the treatment is entirely symptomatic. (Stuhr).

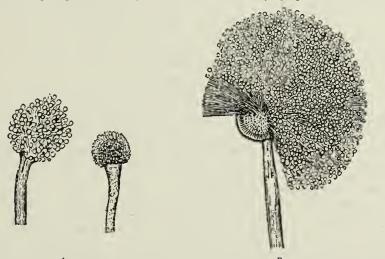


Fig. 5. A. Aspergillus fumigatus showing coniliophore on right with sterigmata and spores attached on left. B. A. niger showing conidiophore, sterigmata, and spores attached in chains. After Siebenmann.

Aspergillosis animals caused chiefly by the mould, Aspergillus fumigatus, although the Aspergillus niger is also pathogenic for birds. This disease is most frequent in birds, both domestic and wild, occasionally observed in horses and cattle, and rarely in man. Respiratory diseases and lowered vitality predispose. In all species the disease is characterized by purulent local inflammations in the lungs or other tissues, and a purulent and necrotic pseudomembrane upon the bronchial, tracheal, and other mucous membranes upon which it grows. The appearance of the pulmonary lesions sometimes resembles tubercle, sometimes actinomycosis.

Pneumonomycosis has been experimentally produced in birds (pigeons and geese) by compelling them to inhale aspergillus spores for a few minutes, after which they usually die of pneumonia in a few days. Rabbits have also been successfully inoculated by intravenous injection of spores.

Etiology. In mammals the Aspergillus fumigatus and in birds the Aspergillus fumigatus, niger and flavescens seem to be pathogenic species.

Infection takes place most commonly by inhalation of the spores which often are suspended in the air, or by taking them in with the food. Intestinal infection has not been observed. The spores are widely distributed in nature and exist in vegetable matter and grain abundantly. They possess remarkable vitality and exhibit considerable resistance to destructive agencies. The pathogenic power of the mould does not depend upon any product which it elaborates but upon the reactions which result from its penetration into the tissues. Peck observed the disease in seven subjects, in a stable where horses were fed on mouldy hacked hay.

Symptoms. The disease is of slow development in the larger animals and may not be observed until well advanced. In general the symptoms are of a pneumonic nature and in addition there is progressive emaciation. A case in a

Jersey cow, described by Pearson and Ravenel presented the following symptoms; the animal had been in poor condition for six months before it was examined. It was weak and depressed, did not eat, breathed with difficulty and, at times, coughed violently. Percussion of the chest gave sounds clearer and louder than normal and auscultation revealed the lung and bronchial sounds much intensified. Six days later these symptoms became more pronounced, the respiration and pulse very rapid. The animal grew rapidly weaker and died ten days after first being seen. The symptoms in birds are much the same as those in mammals except, that the disease runs a more rapid course. Emaciation advances rapidly and fetid diarrhoea may set in and continue until death in from a week to two months. At times emaciation is the only symptom. Fowls emit a glairy discharge from the nostrils which may contain the spores. In the prevention of the disease in fowls therefore, it is necessary to isolate or destroy the sick fowls together with the carcasses and fumigate the poultry houses. The roosts may be whitewashed.

Lesions. The lesions take the form of a miliary suppurative process, the foci varying in size from very small up to that of a pea. These may exist in large numbers and be scattered throughout the entire lung. Sometimes they become confluent and produce large areas of disease. The process starts in the bronchial mucous membrane, and later involves the bronchioles and alveoli. A very important feature is the intense amount of emphysema which is apparent on external examination of the lung. The lobules are often widely separated and can be readily seen in outline when a portion of the tissue is examined by transmitted light. In these emphysematous interlobular spaces, and in the air passages are seen whitish, mouldy looking patches. They are composed of denuded epithelium, inflammatory exudate, fruit hyphae and spores.

The lesions spread by penetration of the mycelium causing a destruction of tissue. Spores are not found within the tissues. In rare cases there is diffuse pneumonia characterized by hepatization and interstitial infiltration. On this latter account the disease has been described as being similar to contagious pleuro pneumonia of cattle. There may be pulmonary gangrene from secondary invasion of putrefactive organisms acting upon the devitalized tissue. An interesting feature is that this disease may interfere with the tuberculin test. This was shown in the case, above referred to, in which the test was used without success, and lesions of tuberculosis found in the lung on postmortem examination.

Treatment. This must of necessity be unsatisfactory since it is quite impossible to destroy the moulds which have penetrated the lungs. (Stuhr).

CHAPTER V

POISONING FROM FUNGI

That fungi of various kind are injurious, was known to the ancients. Prof. Ford 1 says, "The most interesting cases of mushroom or, as commonly described, toadstool poisoning and one of the first authentic cases on record, occurred in the family of the Greek poet, Euripedes, who lost in one day, wife, daughter, and two sons, who in the poet's absence partook of the deadly species. Among the great ones whose lives were sacrificed to the same ignorance may be mentioned Pope Clement VII., the Emperor Jovian, the Emperor Charles VI., Berronill of Naples and the widow of Tsar Alexis. The death of the Emperor Claudius is also assigned to this cause, but the reason and manner of the accident are not certain.

In addition to poisoning from toadstools, it has long been known that Ergot (Claviceps purpurea) is injurious to man and lower animals. In recent years Ergotism has not been so serious as formerly.

Other fungi also may be responsible for the death of animals by poisoning. The Fly Agaric (Amanita muscaria), a beautiful species, is common in many parts of the United States. I have described it in detail in another part of this work. In this connection I shall quote freely from the detailed and excellent account of poisoning as given by Prof. V. K. Chesnut, and the excellent report given of A. phalloides by Prof. Ford, who has written the most recent account of poisoning from this fungus.

The symptoms and treatment are thus described by Mr. V. K. Chesnut:

"The symptoms of poisoning from the fly amanita, as deduced from a number of cases, are varied. In some instances they begin only after several hours, but usually in from one-half to one or two hours. Vomiting and diarrhoea almost always occur, with a pronounced flow of saliva, suppression of the urine, and various cerebral phenomena beginning with giddiness, loss of confidence in one's ability to make ordinary movements, and derangement of vision. This is succeeded by stupor, cold sweats, and a very marked weakening of the heart's action. In case of rapid recovery the stupor is short and usually marked with mild delirium. In fatal cases the stupor continues from one to three days and death at last ensues from the gradual weakening and final stoppage of the heart's action.

"The treatment for poisoning by Amanita muscaria consists primarily in removing the unabsorbed portion of the amanita from the alimentary canal and in counteracting the effect of muscarin on the heart. The action of this organ should be fortified at once by the subcutaneous injection, by a physician, of atropin in doses of from one one-hundredth to one-fiftieth of a grain. As a stimulant emetic, mustard is particularly valuable. If this is not effective apomorphin should be administered by a physician. In case of profound stupor, however, even this may not produce the desired action. Tannin is of little

¹ Science N. S. 30: 97, 98.

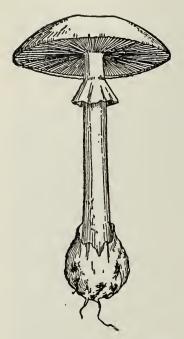


Fig. 6. Deadly Amanita (Amanita phalloides). U. S. Dept. Agrl.

or no value in rendering the muscarin insoluble in the stomach. If vomiting has not taken place, recently burned charcoal or two grains of a one per cent alkaline solution of permanganate of potash may then be administered, in order, in the cases of the former substance, to absorb the poison, or, in case of the latter, to decompose it. This should be followed by oils and oleaginous purgatives, and the intestines should be cleaned and washed with an enema of warm water and turpentine.

"Experiments on animals poisoned by the fly amanita and with pure muscarin show very clearly that when the heart has nearly ceased to beat it may be stimulated to strong action almost instantly by the use of atropin. Its use as thus demonstrated has been the means of saving many lives. We have in this alkaloid an almost perfect physiological antidote for muscarin, and therefore in such cases of poisoning its use should be pushed as heroically as the symptoms will warrant. The presence of phallin in Amanita muscaria is possible, and its symptoms should be looked for in the red color of the blood serum discharged from the intestines. Its treatment, which difficult, is discussed under Amanita phalloides.

"It is well known that in some parts of Europe the fly amanita, after the removal of the poison by treatment with vinegar, is a common article of food. It was interesting to discover not long since that among some of our own people a similar practice prevails. Though most of the colored women of the markets look upon the species with horror, one of them recited in detail how she was in the habit of cooking it. She prepared the stem by scraping, the cap by removing the gills and peeling the upper surface. Thus dressed the mushrooms were first boiled in salt and water, and afterwards steeped in vinegar. They were then washed in clear water, cooked in gravy like ordinary mushrooms and served with beefsteak. This is an exceedingly interesting operation from the fact that although its author was wholly ignorant of the chemistry of mushroom poisons, she had nevertheless been employing a process for the removal of these poisons which was scientifically correct. The gills, according to various pharmacological researches, are the chief seat of the poisonous principles in this plant and their removal at once takes away a large part of the poison. The salt and water would remove phallin or any other toxalbumin the mushroom contained, and although the presence of phallin or any of this class of poisons has not been demonstrated in Amanita muscaria, there is a strong suspicion that it may occur in slight amount. The vinegar, secondly, removes the alkaloid poison, muscarin, and the mushroom after the two treatments is free from poisons. This process is cited, not to recommend its wider use, but as a matter of general interest. The writer's recommendation is that a mushroom containing such a deadly poison should not be used for food in any form, particularly at a season when excellent non-poisonous species may be had in abundance.

"It is surprising that cases of poisoning are not more frequent. At Tacoma Park, D. C., on November 9, of last year, a lady who has a thorough knowledge of edible and poisonous mushrooms met a family, consisting of a man, woman, and two children, who had just completed the gathering of a basketful of the fly amanita and the death cup, described below, which they were taking home to eat. In reply to questions the woman stated that they had often eaten this kind purchased dry at an Italian store, but that they had never gathered fresh ones before. Of course they had mistaken the species, or possibly the dried ones were fly amanitas from which the poison had been removed by treatment with vinegar. After considerable persuasion the people consented to throw the lot away.

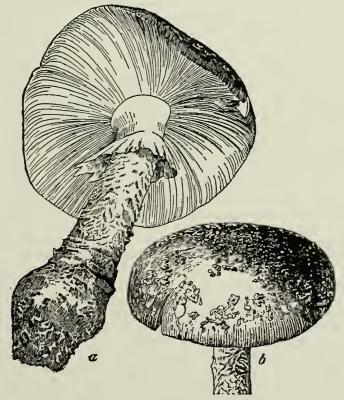


Fig. 7. Fly Agaric (Amanita muscaria). U. S. Dept. Ag.l.

"It is impossible to say what amount of the fly amanita would prove fatal, but in this connection it is of interest to note the custom reported by Krasheninnikoff, a Russian who travelled in Siberia and Kamchatka from 1733 to 1743, namely that the natives of the latter country, particularly the Koraks, used the fly amanita as an intoxicant, three or four specimens constituting a moderate dose for one habituated to its use, but ten being required for a thorough drunk.

The same observations, with varied details, have been made by others, particularly by Langsdorff, who traveled around the world with the Russian navigator Krusenstern from 1803 to 1806, and in more recent times by Kennan in his first Siberian journey of 1865-67.

"The plant may be taken fresh, but its taste is so disagreeable that only with great difficulty can a sufficient amount be eaten to produce the intoxicating effect. The Koraks have two principal methods of taking it: First, by swallowing pieces of the dried caps without chewing them; second, by boiling the dry caps in water and then drinking the liquor thus produced mixed with the juice of berries or herbs to disguise the taste. The intensity of the poisonous character of the fly amanita undoubtedly varies at different ages, with different individuals, and with different methods of preparation. The amount of the poison that can be taken into the system with impunity varies, too, with the person who takes it. The fact that a Korak, who has long used the plant as an intoxicant, can eat ten specimens and merely become drunk, does not prove that a similar number would not be fatal to an American who had never eaten it before.

"Very diverse statements concerning the properties of this fungus have been recorded. While some have attributed to it edible qualities, others have asserted that it is a most active poison and has caused numerous accidents by being confused with the Orange amanita. It is said to have caused death even when eaten in small quantities, and again it is said to have been eaten in abundance without any evil results. According to Quelet, it acts as a cathartic if eaten in small quantity, but causes death if eaten freely. One of my own correspondents assures me that he has eaten of the yellow variety, Var. formosa, without evil results, and that he regards it as very good. But there is no disputing the fact that the species possesses intoxicating and poisonous properties. It has long had the reputation of possessing properties fatal to flies that sip its juice. This suggests the names muscaria, Fly amanita, Fly agaric and Fly killer by which it is known. I have myself seen the cap of a single specimen surrounded by a circle of lifeless flies that had sipped the viscid juice from its moist surface and fallen victims to its virulent properties before leaving the place of their fatal repast.

"Some have attempted an explanation of the contradictory statements concerning this plant by supposing that its poisonous properties are not always developed, that in some localities or under favorable circumstances it is harmless. This explanation violates our sense of the constancy of Nature, and is not at all satisfactory. In the case of my own correspondent, the caps were peeled before cooking. May it not be that much of the noxious quality resides in the epidermis and the viscid substance upon it, and that by discarding this the dish is rendered less dangerous? In some cases it is said that those who eat it freely and without harm boil it a long time in water and throw away the water. In this way, doubtless, much of the poison is abstracted. Long soaking in salt and water, also in vinegar, have been recommended as a means of rendering suspected or noxious species harmless, and may have been practiced in some of the cases in which this fungus has been eaten with impunity. Whatever may be the explanation of the contradictory statements, the only safe way is to consider this species as deleterious and avoid its use under all circumstances. There is no need of taking any risks, with suspected species,

since there are so many good ones against which no charge of evil has ever been established."

A second very poisonous species is the White or Deadly Amanita (Amanita phalloides), common also in some parts of the United States. This species is described in another part of this work. This and allied species are eaten ignorantly by persons who do not know the nature of the powerful poison found in the plant. Prof. Ford says, "A small amount of the fresh material is sufficient to cause profound illness with fatal outcome, so potent is the poison contained in its meshes, and the raw plant seems usually more toxic than the cooked specimens.

"Two or three 'deadly amanitas' suffice to bring on disastrous results, and Plowright reports the death of a child of twelve from eating a third of the pileus of a small raw plant. The extreme toxicity of this species illustrates the dangerous consequences which the admixture of two or three specimens to a dish of edible mushrooms entails.

"Following the consumption of the fungi there is a period of six to fifteen hours during which no symptoms of poisoning are shown by the victims. This corresponds to the period of incubation of other intoxications or infections. The first sign of trouble is sudden pain of the greatest intensity located in the abdomen, accompanied by vomiting, thirst and choleraic diarrhoea with mucous and bloody stools. The latter symptom is by no means constant. The pain continues in paroxysms often so severe as to cause the peculiar Hypocratic facies, "la face vulteuse" of the French, and though sometimes ameliorated in character, it usually recurs with greater severity. The patients rapidly lose strength and flesh, their complexion assuming a peculiar yellow tone. After three to four days in children and six to eight in adults the victims sink into a profound coma from which they cannot be roused and death soon ends the fearful and useless tragedy. Convulsions rarely if ever, occur and when present indicate, I am inclined to believe, a mixed intoxication, specimens of Amanita muscaria being eaten with phalloides. The majority of individuals poisoned by the "deadly amanita" die, the mortality varying from 60 to 100 per cent. in various accidents, but recovery is not impossible when small amounts of the fungus are eaten, especially if the stomach be very promptly emptied, either naturally or artificially."

Kobert isolated from the fungus, a substance which he called *phallin*, and which had the property of disolving the red blood corpuscles. Such substances are called hemolysins. Prof. Ford says, "Very minute traces of this substance brought in contact with the red blood cells of man or with those of animals, produced within a short space of time, fifteen minutes to one or two hours, a complete solution of these corpuscles—a laking of the blood. So powerful was the hemolytic action that even in a dilution of 1-125,000 it was still operative upon the red cells of ox blood."

In a recently published statement by Prof. Ford it appears that 1 the fungus always contains another poison which differs from hemolysin in being resistent to heat and digestion, "the blood-laking substance phallin, being destroyed by heating to 70° C., and by the action of the digestive ferment. This substance he called Amanita-toxin, and the blood-laking substance Amanita-hemolysin. Abel and Ford 2 have shown that the so-called phallin, regarded by Kobert as

¹ Science N. S. 30:101. 2 Jour. Biol. Chem. 2:273; 1907.

a toxalbumin is a glucoside. Prof. Ford has obtained an anti-poison or an anti-hemolysin with a high grade of immunity. According to Schlesinger and Ford 1 the Amanita-toxin in a purified state is one of the most powerful of organic poisons — four tenths of one milligram killing a guinea pig within twenty-four hours. Ford believes that the hemolysin plays no part in human intoxication, but that the toxin is the active principal which resists the action of the gastric juice and boiling. He finds that the Amanita rubescens considered an edible species by some, contains an hemolysin as powerful as the Deadly Amanita. He found a toxin and an hemolysin in Amanita virosa. The latter substance in a dilution of 1-200 killed a guinea pig. The A. spreta produced intoxication and according to Ford must be classified with the "deadly poisonous" as the A. verna.

The A. strobiliformis, A. chlorinosma, A. radicata, and A. porphyria, do not contain hemolysins but small quantities of a toxin probably identical with amanita-toxin. The Amanita solitaria, regarded as edible, causes the blood corpuscles to adhere in clumps much as agglutination occurs with typhoid bacilli when brought in contact with the blood of a typhoid patient.

¹ Jour. Biol. Chem. 3:279. 1909.



Anemonella and Stemless Loco Weed. Photo by Charlotte M. King,



CHAPTER VI

POISONING FROM OTHER PLANTS. EQUISETOSIS, LOCOISM, AND LUPINOSIS

Equisetosis. It has been recently proven by direct experimentation that the common horsetail (Equisetum arvense) when ingested in sufficient amount, is capable of producing fatal poisoning among horses. This discovery is of great importance since the plant has a wide distribution, and at times is the cause of extensive losses. The common horsetail thrives best in moist sandy soils or in low, damp meadows, which are not frequently cultivated, and often constitutes a large part of wild hay. The dried plant alone seems to be poisonous. Young horses seem to be the most susceptible. Sheep are supposed to be slightly susceptible although cattle eat the hay in which the plant occurs in large proportion, with impunity. The toxic principle of the plant has not been determined.

Symptoms. The effects of poisoning from eating horsetail appear at times varying from two to five weeks, depending upon the age of the animal, and the amount of contaminated hay ingested.

The first symptoms are usually unthriftiness, general bodily weakness and emaciation. The animal seems to have a depraved appetite, preferring the plant to wholesome feed. As the disease progresses the muscular weakness becomes more pronounced, the animal loses muscular control and exhibits incoordinate movements. During this stage the pulse and temperature are depressed, extremities are cold and the visible mucosae are pale. Appetite usually remains good until the end and consciousness is apparently retained. Finally the animal falls, manifests nervous excitement, paroxysms of convulsions appear and death results from exhaustion. In the final stage the pulse becomes accelerated and the temperature elevated.

Hypostatic pneumonia is a frequent complication.

Treatment. The first step in the treatment is the removal of the cause. A cathartic should be administered to rid the bowels of the irritant and nerve and heart stimulants given to combat the symptoms of depression. In case the patient is unable to stand, it would be advisable to give some support. When the animal is down it becomes necessary to guard against the development of hypostatic pneumonia.

Where cases are not too far advanced and appropriate treatment is instituted, recovery is the usual result. (Stuhr).

Stock-poisoning by the loco weed is a frequent and serious con-Locoism. dition with which the stock-owners of the western half of the United States have to contend. Montana and Colorado, especially, sustain heavy annual losses. Similar diseases occur in other parts of the world. In Australia other plants of the order Leguiminosae like Gastrolobium produce similar symptoms. Maiden 1 states that the "Nenta Lessertia disease of S. Africa is identical with a disease of the Pea-eating animals of Australia and

¹ Miscell. Pub. Dept. Agrl. N. S. Wales, 477: 11.

with the Loco disease of the United States. Many forage plants of excellent repute such as white clover, alfalfa, lotus and other plants, may produce tympanites.

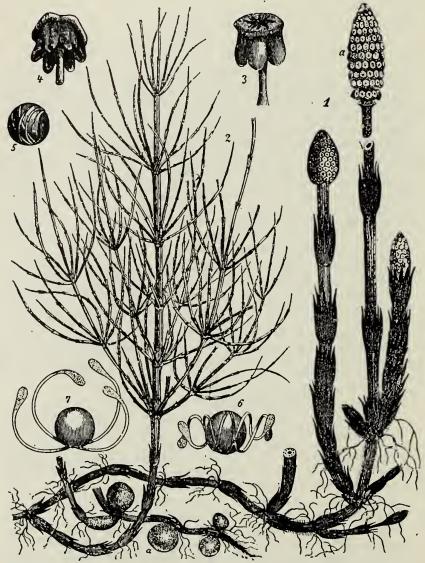


Fig. 8. Common Horsetail (Equisetum arvense), the plant causing Equisetosis. 1. Fertile stems terminating in cones a. 2, Sterile stem. a. rhizome tubers. 3. Sporophyl with sporangia. 4. Sporangia opened to discharge spores. 5, 6, 7. Spores with spiral elaters. After Wossidlo.

Symptoms: difficult breathing; the poison enters the circulation and stops the action of the lungs and heart when the animals stagger and die.

In the advanced stages the animals become frantic, hence the name "loco" or crazy. Horses and sheep are the most susceptible, although cattle are also affected.

Of the various species of loco weed, the stemless loco Oxytropis Lamberti Pursh), and the woolly loco weed (Astragalus mollisismus Torr), are the most injurious. These weeds grow luxuriantly on sandy ranges and appear early in the spring when other vegetation is scarce, and since they retain their fresh



Fig. 9. Loco Weed (Astragalus mollissimus), U. S. Dept. Agrl.

green color during the entire summer they prove especially attractive to stock.

Recently the poisoning has been attributed by Dr. Crawford and others of the U. S. Dept. of Agrl., to mineral salts in the plant.

The period of greatest danger is chiefly during the month of May.

Symptoms. The symptoms, which are referable to the nervous system, are attributable to the narcotic effect of the plant. They appear slowly and are apparently divisible into two stages. The first stage is characterized by the following symptoms: Stupor, defective vision, unnatural movements and apparent hallucinations. When excited the animals become frenzied. The coat becomes shaggy, the teeth grow long and become loose, and a depraved appetite which is very marked, is developed. The animals prefer the loco weed to wholesome food, and will dig up the roots and cat them to satisfy their craving.

In the second stage there is emaciation, exhaustion, feeble movements and finally death from starvation. The course of the disease is quite variable

and may last from a few months to one or two years. Sheep manifest symptoms very similar to those above described.

Treatment. When the disease has reached the advanced stage, treatment is of no avail as recovery does not occur. If, however, the afflicted animals are taken early in the course of the disease and placed on pasture where loco weeds do not exist, and are given good nourishing food, there is hope of recovery.

Prevention, which of course is the most desirable, is not always practicable. Animals do not as a rule become addicted to the loco habit when they have plenty of wholesome food and salt.

There are no demonstrable lesions other than emaciation. (Stuhr).

The recent investigations of Marsh and Crawford lay considerable stress on the presence of barium in the plants and the Bureau of Animal Industry, Washington, D. C., recommends the following treatment: for cattle, strychnin in doses of three-twentieths to four-twentieths of a grain daily, administered hypodermically; for horses, Fowler's solution of arsenic in half-ounce doses daily in the drinking water or in the grain. This treatment should be continued for at least a month. To correct the constipation which is almost universal in locoed animals, magnesium sulphate (Epsom salt) may be administered as a drench in two-ounce doses. Epsom salt may also serve to some extent as an antidote to the poison produced by the weeds. Beneficial results have also been obtained by giving horses daily a drench containing two ounces of Epsom salt with ten drops of dilute sulphuric acid, and by giving cattle tri-weekly three or four ounces of Epsom salt with a proportional increase in the quantity of dilute sulphuric acid.

As the foregoing treatments are in the experimental stage, the Bureau of Animal industry of Washington, D. C., would be glad to receive reports from their use.

The value of keeping stock away from these poisonous plants is indicated in some investigations that have been carried on by the Bureau of Forestry and the Bureau of Plant Industry. In many cases the ranges are becoming practically useless on account of these poisonous plants and if used the losses are so heavy as to materially reduce the profits of the business. In the Manta Forest Reserves in Utah for instance, it was found that the death of sheep was due to their browsing upon the chokecherry. Certain portions of the old trail were abandoned, and along other portions the chokecherry bushes were cut out. The method of handling the sheep was also changed. Instead of large bands which could be moved but slowly, smaller bands were trailed, and so far as possible they were allowed to fill up on healthy forage before entering the dangerous area. The trail was also improved wherever practicable and by this means it was possible to get the sheep through in much better shape and with little or no loss. The Department has also, in some instances, adopted the plan of flagging the area in which these injurious plants occur.

This is a disease of sheep and horses especially, caused by Lupinosis. eating the seeds and straw of the lupine. Cattle and goats are also susceptible and the dog has been poisoned experimentally. There are many species of the lupine growing in various parts of the United States, although the yellow lupine (Lupinus luteus) is the most toxic. The nature of the toxic agent found in the lupines has not as yet been determined. Arnold and Schneidemühl succeeded in isolating a chemical poison and



Fig. 10. Lupine (Lupinus leucophyllus) causes lupinosis U. S. Dept. Agrl.

gave to it the name lupinotoxin. They described its physical properties but failed to determine its chemical composition. Attempts to associate a fungus with the plant have failed. Lupinosis is characterized by jaundice, acute yellow atrophy of the liver, and parenchymatous inflammation of other internal organs.

Symptoms in Sheep. The disease appears in either the acute or chronic form, depending upon the amount of poison ingested. These two forms have been experimentally reproduced by giving carefully regulated amounts of lupinotoxin.

In the acute form the disease appears suddenly. There is loss of appetite, fever, hurried and difficult breathing, rapid pulse, stupor, vertigo, and not infrequently swelling of the lips, ears or face. The initial temperature may be as high as 104° to 106° Fahr., but is intermittent and gradually falls just before death. The pulse may reach 130 per minute and the respirations 100. A bloody froth may issue from the nostrils. Icterus which may be detected in the conjunctiva and the urine, usually appears on the second or third day. In certain cases this latter symptom fails to manifest itself and therefore is not constant. There is grinding of the teeth and sometimes trismus. The animal apparently prefers the recumbent position, extends the head on the ground and seems entirely oblivious to all surroundings. At first there is constipation, the faeces being hard and scanty and covered with yellow mucous. Later diarrhoea may set in and the excreta be tinged with blood giving them a dark brown color. Emaciation develops rapidly. In case of recovery the symptoms gradually abate and improvement takes place slowly. Cachexia is a common sequel. In the chronic form the symptoms are not so violent. Jaundice may be entirely absent and emaciation and anemia may be the chief signs. Inflammatory tumefaction of the lips, eyelids, and ears with the formation of ulcers and scabs is described by various writers.

Course. Death may supervene within twenty-four to forty-eight hours, although frequently the disease lasts four or five days. The immediate cause of death is rapid emaciation and extreme weakness. Horses contract the disease from eating oats contaminated with the seeds or from eating the straw of the plant. The symptoms which they manifest are essentially the same as those above described. Horses seldom die from the effects of lupines.

Lesions. The cadavers are emaciated and decompose rapidly. The muscles are of a grayish yellow color, the fibers having become fatty and having lost their striations. The subcutaneous tissue of the abdomen and the omentum and mesentery are yellowish. The most important lesion in both the acute and chronic forms is found in the liver. The alterations in this organ are those of acute hepatitis. The liver cells have become swollen and granular on account of the parenchymatous change, or they may be more or less completely degenerated into fat. The gland is soft and friable and may be somewhat swollen. The interlobular connective tissue is greatly increased in amount due to inflammatory hyperplasia. In the course of a few days the liver undergoes acute yellow atrophy as a result of the absorption of the degenerated cells and the contraction of the hyperplastic stroma. In the chronic form the changes are those of chronic interstitial hepatitis. The icterus is of hepatic origin and due to catarrh of the bile ducts. 'The gall bladder is distended with bile and its lining membrane is congested and swollen. The kidneys and bladder may show changes, more or less marked, due to inflammation. The bladder is, as a rule, empty. In the digestive tract we observe frequently yellowish discoloration of the mucosa, hemorrhages in the small intestine with catarrhal lesions of the entire canal. The heart is pale and friable and the blood which it contains is dark and thick. Capillary hemorrhages are quite generally observed throughout all of the tissues.

Treatment. This is chiefly preventive since there is no specific antidote. Attempts should be made at once to prevent further absorption of poison by administering some acid as acetic or hydrochloric, well diluted with water. Alkalis should be strictly avoided as the poisonous principle is very soluble in alkaline solutions. It is advisable to evacuate the bowels by giving a purgative, preferably oil. Potassium permanganate is recommended by some as an antidote. Further than this the treatment is entirely symptomatic. (Stuhr).



Fig. 11. A. Larkspur ($Delphinium\ tricorne$). B. D. Carolinianum. Larkspur poisoning (Delphinosis) is caused by various species of Delphinium. A. U. S. Dept. Agrl. B. Ada Hayden.

CHAPTER VII

DELPHINOSIS, LATHYRISMUS (LATHYRISM), ACONITISM, VERATRISM, UMBELLIFERAE, CONIUM, CICUTA.

The purple larkspur (Delphinium Menziesii D C.), and Delphinosis. other species which are found in the northwestern part of this country and especially in Montana are plants very dangerous to stock. Drs. Chesnut and Wilcox have proven the toxicity of the above species by direct experiment and have called attention to the serious losses which they occasion annually.

The latter experimenter has fatally poisoned a yearling lamb within two hours by administering, per os, the extract made from less than an ounce of the dried leaves. The weed appears early in spring, in advance of the forage plants and it is during this period that the greatest harm results. The poison is found both in the leaves and the roots although the latter are not frequently eaten on account of their woody fibrous nature. The poisonous principle has not been isolated. Cattle and sheep are most susceptibl, e although horses frequently suffer.

Symptoms. The first indication of poisoning is a general stiffness and a straggling gait, especially in the posterior limbs. Walking appears to be difficult, and is evidently painful. At this stage the pulse and respiration are much depressed, and the temperature is lowered. The appetite is retained in most cases until the appearance of the final stage of the poisoning. This is manifested by irregular muscular twitching of all of the muscles which finally becomes frequent and violent, and by incoordinate movement. There are attempts at vomiting and the animal froths at the mouth. Finally all of the muscles of the body contract spasmodically, the animal falls and dies in violent spasms. The pulse and respiration become very weak and rapid just before death. There is no aberration of the special senses. The course of the poisoning is quite rapid and death usually takes place within a few hours.

Lesions. The direct cause of death from larkspur poisoning is probably failure of respiration due to paralysis of the centre and the alterations therefore are those of asphyxia. The lungs are congested and dark-colored and the right heart, veins and capillaries are distended with dark colored blood. Other organs and tissues are normal with the exception of the general venous and capillary congestion.

Treatment. Potassium permanganate is the chemical antidote and should be given as early as possible in doses of fifteen to twenty grains to horses, thirty to fifty grains to cattle, and five to ten grains to sheep, dissolved in a copious amount of water. To combat the extreme depression of the circulation and respiration, atropin sulphate has proven very efficacious. Even after the final convulsions have begun this drug has been of good service. It should be hypodermically administered in doses of three-fourths to one grain to the larger animals and one-twentieth to one-fifteenth of a grain to sheep. In the late stages sheep should be given as much as one-sixth to one-

fourth of a grain. In the convulsive stage when there is impending failure of respiration, inhalations of ammonia may be resorted to with good results.

A feature of the treatment quite as important as the medication is the care of the animals. Complete rest and freedom from excitement are very essential since exercise or fright is likely to induce fatal spasms.

Finally the danger from poisoning can be largely obviated by preventing access to the young plant in the early spring.

In ancient times, this disease was quite common, affecting Lathyrismus. both man and the horse. It is very frequent today in Spain, Italy, Russia, and India.

Etiology. It is caused by eating bread made of flour derived from some species of Lathyrus or Vetch (L. Cicera, L. Clymenum, or L. sativus), or, in case of horses, by the consumption of Fodder Pea. The seeds of these species contain a toxic substance formed without the action of bacteria. Man, the horse and the pig are subject to this disease.

Symptoms. In domestic animals, the symptoms are debility of the rear and lower extremities, producing motor paralysis. In lower animals, the normal functions of the larynx become impaired because of paralysis.

Treatment. Change of fodder, providing good food, application of massage and electric treatment.



Agrl.).

Prognosis. Fatal cases are not frequent.

Post-mortem. The ganglion cells of the anterior horns are atrophied; also the recurrent nerves and the muscles of the larynx.

(Adapted from Friedberger and Fröhner).

Various species of aconite (Aconitum) are known to be Aconitism. poisonous. One species, the Aconitum Napellus, has long been used jn medicine. The most common species in North America is the Columbia Aconite (A. columbianum), which is found in the Rocky Mountains and on the Pacific coast. Several other species occur in North America, one extending into northeastern Iowa; the European Aconite (A. Napellus) is frequently cultivated in gardens. Cases of poisoning are largely due to the administration of over doses in medicine. In the Rocky Mountains accidental poisoning among live stock is confined to animals that graze at higher altitudes. All parts of the plant are poisonous; one tenth of a grain of the drug is a poisonous dose for some animals. The smallest fatal dose recorded in man is a teaspoonful of the tincture of aconite, which is equivalent to about XXX gr. of the crude drug. The minimum lethal quantity is 1-16 of a gr. for man.

Symptoms. The effects of the poisoning are a tingling sensation on the end of the tongue, which shortly gives rise to a burning sensation followed by pronounced constriction in the throat. It reduces the pulse and frequency of the cardiac pulsations. The action of the heart is lessened and the pulse is weak, irregular and intermittent, at first slow and then rapid; tingling and prickling over the entire body is characteristic; vision is confused, there are abdominal pains, vomiting and diarrhoea. Death is caused by the stoppage of respiration, but is preceded by numerous twitchings; in the case of the horse the animal falls and is unable to rise. The symptoms are different in cats and rabbits.

Treatment. No specific antidote is known, but physicians use atropin or digitalis and nitrite of amyl. The stomach should, however, be evacuated at once; cardiac and respirative stimulants are given subcutaneously.

The Hellebores belonging to the genus Veratrum are common Veratrism. in the mountain regions of the west and one species is abundant in the east. They are found in swampy places, in wet meadows and along brooks. The most frequent cases of poisoning come through the administration of the drug; although in the Rocky Mountains considerable quantities of the plant are consumed by sheep where grazing is close. All parts of the plant are poisonous although the root is more poisonous than the seeds and leaves which contain several alkaloids.

Symptoms. Veratrin is a powerful irritant and when inhaled in minute quantities produces sneezing. When injected under the skin it causes restlessness, when consumed in large quantities it produces salivation, frequent vomiting with purging pain, and collapse, the temperature falls. Veratrin is a drug poisonous to the motor and sensory nerves. Death is caused by paralysis of the heart; 1/16 of a grain has produced alarming symptoms in man and 1 gr. injected subcutaneously produces poisonous symptoms in a horse.

Treatment. The stomach should be emptied immediately, then give stimulants, such as brandy and strong coffee.



Fig. 14. Larkspur (Delphinium glaucum) causes Delphinosis (U. S. Dept. Agrl.).



Fig. 15. Cowbane (Cicuta occidentalis), a deadly poisonous plant. (U. S. Dept. Agrl.).

The Cowbane (Cicuta maculata) and related species be-Poisoning from long to the Carrot family. The Cowbane is common in low Umbelliferae. grounds in the Northern States and in the Rocky Mountains; another species (C. vagans) occurs on the Pacific Coast; and

a third species (C. bulbifera) is a common bog plant in the Northern States.

The Cowbane is frequently mis-called Wild Parsnip. The latter, however, has a conical root and is the feral form of our cultivated species, and, although it may be somewhat injurious, as indicated elsewhere, is not poisonous like the Cowbane (Cicuta maculata).

Cowbane is a smooth, marsh perennial plant from 2-5 feet high, with fleshy, fascicled roots and a pungent odor; leaves are pinnately compound with coarsely-serrate leaflets; flowers are white and small; fruit broadly ovate to oval and small. The European species has long been recognized as poisonous. Many cases of poisoning of man and lower animals by this plant are on record.

Cases of poisoning of children by Cowbane are not of infrequent occurrence, several being reported each year in the daily press.

The following item appeared in the Des Moines Register and Leader of May 23, 1909, and is but one of many that have come under the observation of the writer during the last twenty years.

Boone, May 22.—"Virgil Hyatt, a high school boy, was poisoned last night while walking to the Ledges, a summer resort near Boone. He fell to the ground unconscious, and a companion carried him to a nearby farm house, and summoned medical aid and his mother from Boone. The trip was made in an automobile at record breaking speed. The boy was brought to a hospital here (Boone) but died just as he was being carried into the institution."

It seems that the young man, while walking across a plowed field with a companion picked up some of the weed whose roots had been exposed and ate freely of them. In an hour he complained of illness, and fell into the creek from which his companion rescued him. However, as reported above, all attempts to save his life were unavailing and he died from convulsions in a few hours.

As a sequel to the above, the daily papers of June 3, of the same year report a second death in the same locality from the same cause. In this case a young man who was sent to secure some of the weed for examination, became poisoned in some way, possibly by the juice of the plant coming in contact with some abrasion of the skin. The lad died after a short illness, having shown symptoms similar to those of the previous case.

Symptoms. The first symptoms are pain in the bowels, urging to ineffectual attempts to evacuation, burning in the stomach, nausea, vomiting, tetanic convulsions which may be severe resembling those produced by strychnin, or there may be coma without convulsions,

Dr. Hazeltine says that, in one case, he found the patient "showing convulsive agitations consisting of tremors, violent contractions and distortions with imperfect relaxations of the whole muscular system, astonishing mobility of the eyeballs and eyelids, with wide dilated pupils, frothing at the mouth and nose mixed with blood, and, occasionally, genuine, violent epilepsy."

The convulsive agitations were so violent that the pulse could not be examined with sufficient accuracy to determine its character. There is a profuse sweat. In fatal cases, the respiration is stertorious, the pulse small, and the face cyanotic. Not many cases among animals have been recorded. In a case

reported to the writer by an Iowa farmer, a cow which had eaten freely of the roots, fell into a spasm when brought into the barnyard. The animal, however, rose, walked one hundred feet and fell again, got up again, walked about thirty rods, fell and died in about thirty minutes. In a second case, a yearling owned by the same man had been in good healthy condition but began to display the same symptoms and died in twenty minutes.

Dr. E. S. McCord, on September 31, of the same year, gave an old horse six drachms, hypodermically, of a strong decoction of the root. In fifteen minutes the animal showed uneasiness; pulse was full and fast; in a short time the animal laid down, and the pulse decreased; the horse was in great pain and kept moving the extremities; the pulse was weak but the patient finally recovered. The botanist of the Oregon Experiment Station found that the root has less of the toxic substance in the summer than in the winter and spring, which may account for the failure in this last case to produce death. In frogs, frequency of breathing is increased, tetanic convulsions follow, gradually paresis of the extremities, and lastly full paralysis and death. Cicutoxin,



Fig. 16a. Poison Hemlock (Conium maculatum), native to Europe; naturalized in the U. S. (U. S. Dept. Agrl.).

the characteristic poison of Cicuta, acts especially upon the medulla oblongata; the brain and spinal cord are merely secondary seats of its action.

Treatment. The stomach should be effectually evacuated by the use of the stomach pump or by a strong emetic. External and internal stimulants such as whisky should be applied; anaesthetics and narcotics used to control the spasms; hypodermic injection of morphin aids in recovery. It usually hap-

pens, however, that the veterinarian or physician is called too late to accomplish much.

The Poison Hemlock (Conium maculatum) is indigenous to The Poison

Europe and has long been known as a poisonous plant. It is Hemlock.

a fetid smelling herb from 2-5 feet high, with a spotted stem, compound leaves, and small, white flowers in umbels. The plant is not uncommon in waste places in the East and in the Rocky Mountains, especially in Utah. It has long been used as a poison.

Symptoms. In lower animals, there is observed a dilatation of the pupil, followed by weakness of the limbs, passing into paralysis; labored respiration, frequency of breathing diminished, heart action irregular; death preceded by convulsions. In man, there are weakness in the lower extremities, staggering gait, in two hours paralysis of upper and lower extremities and slight convulsions; death occurs in a few hours usually caused by cessation of respiration.

Treatment. The stomach should be evacuated by means of a pump or tube; or a hypodermic injection of 4-5 drops of a solution of apomorphin given; or emetics of sulphate of zinc or mustard administered. The temperature of the body should be kept up by hot applications.

Stimulants may be given, and, if necessary, artificial respiration applied. As a drink, strong tea, tannin, or any harmless vegetable decoction containing tannin may be administered.

CHAPTER VIII

FISH AND ARROW POISONS, HYDROCYANIC POISONING-TOXALBUMINS-BLACK LOCUST, CASTOR OIL, AND JEQUIRITY.

Fish and arrow poisons have played an important part Fish and Arwith the aborigines of all countries and they are still used row Poisons. to a considerable extent by primitive people. Thus Merrill 1 mentions the use of the Antiaria toxicaria in the Philippines and other plants used in the same way which are being worked up by Dr. R. F. Bacon.

Radlkofer 2 some years ago published a long list of plants which are used to poison fish, and added a history of the earlier literature. He lists some 154 species which have been used in various parts of the world for this purpose and these plants belong to the following orders and genera. The species are listed under the poisonous species in another part of this work.

Dilleniaceae (Tetracera), Menispermaceae (Anamirta, Abuta, Pachygone); Cruciferae (Lepidium), Capparideae (Cleome), Bixaceae (Pangium, Hydnocarpus), Ternstroemiaceae (Caryocar), Tiliaceae (Grewia), Meliaceae (Walsura); Chailletiaceae (Chailletia Tapura); Rhamneae (Gonania); Sapindaceae (Serjania, Paullinia, Sapindus, Dodonaea, Harpullia, Magonia); Hippocastaneae (Pavia); Leguminosae (Tephrosia, Milletia, Orobus, Abrus, Centrosema, Clitoria, Camptosema, Phaseolus, Lonchocarpus, Derris, Piscidia, Bowdichia, Cassia, Bauhinia, Leucaena, Albizzia); Myrtaceae (Barringtonia, Gustavia); Compositae (Clibadium, Ichthyothere); Campanulaceae (Tupa); Ericaceae (Rhododendron); Primulaceae (Cyclamen); Myrsineae (Aegiceras, Jacquinia); Sapotaceae (Bassia); Ebenaceae (Diospyros); Apocyneae (Melodinus, Thevetia, Cerbera, Aspidosperma); Loganiaceae (Buddleia, Strychnos); Solanaceae (Hyoscyamus, Nicotiana); Scrophularineae (Verbascum, Digitalis); Bignoniaceae (Bignonia, Tecoma, Jacaranda); Labiatae (Eremostachys); Chenopodiaceae (Chenopodium); Polygoneae (Polygonum); Aristolochiaceae (Aristolochia); Piperaceae (Piper); Thymelaeaceae (Daphne, Wilkstroemia); Euphorbiaceae (Euphorbia, Phyllanthus, Securinega, Piranhea, Croton, Joannesia, Manihot, Jatropha, Excoecaria, Hura); Coniferae (Taxus); Liliaceae (Veratrum).

Ernst 3 lists only sixty species that are used as fish poison. There must, however, be considerably more as indicated by Radlkofer.

W. M. I. Brost Pauwels 4 in his contribution on the Surinamic fish poisoning 5 contributes an interesting article on the subject.

Pauwels who made an investigation of Nekoe (Lonchocarpus violaceus) states that it is a powerfully toxic substance. He found that Nekoeid will poison fish in proportion of 1,5,000,000, and that a second substance B.

¹ Philip. Journ. of Sci. 2:111, Sect. C.
2 Sitz. Math-Phys. Classe k. b. Akad. d. Wiss. München., 1886, 379.
3 Memorio Bot. el Embarbascar o sea la Pesca por media de Plantis venenosas.
4 Bijdrage tot de Kennis der Surinaamsche Vischvergiften. M. Greshoff has likewise published a number of works on fish poisoning plants.
5 Hart and Swatters found in the Piscidia Erythrina piscidin C₁₆H₁₂O₄, and Greshoff found in Pachyrhizus angulatus, pachyrhizid C₂₆H₁₈O₆ (OCH₃)₂.

Nekoeid will poison fish in proportion of 1-10,000,000. The poison will take effect in one hour. The water poisoned with the substance will cause the fish to make an effort to get away from the poison, they are in a horizontal position, breathe heavily, come to the surface of the water and try to jump out and finally breathing becomes increasingly difficult and at last they turn on their backs and die.

Under poisoning from cherry, sorghum and a few other Hydrocyanic plants, an account has been given of poisoning from hydrocyanic acid. It may be convenient to bring together some of the plants from which the very poisonous substance, prussic acid, has been obtained.

Maurits Greshoff of the Colonial Museum in Holland has taken the pains in a paper on Cyanogenesis to give the distribution of Prussic acid in the vegetable kingdom, the Hydrocyanic acid being found in a great many different plants. The following list gives the orders in which this substance occurs.

Ranunculaceae (Aquilegia vulgaris, Thalictrum aquilegifolium). Berberidaceae (Nandina domestica). Cruciferae (Lepidium sativum), Bixaceae (Gynocardia odorata, Hydnocarpus venenata, Kiggelaria africana, Pangium edule, Ryparosa caesia, Taraktogenos Blumei, Trichadenia zeylanica); Sterculiaceae (Sterculia); Tiliaceae (Echinocarpus); Linaceae (Linum usitatissimum); Rutaccae (Citrus medica); Dichopetalaceae (Chaillctia cymosa); Olacaceae (Ximenia americana); Celastraceae (Kurrimia zeylanica); Rhamnaceae (Rhamnus Frangula); Sapindaceae (Cupania, Schleichera trijuga); Anacardiaceae (Corynocarpus lacvigata); Leguminosae-Papilionaceae (Lotus arabiscus, Indigofera galegoides, Phaseolus lunatus, Vicia sativa, Dolichos Lablab); Rosaceae (Amclanchier vulgaris, Chamaemeles, Cotoneaster integerrima, Crataegus Oxyacantha, Eriobotyra japonica, Nuttallia cerasiformis, Ostcomeles, Photinia, Pyrus, Prunus Amygdalus, Pygcum africanum, Spiraea Aruncus); Saxifragaceae (Ribes aureum); Combretaceae (?Combretum constrictum); Myrtaccae (?Psidium montanum); Melastomaceae (Memecylon); Samydaceae (Homalium); Passifloraceae (Passiflora quadrangularis, Tacsonia); Caprifoliaceae (Sambucus nigra); Rubiaceae (Plectronia dicocca); Compositae (Chardinia xeranthemoides, Xcranthemum annuum); Sapotaceae (Isonandra, Lucuma bonplandia, Payena latifolia); Asclepiadaccae (Gymnema latifolium); Convolculaceae (Ipomoea dissecta); Bignoniaceae (Osmohydrophora nocturna); Euphorbiaceae (Bridelia ovata, Elateriospermum Tapos, Hevea brasiliensis, Jatropha augustidens, Manihot utilissima, Ricinus communis); Urticaceae (Sponia virgata); Araceae (Arum maculatum, Colocasia gigantca, Cyrtosperma lasioides, Lasia aculcata); Gramineae (Glyceria aquatica, Panicum, Sorghum vulgare, Stipa hystricina); Fungi (Hygrophorus agathosmus, Marasmius oreades, Phaliota radicosa, Russula foetens). He makes the following statement with regard to the presence of this substance in plants:

"Many plant physiologists in Europe, with more experience with Prunus or amygdalin than with the tropical Pangium, incline to the view that hydrocyanic acid in these plants has nothing to do with either the building-up or the breaking-down of proteids, but that this substance is made by the plant from sugar and nitrate by a special process, and serves no other purpose than to defend the plants against the attacks of animals. It is above all the incompleteness of our physiological knowledge which makes decision between these theories difficult.

"In the study of this question it is important to remember the possible diversity of origin of this body, and every cyanogenetic plant will be required to be examined on the lines laid down by Treub."

The wide distribution of glucosides that yield hydrocyanic acid is evident from the list above. Dunstan and Henry 1 discovered three glucosides, dhurrin $C_{17}H_{17}O_7N$ in the common sorghum, lotusin $C_{28}H_{31}O_{16}N$ in a species of lotus of Egypt and phaseo-lunatin $C_{10}H_{17}O_6N$ in wild beans of Phaseolus lunatus, the common lima bean. Brunnich 2 attributed death from the feeding of immature sorghum to dhurrin. Power and Lees 3 isolated from the seeds of Gynocardata odorata a glucoside to which they gave the name gynocardin C₁₃H₁₉O₀N. All of the above glucosides yield on hydrolysis, hydrocyanic acid. The most important and best known of all the glucosides that yield hydrocyanic acid is amygdalin.

Greshoff discovered an amygdalin-like glucoside in two tropical trees, Pygeum parviflorum and P. latifolium. The same author found glucosides. in a member of the milkweed family Asclepidaceae. The Pangium edule of the tropics contains a large quantity of\a glucoside capable of being converted into hydrocyanic acid and a large amount can be prepared from a single plant. The Hydnocarpus inebrians also contains a large quantity of a glucoside which yields hydrocyanic acid. It is used to destroy fish. The common linseed cake contains a glucoside which yields prussic acid. Francis found prussic acid in the sweet cassava root, .0168 per cent, and in the bitter cassava .0275 per cent. It is well known that fresh bitter cassava root is bitter poison. The above facts are brought together by Blyth in his work on poisons and may be consulted for more of the details.

The statistics on poisoning seem to indicate that it occupies third place among poisons in the order of frequency in Great Britain. In that country there are about forty deaths annually from this poison according to Blyth. It is responsible for the loss of a great many cattle in sections of the country where the wild cherries are abundant and also from sorghum poisoning. It is frequently used for criminal poisoning, at one time more frequently than now. It is nearly always taken by the mouth into the stomach, but occasionally the vapors produce death. It is generally used by Entomologists to kill insects.

Blyth gives the symptoms of poisoning as follows: Cold blooded animals require a larger relative dose than warm blooded animals except the birds which are slightly less sensitive but the action is essentially the same. Hydrocyanic acid acts in two ways:

1. It profoundly interferes in the ordinary metabolic changes in animals. 2. It causes a paralysis of the nerve centers.

Normal blood decomposes with great ease hydrogen peroxide into oxygen and water. If it is normal venus blood and a little hydrogen is added it becomes bright red, but if a trace of prussic acid be present it is a dark brown color.

The blood corpuscles lose their power of conveying oxygen to all parts of the system and asphyxia results. The main symptoms in animals are as follows:

The main differences between the symptoms induced in cold-blooded and warm-blooded animals, by a fatal dose of hydric cyanide, are as follows:

The respiration in frogs is at first somewhat dyspnoeic, then much slowed, and at length it ceases. The heart, at first slowed, later contracts irregularly, and at length gradually

¹ Proc. Roy. Soc. Ixviii and Ixxii (See Blyth "Poisons" p. 204). 2 Ib., Ixxxiii. (See Blyth "Poisons" p. 204). 3 Journ. Cham. Soc. Ixxxix. (See Blyth "Poisons" p. 204).

stops; but it may continue to beat for several minutes after the respiration has ceased. But all these progressive symptoms are without convulsion. Among warm-blooded animals, on the contrary, convulsions are constant, and the sequence of the symptoms dyspnoea, slowing of the pulse, giddiness, falling down, then convulsions with expulsion of the urine and faeces.

When the dose is short of a fatal one, the symptoms are as follows: Evident giddiness and distress; the tongue is protruded, the breath is taken in short, hurried gasps, there is salivation, and convulsions rapidly set in, preceded, it may be, by a cry. The convulsions pass into paralysis and insensibility. After remaining in the state some time, the animal again wakes up, as it were, very often howls, and is again convulsed; finally, it sinks into a deep sleep, and wakes up well.

Dr. K. Winslow in his work on Veterinary Materia Medica and Therapeutics, gives the Toxicology of Hydrocyanic Acid as follows: "Three stages may be distinguished in fatal poisoning. First: a very short period elapses before the symptoms appear. There are giddiness, difficult breathing, and slow pulse in this stage. Second: the pupils dilate, vomiting may occur, and the animal utters loud cries. Spasmodic defacation, micturition and erections may be present, with convulsions and unconsciousness. Third: the last stage is characterized by collapse, spasms, general paralysis and death. The subacute form of poisoning may ensue and prove fatal, or, owing to the volatile character of the drug, complete recovery may take place within one-half or three-quarters of an hour. Occasionally dogs continue to be paralyzed for several days and get well. The minimum fatal dose recorded in man is 9/10 of a grain of pure acid, or about 50 drops of the medicinal solution. Four to five drachms of the diluted acid frequently, but not invariably, causes subacute poisoning and death, in horses, within an hour. One or two drachms of the pharmacopoeial preparation usually kills dogs within ten minutes."

Poisoning from Toxalbumins, Black Locust, Ricinus and Abrus.

In recent years much work has been done with a class of poisons, known astoxalbumins. These are of especial interest because many of the bacteria produce such poisons. Some of the fungi responsible for "forage poisoning" produce, it is thought, toxalbumins. In recent years a number of cases of horse poisoning from Black Locust bark have been reported. The poisoning from castor oil bean (Ricinus) and from Abrus are also of this class.

Castor Oil Seed and Abrus. One of the best known of the toxalbumins is that occurring in the castor oil seed, known as ricin. This albuminous substance is very poisonous, more so than strychnin and prussic acid. Ricin coagulates the blood. Blyth in his work on poisons states:

If castor-oil seeds are eaten, a portion of the poison is destroyed by the digestive processes; a part is not thus destroyed, but is absorbed, and produces in the blood-vessels its coagulating property. Where this takes place, ulcers naturally form, because isolated small areas are deprived of their blood supply. These areas thus becoming dead, may be digested by the gastric or intestinal fluids. and thus, weeks after, death may be produced. The symptoms noted are nausea, vomiting, colic, diarrhoea, tenesmus, thirst, hot skin, frequent pulse, sweats, headache, jaundice, and death in convulsions or from exhaustion. Animals may be made immune by feeding them carefully with small doses, gradually increased.

The post-mortem appearances are ulceration in the stomach and intestines. In animals the appearances of haemorrhagic gastro-enteritis with diffuse nephritis, haemorrhages in the mesentery, and so forth have been found.

A toxalbumin also occurs in the Jequirity seed (Abrus precatorius) which causes similar effects and symptoms. That the poisons are not the same have been shown by experiments with animals. It is known that animals may become immune by repeated doses of Jequirity against abrin and the principle of castor oil does not produce immunity against abrin, nor does abrin confer immunity against the ricin of the castor oil bean. The abrin when applied to the con-

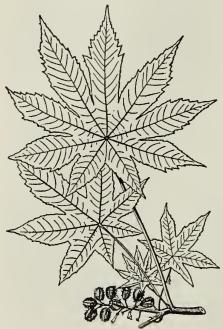


Fig. 16b. Castor oil plant (Ricinus communis). The seeds furnish the castor oil of commerce, and also contain an acrid poison. (Chestnut, U. S. Dept. Agr.).

junctiva causes coagulation in the vessels and a secondary inflamation. The disease is known as Jequirity opthalmia. More details in regard to the poisons of these plants are given under the plants of the families in which they occur.

Black Locust Poisoning. The Black Locust which is commonly planted as an ornamental tree has in a number of instances caused death.

Dr. Waldron in the American Veterinary Rewiew, writes thus of locust bark poisoning, referring especially to the beating of the heart of a horse that had been poisoned by the locust bark. This beating shook the horse and could be heard outside the stable.

The sound was caused by the action of the diaphragm. It was greater when the ribs were at their fullest expansion and could be heard most distinctly at a distance of ten feet. I tried to locate or rather find out what produced the sound, but in that I am as ignorant as I was then. . . . In questioning where the team was hitched at the mill, it was found that the driver had tied them to a young locust tree that had been cut down a few years

before. This was a sapling of about four inches in diameter and had probably made a very rapid growth and the bark, from this reason was tender and easily peeled. They had done a good job of peeling, but as they had their bits in they were not able to swallow much. The poison obtained from this bark is, in my opinion, the cause of the trouble.

Dr. Waldron says that the symptoms otherwise are about the same as those occurring in cases of belladonna poisoning and are about as follows:

Extreme lassitude, which includes almost imperceptible pulse and which, when found, is weak and prolonged; respiration less than normal by one-third and sonorous; temperature normal; no pain, no appetite, mucous membrane congested, of a blue, rusty, or yellow color. Mucous membrane of the mouth some swollen, caused by the congestion of the capillaries; slight ptyalism, and above all, the dilation of the pupil of the eye; in fact I should judge, we have nearly the same symptoms we get in belladonna poison.

He also says that he had not known before that locust bark was poisonous and although he had searched for literature upon the subject had found but one reference, that being in the U. S. Pharmacopoeia ,which records a case reported in Jan., 1887, when 33 children were said to have been poisoned by chewing locust bark. In mild cases there were "flushed faces, dryness of the throat and mouth, and dilation of the pupils. In severe cases, were added epigastric pain, extremely intermittent heartbeats, and stupor." It is evident from the fact that there is not much literature on the subject that such poisoning does not occur often.

Dr. H. S. Murphy has kindly contributed the following case on locust bark poisoning:

Anamusis: Gray mare twelve years old, pregnant ten months, has been at light work

continuously, is driven to town afternoon of March 31, and tied to a locust tree which is 6-8 inches in diameter; owner noticed that animal had peeled bark from tree also noticed animal pant on road home. Anorexia and stupidity were all owner noticed until 8 p. m. when a faint noise was heard this gradually grew worse until I arrived at 7 a. m. April 1.

Symptoms: 1. Animal stands back from manger with legs well apart resting nose and mouth on floor, most of the time, a part of the time head is held 18-24 inches from floor.

- 2. A continual "thud" is heard which is not synchronous with respiration but is synchronous with a vibration of thorax.
- 3. Animal will not obey commands and when pushed over in stall loses balance and nearly falls.
 - 4. Fetus is seen kicking in mare's left hypo-chondriac region.
 - 5. Temperature 99.C Farh.
 - 6. Pulse 54 full, bounding.
 - 7. Respiration, 14, very feeble.



Fig. 16c. Black Locust (Robinia Pseud-acacia) Bark, leaves and flowers poisonous. (After Faguet).

- 8. Mucous membranes dry, very yellow a dirty yellow (in distinction from bright yellow of icterus) swollen, only moderately sensitive (probably due to general depression).
 - 9. Intense dilation of pupil.
 - 10. No evacuation of feces; auscultation reveals paresis of bowels; ropy urine is passed.
- 11. Internal cause of thud was determined as due to heart beat in the following manner:

 1. Pulse over maxillary artery is syncbronous with "thud" and thoracic vibration. 2.

 Owner counts aloud while auscultation of heart is conducted and beart beat is synchronous with thud. 3. Ausculation of heart and radial pulse are syncbronous, also beart sounds are muffled but very loud. (Palpitation).

Treatment: (Symptomatic) 1/4 gr. of Digitalin (P. D.) every fifteen minutes subcutan-

eously until four doses are given, also & I potassi iodidum is given per orem at once and the following prescription left.

Ex Strychnini sulphatis gr. IV.

Fld. ext. digitalis &I.

Potassi iodidum II.

Aqua ξXVI. Sig.: A table spoon full every 2 hours.

Result: After the second hypodermic was given a slight improvement noticed (lessening in force and frequency of thud, which was quite marked after fourth hypodermic. Owner reports thud gone at end of 24 hours but stupidity still present though not so marked.

Recovery uneventful and at end of gestation parturition proceeds normally, and foal

Toxicity of bark proven in the following manner: Two cc of 10% tincture (bark grated) killed half grown kittens which showed above symptoms in an aggravated form.

One ounce necessary to kill a 25 pound dog.

This tincture dropped in eye acted much as atropin but no toxic symptoms developed. Simply a dilation of pupil. Grated root in lard produced slight symptoms in two dogs. P. m. on cats revealed a generalized dirty m. m. but only a very slight yellow on cartilages. Swollen liver and a few petechia on serous membranes. Blood quite dark.

The cadaver, resembled somewhat that of one dead of septicaemia.

CHAPTER IX

POISONING FROM OPIUM, SOLANACEAE AND PLANTS THAT CONTAIN SAPONINS.

The use of opium by Chinese and other races is as Flückiger and Hanbury say, "in the words of Pereira, the most important and valuable used in medicine of the whole Materia Medica; and we may add, the source by its judicious employment of more happiness by mankind." Blyth in his work on Poisons, states that in England and Wales 1505 deaths were attributed to the use of opium or its active constituents between the years 1898-1903. Of these 882 were accidental or because of negligence, 621 were suicidal. In France opium and morphin poisons are said to cause about 1 per cent of the cases of poisoning. Various patent medicines contain opium or some of its products and in the past have been the cause of frequent cases of poisoning. The use of the drug in patent medicines for children in the United States in the form of soothing syrups was once more common than now. The use of opium for infants is a common practice in India, according to Blyth who quotes from Dr. Chevers.¹

In general the opium and morphin poisoning are as follows:. The beats of the heart are at first accelerated and then diminished. Large doses introduced into the circulation diminish the pulsations without acceleration and may even cause heart paralysis. "The arterial blood pressure, at first increased is afterwards diminished. If morphin is in sufficient quantity thrown into the circulation, then tetanus at once occurs. Depression and stimulation depend on dosage. The common form occurring in 99 per cent of the cases; excitement, narcosis, and coma, bowels nearly always constipated. (2) A very sudden form in which death occurs rapidly, the person sinks into a deep sleep almost immediately. (2) An abnormal form in which there is no coma but convulsions.

Blyth in referring to opium eating says:

The consumption of opium is a very ancient practice among Eastern nations, and the picture, drawn by novelist and traveler, of poor, dried-up, yellow mortals addicted to this vice, with their faculties torpid, their skin hanging in wrinkles on their wasted bodies, the conjunctivae tinged with bile, the bowels so inactive that there is scarcely an excretion in the course of a week, the mental faculties verging on idiocy and imbecility, is only true of a percentage of those who are addicted to the habit.

In the case of opium poisoning the stomach tube should be used to empty the stomach, and wash with warm water, then coffee may be given. Permanganate of potash is a perfect antidote and should be given when at hand.

The alkaloid codein also found in opium produces sleep but its effects are different. Large enough doses produce death and cause epilepti-form convulsions. Thebain found in opium produces symptoms that resemble those produced by strychnin, namely tetanic spasms. Apomorphin found in opium is an active emetic. Papaverin causes paralysis of respiration in painea pigs.

¹ Jurisprudence 232 (3rd ed.).

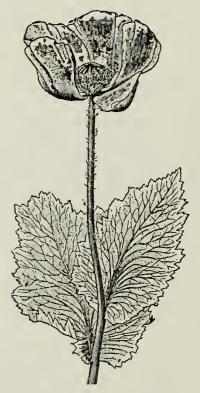


Fig. 16d. Garden Poppy (Papaver somniferum). A poisonous plant producing a large number of alkaloids like morphin, codein, etc., (Strasburger, Noll, Schenek and Schimper).

Poisoning from Solanaceae.

A number of plants of the Solanaceae are known to be poisonous; among them the common thorn-apple or Jimson weed (Datura Stramonium), the atropa (Atropa Belladonna) and hyoscyamin (Hyoscyamus niger) besides such suspected plants as the common black nightshade (Solanum nigrum), horse nettle (Solanum carolinense), bittersweet (Solanum dulcamara) and scopiola. The cases of poisoning from atropin are more frequent, perhaps, than statistics seem to indicate. The English death statistics for ten years, ending 1903, according to Blyth show 95 per cent of the deaths from atropin; 35 per cent were suicidal. Most of the accidental cases arise from mistakes made by the pharmacist or physician. Criminal poisoning is carried on to a less extent in Europe and America than in India. Blyth states that of the 120 cases recorded in works on Indian toxicology no less than 63 per cent were criminals, 19 per cent suicidal, and 18 per cent accidental. The most important alkaloids found are atropin, hyoscyamin, scopalamin and solanin. Solanin is poisonous and is regarded as a nitrogenised glucoside. In man the symptoms of atropin poisoning are: Dilating of the pupils, dryness of the mouth and throat; the

mucous membrane is reddened, inability to swallow, deranged vision, breath at first a little slow and then rapid; the nervous system is affected in a marked degree; the lower extremities are often partly paralyzed. There is want of coördination. "The person reels like a drunken man." In adults this takes on a hilarious pleasing form. The symptoms of poisoning from hyoscyamin are similar to those of atropin. The absence of delirium and excitement, however, makes it decidedly different. The symptoms in animals for both of these substances do not differ essentially from those given above

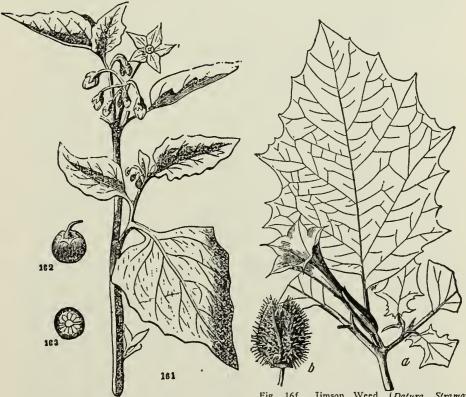


Fig. 16c. Nightshade (Solanum nigrum)
From Darlington's Weeds and Useful Plants.

Fig. 16f. Jimson Weed (Datura Stramonium). a, leaf and flowers; b, fruiting capsule.

(U. S. Dept. Agrl.).

Atropin may be absorbed by the skin, and enough may be absorbed, if it is broken, to cause death. Blyth quotes Ploss 1 to the effect that atropin sulfate applied as an ointment to the abraded skin was fatal. Atropin has also been absorbed from the bowel as recorded by Blyth:

A clyster containing the active principles of 5.2 grms. (80 grains) of belladonna root was administered to a woman 27 years of age, and caused death. Allowing the root to have been carefully dried, and to contain .21 per cent of alkaloid, it would seem that so little as 10.9 mgrms. (.16 grains) may even prove fatal, if left in contact with the intestinal mucous membrane. Belladonna berries and stramonium leaves and seeds are eaten occasionally by children. A remarkable series of poisoning by belladonna berries occurred in London during the autumn of 1846.

¹ Zeitschr f. Chir. 1863. Blyth, Poisons; Their Effects and Detection.

Poisoning from Plants that contain Saponin.

In recent years our knowledge of the Saponins has been greatly extended; many of these studies have been made by Kobert or his students in the laboratory at Dorpat. The term saponin has been applied to a class of substances of a glucosidal nature which are poisonous and when dissolved in water form a solution which froths much like soap-suds. These substances are not all the same chemically, but have the general formula CnH2nO10. Blyth gives the following list with their formulae:

Saponin 1.

Saponin senegin
Quillaja sapotoxin
Sapindus sapotoxin
Cypsophila sapotoxin
Agrostemma sapotoxin

Saponin 2.

Asamin
Digitonin saporubrin

Saponin 3.

Quillagic acid
Polygalic acid
Herniaria saponin
Cyclamin
Sarsaparilla saponin

Sarsa saponin.

Cap $H_{30}O_{10}$. $C_{18}H_{28}O_{16}$. $C_{18}H_{28}O_{16}$. $C_{19}H_{30}O_{10}$. $C_{20}H_{30}O_{10}$.

Parillin $C_{20}H_{30}O_{10}$. $C_{20}H_{30}O_{10}$.

The suggestion is made that possibly dulcamarin $C_{22}H_{34}O_{10}$, and syringin $C_{12}H_{50}O_{10}$ may belong to the same series.

One of the oldest of the known saponins was isolated from the Bouncing Betty, Saponaria officinalis, and later from the corn cockle Agrostemma Githago and many other plants. This saponin is a white amorphous powder, very soluble in water, is neutral and reacts without odor; it causes sneezing when applied to the mucous membrane of the nose; tastes at first sweetish, then becomes sharp and acrid. The saponin when rubbed on the skin exerts no action because not absorbed; when injected subcutaneously into frogs it becomes quickly absorbed and acts upon the nerves and muscles. In warm blooded animals there is little or no absorption because of an aseptic abscess which forms. Intravenous injections in small amounts in the laboratory of Kobert proved fatal for cats and dogs. It acts injuriously on the striated muscle and heart muscle. The sensor and motor nerve fibers are also affected in a serious way. On the digestive tract it causes inflammation and peristalsis. The saponin substances dissolve the blood corpuscles of all animals and thus penetrate the corpuscles. It is thought that the haemolytic action of these substances is due to the liquefaction of the cell membrane.

Ransom found that the saponin may become bound to the corpuscles and the serum. That this action depends on the cholosterin, saponin so bound will not act on the red corpuscles. The saponin cholosterin mixture exerts no action on dog's blood.¹ Dr. R. F. Bacon and H. T. Marshall who made a study

¹ The toxic action of saponin. Phil. Jour. Sci. 1:1037. Dr. R. F. Bacon and H. T. Marshall.

of the saponin found in *Entada scandens* proved that it was highly toxic for rabbits and guinea pigs.

When diluted with normal salt solution to a concentrated form 1-200 and injected into the peritoneal cavity, 2-5 mgrms. of saponin to 100 grms. of animal was uniformily fatal, while a quarter of this amount killed in several cases. Where smaller quantities were used and animals living for a longer period of time localized peritotinitis was found.

"The saponin is powerfully haemolytic" 0.005 cubic cm. of ½ of 1% solution (0.025 mgrms.) completely dissolved one cubic cm. of a 5% suspension of rabbits corpuscles which were freed from serum and the half of this amount produced haemolysis of the serum of the free corpuscles of the guinea pig. "Saponin, however, loses its haemolytic power after the addition of serum." Immunity could not be produced in rabbits or guinea pigs recently treated with intraperitoneal doses of saponin or saponin serum mixture.

Blyth has studied the general action of saponin on kittens. He states that when 13 to 22 mm. (1/5 to ½ gr.) is injected underneath the skin of a kitten immediately symptoms of local pain occur, in 5 to 10 minutes the respiration is quicker and the animal falls into a lethargic condition with signs of muscular weakness; just before death breathing became rapid with all the signs of asphyxia. The appearances after death were fullness in the right side of the heart and congestion of the intestinal canal. In man the taking of saponin causes an increase of mucus secretion and nausea.

Saponin or saponin-like substances occur in the following families:

Phytolaccaceae, (Phytolacca abyssinica); Caryophyllaceae, (Gypsophila struthium, Agrostemma Githago, Lychnis, Saponaria officinalis, S. rubra, Herniaria); Berberidaceae, (Caulophyllum thalictroides); Leguminosae, (Entada scandens, Gymnocladus dioica, Gleditschia, Enterolobium): Oleaceae, (Chionanthus virginica, Syringa vulgaris); Amaryllidaceae, (Agave); Liliaceae, (Yuccaglauca?, Chlorogalum pomeridianum); Rosaceae, (Quillaja Saponaria); Sapindaceae, (Sapindus trifoliatus, S. saponaria, Pometia pinnata, Magonia); Hippocastanaceae, (Aescula Hippocastanum, A. Pavia); Theaceae, (Thea Sasanqua, T. assanica); Sapotaceae, (Omphalocarpum procerum); Polygalaceae, (Polygala Senega).

In all more than 200 species of plants contain saponin. See Chapter XIV, and Pt. II, for a list of others.

CHAPTER X

POISONING FROM FLOWERS, POISONING FROM HONEY, MECHANICAL INJURIES

Poisoning from Flowers. The odors from a large number of flowers are more or less injurious, especially to some individuals. The flowers of the common wild black cherry (Prunus serotina) when past their prime give off a cyanogentic odor that is quite objectionable to many people, causing headache. The carion-like odors, like those produced by Stapelia, the carion flower (Smilax herbacea) and Aristolochia are sickening to many people, causing headache and a depressing feeling. Many flowers that are ordinarily sweet scented like the tuberose (Polyanthes tuberosa) often give people the headache if the flowers are abundant in the room they are sitting in. The flowers of Wistaria are injurious to some people. The flowers of Magnolia grandiflora are "overpowering" according to some authorities.

Poisoning from Honey. The honey obtained from the flowers of some plants is said to be poisonous; for instance the honey collected by insects from the oleander was long ago recorded as being injurious, and it is said that the honey collected from the mountain laurel (Kalmia latifolia) appears to be poisonous under some conditions according to Chesnut. According to Chesnut the honey collected from the flowers of the snow-on-the-mountain, (Euphorbia marginata) is bitter and disagreeable, but does not appear to be a serious poison. Several cases of poisoning from the nectar of the flowers of Datura metel and D. Wrightii are on record and the flowers of the Brazilian magonia of the family Sapindaceae produces poisonous honey; also the flowers of Rhododendron are said to contain andromedotoxin. Toxic honey has also been gathered from the flowers of Black Locust (Robinia pseud-acacia) and Lily of the Valley (Convallaria majalis).

Prof. Lyman F. Kebler who has made a somewhat extended investigation with poisonous honey 1 has given an excellent bibliography with reference to the earlier literature on the subject. It has been known for centuries that the honey collected from Ericaceae acts as a narcotic irritant, producing giddiness, vomiting, and purging. Poisonous honey was described by Xenophon. He gives a fairly accurate description of how the soldiers of his army acted that ate honey that was poisoned. He states that they lost their senses, vomited and were affected with purging, and those who had eaten but little were intoxicated, but when they had eaten much they were like mad men. Strabo and Pliny spoke of poisonous honey, the latter writer, an early naturalist noted for his accurate observations, records poisonous honey which he called "aegolethron" (goat's death), which bees collected at Heraclea. He gives a description of the honey which is said to have had a peculiar smell and produced sneezing. It is generally supposed that this honey came from a species of Rhododendron, the R. pontica. This and allied species are the chief source of poisonous honey in Asia and Asia Minor,

¹ Poisonous Honey. Proc. Amer. Pharm. Assoc. 1896: 167-173.

but it may be said in this connection that honey collected from the Heather in Scotland is not poisonous.

Barton, an early American botanist, reported poisonous honey in New Jersey as early as 1794. Subsequently Coleman reported a large number of cases in 1852, and Gammer in Gleanings in Bee Culture and several writers in the American Bee Journal reported poisonous honey. Other writers like Chesnut and Crawford have reported on the occurrence of poisonous honey in the United States and Kebler reports no less than eight cases for New Jersey in 1896 and . believes that it is much more common than the records seem to indicate. Kebler was fortunate enough to investigate some of the poisonous honey following a case of poisoning in New Jersey. He examined a part of the comb of the dark honey which had a light brown color and a nauseating odor, pungent taste, caused a burning sensation in the back of the mouth similar to that of aconite. Persons who partook a small amount of this honey began to cough immediately. He also made a chemical analysis of the non-suspicious honey, digesting with alcohol then evaporated, the residue was again treated to alcohol and evaporated and administered to two cats. One received a small dose and the other a larger one. The results from the cats were so interesting that I quote from Prof. Kebler.1

The small dose produced partial exhaustion, relaxation of the voluntary muscles and general depression. The large dose in a short time produced restlessness, vomiting, purging, prostration and almost complete loss of the voluntary muscles, showing that the honey contained a prompt and potent poison. The animal could scarcely be induced to move, and when motion was attempted, first the fore-limbs would fail, and then the back limbs would give way. First one portion of the body would sway in one direction, then the other portion in another, reminding one of a highly intoxicated person. Had the entire dose been retained, death undoubtedly would have followed. As it was, the cat had regained her normal condition only at the end of twenty-four hours.

Along with this we may append the symptoms as recorded by the physician who attended the persons who were poisoned:

Mr. and Mrs. Chambers took but a small quantity, yet each noticed a peculiar, pungent, burning taste in the comb as soon as it had passed their lips. In fifteen or twenty minutes afterwards, Mrs. C. was taken with nausea, abdominal pain, and vomiting, soon followed by loss of consciousness, coldness of extremities, feebly acting heart, and complete collapse. While ministering to her, Mr. Chambers, who had also experienced the initiatory symptoms of pain and nausea, suddenly exclaimed, "I cannot see," and soon sank in a state of syncope to the floor. In each case the symptoms were similar. Retching, vomiting, purging, acute gastric and abdominal pain, and continued cramps for some hours, with surface coldness, and deadly pallor, and the general symptoms of collapse.

Kebler was, however, unable to definitely locate the andromedotoxin found by Plugge.² This author recorded it for a large number of plants as follows: Andromeda japonica, A. polifolia, A. Catesbaei,, A. calyculata, Kalmia latifolia, Monotropa uniflora, Pieris formosa, P. ovalifolia, Rhododendron grande, R. barbatum, and R. fulgens. It has been recorded for additional plants by Greshoff who mentions the following plants which produce poisonous honey, Nerium oleander, Cytisus Laburnum, Pieris ovalifolia, Callotropis procera, Daphne, Pontiea, Buxus balearica, Clerodendron serratum, C. Bhramaramari, Sapindus emarginatus. (It is said that thousands of bees are killed by this honey.) Centaurea seabiosa, Carduus natans, Scabiosa suceisa. A South African species of Euphorbia also produces a poisonous honey which was not noted by Greshoff.

Proc. Amer. Pharma. Assoc., 1896:170.
 Arch. d. Pharm., 229; 552, 1891; Am. Jour. Pharm. 63: 603. Shresh, J. C. Notes on Trebizonde Honey. Pharm. Jour. and Trans., 18:397, 404. 1887-88.



Fig. 16g. The flowers of Black Locust (Robinia pseud-acacia) produces a toxic nectar. (Ada Hayden).

Fig. 16h. The wilted and old flowers of Wild Cherry (Prunus) produce a cyanogentic poisonous odor. (Ada Hayden).

We have a long list of plants that act injuriously Mechanical Injuries in a mechanical way. Among the best known of these are Wild Barley or Squirrel-tail Grass (Hordeum jubatum) and the related species, which, by mechanical means, injure sheep, horses, and cattle. The awned heads, when eaten with hay or grass, break up into sections, the awns working their way into the mucous membrane, insinuating themselves around the teeth, thus causing inflammation and deep ulcerating sores, with the formation of pus. The teeth may consequently become loosened and fall out.

A Cheat or Brome Grass (Bromus tectorum) which is common in Utah, parts of Colorado, and westward, produces similar injuries.

The Needle Grass, common in the dry gravelly hills and sandy plains of Northern Mississippi Valley has a fruit with a sharp pointed callus, with hairs above the pointed callus projecting upwards. The sharp-pointed callus of the "seed" enters the skin, especially of sheep, where it produces an irritation which is sometimes followed by death. This has sometimes been eaten with forage, thus entering the intestinal tract, perhaps perforating it, causing death, especially when they have pierced the walls of the intestines. Several allied species as Black Oat (Stipa avenacea) produce similar injuries. The Western Needle Grass (Stipa comata), however, is less injurious than our Common Needle Grass.

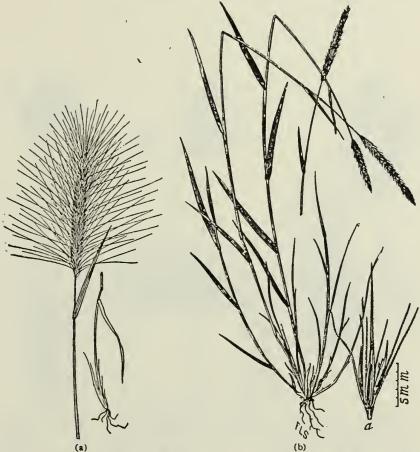


Fig. 16i. Wild Barley (Hordcum jubatum). b. Wild Barley (H. nodosum), cause of mechanical injuries to animals.

Similar and allied species in other parts of the world are known to be injurious in the same way. Other plants are injurious by means of their sharp-pointed fruit. Among those of the last named type are members of the Geranium family notably the Stork's bill (*Erodium cicutarium*), common in the west, which frequently gets into the wool of sheep and may produce local irritation. Mechanical injuries are also produced by the Sand Bur (*Cenchrus tribuloides*) whose spiny involucre may work into the flesh of animals and induce an irritation followed by inflammation, and the formation of pus. Hogs and cattle often receive mechanical injuries from the Cocklebur (*Xanthium canadense*) and allied species.

A recent number of Horticulture (Jan. 1, 1910) notes that rose thorns have caused the poisoning of the hands of clerks who handle the roses.

Prof. J. Davy states that in South Africa the burrs of a clover Trifolium terrestris var hispidissimus are said to injure young lambs.

The Burdock (Lappa major) sometimes produces no little irritation, and the same may be said of the Spanish Needle (Bidens frondosa). The sharp, stiff

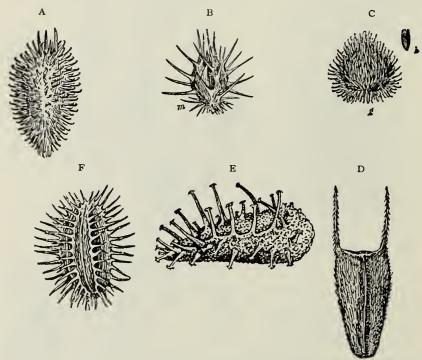


Fig. 17. A. Fruit of the Cocklebur (Xanthium canadense), causes mechanical injuries to hogs when eaten by them. B. Sandbur (Cenchrus tribuloides). C. Burdock (Arctium major). D. Boot Jack (Bidens frondosa). F. One of the Borages (Cynoglossum), F. Carrot (Daucus Carota).

branches of Greasewood (Sarcobatus vermiculatus) easily penetrate the skin and sometimes induce pus infection. The bristles and spines of the rose and stiff bracts of the Russian Thistle may be the cause of injury to animals and men. The small, barbed trichomes of the calyx of Crimson Clover (Trifolium incarnatum), according to Prof. Coville, produce phytobezoars similar to those commonly produced by hair. Dr. Trelease has described similar bezoars from the barbed trichomes and spines of cacti. Millet and barley awns are known to produce similar balls in horses as well as in cattle. Corn stalks, when eaten with an insufficient amount of water, produce impaction. This has, however, been attributed to other causes also. Bloat is known to follow the use of such forage as white and red clover, alfalfa, pigweed and many other plants consumed under the same conditions.

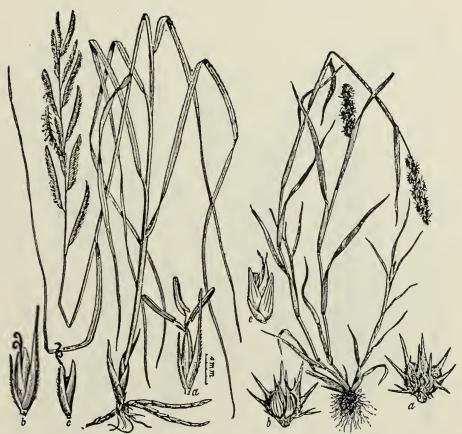


Fig. 17a. Fresh water cord grass (Spartina cynosuroides) with sharp leaves often injurious. 17b. Sand Bur (Cenchrus tribuloides) U. S. Dept. Agrl.

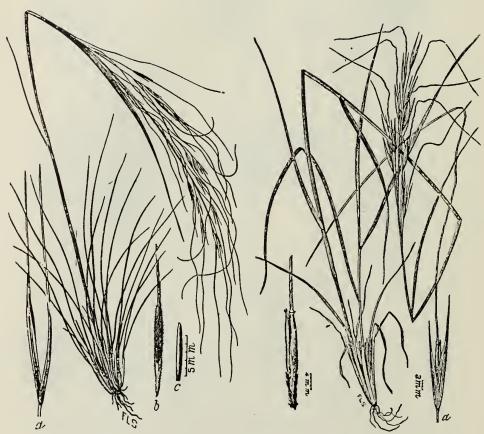


Fig. 18. Needle Grass (Stipa comata). U. S. Dept. Agrl. Fig. 18a. Needle Grass (Stipa spartea) inflicting mechanical injuries. U. S. Dept. Agrl.

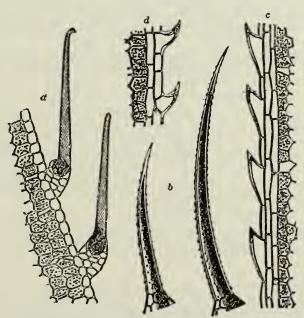


Fig. 18b. Urticating hairs and cutting leaves. a. Urticating hair of nettle. b. Bristles of bugloss. c. barbed margin of a leaf of sedge. d. barbed margin of a leaf of grass.

CHAPTER XI

CLASSIFICATION OF POISONS, SYMPTOMS AND ANTIDOTES

Blyth classifies poisons as follows:

- A. Poisons causing death immediately or in a few minutes. Prussic acid, cyanides, oxalic acid and occasionally, strychnin.
- B. IRRITANT POISONS. Symptoms mainly pain, vomiting, and purging. Savin, ergot, digitalis, colchicum, yew, laburnum, and putrid substances.
- C. IRRITANT AND NARCOTIC POISONS. Symptoms of an irritant nature, with more or less cerebral indications. Oxalic acid or oxalates.
 - D. Poisons more especially affecting the Nervous System.
- 1. Narcotics. Chief symptoms: insensibility which may be preceded by more or less cerebral excitement. Opium.
- 2. Deliriants. Delirium, for the most part, a prominent symptom: Belladonna, hyoscyamus, stramonium, and other Solanaceae, poisonous fungi, Indian hemp (Cannabis), darnel (Lolium temulentum), camphor and Oenanthe crocata.
- 3. Convulsives. Almost every poison has been known to produce convulsive effects, but the only true convulsive poisons are the alkaloids of the strychnin class.
- 4. Complex nervous phenomena. Aconite, digitalis, poison hemlock, Calabar bean, tobacco, *Lobelia inflata*, and curare.

Kobert's classification is as follows:

- I. Poisons which cause coarse anatomical changes of the organs.
- A. Those which especially irritate the part to which they are applied.
 - 1. Acids.
 - 2. Caustic alkalies.
 - 3. Caustic salts, especially those of the heavy metals.
- 4. Locally irritating organic substances which neither can be classified as corrosive acids nor alkalies, nor as corrosive salts; such are: cantharadin phrynin, and others in the animal kingdom, croton oil and savin in the vegetable kingdom; locally irritating colors, such as the anilin dyes.
- 5. Gases and vapors which cause local irritation when breathed; such as ammonia, chlorin, iodin, bromin, and sulphur dioxid.
- B. Those which have but little effect locally, but change anatomically other parts of the body; such as lead, phosphorus, and others.

 II. BLOOD POISONS.
- 1. Blood poisons interfering with the circulation in a purely physical manner; such as peroxid of hydrogen, ricin, abrin.
- 2. Poisons which have the property of dissolving the red corpuscles; such as the saponins.
- 3. Poisons which, with or without primary solution of the red blood corpuscles, produce in the blood methaemoglobin; such as potassic chlorate, hydrazine, nitrobenzene, anilin, picric acid, carbon disulphid.

- 4. Poisons having a peculiar action on the coloring matter of the blood, or on its decomposition products; such as hydric cyanid, and the cyanides and carbon monoxid.
- III. Poisons which kill without the production of coarse anatomical change.
- 1. Poisons affecting the cerebro-spinal system; such as chloroform, ether, nitrous oxid, alcohol, chloral, cocain, atropin, morphin, nicotin, coniin, aconitin, strychnin, curarin, and others.
 - 2. Heart poisons; such as digitalis, helleborin, muscarin.
 - IV. Poisonous product of tissue change.
 - 1. Poisonous albumin.
 - 2. Poisons developed in food.
 - 3. Auto-poisoning, e. g. uraemia, glycosuria, oxaluria.
- 4. The more important products of tissue change; such as, fatty acids, oxy-acids, amido-fatty acids, amines, diamines, and ptomaines.

CLASSIFICATION OF POISONS AND POISONOUS SYMPTOMS, ACCORDING TO BERNHARD H. SMITH

POISONS ACTING ON THE BRAIN

Ι

NARCOTICS

Symptoms. Giddiness; dimness of sight; contracted pupils; headache; noises in the ears; confusion of ideas, and drowsiness, passing into insensibility.

Treatment. (Immediate):

EMETICS (especially mustard, a tablespoonful in half a tumbler of warm water).

STOMACH TUBE. (Wash out at half-hourly intervals with Potash Permanganate solution B. P. 1 per cent solution diluted three times with warm water). Dash cold water on face and chest. Ammonia or Amyl Nitrite to nostrils.

(Later:)

TANNIN. (Hot, strong coffee, tea, or infusions of nut-galls or oak-bark). Strychnin solution B. P. (1per cent solution) two minims subcutaneously. Atropin, half grain subcutaneously, repeated if necessary.

Faradic current. Oxygen.

Artificial respiration if necessary.

DEMULCENTS. (Milk, eggs, oil, etc., swallowed and injected).

Note—All vegetable poisons act more quickly on the empty stomach; our treatment therefore is directed towards (1) diluting and evacuating the poison by means of washing out with warm water containing powdered charcoal; (2) neutralizing the poison by means of an antidote such as Tannin.

Plant producing poisons that act on the brain as narcotics—Poppy (Papaver somniferum).

Η

DELIRIANTS

Symptoms. Spectral illusions; delirium; dilated pupils; thirst, and dryness of the mouth; inco-ordination. Occasionally, though rarely, there are paralysis and tetanoid spasms.

Treatment. (Immediate:)

EMETIC and STOMACH TUBE.

Pilocarpin Nitrate, one-third of a grain subcutaneously, repeated if necessary.

Ammonia or Amyl Nitrite to nostrils.

TANNIN.

(Later:)

Stimulants, Castor Oil.

DEMULCENTS (swallowed and injected).

Note—Muscarin poisoning give Belladonna Tincture B. P. fifteen minims Plants furnishing poisons that act as deliriants—Thorn Apple (Datura Stramonium). Black Nightshade (Solanum nigrum), Hemp (Cannabis sativa), Darnel (Lolium temulentum), Several Fungi as Fly Agaric (Amanita muscaria).

Ш

INEBRIANTS

Symptoms. Excitement of cerebral functions, and of the circulation; loss of power of co-ordination, and of muscular movements, with double vision; leading to profound sleep, and deep coma.

Treatment. (Immediate:)

EMETIC and STOMACH TUBE.

TANNIN.

(Later:)

Epsom Salts.

DEMULCENTS (swallowed and injected).

Plants furnishing poisons that act on the brain as inebriants: Wormwood (Artemisia Absinthium), Jamaica Dogwood (Piscidia Erythrina).

POISONS ACTING ON THE SPINAL CORD

CONVULSIVES

Symptoms. Clonic (intermittant) spasms, extending from above downwards. Opisthotonos very violent; but trismus (lock-jaw) rare. Swallowing spasmodic. Death, usually, in less than three hours, or rapid recovery.

Treatment. (Immediate:)

EMETIC and STOMACH TUBE. TANNIN.

Chloral Hydrate five grains subcutaneously (a weak solution as it is an irritant), repeated if necessary. Chloroform inhalation. Artificial respiration.

Potassium Bromide, one drachm in water every half-hour. Morphia (?).

DEMULCENTS (swallowed and injected).

Castor Oil. Chloroform inhalation if convulsions return.

Plants furnishing poisons that act upon the spinal cord as convulsives: Nux-vomica Tree (Strychnos Nux-vomica), St. Ignatius's Bean (Strychnos Ignatii).

POISONS ACTING ON THE HEART

Ι

DEPRESSANTS

Symptoms. Vertigo; vomiting; abdominal pain; confused vision; convulsions; occasional delirium; paralysis; syncope; sometimes asphyxia.

Treatment. (Immediate:)

EMETIC and STOMACH TUBE. TANNIN.

STIMULANTS. Strychnin solution B. P., two minims subcutaneously.

Atropin, half a grain subcutaneously, repeated if necessary.

(Later:)

Stimulants. Hot fomentations.

Artificial respiration if necessary. Castor Oil.

DEMULCENTS (swallowed and injected).

Plants furnishing poisons that act on the heart as depressants: Tobacco (Nicotiana Tabacum), Hemlock (Conium maculatum), Indian Tobacco (Lobelia inflata).

H

ASTHENICS

Symptoms. Numbness, and tingling in the mouth; abdominal pain; vertigo; vomiting; purging; tremor; occasional delirium; paralysis; dyspnoea, ending in syncope.

Treatment. (Immediate:)

EMETIC and STOMACH TUBE. TANNIN.

STIMULANTS. Cold affusion.

Faradic current. Atropin, half a grain subcutaneously.

(Later:)

DEMULCENTS (swallowed and injected).

Continued recumbent position.

Artificial respiration if necessary. Castor Oil.

Note—In Aconite poisoning inject Digitalis Tincture B. P., twenty minims subcutaneously (12½ per cent strength).

Plants furnishing poisons that act on the heart as asthenics: Lima Bean (Phaseolus lunatus), Tapioca (Jatropha manihot), Aconite (Aconitum Napellus), Cohosh (Cimicifuga racemosa), Oleander (Nerium Oleander), Fox-glove (Digitalis purpurea), White Hellebore (Veratrum album), Green Hellebore (Veratrum viride).

VEGETABLE IRRITANTS

I

PURGATIVES

Symptoms. Abdominal pain; vomiting, and purging; cramps; strangury and tenesmus, followed by collapse, and sometimes accompanied by drowsiness, and slight nervous symptoms.

Treatment. (Immediate:)

EMETIC and STOMACH TUBE. TANNIN.

(Later:)

Opium to relieve pain.

Stimulants to counteract collapse.

DEMULCENTS (swallowed and injected).

Plants producing poisons that act as purgatives: Castor Bean (Ricinus communis), Green Hellebore (Helleborus viridis), May Apple (Podophyllum peltatum), Marsh Marigold (Caltha palustris).

Π

ABORTIVES

Symptoms. Nausea; vomiting; stupor; polyuria; sometimes tenesmus. Abortion may or may not occur; coma.

Treatment, (Immediate:)

EMETIC and STOMACH TUBE. TANNIN.

Ammonia or Amyl Nitrite to nostrils.

(Later:)

Opium to relieve pain.

Stimulants to counteract collapse.

DEMULCENTS (swallowed and injected).

Plants producing poisons that act as abortives: Ergot (Claviceps purpurea), Herb of Grace (Ruta graveolens), Cotton root (Gossypium herbaceum), Pulsatilla (Anemone patens, and its variety).

III

IRRITANTS WITH NERVOUS SYMPTOMS

Symptoms. Abdominal pain; vomiting and purging; dilated pupils; headache; tetanic spasms; occasional convulsions; sometimes rapid coma.

Treatment. (Immediate:)

EMETIC and STOMACH TUBE. TANNIN.

Opium to relieve pain.

Stimulants to counteract collapse.

Bleeding if necessary.

(Later:)

Castor Oil.

DEMULCENTS (swallowed and injected).

Plants furnishing poisons that act as irritants, causing also nervous symptoms: Indian Pink (Spigelia marilandica), Cut-leaved Water Parsnip (Berula erecta), Fool's Parsley (Aethusa Cynapium).

IV

SIMPLE IRRITANTS

Symptoms. Burning pain in the throat and stomach; thirst; nausea; vomiting; tenesmus; purging; dysuria; dyspnoea and cough occasionally. Death through shock; convulsions; exhaustion; or starvation due to injury to throat or stomach.

Some few (i. e. the Nettles) cause smarting pain on the merest contact with the secretions of the plant; quickly followed by erythema and urticarial rash, which slowly subsides.

Treatment. (Immediate:)

EMETIC and STOMACH TUBE. TANNIN.

Opium to relieve pain.

Stimulants to counteract collapse.

Chloral and Bromides if convulsions.

(Later:)

Castor Oil.

DEMULCENTS (swallowed and injected).

Plants producing poisons that cause the above symptoms: Arrow Root

(Arum maculatum), Wood Anemone (Anemone nemorasa), Cursed Crowfoot (Ranunculus sceleratus), Buttercup (Ranunculus acris), Bouncing Betty (Sapanaria afficinalis), Kinnikinnik (Arctastaphylos Uva-ursi), Sundew (Drosera rotundifolia), Poison Ivy (Rhus Toxicodendron), Nettle (Urtica dioica U. gracilis), Wood Nettle (Laportea canadensis), Bull Nettle (Jatropha stimulosa).

V

SIMPLE IRRITANTS WHEN TAKEN IN LARGE QUANTITIES

Symptams. Burning pain in throat and stomach. Vomiting; purging; difficulty in swallowing. Recovery usual.

Treatment. (Immediate:)

EMETIC and STOMACH TUBE. TANNIN.

Opium to relieve pain.

Stimulants to counteract collapse.

(Later:)

Castor Oil.

DEMULCENTS (swallowed and injected).

Plants affording poisons that act as simple irritants when taken in large quantities only: White Mustard (Brassica alba), Black Mustard (B. nigra), Black Pepper (Piper nigrum), Common Ginger (Zingiber officinalis), Cayenne Pepper (Capsicum annuum).

TABLE OF SYMPTOMS OBSERVED AFTER THE ADMINISTRA-TION OF POISONS, ADAPTED FROM CATTELL AFTER THE WORK OF KOBERT.

ACUTE SYMPTOMS PRESENT

- 1. Death within a few seconds or minutes.
- 2. Deep coma.
- 3. Collapse.
- 4. Feverish rise of tempera-
- 5. Mania; furious delirium; psychic excitement.
- 6. Mental disturbance of the most diverse kind.
- 7. Violent at times, tetanic convulsions.
- 8. General paralysis, for the most part ascending.
- 9. Dilation of the pupil.
- 10. Contraction of the pupil.
- 11. Aneurosis.
- 12. Diplodia and ptosis.
- 13. Conjunctivitis.

WE SHOULD THINK OF

Hydrocyanic acid; potassium cyanid; carbonic acid; carbolic acid.

Alcohol; morphin; opium.

Nicotin; colchicin.

Cocain; enzymes.

Chronic alcoholism; atropin; camphor; physostigmin; veratrin.

Alcoholism; morphinism; cocainism; pellagra; ergotism.

Strychnin; toxin of tetanus; cytisin; cornutin; picrotoxin; cicutoxin; active principle of digitalis; cocain; santonin; acouitin; gelsemin; filicic acid.

Coniin; curarin; colchicin.

Atropin; hyoscyamin; scopolamin; cocain; ephedrin; aconitin.

Muscarin; pilocarpin; nicotin; codein; opium; physostigmin.

Quinin; extract of male fern; belladonna; uraemic poisoning.

Sausage (botulism) and fish poisoning.

Irritating vapors; ethereal oil of mustard;

- 14. Moist skin.
- Skin conspicuously dry even in a warmed bed. Mouth and throat parched.
- 16. Urticaria or scarlatiniform erythema.
- 17. Eczematous eruptions of skin.
- 18. Diffuse dermatitis with perspiration of the hands.
- 19. Acne pustules.
- Blisters on the skin or the mouth, containing clear serum.
- 21. Gangrenous ergotism; carbolism.
- 22. Cyanosis.
- Yellowish-brown discoloration of the conjunctiva, in combination with that of the skin.
- 24. Discoloration primarily of the tongue and mucous membrane of the mouth.
- 25. Salivation.
- Metallic cough and aphonia.
- 27. Oedema of the glottis.
- 28. Oedema of the lungs.
- 29. Increased dullness of the liver.
- 30. Diarrhoea with vomiting.
- 31. Vomiting without diarrhoea.
- Diarrhoea without vomiting.
- Pulse continuously and markedly becoming slower.
- Pulse first slower, then irregular, finally accelerated.
- 35. Pulse greatly accelerated.
- 36. Abortion.

croton oil vapor; irritating kinds of dust as roots of ipecac, quillaja bark, pepper, formalin, etc.

Opium; morphin; aconitin; pilocarpin; nicotin; physostigmin; lobelin.

Atropin; belladonna; stramonium; hyoscyamus; hyoscyamin; scopalamin; sausage and fish poisoning.

Atropin; hyoscyamin; antipyrin; quinin; balsam of copaiba; cubebene; morphin; handling of nettles; buckwheat; smartweed.

Croton oil; curcas oil; cardol; Rhus Toxicodendron; powdered cinchona bark; carbolic acid; tar.

Anilin colors; aurantia; butter yellow.

Powdered ipecac.

Ranunculus acris; R. sccleratus, etc.

Ergot.

Antifebrin; exalgin; anilin.

Helvellie acid; lupinotoxin (ietrogen).

Carbolic acid.

Pilocarpin; muscarin; arccolin; nicotin; cornutin; physostigmin; cytisin; saponin.

Atropin; hyoscyamin; scopolamin; sausage poisoning.

All caustic poisons.

Muscarin; morphin; pilocarpin; nitric acid vapors.

Agaricus bulbosus; poley oil; alcohol.

Digitalin; pilocarpin; nicotin; muscarin; colchicin; corrosive poisons; colocynthin; emetin; cephalin; croton oil.

Apomorphin; lobelin; cytisin.

Jalap; podophyllotoxin; croton oil; calomel.

Opium; morphin; muscarin; arecolin; physostigmin; all narcotics.

Digitalin; hellebore; adonis; coronilla; cheirathin; nervin; scilla; strophanthus; convallaria; pilocarpin; nicotin; scopolamin.

Belladonna; hyoscyamin; atropin.

Sabina; thuja, rue; mentha; pulegium; ergot; cotton root.

37. 6-12 hour period of good health between the poisoning and the appearance of the symptoms.

Most of the poisonous fungi, but especially Amanita phalloides.

TREATMENT FOR POISONING. (CHIEFLY AFTER KOBERT).

KIND OF POISON

Aconite.

Alcoholism, acute.

Aspergillosis

Atropin

Carbolic acid.

Cicuta virosa. C. maculata cowbane.

Cicutoxin.

Cocain.

Colchicin.

Coniin.

Conium. Cytisin.

Delphinosis.

Dermatitis. Rhus.

TREATMENT.

Usc stomach pump at once; give emetics of sulphate of zinc, or a hypodermic injection of apomorphin; patient should recline; when stomach has been evacuated, give atropin (4 drops U. S. P. solution) hypodermically or by the mouth, or 20 drops of tincture of belladonna; if there is a tendency to heart-syncope, give tincture of digitalis, in ½ drachm doses by mouth, or hypodermically in doses from 10 drops upwards; apply mustard plasters to pericardium; aid vomiting by plenty of water; if necessary apply artificial respiration.

Wash out the stomach with a siphon tube; cause vomiting by emetics, cold and hot effusions alternated; strychnin hypodermically.

Treatment unsatisfactory; moulds that have entered lungs cannot be destroyed.

Wash out the stomach with a solution of tannic acid or cause evacuation with an emetic; hypodermic injection of strychnin to stimulate respiration; administer tea or whiskey.

A liberal dose of whiskey or alcohol as a diluent; use soft stomach tube to wash out the stomach with sodium sulfate; sodium sulfate, raw eggs, milk and saccharate of lime are antidotes.

Tannin and narcotics according to symptoms, especially chloral hydrate.

Chloroform, chloral hydate, artificial respiration, stimulants like whiskey.

Evacuate the stomach; then give vegetable astringents, iodin 1 gr. and potassium iodid 10 gr. dissolved in water; digitalis and amyl nitrite are given for syncope; give stimulants like oxygen and whiskey for cyanosis.

Mucilaginous potions, morphin, warm compresses on the abdomen.

Wash out the stomach after giving tannic acid or some other astringent; strong coffee and whiskey; strychnin hypodermically; apply artificial respiration if necessary.

Tannin, stimulants, especially camphor.

When spasms occur, give narcotics; artificial respiration in case of paralysis of the organs of respiration.

Potassium permanganate is the antidote; in extreme depression of circulation and respiration, atropin sulfate administered hypodermically.

Antidote is acetate of lead.

Dermatomycosis.

Digitalis. Equisetosis.

Ergotism.

Fagopyrism.
Forage Poisoning.
Formic Acid. (In stinging nettles).
Gelsemin.

Helleborus niger. Hydrocyanic Acid.

Lathyrism Locoism.

Lupinosis.

Maydism. Morphin.

Muscarin.

Nicotin.

Phallin.

Picrotoxin.

Pilocarpin.

Cleanliness; sanitary surroundings; disinfection; preparation of bichloride of mercury.

No antidote; treat symptomatically.

Administer cathartic; also nerve and heart stimulants to combat symptoms of depression; change of food.

Administer purgatives and stimulants, especially camphor; tannic acid is chemical antidote and will neutralize unabsorbed portions of poison; chloral is physiological antidotc; further treatment symptomatic.

Change of food.

Change of food.

Cooling compresses externally; chalk, soda, or magnesia internally.

Wash out the stomach thoroughly; give stimulants and hot applications to the epigastrium and extremities; digitalis to strengthen the heart action and atropin to increase respiration.

Use stimulants, especially camphor.

Wash stomach with 0.5 per cent potassium permanganate solution or with hydrogen peroxid; the latter may be given hypodermically in small doses but with great caution; artificial respiration.

Treat paralyzed parts electrically; massage. In advanced stage of disease, treatment of no avail; in early stage, removal to an uninfected pasture with plenty of good nourishing food besides may benefit.

Chiefly preventive; no specific antidote; to prevent further absorption of poison, administer an acid; also give purgative.

Change of diet; transfer to hospital.

Wash out stomach with siphon tube using water containing potassium permanganate in the proportion of 20 gr. of the permanganate to 1 tumbler full of water; or use in the same manner an infusion of tea or tannic acid; emetics like mustard, using one or two teaspoons of each; apomorphin 5-10 minims of a 2 percent solution; hypodermic doses of strychnin 1-20 gr. or sulphate of atropin 1-60 gr.

Atropin used hypodermically; stimulants like strychnin may be given; wash out stomach.

If free vomiting has not occurred, wash out stomach with warm water or tea; give stimulants like whiskey or use hypodermic injection of strychnin nitrate 1-25 gr.

No known antidote; undigested material should be removed from stomach and same remedics as those suggested in muscarin poisoning may be administered.

Chloroform; chloral hydrate; artificial respiration.

Evacuate the stomach and wash out with so-

Physostigmin.

Ranunculus. Santonin.

Strychnin.

Toxins.

Veratrin.

lution of tannin; atropin is antagonistic, give 1-60 gr. hypodermically; use whiskey and ammonia as stimulants.

Scopalamin hypodermically; artificial respiration.

According to symptoms; tannin.

Chloroform; chloral hydrate; artificial respiration.

Control spasms by inhalation of chloroform; use stomach tube with warm water containing potassium permanganate, 4 gr. in 11 fl. oz. water.

Botulism produced in sausage poisoning may be treated by removing the poison, by giving free potations of warm water and salt, irrigation of intestines with enemas; vomiting and purging can be relieved by hypodermic injections of morphin; inject normal salt solutions.

Wash out stomach thoroughly with the siphon tube, or give emetics; give tannic acid or vegetable astringents which will precipitate the alkaloids; atropin or strychnin will counteract the cardiac depression.¹

¹ In the above table use has been made of the English translation of Kobert's Practical Toxicology by Friedburg, as well as the original.

CHAPTER XII

THE PRODUCTION OF POISON IN PLANTS

THE USE AND ACTION OF POISONS ON PLANTS

An extraordinarily large number of poisonous substances belonging to the alkaloids, glucosides, saponins, and toxins, occur in the vegetable kingdom. In addition there is a larger number of substances not strongly poisonous which are curative in their nature. These substances so widespread in the vegetable kingdom are the products of metabolism and probably in some cases, waste products, although according to Weevers, may act as reserve food substances. Treub 1 states that the hydrocyanic acid in the Pangium edule is of importance in the metabolism of the plant. It occurs not only in certain parts of the fibrovascular bundle but in the cells from the leaf and certain specialized cells of the epidermis, occurring both in a free and in an unstable combination. His conclusion is that the hydrocyanic acid is the first recognizable product of nitrogen assimilation. It is certainly true that in some cases the poisonous products formed in the plant do not undergo any further change. Undoubtedly the substances serve as a protection to the plant and it may be interesting to note that large quantities of these substances may be excreted and occur in an insoluble form in the cell sap and do no injury to the plant. Digitalin, morphin, atropin, eserin, muscarin, and veratrin, seem to exert little or no poisonous action on most plants; while strychnin may act as a strong poison.2 There are also other alkaloids that when applied to the plant are poisonous to the plants from which they have been obtained. Morphin is said to poison the poppy, and the motile spores are speedily killed by the same substance according to Strasburger. Too little, however, is known about this subject to make any extended remarks. It is interesting, however, to observe in this connection that as in the case of man and other animals, plants can be gradually accustomed to doses which would probably prove fatal in many cases. The Blue Mould (Penicillium glaucum) and some species of Aspergillus can accommodate themselves to strong solutions of copper and formalin. There are some reasons for believing that the protoplasm of different plants is not a uniform substance but varies, and that one substance may be toxic to the plant while harmless to another and even act as a stimulant.

DISTRIBUTION OF POISONOUS SUBSTANCES IN PLANTS

The seed may contain a toxic substance and upon the germination the poisonous material may occur not only in its juvenile stage but at maturity. In some cases the seed and the juvenile form may be non-poisonous, but as the plant becomes older the poisonous substance is elaborated as in the *latex* of some plants that contain narcotic principles. In some cases the seed is poisonous and the young plants apparently do not contain a toxic material; the

¹ Ann. du Jardin Bot. de Buitenzorg 13. Pharm. Review. 14:278.
2 Pfeffer, Physiology of Plants. English Translation. 2:260; Schwarz, Wirkungen von Alkaloiden auf Pflanzen, Erlanger Dissertation. 1897.

poisonous substance apparently being broken up to serve as a nourishing material for the growth of the plants.

Cornevin in his work upon poisonous plants says: In one group "the poisonous substance does not exist in the plants themselves, but, in some parts or tissue, elements are present which are not really poisonous in themselves but become so when the parts or tissues come in contact with one another. An example is seen in the glucoside amygdalin which in contact with emulsin forms hydrocyanic acid."

The activity of vegetable poisons may depend upon the age of the plant producing them. No definite rule can be established in regard to the age at which a plant produces its poisonous substance. Sometimes a younger plant is more actively poisonous than when older, sometimes, also, the poison is stored in certain tissues.

Poisonous principles are found in various parts of the plant, such as the root, stem, flowers, fruit, leaves, bark, tubers, seed and bulbs.

In many aerial parts of plants the poisonous substances are more ephemeral than they are in organs of the plant that serve as store houses of food.

Cornevin says:

It sometimes happens that the subterranean part only is poisonous as in Atractylis gummifera. This is true, also, of the common European Violet. On the other hand, when subterranean organs of certain plants come in contact with the light a poisonous substance may be produced. This is true of the tuber of potato which when green is poisonous. Poisonous substances are elaborated both in evergreen and deciduous leaves; no rule of comparison has, however, been established.

Certain variations of the plant are dependent upon its environment, certain conditions of which play an important part especially in the elaboration of poisonous substances.

These conditions are light, heat, season, climate, soil, culture and fertility.

LIGHT. It is a well knewn fact that light seriously interferes with the growth of parasitic fungi and bacteria so that pathogenic species may become quite harmless when placed in direct sunlight. The poison atractylin is formed only in darkness, on the other hand solanin is formed only in the light, as in the case of the green potato.

HEAT. Cornevin says:

The action of heat upon plants should be considered with that of light, of the seasonal variation, and of climate, and not as an isolated cause. When it is prolonged it induces dessication of the plant and as a consequence evaporation follows and destruction of the poison when it is volatile. This result is produced in some Ranunculaceae, Chenopodiaceae, etc.

'Moist heat, that is boiling, conduces to the same result in some poisons. Three-seeded mercury (*Mercurialis annua*) becomes inoffensive when it has been submitted to the action of heat, because of the volatilization of its toxic substance mercurialin.

SEASONS. Seasons cause considerable variation in the poisonous material produced in plants, the amount of poison contained often varying with the advance of the season. In aconite the poison, at first contained in the leafy organs becomes concentrated little by little in the seed.

The time of the year may have an important bearing upon the amount of poisonous material found in the plant. According to Prof. Hedrick the Cowbane or Musquash root (Cicuta vagans) of the west is much more toxic in the fall, winter, and spring, than in the summer, and this is partially confirmed by an experiment conducted with our common Cowbane (Cicuta maculata). It is also well known that the mature bulbs of Colchicum contains a much larger amount of the toxic substances than the growing bulbs and that for medicinal

¹ Des plantes vénéneuses et des empoisonnements qu'elles deterrminent, 524. Paris, 1893.

purposes the bulbs are usually collected between the decay of the foliage and the production of the flowers.

Opium is obtained from the capsules of the opium plant a few days after the petals have fallen, the seed containing comparatively little of the narcotic substances. It is well-known also that the poisonous principles of the Larkspurs are much more active in the spring than in the summer. Dr. Albert C. Crawford 1 says referring to the *Delphinium camporum*, with which he conducted some experiments and was able to kill several guinea pigs with toxic material obtained from the plant collected on April 26th and May 16th, but failed to get positive results of material collected in June.

There is no question as to the fact that Delphinium when injected subcutaneously will kill, and these experiments also establish the fact that the plant loses much of its toxicity as it approaches the flowering stage. It has been noted that Delphinium consolida² is also less active when mature.

Just after flowering, the purple larkspur turns yellow and ceases to be attractive, so that there is less danger of poisoning, although Chesnut and Wilcox report death in cattle from eating Delphinium glaucum in September. The great danger early in the season seems to arise from the fact that the Delphinium appears early in the spring, and the ground may again be covered with snow, so that it is the only green plant in sight, and therefore when in an especially poisonous stage it is eaten by cattle.

Botanical and other writers have frequently called attention to the fact that the greatest amount of poisoning in the west occurs in early spring. Of course, this may be because there is less green food and live stock may consume more of this plant than at other seasons. However, there seems scarcely any reason to doubt that the plant does contain a larger amount of the aerid toxic substances in the spring than in the summer, as proved by the experiments of Dr. Crawford.

The same author who investigated the Mountain Laurel ³ calls attention to the well-known fact that most of the cases of poisoning from Mountain Laurel occur in the winter. Undoubtedly the animals will eat more of the tough and leathery leaves in the winter because there is very little green for them; but may they not also have a larger amount of the toxic material? The plant is evidently also poisonous in the summer, as indicated by numerons reports of the experiments by Dr. Crawford, who conducted an experiment with material collected in the summer, in May and June, death occuring in a sheep weighing 49 lbs⁶, that had received 90 grams of powdered dried laurel leaves.

The late lamented Dr. Greshoff has called attention to the peculiar distribution of hydrocyanic acid in plants. In referring to a species of Hydrangea of the Saxifragaceae he states that he sometimes found considerable quantities of hydrocyanic acid in some of the well-known ornamental plants like the II. hortensia and sometimes he did not find it. He surmises that the cyanogentic principle disappears from the leaves in the autumn and that the young leaves have much more of the HCN than the older ones. In the case of the Plane Tree (Platanus) he found considerable of the same acid in the young leaves but as the leaves grow older the HCN content falls off to small traces. He states further that in the ordinary plane tree of the London Streets there is so much IICN that the amount from each London Plane Tree leaf would be enough to kill a London sparrow.

¹ Bull. Bur. Plant Ind., U. S. Dept. of Agric. 111: Part II; 7. 2 Dammann, C. Gesundheitspflege. 1886:1072, 3 Bull. U. S. Dept. of Ag., Bur. Plant Industry. 121:21.

Miss Alice Henkel in a paper on American Root Drugs 1 notes the importance of collecting medicinal plants and drugs at the right time. The roots of the American Hellebore (Veratrum viride) should be collected in the autumn after the leaves are dead. Generally speaking the drugs contain more of the medicinal virtues after the period of cessation of growth of the plant, Dr. C. Mueller 2 calls attention to the fact that Colchicum is much better if the rhizome is collected at time of flowering. On this point all authorities do not agree. Corncvin who has made a study from month to month of the migration of poison in plants such as the Laburnum found the following conditions: On May 20th, 2 grammes of the dried leaves of Cytisus Laburnum, administered to some carnivorous animal, were sufficient to induce vomiting. On June 10 (at which time pods were forming) 4 grammes were required to produce the same rcsult. July 28 (when the pods were fully formed) 12 grammes were required. September 28 (pods were beginning to dry) 20 grammes were required.

The experiments show that at all periods the leaves were poisonous but as the poison became concentrated in the pod the leaves became less toxic.

Similar experiments resulted in demonstrating that while the poison concentrates in the pod it also loses toxicity as the season advances, enormous doses taken from dried seed in October failing to produce death, while a 2gramme dose from a June pod proved fatal.

CLIMATIC. Latitude has an influence upon the formation of poisons. There are more poisonous plants in tropical regions than in colder regions. Certain plants which are poisonous in temperate regions lose their poisonous properties when taken into colder regions. Examples of this are aconite and cherry laurel.

It is difficult to make a comparison between the total number of poisonous plants in tropical and temperate regions, but it is probable that the warmer regions will show the greater number of poisonous species.

It might be well, however, to note in this connection that the poisonous properties of Rhododendron Chrysanthemum and R. catawbiense as well as of Aconitum are developed in colder regions while such toxic plants as the calabar bean (Physostiama venenosum). Strychnos nux-vomica, the Upas tree (Antiaris toxicaria) and numerous others are inhabitants of warm climates.

It is well known that the sorghum grown in dry climates produces a greater amount of a glucoside which is capable of being converted into hydrocyanic acid than in more moist regions. Some writers even assert that in dry seasons the drought depauperates the plant and that in the nodes there are considerable quantities of potassium nitrate.3

Again Dr. Ludwig Bernegau notes that the amount of alkaloid in Cola Nut varies considerably, it depending upon the source of the nut.

With reference to the presence of alkaloids in different kinds of opium, it is known that the Smyrna opium is of superior quality for medicinal purposcs. It is said to be superior to the opium obtained from India, which is in part attributed to climatic conditions, and probably in part to the method of collection. According to Blyth, the amount of morphin varies as follows:

Crude morphin (containing about 7-10 of pure morphin). Highest, 12.30; lowest, 6.76; mean, 9.92 per cent, which equals 12.3 per cent of the dried drug.

¹ Bull. U. S. Dept. Agrl., Bur. Plant Industry, 107. Farmers Bull. 188.
2 Pharm. Review. 14:113. Die Natur. 44:550.
3 An English writer, Pease, attributed the death of animals to this substance rather than to the hydrocyanic acid. Pharm. Review, 15:208.

The nicotin of tobacco depends largely on climatic and soil conditions, The Havana, Porto Rico, Sumatra, Connecticut Seed Leaf and Wisconsin have qualities all peculiarly their own.

Blyth records the following percentages of nicotin in various tobaccos as given by Cox.1

| Variety examined | | | Nicot | in pe | er cent |
|---------------------------------|--|--|-------|-------|---------|
| 1. Syrian leaves (a), . | | | | | .612 |
| 2. Syrian leaves (b), . | | | | | 1.093 |
| 3. Gold Flake (Virginia), | | | | | 2.501 |
| 10. "Navy-Cut" (Light colored), | | | | | 3.640 |
| 15. "Best Shag" (b), . | | | | | 5.000 |
| 17. Algerian tobacco (a), | | | | | 8.813 |

According to Professor Garner 2 the nicotin contents vary as follows:

Nicotin soluble in petroleum ether in domestic filler tobacco 2.20 percent; Imported Cuban Santa Clara tobacco 1.33 percent.

That climate plays an important part in connection with chemical products is illustrated in the essential oils. The most important of these such as bergamot, cassia, cinnamon, jasmine, fennel, lavender, orange, rosmary, attar of roses, and many of the others are imported into the United States. These are produced in localities that are more favorable for their production than in many parts of this country. However, some of the essential oils like those from peppermint, sweet birch, and sassafras are produced extensively in the United States. The time of collecting and many other factors as well as the matter of labor enter into the production of these oils. The variation in composition has been indicated by Dr. Edward Kremers and his students in various publications.3

It is well known that the locality for attar of roses which supplies most of the commerce of the world is a small district of country on the southern side of the Balkan Mountains. The best localities according to those best informed on the subject are those occupying southern or southeastern slopes. The flowers attain perfection in April or May and are gathered before sunrise. It is concluded from some experiments carried on in some of the northern countries in Europe, that a cool northern climate is not conducive to the production of highly odorous oils.4

It is interesting to note that the oil of orange flowers comes chiefly from the southern part of France, and that the flowers of sweet orange afford about one-half the amount of oil that those of the bitter orange do. Lavendar oil, made from Lavandula vera, is very variable in quantity, depending upon its source, although not grown to any great extent in England. Flückiger and Hanbury state that the Mitcham oil grown in the Surrey, is of a superior and high quality. The above is used here to illustrate the fact that the chemical products of plants vary depending upon the different climatic conditions.

¹ Pharm, Journ., Jan. 20, 1894. Blyth, Poisons; Their Effects and Detection. Ed. 2.274.
2 Bull. U. S. Dept. Agr., Bur. Plant Ind. 141; Pt. I, 12.
3 Kremers and Schreiner. The Quantitative Estimation of Carvone in Volatile Oils.
Pharm. Review. 14:76.
Edward Kremers and Florence M. Gage. Notes on Two Oils Containing Pulegeone.
Pharm. Review. 16:412.
Kremers and Sievers. Oils from Milfoil. Pharm. Review. 25:215.
The Volatile Oils, by E. Gildemeister & Fr. Hoffman. English Translation, E. Kremers.
4 Flückiger & Hanbury, 262.

Dr. Rodney H. True in an article in the Cyclopedia of American Agriculture,1 says:

The sources of our crude drugs and condiments are very widely separated, depending in large part on climatic conditions. Common drug plants belonging to the temperate zone, such as digitalis, burdock and caraway, are in very large part produced in northern and central Europe, frequently in more or less localized regions. Caraway comes chiefly from Holland, in small quantities from Norway, east Prussia and southern Germany. Feuncl is cultivated in Saxony, Galicia, Macedonia and Italy. Digitalis leaves and belladonna reach the market of northern Germany, Austria, Belgium, Holland and England. Peppermint oil is produced chiefly in Japan and the United States. Other plants demanding tropical conditions are obtained from regions in which their culture has been undertaken. Cinchona bark, from which quinine is obtained, came formerly from the slopes of the Andes. Cultivation of this plant in India, Java, and other parts of the Orient has succeeded in so far as to cause the practical disappearance of the wild barks of South America from the market. Ipecacuanha, likewise a native of northern South America, is apparently repeating this history. Black and white pepper are chiefly produced in southeastern Asia, coming on the market through Singapore and Penang. Cloves are in large part supplied by Zanzibar, where the crop constitutes one of the royal monopolies. Some products are derived from still more localized regions, as buchu leaves from the vicinity of Cape Town, South Africa, and aloes from South Africa, the island of Socotra in the Red Sea, and the Barbadoes islands. Some are cultivated, as may be seen in numerous cases cited above, and some are wild products. Camphor until recently has been derived from an essentially wild tree growing in Japan, China and Formosa. The great depletion of the natural forests has led the Japanese government to make extensive plantings. Several African sorts of the red peppers of the market are collected by natives from the wild plants and brought long distances to market.

CULTURE. Cultivation often affects the amount of poison contained in a plant. A wild vetch of Europe (Laythrus) is extremely bitter, but through cultivation the poisonous material has been largely eliminated and the vetch has become a useful cultivated forage plant. It is stated also that the aconite (Aconitum Napellus) when cultivated loses some of its toxic properties and that this loss of toxic action occurs in a few generations. Prof. S. M. Tracy informs me that the cultivated forms of the Cassava are not injurious to stock but the wild form, as is well known, contains toxic materials. The wild forms of the lima bean (Phaseolus lunatus) contains much more HCN than the cultivated forms.

Dr. Rodney H. True in speaking of the physiological action of the betel nut states that its physiological action 2 depends on several factors. The green nuts produce temporary dizziness. The poisonous variety according to Indian authorities is one that is reverted to its former wild condition, while the common or ordinary betel nut which has been cultivated for hundreds of years is a mild narcotic stimulant bringing about a feeling of general comfort, good humor and exhilaration.

It is also well known that when certain toxic bacteria as Streptococcus pyogenes are cultivated they lose some of their pathogenic properties.

We must not, however, conclude that because a plant is cultivated it loses its poisonous properties, because there are certain cultivated ornamental plants that are as poisonous in their cultivaed form as when grown wild. For example the tobacco which has been cultivated for several hundred years contains as much nicotin in its cultivated as in its primitive form.

SOIL. It is believed also that the soil plays an important part in connection with the amount of poison produced in the plant. There can be no question that the soil plays an influence upon the quality of the fruit as well as upon

¹ Cyclopedia of American Agriculture. 2:458. 2 Pharm. Review, 14:130.

the quality of the grain. Why should it not also influence the poisonous constituents of the plant? Cornevin mentions that soil affects the color of Digitalis, it being much paler when grown in calcareous than when grown in granitic soil.

RELATED SPECIES AND TOXIC SUBSTANCES. In some cases the same toxic substance is widely distributed in different families of plants. On the other hand closely related plants frequently have entirely different amounts of toxic substances. The bitter and sweet almond illustrate this in a very marked degree, the bitter almond having considerable amounts of the glucoside amygdalin, while the sweet almond is entirely harmless.

Frank Rabak 1 has made an investigation of the amount of the chemical substances found in the kernels of the fresh apricot and plum, in which it appears that the amount of hydrocyanic acid by per cent contained in these plants is as follows:-

| | | | | | Per | |
|------------|------|--|--|--|-----|------|
| Peach | | | | | | 2.20 |
| Apricot | | | | | | 2.40 |
| Apricot | | | | | | 2.05 |
| Prune | | | | | | 1.75 |
| Bitter Aln | nond | | | | | 4.80 |

According to Dohme and Engelhardt 2 the thin green or young bark of Prunus serotina is richer in hydrocyanic acid, than the dark brown or older bark.

It may be interesting to note that certain species of Cacti are used for stock food,3 and others contain powerful drugs. For instance, the Mescal Bean, which is derived from several species of Anhalonium which contain peliotin, is a narcotic of considerable potency.4

According to Dr. Peinemann 5 the alkaloidal-content of Datura varies as follows; the seeds, 0.541 percent.; root, 0.315; leaves, 0.41 percent. of atropin, The Datura alba is richer in alkaloids than the native species, D. Stramonium.

Dunstan and Henry are authorities for the statement that two forms of Mandrake, the American (Podophyllum peltatum) and the Indian (P. emodi), produce the same principle podophyllotoxin C₁₅II₁₄O₆ a strong purgative and also the so-called podophyllin which consists of a resin. The podophyllotoxin occurs in the Indian plant from 9-12% and in the American plant from 4-5%.

DISTRIBUTION OF CHEMICAL SUBSTANCES. The same chemical substance is frequently found in plants that are not closely related. For instance, Edward Kremers and many other pharmaceutical chemists have found the essential oils in widely separated families, and Greshoff states that a leaf of Five-finger (Potentilla davurica of China) which is closely related to our shrubby Cinquefoil (P. fruticosa) produces a powerful odor of roses. Geranoil occurs, for instance, in the young foliage of willow and many other plants.

The substance berberin is found in a large number of plants like the Barberry, the Mandrake, Twin Leaf (Jeffersonia diphylla), Meadow Rue (Thalictrum flavum), Toddalea aculeata, Hydrastis canadensis, Argemone mexicana,

¹ Bull. U. S. Dept. Agr., Bur. Plant Industry. 133. 2 Pharm. Review. 14:13. 3 Griffiths and R. F. Hare, The Tuna as Food for Man. Bull. U. S. Dept. Agrl. Bur. Plant Ind. 116. 4 Pharm. Review. 14:153.

⁵ Pharm. Review. 14:233.6 Proc. Chem. Soc. 189:42-44.

Greshoff 1 has indicated that a species of Thymus, the *Thymus Serpyllus* can be readily distinguished from the variety *album* phytochemically; the common form of this species (*Thymus Serpyllus*) produces thymol in its leaves, whereas the other has the odor of lemons.

SAPONINS. We may note here the very wide distribution of saponins in different plants. Greshoff in several papers 2 gives a long list of the orders in which saponin occurs, 1 many of the others are mentioned in the list of species at the end of this manual.

The substance saponin occurs in many different plants of which the following is a partial list:

Amaryllidaceae: Agave, Fourcroya. Araceae: Arum. Araliceae: Aralia. Begoniaceae: Begonia. Berberidaceae: Berberis, Caulophyllum, Leontice. Bromeliceae: Bromelia. Caryophyllaceae: Acanthophyllum, Arenaria, Dianthus, Gypsophila, Lychnis, Melandryum, Polycarpaea, Saponaria, Silene. Chenopodiaceae: Chenopodium. Commelinaceae: Tradescantia. Compositae: Arnica, Grindelia, Mutisia. Convolvulaceae: Ipomoea. Cucurbitaceae: Echinocystis, Luffa, Trichosanthes. Dioscoreaceae: Dioscorea. Euphorbiaceae: Phyllanthus? Ficoideae: Trianthema. Filices: Polypodium. Gramineae: Panicum, Lolium. Hippocastanaceae: Aesculus. Illecebraceae: Herniaria. Leguminosae: Acacia, Albizzia, Cacsalpinia, Entada, Enterolobium, Gleditschia, Gymnocladus, Milletia, Mimosa, Pithecolobium, Prosopis, Calliandra, Tetrapleura, Xylia e. a. Liliaceae: Chamaelirium, Chlorogalum, Helonias, Medeola, Muscari Paris, Polygonatum, Smilax, Trillium, Yucca, e. a. Loganiaceae: Buddleia. Magnoliaceae: Illicium. Meliaceae: Trichilia, Walsura. Melianthaceae: Bersama. Menispermaceae: Stephania? Cocculus? Coscinium? Myrtaceae: Barringtonia, Careya. Oleaceae: Chionanthus, Syringa. Passifloraceae: Carica? Modecca. Phytolaccaceae: Pircunia. Pittosporaceae: Pittosporum. Polemoniaceae: Cantua. Polygalaceae: Monnina, Polygala. Primulaceae: Anagallis, Androsace, Cyclamen, Primula, Soldanella, Trientalis, e. a. Ranunculaceae: Ficaria, Nigella. Rhamnaceae: Gouania, Zizyphus. Rosaceae: Quillaja, Spiraea. Rubiaceae: Basanacantha, Cephalanthus, Mitchella, Mussaenda, Randia. Rutaceae: Zanthoxylum. Sapindaceae: Blighia, Dodonaea, Ganophyllum, Koelreuteria, Magonia, Nephelium, Paullinia, Pancovia, Pometia, Sapindus Serjania. More than 100 species of this family contain saponin. Sapotaceae: Achras, Bassia, Lucuma, Omphalocarpum. Saxifragaceae: Hydrangea. Scrophulariaceae: Digitalis, Leptandra, Limosella, Simarubaceae: Balanites, Solanaceae: Acnistus, Lycopersicum, Scopolia, Solanum. Ternstroemiaceae: Camellia, Caryocar. Tiliaceae Grewia. Urticaceae: Ficus. Zygophyllaceae: Guapacum, Tribulus.

HYDROCYANIC ACID. A list of the plants containing a glucoside capable of being converted into HCN has been given in another connection, but we may note in this connection plants belonging to many different orders like the Compositae, Euphorbiaceae or Spurge Family, Gramineae or Grass Family, Raunnculaceae or Buttercup Family, Fern, Filices, Droseraceae or Sundew Family, Saxifragaceae or Saxifrage Family. A common Rocky Mountain shrub, the Jamesia, of this family, is said to contain considerable quantities of HCN, and the Mountain Mahogany (Cercocarpus parvifolius) of the Rose Family (Rosa-

¹ Bull, Misc. Information, Royal Bot, Gardens Kew. 1909:415.

2 Kew Bot. Garden 1909:397. Phytochemical Information at Kew. Mededeelingen uit 'S Lands Plantentuin. 29:1909.

ceae), contains HCN, and Greshoff says must henceforth be counted among the poisonous plants. Of the Grass Family (Gramineae) quite a number of species such as the Stipa lessingiana, are now known to contain HCN. Some of them like our Stipa robusta, have been known to be toxic for a long time, but such genera as the Quaking Grass, Briza (Catabrosa), Wild rye (Elymus), Manna Grass (Glyceria), Salt Grass Holcus lanatus all contain HCN.

CUMARIN: The substance cumarin also occurs in many different plants which we may list as follows:

Talauma (Magnoliaceae); Phoenix (Palmae); Dipteryx, Toluifera, Melilotus (Leguminosae); Prunus (Rosaceae); Ceratopetalum (Saxifragaceae); Asperula, Basauacantha, Borreria, Diodia, Galium, Mitracarpum, Spermacoce (Rubiaceae); Ageratum, Chrysanthemum, Eupatorium, Humea, Trilisa, Liatris (Compositae); Alyxia (Apocynaceae); Hemidesmus (Asclepiadaceae); Rhinacanthus (Acanthaceae), Aceras, Augraecum, Orchis (Orchidaceae); Hierochloe, Anthoxanthum (Gramineae); Adiautum, Cheilanthes, Polypodium (Filices); Lactarius, Russula (Fungi).

CYTISIN. Greshoff reports cytisin as occuring in the following plants of the Pulse family: Cytisus Adami; C. alpinus; C. Alschingeri; C. biflorus; C. hirsutus; C. Laburnum, seed 1.8 per cent; C. nigricans; C. polytrichus; C. proliferus; C. Weldeni; C. Attleanus; C. candicans; C. formosissimus; C. monspessulanus; C. Ruthcnicus; C. scoparius; Ulex europaeus, seed 1 per cent; U. Jussiae; Genista ephedroides; G. monosperma, seed 1.9 per cent; G. florida; G. germanica; G. spicata; G. tinctoria; G. ramosissima; Sophora secundiflora, seed 3.5 per cent; S. tomentosa; S. flavescens; S. sericea; Baptisia perfoliata; B. tinctoria; B. alba; B. australis; B. leucantha; B. versicolor; Euchresta Horsfieldii, Anagyris foetida, Rauwerda reported it in 28 species of Lotus, L. suaveolens; Thermopsis Caroliniana; and Th. montana.

In some instances apparently the same alkaloidal substance was first reported in some plants, but later investigations have shown that these substances are not identical. For instance Schlotterbeck who investigated the alkaloids found in the poppy family, especially Argemonia Mexicana, reported that this plant does not contain morphin but protopin and berberin.

CHAPTER XIII

ALGAE IN WATER SUPPLIES

Algae have long been known to be injurious to water supplies and numerous papers in the United States and Europe have discussed some of the problems arising therefrom. Algae are generally small, microscopic plants found in water and belong to the great class known as Thallophyta. These frequently cause the pollution of water supplies and are therefore of first importance in this connection. Some algae contain a green coloring matter called chlorophyll and hence they are able to make their own food out of water and carbon dioxid. Many of the algae, however, are not green, some being blue, others red, and still others brown. The Sea Mosses are algae also, and occasionally attain great size and length. Some algae are supposed to be injurious to stock, Mr. George Francis in Australia having attributed a disease of horses, sheep, dogs, and pigs to some blue-green algae. The organism causing the trouble in this case was referred to Nodularia spumigera, which floats on the water and, being wafted to the lea shores, forms scums from two to six inches thick. He says that, when animals drink the water, it acts as a rapid poison and causes death. It first produces stupor and unconscioueness, the animal falling and remaining quiet, as if asleep, unless touched, when convulsions come on, with head and neck drawn back by rigid spasms which subside before death. This, in sheep, takes place in from 1-6, or 8 hours; in horses, in 8-24 hours; in dogs, in 4-5 hours; and in pigs, in 3-4 hours. A post mortem was made on a sheep that had 30 ounces of fresh scum administered by mouth, death occurring in 15 hours. The post mortem, 6 hours later, showed that the stomach had none of the green scum left, all having been absorbed; the abdominal cavity contained 2 pints of yellowish material; the heart was flaccid with effusion of serum around it; lungs, liver, kidneys, and brain normal; dura mater enlarged; blood of ventricles and arteries black. It was thought that poisoning was due to the decomposition of the algae which smelled like putrid urine, previous to this it had an odor like butyric acid. Account was not taken of the products of decomposition by bacteria. The trouble may have been caused by the poisonous products of bacteria rather than these algae. This seems not have occurred to Dr. Francis.

Many of the algae, especially the following, Anabaena, Clathrocystis and bacteria-like Beggiatoa, produce what is known as the working of the lakes and pig pen odors. Dr. Farlow has described these from Massachusetts and Dr. Trelease has described them for the lakes about Madison, Wisconsin. The greenish-yellow scum occurs on the surface of the lakes, especially noticeable during hot weather of the summer. Dr. Trelease says:

When a lot of it is present, it appears as fine granules suspended in the water, scarcely visible to the naked eye except as they reflect the light, when they call to mind the dancing motes in a beam of sunlight. Under the influence of a gentle, but continuous breeze, these particles are collected into fleshy masses and are driven ashore, so that they accumulate along the margins of the lake, forming a slimy scum, which quickly putrefies, giving off a very disagreeable odor. During this change, its color changes to a decided

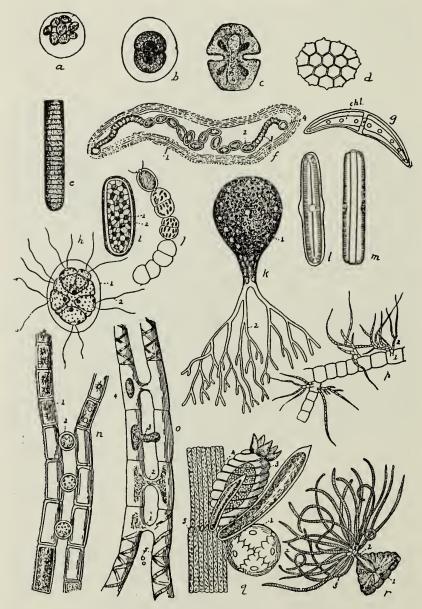


Fig. 19. Algae found in water supplies. a. Protococcus. b, c. Casmarium, one of the clesmids. d. Pediastrum. e. Oscillatoria. f. Nostoc paludosum. g. Closterium, chr—chromatophore. h. Pandorina; 1, gelatinous envelope; 2, cilia. i, j. (Anabaena flas-aquae). k. Botrydium granulatum; 2, rhizoids. I and m. Diatoms. n. Zygnema; 1, chromatophore; 2, zygospores. o. Spirogyra; 1, 2, 3, 4, different stages in the formation of the zygospores. p. Draparnaldia; 2, thallus. q. Stoneworts (Chara); 1, antheridium; 3, archegonium; 5, nodal cells. r. Portion of antheridium with sperm cells.

blue green, which stains the pebbles, sticks, ctc., over which it is smeared. The appearance of this scum is sometimes spoken of as the working of the lakes, from the resemblance to the collection of scum on cider, etc., when fermenting, or, as an idiom expresses it, "working." The odor given off by this putrefaction can be detected from some distance.

In speaking of the water bloom, Prof. Dwight C. Marsh says:

This phenomenon is especially marked in Lake Winnebago in some summers. It is due, of course, to the enormous growth of the plants of the plankton, that growth being particularly fostered by the hot weather of midsummer. The plants especially concerned in forming the bloom are Clathrocystis, Anaboeno, Aphanizomenonfi Oscillorio, Lyngbya, and Gloiotrichia. .The times of occurrence of these plants have already been noticed in the discussion of the individual constituents of the plankton. At the middle of August, in some summers, on a still day, the surface of Lake Winnebago is apparently a solid, opaque green. Some of this material decomposes, and as the currents slowly move along the surface material, it shows a wavy streaked appearance like the surface of polished malachite. Its intrinsic beauty, however, does not attract the average person, for he looks upon it as "scum," and he thinks of it simply as an evidence of filth. This material is thrown upon the shores by the waves until the rocks alongshore are completely eovered with it, and it may in its decay become very offensive. Following the maximum period of the "bloom," Cladophora appears and covers the littoral rocks with a thick mat of green. This great growth of "bloom" naturally attracts the attention of the non-scientific observer, and many absurd explanations of its appearance are given. The most common one in Oshkosh is that it is a mass of seeds coming from the marshy shore of the Fox and Wolf above Ohkoslı. Doubtless the Anabacno and Gloiotrichia have given rise to the supposition that the bloom is a mass of seeds. The decomposition of Gloiotrichia produces a blood red coloring matter which is sometimes very noticeable on the shores of Lake Winnebago, and has led people to question as to whether the lake is not affected by one of the plagues of ligypt.

When the water is still the plants of the bloom are in greatest abundance, close to the surface, and are distributed very uniformly over the lake. Frequently, in the latter part of July and in August, there may be seen floating about yellowish green masses of a more or less spherical outline, perhaps as much as three inches or more in diameter. These masses, which are composed of aggregations of Aphanizomenon mingled with scattered fronds of Gloistrichio and Anaboena, have very little coherence and elude the collector by falling in pieces almost at a touch. As is evident from the discussion of the occurrence of the algal constituents of the plankton, the bloom is not a prominent feature of the deep lakes,—in fact, in some years the growth of these algae is hardly noticed by the ordinary observer,—and of the shallow lakes few seem to produce as large an amount as is seen in Lake Winnebago. Of the lakes under observation, Shawano and Pelican were the only ones that could be compared at all with Lake Winnebago.

Odors produced by the decomposition of organic matter in water are easily detected. Sometimes they are grassy and sometimes mouldy or musty. Sometimes the odor is decidedly fishy, which is especially true of some of the higher plants like Myriophyllum or Potamogeton. Mr. Whipple* says that the cucumber taste found in the Boston water supplies was due to a fresh water sponge. He made a series of experiments with oil of peppermint and other oils and he found that the oil of peppermint could be recognized when the dilution was 1 to 50,000,000; oil of cloves, 1 to 8,000,000; cod liver oil, 1 in 8,000,000. Kerosene oil could not be detected when diluted to 1 in 8,000. According to Whipple, aromatic odors are produced by certain diatoms; grassy odors are produced by certain blue-green algae like Anabaena and Rivularia; the sweet grassy odor is produced by Clathrocystis; Volvox produces a fishy odor, while Pandorina and Eudorina produce a faintly fishy odor. Certain protozoa produce fishy and oily odors.

Perhaps the most extensive investigation of microscopical organisms in water are those reported by the Massachusetts State Board of Health. Special methods for determining the amount of impurities found in water are given in these reports. Prof. Parker has shown that when the odors are pronounced the organisms are always present in considerable numbers. He says:

The majority of organisms which have been recorded are of microscopic size. At first sight it might appear that these organisms would be insignificent in proportion to their mass, and that more attention should have been devoted to the larger plants and animals. Very few cases have been observed, however, in which really serious trouble in water supplies can be referred directly to the growth of large plants or animals; whereas, in many instances, waters with very objectionable qualities contain nothing to which these qualities can be attributed except microscopic organisms. Further, although the organisms here dealt with are of such small dimensions, the immense numbers in which they occur more than counterbalance their small size; and I am therefore, of the opinion that, of those organisms which cause objectionable qualities in water, the microscopic ones are the more important.

HOW GROWTH OF ALGAE IN WATER MAY BE PREVENTED

The growth of algae may be prevented by covering the reservoir; since light is essential for their growth, they will not develop in a closed reservoir, and we hear of little complaint under such conditions. Large reservoirs for public water supplies are not generally covered. Often reservoirs are much troubled by algal contamination; in some cases it becomes necessary to remove these by a laborious method. The removal of organic matter by keeping the source of the water supply in as pure condition as possible will no doubt do something toward keeping algae out, but nearly all water contains sufficient organic matter for the growth of algae, especially water coming from water sheds.

THE USE OF ALGICIDES

Within recent years much work has been done in preventing the growth of algae and bacteria by copper sulphate solutions. Moore and Kellerman in a recent paper arrive at the following conclusions:

The disagreeable odors and tastes so often present in drinking water are due almost exclusively to algae, although the economic importance of studying these plants has not been recognized until recent years. These algal forms are widely distributed, and reservoirs in many states have been rendered unfit for use by their presence. It has been found that copper sulphate in a dilution so great as to be colorless, tasteless, and harmless to man is sufficiently toxic to the algae to detroy or prevent their appearance. At ordinary temperatures one part of copper sulphate to 100,000 parts of water destroys typhoid and cholera germs in from three to four hours. The ease with which the sulphate can then be eliminated from the water seems to offer a practical method of sterilizing large bodies of water when this becomes necessary. The cost of material for exterminating algae will not exceed fifty to sixty cents per million gallons and will usually be less. The destruction of pathogenic bacteria requires an expenditure of from \$5.00 to \$8.00 per million gallons, not including the cost of labor.

It has been found that Spirogyra will die in water containing one part of copper to one billion parts of water. Some of the algae like Anabaena are destroyed in dilutions from one to five million, although Moore and Kellerman found that one species of Spirogyra requires a greater strength of solution. There is a wide-spread opinion that metallic copper and copper salts are injurious, which is certainly true when the same are taken in larger quantities. According to Tschirsch, .05 to .2 of copper sulphate causes vomiting and diarrhoea. In the paper cited, Moore and Kellerman write as follows:

It is evident that there is still a considerable difference of opinion among eminent authorities as to the exact amount of copper which may be injurious, but as a very conservative limit we may accept 0.02 gram as the amount that may with safety be absorbed daily. According to Merck's Index, the National Dispensatory, and the United States Dispensatory, the dose of copper sulphate for tonic and astringent purposes is one-fourth grain, or 0.016 gram; as an emetic, a dose of five grains, or 0.33 gram. Thus it is seen that even if the maximum concentration of copper sulphate necessary to destroy algae in reservoirs were maintained indefinitely, the absorption from daily use would be very far below an amount that could produce the least unpleasant effect. Taking a dilution of one

to one million, which in all cases would be sufficient to prevent the growth of a polluting algal form, it would be necessary to drink something over twenty quarts of water a day before an amount which is universally recognized as harmless would be introduced into the system, while more than fifty quarts would have to be consumed before there would be danger of producing an unpleasant or undesirable effect. As will be seen from the preceding tables the use of copper sulphate at this maximum strength of one to one million would need to be resorted to only in extreme cases, and for a very short length of time, for, the reservoir once entirely free from the organisms, a very much weaker solution would be sufficient should any further application be necessary.

The Report of the Massachusetts State Board of Health for 1905 stated as follows:

The objectionable tastes and odors of pond and reservoir waters, which are often attributed to decaying fish and other causes, are, in practically all cases which have been brought to the attention of the Board, caused by the presence of organisms, in some cases of kinds which attach themselves to the sides and hottom of the reservoir, but in the majority of cases of those kinds which live in suspension in the water. Early in 1903, Dr. George T. Moore formerly Algologist of the United States Department of Agriculture, brought to the attention of the Board the results of investigations which he had made, indicating that the microscopic organisms which are the chief cause of objectionable tastes and odors in the waters of ponds and reservoirs could be destroyed by applying sulphate of copper or blue vitrol to the water in very small quantities; and information was also submitted tending to show that bacteria were also destroyed in water brought in contact with metallic copper.

The results of some of the experiments and investigations so far as obtained tend to support the conclusions which had been reached when the matter was first brought to the attention of the Board as to the practicability of the copper treatment for the removal of growths of organisms and bacteria, but the results of other experiments conflict with some of these conclusions. Further study and experiment are necessary before the probable results of the use of copper in preventing objectionable conditions resulting from growths of organisms, or the probable effect of the use of this substance in public water supplies upon the public health can be conclusively determined.

Dr. Moore states that:

Spirogyra is often the cause of considerable trouble in a mechanical way, and on account of its method of forming resting sporcs is usually able to withstand the most unfavorable conditions to which it may be subjected in a pond or reservoir. In at least one instance this alga has been the cause of the loss of thousands of dollars by the damage it produced in smothering out the young water-cress plants in the artificial heds constructed for the winter propagation of this vegetable. When the cress is cut for market, it necessarily leaves the plants in a weakened condition, and if the Spirogyra gets a start it will form a thick, beavy mat over the water, which is sufficient to prevent the growth, if not entirely to kill, the cress plants.

CHAPTER XIV

A CATALOGUE OF THE MORE IMPORTANT POISONOUS PLANTS OF THE UNITED STATES AND CANADA

For the purpose of facilitating a study of the poisonous plants of this country a partial catalogue is given here. It is based in part on a catalogue of the poisonous plants of Iowa,* issued by the writer and Estelle D. Fogel (Buchanan). To further facilitate the study of the poisonous plants the following papers on the subject should be mentioned:

· Chesnut, 1 Chesnut & Wilcox, 2 Wilcox, 3 Nelson, 4 O'Gara, 5 Kennedy, 6 Peters, Slade & Avery,7 Glover,8 Crawford,9 Heald & Peters,10 White,11 Johnson,12 Bessey,13 Rusby,14 Guttenberg,15, Pammel,16 Millspaugh,17 Coville,18 Peck, 19 J. U. & C. G. Lloyd, 20 Stebler & Schroeter, 21 Schaffner, 22 R. Schimpfky, 23 Winslow,24 Farlow,25 Maiden,26 Atkinson,27 Greshoff,28 Wilson,29 nevin,30 Halsted,31 Kobert,32 Crawford,33 and Marsh.34

*Proceedings of the Iowa Academy of Science. 14:147-172. Contributions, Department of Botany, Iowa State College. 37:147-176.

1 Principal Poisonous Plants of the United States, Div. Bot. U. S. Dept. Agr. 20-60, f34.

Thirty Poisonous Plants of the United State, U. S. Dept. Agr. Farmers Bull. 86:32, f24. Rep. Bu. An. Ind. 15:387-420, f69.

- An. Ind. 15:387-420, f69.

 2 The Stock Poisoning plants in Montana, Bull. Div. Bot. U. S. Dept. Agr. 26:150, pl. 36.

 3 Rep. Bu. An. Ind. 17:91-121, p. 32.

 4 Feeding Wild Plants to Sheep, Rep. Bu. An. Ind. 15:421-425.

 5 Rep. Nebr. Agr. Expt. Sta. 16:14-84, f. 13.

 6 Bull. Nev. Agr. Expt. Sta. 51-57, pl. 26.

 7 Poisoning of Cattle by Common Sorghum and Kafir Corn, Neb. Agr. Expt. Sta. 77:16.

 8 Bull. Col. Agr. Expt. Sta. 113:24, pl. 8.

 9 The Poisonous Action of Johnson Grass, Bull. Bur. Pl. Ind. 90, part 4.

 10 Ergot and Ergotism, Press Bull. Nebr. Agr. Expt. Sta. 24, f. 3.

 11 Dermatitis Venenata, An Account of the Action of External Irritants upon the Skin, 1887.
- 11 Dermatitis Venenata, An Account of the Action of 216:1887.

 12 A Mannal of the Medical Botany of N. Amer. p. 292, pl. 9, f. 160.

 13 Bull. Dept. Bot. 1884. Proc. Soc. Prom. Agr. Sci. 23:35-41.

 14 Poisonous Plants of the Vicinity of New York City, 19.

 15 Poisonous Plants which Grow in and around Erie, 21:25.

 16 Bull. 1a. Agr. Expt. Sta. 70:423-448. Poisoning from Cowbane, Bull. Ia. Agr. Expt. Sta. 28:215-225, f. 5.

 17 Medicinal Plants, 1:99, pl. 99. 2:100-180, pl. 180.

 18 Poisonous Mushrooms, Cir. Div. Bot. 13:24.

 19 Edible and Poisonous Fungi of New York, Ann. Rep. State Bot. of State of N. Y. 1896: 105-248, pl. 43. Rep. State N. Y. Mus. Nat. Hist. 48.

 20 Drugs and Medicines of N. Amer., Ranunculaceae 1:304, pl. 24, f. 105.

 21 Beiträge zur Kenntniss der Matten und Weiden der Schweiz. Landw. Jahrb. d. Schweiz 5:141-225, pl. 20, f. 4.

22 Poisonous and other Injurious Plants of Ohio, Ohio Naturalist 4:16, 32, 69. 28 Deutschlands Wichtigste Giftgewächse in Wort und Bild nebst eine Abhandlung ueber

- Pflanzengifte, pl. 27.

 24 Veterinary Materia Medica and Therapeutics, 775.

 25 Some Edible and Poisonous Fungi. Bull. U. S. Dept. Agr. Div. of Veg. Path. and Physiology. 15:453-470.

 20 Plants Reported to be Poisonous to Stock in Australia. N. S. W. Deut. of Agr. Misc. 20 Plants Reported to be Poisonous to Stock in Australia. N. S. W. Dept. of Agr. Misc.
- 27 Mushrooms; Edible, Poisonous, etc. 322. Ithaca, N. Y. (2nd Ed.).
 28 Phytochemical Investigations at Kew. No. 10, 1909. Monog. de plantis venenatis et sapientibus, etc. Batavia, 1900.
 29 Oleander Poisoning of Live Stock.

Sapientibus, etc. Batavia, 1900.
20 Oleander Poisoning of Live Stock. Bull. Ariz. Agr. Expt. Sta. 59:381-397.
30 Des plantes vénéneuses et des empoisonnements qu'elles determinent. 524. Paris, 1893.
31 The Poisonous Plants of New Jersey. Bull. N. J. Agr. Expt. Sta. 135:28.
22 Lehrbuch der Intoxikationem. Stuttgart, 1902.
Practical Toxicology for Physicians and Students. Trans. and ed. by I., II. Friedhurg.
33 Laboratory work on loco-weed Investigation. Bull. U. S. Dept. Agr. Bur. Pl. Ind.
121:39-48.

34 The Loco-weed Disease. Far. Bull. U. S. Dept. Agr. 380:16.

Many of the plants in this list are certainly not strongly toxic but they frequently produce injurious symptoms in man and animals. The list is by no means complete; it will, however, serve the purpose of calling attention to some of the injurious plants in North America.

It may be noted in this connection that experiments recorded by no means agree as to the effect of certain poisonous plants upon animals. Notice the very discordant results obtained with reference to the poisoning from loco weeds. We may note also the opposite results obtained by experiments reported by Dr. S. V. Nelson 1 with reference to feeding Delphinium Menziezii to sheep. This writer concluded that when fed to sheep early in May it is not poisonous to them. In the same report 2 Dr. E. V. Wilcox reports a very different experience. This writer reports an arrested heart action and respiration and paralysis of the spinal cord. The animals were fed early in the season, about the same time of the year that the unsuccessful experiments were reported by Dr. Nelson. Either these writers were dealing with different plants or the plant is more toxic in some localities than in others. Future experiments only can determine

EUTHALLOPHYTA SCHIZOPHYTA

SCHIZOMYCETES. BACTERIA.

Bacteria produce disease in two ways:-First, as parasites when they derive their nourishment from the living animal; in this case they may cause embolism as in the case of Bacillus anthracis, or they may produce within the body products, toxins that are poisonous, as the diptheria bacillus does. The tetanusbacillus though parasitic produces powerful poisons that when injected even in minute doses cause a fatal termination, producing all the symptoms found in animals having the disease. Second, many saprophytic bacteria produce poisonous substances, especially such as occur in putrid flesh, fish, and other decaying substances. The list of such organism is a long one and cannot be given in this connection.

SCHIZOPHYCEAE.

Oscillatoriaceae. Blue-green algae.

Oscillatoria.

Several species in the U.S., probably somewhat injurious.

Nostocacae.

Nostoc caeruleum Lyngb.

N. muscorum, Ag.

N. commune Vauch.

Dr. J. C. Arthur some years ago thought that one of the common bluegreen algae, a species of nostoc, found in lakes in Northern Iowa and Southern Minnesota, was poisonous. Its poisonous nature was not conclusively demonstrated. These algae have been suspected in other parts of the world. Mr. George Francis calls attention to the Nodularia occurring in a fresh water lake in Australia. Thirty ounces of a scum fed to sheep produced death. It is also poisonous to horse, dogs and pigs. Many of the algae of this group

¹ Rept. Bur. An. Ind. 1898:421. 2 1.c. 437.

produce very disagreeable odors when decomposition occurs, and this plant is no exception to the rule. A few of these algae may be mentioned.

Anabaena flos-aquae Breb.

Anabaena stagnalis, Kg. Both of these are found floating on the water. In their decomposition they produce pig-pen odors.

EUPHYCEAE—Algae CHLOROPHYCEAE.

Volvocaceae.

Pandorina. Common in stagnant pools, especially in barnyards. The water is repulsive. Cattle will not drink it unless driven to do so. May be injurious. Volvox may be placed in the same category.

EUMYCETES---Fungi

PHYCOMYCETES. BLACK MOULDS AND DOWNY MILDEWS.

Mucoraceae.

The species of this family are common; among them are: Mucor mucedo L., found on horse manure.

Mucor corymbifer F. Cohn, and Mucor rhizopodiformis F. Cohn. Both species are pathogenic.

Mucor stolonifer Ehrb.

Common in the United States. Not pathogenic. Mouldy vegetables, etc. Mucor racemosus A. Fres.

Is widely distributed in North America. Mouldy vegetables, etc.

BASIDIOMYCETES.

Ustilaginaceae.

Ustilago Zeae (Beckm.) Ung. Corn smut is supposed to be poisonous to cattle, but the evidence is not very conclusive.

Ustilago avenae (Pers.) Jens. The common loose smut of oats is supposed to be injurious in large quantities, the same may be said of other smuts occurring upon cereals. Among these are barley smut *Ustilago hordei*, *U. nuda*, *U. tritici*.

Ustilago neglecta Niessl.

Pigeon Grass smut is thought by many farmers in Iowa to cause poisoning, especially abortion. Prof. Power, formerly of the University of Wisconsin, found present in this smut a small quantity of ergotin.

Ustilago utriculosa Tul. Smartweed Smut.

This common smut is said to produce irritation and frequently is the cause of trouble at husking time. This fungus is widely distributed in North America.

Tilletiaceae.

Tilletia foetens (B. & C.) Trel.

Stinking smut of wheat is not common in this state, but when it occurs in flour it causes bad odors. The spores give to the flour a dark color and make it unsalable.

Melampsoraceae.

Coleosporium solidaginis (Schw.) Thum.

Parasitic on golden rod and some other plants of the order. Common in

the state. A number of horses in Black River Falls, Wisconsin, a few years ago, became diseased, it is thought, by means of this rust. It may produce a form of Mycotic stomatitis.

Pucciniaceae.

Puccinia graminis Pers.

Wheat and oat rust, especially the uredo stage, produces inflammation of the mucus membrane of the mouth and nose. The dust coming from the straw when the grain is threshed often causes serious disturbances. Other rusts might be mentioned in this connection, like *Puccinia coronata*, Cda., the uredospores of which have an effect similar to that of the common grass rust. The above rusts are widely distributed in North America. *Uromyces trifolii*, Clover Rust, is widely distributed on red clover and, according to several authorities, is responsible for mycotic stomatitis.

Polyporaceae.

Bolctus felleus Bull.

It has a bitter taste and is poisonous.

Agaricaceae.

Lepiota morgani Pk.

This fungus is very common in the fall in meadows, pastures, and lawns. The cap is from 5 to 11 inches across. The gills or radiating plates beneath the cap are brown when mature. The lower part of the stipe is somewhat enlarged but no cup occurs. This when eaten by some people is known to produce poisoning.

Amanita muscaria L. Fly agaric.

Used to poison flies in Asia. Poisonous to persons.

Amanita phalloides Fr. Amanita.

This species is very poisonous and no one should eat so-called mushrooms unless he is familiar with them.

Amanita sprcta Pk.

This species is said to be poisonous in Eastern North America.

Amanita rubescens Fr.

This species is said to be edible, but authorities recommend that it should be used with great care. The same is true of A. solitaria. Both species are found in E. North America.

Amanita verna Bull.

This species is closely related to A. phalloides, but it is pure white in color; it is deadly poisonous. A. virosa Fr. belongs to the same category. A. frostiana Peck is also poisonous.

Clitocybe illudens Schw. Jack-my-Lantern.

This species is common in Eastern North America and is phosphorescent. While it is not dangerously poisonous, it is not edible.

Phallaceae.

Ithyphallus impudicus (L.) Fries.

The common stinkhorn is probably poisonous. Its disagrecable odor, however, would seem to render it distasteful to animals.

Lycoperdaceae.

Lycoperdon Bovista L.

The giant puffball is edible in fresh condition, when the flesh is white, but

in the mature form in considered poisonous; the same may be said of other puffballs.

ASCOMYCETES

Aspergillaceae.

Aspergillus glaucus Link.

This fungus is supposed to produce staggers. Frequently found in mouldy hay and gives rise to digestive disorders. Mycotic stomatitis.

Aspergillus niger van Tieghem.

This mould also occurs in mouldy hay and other mouldy substances and like the preceding species is injurious.

Aspergillus fumigatus Fr.

Commonly found on decaying substances, especially mouldy hay. It is pathogenic. Found in the ear of man as a parasite.

Aspergillus flavus Link.

Also pathogenic.

Hypocreaceae.

Claviceps purpurea Fr.

Found on many different grasses, especially wild rye, cultivated rye, timothy and quack grass. Produces a disease known as crgotism. The fungus is very poisonous, causes dry gangrene and abortion.

FUNGI IMPERFECTI

Many of the imperfect fungi undoubtedly produce Mycotic stomatitis. Oidium albicans Robin.

Commonly found in the mouth of sucking animals of different kinds, especially calves. Blastomycosis produced by Oidium or Saccharomyces.

Fusarium equinum Norg. Umatilla horse mange.

Dr. Mohler and others have demonstrated that the fungus is parasitic and produces this mange.

Fusarium roscum is commonly found on corn and other mouldy grain. It is probably concerned in forage poisoning. Dr. Burrill and Prof. Barrett report several forms of Fusarium on corn in Illinois.

Diplodia zeae Lev.

This disease is widely distributed on the sheaths and blades of corn in the United States. It is reported as common in Illinois by Burrill and Barrett and in Nebraska by Heald and Wilcox. It is also common in the state of Iowa. Dr. Erwin F. Smith suggests some connection between pellagra and this mould.

Sporotrichum Furfur Rob. Favus.

Sporotrichum tonsurans. Barber's Itch.

Polythrincium trifolii Kunze. Helminthosporium gramineum and other fungi may be responsible for mycotic stomatitis. The rape fungus, Polydesmus exitiosus Kühn, has long been associated with this disease.

EMBRYOPHYTA PTERIDOPHYTA

Polypodiaceae. Ferns.

Pteris aquilina L.

The common brake is found in eastern states and across the continent;

Oregon and Washington. It produces a large, strong root-stock and is said tobe poisonous to cattle and horses.

Adiantum pedatum L. Maiden Hair Fern.

Said to be poisonous. Widely distributed in Northern States.

Aspidium marginale L.

The root-stock is used as a remedy for the expulsion of tapeworm and undoubtedly sometimes produces poisoning. E. N. Am., Canada to Mo.

Cystopteris fragilis (L.) Bernh. Bladder fern.

This and many other ferns, according to Greshoff, contain HCN.

Osmundaceae. Royal Ferns.

Osmunda claytoniana L.

This fern is widely distributed in the eastern states. The odor is not pleasant and the plant is undoubtedly more or less poisonous.

Equisetaceae. Horsetails.

Equisetum arvense L.

The common horse-tail is supposed to be injurious to horses, as reported in Vermont by Dr. Richman and Professor Jones, and reports of poisoning in other parts of the country are recorded. In Europe it has long been suspected of being poisonous.

Equisetum robustum A. Br.

It is common in the state and, like the preceding, is considered poisonous.

A suspected case of poisoning recently was reported to the writer from Iowa.

EMBRYOPHYTA SIPHONOGAMA

(Flowering Plants)

GYMNOSPERMAE Conifers and Allies

Taxaceae. Yews.

Taxus canadensis Marsh. American yew.

Found in northern U. S., especially on calcareous sandstone rocks or insandy woods. The European species has long been regarded as poisonous tostock. Cases of poisoning have been reported in this country as well. It contains toxic substances, one of which is known as taxin.

Taxus brevifolia Nutt. Yew. Common on Pacific Coast.

Pinaceae. Pines and Junipers.

Juniperus communis L. Common juniper.

Poisonous, especially the oil obtained from the juniper berries. Rusby and others refer to the poisonous nature of this plant, and Schaffner records that goats are poisoned by eating the leaves. Common only in lime and sandstone-rocks in Northern U. S. and Rocky Mountains.

Juniperus virginiana L. Red Cedar.

Common in Northern and Eastern N. Am. to Texas. The leaves contain the same principle as that found in the juniper, and according to Schaffner, are poisonous to goats. The oil produces abortion and poisoning has also resulted from its use. The oil of Cedar has well known antiseptic properties. It contains an aromatic body, cedren, oil of cedar, from which cedren-camphor has been obtained.

Juniperus scopulorum Sarg.

Probably poisonous, like the preceding.

Juniperus sabina L. Swedish juniper.

Poisonous, like the preceding species. Cultivated.

Juniperus horizontalis Moench. American Savin.

Poisonous, Canada to Minn., in swamps.

Juniperus occidentalis Hook and vars. Red cedars.

Common from the western slope of the Rocky Mountains westward.

Sequoia sempervirons Endl.

Redwood leaves are said to be poisonous.

ANGIOSPERMAE

Typhaceae.

Typha latifolia L. Common Cat-tail. Said to be poisonous to cattle.

MONOCOTYLEDONEAE.

Alismaceae.

Alisma Plantago-aquatica L. Water plantain.

Recorded as being poisonous. Swamps.

Sagittaria latifolia Willd. Large arrow head.

The milky juice is somewhat bitter. The plant is edible, when cooked. The root stocks of several species of the genus are caten by the Indians and in China. Swamps.

Gramineae.

Zea Mays Gärtn. Numerous cases of poisoning have been recorded from the consuming of corn. This, however, is probably due to the production of a poison from the growth of mould or bacteria. The feeding of large amounts of dry fodder produces impaction of the stomach.

Andropogon sorghum Brot. Sorghum.

Second growth sorghum has frequently been reported as poisonous to live stock; this is due to the formation of HCN in the wilted leaves. *Calamagrostis, Milium* and some other grasses contain the same substance.

Setaria italica, Kunth. Millet.

Both the German Millet and the Hungarian Grass are poisonous to horses, acting especially on the kidneys. The poisoning is probably due to a glucoside.

Stipa spartea Trin. Needle Grass, or Porcupine Grass.

The sharp pointed callus often inflicts serious injuries; the fruits work their way under the cuticle into the flesh of the animals, and in some instances they have even penetrated the intestines.

Stipa comata Trin. Western needle grass.

Somewhat injurious, like the preceding. West of Missouri River.

Stipa robusta Vasey. Sleepy grass.

This grass has been suspected of producing stupor in horses. Rocky Mountains. Some species produce HCN.

Avena sativa L. Common oats.

The chaff of this grass sometimes produces balls in the stomachs of horses, known as phytobezoars.

Avena fatua L. Wild oats.

Common in a few counties in northern Iowa. Common in N. West., Cali-

fornia, Rocky Mts. Sometimes causes mechanical injuries on account of the pointed callus of the fruit.

Lolium temulentum L. Darnel.

The grain of this grass is injurious when ground in with flour. It produces stupor and symptoms resembling drunkenness. The poisoning is due to the fungus found in the seed. Principle loliin, a glucoside.

Hordeum vulgare L.

The chaff and awns of barley are often injurious, especially when coming in contact with the mucous membrane, not only in man, but in lower animals.

Hordeum jubatum, L. Squirrel-tail. Wild Barley.

Common throughout the west. This grass produces mechanical injuries in animals that feed on hay containing it, the awns working their way in between the teeth and maxillae, where they cause inflammation and the formation of pus. G. gussonianum, H. caespitosum, H. secalinum, etc., produce similar injuries.

Sitanion Elymoides Raf. Squirrel tail.

Common Rocky Mts. to Pacific Coast. Troublesome like Wild Barley.

Agropyron repens Beauv. Quack grass.

Widely distributed in northern Iowa. Produces a slight irritation of the mucus membrane. Contains triticin.

Araceae.

Arisaema triphyllum (L.) Schott. Jack-in-the-pulpit.

It is widely distributed in the north. The corm is known to be very acrid and poisonous, but when boiled or roasted the poisonous substance is expelled.

Arisaema Dracontium (L) Schott. Dragon's head.

Widely distributed, especially in eastern and central Iowa and northern states. The corm is somewhat acrid and is used to destroy insects; it is said to be a good vermifuge. The action of the plant in fresh condition is somewhat similar to that of ammonia.

Symplocarpus foetidus (L.) Raf. Skunk Cabbage.

Local only in a very few places. Said to be poisonous, causes vomiting, and temporary blindness. The juice is acrid and the plant has a very disagreeable odor.

Liliaceae.

Zygadenus elegans Ph. Swamp Camas.

Common in northern Iowa, Minnesota and westward. In the western states it is regarded as poisonous to cattle and sheep, occasionally causing death. It is not as poisonous as some other species of the genus.

Zygadenus venenosus Wats. Death Camas. Common in the Rocky Mountains and westward. The bulb is especially poisonous.

Melanthium virginicum L. Bunch-flower.

Common on low grounds in eastern Iowa and South and East. The root stocks are regarded as poisonous, but reports have come to us of the poisonous effect on horses, of the leaves and stems, when occurring in hay.

Veratrum woodii Robb. False Hellebore.

Southeastern Iowa to Mo. Poisonous like the eastern white Hellebore and the western California Hellebore. Probably contains jervin, cevadina and cevadin.

Veratrum viride Ait. Common swamp hellebore.

Eastern North America; swampy places. Seeds are poisonous, also herbage under some conditions.

Veratrum californicum Durand. California hellebore.

Rocky Mountains to Pacific Coast.

Amianthium Muscaetoxicum (Walt.) Gray. Fly poison. Crow poison.

Common along the Atlantic Coast and Long Island southward.

Erythronium purpurascens S. Wat. Dog-tooth Violet.

California. Contains saponin. Said to be poisonous.

Leucocrinum montanum Nutt. Leucocrinum.

Northwestern United States. Thought to be poisonous to sheep.

Nothoscordum bivalve (L.) Britton. Crow poison.

Southern United States.

Aletris farinosa L. Colic root.

Said to be poisonous. Sandy soil. E. N. Am.

Medcola virginiana L. Indian cucumber root.

From New Brunswick to Minn., and Florida. Said to be poisonous.

Chamaelirium lutcum (L.) Gray. Blazing Star.

Said to be poisonous. Mass. to Ark. and Fla. low grounds.

Chlorogalum pomeridianum Kunth. Soap plant.

Pacific Coast. Contains saponin.

Allium canadense L. Wild onion.

Widely distributed; common in low pastures, N. U. S. Milk is flavored where cattle feed on the plant. The A. unifolium of California is poisonous.

Allium tricoccum Ait. Wild Lcck.

Eastern and northern States. Taints milk like preceeding.

Lilium superbum L. Turk's-cap lily.

According to Schaffner, this species produces dermatitis. The bulbs produce mental exhaustion and headache.

Asparagus officinalis L. Asparagus.

According to Dr. White, in his Dermatitis Venenata, persons who constantly work with asparagus may have the skin somewhat blistered.

Convallaria majalis L. Lily-of-the-valley.

All parts of this plant are very poisonous to man and domestic animals. Contains two glucosides, convallamarin $C_{4n}H_{44}O_{24}$, and convallarin $C_{34}H_{31}O_{11}$. Trillium grandiflorum, Salisb. Large flowered Trillium.

This is used as an emotic and contains a principle which has been called trillin, found in a few other species of the genus.

Trillium erectum L. Erect Wake-robin.

The root stock of this species is somewhat poisonous.

Smilax rotundifolia L. Round-leaved Greenbrier.

Widely distributed in the northern states. Dr. Schaffner reports a case of poisoning from eating the young leaves of these plants. The spines are injurious in a mechanical way; they cause inflammation and pus formation.

Haemodoraceae.

Lacnanthes tinctoria (Walt) Ell. Red root.

Mass. to Florida. It is said that white hogs are subject to poisoning from this plant.

Amaryllidaceae.

Narcissus poeticus L. Narcissus.

This plant and other species regarded as poisonous in Europc.

Agave americana L. Century plant.

Used as a fish poison in some countries. Cultivated.

Zephyranthes atamasco (L.) Herb. Atamasco lily.

From Pa. to Florida. Said to produce the disease "staggers."

Belamcanda chincusis (L.) D C. Blackberry lily.

Rhizomc acrid, said to be deleterious, E. N. Am. Naturalized.

Dioscoreaceae.

Several species of Dioscorca or yams are poisonous.

Iridaceac.

Iris versicolor L. Large Blue-flag.

The underground root stocks of this plant are known to be very poisonous. Dr. Rusby thinks there is some danger "that it might be eaten in mistake for Calamus, which is commonly known as Sweet-flag. If so, it would prove seriously if not fatally poisonous, as its well-known emetico-cathartic properties, even when toned by drying and keeping, are powerful, and in a fresh state would be decidedly violent." It contains the resinous body iridin.

Orchidaceae.

Cypripedium spectabile Swz. Showy Moccasin flower.

Causes dermatitis resembling that produced by Poison Ivy. A great many persons are more or less susceptible to this form of dermatitis. Swamps in Northern States.

Cypripedium pubescens Willd. Yellow Lady-slipper.

Poisonous like the preceding. This species is more widely distributed.

Cypripedium candidum Muhl. Small white Lady-slipper.

Less poisonous than the preceding. At one time common in the marshes in the north, but rapidly disappearing.

DICOTYLEDONEAE.

Salicaceae

Populus balsamifera L. Balsam Poplar. Said by Dr. Halsted to be poisonous.

Juglandaceae.

Juglans nigra L. Black walnut.

It is commonly believed that the bitter principle Juglandin found in the fruit and leaves of this black walnut are more or less poisonous, at least it is thought that different species may poison the soil and prevent the growth of other plants; this is, however, not well substantiated. Probably contains nucin, an acrid body, causing an irritation which resembles scarlatina.

Fagaceae. (Beech Family).

In Europe beech nuts are known to produce some form of enteritis and the burs of the chestnut, Castanea dentata, produce mechanical injuries. Species of oak(Quercus) particularly the acorns of Quercus rubra are injurious to cattle and the nuts of European oaks produce death in hogs and cattle. This may possibly be due to the tannin and glucoside contained in the seed. The Q. Chrysolepis of California is poisonous.

Urticaceae.

Maclura pomifera (Raf.) Schneider. Osage orange.

Cultivated especially in the southern part of Iowa to Nebraska and Southward. The leaves and fruit are more or less poisonous. The thorns upon the plant produce serious injuries giving rise to inflammation.

Humulus Lupulus L. Common Hop.

Hop pickers often have an inflammation of the hands. The plant is a sedative and contains cholin, lupulic acid and oil of humulus.

Cannabis sativa L. Hemp.

Naturalized in many parts of the North. The narcotic effect of the resin of the plant is well known and in India an intoxicating drink is made from the juice of the leaves. Contains the substances cannabin, and oxycannabin.

Urtica dioica L. Stinging nettle.

The urticating properties of our common nettle are known to all who have had any experience in collecting the plants; there is at first a reddening, followed by a swelling, intense burning, and a small amount of itching. It is said to contain formic acid. The Western Nettle *U. holosericea* of Utah has similar properties.

Urtica gracilis L. Slender nettle.

Injurious like the preceding but more widely distributed.

Laportea canadensis Gaud. Wood nettle.

This plant is even more widely distributed than the preceding nettles. It is found in deep woods. It produces an irritation of the skin like that caused by common nettle.

Loranthaceae.

Poisoning has been reported from the European mistletoe Viscum album and there are a few references in America regarding poisoning by the false mistletoe Phoradendron flavescens. Possibly the Arceuthobiums may also be injurious.

Aristolochiceae.

Artistolochia Clematitis L.

Atlantic States from New York to Md. Some of the European species of Aristolochia are poisonous like the A. Clematitis which is naturalized along the Atlantic Coast.

Polygonaceae.

Rumex acetosella L. Sheep sorrel.

The plant is widely distributed in the U. S. and is becoming more common. Said to be poisonous to horses and sheep. Contains oxalic acid.

Rumex crispus L. Sour dock.

The plant is an astringent and is looked on with suspicion as are some of the other species of the genus.

Fagopyrum esculentum Moench. Buckwheat.

A dermatitis produced by the eating of buckwheat cakes is well known to most people and occasionally where screenings of this material are fed in quantities to hogs a similar rash is produced. Buckwheat straw is also considered poisonous. The plant contains the glucoside indican.

Fagopyrum tataricum Gaertn.

Poisonous like the preceding.

Polygonum acre H B K. Smartweed.

The acrid properties of many of the species of Polygonaceae are well known. This species is widely distributed in the north. Contains probably polygonic acid.

Polygonum hydropiper L. Smartweed.

Poisonous like preceding.

Chenopodiaceae.

Chenopodium anthelminticum L. Worm-seed.

Occasionally reported in the state. Cases of poisoning from the oil of the seeds have been reported in medical literature. Contains the volatile oil of worm-seed. This is a narcotic-acrid poison.

Chenopodium ambrosioides L. Mexican Tea.

This species is occasionally reported with properties like the preceding.

Kochia scoparia (L.) Schrad. Summer cypress.

This plant contains saponin and, according to Greshoff, the extracts from the seeds of another species froth in a solution up to 1 to 700. These plants must be regarded as slightly toxic in their effect. We may add here that saponin has also been found in *Eurotia ceratoides*. Eurotia lanata is used as a forage plant in the West; it is commonly called winter fat.

Chenopodium mexicana Moq. Mexican Lamb's Quarters.

This plant contains saponin and is known to be poisonous.

Amaranthaceac.

Amaranthus retroflexus L. Green pigweed.

Common everywhere in eastern north America, also in the great basin. O'Gara reports bloat from it in Nebraska.

Amaranthus spinosus L. Thorny pigweed.

The species are injurious.

Greshoff states that the leaves of A. hypochondriacus give an extract which froths strongly and contains saponin. The saponin is only slightly toxic.

Atriplex Nuttallii S. Wats. Salt bush.

The leaves of the salt bush contain saponin and Greshoff also found the same substance in A. halimus, A. hortensis and A. laciniata. He states that the haemolysis of the seeds of some of the species is moderately great. In China a skin disease known as Atriplicismus is caused by a species of Atriplex.

Beta vulgaris. Mangolds and Sugar Beets.

The feeding of mangolds and sugar beets to sheep causes renal calculi.

Sarcobatus vermiculatus Hooker. Grease wood.

Frequently produces mechanical injuries. According to Chestnut one man lost over 1000 sheep, probably due to bloat, caused by this plant.

Phytolaccaceae.

Phytolacca decandra L. Pokeweed.

The roots and seeds contain a very poisonous substance. The young shoots are eaten as greens; probably the poisonous principle is dissipated on boiling the plant. Found from southern Iowa Eastward and Southward. Contains phytolaccin.

Caryophyllaceae.

Stellaria media L. Chick-weed.

This has been reported as poisonous, although the seeds are eaten by birds. Agrostemma githago L. Corn cockle or cockle.

Generally found in wheat fields. Screenings are often sold as stock food

and several cases of poisoning from food that contained screenings of cockle have been reported. When cockle is in flour, it is poisonous. Several cases of poisoning from flour containing cockle are on record. Cockle is said to be especially poisonous to poultry. Contains the substances saponin, sapogenin and the alkaloid agrostemmin.

Silene antirrhina L. Sleepy catchfly.

Very widely distributed in the north. Said to be poisonous.

Silene noctiflora L.

Widely distributed in the north. Said to be poisonous.

Widely distributed in northern states. Clover fields. Native to Europe, probably also poisonous.

Saponaria officinalis L. Bouncing Betty.

This plant is said to be somewhat poisonous. Naturalized in the East. Ry Mts. and Pacific Coast.

Vaccaria vulgaris Host. Cow cockle.

Common only in grain fields, seeds said to be poisonous, like corn cockle.

Nymphaeaceae.

Nelumbo lutea Pers. The American Nelumbo.

The root stock is used by the Indians for food. According to Schaffner it is said to be used to destroy cockroaches. Roasting dispels the poisonous principle.

Ranunculaceae.

Hydrastis canadensis L. Orange Root. Golden Seal.

In northeastern Iowa and eastward. Contains the alkaloids hydrastin, berberine, and xanthopuccine. Hydrastis causes severe ulceration and catarrhal inflammation.

Caltha palustris L. Marsh marigold.

The leaves of the marsh marigold are eaten, but the poisonous principle is dissipated on boiling. Plant found on low grounds, especially in northern states. The related species with whitish flower R. leptosepala of the Ry Mts. must be regarded with suspicion.

Actaea alba Mill. White baneberry.

More or less poisonous, but generally not eaten by live stock. Found in woods more or less widely distributed in northern states.

Actaea rubra Willd. Red baneberry.

Widely distributed in the state, but never abundant. Berries poisonous.

Delphinium consolida L. Field Larkspur.

Naturalized from Europe. Poisonous and fatal to cattle, frequently cultivated as an ornamental plant. It contains several poisonous alkaloids. The alkaloids, delphinin, delphisin, delphinoidin and staphisagrin occur in D. Staphisagria and may be looked for in some of our native larkspurs.

Delphinium carolinianum Walt. Carolina Larkspur.

Native to prairies, especially gravelly knolls. Reported as fatal to cattle.

Delphinium exaltatum Ait. Tall Larkspur.

Frequently cultivated, native to Europe.

Delphinium tricorne Michx.

Produces fascicled tuberous roots. Common in southern Iowa. Very poisonous to cattle.

Delphinium hesperium Gray. Larkspur.

California. Said to be poisonous to cattle. Also the *D. nudicaule* which has narcotic properties.

Delphinium menziesii DC. Western purple larkspur.

Common in Montana and Westward. According to Drs. Wilcox and Chestnut it is poisonous, although Dr. Nelson fed 2434 pounds of fresh leaves to sheep without injury. Montana and west.

Delphinium geyeri Greene. Wyoming Larkspur.

Common in Wyoming, Colorado, and Nebraska. It is known as the poison weed.

Delphinium recurvatum Greene. Larkspur.

Common in Southern California. Said to be fatal.

Delphinium scopulorum Gray. Tall Mountain Larkspur.

Rocky Mountains to Canada. The Canadian Department of Agriculture states that it is poisonous to cattle.

Delphinium trolliifolium Gray.

Common along the Pacific Coast, especially California, Washington, and Oregon. It is known as cow poison in Humboldt county, California. Chesnut says perhaps it is not equally poisonous throughout all stages of its growth.

Aconitum uncinatum L. Wild Monk's-hood.

Native to a limited area in northeastern Iowa, Wisconsin and Eastward Contains aconitin. Root, flowers and leaves are poisonous.

Aconitum napellus L. European aconitc.

Commonly cultivated in gardens. This plant is powerfully toxic; it contains several important alkaloids, as aconitin, pseudoaconitin, and aconin. Bodily heat is reduced by aconite. Winslow states that the smallest fatal dose to a man is a teaspoonful of aconite, equivalent to about gr. XXX of the crude drug. The minimum lethal quantity of aconitin is 1-16 of gr. for man. Large doses produce death by paralyzing the heart.

Aconitum columbianum Nutt.

Common in the Rocky Mountains and westward. In swamps near springs and banks at higher altitudes in Colorado and Utah.

Aconitum noveboracense Gray. Aconite.

Northern States.

Aconitum reclinatum Gray. Trailing Wolf's-bane.

Alleghany Mountains.

Helleborus viridis L. Green Hellebore.

This plant is sometimes an escape from cultivation from Long Island to West Virginia. All parts of the plant are poisonous and have long been so recognized in Europe.

Anemone nemorosa L. Wood Anemone.

The common wind flower is said to be a local irritant. Common in woods of N. States.

Anomene patens, var. Wolfgangiana, (Bess.) Koch. Crocus, Sand flower. Pasque flower. Wind flower.

This plant is local in central and southern Iowa, but in northern Iowa it is common on gravelly knolls, also in Ill., Wis., British America, Rocky Mountains, Nebr., Col., New Mexico. It is a well known irritant containing the bitter substance anemonin.

Clematis virginiana L. Virgin's bower.

This plant is widely distributed in the state along with other species of the

same genus. The herbage is said to be acrid and caustic. The juice of some species of the genus causes blisters, or even ulcers. The fresh leaves of the *C. erecta* are used as a vesicant in Europe, especially by beggars, hence sometimes called beggar's weed.

Clematis Fremonti Wat. Clematis.

Common from Missouri to Kansas,

Clematis Pitcheri Torr. & Gray.

From Southern Indiana to Texas.

Clematis ligusticifolia Nutt.

Very abundant in the Rocky Mountains west to the Pacific Coast along streams at a lower altitude. Greshoff reports saponin in a large number of species, notably C. Pitcheri, C. recta, and others; also hydrocyanic acid in C. Fremonti.

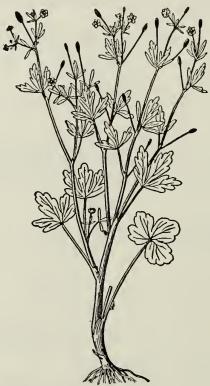


Fig. 19a. Common Crowfoot (Ranunculus acris). An acrid poison. (U. S. Dept. Agr.).

Ranunculus acris L. Tall buttercup.

Poisonous, causes inflammation when it comes in contact with mucous membrane. Rarely found in Iowa, but sometimes naturalized; common eastward.

Ranunculus septentrionalis Poir. Creeping Buttercup.

Widely distributed in low grounds. Acrid like the preceding species.

Ranunculus fascicularis Muhl. Tufted buttercup.

Found in eastern Iowa, as far west as the Iowa River; common north and east. Probably poisonous like the preceding species.

Ranunculus bulbosus L. Bulbous Crowfoot.

Common eastward, naturalized from Europe.

Ranunculus abortivus L. Crowfoot.

Common weed in many parts of the north, the leaves are quite acrid and have a sharp, peppery taste.

Ranunculus sceleratus L. Cursed crowfoot.

A very poisonous species, especially to cattle, since it grows in marshes along with other herbage and is often caten with other forage plants. Contains anemonol and anenmonic acid.



Fig. 19b. Common Poppy (Papaver somniferum). Narcotic. (Lois Pammel).

Anonaceae. Custard Apple Family.

Asimina triloba Dunal. Papaw.

It occurs in eastern Iowa as far north as Clinton and Dubuque. Common Southward. The pawpaw is commonly eaten, but a case of poisoning is reported.

Trollius laxus Salisb. Spreading globe flower.

From western Connecticut to Michigan and the Rocky Mountains. Several species contain saponin and T. Europaeus is considered poisonous in Europe. It contains saponin.

Thalictrum revolutum DC. Meadow Rue.

Common in low moist meadows in Western North America. All of the species of meadow rue arc more or less acrid. The *T. polycarpum* of California is poisonous to man according to Chesnut.

Magnoliaceae. Magnolia Family.

Magnolia grandiflora L. Magnolia.

The flowers of this plant are said to be injurious.

Illicium floridanum Ellis. Anise tree.

Native to Florida. The leaves are poisonous to stock.

Berberidaceae. Barberry Family.

Berberis repens. Lindl. Trailing Mahonia.

Cultivated. Poisonous. According to Schaffner, the berries are injurious to birds. Probably contain the alkaloids berberin and oxyacanthin which occur in the common barberry.

Berberis Aquifolium Pursh. Oregon grape.

This plant also causes poisoning. Cultivated. In the Cascade Mts., Oregon, Washington.

Podophyllum peltatum L. May Apple or Mandrake.

This plant is widely distributed east of the Missouri divide. The roots and leaves are drastic and are known to be poisonous. It is said also that when the leaves are eaten by cows, they produce injurious milk. The roots, according to Dr. White, are irritating to the eye, nose, mouth and skin, and contain picropodophyllin, podophyllotoxin. The podophyllin is a resinous inixture.

Caulophyllum thalictroides Mich. Blue Cohosh.

Widely distributed from Iowa eastward and northward. Said to be poisonous.

Menispermaceae.

Menispermum canadense L. Canadian Moonseed.

This plant is widely distributed in woods in Iowa and the north. Contains menispermin, and menispin. A case is reported of the death of three boys from eating the berries in mistake for grapes. The *Cocculus indicus* is a well-known remedy for the destruction of pediculi and is known to be poisonous. It contains picrotoxin, cocculin and an alkaloid menispermin.

Calycanthaceae. Allspice Family.

Calycanthus floridus L.

Cultivated in Southern Iowa and in the southern states. The aromatic properties of the flowers resemble those of strawberries. This makes it a very desirable cultivated ornamental plant. It contains an active principle calycanthin. Chesnut records it as poisonous.

The other species of Calycanthus may also be regarded as poisonous.

Lauraceac. Laurel Family.

Sassafras officinale Nees. Sassafras.

According to Schaffner the berries of this plant are reported to be poisonous. This species is native to southeastern Iowa, southward and eastward.

Umbellularia Californica (Arnott) Nutt. California Laurel.

California. Leaves smart. Used to drive fleas away.

Papaveraceae. Poppy Family.

Papaver somniferum L. Opium Poppy.

Opium is obtained from the common garden poppy. This species is widely cultivated in the state, and is frequently spontaneous. The more common alkaloids found in the poppy plant are morphin, codein and narcotin. The seeds of poppy are sometimes used to spread on top of cookies and bread.

Papaver rhoeas L. Corn Poppy.

This species is occasionally cultivated and sometimes spontaneous; the milky juice contains some poisonous alkaloids, as papaverin and codein.

Papaver dubium L. Long Smooth-fruited Poppy.

Cultivated; native to Europe, naturalized in Eastern North America.

Papaver orientale L.

This handsome, showy plant is frequently cultivated for ornamental purposes and contains a large amount of milky juice. This plant must be regarded as suspicious.

Argemone mexicana L. Mexican Prickly Poppy.

The Mexican prickly poppy, with pale yellow or yellowish petals, is cultivated in gardens. The latex not only contains narcotic principles, but the prickly pods sometimes cause mechanical injury and set up inflammation. The latex contains morphin: common in Texas to Kansas.

Argemone intermedia Sweet.

This prickly poppy of the plains contains a narcotic substance in its latex. The prickly leaves of the plant cause serious mechanical injury and inflammation. Common from Nebraska to Utah, Kansas and west.

Sanguinaria canadensis (Dill) L. Bloodroot.

This widely distributed plant of Eastern North America is well known to nearly everyone. The root is poisonous and contains the principle sanguinarinae, found in the red latex. It is sharply irritating, especially to mucous surfaces. Contains the alkaloid sanguinarin.

Chelidonium majus L.

Celandine is occasionally cultivated and spontaneous eastward. The orange yellow juice or latex is an irritant. The fresh juice produces inflammation and blisters, and it poisons the skin if handled so as to crush the leaves or stem. Contains chelerythrin $C_{21}H_{12}NO_4$, with a burning taste, and glaucin.

Stylophorum diphyllum (Michx.) Nutt. Celandine Poppy.

Common from Wisconsin to Tennessee and Pennsylvania. According to Schlotterbeck this plant contains the following substances which are regarded as injurious: protopin, stylopin, sanguinarine and others.

Eschscholtzia Douglasii (Hock & Arn.) Walp.

Narcotic used to stupefy fish. California.

Fumariaceae.

Fumaria officinalis L. Fumitory.

It has long been regarded as poisonous in Europe. Naturalized in waste places eastward.

Corydalis aurea Willd. Golden Corydalis.

A related exotic species which is said to be very poisonous. The above species and others of the family have been considered poisonous.

Cruciferae. Mustard Family.

Lepidium sativum L. Garden Cress.

Native to Europe, but occasionally escaped from cultivation. The Garden Cress is used as greens. It is a local irritant.

Lepidium apetalum Willd. Small Pepper-grass.

Widely distributed in this state, and like the other pepper-grasses, may produce sinapism.

Radicula armoracia (L.) Robinson. Horse-radish.

This is poisonous, like the preceding species of this order that have been

mentioned. Dr. Rusby refers to its irritating properties when taken in excess especially because of its action upon the urinary organs. One case referred to by Dr. Johnson was extreme and serious.

Sisymbrium officinale Scop. Hedge Mustard.

This plant is widely distributed in this state, as a weed, and produces more trouble than mustard.

Sisymbrium altissimum L. Tumbling Mustard.

This weed is common in grain fields in the north and may thus find its way into wheat screenings which should be fed with caution. It probably produces sinapism, as do the other plants of the order. It is also found in Iowa.

Brassica arvensis (L.) Ktze. Charlock.

This weed produces sinapism. After the application of the powdered material, there is a sense of burning. The volatile oil of mustard is a powerful irritant, and caustic, and should be used with caution.

Thlaspi arvense (Tourn.) L. Penny Cress.

This weed is said to be injurious to animals in Canada and is more or less avoided by them in pasture. It is widely distributed in Northern United States, but is more abundant in Canada.

Capparidaceae. Caper Family.

Cleome serrulata Pursh. Stinking Clover.

This plant is more or less pungent and acrid; it is widely distributed from Missouri River westward across the continent, especially along railways, etc. The flowers are showy and purple.

Cleome lutea Hook. Western Cleome.

This plant is a western species with yellow flowers and has similar properties to the one named above.

Polanisia graveolens Raf.

Fetid annual, with glandular hairs and common in sandy soils in the castern States. The *P. trachysperma* T. & G. is similar to the preceding. This plant is found from Iowa to Kansas.

Sarraceniaceae.

Sarracenia purpurea L. Side-Saddle Flower.

Probably not native to Iowa, although it is found distributed with Drosera in sphagnum bogs. It contains the substance sarracenin. The root produces diurcsis, gastric excitation, and an increased, irregular action of the heart. It produces papular eruptions changing to vesicular with depression as in smallpox. The plant was formerly used medicinally by the Indians.

Droseraceae. Sundew Family.

Droscra rotundifolia L. Sundew.

Sundew is said to be poisonous to cattle. In bogs in the northern states. Greshoff reported hydrocyanic acid in this species and in *D. intermedia*. Several other species in Europe were reported as harmful and toxic to cattle.

Crassulaceae. Orpine Family.

Sedum acre L. Stone Crop.

Produces inflammation when applied to the skin of many persons. The juice is acrid and biting.

Other species of this genus are known to be acrid. Several species with yellow flowers are common in rocky soils in the Rocky Mountains.

Saxifragaceae.

According to Greshoff several species of the genus Saxifraga of Europe contain hydrocyanic acid. The order also contains the currant and goose-berry which are well known food plants.

Hydrangea arborescens L. Wild Hydrangea.

Common in the Southern States and woods from Southern Iowa eastward to New York. According to Greshoff this species contains saponin; hydrocyanic acid occurs in the well known ornamental plant H. Hortensia.

Jamesia americana T. & G. Jamesia.

This plant is widely distributed from New Mexico to Montana. The leaves of this Rocky Mountain shrub contain hydrocyanic acid.

Philadelphus grandiflorus Willd. and P. coronarius L.

Mock Orange or Syringa. Native in the mountains from Virginia to Florida. The well known cultivated syringa contains saponin; according to Greshoff the Rocky Mountain P. microphyllus also contains saponin.

Deutzia. Several species of Deutzia, as D. staminea, contain saponin.

Chrysosplenium oppositifolium L.

This plant contains some saponin and the C. tetrandum Frics., is known to be poisonous to sheep.

Rosaceac. Rose Family.

Fragaria vesca L. European Strawberry.

Found on sand-stone and limestone rocks. In some people it produces irritation of the stomach.

Fragaria virginiana Mill. Wild Virginia Strawberry.

Similar to the preceding.

Fragaria chiloensis Duchesne. Cultivated Strawberry.

Like the preceding. There are people who can not eat strawberries or pick them without being irritated.

Rosa arkansana Porter. Arkansas Rose.

Widely distributed in the west, especially in prairie regions. Not poisonous, but the bristles and prickles often enter the skin and produce serious inflammation. Other species produce mechanical injuries.

Rosa rubiginosa L. Sweetbriar.

A frequent escape in pastures. The recurved spines and prickles are injurious like those of the preceding species.

Pyrus Aucuparia Meyer. Mountain Ash.

The berries are poisonous to man, but not to birds. However, they are readily disseminated by birds.

Pyrus communis L. Pear.

Dr. Schaffner states that horses are reported to have been killed by eating rotten pears.

Nuttallia cerasiformis T. & G. Oso Berry.

It is found in moist places and on north slopes in western U. S., and contains amygdalin. It is poisonous.

Gillenia stipulata (Muhl) Trel. Indian Physic.

From New York to Kansas. This species and G. trifoliata, known as Bowman's Root, are said to contain a poisonous glucoside.

Filipendula ulmaria (L.) Maxim Qucen of the meadow.

It is cultivated and occasionally an escape; said to be poisonous. Sanguisorba canadensis L. Burnet.

From Labrador to N. Mich, and Alleghany Mts. Said to be poisonous.



Fig. 19c. Coffee Bean (Gymnocladus dioica (L) Koch. The sweet juice in pod is poisonous. (Ada Hayden).

Pyrus malus L. Apple.

The seeds are poisonous and contain the glucoside which is changed into hydrocyanic acid.

Crataegus mollis L. Haw.

Cases of poisoning are reported by persons eating the fruit of *Crataegus mollis*, the injurious effects being probably due more to the incdible seeds than to the fruit.

Prunus americana Marsh. Wild Plum.

The shoots and seeds contain the principle amygdalin, which is converted into hydrocyanic acid. The fruit, it should be said, is entirely harmless.

Prunus pumila L. Sand Cherry.

This cherry is not widely distributed in the state, but is found along the Missouri river and in sandy soil eastward. The fruit is slightly acid and somewhat astringent, but is not poisonous, except the wilted leaves and the seeds, The astringent qualities in our wild fruit are undesirable. The same is true of P. Besseyi of Nebraska, the Dakotas and the Rocky Mountains.

Prunus pennsylvanica L. Wild Red Cherry.

Common, especially in the castern part of Iowa and northern states. The leaves are poisonous, as well as the seeds. The fruit is edible.

Prunus virginiana L. Choke Cherry.

The leaves and seeds are poisonous. The fruit is so astringent it often produces very unpleasant conditions when caten in any considerable quantity. Choke Cherry is widely distributed in the north. The leaves in the wilted condition contain hydrocyanic acid.

Prunus demissa (Nutt) Walp. Choke Cherry.

Common in the Rocky Mountains. Cases of poisoning from this species have frequently been reported.

Prunus serotina Ehrh. Wild Black Cherry.

Most poisonous species in the north. The half wilted leaves are much more poisonous than the fresh leaves, and the seeds are very poisonous, all producing hydrocyanic acid.

Prunus caroliniana (Mill) Ait. Laurel Cherry.

Laurel cherry is distributed in Southeastern United States. It is frequently cultivated for hedges. The leaves contain prussic acid.

Cercocarpus parvifolius Nutt. Mountain Mahogany.

It is poisonous and the same may probably also be true of *C. ledifolius*, both being common in the Rocky Mountains; the former contains hydrocyanic acid.

Amygdalus persica L. Peach.

The leaves and seeds are poisonous. They contain amygdalin, from which hydrocyanic acid is derived.

Leguminosae. Pulse Family.

Cassia chamaecrista L. Partridge Pea.

Widely distributed in the north especially on sandy, gravelly soils. A case was reported where a great many sheep had the scours; the ailment being attributed to this plant. Many species of the genus are known to be laxative.

Cassia marylandica L. Wild or American Senna.

Found in the southeastern part of the state, and probably acts like the preceding. This plant contains saponin.

Gymnocladus dioica (L) Koch. Kentucky Coffee-tree.

This is widely distributed in the Miss. Valley, especially along the river courses in Eastern Iowa, although growing as far north as Sioux City in the northwestern part of the state, and along the Mississippi into Minnesota. It is abundant in Missouri and Illinois. The fruit contains a sweetish, but disagreeable pulp which, as well as the leaves, is poisonous. In the South the leaves are used as fly poison.

Baptisia leucantha T. & G. Large White Wild Indigo.

It is widely distributed in prairie regions but is generally avoided by stock. Two eastern species are regarded as poisonous, being emetics. It is probable that this species must be regarded as suspicious.

Baptisia bracteata (Muhl) Ell.

This plant contains the same active substances as the preceding species, including baptitoxin. It is common in Eastern North America. The *B. australis*, or the blue indigo of the Southern States, is also regarded as poisonous.

Thermopsis rhombifolia (Nutt.) Richards.

Common in the Rocky Mountains and in the plains region. The seeds are said to be poisonous and the plant is very bitter. Several other Rocky Mountain species and one Eastern species must be put in the suspected list. These plants produce handsome yellow flowers.

Sophora secundiflora (Cav.) DC. Coral Bean.

Common in Central and Southern Texas. It contains a powerful poisonous substance. Reports of its poisoning occur in Mexico and Texas.

Sophora sericea Nutt. Silky Sophora.

Common on the plains of Colorado also from the Dakotas to Mexico Said to be poisonous.

Crotalaria sagittalis L. Rattle-box.

This is found in the western part of the state in the Missouri River Bottoms and produces a disease known as the Missouri Bottom disease, called crotalism, by Dr. Stalker. The seeds contain an unnamed alkaloid found by Dr. Power. The plant is not only poisonous in the meadow, but also in hay.

Cytisus scoparius (L.) Link. Scotch Broom.

Found along the Pacific and Atlantic oceans. It is common in Washington and Oregon; also in Massachusetts and Virginia. The flowers are yellow. The plant contains cytisin which also occurs in several other genera of this family like *Ulex*, *Thermopsis* and *Baptisia*.

Lupinus albus L. Lupine.

The European lupine is occasionally cultivated and in Europe it produces a disease known as lupinosis. Our native species, *L. perennis*, is also regarded as poisonous. Contains lupinin; lupinidin; lupinin.

Lupinus leucophyllus Dougl. Western Lupine.

It is said to be poisonous in Montana, although this is disputed by some. Other suspected species of the blue lupines are L. argenteus and L. argophyllus.

Trifolium incarnatum L. Italian or Crimson Clover.

According to Coville, it produces "hair" balls. This plant is cultivated as a cover crop in the south and east.

Trifolium repens (L.) White clover.

This plant and several other species such as T. pratense, at times, produce bloat.

Melilotus alba Desv. White Sweet Clover.

Widely distributed in the U. S. as a weed. The honey bees collect considerable quantities of honey from the Sweet Clover blossoms. It has been looked upon with suspicion. Dr. Schaffner states that the seeds impart a foul odor to flour.

Melilotus officinalis Willd. Yellow Sweet Clover.

This is also widely distributed in the state and is objectionable like the preceding.

Medicago sativa L. Alfalfa.

Alfalfa may cause bloat.

Psoralea tenuislora Pursh. Slender Psoralea.

Common from Illinois to Texas. Perennial herb sprinkled with little glandular dots. The Silvery Psoralea, P. argophylla, was reported from Iowa as poisonous. The tuberous roots of P. esculenta Pursh, were eaten by the pioneers and Indians.

Tephrosia virginiana Pers. Goat's Rue.

It grows only in sandy soil from Wisconsin to Iowa, east and south. It was used by Indians as a fish poison. Several other species in South America and Mexico have been used in a similar way. One is called *T. toxicaria*, and is a well known fish poison. Other N. Amer. species probably also poisonous.

Sesbania vesicaria Muhl.

An annual vine of the Southern States, Carolinas and Westward. The seeds are said to be poisonous.

Robinia neo-mexicana A. Gray.

Common in New Mexico and frequently cultivated as a hedge plant in Southern Colorado. The leaves somewhat resemble those of the black locust. Robinia pseudacacia L. Locust-tree. Black locust.

This plant is frequently cultivated in the north and west, and in numerous places is an escape from cultivation. Native to N. Y. and Alleghany Mts. The roots, leaves and bark are very poisonous to man. Contains robinin.

Robinia viscosa Vent. Clammy locust.

Cultivated as an ornamental plant; the roots are somewhat poisonous.

Astragalus mollissimus Torr.

A loco-weed from Neb. to New Mexico and Wyoming.

Astragalus hornii A. Gray.

Poisonous in Arizona and adjacent regions.

Oxytropis lamberti Pursh. Stemless Loco Weed.

Found in the western part of Iowa, along the Missouri River and its tributaries, very abundantly. One of the conspicuous loco or crazy weeds of the west.

Coronilla varia L. Coronilla.

An escape from cultivation in the West. It has long been regarded as a poisonous plant in Europe.

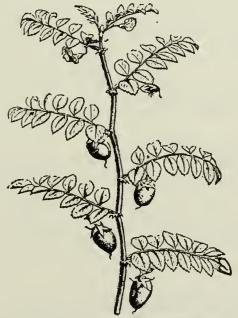


Fig. 19d. Chick Pea (Cicer arietinum) said to be poisonous.

Phaseolus lunatus Linn. Lima bean.

Investigations carried on in Europe seem to indicate that the lima bean leaves in the wilted condition contain hydrocyanic acid. According to Guignard practically all varieties, whether wild or cultivated, were found to contain a principle which when acted upon by an enzyme yields hydrocyanic acid. Prolonged boiling, however, extracts the greater part of it, but it is not destroyed, consequently this water should not be used, as it contains the substance which is converted into hydrocyanic acid.

Phaseolus multiflorus Willd. Scarlet Runner.

Commonly cultivated in gardens. The root is poisonous.

Vicia sativa L. Common Vetch.

A frequent weed in grain fields. The seeds of this are said to be injurious to pigs. It is not injurious to cows. Contains vicin.

Prosopis juliflora DC. Mesquit Tree.

The seeds of this tree contain a small amount of saponin, probably it is not very strongly poisonous as it is used as food for cattle in Texas.

Cicer arietinum L. Chick Pea. Idaho Pea.

Commonly cultivated in Southwestern U. S. Said to be poisonous in Europe.

Geraniaceae. Geranium Family.

Erodium cicutarium (L.) L'Her. and E. moschatum (L.) L'Her. Storksbill.

Both of these plants are widely distributed, especially on the Pacific Coast. The former is becoming abundant in Utah. The pointed callus of the seed sometimes inflicts mechanical injuries.

Oxalidaceae. Wood Sorrel Family.

Oxalis violacea L. Wood Sorrel.

Dr. Schaffner reports a case of a boy who was poisoned from eating a considerable quantity of the leaves. The leaves are frequently caten as a salad.

Tropaeolaceae. Nasturtium Family.

Tropaeolum majus. L. Nasturtium.

This plant is commonly cultivated and has more or less acrid properties.

Linaceae. Flax Family.

Linum usitatissimum L. Flax.

Said to produce death to cattle, probably due to the formation of HCN in the wilted leaves. People working with the fiber of the plant often have a form of dermatitis. Flax-seed, when fed in considerable quantities to live stock, especially hogs, produces death. Linum catharticum contains a bitter principle linin, and linamarin.

Linum rigidum Pursh. Large-flowered Yellow Flax.

This plant is reported as poisonous to sheep in some parts of the country. Found westward.

Zygophyllaceae.

Tribulus terrestris (L.) Caltrop.

Caltrop is found chiefly from Nebraska to Kansas and occasionally eastward. T. maximus, sometimes called soap-brush, is found in the west. The prickly fruit is more or less injurious; it presumably contains saponin. This plant should be looked upon with suspicion as one exotic species is regarded as poisonous.

Rutaceae. Rue Family.

Ruta graveolens L.

This plant is more or less acrid and produces blisters. Several species of the family are regarded as poisonous.

Zanthoxylum scandens and Z. alatum are used as fish poisons in India. It is not known whether our native prickly ashes are poisonous or not, but they may be looked upon with suspicion.

Simarubaceae.

Ailanthus glandulosa Desf. Tree-of-Heaven.

This plant is occasionally spontaneous in the United States, southcastern Iowa to Texas and eastward, where it is cultivated as an ornamental plant. It is supposed to produce poisoning when people come in contact with it. The odor of the flowers is very disagreeable. It is said also, according to Dr. Rusby, that water coming in contact with the leaves is poisonous.

Polygalaceae.

Polygala Senega L. Seneca Snakeroot.

The roots of this plant are used in medicine as an emetic. The plant is common in eastern North America. Probably other species of the genus Polygala are injurious.

Meliaceae.

Melia azcdarach L. Chinese Umbrella Tree.

This plant is commonly planted as an ornamental tree in the South and on the Pacific Coast; it has long been regarded as poisonous in Europe. Prof. Fawcett writes that it is so regarded in Florida and Chesnut reports that hogs have been poisoned by eating the seeds.

Euphorbiaceae.

Croton capitatus Michx. Hogwort, Croton.

Native to Southeastern Iowa and south. Many species of the genus contain very active poisonous principles. While this plant has not been suspected, the related species, *C. texensis*, is known to be poisonous. Croton texensis (Klotzsch) Muell. Texas Croton.

This plant is common from Western Nebraska to Colorado and South. It is poisonous. The S. setigerus of California is used as a fish poison.

Ricinus communis L. Castor-Oil Plant.

It is widely cultivated as an ornamental plant. The seeds contain a deadly poisonous substance ricin.

Euphorbia corollata L. Flowering Spurge.

Widely distributed in the Mississippi Valley, upon sandy or gravelly soil. Produces inflammation of the skin.

Euphorbia Preslii Guss. Spurge.

Widely distributed in meadows and fields. It has been sent to the writer as supposedly poisonous to live stock.

Euphorbia marginata Pursh. Snow-on-the-Mountain.

The honey coming from the plant is poisonous. The milky juice produces dermatitis. At one time the plant was used for branding cattle.

Euphorbia lathyris L. Caper or Myrtle Spurge.

Native to Europe, occasionally cultivated. The seeds of this plant are poisonous. Euphorbia resinifera contains euphorbon.

Euphorbia cyparissias L. Cypress Spurge.

Frequently escaped from cultivation, especially near cemeteries. Poisonous to the skin, producing dermatitis.

Probably we should add to this list other species like E. maculata which has several times been suspected of being poisonous in the state of Iowa. E. obtusata, common East of the Rockies, and E. heterophylla, known in the East as painted leaf, and common from Central Iowa eastward to Florida and

westward to Texas. E. maculata is common everywhere east of the Rockies. The E. Cyparissias was recently sent to me from Logan Iowa, where it is said to have killed lambs.



Fig. 19e. Flowering Spurge (Euphorbia corollata). (Lois Pammel). Fig. 19f. Three Seeded Mercury (Acalypha virginica). Both plants poisonous.

Acalypha virginica L. Three Seeded Mercury.

Common in fields and open places from Canada and Minnesota to the Gulf. The leaves of this plant turn purple in the Autumn.

Jatropha stimulosa Michx. Spurge or Bull Nettle.

This plant has stinging bristles and is common in sandy soil from Texas to Missouri, Virginia and Florida.

Tragia nepetaefolia Cav. Nettle Spurge.

With stinging hairs, common in the south. Other species in the Rocky Mountains and south also have stinging hairs.

Buxaceae.

Buxus sempervirens L. Box.

Cultivated in the eastern states for hedges. It is a well known poisonous plant.

Anacardiaceae.

Rhus toxicodendron L. Poison Ivy.

The leaves and stems are poisonous to many people. The form of dermatitis produced, and the seriousness of the case varies according to the susceptibility of the individual. The plant is widely distributed in the state.

Rhus vernix L. Sumac.





Poison Ivy. (After Halsted)

This plant occurs in swamps in the Northern States and is the most poisonous species.

Rhus Rydbergii Small. Poison Ivy.

This plant is common from Montana and British Columbia to Colorado and Nebraska. It is poisonous to touch.

Rhus diversiloba Torr & Gr. California Poison Ivy.

Common on the Pacific Coast. It is poisonous to the touch.

Celastraceae. Bitter Sweet Family.

Euonymus atropurpureus Jacq. Burning Bush, Waahoo.

The European species has long been regarded as poisonous.

Celastrus scandens L. Climbing Bittersweet.

The aril is red and has a somewhat disagreeable, sweetish taste. The leaves are said to be poisonous to horses. Bittersweet is quite widely distributed in Eastern North America.

Aquifoliaceae. Holly Family.

The berries of the European Ilex Aquifolium commonly cultivated in the East are said to be poisonous. This plant is called Poison Berry. The Ilex Cassine Yaupon or Black Vomit may be poisonous. This occurs from Virginia to Texas.

Hippocastanaceae. Buckeye Family.

Aesculus Hippocastanum L. Horse-Chestnut.

The seed of this species has long been recognized as poisonous in Europe. Frequently cultivated in the U. S. It contains aesculin, and argyraescin.

Aesculus glabra Willd. Ohio Buckeye.

Found in Southern Iowa, along river courses as far north as Fort Dodge and common South and East. Cases of poisoning have been reported, especially where cattle eat the young shoots and seeds of the plant.

Aesculus Pavia L. Red Buckeye.

Common from Virginia to Missouri.

Aesculus octandra Marsh. Buckeye.

Common Southward.

Aesculus californica (Spach.) Nutt. California Buckeye.

The leaves and fruit of this tree are regarded as poisonous to stock. The California Buckeye especially is said to cause abortion.

Sapindaceae.

Many members of this family are poisonous; among them several species of Paulinia sometimes cultivated in North America; also a species of the genus Sapindus. One species occurs in the Southern States, S. marginatus, known as Soap-tree or Soap-berry.

Balsaminaceae.

Impatiens fulva Nutt. Spotted Touch-me-not.

Widely distributed in the Northern States, especially on low grounds. The leaves are acrid and the plant is suspected of being poisonous to live stock.

The cultivated *I. Noli-tangere* is alterative and is regarded with suspicion. The exotic I. Royalii is considered poisonous.

Rhamnaceae.

Rhamnus cathartica L. Buckthorn.

Frequently cultivated and used as a hedge plant. The ripe fruit is said to be poisonous. It contains the glucosides rhamnin, rhamnetin, and rhamnocathartin.

Rhamnus lanceolata Pursh. Buckthorn.

Native to southern Iowa, east and south. Has the same properties as the preceding species.

Rhamnus Frangula L.

This plant is considered as poisonous. The R. Frangula of Europe is commonly cultivated. The tropical Colletia is poisonous.

Karwinskia Humboldtiana Zucc. Cajotillo.

Southwestern U. S. Dr. Mitchell of the 3rd U. S. Cavalry says that it is poisonous to goats. The berries have long been regarded as very poisonous.

Ceanothus velutinus Dougl. Ceanothus.

This species with shiny leaves and small white flowers occurs abundantly in the Rocky Mountains from New Mexico to British America, to Oregon and Washington. According to Greshoff, it contains saponin in considerable quantities. The same authority states that several other species examined by him contained the same substance, among them were C. azureus Desf., and C. thyrsiflorus Eschw. He also found the same substance but in less quantity in the seeds of the New Jersey tea C. americanus L. and C. ovatus Desf., two shrubby plants of Eastern North America, the latter being also common on the east slope of the Rockies.

Vitaceae. Grape Family.

Psedera quinquefolia (L.) Greene. Virginia Creeper.

Common from New England to the Rocky Mountains, especially east of the Missouri River. This plant is regarded as poisonous by some. The leaves and fruit abound in raphides. In this connection it may be of interest to state that the fruit of Cissus nivea of the old world produces poisonous fruits, and that the C. pruviens with a pleasant, acid, sweet taste produces a painful, burning sensation later. The same is true of the southern Mustang Grape (V. candicaus).

Malvaceae.

Abutilon Theophrasti (Medic) Gaertn. Velvet-Leaf. Indian Mallow. Butter Print.

Widely naturalized in the state. The plant gives off a very disagreeable odor, and is suspected of being poisonous.

Gossypium herbaceum L. Common Cotton.

The root of cotton is well-known as an abortive. Feeding excessive amounts of cotton seed meal frequently produces death in animals, particularly in hogs.

Dr. Crawford states that this is due to meta and pyrophosphoric acid in cotton seed meal.

Sida urens L. The Stinging Sida.

Occurs in tropical America and produces mechanical injuries because of the hairs with which it is covered. This is also true of the *S. paniculata* L. Probably other species of Sida of the southern states may produce similar mechanical injuries.

Hypericaceae.

١

Hypericum maculatum Walt. Spotted St. John's-Wort.
All the species are suspected of being poisonous to horses. Vesicant.

Hypericum perforatum L. Common St. John's-Wort.

Naturalized in Eastern U. S. A well known vesicant.

Hypericum Ascyron L. Great St. John's-Wort.

Most widely distributed species in the state, in woods or borders of woods.

Violaceae.

Viola odorata L. Sweet Violet.

Commonly cultivated in greenhouses. Said to be somewhat poisonous. Underground parts of the plant are emetic.

Viola cucullata Ait. Common Blue Violet.

The most widely distributed species in the eastern states; the roots, like the preceding, are emetic.

Loasaceae.

Mentzelia ornata Torr. & Gray. Mentzelia.

The backwardly-barbed trichomes sometimes produce mechanical injuries. Found in Northwestern Iowa on the Big Sioux near Sioux City, and west and southwest.

Datiscaceac.

Datisca glomerata (Presl) Benth & Hook.

California fish poison.

Cactaceae.

Opuntia Rafinesquii Engelm. Cactus.

Found in sandy soil in the state. The barbed trichomes cause mechanical injuries.

Other spiny species of Opuntia, Mammillaria and Cereus, largely developed in southwestern United States, produce similar mechanical injuries.

Anhalonium Lewinii Henn. Mescal Bean.

This cactus contains a narcotic substance which has long been used by the Indians of Mexico and adjacent regions to produce narcosis.

Cercus grandiflorus Mill. Night-blooming Cereus.

It has been regarded with suspicion where the plant is cultivated and native. Frequently cultivated in the United States.

Thymelaeaceae. Mezereum Family.

Dapline Mczercum L. Spurge Laurel.

This is a well-known poisonous plant of Europe and is occasionally cultivated. It contains the glucosides daphnin and daphnetin.

Other cultivated species in North America are known to be poisonous and several of the related genera native to Asia and Australia are poisonous.

Dirca palustris L. Leather-Wood, Moose-Wood.

Found in the North to Iowa. The bark is acrid, the berries narcotic and poisonous.

Elaeagnaceae. Oleaster Family.

Hippophae rhamnoides Linn.

A native of Europe but cultivated in the United States and said to be narcotic.

Araliaceae. Ginseng Family.

Fatsia horrida (Smith) B. & H.

From Isle Royal, Lake Superior region, to the Rocky Mountains and Alaska, and California. It causes mechanical injuries.

Umbelliferae. Parsley Family.

Conium maculatum L. Poison Hemlock.

Introduced here and there in Iowa and eastward, common in Utah. The plant contains the alkaloids coniin, conydrin, methylconiin, and a bitter principle cicutoxin. A very poisonous plant both to man and lower animals.

Petroselinum hortense Hoffm. Parsley.

Some people are suspicious of parsley. Dr. Schaffner states that the seeds are injurious to birds. He reports a case of poisoning of several parrots from eating the leaves of this plant. Cultivated.

Apium graveolens L. Celery.

Several cases are known where persons who have handled celery have had a form of dermatitis. Some persons cannot eat celery because a rash forms.

Cicuta maculata L. Water Hemlock. Cowbane.

The roots of this plant are very poisonous. The plant is widely distributed in the north, especially in low grounds. The European cowbane, *C. virosa*, contains the alkaloid coniin, a substance which probably also occurs in our plant The poisonous principle is cicutoxin.

Cicuta bulbifera L. Bulb-bearing Hemlock.

The roots of this, and the whole plant are supposed to be very poisonous. In swamps and northern states.

Cicuta vagans Greene. Cowbane.

Occurs in Washington, Oregon and California. Occasionally said to poison cattle drinking water in which they have trampled roots of this plant, thus expressing the extract. Very poisonous.

Cicuta Bolanderi A. Grav.

Found in marshy regions in California.

Cicuta occidentalis Dougl. Cowbane.

Found in the Rocky Mountains and considered poisonous.

Sium cicutaefolium Gmelin. Water Parsnip.

Common in many parts of the north in low grounds. Said to be poisonous. Aethusa Cynapium L. Fool's Parsley.

A poisonous herb native to Europe, with a disagreeable odor. Possibly occurs in a few places in the state. Contains the alkaloid cynapin and another coniin-like alkaloid.

Angelica atropurpurea L. Purple-stemmed Angelica.

Found in low grounds in North and Northeastern Iowa. Possibly poisonous. Cattle do not relish it.

Oxypolis rigidior (L.) Coult. & Rose. Cowbane.

Aquatic herb with white flowers, leaves simple pinnate with 3-9 linear-lanceolate leaflets.

Pastinaca sativa L. Parsnip.

Persons are often poisoned by handling the plant, which causes inflammation and vesication. Mr. F. C. Stewart, in a letter to the writer, states that in one case, the eyes became swollen, vesication occurred from poisoning caused by the flowers.

Heracleum lanatum Michx. Cow Parsnip,

Supposedly poisonous, although the leaves of the fresh plant are eaten by the Indians. This species is widely distributed in Iowa, especially in rich woods. Contains the bitter principle heraclin. Daucus carota L. Carrot.

The carrot, like the parsnip, causes vesication. Dr. Schaffner says that persons handling the plant are often poisoned, especially when the plant is wet with dew.

Cornaceae. Dogwood Family.

Cornus paniculata L'Her. Dogwood.

Widely distributed in the north. Some regard it as poisonous.

Cornus paniculata L'Her.

The fruits of this species are considered by some people to be poisonous, but there are no authentic cases.

Garrya Fremontii Torr. Dogwood.

Contains an alkaloid. It may be of interest here to state that the Marlea vitiensis of Australia and the Pacific Islands is poisonous. It would not be strange, therefore, to find that some of the other members of this family are poisonous.

Ericaceae. Heath Family.

Epigaea repens L. Trailing Arbutus.

North and East, Alleghany Mountains. Supposed to be poisonous. Contains the glucoside ericolin.

Andromeda polifolia L. Wild Rosemary.

Arctic America, Adirondack Mountains and in Europe and Asia A. glau-copliylla Link, is the Bog Rosemary, with whitish leaves found in bogs and wet shores from Labrador to Minnesota, and the A. floribunda Pursh. is found in moist hillsides in the Alleghany Mountains from Virginia to Georgia. These plants contain andromedotoxin.

Rhododendron maximum L. Large Rhododendron.

Small shrub or tree from Nova Scotia to Ohio and the Alleghany Mountains. *R. catawbiense* Michx., in the high Alleghanies from Virginia to Georgia, known as the Mountain Rosebay, and the California Azalea (*R. occidentale*) are known to be poisonous, the latter especially to sheep.

Rhododendron californicum Hooker. California Rhododendron.

From San Francisco to British Columbia. Cases of poisoning have been reported from the Pacific Coast.

Kalmia angustifolia L. Lambkill. Laurel.

Common on hillsides, pastures, and bogs, from Labrador southwest through the Alleghanies. Known to be very poisonous. The leaves contain andromedotoxin; frequent cases of poisoning on record.

Kalmia latifolia L. Laurel.

From New Brunswick to Ontario to Pennsylvania and south to the Alleghanies. A well-known poisonous shrub. Many cases of poisoning of sheep and cattle are on record from this species, probably the most poisonous of all the members of this genus. K. polifolia Wang., found in bogs, with rose-purple flowers and pale colored leaves, is poisonous.

Ledum glandulosum Nutt. Labrador Tea.

Common in bogs of the Northern Rockies, the Cascade Mountains to California. It is said to be poisonous. L. groenlandicum Oeder., with white flowers and rusty wool underneath, occurs in bogs from New England to Wisconsin and Minnesota. Said to be poisonous.

Leucothoe Catesbaei (Walt.) Gray.

With white flowers in axillary or spiked racemes; shrub. From Virginia to Georgia in the mountains; an allied species L. racemosa (L.) Gray, occurs in moist thickets from Massachusetts to Louisiana, near the coast. Both species are said to be fatal to stock, the latter especially to calves.

Lyonia mariana (L.) D. Don. Stagger-bush.

From the Atlantic Coast to Tennessee and Arkansas. Said to produce intoxication, hence the name.

Primulaceae. Primrose Family.

Anagallis arvensis L. Poor Man's or Shepherd's Weather Glass. Pimpernel. Possibly growing in the state. Known to be poisonous. Contains glucoside cyclamin.

Primula obconica Hance.

Poisonous to the touch; very much like poison ivy. This plant is commonly cultivated in greenhouses.

Primula Parryi A. Gray. Parry's Primrose.

Common in higher altitudes in the Rocky Mountains, especially near brooks or springs. The root has the odor of musk, and is said to be poisonous.

Cyclamen Europaeum L. Cyclamen.

This beautiful cultivated plant has long been regarded as poisonous in Europe. The C. persiclum Mill, is also regarded as poisonous.

Plumbaginaceae. Plumbago Family.

Limonium carolinianum (Walt.) Britton.

It is not known whether this plant is poisonous or not but a related plant, Statice pectinata Ait, of the Cape Verde Islands is poisonous.

Sapotaceae. Sapodilla Family.

Achras Sapota L.

A tropical fruit cultivated in Florida. It is said to be injurious and according to Greshoff contains saponin. Lucuma forms a well-known genus of West Indian fruit trees. In L. Bonplandia, H. B. K., Altamirans demonstrated amygdalin as early as 1876, and in another species a cyanogenetic glucoside was suspected, but the experimental proof was wanting. Greshoff reports hydrocyanic acid in the L. mammosa.

Ebenaceae. Ebony Family.

Diospyrus virginiana L. Persimmon.

Common in the Southern States as far north as southern Iowa. It is used as an anthelmintic, but it is not positively known whether any part of this plant is poisonous; several exotic species, however, blister the skin and one species in Madagascar is said to be very poisonous.

Oleaceae. Olive Family.

Ligustrum vulgare L. Privet.

The privet is frequently cultivated in the U. S. The leaves and fruit of the plant are said to be poisonous. Prof. Chesnut says that accidents have been occasioned in children, both by fruit and the leaves. It contains the bitter principle syringopicrin.

Forsythia suspensa Vahl. Forsythia.

Commonly cultivated in northern states. The root is slightly poisonous. The F. viridissima Lindl. is very bitter. Greshoff has found saponin in the seeds of the former species.



Mountain Laurel (After Halsted)





Fig. 19g. Oleander (Nerium Oleander). Cultivated poisonous. Fig. 19h. False Jessamine (Gelsemium sempervirens). Contains several poisonous alkaloids. (Chesnut, U. S. Dept. Agri.).

Chionanthus virginica L. Fringe Tree.

Common in the Alleghany Mountains, New Jersey to Florida and Missouri. This is said to be a narcotic.

Loganiaceae. Logania Family.

Gelsemium sempervirens (L.) Ait. Yellow Jessamine.

This climbing shrub with yellow fragrant flowers is common in the South. It contains several potent alkaloids, among them gelsemin. The symptoms of poisoning from consuming this plant are muscular weakness, especially in the fore legs, followed by convulsive movements of the head, fore legs and hind legs.

Spigelia marilandica L. Indian Pink or Pink Root.

Common from Ohio to Missouri, Florida and Texas. This plant is used in medicine and is known to be toxic. Strychnin belongs to this same family.

Gentianaceae. Gentian Family.

Menyanthes trifoliata L. Buckbean.

The plant has bitter properties and is nauseous. Contains menyanthin,

Apocynaceae. Dogbane Family.

Apocynum androsaemifolium L. Spreading Dogbane.

Widely distributed in Iowa, probably poisonous.

Apocynum cannabinum L. Indian Hemp.

Like the preceding. Contains apocynin, which is poisonous.

Nerium Oleander L. Common Oleander.

Cultivated. The leaves are deadly poisonous to stock. Contains conessine, and neriin, which has the properties of digitalin. Nerianthin bears a resemblance to digitalin.

Dr. Wilson of the Arizona Experiment Station has recently demonstrated the very poisonous nature of this plant in Arizona.

Asclepiadaceae. Milkweed Family.

Asclepias tuberosa L. Pluerisy-Root.

Widely distributed in Iowa, especially on gravelly knolls and prairies. The leaves are more or less poisonous to stock. However, honey bees collect considerable honey from this plant.

Asclepias incarnata L. Swamp Milkweed.

Poisonous probably like the preceding. The root is emetic and cathartic. Asclepias syriaca L. Milkweed,

Poisonous. Contains the glucoside asclepione, an amorphous bitter substance.

Asclepias speciosa Torr. Showy Milkweed.

Poisonous. This species is found in Western and Northwestern Iowa to Utah.

Asclepias campestris, Decne. Milkweed.

Commonly cultivated in gardens southward. It is said to be poisonous.

Asclepias eriocarpa Benth, Milkweed,

Common in California and adjacent regions. According to Chesnut sheepmen in California very much fear this weed. It has broad mullein-like leaves. Asclepias mexicana Cav. Narrow Leaved Milkweed.

Native to California, Oregon and Nevada. According to Chesnut sheep and calves are not infrequently poisoned by eating this plant and cows have been poisoned by eating hay contaminated with it.

Convolvulaceae. Convolulus Family.

Ipomoea pandurata Meyer. Wild Potato Vine. Man of the Earth.

The large root is poisonous. Contains the gulcoside impomoein.

Convolvulus sepium L. Hedge Bindweed.

The plant produces a somewhat disagreeable odor. Dr. Schaffner states that it is supposedly poisonous to swine. Jalap contains several glucosides which also probably occur in our Morning Glory. One is convolvulin.

Cuscuta epithymum Murr. Clover and Alfalfa Dodder.

Dr. J. Q. Taylor of Lisbon, Ohio, in writing to Prof. A. D. Selby stated that dodder produced a bowel trouble in horses, and Dr. Jenkins of New Haven writes that clover hay containing a great deal of dodder produced scours. He added that the hay had moulded badly, although the injury could not be definitely traced to dodder. Some exotic dodders are poisonous. The dodder must, therefore, be looked upon with suspicion.

Polemoniaceae. Polemonium Family.

Gilia aggregata Spreng. Cypress plant.

Common in the Ry. Mts. to Neb. This species according to Greshoff contains a considerable amount of saponin and is very poisonous. He lists several other species which contain this substance.

Hydrophyllaceae. Waterleaf Family.

Phacelia circinata Jacq. Rough Phacelia.

This plant is common in the Rocky Mountains. It produces rough bristles and causes considerable irritation and inflammation; other species act in a similar way.

Boraginaceae. Borage Family.

Heliotropium Europaeum L. Wild Heliotrope.

This plant contains a poisonous alkaloid and as well as the H. indicum L. has long been regarded as poisonous.

Cynoglossum officinale L. Hound's Tongue.

Poison acts much like Curare.

Cynoglossum virginicum L. Wild Comfrey.

Supposed to be poisonous.

Lappula officinalis Lehm. Stickweed.

The fruit of this plant gets into the wool of sheep and sometimes produces mechanical injuries.

Echium vulgare L. Viper's Bugloss.

Occasionally spontaneous, probably in the Eastern part of the state. Contains a poisonous alkaloid.

Verbenaceae. Verbena Family.

Callicarpa americana L. Mexican Mulberry.

Common in the southern states. Several Asiatic species of the genus used as fish poisons. No report of poisoning from the American species is recorded.

Labiatae. Mint Family.

Scutellaria galericulata L. Marsh Skullcap.

Common in wet places in the North. Suspected of producing poisoning. Stachys arvensis L. Corn or Field Woundwort.

Waste places, especially along the Atlantic Coast. Suspected.

Galeopsis tetrahit L. Dead Nettle.

Common bemp nettle with purplish flowers and stiff calyx bristles. Common Eastward and in the Northwest in British Columbia and Washington. Causes irritation.

Nepeta hederacea (L.) Trevisan. Ground Ivy.

Widely naturalized in the state. It is said to be poisonous to horses.

Hedeoma pulegioides Pers. Pennyroyal.

Common, especially in clay soils in Eastern Iowa. The oil is known to cause poisoning.

Leonurus Cardiaca L. Common Motherwort.

Known to produce mechanical injuries and dermatitis. Widely distributed in the U. S., naturalized in Europe.

Solanaceae. Ninghshadt Family.

Nicandra Physalodes (L.) Pers. Apple of Peru.

Cultivated here and there in Iowa. Said to be poisonous. Used as a fly poison in parts of the United States.

Solanum nigrum L. Black Nightshade.

The leaves and other parts of the plants are reputed to be poisonous to calves, sheep, goats and swine, and the green berries are known to be poisonous to man. The fruit of a form of this species is cultivated as an esculent.



Fig. 19i. Black Nightshade (Solanum nigrumy). Contains a poisonous alkaloid. (Chesnut, U. S. Dept. Agr.).

The writer has not only eaten berries of this, but has seen others eat berries of this and the common Black Nightshade without injurious results. Contains the alkaloid solanin, with a hot, bitter taste.

Solanum triflorum Nutt. Nightshade.

Common from Nebraska to Rocky Mountains to Alberta, south to Texas. Poisonous.

Solanum tuberosum L. Potato.

At certain times the tubers of the potato are poisonous, especially when green. The writer knows of an instance where the eating of potatoes acted as a poison. The substances produced in the young shoots of the potatoes are solanin and solanadin.

Solanum carolinense L. Horse-nettle.

Southern United States North to Minnesota and East to New York. Fruit with disagreeable odor. Said to be poisonous. Narcotic.

Solanum Dulcamara L. Bittersweet.

The berries are poisonous, as are also the leaves. Cattle are known to have been poisoned by it. The bitter substance contained in it is known as dulcamarin.

Lycopersicum esculentum Mill. Tomato.





Jimson Weed. (After Halsted)

The green parts of the plant contain saponin and solanin.

Physalis heterophylla Nees. Ground Cherry.

From New Brunswick southward and westward. Suspected plant as is P. virginiana Mill.

Cestrum cauliflorum Jacq. Cestrum.

West Indies. The C. nocturnum L., frequently cultivated in greenhouses. Several species are poisonous.

Atropa Belladonna L. Belladonna, Deadly Nightshade.

Occasionally cultivated and possibly also an escape. Very poisonous.

Nicotiana Tabacum L. Tobacco.

Cultivated. Narcotic and poisonous, and produces the alkaloid nicotin, a very poisonous substance.

Nicotiana alatum Link & Otto. Flowering Tobacco.

Poisonous like the preceding.

Nicotiana glauca R. Grah. Smooth Tobacco.

An escape from cultivation in California. According to Dr. G. Burtt Dayy poisonous.

Hyoscyamus niger L. Black Henbane.

Probably occasionally found in Iowa, Utah, Mont., Atlantic States. Known to be poisonous to stock and also to logs. Universally recognized as a poisonous plant in Europe and this country. Probably one of the most deadly poisonous plants in the United States. Seeds are poisonous to chickens. Contains the alkaloid hyoscyamin.

Datura Stramonium L. Jimson-weed.

Naturalized in various parts of the U. S. All parts of the plant are narcotic and poisonous, especially the seed. Several cases of poisoning in children are reported in Iowa. The plant produces a very disagreeable odor, and the hay containing the plant is poisonous to cattle. It contains the alkaloid, atropin, and hyoscyamin.

Datura Tatula L. Purple Jimson-weed.

Poisonous like the preceding, and the following species. Naturalized in U. S.

Datura Wrightii, DC. Wright's Datura.

Frequently cultivated as an ornamental plant and known to be poisonous. The nectar from the flowers which is produced in great abundance is known to produce poisoning in children in this country.

Datura alba Nees. Thorn Apple.

Cultivated. Plant said to have been much used in India for criminal purposes. Very toxic.

Capsicum annuum L. Red or Cayenne Peppers.

Well known remedy used as a stimulating plaster externally; if the pepper is applied long enough it produces vesicles. Red pepper is often injurious when taken in too large doses internally. The active poison is capsicol with a strong odor and burning taste.

Capsicum frutescens L. Shrubby Pepper.

Southwestern United States, said to be poisonous. The fruit has a sharp pungent taste.

Lycium halimifolium Mill. Matrimony Vine.

Cultivated and commonly naturalized. Somewhat spiny thorns. Said to be poisonous.

Scrophulariaceae. Figwort Family.

Verbascum Thapsus L. Moth Mullein.

Naturalized on the Atlantic coast and common in Utah and the west. Said to be poisonous.

Linaria vulgaris Hill. Butter-and-eggs,, Toad Flax.

A weed, especially northward. Suspected of being poisonous.

Scrophularia marilandica Gray. Simpson Honey Plant,

Widely distributed in E. U. S., pastures and woods. Not eaten by stock. According to Millspaugh, the physiological effect of this plant is bleeding of the gums, colic, and sleepiness. Contains a crystalline bitter substance, scrophularin.

Digitalis purpurea L. Purple Foxglove.

This plant is widely cultivated in the U. S., and naturalized on the Pacific Coast.

Is poisonous to man and live stock, especially horses. It contains the glucosides digitalin, which dilates the pupil; digitoxin, and digitonin.

Gerardia tenuifolia Vahl. Slender Gerardia.

Said to be poisonous to sheep and calves. Probably other species are likewise poisonous, like G. grandiflora and G. purpurea.

Pedicularis lanceolata Mx. Lousewort.

Widely distributed in low grounds and swamps. Said to be poisonous.

Pedicularis canadensis L. Lousewort.

Widely distributed in the state in gravelly soils and on knolls. Said to be poisonous. Sheep, however, eat large quantities of the P. groenlandica without apparent injuries.

Pedicularis groenlandica Retz. Mountain Lousewort.

Common at higher altitudes in swamps. Suspected of being poisonous.

Melampyrum lineare Lam. Cow Wheat,

Common in open woods eastern states to Tennessee. The European species, *M. silvaticum*, is regarded as poisonous in Europe. Our American plant has not, however, been reported.

Gratiola sp. Hedge Hyssop.

The European G. officinalis is said to be poisonous to cattle. Several species common in eastern North America.

Veronica virginica L.. Culver's Root.

Common in swamps in eastern North America especially northward. Contains saponin.

Chelone glabra L. Balmony.

In swamps northern United States, Contains an alkaloid, Suspicious,

Bignoniaceae. Bignonia Family.

Catalpa speciosa Warder. Hardy Catalpa.

Widely cultivated in U. S. Odor coming from the fragrant flowers is poisonous and Dr. White in his Dermatitis Venenata states that the flowers are irritating to many persons. Dr. Millspaugh, on the other hand, states that it is said to be dangerous to inhale the odor of the flowers for a long time, which, however, is probably not generally true. The allied Caroba contains the bitter principle carobin.

Catalpa bignonioides Walt. Common Catalpa.

Occasionally cultivated in the north, but scarcely hardy. Poisonous like the preceding.

Bignonia capreolata L. Cross Vine.

A tall climbing vine with large orange colored flowers; from Virginia to Louisiana. Said to be poisonous.

Tecoma radicans (L) Juss. Trumpet Creeper.

Climbing vine, large tubular orange and scarlet flowers. New Jersey to Iowa, Texas to Florida. Said to be poisonous, several exotic species are poisonous.

Rubiaceae. Madder Family.

Cephalanthus occidentalis L. Buttonbush.

This plant commonly occurs in low grounds, especially along river courses. The leaves contain a poisonous principle, cephalanthin, a very bitter glucoside.

Caprifoliaceae. Honeysuckle Family.

Triostenni perfoliatum L. Feverwort, Horse-gentian.

Widely distributed in woods. Some species of the genus were used by the Indians as a cure for fevers and early practitioners in this country used the root as an emetic. The physiological action of the plant is to produce vomiting.

Sambucus canadensis L. Elderberry.

Dr. Rusby states that the plant is poisonous. The elderberry is widely distributed in the U. S. and the flowers of this are commonly used to prepare a tea. Contains coniin.

Sambucus racemosa L. Red-berried Elder.

Northern states and Rocky Mountains. Northeastern Iowa. Suspected.

Symphoricarpos orbiculatus Moench. Coral-berry.

Common in the south, occurring on sterile or rocky soils and on borders of woods. Reported as poisonous.

Symphoricarpos mollis Nutt. Coral Berry.

This plant is said to contain saponin according to Greshoff. The S. race-mosus Michx. Snowberry. Across the continent northward. Greshoff found saponin in leaves but not the fruit. Cases of poisoning have been reported in the old world.

Lonicera involucrata (Richards) Banks. Rocky Mountain Honeysuckle.

Common in the Rocky Mountains. It is said to contain saponin in considerable amounts. Several other species contain saponin and may be regarded as poisonous.

Diervilla Lonicera Mill. Bush Honeysuckle.

Common in woods, north. Not reported as poisonous, but the Japanese D. Japonica DC is regarded as poisonous. It contains saponin.

Cucurbitaceae. Gourd Family.

Some of the exotic genera of this family, like the Bryonia dioica known as the poison berries, and the Trichosanthes palmata, are poisonous.

Sicyos angulatus L. Bur Cucumber.

Eastern North America. The barbed prickles are irritating and injurious to many persons according to Dr. Halsted.

Boleria sp.

One or more species of the genus regarded as poisonous in California ac cording to Chesnut.

Campanulaceae. Bluebell Family.

Lobelia cardinalis L. Cardinal-flower.

This is listed as one of the poisonous plants by Dr. Schaffner. Cardinal-flower is very abundant in the swamps along river courses in Eastern Iowa, and N. Miss. Valley.

Lobelia siphilitica L. Blue Lobelia.

Also listed as a poisonous plant by Dr. Schaffner.

Lobelia spicata Lam. Pale Spiked Lobelia.

Everywhere on prairies of the northern states. Probably poisonous like the preceding.

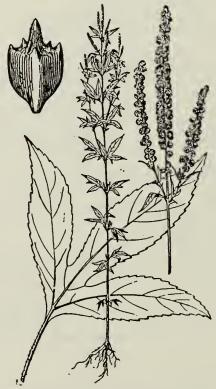


Fig. 19j. Greater Ragweed (Ambrosia trifida). Said to cause hay fever. (Dewey U. S. Dept. Agrl.).

Lobelia inflata L. Indian Tobacco.

Widely distributed, occurring in woods. The leaves of this plant were used by the Indians. The plant has long been used in medicine. L. inflata is very poisonous and is used for its action upon the pneumogastric nerve; the toxic doses produce exhaustion and dilation of the pupils. Death is usually preceded by insensibility and convulsions. Contains the acrid lobeliin.

Lobelia Kalmii L. Kalm's Lobelia.

Found in swamps from Nfd. to N. J. west to northern Iowa, Minnesota, and Man. Probably poisonous like the preceding.

Compositae. Composite Family.

Lactuca Scariola L. Prickly Lettuce.

Common across the continent, also the var. integra. Said to be poisonous. Cichorium Intybus L. Chicory.

It has become widely naturalized in the north and west. When fed in large quantities to dairy cattle it imparts a bitter flavor to the milk and butter. It contains the bitter glucoside chicorin.

Iva xanthiifolia Nutt. Marsh Elder. Half-breed Weed.

Western Wisconsin, Red River Valley, south and west to the Rocky Mountains. Said to cause hay fever.

Ambrosia artemisiaefolia L. Common Ragweed.

The pollen of this plant is suspected of causing hay-fever.



Fig. 19k. Spiny Clotbur (Xanthium spinosum). Injurious in a mechanical way. Fig. 19l. Prickly Lettuce (Lactuca Scariola). Said to be poisonous. (Bentham).

Ambrosia trifida L. Great Ragweed.

The pollen of this species is said to produce an irritating action upon the mucous membrane.

Xanthium spinosum L. Clotbur.

Maine to Texas. It causes mechanical injuries.

Xanthium canadense Mill. Cocklebur.

Young seedlings of this plant are poisonous to horses. Several cases of poisoning to hogs have been reported in this state.

Xanthium strumarium L. Cocklebur.

Poisonous like the preceding. This species is not common in the state. Contains the glucoside xanthostrumarin.

Eupatorium perfoliatum L. Boneset.

Commonly found in low grounds and marshes. It is an emetic when given in large doses.

Eupatorium urticaefolium Reichard. White Snake-root.

Widely distributed in woods in this state. It is said to produce a disease known as milk fever. No reports of this kind of poisoning have come to us in Iowa. The Σ cannabinum contains the alkaloid eupatorin and the glucoside eupatorin.

Trilisa odoratissima (Walt.) Cass.

It is said to be injurious and has the odor of sweet clover.

Grindelia squarrosa (Pursh) Dunal. Tar Weed.

From Wisconsin, Iowa, westward and northward. Is said to be injurious. The G. lanceolata Walt., occurs from Tennessee to Texas. It has large heads and resinous viscid leaves.

Xanthisma texana D. C.

It is said to be poisonous and contains saponin. Southward.

Erigeron canadensis L. Horse Weed.

It is a widely distributed troublesome weed in the north. The physiological action of the drug obtained from this plant is to produce smarting of the eyes, soreness of the throat, and prostration.

Erigeron ramosus (Walt.) B. S. P. White Weed.

Common in meadows westward. The E. annuus of the same distribution is found also in meadows. Both are regarded with suspicion.

Baccharis halimifolia L. Salt Groundsel.

The European B. cordifolia is said to be poisonous. It occurs in salt marshes along the Atlantic sea coast.

Solidago canadensis L. Golden-rod.

It is widely distributed in the U. S., and is one of the most common of our golden-rods. The golden-rods are generally regarded as harmless plants, but in a few cases they are suspected of being poisonous. A disease of horses near Black River Falls, Wisconsin, was attributed to a golden-rod. Chesnut thinks the disease due to a rust on the plant. As a general thing stock does not relish the golden-rod.

Madia glomerata Hooker. Tarweed.

A glandular viscid, heavily scented herb common from Saskatchewan to Colorado, Utah, Oregon and Washington. Probably poisonous or at any rate it is avoided by cattle.

Hemizonia macradenia DC. Tarweed.

Common on the Pacific Coast from San Francisco southward. A strongly and unpleasantly scented herb avoided by stock. Many of the species of the genus occur in California and most of them are strongly scented.

Aster Parryi Gray. Woody Aster.

Common in the Rocky Mountains, Utah and Wyoming. Thousands of sheep in Wyoming where this plant occurs suffer with the disease which has been attributed to "grub in the head." Healthy sheep often die within a very short time after the first symptoms appear. This plant is affected with a fungus, Puccinia xylorrhizae, and Prof. Aven Nelson suggests in his account of this disease, "The chances are rather better that the suggested poisonous qualities are due to the fungus. Some other parasitic fungi have been proven poisonous

and we may well in the absence of evidence suspect this one." The aster is confined to Gumbo clay soil intermixed with gravel and soil that contains more or less alkali and other salts.

Dr. O. L. Prien is investigating the disease.



Fig. 19m. Fetid Marigold (Dysodia chrysonthemoides). Pungent odor said to be injurious. (Charlotte M. King). Fig. 19n. Dog Fennel (Anthemis cotula). Contains a pungent principle.

Rudbeckia lacinata L. Cone-flower.

In moist grounds throughout the north. Dr. Schaffner says it is supposed to be poisonous to sheep.

Bidens frondosa L. Black Beggar-ticks.

Common in the north. The downwardly barbed awns are irritating.

Coreopsis discoidea T. & G. Small Beggar-ticks.

Very common in the east. It is a local irritant.

Helenium autumnale L. Sneezeweed.

It is common in low grounds throughout Iowa and is used by the Indians to produce sneezing. The whole plant and flowers are poisonous to cattle and sheep.

Helenium tenuifolium Nutt. Sneezeweed.

Common in the south and said to be fatal to horses and mules. It imparts ats bitter flavor to milk.

Helenium Hoopesii A. Gray. Mountain Sneezeweed.

Common in the Rocky, the Uintah and Wasatch Mountains. Said to be injurious to sheep.

Achillea millefolium L. Yarrow.

It is used as a forage plant, but causes an irritating sensation of the membranes and much pain in the gastric and abdominal regions. It contains the glucoside achillein, an amorphous bitter substance, and the alkaloid moschatin.

Anthemis Cotula L. Mayweed.

Has a very disagreeable odor and causes blistering of the skin. The plant \bar{a} s carefully avoided by stock.

Anthemis arvensis L. Corn chamomile.

Occasionally escaped from cultivation. Seeds of this and other species contain HCN.

Dysodia chrysanthemoides Lag. Fetid Marigold.

Common in the west, Dak., Ia., Neb. to Mo., Tex. The leaf bracts and other parts of the plant are provided with large pellucid glands which produce the characteristic odor of the plant.

Tanacetum vulgare L. Common Tansy.

Introduced into many parts of the north. Many serious and a few fatal cases of poisoning are recorded by the use of tansy oil. The symptoms of poisoning are varied, convulsions, violent spasms, dilation of the pupils, frequent and feeble pulse. Eleven drachms of the oil in a girl produced death in three and one-half hours. The effect on animals is salivation, vomiting, dilation of the pupils, muscular twitchings, followed by chronic spasms, death appears to be caused by paralysis of the heart and lungs.

Artemisia biennis Willd. Biennial Wormwood.

Probably poisonous.

Artemisia tridentata Nutt.

Sage brush used as forage by sheep.

Artemisia Absinthium L. Common Wormwood.

Occasionally cultivated. The volatile oil of the plant is a violent, narcotic poison, and contains the glucoside absinthiin, the alkaloid abrotin, the bitter principle santonin.

Other species probably also poisonous.

Arnica cordifolia Hook. Arnica.

This species with yellow flowers is common in the Rocky Mountains. The bruised leaves give off the odor of arnica. The European A. montana is suspected.

Senecio Jacobaea L. Staggerwort.

Occasionally found in the east and causes the Pictou disease. The Squaw Weed (S. aureus) is common in the north. The S. plattensis, common in western Iowa to Montana and east to Ontario, has been associated by Dr. Day with the Missouri Bottom disease. The species are numerous in the Rocky Mountains and may be responsible for some diseases, It is to be noted that the S. guadalensis of Mexico is fatal to stock. The exotic S. toluccanus contains an alkaloid with tetanus like action.

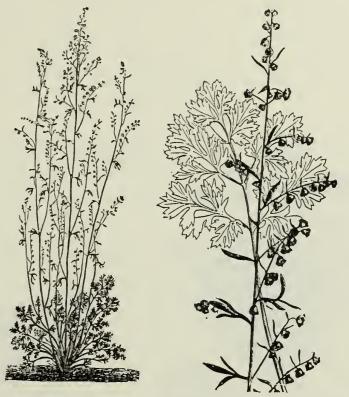


Fig. 190. Absinthium (Artemisia absinthium). Plant, leaves, and flowering branch. (Faguet).

Arctium Lappa L. Burdock.

Produces itching. Contains the alkaloid lappin. Common weed in the U. S.

Cirsium lanceolatum (L.) Hill. Bull Thistle.

Commonly naturalized in the north from the Atlantic to the Pacific. It acts injuriously in a mechanical way. Other species like C. Nelsoni, Canada Thistle (C. arvense), C. scariosum, etc., all act in a similar way. Some species contain HCN.

Centaurea solstitialis L. Knapp Weed.

Common in alfalfa meadows westward. Acts injuriously in a mechanical way. Some species, according to Greshoff, contains HCN.



Fig. 19p. Sneezeweed (Helenium tenuifolium). Poisonous. Fig. 19q. Bull Thistle. (Cirsium lanceolatum). Mechanical injuries.

CHAPTER XV

CHEMISTRY OF ALKALOIDS.

PROF. A. A. BENNETT

The term alkaloid is a relic of an early method of nomenclature, namely, the method of naming substances without reference to their fundamental properties. For example, the name, oil of vitriol, does not describe sulfuric acid except that it has a deceptive resemblance to an oil, and was originally produced from green vitriol. The word alkaloid literally signifies a substance resembling an alkali. They do form salt-like compounds with acids but here the likeness as to specific properties ends.

It is only about one hundred years since the facts as to alkaloids began to accumulate. The first separation of these compounds was made in 1803 by Derosne but their basic character was not noted until three years afterwards by Serturner while studying opium. Before this time many plant extracts were known to contain some very active compounds usually called principles, but their isolation and the determination of their composition and properties date from the first quarter of the nineteenth century.

The first alkaloid that was prepared and reported according to the usual method of procedure of the chemists, namely, obtaining the pure compound and then determining its chief properties, was morphin. Although this was done about 1806 ft was not until some cleven years afterwards that the report attracted the attention of chemists, sufficiently to start the investigation of other substances for the presence of similar compounds with the result that new compounds of this class have been separated and described each year since this date. The property that especially characterised these compounds was their basicity, i. e., they formed salt-like compounds with acids, although they were but weakly alkaline to the usual indicators of alkalinity.

COMPOSITION AND GENERAL PROPERTIES OF ALKALOIDS.

The alkaloids all contain nitrogen, carbon and hydrogen and all but two of them contain oxygen. A large proportion of the alkaloids are non-volatile, solid, crystalline compounds, while a few are volatile liquids but contain no oxygen. They are generally insoluble in water but are generally soluble in alcohol and possess varying degress of solubility in ether, chloroform, amylic alcohol and carbon disulfid. These latter facts of solubility are often made use of to separate the alkaloids from each other, and from other substances. The salt-like compounds, on the other hand, generally possess a measurable degree of water solubility but not in the other solvents mentioned. They differ from the true alkalis, like potassium and sodium hydroxids, in the fact that the molecules of acids and alkaloids unite with each other without forming other products. In this property, they resemble ammonia, HN₃, when it forms such salts as ammonium chlorid, NH₃, HC1 or NH₄C1. However, this property does not characteristically belong to the so-called true alkaloids, since the amins, put in bases,

and ptomains react in a similar manner. The volatile alkaloids, represented by nicotin and coniin, possess disagreeable odors suggesting in part that of ammonia. The solid alkaloids, represented by strychnin, morphin, are odorless, but have a bitter taste, often very characteristically bitter.

Physiologically, the alkaloid is quite generally a very active intoxicant, acting directly on the nervous tissues and producing results all the way from the atonic to the strongly tonic effects. The extreme effects are seen in the muscular excitation of strychnin and the depressing action of morphin, or, in the heart stimulation of atropin, and the depressing effects of cocain.

OCCURENCE OF THE ALKALOIDS.

The alkaloids are peculiarly a plant product and probably may be regarded as a protective agency to preserve a given species of plants. They are deposited in various parts of the plant but commonly in the seeds. The seeds containing, as they do, the vital parts of annual plants, (and also perennials) are protected from destruction by micro-organisms and by animals using them for food, by the intoxicating property, just referred to. A similar case is that of the glucoside amygdalin, as found in the seeds of several of the Rosaceae. The enzyme present in the seed, under proper conditions of temperature and moisture, decomposes it into glucose benzaldehyde and hydrogen cyanid or prussic acid, this latter compound acting as the intoxicant.

The alkaloids are not widely distributed in the plant world, although they are found in several orders or families of plants. Three families are especially characterized by the presence of alkaloids, namely, the poppy family or Papaveraceae, night shade family, Solanaceae, and the Rubiaceae.

The basic property of the alkaloids suggests the probability of their occurrence in combination with acidic compounds more or less characteristic of the plants in which they are found. In some cases the so-called alkaloid appears to be similar to glucoside, i. e. it can be hydrolyzed. For example, cocain can be hydrolyzed into ecgonin, $C_9H_{15}NO_3$, benzoic acid, $C_0H_5CO_2H$ and methyl alcohol. Others are real glucosides like digitalin and solanin. It is undoubtedly true that the latter two should not be classed with true alkaloids, but with the glucosides, and are like caffein theobromin in this sense, i. e., that they are substances that have been classed with the alkaloids on superficial grounds, such as bitter taste, but really have no close chemical relation with them. Caffein and theobromin are now known to belong with the purin compounds. The purin compounds are basic, and hence their classification with the alkaloids.

The acids with which the alkaloids are often united are somewhat common in plants, or in a few cases they are characteristically found in combination with the given alkaloid, e. g., meconic acid in combination with morphin in opium, or aconitic acid united with aconitin. Other acids form compounds with the alkaloids in the various plants in which they are found. Among these acids are tannic, citric, malic and quinic. The combinations are easily broken up by strong bases, like potassium or sodium hydroxid, and in this way they may be separated from the acids. Since the true alkaloids are generally soluble in water while alkali salts of the acids are not soluble, the alkaloids may be separated by treatment with potassium or sodium hydroxid and filtering out the insoluble alkaloid. They are then further purified by the formation of the soluble salt and re-precipitating the alkaloid by an alkali. The alkaloid is then dissolved by the appropriate solvent and crystallized. Again it is interesting

to note that an alkaloid rarely exists alone in a given plant, but is accompanied by several others. For example, aconitin, as extracted from the roots of the aconite plant, Aconitum napellus, contains nine alkaloids; in the extract from poppies, called opium, upwards of seventeen alkaloids have been separated and studied; in the so-called chinchona or Peruvian bark extracts, some thirty-three distinct alkaloids have been isolated; strychnin is accompanied by brucin in Strychnos ignatii, or Saint Ignatius bean.

CLASSIFICATION OF THE ALKALOIDS.

Owing to the basic character of the alkaloids, and the fact that they always contain nitrogen, it was suggested that they were connected with ammonia in some manner, and if so, that they might be readily broken down by distilling them with potassium hydroxid or caustic potash. They were regarded as derived or at least connected with ammonia. Hoffman, who added much to our knowledge of the amin compounds, considered that they were of the ammonia type and were tertiary amins. In attempting to find some reaction characteristic of the amins, Gerhardt and others, heated the alkaloids with caustic potash, but were unsuccessful in obtaining any results that showed that their basic character was due to this structural cause, although some of them do possess some properties resembling the amins. Others more closely resemble the ammonium compounds. Products obtained by heating some of the alkaloids with potassium hydroxid and distilling the volatile products were found to be the same as were obtained from the destructive distillation of bones. Later, 1834, a study of bone oil by Runge led to the separation of a pure compound which was shown to have the formula C5H5N, known as pyridin. This was later shown to be a cyclic (hetero-cyclic) compound, like benzene, one group "CH" being substituted by nitrogen, i. e., trivalent nitrogen. A number of alkaloids have been shown to be constructed on this nucleus by substituting various hydrocarbon groups and are known as the pyridin alkaloids.

Among the pyridin alkaloids and derivatives from them are nicotin, coniin, atropin, cocain. The study of the alkaloids by noting the action of potassium hydroxid proved to be a fruitful one. In 1842, Gerhardt obtained a compound from the destructive distillation of quinine that was a new substance. This was named quinolin because of its origin from this alkaloid. It was later shown to be structurally composed of a benzene and a pyridin nucleus joined by two atoms of carbon in common; the further complexity is due to substituting in this nucleus. Most of the alkaloids are esters and are consequently quite readily separated into the two parts of such compounds namely the acid and basic parts. A study of these constituent parts gives the complete facts as to the structure of the original compound. The esters may be decomposed by acids, alkalis, and water.

The nucleus quinolin has been found in quinin, cinchonin, cinchonidin, strychnin, and brucin, and hence these compounds are known as the quinolin alkaloids.

THE ISO-QUINOLIN GROUP.

This group of alkaloids have a nucleus isomeric with quinolin known as the quinolin group. Like the quinolin group, iso-quinolin has the empirical formula, C_9H_7N . The difference between this base and quinolin, so far as structural constitution is concerned, appears in the position of the atom of nitrogen. Not

only is the fundamental nucleus different in the two classes of alkaloids, but groups that enter into these nuclei are different, thus leading to a large number of possible compounds. To this group of alkaloids belong especially the alkaloids of opium. For example, morphin, thebain, narcotin, narcein, papaverin, and codein. There are also hydrastin, hydrastium and berberin.

THE PURIN GROUP OF ALKALOIDS.

These compounds are not properly included with the alkaloids but they belong with the purin compounds or xanthin bases. The nucleus (purin) or atomic framework, shown by Fischer to characterize the xanthin bases, such as uric acid, the xanthin derivatives, guanin and adenin, is found in caffein, theobromin, theo-phyllin, compounds that are yet classed with the alkaloids. The structural and mutual relationship of these to each other and to the xanthin bases has been determined synthetically and hence their classification is not a doubtful question. The alkaloids are methyl xanthins. Caffein is a tri-methyl xanthin and theo-bromin and theo-phyllin are di-methyl xanthins, the latter being an isomer of theo-bromin. The alkaloid of tea is sometimes called thein although its identity with the caffein of coffee has been recognized for a long time.

There are several alkaloids whose structural relations have not been determined. Among these are pilocarpin, colchicin, and physostigmin. For the description of these alkaloids see statement in this text under their appropriate heads.

PHYSIOLOGICAL ACTION OF ALKALOIDS AS DETERMINED BY THEIR STRUCTURAL COMPOSITION.

It is within comparatively recent time that pure alkaloids have been prepared and consequently that their physiological effects could be determined. Some of these like quinin were prepared in a fairly pure condition during the first half of the last century but most of them belong to a later period. Attention has been called to the fact that a single alkaloid is rarely produced by the plant and hence in the usual extracts from the plant there are several alkaloids with varying physiological effects. These constitutents vary in amount according to a variety of conditions under which the plant may produce them. Acordingly the extracts of such plants will vary in the proportion of the alkaloid present. For example, in cinchona bark the amount of quinin may vary from 2 to 13 percent. The physiological effect of the cinchona extracts will be markedly different in such extremes of composition, i. e. in reference to the alkaloid quinin.

In 1869, Crum, Brown, and Frazer called attention to the relation of the structure of organic compounds to their physiological effects. They were studying the comparative action of strychnin and brucin and although their knowledge of the structure of these two compounds was not complete yet they were able to trace a relation in this case. They found that the presence of methyl or ethyl groups strongly affected the active properties of these compounds. For example, brucin is regarded as the dimethoxy-deriative of strychnin. The discovery that most alkaloids are built about three nuclei, namely pyridin, quinolin, and iso-quinolin added new zest to the study of the physiological effects of certain organic groups when substituted in organic compounds.

Quinolin is a strong antipyretic and antiseptic but produces other results

that are decidedly toxic. By introducing the organic group, methoxy CH,O, a compound is produced from which several derivatives have been formed, which possess antipyretic properties that have led to their use in medicine. These are known as analgen, kairin, kairolin, and thallin. Quinin differs from cinchonin in containing the methoxy-group in place of a hydrogen atom. This addition produces a much stronger antipyretic. The antipyretic effect of the methoxy group is shown in some of the derivatives of anilin. For example, anilin is a highly poisonous compound although it has good antipyretic properties. The addition of a similar group to the one in question produces a compound known as acetanilid or antifebrin. The group added, CH, CO, is known as acetyl. In introducing the methyl group into acetanilid its antipyretic properties are somewhat reduced but its anti-neuralgic properties are increased. The addition of hydrogen to quinolin increases its toxic properties while the hydroxyl group increases the antifebrin results. The antipyretic properties of antipyrin are probably due to the two methyl groups present in the molecule. The ethyl group when added to organic compounds that are poisonous, in many cases at least, decrease their toxic character. The same is true of the acid or carboxyl group. When morphin is heated with hydrochloric acid, water is formed, as in hydrolysis, and a new compound known as apo-morphin. This treatment has developed two hydroxyl groups. This new compound is an excellent emetic The introduction of two acetic acid groups changes morphin into a mildly acting sedative known as heröin. Finally the purin alkaloids owe their properties to the methyl groups introduced into xanthin i. e. as distinct from the zanthins.

The accumulation of facts in this direction is increasing rapidly and some generalization from them will do for medicine what antisepticism and anesthetics have done for surgery, and change it from an empirical to a scientific basis.

Blyth in his work on "Poisons; Their Effects and Detection" has classified the alkaloids and other organic poisons into the following groups.

ALKALOIDS.

First group, liquid volatile alkaloids, under which there are grouped the alkaloids of Conium (coniin, conhydrin), tobacco (nicotin), piturie from Duboisia Hopwoodii belonging to the same family as tobacco, and spartein from the Common Broom (Spartium scoparium).

Second group, the opium group, contains the alkaloids from the Poppy, the more important being morphin, thebain, codein, of the Morphin group; and papaveramin, codamin, laudanin, narcotin, papaverin of the Papaverin group.

Third group, the strychnin or tetanus-producing group of alkaloids. Under this head are included the alkaloids strychnin and brucin, derived from Strychnos Nux vomica of the family Loganiaceae; the alkaloids aspidospermin, quebracho obtained from the Querbracho (Aspidosperma quebracho blanco) of the family Apocynaceae, and at least four others; the alkaloids from the Pereira bark Geissospernum Vellosii, pereirin; the alkaloids from Gelsemium or Carolina Jessamine (Gelsemium sempervirens) of the family Loganiceae, gelsemin and gelseminin; the cocaine alkaloids obtained from Erythroxylon coca, cocain hygrin, etc.; the alkaloids from the roots of the European Corydalis cava, corydalin, corybulbin, and six others. The corydalin in large doses causes epileptiform convulsions.

Fourth group, alkaloids aconitin, atisin, and japaconitin of the aconite group, obtained from several species of Aconitum, as Aconitum Napellus, A. heterophyllum, etc.

Fifth group, the mydriatic group of alkaloids. The alkaloids of this group are atropin, obtained from Atropa, Belladonna, and Datura; hyoscyamin obtained from Datura, Hyoscyamus, Scopolia carniolica, and Duboisia (Hyoscin pseudohyoscyamin being also obtained from these plants); scopolamin from some of the same plants as the preceding; solanin which is, however, regarded as a nitrogenized glucoside, obtained from various species of Solanum; solanidin with stronger basic properties than solanin, obtained from plants of the same family; cytisin obtained from Laburnum (Cytisus Laburnum) of the family Leguminosae, found also in quite a number of other plants of the same family.

Sixth group, the Veratrum alkaloids containing the alkaloids jervin, pseudojervin cevadin, etc., obtained from various species of Veratrum as V. album, V. viride, etc.

Seventh group physostigmin, the most important alkaloid of the group derived from the Calabar Bean (*Physostigma venenosum*), and calabarin.

Eighth group, containing pilocarpin, obtained from the leaves of jaborandi (Pilocarpus pennatifolius) and four other alkaloids, jaborin, pilocarpin, isopilocarpin, pilocarpin. Jaborandi belongs to the family Rutaceae.

Ninth group, taxin, obtained from the yew tree (Taxus baccata).

Tenth group, the curare alkaloids which are obtained from the Curare plants (Strychnos toxifera and S. Castelnaei of the family Loganiaceae. The alkaloids are tubo-curarin, curin, etc. Protocurin obtained from the latter species is a slightly toxic substance.

Eleventh group, colchicin alkaloid; this alkaloid is obtained from the seeds and roots of the common meadow-saffron or Colchicum (Colchicum autumnale).

Twelfth group, muscarin from the Amanita Muscaria or Fly agaric.

GLUCOSIDES.

The glucosides widely distributed in plants are compounds of glucose and organic acids and are certainly of great importance in connection with the poisonous principles found in plants. They have been grouped by Blyth into: A first group the digitalis group, consisting of digitalin, digitonin, and digitogenin, all found in the common fox glove (Digitalis purpurea); (2) second group of glucosides acting on the heart and containing antiarin obtained from Antiaris toxicaria, the upas tree; the helleborin and helleboretin found in Helleborus niger, H. viridis, H. foetidus and euonymin, a resinous substance found in Wahoo (Euonymus atropurpureus) which is a powerful heart poison; the third group containing thevetin, obtained from Thevetia neriifolia; strophantin from Strophanthus hispidus of the Dogbane Family belong to this group of heart poisons, but it is not a glucoside and only partly crystallizable; scillain from squill; adonidin from the root of Adonis vernalis of the Crowfoot Family; oleandrin from the Oleander; neriin also from the Oleander, sometimes called the Oleander digitalin and the poison of the Madagascar Ordeal plant (Tanghinia venenifera). The fourth group contains the digitalin-like apocynin from the common Dogbane, and other Apocynums; erythrophlein, convallamarin, a glucoside from the Lily-of-the-valley; coronillin from Coronilla; and cheiranthin from Cheranthus. These behave like the Digitalins.

GLUCO-ALKALOIDS.

The gluco-alkaloids represent a class of compounds intermediate in connection between the alkaloids and glucosides. The achillein found in Yarrow (Achillea millefolium) and solanin in various species of Solanum should be mentioned. The latter substance has, however, been referred to in connection with the alkaloids.

SAPONINS.

The Saponins have been treated fully in another connection. They are poisonous and when dissolved in water form solutions which froth. Of these mention may be made of saponin and senegin.

OTHER VEGETABLE POISONS.

A third division of poisonous substances includes those which cannot be readily classified and under this head is santonin, a lactone found in the heads of Artemisia sp. A second division of this group is mezereon obtained from Daphne Mezereum. A third group is ergot of rye, containing ecbolin, secalintoxin, and other substances referred to at length in another connection.

PICROTOXIN, CICUTOXIN AND TOXINS.

Picrotoxin is the active principle of the Indian Berry (Cocculus indicus or Menispermum cocculus), which contains the active principle picrotoxinin, picrotin and menispermin.

Tutin, a non-nitrogenous glucoside, is obtained from Coriaria sarmentosa and other species. Another poison belonging to the picrotoxin class has been isolated from the Japanese Illicium anisatum, a member of the Magnolia Family. The plant is sometimes called the Japanese Star Anise. To this group may also be added cicutoxin obtained from the Cowbane, the oil of savin obtained from the common savin (Juniperus sabina); croton oil expressed from the seed of Croton tiglium; the toxalbumins of castor oil seed and of Abrus; ictrogen from various species of lupines (Lupinus luteus, L. hirsutus, etc.); the toxic substances in the cotton seeds; toxic substances in various species of Lathyrus; the toxic substances in Arum; in the black bryony (Tamus communis); the toxalbumin of the black locust; and the poisonous substances of the male shield fern.

ANIMAL AND PLANT TOXINS.

Another group of poisonous substances is included under the head of ptomaines and animal toxins. The word ptomaine is used in a rather indefinite way, and is open to objection, but the classification given by Blyth as animal toxin is also objectionable. Many of these toxins are the products of bacteria, some of these poisons are, however, the products of higher plants, (as toxin of the black locust, Abrus), etc. The groups given by Blyth are the Amins, under which head we have methylamin, found in the cultures of the Comma bacillus, and the trimethylamin, non-toxic, found in a variety of putrefying substances.

AMINS.

The Amins are basic and originate from ammonia; they include the diamins, belonging to the amin series, which are formed in putrefactive substances. Of these we have neuridin in putrefying substances; cadaverin, found

in cultures of Spirillum and putrid animal matter; mydalein, guanidin. The cholin group includes neuridin, betain, and muscarin. The muscarin has been referred to elsewhere. The neurin is intensely poisonous, and atropin is an antidote to neurin. Tetanin produces tetanus. Tetanotoxin from tetanus produces tremor paralysis and violent convulsions. Mydatoxin contained in putrid horse flesh is poisonous in large doses, causing lachrymation, diarrhoea, and convulsions. Tyrotoxicon, isolated by Vaughan from milk, is toxic. The susotoxin isolated from hog cholera is said to be quite toxic.

ORGANIC ACIDS.

The last group of poisonous substances includes the organic acids, the most important of which is oxalic acid. This is widely distributed both in the free state and in combination with lime soda and potash in the vegetable kingdom. It occurs in some species of the geranium, spinach, Phytolacca decandra, pie plant, Rumex Acetosa and in Atropa Belladonna, in connection with potash. In Russian Thistle and Salicornia it occurs in combination with sodium. In clover, apple twigs, begonia, and many other plants it occurs in the form of so-called compound aggregate crystals, or rosettes of calcium oxalate, in the onion and some other plants of simple crystals. In aroids and Virginia Creeper it appears in the form of needle shaped crystals, known as raphides, which are formed during the metabolism of the plant, the oxalic acid being set free and uniting with the lime in the plant to form calcium oxalate.

Oxalic acid is commonly used by dyers and calico-printers and also by curriers and harness makers for cleaning leather, to remove iron stains, to bleach straw, etc.

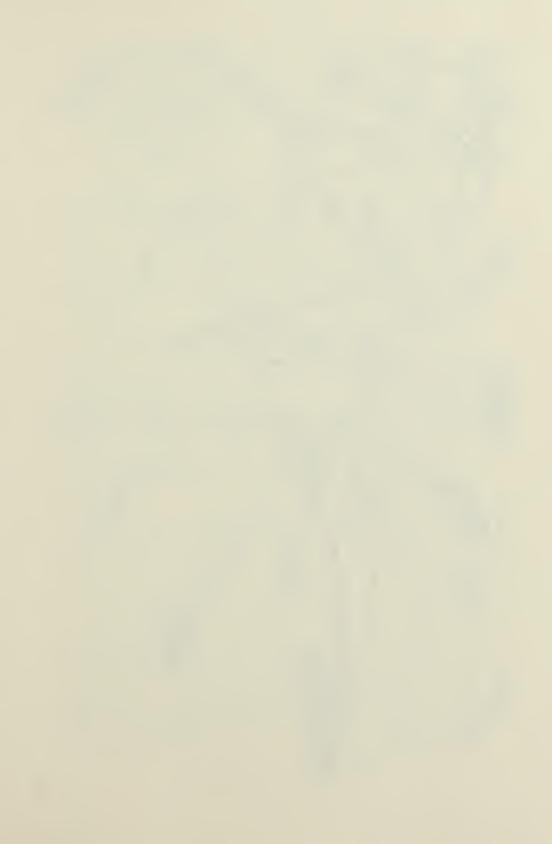
Several cases of poisoning have been attributed to the use of plants (like the sheep sorrel, oxalis, etc.), that contain large amounts of oxalates.

Quite a number of cases of poisoning from this acid are reported, especially in Europe. The smallest dose of oxalic acid known to have destroyed life, according to Dr. Taylor, is 60 grains. Oxalic acid acts upon the central nervous system. There is temporary loss of voice, burning in the throat, burning in the stomach, vomiting, especially bloody matter, pulse weak, locally it acts on the mucous tissues.

Part II

A Descriptive Manual of the Poisonous Plants of North America Chiefly of the United States







Cowbane-Very Poisonous. (After Vasey)

KEY FOR THE PLANT KINGDOM

Organisms without chlorophyll, the vegetative body a naked mass of protoplasm with many nuclei; reproduction asexual, spores free or enclosed in sporangia; spores produce motile swarm spores or amoeboid bodies.

A. Myxothallophyta. 158

Cells generally with cell membrane, with one or more generations; sexual reproductions frequently absent, the fertilized spores when present with one cell, which later separates from the mother plant, or a several celled body resulting from the fertilization of the female fructifying body, which later develops into a new plant.

B. Euthallophyta. 160

Small unicellular organisms, never green but frequently of other colors, blue greens, etc.; reproduction asexual by fission; spores formed in the interior of the cell or by transformation of vegetative cells into the endospores or arthrospores; nuclei absent, but a so-called "Central body" occasionally present; coloring matter equally distributed.

I. Schizophyta. 160

Unicellular organisms generally colorless at least never green; membrane, consisting of a chitin-like substance, occasionally gelatinous; cells frequently with cilia; reproduction by fission; spores when present endospores or arthrospores.

Schizomycetes. 161

Unicellular organisms like the preceding, cells contain chlorophyll and phycocyanin consisting of blue, blue-green, violet, or reddish pigments; swarm spores absent.

Schizophyceae. 184

One-celled organisms with nucleus sharply differentiated; protoplasmic body with a simple denser protoplasmic membrane, pseudopodia absent, motile during most of their existence; cilia 1 or more, and with 1 or 2 pulsating vacuoles; chromatophores occasionally absent; reproduction asexual by longitudinal division.

II. Flagellatae. 188

Plants occurring mostly in water, always with cell-membrane and nucleus; green or other colors (brown or red) mixed with the green.

III. Euphyceae, 188

Small unicellular organisms occasionally forming chain-like colonies. Cells possess two long cilia which arise from a furrow in the ventral surface. Found mostly in the plankton of salt water.

*Peridinales. 188**

Small one-celled organisms of brown color, the chlorophyll masked by diatomin found in chromatophores; cell-wall consisting of silica with a girdle and fine lines; reproduction asexual; division parallel to the long axis of the organism, and the formation of auxospores and sexual, by the formation of auxospores by conjugation.

Bacillariales. 188

Chlorophyll green algae; membrane without silica; reproduction by division,

swarm cells absent; sexual reproduction by zygospores through the union of two equivalent gametes. (Aplanogametes).

Conjugatae. 189

Chlorophyll green plants, occurring singly or in colonies (threads or flattened bodies); one or more nuclei, reproduction asexual by producing multilateral zoospores and nonmotile aplanospores; sexual by the copulation of zoogametes, or spermatozoids and oospheres; the spores produce a new plant directly or generally produce swarm spores.

Chlorophyceae. 190

Plants of brackish or fresh water, consisting of internodes, short whorls of cylindrical branches, cells nucleated; growth from an apical cell; asexual reproduction by means of bulbils and vegetative threads; sexual reproduction by egg-cells and spermatozoids, the latter are spirally coiled in the cells of the antheridium; the egg cell is contained in a spirally coiled structure and after fertilization becomes an oospore.

Charales. 193

Brown algae, chlorophyll masked by a brown coloring matter, phycophaein; reproduction sexual and asexual, swarm spores, sperm cells and egg cells; marine; tetragonidia absent.

Phaeophyceae. 194

Brown algae; reproductive bodies without motion; tetragonidia present.

Dictyotales. 194

Red or violet algae; chromatophores contain chlorophyll and a red coloring matter (phycocrythrin and rhodophyll); reproduction sexual and asexual; mostly marine.

Rhodophyccae. 194

Parasitic or saprophytic plants with one or more cells, chlorophyll absent, with apical growth; mycelium usually evident; reproduction sexual and asexual, generally the latter; asexual by the formation of zoospores, conidia or spores.

IV. Eumycetes. 195

The vegetative body mostly 1-celled, tubular, asexual by the formation of spores or endospores sexual by the formation of zygospores.

Phycomycetes. 195

Copious nonseptate branched mycelium, asexual reproduction by endospores or chlamydospores; sexual by zygospores. Zygomyceles. 195

Mycelium occasionally sparingly developed, tubular, asexual; reproduction by swarm spores or conidia; sexual by the formation of oospores.

Oomycetes. 204

Mycelium, many celled; reproduction asexual or sexual by union of nuclei; conidia borne on basidia, number various.

Basidiomycetes. 209

Mycelium many celled; reproduction sexual and asexual; the latter by conidia; pycnidia and spermogonia with spermatia; sexual spores in sacs known as asci; spores called ascospores.

Asconycetes. 247

Fungi whose spores are not in sacs, or consist of sterile mycelium. Forms like Oidium, Ozonium, or Mycorrhiza. Fungi Imperfecti. 281

Organisms consisting of a fungus and an alga. Spores either in sacs (Ascolichenes) or borne like toadstools (Hymenolichenes). Lichenes. 307

Plants with stem, root and leaf; cormophyte or in some cases thalloid. Two generations, gametophyte and sporophyte; antheridium with sperm cells; tube cell absent.

C. Embryophyta Zoidiogama. 308

Many celled differentiated structure frequently with leaves and stem or

thalloid in some cases. Male (antheridium) and female (archegouium) organs are produced. Asexual spores in spore cases which open at the top, in true mosses.

I. Bryophyta. 308

Spores alike or unlike, microspores and macrospores developing into flat or irregular prothallia; these bear the reproductive organs, (antheridia and archegonia); flowers and seeds absent; usually a well developed vascular system.

II. Pteridophyta. 312

Plants with a microsporangium (anther) containing the microspores (pollen grains) which develop a tubular body, the prothallium (pollen tube) a macrosporangium (ovule) containing the macrospore (embryo-sac) which develops into a minute prothallium; this remains enclosed in the macrosporangium; after the fertilization of the egg cell in the macrospore a seed develops; plants with flowers and usually well developed tissues, the epidermis, parenchyma and vascular. (Embryophyta Siphonogama). D. Spermatophyta. 325

Ovules not enclosed in an ovary.

I. Gymnospermae. 325

Resinous trees or shrubs; wood with tracheids, tracheae usually absent; fruit a cone of dry or fleshy scales.

Coniferae. 327

Ovules enclosed in an ovary.

II. Angiospermae. 332

Embryo with 1 cotyledon; stem without distinction into pith, wood, and bark; endogenous; leaves usually parallel veined; flowers generally on the plan of 3.

1. Monocotyledonae. 332

Flowers generally small, unisexual, regular with persistent perianth; 6 or 3 stamens; carpels free or rarely united; fruit a berry, drupe or nut; embryo small; endosperm copious.

Principes. 369

Palm-like plants with palm-like leaves; flowers naked or with thick leaves of perianth; carpels 2 or 4 with 2 or 4 placentae.

Synanthae.

Mostly fleshy herbs or thalloid floating plants; inflorescence a fleshy spadix subtended by a spathe or naked or reduced to few or solitary flowers on the margin or back of a thalloid body.

Spathiflorae. 370

Herbs generally with narrow leaves; flowers usually complete, their parts mostly on the plan of 3; corolla regular or nearly so; ovary compound superior; endosperm mealy.

Farinosae. 372

Mostly herbs; flowers with a well developed perianth, usually regular and complete; usually on the plan of 3; ovary superior or inferior, compound; endosperm fleshy or horny.

Lilitforae. 374

Large herbs; flowers irregular; ovary inferior, compound; composed of several united carpels; seeds generally arillate, frequently with perisperm and endosperm.

Scitamineae. 390

Herbs, tropical species frequently epiphytes; flowers very irregular, or in one family regular, generally complete and perfect; parts of the perianth in 3's or 6's; ovary inferior, compound; seeds numerous; endosperm present or absent.

Microsperimae. 392**

Embryo usually with two cotyledons; stem usually with wood, pith and bark marked, usually exogenous; leaves mostly netted veined; flowers frequently on the plan of 5.

2. Dicotyledoneae. 395

Petals separate or distinct from each other or wanting; occasionally some

carpels united; in one division ovules with many megaspores. In most plants of the other division there is a single megaspore with synergidal and antipodal cells. (Choripetalae and Apetalae).

a. Archichlamydeae. 395

Tropical plants with monoecious flowers and branches with a longitudinal ridge in which the stomata are hidden.

Verticillatae. 395

Dicotyledonous herbs; petals and sepals absent; flowers small spicate with bractlets.

*Piperales. 396**

Trees or shrubs; flowers small, in catkins, imperfect; sepals and petals none; leaves simple; fruit a many-seeded capsule; seeds with tufts of hairs at one end.

Salicales. 396

Trees or shrubs; leaves simple; flowers small, monoecious or dioecious, in catkins; perianth absent; ovary 1-celled; style short; stigmas 2; endosperm none.

Myricales. 397

Woody plants with simple leaves; flowers, staminate spicate, pistillate solitary.

Balanopsidales.* 399

Shrubs or trees; flowers small, dioecious, in catkins; leaves simple, alternate, petioled, perianth absent in staminate flowers; pistillate, subtended by bractlets; ovary 1-celled; endosperm thin.

**Leitneriales.* 400*

Trees with alternate, pinnately-compound leaves; flowers monoecious, with bractlets, staminate in catkins, pistillate, solitary or several; ovule solitary, erect; fruit a drupe, indehiscent or dehiscent, with woody husk, seed large 2-4 lobed; endosperm none.

Juglandales. 400

Trees or shrubs; flowers small; calyx usually present; monoecious, or rarely dioecious, in catkins; pistillate flowers subtended by an involucre which becomes a bur or cup in fruit.

Fagales. 402

Shrubs, herbs or trees; calyx present but corolla absent; flowers small, not borne in catkins, monoecious, dioecious or polygamous; ovary 1-celled superior.

Urticales. 404*

Shrubs, trees or herbs with scattered leaves; flowers in spikes, racemes or panicles usually perfect; single carpel.

Proteales.* 415

Herbs or shrubs; generally parasitic; calyx present; corolla absent; flowers perfect, or imperfect; a single inferior ovary; fruit various. Santalales. 415

Generally vines or herbs; leaves cordate, or reniform; corolla absent; calyx inferior; tube wholly, or partly adnate to ovary; flowers perfect.

Aristolochiales, 416

Generally herbs, occasionally trees, shrubs or twining vines; leaves simple, mostly entire; flowers small, regular, perfect, dioecious, monoecious or polygamous; petals absent; stamens 2-9; filaments filiform or subulate; ovary superior one-celled; ovule solitary; fruit an achene; endosperm mealy.

Polygonales. 417

Generally herbs, occasionally shrubs; flowers perfect; corolla usually absent, when present polypetalous; calyx present; ovary superior; embryo coiled curved or annular; albumen present.

Centrospermae. 423

Herbs, shrubs or trees; calyx usually of separate sepals; corolla generally present, polypetalous; ovary superior; carpels many usually separate; stamens generally free and more numerous than sepals.

Ranales. 444

Generally herbs; flowers regular and perfect; petals generally separate; stamens free; ovary compound, superior; free from calyx. Rhocadales. 479

Carnivorous plants; flower seapose; corolla with separate petals or nearly so sepals generally distinct; stamens free; ovary compound superior.

Sarraceniales. 497

Herbs, shrubs or trees; usually with petals which are separate; stamens generally perigynous or epigynous; sepals generally united or confluent with receptacle which is concave; earpels 1 or more distinct or united into a compound ovary.

Rosales.* 498

Trees, shrubs, or herbs; usually with petals which are separate,, united in some or entirely wanting; sepals mostly distinct; stamens few or occasionally more than twice as many as the sepals; alternate or opposite with them; ovary compound, superior.

Geraniales. 574

Trees, herbs or shrubs; petals usually present and separate; sepals generally distinct; stamens opposite usually fewer than sepals or as many, occasionally more than twice as many; ovary superior, compound; ovules pendulous.

Sapindales. 604

Shrubs, small trees or occasionally vines; leaves generally alternate; flowers, small, regular; stamens as many as sepals or calyx lobes alternate or opposite with them; ovary compound superior; ovules erect.

Rhamnales. 620

Trees, shrubs or herbs; leaves simple, mostly alternate; flowers regular, usually perfect; sepals separate, or more or less united; petals separate, or wanting; stamens usually numerous; ovary superior, compound; disk inconspicuous or none.

Malvales. 621

Shrubs, trees or herbs; flowers generally complete, perfect, and regular or irregular in some; sepals distinct, or more or less united; petals almost always present and distinct; stamens usually numerous; ovary compound, superior; placentae mostly parietal.

Parietales. 627

Fleshy plants, leafless, or with small leaves, generally spiny; flowers mostly solitary, sessile, regular, perfect and showy; calyx tube adnate to ovary; limb many-lobed; petals numerous; stamens numerous; ovary 1-celled; ovules numerous; fruit a berry.

Opuntiales. 634

Shrubs, trees or herbs; leaves simple; petals usually present and distinet; calyx 4-5 lobed or entire and petals wanting in Thymeleales, superior or inferior; ovary 1 or more celled; ovules 1 or numerous. * Myrtiflorae. 637

Herbs, shrubs or trees with petals; leaves of calyx usually 5; stamens 4 to 5; ovary epigynous, adnate to calyx; ovule, 1 in each cavity.

Umbellales. 645

Petals partly or wholly united rarely separate or wanting; coherence variable in some cases; tubular or funnel-form. (Sympetalae or Gamopetalae).

b. Metachlamydeae. 664

Flowers complete, regular with lobed or distinct calyx; corolla cleft gamopetalous; stamens free from corolla; ovary compound. Ericales. 664

Mainly herbs; corolla gamopetalous; calyx generally free from ovary; stamens borne on corolla, as many as its lobes, or twice as many, or more.

Primulales, 675

Trees or shrubs; alternate, simple leaves; flowers generally regular; calyx free from ovary, inferior; corolla gamopetalous or polypetalous; stamens borne on tube, at base of corolla.

Ebenales. 679

Trees, shrubs, herbs or vines, generally with opposite leaves; flowers regular; corolla generally gamopetalous, or rarely polypetalous or wanting; stamens borne mostly in lower part of corolla, as many as lobes or fewer, alternate; ovaries 2 and distinct.

Contortae. 670

Rarely trees, shrubs, generally herbs; corolla nearly always gamopetalous, regular or irregular; stamens adnate to corolla tube; ovary one, superior compound.

Tubiflorae. 698

Herbs frequently acaulescent or caulescent with opposite or alternate leaves; flowers small, perfect, polygamous or monoecious; calyx 4-parted; corolla free; stamens 2 or only 1; ovary sessile, superior, 1-2-celled or falsely more celled; fruit a pyxis.

Plantaginales. 739

Plants with gamopetalous corolla; stamens as many as corolla lobes; and alternate with them, or occasionally fewer, or twice as many; ovary compound inferior, adnate to calyx tube or ovary 1 or more celled; ovules 1 or more in each cavity of ovary; leaves opposite or verticillate.

Rubiales. 740

Herbs or rarely shrubs with gamopetalous corolla or occasionally petals separate; stamens as many as corolla lobes or fewer; anthers generally united; ovary inferior.

Campanulatae. 748*

MYXOTHALLOPHYTA

Fungus-like organisms without chlorphyll, regarded by some as animals; intermediate, in some respects, between animals and plants and hence called Mycetozoa by Rostafinski. In their vegetative condition, they consist of naked masses of protoplasm with many nuclei, the mass of protoplasm being called the plasmodium which creeps about on the substrata changing in form and thrusting out processes called pseudopodia which may later coalesce. After

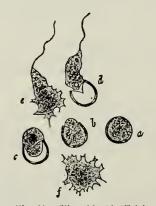


Fig. 20. Slime Mould (*Trichia varia*). a. Before germination. b, c, d. Different stages in germination. d, e. Amoeboid body with flagellum. After DeBary.

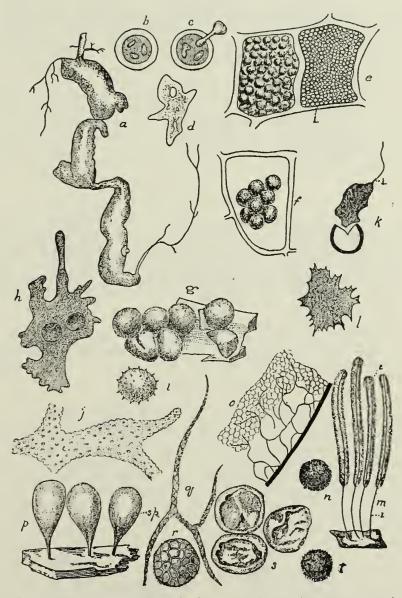


Fig. 18. Various slime moulds. a-f. Club root of Cabbage; Plasmodiphora Brassicae: a. Swollen root; b. Spore; c. Spore germinating; d. Plasmodium; e. Cells showing aggregated masses; f. Spores in cells; g. Lycogola epidendron; h, j. Plasmodium with branches; i. Spore; k. Spore germinating showing cilium; m. Stemonitis; 1. Stipe; 2. Columella; o. Capillitium; p. Trichia decipiens; sp. Sporangia; q. Elater; r. Spore.

a shorter or longer period the protoplasm contracts forming little heaps which contain the spores; the parts of the reproductive body are called the *sporangium* or spore case, the *peridium* or the wall of the case, the *stipe* or stalk, the *columella* or central axis in the spore case, the *capillitium* or fine threads, and the *spores*. The spores after absorbing water, germinate by breaking the wall and move about by means of cilia; sexual reproduction is entirely absent.

The division Myxothallophyta includes three classes: Acrasieue without swarm cells; Plasmodiophorales of which the club root of Cabbage, Plasmodiophora Brassicae is an example (a very destructive parasite upon cabbage, turnip, etc., in Europe and the Eastern States); and Myxogasteres which contains a great many species and genera common on spent tan bark, rotten logs, and the ground. Of the third class Stemonitis, Physarum, Lycogola and Fuligo are common genera. No species of this class is poisonous so far as known.

EUTHALLOPHYTA

Cells generally with cell membrane, with one or more generations, sexual reproduction frequently absent, the fertilized spores when present, with 1 cell which later separates from the mother plant, or a several-celled body resulting from the fertilization of the female fructifying body, which later develops into a new plant. This division includes such plants as bacteria, blue green algae, the green algae, rusts, smuts, mildews, moulds, puffballs, mushrooms and toad-stools.

SCHIZOPHYTA

Small unicellular organisms, never green but frequently of other colors, blue greens, etc., reproduction asexual by fission, spores formed in the interior

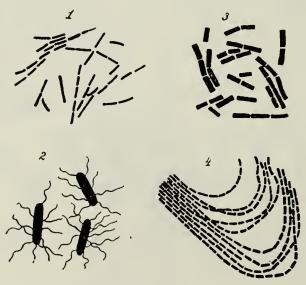


Fig. 19. Schizophyta. Schizomyceles Bacteria. 1 and 2. Bacillus subtilis 3 and 4. Bacillus anthracis. 1, 3 and 4x1000. 1, 3, and 4 after Fränkel and Pfeiffer. 2 after Migula.

of the cell by the transformation of vegetative cells into endospores, or by the transformation of ordinary vegetative cells into arthrospores; nuclei absent, but a so-called "central body" occasionally present; coloring matter equally distributed. This sub-division includes the Bacteria or Schizomycetes and the Blue-green Algae or Schizophyceae.

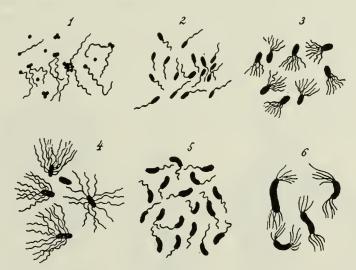


Fig. 20. Schizophyta. Shizomycetes. Bacteria with flagellae. 1. Planococcus citrus 2. Pseudamonas pyocyanea. 3. Pseudamonas syncyanea. 4. Bacillus typhi. 5. Spirillum comma. 6. Spirillum rubrum. Fig. 1-6x1000; all after Migula.

SCHIZOMYCETES

Schizomycetes is one of the two classes of the sub-division Schizophyta. All the members of this sub-division are characterized by having no known sexual method of reproduction, multiplying by means of simple fission or cell division. The bacteria are distinguished from the first, or Schizophyceae, by the absence of the blue-green coloring matter which is characteristic of these forms. The two sub-divisions approach each other very closely at some points, particularly among the branched bacteria. The shape of the bacteria is used as the character in the separation of the families. They are either rod-shaped, and unbranched, spherical, bent, or spiral and straight and branched, and with or without sheathing, covering, or membrane. Five families are distinguished by Migula. Some of these contain considerable numbers of bacteria important from their toxigenic properties.

Bacteria are among the smallest of living beings, some undoubtedly being so small that they cannot be seen with the highest powers of the microscope. Others are large enough so that they may be seen as minute specks by the naked eye. In other words, they vary from less than $1/10~\mu$ to $100~\mu$. They may be arranged in the case of the rod-shaped forms or bacilli, either singly or in chains. The same is true of the spirilla, or spiral forms. The cocci or spherical forms may be single, in pairs, in regular mass of 4 and multiples of 4, in chains, in irregular clusters, or imbedded in gelatinous mass forming zoogloeae. Multi-

plication, as before stated, is by simple fission or cell division. Growth takes place very rapidly in many forms, some being capable of growing to their full size and dividing to form two individuals in twenty minutes to half an hour. Under favorable conditions, this rapidity of multiplication explains the importance of the results obtained, and the products formed from such small plants. Some forms are capable of moving or swimming about by means of whips or flagella placed on all sides or simply at the ends. Other forms move by a sinuous or snake-like bending of the body. Many forms are incapable of motion. Spores are formed by many species; they are called endospores when formed singly within the bacterial cell, and arthrospores when formed by increase in size of the cells of the filament or part of a filament and its splitting into a

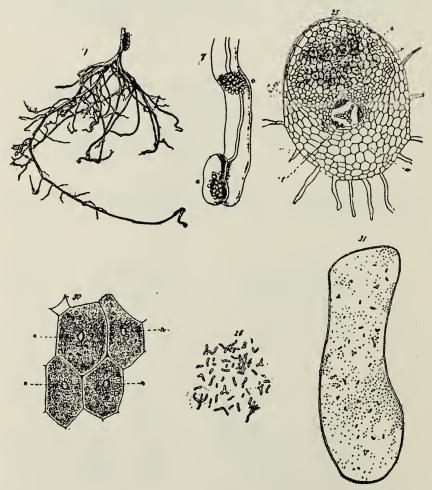


Fig. 21. The root tubercle organism ($Rhizobium\ leguminosarum$). 1. General view of root showing tubercles. 7. Root hair and strand with enlargements at a and e. 25. Cross-section of root at b bacterial tissue. 30. Cells of clover plant filled with the organism nucleus at n, 26. Rod and y shaped organisms from 30 more enlarged. 31. Single cell containing bacterioids. After Frank.

number of small cells. These spores serve, on account of their great resistance to dessication, and other unfavorable conditions, to tide the organism over until suitable conditions once more obtain. Bacteria are universally distributed, abounding in the soil, in the water, and being present often in the air, except at high altitudes. Normally, they are absent from the tissues of living animals and plants, but are to be looked for practically everywhere else. Their food requirements are as various as their habitats. Some require the most complex organic compounds, while others cannot live in the presence of such, but manufacture their own food from inorganic substances. Most bacteria lie between these two extremes. In respiration, some bacteria require oxygen or air, others will not develop in its presence. Most species require an abundance of moisture for their development, but many species will withstand a considerable amount of drying. Light inhibits the growth and in many cases destroys the bacteria. As to heat requirements, some live only in hot water, others will develop upon the surface of ice, some best at blood heat, while most develop between 15° and 22° C.

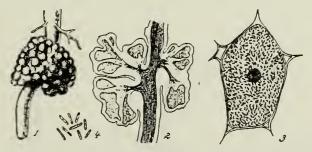


Fig. 22. Schizophyta. Schizomycetes. Nodule forming bacteria. Rhizobium leguminosarum. 1. Root tubercle of Lupine. 2. Cross-section of nodule. 3. Cell showing bacteria x 600. 4. Bacteria x 1500. After Woronin and Fischer.

Bacteria are also important in connection with the decomposition of organic matter. The nitrifying bacteria in the soil change the complex albuminous substances into nitric acid. This uniting with a base forms nitrates. The tubercle bacteria like Rhizobium leguminosarum are in mutual relation with clover and other leguminous plants and are important in the acquisition of nitrogen. Some bacteria play an important part in the dairy industry, the aroma and flavor of butter being due to these. Some, like the red milk organism (Bacillus prodigiosus), produce bad and disagreeable odors or cause the milk to become viscid or colored. Vinegar is produced by the acetic acid bacillus (Bacillus aceticus). Some bacteria produce diseases of plants like Fire blight of apples (Bacillus amylovorus), Cabbage rot (Pseudomonas campestris), Sorghum Blight, Corn wilt, etc. Some bacteria produce diseases of insects like Foul brood of bees, Silk worm disease, etc.

BACTERIA; POISONOUS PROPERTIES. It is believed best to consider in a general way, the various poisonous principles which are developed by bacteria before the discussion of the specific organisms and their specific poisons. Inasmuch as bacteria play a very important part in nature in breaking down dead tissues of all kinds, destroying them and returning them to their elements, or forming simple compounds, it is to be expected that among the multitude of chemical substances which are developed, there would be some which would be harmful

when taken into the body of man or animal. In fact, such decomposition products are known and as most of them, probably all, are basic, containing nitrogen, they have been grouped with that general class of vegetable alkaloidal substances called Ptomains. If other poisonous substances than Ptomains are developed, they are not known at the present time. In addition to these poisonous substances that arise as decomposition, other poisonous substances are produced by certain bacteria which are strictly synthetic, that is not produced by the breaking down of complex compounds into more complex forms. The exact chemical nature of these substances is not understood, the reasons for this being that they are extremely unstable, it being impossible to heat them without destroying, and they cannot be recognized by any known chemical means. They must be distinguished and differentiated, and often detected only by animal inoculation and experimentation. These soluble substances excreted by the bacteria are called toxins. The term toxin is rather an unfortunate choice, because it refers simply to their poisonous properties. In the broad sense, any poisonous substance is a toxin, but in the sense in which it will be here used, toxin indicates specific bacterial poisons excreted into the medium in which the

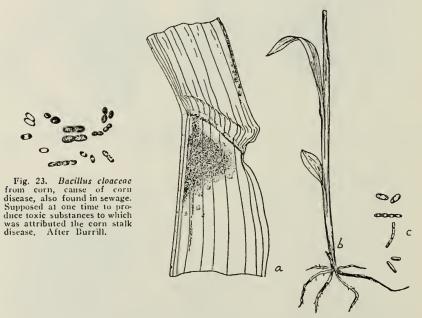


Fig. 24. Sorghum Blight (Bacillus Sorghi). b Young plant infected with the organism, a leaf and sheath. c Bacilli. Modified after Kellerman and Swingle.

organism is growing, and producing upon inoculation, anti-toxins. In addition to the products above mentioned, many bacteria undoubtedly owe their poisonous or intoxicating qualities to the fact that the protoplasm of living matter of the organism is poisonous or contains poisonous substances which are not excreted into the surrounding medium. When bacteria of this type are allowed to grow in favorable culture media for a considerable length of time, there is a certain amount of self-digestion or autolysis which takes place and these

poisonous contents of the cells are liberated and then go into solution. They may be liberated, also, by grinding the bacteria, and extracting with water. These poisonous protoplasmic substances have been called toxalbumins, but this term commits one to the supposition that all of this type of poisonous substances are proteid in nature. This has been by no means proven, consequently the term endotoxins is to be preferred.

The following terms used in discussions of immunity will need defining.

An Antitoxin is a substance capable of neutralizing a toxin by combining with it, and is produced in the animal body as a reaction to the introduction of a toxin in non lethal doses.

A Bacterial Agglutinin is a substance produced in the animal body as a reaction to the presence of certain bacteria or their products. When introduced into a suspension of the organism the agglutinin will cause the bacteria to clump (agglutinate) into groups.

A Bacteriolysin is a substance produced in the animal body as a reaction to the presence of bacteria or their products which will destroy and dissolve the corresponding (homologous) organisms.

An Opsonin is a substance found in the blood serum which will unite with bacteria and render them positively chemotactic for the white blood cells. This preliminary union of opsonin and bacterium seems to be necessary before engulfment and destruction of the bacteria by the white blood cells can take place.

In discussing the specific effects and products of the various species of bacteria, there would be an advantage in grouping these bacteria, according to the substances produced, and their effect upon animals and man. However, that this portion may be in keeping with the remainder of the text, the specific effect will be discussed and noted under each organism, and the organisms put in their correct place in the genera of Migula's system of classification. In many instances, bacteria not closely related produce effects that are very similar; in some of these cases, the discussion will be under the first of that group reached, the remainder of the group will contain simply the reference to the form under which the discussion is given.

COCCACEAE

Organisms globose or spherical in a free state, not clongated in any direction before division into one, two, or three planes, when united in pairs or groups, sometimes flattened on the proximal sides, containing five genera, three of which are of importance from our point of view.

BACTERIACEAE

Cells cylindrical or oval, dividing only in one plane, cells straight, rod-shaped, without sheath, either non-motile or motile, by means of flagella; contains three genera.

SPIRILLACEAE

Cells cylindrical, dividing in one plane, not straight, being bent or spiral, and without sheath; contains four genera.

CHLAMYDOBACTERIACEAE

Cells cylindrical, dividing in only one plane, enclosed in a sheath; contains five genera.

BEGGIATOACEAE

Cells cylindrical, dividing in only one plane, destitute of the sheath, united into threads containing sulphur granules, usually motile by means of the undulating membrane. One genus only.

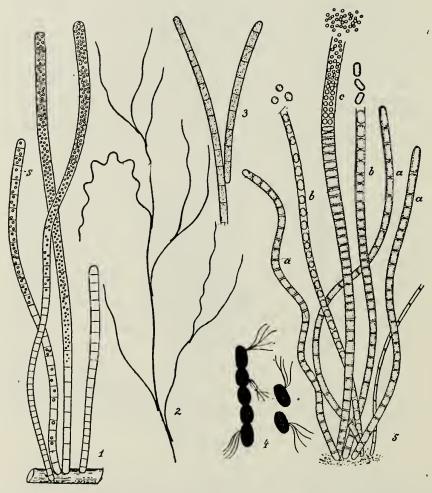


Fig. 25. Chlamydobacteriaceae and Beggiatoaceae. 1. Beggiatoa alba, s Sulfur granules x 800. 2.4. Cladothrix dichotoma; 2 Caenobium x 350. 3, Part of filament magnified 1000 times. 4. Cells with fiagellae x 1000. 5. Crenothrix polyspora; a, young threads; b, threads separating into bodies; c, older threads with spores x 1000. Fig. 1, 2, and 5, after Zopf, Fig. 3 and 4 after Migula.

MYXOBACTERIACEAE

In the vegetative stage these occur as swarming rod-shaped organisms held together by a gelatinous substance secreted by the cells; they show slow creeping movements. They form cysts in which the spores occur. This very peculiar

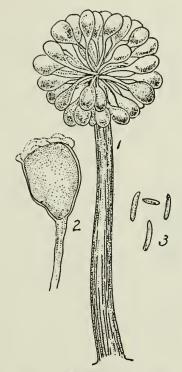


Fig. 26. Myxobacteriaceae. Chondromyces pedunculata. 1. Genereral form of fructification, cyst and pedicel. 2. Single cyst. 3. Rods forming the mass in cyst. After Thaxter.

group first described by Thaxter lives on the dung of animals and in habit resembles the Slime Moulds or Myxomycetes.

COCCACEAE

Micrococcus pyogenes, Var. albus, Roscubach

White-pus Coccus.

This organism, when grown upon artificial media, produces no pigment, otherwise it is identical with the following.

Micrococcus pyogenes, Var. aureus, Rosenbach

Golden-pus Coccus.

Infections produced. This organism, or the preceding, or both, are found very generally associated with, and usually as the cause of wound infection and suppuration in general. They are usually found in furuncles, abscesses, carbuncles, and other inflammatory processes affecting the surface of the body. When present under certain conditions in the blood or various internal organs, they cause pycmia, septicemia, ostemyelitis, inflammations of serous membranes such as pleuritis, peritonitis, ulcerative endocarditis, etc.

Pathogenesis. Infection with this micrococcus causes a marked hyper-leucocytosis. Its presence in tissues is generally followed by the production of pus made up of serum, polymorphonuclear leucocytes, disintegrated tissue, and the bacteria. Ordinarily, the area of infection and inflammation is walled off by an infiltration of the surrounding tissues by these polymorphonuclear leucocytes, forming the so-called pyogenic membrane.

Poisonous properties. In 1894, Van der Velde discovered that sterile filtrates from cultures of Micrococcus contain an hemolysin, which he termed staphylotoxin. In 1901, Neisser & Wechsberg studied this hemolytic substance and gave to it the name staphylolysin. This substance will cause the erythrocytes to dissolve whether within or without the body. A true toxin, leucocidin, is



produced under certain conditions. It causes the leucocytes to swell up and their nuclei to disappear. Leucocidin and staphylolysin differ from the true toxins in that they are not capable of producing anti-toxins after having been heated, i. c., toxoids are not formed. The normal blood serum, however, of man and animals contains more or less of this anti-toxin. The presence of an endotoxin has not been demonstrated.

Immunity. The resistance of the body may Fig. 27. Pus organism (Staphy be heightened by immunization with pure cultures locaceus pyogenes aureus) shown in irregular bunches. After Flügge, of the organism. Immunity is undoubtedly due in large part to the phagocytic activity of the leucocytes. It seemed that virulence of the organism has very little relation to the production of toxins by them for some very virulent types produce very small quantities of toxin. There is some substance secreted which is positively chemotactic to the phagocytes. Immunization is not due to the production of bacteriolysins in the blood. Immune sera have been produced, but have not proven to be of any practical importance. Vaccination is held by Wright to raise the opsonic index of the blood; to this he attributes increased resistance. Agglutinins are present in normal sera in most cases, but systematic injection greatly increases the agglutinating power of the blood.

Micrococcus caprinus, Mohler and Washburn

Disease produced. Takosis.

Animals originally infected. Angora goats.

Susceptible animals. Mouse, guinea pig, rabbit.

Animals naturally immune. White and brown rat, chicken, dog, and sheep. Pathogenesis. Anatomically characterized by emaciation and anemia, congested pneumonic areas in the lung, splenitic atrophy, and induration, spleen often being attached to the diaphragm and neighboring organs by fibrous tissues. The mucous membranes exhibit necrosed areas of mucosa and bacteria are distributed through the blood, consequently may be isolated from any of the internal organs.

Poisonous properties. Microscopically, the lungs are found with many of the terminal bronchioles and alveolae filled mucus and desquamated epithelium, in the liver, hyperaemia with fatty degeneration of periphery of many of the acini, catarrhal nephritis, localized areas of parenchymatous degeneration in the heart; capsule of the spleen thickened and contracted. Small intestines with a local superficial or completed necrosis of the glandular areas. The blood examination shows polycythemia, leucocytosis, and in advanced stages, general poikilocytosis. True toxins not produced.

Immunity. Filtrates seem to have some immunizing power, probably either bactericidal or opsonic. The serum of immune animals has little or no immunizing power. Sterilized cultures and filtrates heated for thirty minutes at 60° C. lose their immunizing power.

Miccrococcus meningitidis, (Diplococcus intracellulus meningitidis, Weichselbaum)

Name of disease. Epidemic cerebro-spinal meningitis.

Animals infected naturally. Man.

Animals susceptible. None, except when injected into a cavity in large quantities. Subdural inoculations in most cases produce a meningitis which, however, does not agree with that produced in man.

Pathogenesis. Severe inflammation of the meninges of both brain and spinal column characterized by the production of considerable quantities of pus. The lumbar puncture and a microscopic examination of the pus will show the organisms present in large numbers. Probably the organisms sometimes reach the blood stream, and secondary infections are produced in various parts of the body by metastisis. It is very probable that the organism is present in acute rhinitis, and that the infection of the brain and spinal cord is secondary.

Poisonous properties. Lipierre extracted what he called a toxin with glycerine, from old culture.

Immunity. Second attacks of the disease are very rare. Lipierre claims to have immunized animals with the toxin and with cultures produced a preventive curative serum from immunized animals. Davis states that there is developed a bactericidal property in the serum, and also agglutinins.

Micrococcus lanccolatus, Fraenkel

Disease produced. Acute infectious pneumonia.

Animals infected naturally. Man (and domestic animals).

Animals susceptible. Rabbit, guinea-pig, dog, and mouse.

Animals immune. Chicken and pigeon.

Pathogenesis. The lungs are most frequently the seat of infection. The infected portion passing through several stages; first, that of congestion, in which the air cells become filled with blood serum and red corpuscles, the former coagulates, the tissues become liver-like in consistency; this is followed by a marked invasion of leucocytes; the contents of the air cells soften and are absorbed and discharged. In most cases of the disease, blood infection probably occurs, consequently, in many cases, infection of various other organs. Pleuritis is most common, then pericarditis, and even generalized peritonitis, endocarditis, arthritis, meningitis, otitis media, conjunctivitis, osteomyelitis, and degenerations in various internal organs, particularly the kidneys and liver.

Poisonous properties. Poisonous substances are produced in greater or smaller quantities in culture media, but no true soluble toxin. Presumably, there is present an endotoxin. The pneumotoxin seems to be toxic toward all the organs of the body.

Immunity. Immunity is probably due largely to phagocytosis, and it is

likewise probable therefore, that it is due in large part to the opsonin content of the blood. There are no bactericidal or anti-toxic substances formed in immune blood. The blood serum of an immunized individual exhibits an increased agglutinating power. Immune sera for the prevention of the disease have not proven a success.

Micrococcus tetragenus, Gaffky

Disease produced. Associated with the tubercle-bacillus, and probably of importance in complicating the pus infections.

Animals infected. Man and animals.

Animals susceptible. White mouse, and guinea-pig.

Animals immune. House-mouse, field mouse, dog, and rabbit.

Pathogenesis. Inoculation of white mouse causes fatal bacteremia. The organism is found in tubercular infections, and probably hastens the necrosis of infected tissue. In some cases it may be the primary infecting agent.

Poisonous properties. Not known.

Immunity. Not known.

Micrococcus catarrhallis, Seifert

Disease produced. Superficial inflammations of the respiratory tract and conjunctiva.

Animals infected. Man.

Animals susceptible. None of the laboratory animals are susceptible, except when the organism is introduced in very large quantities.

Pathogenesis. Probably the primary cause in some cases of conjunctivitis, bronchitis, and catarrh, and in general superficial inflammations of the respiratory passages.

Poisonous properties. Not known.

Immunity. Not known.

Micrococcus gonorrheae, (Diplococcus of Neisser)

Disease produced. Gonorrhea.

Animals infected. Man.

Animals susceptible. None of the laboratory animals are susceptible.

Pathogenesis. Producing a severe inflammation of the mucous membranes of the urethra accompanied by blennorrhea. Secondary infection of fallopian tubes, ovaries, urethra, etc., may occur.

Poisonous properties. Not known.

Immunity. Probably some immunity is developed after infection, but is not lasting. No method of immunizing is known.

Streptococcus equi, Schütz

Disease produced. Strangles.

Animals susceptible. Horses, asses, and their hybrids, and mouse.

Pathogenesis. Producing a severe catarrh of the nasal mucosa, with a swelling of the sub-maxillary, and pharyngial lymphatic glands, abscesses generally form in the latter. May terminate in pharyngitis in a purulent pneumonia or pleuritis. Sometimes cutaneous exanthemata. Metastatic abscesses may appear in various lymph glands.

Poisonous properties. Not fully studied, but probably the same as those of the next organism, Streptococcus pyogenes.

Immunity. Not known.

Streptococcus pyogenes, Rosenbach. (Streptococcus erysipelatos, Fehleisen)

Disease produced. This organism in various forms of inflammation and septic inflammation in general, sometimes alone, sometimes associated with



Micrococcus pyogenes albus, and aureus. Its specific cause, in many instances of septicemia, pyemia, phlegmon, abscesses, boils, erysipelas, ulcerative endocarditis, periostitis, otitis, meningitis, pneumonia, lymphangitis, bronchitis, inflammation of the serous membranes, as pericarditis, pleuritis, peritonitis arthitis, enteritis, endometritis, tonsillitis, salpingitis, has

Fig. 28. Pus organism.bccn held by some authors to cause rheumatic fevers Streptococcus pyogenes. In chains, with pus cells x 800. Af-and also scarlet fever. Probably many so-called terter Flügge. minal infections are produced by this organism.

Pathogenesis. The organism is one of the pyogenic forms reacting much as the Micrococcus pyogenes, as has been described.

Poisonous properties. In many strains of Streptococcus pyogenes, there is present an endotoxin. This is little understood, however, at the present time. It is found that this endotoxin varies greatly; in some cases none at all being found in virulent types. It is susceptible to heat; organisms killed by chloroform being more poisonous than those killed by heat. Virulent streptococci also produce an hemolytic toxin called streptocolysin. This is a true toxin. The blood in fatal cases of streptococcic septicemia is often laked. The toxin is destroyed at a temperature of 70° for two hours and by peptic digestion. Substances which kill the leucocytes are also present in certain strains and inhibit phagocytosis. It is very probable that the pathogenic character of this organism is not entirely explained by its known toxic properties.

Immunity. Immunity against infection of streptococcus is probably due largely to the presence of opsonins in the blood, and the consequent activity of the phagocytes. Sera of animals which have been immunized by inoculation of non-virulent or killed cultures seems to have some protective effect. Such has not come into general use, however. Agglutinins are produced for most strains.

BACTERIACEAE

Bacillus suipestifer, Salmon & Smith

Disease produced. Hog cholera. Probably not the primary cause, but associated with some unknown ultra-microscopic organism.

Animals infected. Swine.

Animals susceptible. Rabbits.

Pathogenesis. Post mortem examination reveals numerous petechiae, ecchymoses, and extravasations of blood into various tissues. This latter is particularly evident beneath the serous membranes. The spleen is enlarged, soft, and engorged. In subacute cases, large intestinal ulcers are formed.

Poisonous properties. Novy gave the name susotoxin to a poison base which he discovered in pure cultures. This is probably not a specific poison of the organism, certainly not a toxic.

Immunity. Vaccination with killed or attenuated cultures develops an immunity, while vaccination against this particular organism is successful, it is not of practical importance, because of the fact that this particular organism is not the primary cause of the disease. Agglutination is well marked, and has been used in diagnosis.

Bacillus piscidus agilis, Sieber

Disease produced. No specific name given.

Animals infected. Fish.

Animals susceptible. Frogs, mice, rabbits, dogs, and guinca-pigs upon inoculation.

Pathogenesis. Disease is marked by shortness of breath, unrest, apathy, and finally paralysis.

Poisonous properties. The filtrate of cultures is poisonous, also the distillate. Cadaverin, and other known ptomains have been obtained from cultures. Immunity.

Bacillus coli, (Bacterium coli commune, Escherich)

Disease produced. This organism is a normal inhabitant of the intestinal tract of man and animals but under certain conditions produces inflammation of the internal organs, such as choleocystitis, peritonitis, meningitis, cystitis, suppurative nephritis, and even generalized septicemia.

Pathogenesis. This organism is not highly pathogenic under ordinary conditions, and when found in inflammatory processes, it is generally associated with other organisms. Undoubtedly, one of the reasons why this organism is found in various organs in post mortems, is the fact that it gains entrance into the blood just before death, producing the so-called agonal invasion.

Poisonous properties. No specific coli-toxin has been produced.

Immunity. Animals may be immunized by the injection of killed cultures. The immunization resulting from the formation of bactericidal amboceptors and agglutinins.

Bacillus enteritidis, Gaertner

Disease produced. Meat poisoning.

Animals infected. Man.

Animals susceptible. Mouse, guinea pig, rabbit, pigeon, lamb, and goat.

Animals immune. Dog, cat, rat, pigeon, and sparrow.

Pathogenesis. It is believed at the present time, that many of the so-called cases of ptomaine poisoning which have occurred in the past, and been recorded as such in medical literature, are due to the presence of this organism, and its soluble toxin. The organism has been isolated repeatedly from spleen of fatal cases. The disease is contracted by eating infected meat, and is characterized by vomiting, and violent diarrhoea, followed by collapse, head-ache, and not infrequently urticarial or herpetic eruptions. Anatomical findings are not specific. Meat undoubtedly is infected only when coming from animals sick with an intestinal or general infection before they were slaughtered.

Poisonous properties. The organism produces in the meat, a soluble heat-resistant toxin in considerable quantities. This toxin in its heat-resisting properties, differs from most toxins and also in the fact that no anti-toxins are produced. Sufficient toxin is usually present to give the first effects of the

disease. The organism itself may or may not gain entrance into the circulation or organs.

Immunity. As before stated, anti-toxins are not developed. Agglutinins, however, are developed.

Bacillus typhosus, Zopf

Disease produced. Typhoid fever.

Animals infected. Man. Inoculation of experimental animals usually negative, except when injected in considerable quantities.

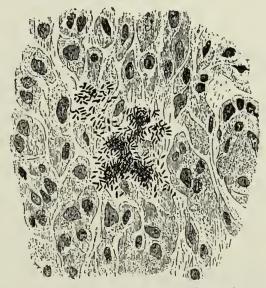


Fig. 29. Typhoid fever bacillus. (Bacillus typhosus). Section from spleen showing bacteria clustered in center. After Flügge.

Pathogenesis. The organism invades the solitary lymph nodes and Peyer's patches in the intestine and produces more or less necrosis and sloughing of tissue. By means of the lymphatic channels, the internal organs are all infected, particularly the spleen which becomes very much enlarged. The bacilli invade the blood, and hence the disease is a true bacteremia. When they lodge in bony tissues, osteitis, periosteitis, and osteomyelitis may be produced.

Poisonous properties. No soluble toxin has been discovered, an endotoxin, however, is present and may be secured through self digestion in cultures, or by grinding and extracting the bodies of the bacteria.

Immunity. No true anti-toxin serum has been produced, inasmuch as no toxin has been discovered. The blood serum of typhoid patients agglutinates the bacteria. Vaccination with killed cultures produces an immunity which lasts probably in most cases several years. The blood serum of animals immunizes against typhoid bacilli, but is not used because it is quickly thrown out of the system when injected, and because it possesses very little curative quality.

Fig. 30.

bott.

Bacillus tctani, Flügge

Disease produced. Animals infected.

Telanus Bacillus. Bacillus tetani. Spores and

rods. a. Vegetative stage. Spore stage. After Ab-

Tetanus, or lock-jaw.

Man, white mouse, rabbit, guinea pig, mouse, rat, horse, to a less degree cattle and most other warm blooded mammals. Most birds, amphibians, and reptiles are immune.

Pathogenesis. There are no gross characteristic anatomie changes, but microscopie degenerative lesions may be found in the ganglionic cells. The disease is characterized usually by a tetanus or rigidity of muscles.

Poisonous properties. A tetanus toxin is produced in quantities in media. It is believed to contain two principles; the first of which, the more important, affects the nerves, and is called tetanospasmin; the second which is hemolytic in its action is ealled tetanolysin. The toxin is destroyed by gastric and panereatic digestion. It has a strong affinity for

nervous tissue; in test tube, practically all of the toxin will become fixed. It is absorbed in the body by the motor ends of the nerves and passes through this by means of the axis cylinders to the ganglionic cells.

Immunity. Natural immunity is probably in part at least, phagocytic in nature, but the presence of a toxin in the blood or in the body causes the production of the anti-toxin. The serum of animals immunized by toxin injections, contains quantities of this anti-toxin, so that it is used in immunization against, and in the cure of tetanus.

Bacillus botulinus, Von Ermengen

Diseases produced. Botulism, or meat poisoning. Animals infected. Man, principally. Animals susceptible. Guinea pig. Animals immune. Dog and rat.

l'ig. 30a. An organism found in sausage. Bacillus botulinus, After Jordan.

Pathogenesis. The ingestion of meat containing bacillus botulinus is followed, in from a day to a day and a half, by salivation, ptosis, bulbar paralysis, and death in from 25 to 50 per cent of the cases. It produces degeneration of glandular organs anid vascular endothelium and consequent hemorrhages.

Poisonous properties. The organism growing in meat produces a characteristic toxin and it is this toxin already formed which produces disease, and not the proliferation of the organism after gaining entrance to the body. It has been found in decomposed hams, and sausages. The toxin differs from that of diphtheria and tetanus in that it is not digested by the gastric juice. It has a special affinity for nervous tissues, but is not so selective as tetanus toxin.

Immunity. Immunization with the toxin results in the formation of an anti-toxic serum which may be used in immunizing against the disease, or in euring. However, this disease is so rare that it is of no commercial importance in this country.

Bacillus alvei, Chesire & Cheyne

Disease produced. European foul brood. Animals infected. Honey bees.

Animals immune. None of the higher animals contract the disease when the organism is inoculated.

Pathogenesis. Destroys the larva of the honey bee.

Immunity. Not known.

Bacillus larvae, White

Disease produced. American foul brood.

Animal infected. Honey bee.

Animals immune. All higher animals.

Pathogenesis. Destroys the larva of the honey bee.

Poisonous properties. Not known.

Immunity. Not known.

Bacillus anthracis-symptomatici, Kruse

Disease produced. Black leg, Quarter evil or symptomatic anthrax.

Animals commonly infected. Cattle.

Animals susceptible. Guinea pig, hog, dog, and rabbit.

Animals immune. Bird, horse, goat.

Pathogenesis. Irregular emphysematous pustules and areas. Muscles contain dark areas with blood serum and gas bubbles.

Poisonous properties. Doubtful, probably an endo-toxin.

Immunity. Established by vaccination with the Bacillus attenuated by exposure to heat and drying.

Bacillus oedematis, Zopf

Disease produced. Malignant oedema.

Animals infected. Horse, sheep, goat, mouse, guinea pig, rabbit, dog, pig, chicken, and pigeon.

Animals immune. Cattle.

Pathogenesis. There is little blood infection by the organism, but a general emphysema of the sub-cutaneous tissues, the gas bubbles being usually very numerous. Any of the body tissues may be affected.



Fig. 30b. Malignant Oedema. Bacillus oedematis maligni. A. From spleen of guinea pig. B. From lung of mouse. Both x 700. After Koch.

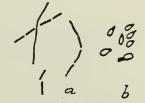


Fig. 30c. Malignant Oedema. Bacillus oedemati maligni, Spores and rods. After Abbott.

Poisonous properties. Not definitely known.

Immunity. Immunization may be affected by vaccination with attenuated cultures. Attenuation is arrived at by passage of the organism through white rats.

Bacillus murisepticus, Koch

Disease produced. Mouse septicemia.

Animals susceptible. Hog, rabbit, mouse, white rat, pigeon, and sparrow. Animals immune. Horse, cow, ass, guinea pig, cat, chicken, and goosc.

Pathogenesis. The organism has been isolated from poisonous meat. Inoculations produce a true bacteremia. Microscopic examination shows the organism to be present principally in the capillaries. The spleen is enlarged, but otherwise, the internal organs show no characteristic lesions.

Poisonous properties. Unknown.

Immunity. Vaccination with killed or attenuated cultures, immunizes, but the serum of immunized animals possesses little curative power.

Bacillus psitticosis, Nocard

Disease produced. Epidemic pneumonia contracted from diseased parrots. Animals infected naturally. Parrots and man.

Animals susceptible. White and gray mouse, pigeon, rabbit, and guinea pig. Animals immune. The dog is partially immune.

Pathogenesis. The disease is a true bacteremia, being associated in man with pneumonia.

Poisonous properties. Not known.

Immunity.

Bactevium influenzae, Lehman & Neumann

Disease produced. Influenza in man.

Animals susceptible. Rabbit, and guinea pig.

Animals immune. Most of the other laboratory animals.

Pathogenesis. Produces purulent bronchitis, and pneumonia. Sometimes there is metastatic infection of other organs producing diseases such as endocarditis.

Poisonous properties. The toxin is intracellular, probably an endotoxin.

Immunity. Vaccination and inoculation do not confer a lasting immunity, in fact, infection in many cases tends to predispose to the disease.

Bacterium cancrosi, Krusc

Disease produced. Chancroid, or soft chancre.

Animals infected. Man.

Animals susceptible. Some of the monkeys.

Animals immune. Other laboratory animals.

Pathogenesis. Disease produced appears first as a small red, papule which becomes larger, and ulcerates. The inguinal and other lymph nodes enlarge and ulcerate. Primary infection most frequently upon the genitalia, other tissues not frequently involved.

Poisonous properties. Not known.

Immunity. Acquired.

Bacterium pneumoniae, Zopf

Disease produced. Pneumonia.

Animals infected. Man.

Animals susceptible. Guinea pig, and rabbit.

Pathogenesis. The organism, probably an avirulent type, is found in normal saliva. Under some conditions, either alone or with other organisms, produces pneumonia or metastatic endocarditis, otitis media, and tonsillitis.

Poisonous properties.

Immunity. Not permanent, probably opsonic in nature.

Bacterium cholerae, Kitt

Disease produced. Chicken cholera, and rabbit septicemia.

Animals infected. Chicken, pigeon, goose, duck, rabbit, and mouse.

Pathogenesis. Generalized septicemia with minute hemorrhages, peritonitis, or the formation of diptheritic areas.

Poisonous properties. Not known.

Immunity. Agglutinins well developed and immunization with killed or attenuated cultures induces the formation of bacterolysins and probably opsonins. In cultural and pathogenic character, this organism is closely related to the four following, all being classed under the general group of organisms producing diseases known as pasteurellosis.

Bacterium suicida, Migula

Animals infected. Hog.

Animals susceptible. Rabbit, guinca pig, and less so chicken and pigeon. See preceding.

Bacterium sanguinarium, Moore

Disease produced. Infectious leukemia.

Animals infected. Chicken. See preceding.

Bacterium bovisepticum, Kruse

Disease produced. Hemorrhagic septicemia.

Animals infected. Cattle. See preceding.

Bacterium avium, Moore

Disease produced. Roup.

Animals infected. Chicken.

Bacterium astheneae, Dawson

Discase produced. Asthenia or going light.

Animals infected. Chicken.

Animals susceptible. Guinea pigs, and rabbits.

Pathogenesis. ?

Poisonous properties. Not known.

Immunity?

Bacterium anthracis, Migula

Disease produced. Anthrax or splenic fever.

Animals infected. Mouse, guinea pig, rabbit, sheep, cattle, man.

Animals immune. Carnivora.

Pathogenesis. Oedema at point of inoculation, spleen very much enlarged, pulpy, internal organs generally hyperacmic. The organism is to be found in

the blood in all parts of the body. Acute degenerative in parenchymatous organs.

Poisonous properties. Neither a soluble toxin nor an endotoxin has been demonstrated, though there is abundant evidence in the tissues of intense intoxication.

Immunity. Immunity may be developed by the inoculation with cultures of the organism grown at high temperatures. The blood serum of animals thus immunized possesses some immunizing power. Opsonins are probably important.

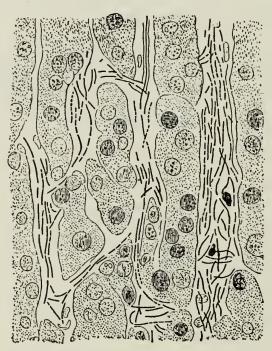


Fig. 30d. A section of liver showing anthrax bacillus (Bacillus anthracis) in blood vessels x 700. After Flügge.

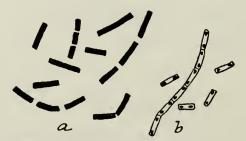


Fig. 30e. Same. a. From culture medium. b. Later stages forming spores.

Bacterium necrophorus, Flügge

Disease produced. Necrosis in various organs.

Animals infected. Calves, lambs, cattle, sheep, goats, antelope, reindeer, red deer, roe, horses, asses, hogs, kangaroos, rabbits, dogs, chickens, kite, guinea pigs, and on experiment mice and pigeons.

Pathogenesis. The local lesion is sharply marked off, usually yellowish, or of a dull brown, of a yeasty consistency, and having a characteristic odor of

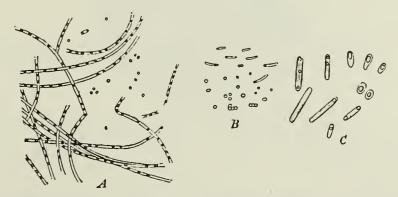


Fig. 30f. Bacillus of Splenic Fever. Bacillus anthracis. Spore formation and spore germination. Λ . From the spleen of a mouse after 24 hours culture in nutrient solution. B. Germination of spores x 650. C. The same x 1650. After Koch.

old cheese and glue. Often produces a diptheritic false membrane. It has been described as producing the following in various animals. Necrotic dermatitis, sheep-pox, abcesses in rabbits, necrobacillosis of the hoof, necrosis in digestive tract, stomatitis, vaginitis, metritis, foot-rot of cattle and sheep, necrotic omphalophlebitis in young animals, jointill, multiple necrosis in the liver, lungs, and avian diptheria, etc.

Disease produced. Diphtheria.

Poisonous properties.

Immunity.

Bacterium diphtheriae, (Flügge)

Animals infected. Man.

Animals susceptible. Horse, guinea pig, rabbit, mouse.

Pathogenesis. Local and general phenomena caused by a soluble toxin, and necrosis of mucous surfaces, and the underlying tissues, and false membrane often forms consisting of fibrin, leucocytes, etc. There is a mild leucocytosis, probably due in most cases to the co-ordinate activity of streptococci.

Poisonous properties. A true toxin is produced in the body and when grown in artificial media. It is this toxin which causes the characteristic clinical picture in diptheria.

Immunity. Injection of non-fatal doses of diptheria toxin results in the production of an abundance of anti-toxin in the blood. This blood serum has powerful curative and prophylactic properties, and has obtained wide usage in general practice.

Bacterium tuberculosis, (Koch)

Disease produced. Tuberculosis, consumption.

Animals infected. Probably no animal is immune to all of the varieties of this organism.

Pathogenesis. The disease generally runs a chronic course, affecting practically all of the organs in the body. In man, characteristic lesions are to be found in the lungs, in cattle generally on the peritoneum. The disease is characterized by the formation of numbers of nodules in the tissues, verying in size from a pin-point to that of an egg. At first, these are hard, and firm throughout, but as they grow larger, generally casease. Microscopically, these tubercules are characterized by the presence in the center of the so-called giant cells with numerous nuclei surrounded by concentric rings of epithelioid and lymphoid cells.



Fig. 30g. Tubercle organisms. Bacterium tuberculosis. After Mohler and Washburn.

Poisonous properties. Not definitely or thoroughly understood, but probably an endotoxin. The killed bodies of the bacteria or the product of their growth in media when injected into animals affected by tuberculosis, cause a characteristic temperature reaction. The material thus injected into animals for diagnostic purposes, is called tuberculin, and is in general used in veterinary practice.

Immunity. Agglutinins are produced in infected animals. Recovery from the disease occurs in many cases doubtless from the walling in of the bacteria. Artificial immunity may be experimentally produced by increasing the opsonic content of the blood, but as yet no accepted method of immunization has been developed.

Bacterium mallei, (Löffler) Migula

Disease produced. Glanders, and farcy glanders.

Animals infected. Usually the horse and ass.

Animals susceptible. Man, goat, cat, hog, field mouse, wood mouse, rabbit, guinea pig, hedge hog.

Animals immune. Cow, house mouse, white mouse, and rat.

Pathogenesis. The lesions usually appear in the horse and ass in the form of discrete, sharply marked ulcers upon the mucous membrance of the nose. The ulcers once formed generally remain open and continue to discharge pus. The lymphatic glands are infected, and generally the submaxillary glands enlarge and ulcerate. Infection of the lungs often occurs through inhalation. Another type of the disease is characterized by infection of the sub-cutaneous lymph channels, which become enlarged and ulcerate, breaking through the skin at various points.

Poisonous properties. This organism produces some poisonous principle similar in a sense to that found in tuberculin and under the name of mallein is used in the diagnosis.

Immunity. Successful immunization of animals against glanders has not been accomplished.

Bacterium leprae, (Hansen) Lehmann Neumann

Disease produced. Leprosy. Animals infected. Man.

Animals immune. Laboratory animals.

Pathogenesis. The organism is found present in leprous tissues, almost filling the cells in many instances. Several types of leprosy are differentiated



Fig. 30h. Glanders bacillus. a. Section through "glandereal" tissue, the small rods massed or single. b. Bacilli stained with methyl-blue. After Flügge.

on the basis of the organ or tissues in the body infected. In many of its aspects, the disease resembles tuberculosis.

Poisonous properties. The organism has not been successfully cultivated. Poisons are unknown.

Immunity.

Bacterium pestis, Lehmann-Neumann

Disease produced. Bubonic plague.

Animals infected. Man.

Animals susceptible. Rat, guinea pig, monkey.

Pathogenesis. Infection usually cutaneous. The lymph glands become swollen, and hemorrhagic and undergo more or less extensive necrosis, generalized septicemia in many cases, pneumonia and hemorrhages in various mucous membranes, especially in the stomach and endothelial surfaces, such as the pericardium and in various parenchymatous organs, with extreme degeneration of the latter. Spleen swollen.

Poisonous properties. The toxic substance is obscure. The filtrates from young cultures usually show little, or no toxicity; the older, more. The toxic substance seems to be susceptible to heat and is present in cultures killed by chloroform.

Immunity. Immunity may be conferred by the injection of sterilized or attenuated cultures, and this vaccination is practiced in some Asiatic countries.

Microspira comma, Schroeter

Disease produced. Asiatic cholera.

Animals infected. Man.

Animals susceptible. Laboratory animals naturally are immune to the disease, but by preventing peristalsis, and neutralizing the acid gastric juice, some experimenters have succeeded in producing the disease. The guinea pig is susceptible to intraperitoneal inoculation.

a



Fig. 30i. Cholera organism. Microspira comma. From the margin of a drop of broth containing pure culture of the Spirillum. a. Long spiral filaments. All x 600. After Flügge.

Pathogenesis. It is essentially an intestinal disease, the organism living within the intestines. The characteristic lesions are produced through the absorption of the poisonous matters there produced. Large and small intestines deeply congested, diarrhoea, Peyer's patches and glands swollen, eventually mucosal necrosis in part. Ulcers eventually form, though perforations are rare. The parenchymatous organs show marked signs of degeneration; the vascular system, the nervous and respiratory systems show no characteristic lesions.

Poisonous properties. The essential poison is intracellular, undoubtedly an endotoxin. It is found in the filtrate of old cultures and in solution of bacterial cells.

Immunity. Produced by considerable quantities of bacteriolysins and probably opsonins. Agglutinins are also produced. Vaccination with killed or attenuated cultures has proven fairly successful, but not the use of the blood serum of immunized individuals as a curative or a prophylactic agent.

SPIRILLACEAE

Spirochaeta pallida

Disease produced. Syphilis.

Animals infected. Man and ape. Animals immune. Other animals.

Pathogenesis. Produces primary lesions in form of ulcers at the point of inoculation, second as gummata in the parenchymatous organs.

Poisonous properties. Not known.

Immunity. Not well understood.

Spirochaeta anserina, Sakharoff

Disease produced. Goose septicemia.

Animals infected. Goose.

Animals susceptible.

Pathogenesis. Producing septicemia.

Immunity.

Spirochaeta Obermeieri, Cohn

Disease produced. Relapsing fever.

Animals infected. Man.

Animals immune. Mouse, rabbit, sheep, and hog.

Pathogenesis. Produces relapsing fever in man.

Immunity.

CHLAMYDOBACTERIACEAE

Cladothrix bovis (Bollinger)

The mass consists of several distinct zones of different elements, the central portion granular with small round bodies radiating out from this tangled mass of thread-like bodies, the outer portion consisting of conspicuous club-shaped colonies. The organism is quite polymorphic. In cultures the threads are from 3 10 to 5-10 μ in thickness with flask-shaped or bottle-like expansions. The organism may be grown upon all the artificial media. The colonies appear as small gray dots with translucent, radiating filaments. If kept for a few days at 37° C, they are opaque and nodular, later they show a whitish downy appearance. In blood serum the nodules are yellowish or blood-red in color; on agaragar the color becomes brownish with age; on potato, reddish-yellow and the white down makes its appearance early.

, Distribution. Widely distributed both in Europe and North America.

Pathogenic properties. The organism was discovered by Langenbeck in 1845, but was not described until 1878 by Bollinger. Israel in 1874-78 called attention to the disease in man, and Boström in 1899 made a careful study of the disease. The disease is not common in man but cases have been described by Murphy and Ochsner and Senn of Chicago. Two of the cases described by Murphy began with tooth-ache and swelling of the jaw.

The disease may be caused by direct inoculation of pus, but there is good reason to believe that not an infrequent source of infection is by means of barley and other grains. There is reason to believe that it occurs in nature as a saprophyte. The history of many cases reported in man seem to indicate

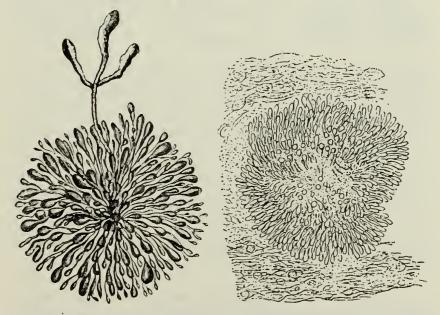


Fig. 30j. Lumpy Jaw Cladothrix bovis showing the radiating masses. At the left, the club-shaped bodies and branches (After Ponfick). At the right, one of the millet like bodies less magnified. After Flügge.

this kind of infection and it is a well-known fact that certain fields are the source of infection. Dr. McFarland says:

When inhaled, the organisms enter the deeper portions of the lung and cause a suppurative broncho-pneumonia with adhesive inflammation of the contiguous plenra. After the formation of the pleuritic adhesions the disease may penetrate the newly formed tissue, extending to the chestwall, and ultimately form external sinuses: or, it may penetrate the diaphragm and invade the abdoninal organs, causing interesting and characteristic lesions in the liver and other large viscera.

Another allied disease is the Mycetoma, or Madura-foot (Cladrothix madureae), which is found in India, especially in the province of Scinde, but occurs also in other parts of Asia, in Europe and northern Africa, and a few cases have even been reported in North America.

Another *Cladrothrix farcinica*, found in Guadaloupe country, is characterized by a superficial lymphangitis and lymphadenitis extending to the tracheal and axillary glands. The glands enlarge, suppurate, and discharge a pus. The internal organs have a pseudo-tubercular appearance.

The organism consists of long delicate filaments, characterized by distinct branching; the old cultures are rich in spores. The organism has been cultivated in the usual media.

It is pathogenic for guinea pigs, cattle and sheep. The culture is virulent for some time.

The papers by Nocard on the farcinica organism and a paper by Musgrove, Clegg and Polk on streptothrix should be consulted.

SCHIZOPHYCEAE

Unicellular organisms common in fresh and salt water. They contain blue, blue-green, violet, or reddish pigments; swarm spores absent; are common in fresh and salt water; simple in structure; existing as a single cell or as a chain held together by a gelatinous envelope, or in small colonies; chlorophyll and other pigments not in definite bodies but distributed throughout the cell-contents or else forming a sheath which lines the cell-wall; reproduction occurs by simple division; some forms produce spores which are thick walled thus enabling the organism to live over unfavorable conditions; after a period of rest these spores germinate and again reproduce in the vegetative way by fission or division. Some of the more common, more or less injurious types found in water are Oscillatoria, Anabaena, Clathrocystis and Nostoc. Cells contain phycocyanin and chlorophyll, the latter not visible because of the former.

CHROOCCACEAE

Cells spherical, singly or collected in colonies surrounded by a copious covering of mucilage forming gelatinous colonies of various sizes. The genus Merismopedia consists of flat rectangular colonies. Cell-division occurs in two directions. The genus Gleocapsa has spherical cells united into colonies, the cell with a thick colorless, brown yellow or violet coat. Some of the species are common in fresh water.

Clathrocystis. Henfrey

This alga occurs in colonies which are at first solid, but later become perforated. The colonies are held together by a gelatinous matrix. The cell contents are blue green, or rose-purple in color. The species most commonly found is the *C. aeruginosa*, occurring not only in Europe, but very widely scattered in

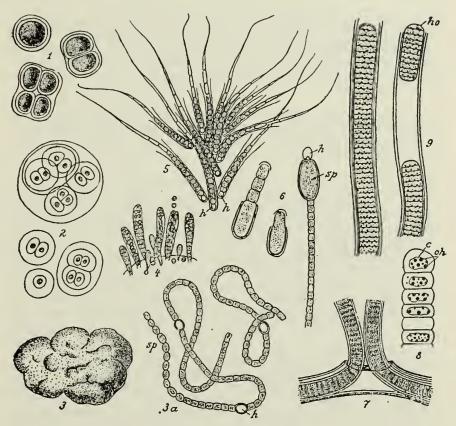


Fig. 31. Schizophyta. Schizophycae. Blue Green Algae. 1. Chrococcus turgidus x 400. 2. Gloeocapsa sanguinea x 400. 3. Nostoc verrucosum. 3a. A pair of chains. 4. Chamaesiphon confervicola x 400; at the right, a caenobium; at the left, germinating arthrospores. 5. Rivularia minutula x 200. 6. Anabaena macrosperma x 100; at the right, a caenobium; at the left, germinating arthrospores. 7. Plectonema Tomasinianum x 200. 8. Filaments of Tolypothrix aegagropila; c—central body, ch—chromatin bodies, x 100. 9. Lyngbya aestnarii x 150; at the right filaments with hormogonia (ho). In all figures, sp—spores, h—heterocyst. Fig. 1, 2, 3, after Cooke; Fig. 4, 5, 7, after Hausgirg; Fig. 8, after Nadson; Fig. 3a, 6, 9, after Wettstein.

North America, especially common in ponds and the plankton of lakes. Other species are *C. roseo-persicina* and *C. Kützingiana*, the former being especially common in ponds and ditches which contain a great deal of decaying vegetable matter. The latter species is now generally referred to the genus *Coleosphacrium*, and the *C. aeruginosa* to the genus *Microcystis*.

OSCILLATORIACEAE

Cells in filaments, apical cells disc-shaped with sheaths variable, sometimes wanting, heterocysts absent; form hormogonia. Common representatives, Oscillatoria and Lyngbya which at times are common in fresh water.

Oscillatoria, Vauch

The plant consists of more than one cell forming a simple filament held together by a common but stout gelatinous sheath, the cells being packed to-

gether like a row of lozenges. A few of the cells fall out of the sheath forming what is called a hormogonium. This starts a new filament which has a characteristic movement hence the name *Oscillatoria*. *Oscillatoria* is common in hot springs sometimes also covering damp soil in greenhouses. It is mainly through decomposition that these algae become noxious.

NOSTOCACEAE

Cells spherical in unbranched chains frequently torulose; sheaths gelatinous frequently forming jelly-like masses; reproduction by hormogonia and spores. Many species of the order are troublesome in water supplies. Some species of the genus *Nostoc* are used as food.

Nostoc, Vaucher

Colonies in flexuose chains united in definite gelatinous investment; cells usually spherical or ellipsoidal; heterocysts terminal or intercalary; spores spherical or oblong. These algae are very common in the lakes in southern Minnesota, Northern Iowa, and elsewhere in the United States. By decomposition, Nostoc produces disagreeable products. Dr. Arthur, some time ago, found Nostoc in quantities in the lakes of southern Minnesota and at one time it was supposed one species produced poisoning of cattle. Dr. Arthur, however, did not attribute the poisoning to this alga. One of the species of this genus frequently found is Nostoc verrucosum common in both the Old and New World.

Anabaena, Bory

Filaments straight or curved, surrounded by a thin sheath united to form a flocculent mass; heterocyst and spores intercalary. This alga also forms filaments which are free or united in a mass. In the filaments occur the vegetative cells, the heterocysts, whose function is not known, and a spore which serves to start the organism again. This organism causes much annoyance in water, not only in North America but in Europe. Dr. Farlow some years ago referred to its injurious properties. It has also been frequently mentioned by Parker as contaminating water supplies in Massachusetts; others have noticed it in New York, and Dr. Trelease has found it in Madison, Wisconsin. The latter writer says in speaking of the Waterbloom and other algae:

After a warm spring, on my return to Madison, June 26, 1887, I observed a considerable quantity of putrid scum on the shore of Fourth Lake, but the south wind scattered it before specimens of it could be obtained. The succeeding fortnight was hot, and after a couple of calm days, succeeding a strong wind from the north-west, the southern half of the lake was filled with suspended particles about a millimeter in diameter. These consisted exclusively of Anabaena Hassalhii, already in full fruit; the spores were the customary Sphaerozyga arrangment, in a collection made June 20th.

This algae is common in many of our northern lakes and is a frequent pest in water reservoirs, producing pig-pen odors and bad taste of water.

RIVULARIACEAE

Filiform filaments attenuated from base to apex, heterocysts basal or rarely absent; sheath tubular, gelatinous, or membranous .

Gloeotrichia, J. Ag.

Free floating colonies solid when young but inflated and hollow when old; the filaments radiating from the centre outwards.

١

Glocotrichia Pisum, (Ag.) Thur.

It forms small green spherical bodies about 1 millimeter in diameter, floating at various depths in the water. It consists of a mass of tapering threads arranged radially in the gelatinous matrix. The apices of the threads protrude more or less, giving it a bristly appearance. The base of each filament contains a heterocyst and above it a slender cylindrical spore and beyond it the ordinary vegetative threads of the alga.

According to Dr. Arthur it is common in Minnesota. He found it common in Waterville, Lake Minnetonka, Lake Phelan in Minnesota and East Okoboji Lake in Iowa. It was thought by the people of Waterville, Minnesota, that this alga caused the death of cattle which drank the water. The history of these cases is recorded by Dr. Arthur as follows:

"That some of the animals had drunk of the water and scum a few hours only before they died was positively known, and that all had done so seemed from circumstances quite probable. After the most careful examination the only plausible hypothesis that could he advanced to account for the death of the animals was that the alga present possessed some toxic or other baneful properties sufficiently powerful to kill a cow in a half hour or more after drinking freely of it. The well-established reputation of all the algae for innocuousness made this hypothesis appear from the very first extremely improbable, but for want of the slightest hint in any other direction it was thought worth while to bear it in mind, and to investigate the matter further.

About the middle of June, 1884, word was received that eight cattle had died on the shore of Lake Tetonka. I at once started for Waterville, arriving on the twentieth and found the algae less abundant than in 1882, but still making the water green some fifty feet or more out from the shore toward which the wind had been blowing several hours. Although the conditions were not the most favorable, yet it seemed best to attempt a direct experiment by giving the animals water charged with algae. After much delay the services of Prof. M. Stalker, state veterinarian of Iowa and professor of veterinary science in the Iowa Agricultural College, were secured to conduct the experiment. A horse and calf were employed. On June 30th, Prof. Stalker, with the assistance of Prof. Edward D. Porter of the university of Minnesota, and in the presence of citizens of Waterville, made the tests, the writer being unable to remain. The animals had not been permitted to drink for some twenty-four hours previous, and were consequently thirsty enough to take a large amount of water well charged with the algae. No bad results of any sort followed.

The thorough and able manner in which the test was made leaves no reasonable doubt of the perfect harmlessness of the algae in a growing condition. I append this last clause, because the citizens of the place still believe that the algae are at the root of the trouble, and that the test did not show it because they were not made at the right stage of their occurrence. Although no sufficient study of the habits of this plant has yet been made to enable one to speak with certainty, yet it does not appear from present data that in some other stage it would give different results, unless it be when decaying, when it turns brown or reddish brown and gives off a peculiar stench. At this time the microscope shows the cells of the algae to be swarming with bacteria. Whether these are other than the common and harmless bacteria of putrefaction it is at present impossible to say. The probabilities are, however, entirely against the hypothesis that the decaying algae or the accompanying bacteria have anything to do with the trouble.

We are therefore obliged to sum up the economic part of this investigation by stating that the death of the animals is probably not due to the suspected algae, and that no clue to the real cause has yet been obtained."

Dr. Arthur in a recent communication states that he has had no evidence so far that these algae are poisonous. That the death of these animals was probably due to bacteria found in the marshes.

OTHER BLUE GREEN ALGAE

Quite a number of additional genera are known to occur in our fresh waters; among them the Glococapsa with cells single or in groups surrounded

by a gelatinous envelope, cell contents bluish green, brownish or reddish; *Merismopedia*, with division in two directions, cells arranged in tabular groups of 4, 8, 16, 32, 64, 128, etc.; *Lyngbya*, with filaments enclosed singly in a sheath, branchless or occasionally branched, cell contents blue and granular.

FLAGELLATAE

One-celled organisms with nucleus, sharply differentiated protoplasmic body, some with a simple membrane, pseudopodia absent, motile during most of their existence; cilia 1 or more and with 1 or 2 pulsating vacuoles; chromatophores occasionally absent; reproduction asexual by longitudinal division. The Euglenales contain the family Euglenaceae. The most common of these is the Euglena viridis which occurs in stagnant pools. Cells are elongated spindle-shaped, cilia 1 and with a red eye spot at one end. Water where these are abundant is not wholesome. The Uroglena forms small sphaeroidal nearly colorless colonies, the central portion of the colony is a hollow space filled with mucilage and the ciliated cells are arranged around the periphery; vegetative multiplication occurs by simple fission and by zoogonidia. The Uroglena, when occurring in large quantities produce a fishy, oily odor. The related Synura produces an odor of ripe cucumbers with a bitter and spicy taste.

EUPHYCEAE

Plants mostly occurring in water, always with a cell-membrane and nucleus. Green or other colors mixed with the green (brown or red). This includes all of the algae or thallophytes that contain chlorphyll which is, however, frequently masked because of other pigments like red and brown.

PERIDINIALES

Small 1-celled organisms of brown color, the chlorophyll masked by diatomin found mostly in the plankton of salt water.

BACILLARIALES

Small 1-celled organisms of brown color, the chlorophyll masked by diatomin of the chromatophores; cell-wall consisting of silica with a girdle and fine lines; reproduction asexual parallel to the long axis of the organism and the formation of auxospores and sexual auxospores by conjugation.

The diatoms are especially important in considering water supplies since they are widely distributed and at times very common. The diatom is like a pill box, made up of two parts, one fitting tightly within the other; the walls are strongly silicified and marked with fine lines; the cell contents are colored brown. Economically, these algae are of some importance as food for fishes, in manufacture of dynamite, and for polishing.

Dr. Moore, speaking of Diatoms in water says:

There are only a few species which are known to give rise to serious trouble in water supplies, but these occur quite frequently and in great quantities. Sometimes the infected water has an odor, variously described as resembling fish or geraniums, and the taste is disagreeable enough to render it quite unfit for use. This condition is often produced by Asterionella. In addition to this effect, however, diatoms are extremely troublesome when contained in water to be used for the manufacture of paper or for laundry purposes, because

of the greenish-brown coloring matter they contain, which stains articles coming in contact with it. Whipple has observed that the growth of diatoms seems to depend upon certain definite conditions of the water—that is, they do not develop when the bottom of the pond or reservoir is quiet; but in spring and fall, when the rising or lowering temperature causes the water to circulate and a good supply of air and nitrates is obtained, the growth is most luxuriant. Thus, it is seen that temperature is only an indirect cause, and not one that need be taken into account oy itself.

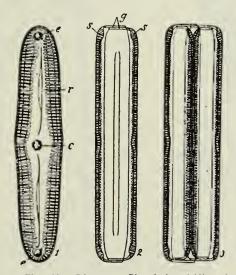


Fig. 32. Diatoms. Pinnularia viridis. 1. View showing markings in wall, c-central nodule; e-polar nodules; r-raphe. 2. Girdle view, s-silicified cell walls; g-girdle bands. 3. In process of division. Fig. 1 and 2 after Pfitzer; Fig. 3 after Wettstein.

CONJUGATAE

Chlorophyll-green algae, membrane without silica; reproduction, swarm cells absent; sexual reproduction by forming zygospores through the union of aplanogametes.

DESM ID1ACEAE

The desinids are green unicellular organisms represented by such genera as Cosmarium, and are found mostly with other algae. They are not especially troublesome. Common genera are Closterium, Cosmarium, and Desmidium.

The filaments of Zygnema are small consisting of a single series of cylindrical cells placed end to end, occasionally with a slight constriction at the points of junction. Each cell has two star-shaped chloroplasts, each one containing a large pyrenoid.

Spirogyra, Link

Spirogyra is a common alga everywhere in our fresh waters, especially in quiet waters as in ponds and ditches. The filaments are simple, occur in bright green masses, often several feet long; cells cylindrical, variable in diameter and relative length; wall smooth and slightly gelatinous; chlorophyll arranged in

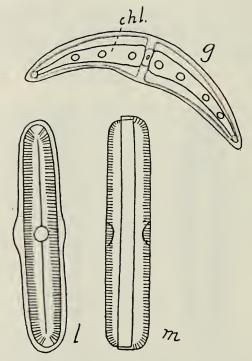


Fig. 32a. Lower figure, two views of diatom. m lateral view; l upper view. Upper figure, a desmid Closterium; chl--choroplastid. Charlotte M. King.

1 or more spiral bands, depending upon the species; nucleus is generally situated in the central portion of the cell; in the chlorophyll band occur what are known as the pyrenoids; reproduction both asexual, by the simple cutting off of the end cell, and sexual, by conjugation through the union of two cells.

ZYGNEMATACEAE

Cells cylindrical, unbranched forming threads, chromatophores present, in masses or spiral bands. Formation of zygospores.

CHLOROPHYCEAE

Chlorophyll-green plants, occurring singly or in colonies, (threads or flattened bodies) with one or more nuclei; reproduction asexual by producing multilateral zoospores and aplanospores; sexual by the copulation of zoogametes, or spermatozoids and oospheres; the spores produce a new plant directly or generally form swarm spores. Water net (Hydrodictyon reticulatum), Pediastrum and Scenedesmus are common in fresh water; Pleurococcus is common on trunks of trees. The Confervales contain the sea lettuce, Ulva latissima, which is used as food, the Conferva with slender green filiaments common in fresh water, the Cladophora fracta in fresh water. Cladophora is a common alga and is quite rough to the touch, and may therefore easily be distinguished from Spirogyra or Zygnema. The frond of Cladophora is branched, with many-

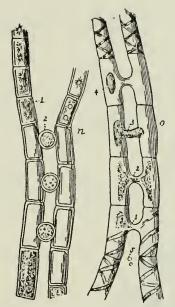


Fig. 33. Fresh Water Green Slimes. Conjugatae. Left hand figure; n—Zygnema. 1. Stellate chlorophyll grain. 2. Zygospore. Right hand figure., s—Spirogyra chlorophyll in spiral band. 1, 2, 3 and 4, in different stages of conjugation. 4. Zygospore.

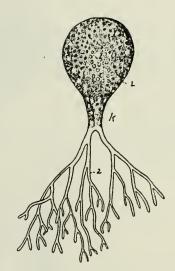


Fig. 33a. Botrydiaceae. Green Scum. Botrydium granulatum. 1. A single large zoosporangium. 2. Rhizoid process.

nucleated cells or, in some instances, only 1 or 2; chloroplasts occur on the margins with a single pyrenoid in each piece of the reticulum. The Siphoneae contain the Vaucheria, found in damp ground, the long unsegmented threads of which produce antheridia and oogonia, and the Botrydium granulatum found on damp ground.

VOLVOCACEAE

Unicellular organisms or forming colonies, each cell with a single chromatophore; forms gametospores and oospores.

Pandorina, Bory

The algae of this genus are collected together in spherical or subspherical colonies known as caenobia. Each caenobium contains about 16 cells closely packed within a gelatinous envelope; the cells are pyramidal in shape and reach almost to the center of the spherical colony; each cell produces 2 cilia.

Volvox, (L.) Ehrenb.

Volvox consists also of globose colonies known as caenobia, each consisting of a large number of small cells from 200 to 22,000, arranged in a single layer within a gelatinous sheath; the caenobium is a hollow sphere, the cells being connected by protoplasmic threads of varying stoutness; each cell has a distinct chloroplast, 2 or more contractile vacuoles and a number of cilia; reproduction occurs through asexual methods or by fertilization; in fertilization, the sperm

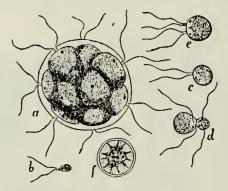


Fig. 34. Chlorophyceae, Pond Scum. Pandorina Morum. a. Floating caenobium. b and c. Zoospores, d. Zoospores conjugating, e. Process of conjugation complete. f. Zygospores. After Luerssen.

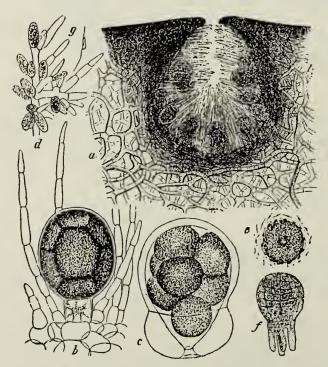


Fig. 35. Bladder Wrack, Fueus vesiculosus, a. Section through conceptacle containing oogonia. b. Single oogonium with egg cells. c. Egg cells escaping. d. Antheridium with sperm cells. e. Egg cells in process of fertilization, surrounded by sperm cells. f. Germination of spores, rhizoid below. g. Single sperm cells. After Thuret.

cell, coming from the antheridium, unites with the egg cell which is contained in the oogonium; the caenobium because of the cilia has a rolling motion. Several species are common like V. globator and V. minor.

Eudorina, Ehrenb.

The eaenobium is globose or subglobose rarely ellipsoid and normally consists of 32 cells arranged within the periphery of a copious mucilaginous mass; each cell contains 1 or more pyrenoids; reproduction takes place as in *Pandorina*. *Eudorina* like *Pandorina* produces a faintly fishy odor.

CHARALES

Plants of brackish or fresh water, consisting of internodes; short whorls of cylindrical branches; cells nucleated; growth from an apical cell; asexual reproduction by means of bulbils and vegetative threads; sexual reproduction by egg-cells and spermatozoids, the latter spirally coiled in the cells of the

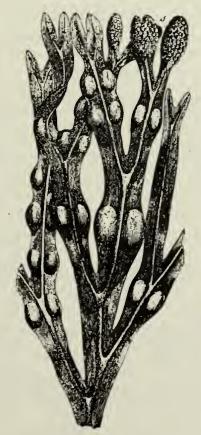


Fig. 36. Bladderwrack, Fucus vesiculosus, Air spaces shown in light areas (I); conceptacles (s) containing reproductive bodies. After Luerssen.

antheridium; the egg cell is contained in a spirally coiled oogonium and after fertilization becomes an oospore. The Stoneworts or Charas are common in brackish water, and though not injurious frequently stop up canals and fill ponds so that it becomes necessary to pull them out.

PHAEOPHYCEAE

Brown algae; chlorophyll marked by a brown coloring matter, phycoplaein, reproduction sexual and asexual, swarm spores, sperm cells and egg cells: marine; tetragonidia absent.

The Phaeosporeae contain the Laminariaceae; the Devil's Apron, Laminaria digitata, and other species from which iodine and mannite are derived. The Macrocystis pyrifera is of great length. The Clyclosporeae contain the family Fucaceae, the common Bladderwrack (Fucus vesiculosus) from which iodine, bromine and soda are obtained. The Sargasso weed (Sargassum bacciferum) found in the Atlantic ocean is also abundant in the Sargasso Sea.

DICTYOTALES

Brown algae; reproductive bodies without motion; tetragonidia present. This group contains a single order Dictyotaceae comprising a few genera.

RHODOPHYCEAE

Red or violet algae; chromatophores contain chlorophyll and red coloring matter (phycoerythrin and rhodophyll); reproduction sexual and asexual; mostly marine. The red sea weeds are divided into several classes and numerous orders. The sub-class Florideae contains most of the species. Food is obtained from several species and the carrageen is furnished by Chondrus crispus, agar agar is obtained from Gracilaria lichenoides found in the Indian Ocean. The Gloiopeltis coliformis and other species are used by the Japanese as food. Many of the species are pretty and are much gathered on the sea coast.

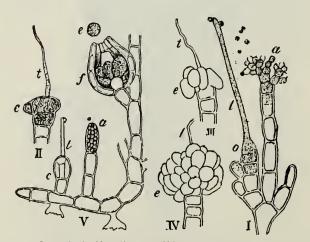


Fig. 37. Red Sea Weed, Nemalion multifidum. 1. Branch with carpogonium and antheridium. 2-4. Different stages of development. 5. Lejolisia mediterranea with antheridium, carpogonium and spores. a—antheridia, c and o—carpogonia, t—trichogyne, s—sperm cells, e—spores, f—fruit. After Thurct and Bornet.

EUMYCETES

Parasitic or saprophytic plants with one or more cells, chlorophyll absent with apical growth; mycelium usually evident; reproduction sexual and asexual, generally the latter; asexual by the formation of zoospores, conidia or spores.

PHYCOMYCETES

Thallus generally of a single branched tubular thread; septa in connection with the reproductive bodies only; threads containing many nuclei; reproduction sexual and asexual, in the latter the spores generally in sporangia (Mucor); conidia in chains (Albugo), or at the end of the hyphae (Plasmopara); reproduction sexual by copulation forming zygospores (Mucor) or oospores in Plasmopara and Albugo.

ZYGOMYCETES

Parasites or saprophytes; mycelium branched not septate, or septa in connection with the formation of the reproductive bodies; reproduction sexual by endospores, acroconidia, or chlamydospores. A group of fungi represented by the Fly Fungus (Empusa) and Common Black Mould (Mucor).

MUCORACEAE

Sporangia with columella, many spored, zygospores between the threads of the mycelium. Few species have the two sexes united on the same plant; generally they are on the separate individuals. According to Blakeslee, Sporodinia contains both sexes (homosporangic, homosporic, homophytic and homothallic). Phycomyces is dioecious, the zygospores producing at germination but one kind of germ tube which gives rise to a sporangium containing both male and female spores, (homosporangic, heterosporic, homophytic, heterothallic) Mucor mucedo has sexes separated on different individuals but two different kinds of germ tubes are formed by the germination of the zygospores, (heterosporangic, heterosporic, heterophytic, and heterothallic). Zygorrhynchus is heterogamic. The same author* has recently reviewed the literature.

About 85 species widely distributed. The Phycomyces was first found in oil kettles, and not infrequently in oil cakes. *Sporodinia* are parasitic on larger fungi. *Pilobolus crystallinus* is common on horse manure, the conidiophore being enlarged. The sporangia look like "fly specks" on the wall. This fungus is not injurious.

Mucor (Micheli) Link. Mucor. Mould

Mycelium creeping, conidiophores simple or branched; sporangia spherical or pear-shaped; columella well developed, wall of sporangium mucilaginous, in some cases chlamydospores, or forming small chains or "cysts"; zygospores produced by the fertilization of two gametes.

A genus of wide distribution of 50 species. The life history of a common species, the *Mucor stolonifer* (*Rhizopus nigricans*) found on bread and decayed fruits is as follows. The gray felted mycelium spreads through the substratum, and on the surface small black bodies, the sporangia, are produced. The conidiophore arises from the felted mycelium and bears an enlarged spherical head, the sporangium, within which, occur the spores. On adding water

^{*} Bot. Gazelte 1900:418. Reprint.

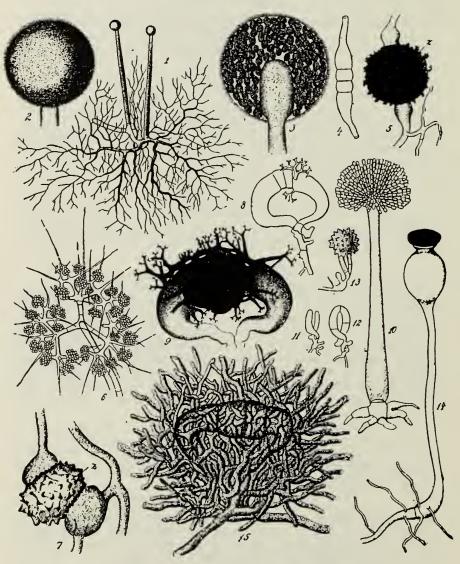


Fig. 39. Black Moulds. Zygomycetes. 1-5. Mucor nucedo. 1. Mycelium and young sporangium x 25. 2. Ripe sporangium x 100. 3. The same in optical view x 100. 4. In process of fertilization x 80. 5. Zygospores (c), greatly magnified. 6-7. Chaetocladium Jonesii. 6. Conidia x 150. 7. Zygospores. 8-9. Fertilization and formation of zygospores of Syncephalis intermedia x 100. 11-13. Fertilization and formation of zygospores in S. cornu x 300. 14. Conidiophore, gelatinous enlargements and black sporangium of Pilolobus crystallinus x 30. 15. Fertilization of Mortierella Rostafinskii x 300. 1-7, 14-15 after Brefeld, 8-13 after Van Tieghem.

to the specimen, the wall of the sporangium collapses and the end of the stalk, known as the columella, turns back, giving it something of the appearance of an umbrella. The columella, before it collapses, projects into the sporangium.

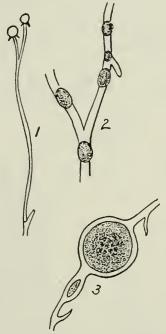


Fig. 40. Mucor Rouxii, Conidiophore. 2. Gemmae. 3. Chlamydospores. All greatly magnified. 1 after Vuillemin. 2 after Calmette. 3 after Wehmer. (Modified by Charlotte M. King).

The spores germinate readily when placed in a moist atmosphere. In addition to the production of a sporangium a stalk may bend over and cause the further extension of the fungus by producing what is known as a stolon.

In some species small, round, or elongated spores are produced in the mycelium which are known as chlamydospores, and spread the fungus. In addition to the formation of spores in the sporangium, zygospores are produced in some species; two threads of the mycelium lie in proximity and nearly parallel, each produces a tube; these meet, the walls are absorbed, and, just back of the meeting point, a cell is cut off. The contents from the old cells pass into the newly formed cell. We also observe that the cell of one arm is somewhat smaller than that of the other. This spore is a resting spore or zygospore. It lies dormant for a period, then germinates by forming directly a coniodiophore with its sporangium containing the spores.

Prof. Blakeslee has shown with reference to the fertilization of some of the species of Mucor that it requires a male and a female plant. In speaking of Mucor mucedo he says:

Mucor mucedo has the sexes separated on different individuals as in Phycomyces, but two different kinds of germ tubes are formed by the germination of its zygospores. While some

germ tubes are male and produce only male spores, others are female and produce only female spores in the germ sporangium. The sporophyte as well as the gametophyte, therefore is unisexual.

"Raggi," used in the manufacture of Arrack, contains Mucor Oryzae (Rhizopus) which transforms rice starch into dextrose, the latter being then fermented by yeasts forms blackish brown sporangia and has a pear-shaped columella. Mucor Rouxii of Calmette is commonly grown in China, where it is found on rice husks and is made from these into Chinese yeast. It changes rice starch into sugar and has been used to some extent for manufacture of alcoholic drinks.

M. racemosus, common in decaying fruit, produces alcoholic fermentation. Mncor fusiger is parasitic on species of Collybia; M. Melittophtorus was found

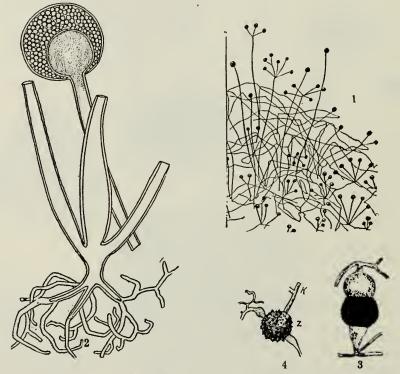


Fig. 41. Common Black Mould (Mucor stolonifer or Rhizopus nigricuns). 1. Sporangia and method of spreading by stolons. 2. Same, showing rhizoids, conidiophore, columella, sporangium and spore. 3. Zygospore, showing method of conjugation. 4. Zygospore germinating; k, conidiophore.

in the stomachs of bees; *M. nigricans* was found by Neumann (1892) and later by Artanet (1893) in the cye of poultry but Barthelat does not consider it pathogenic.

Mucor corymbifer, F. Cohn.

Delicate, white mycelium spreading over the surface of the substratum; conidiophores appressed, spreading; branched sporangia in umbellate clusters, the lower sporangia smaller than the upper, the latter 70μ diameter; wall

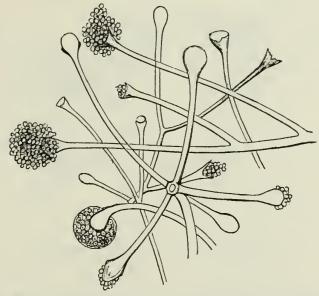


Fig. 42. Mucor corymbiler after the bursting of the sporangium. After Lichtheim.

colorless, smooth, collapsing; spores colorless, small 2 x 3 μ .; columella clubshaped, brownish frequently papillate.

Distribution. Probably tropical, found in tropical drugs, in Europe and the United States.

Pathogenic properties. Lichtheim recognized this species as pathogenic. It grows better at a temperature of blood. When introduced into the circulation of guinea pigs, it produces death in 48 to 72 hours. Mycelium is found in kidneys, spleen and Peyer's patches of intestines which are swollen and ulcerated. Huckel found the organism in the human ear. Dr. Wolffner in Dr. Trelease's laboratory in St. Louis, Mo., cultivated the organism from the human eye. The clinical record of this case was as follows: A farmer near St. Louis, was cutting corn with an old fashioned corn knife. A small piece of corn stalk flew into his eye, later inflammation set in followed by inability to see. Dr. Wolffner found a film over the surface. This film was removed and later Mucor corymbifer developed from it. It has been frequently found in ulcerated portions of the lungs, intestines, nasal cavity, and in the auditory canal. Dogs are immune.

The earliest recorded case of mucor in pneumonomycosis was made by Fürbringer, who had under observation three cases of a disease in two of which he found a Mucor. According to Dr. H. C. Plaut, the cases of otomycosis are not infrequent in India. According to Siebenmann it occurs in .5-1 percent in all diseases of the ear, and males, especially farmers and gardeners, are more predisposed than females. According to Hatch and Row, ear mycosis is common in India, they having observed 22 cases in one month. The most common fungi found in the ear are Verticillium graphii, Aspergillus fumigatus, A. niger, and A. flavus. A. nidulans is somewhat rare, as is Mucor septatus. The Aspergilli will be treated more at length in another connection.

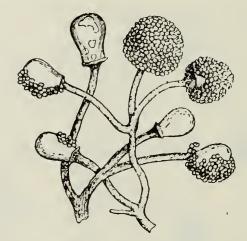


Fig. 43. Black Mould, Mucor rhizopodiformis. Branched conidiophores, large columella and discharged spores around the same. After Lichtheim.

Mucor Trichisi, Lucet and Costantin

This differs from M. corymbifer in a few characteristics of sufficient importance to cause Lucet and Costantin to consider it a distinct species, M. Trichisi having larger spores which are 4 μ in diameter and sporangia 35 μ in diameter. It was isolated from epidermal scabs appearing on a horse affected with tinea, produced by Trichophyton minimum. The M. Regnieri described by the same authors is similar to the preceding.

Mucor rhizopodiformis, F. Cohn

Mycelium at first snow-white then gray, conidiophore single or clustered, brownish, 125 μ long, small rhizoid processes, columella broad, constricted at the base; sporangia spherical, at maturity blackish, spores spherical colorless, 5-6 μ in diameter. Closely allied to M. stolonifer or M. inaequalis.

Distribution. Not uncommon on bread in Europe.

Pathogenic properties. Pathogenic like the preceding. When introduced into the circulation of guinea pigs it produces inflammation and the tissues of the spleen, liver and intestines are found to contain the mycelium of the fungus. The animal becomes inactive, lies on its side and drops its head. Small masses of the mycelium may be found in the kidneys.

Mucor pusillus, Lindt.

Mycelium spreading, with numerous chlamydospores which are capable of germination; conidiophores generally branched; sporangia spherical, brownish, 30-40 μ in diameter; spores ellipsoidal or spherical, 5-8 μ long, 3-5 μ in diameter; columella pear-shaped; zygospores seldom produced, spherical, 70-84 μ in diameter, roughened, chlamydospores abundantly produced: species capable of changing cane sugar into invert sugar, producing the ferment invertase.

Distribution, and hosts. Widely distributed in both Europe and North America, occurring on various decaying objects.

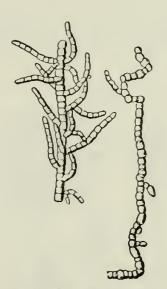


Fig. 44. Mucor racemosus. Fragment of mycelium, which has undergone conversion into chains of oidia x 120. After Brefeld.

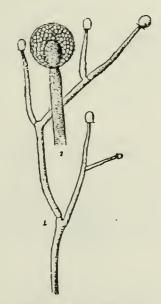


Fig. 45. Mould. Mucor raceniosus. 1. Branched sporophore or conidiophore x 80. 2. Optical section of sporangium x 300. After Brefeld.

Pathogenic properties. Said to be pathogenic for various birds but Pierre Savouré, after some extensive experiments thinks that it plays no part in disease. It was not pathogenic for rabbits and guinea pigs. Bollinger states that it occurs in the respiratory tract of birds where it produces mucormycosis. It has been observed in cutaneous lesions in cavalry horses in France, although culture did not yield this fungus but yielded a trichophyte instead.

Mucor ramosus, Lindt.

Mycelium spreading in the substratum, small, branched, at first white then becoming grayish-white; sporangia black, spherical with marginal spines 60-80 μ in diameter; columella ovate, light brown, 50 μ wide; spores somewhat spherical, 3-3.5 μ in diameter, colorless.

Distribution and Habitat. Found in Europe, not abundant.

Pathogenic properties. Pathogenic for birds. Grows only at a temperature of blood, minimum 20-25° C., maximum 50-58°, optimum 45°.

Mucor ramosus, Lindt

Mycelium branching, abundant in the substratum and superficial conidiophores 5-15 μ wide; sporangia blackish, membrane but slightly colored, 70 μ in diameter; columella rounded at the end or blunt; spores colorless, with delicate membrane, smooth, 3 to 4 μ x 5-6 μ . This resembles M. corymbifer except in the character of the spores.

Pathogenic properties. Pathogenic for guinea pigs, death occurring in 30 to 36 hours.

Mucor septatus, Siebenmann.

Mycelium at first white, later grayish; sporangia light yellowish brown, sporangia small; colmuella colorless; conidiophore branched; spores small 2.5 μ in diameter.

Distribution and Habitat. Found in Europe.

Pathogenic properties. Pathogenic for human beings, found in the ear.

Mucor equinus. (Costantin and Lucet), Pammel

Mycelium branched, at first white or whitish, floccose with simple pedicels withous rhizoid processes, erect or subserect, becoming fascicled, hyphae 8-12 μ in diameter; columella spherical or subspherical 20-50 μ in diameter; spore roundish or slightly angular smooth 4 μ in diameter, chlamydospores numerous especially at blood temperatures.

Distribution. First found in Europe.

Pathogenic properties. Found in horses. Guinea pigs and rabbits inoculated peritoneally die on the 5th or 6th day.

Mucor parasiticus (Lucet and Costantin), Pammel.

Mycelium spreading, branched, brownish fawn color producing stolons and rhizoids; sporangia-bearing peduncles branched; conidiophores 12 to 14 μ wide 1-2 cm. long; columella ovoid pyriform slightly brownish 7-30 μ high; sporangia 8 to 37 μ ; lateral sporangia similar but smaller. Grows readily in nutrient media. The rhizoids sink into the substance, the simple conidiophores rise from the rhizoids. Lucet and Costantin placed this species in a new genus Rhizomucor.

Pathogenic properties. It is essentially parasitic and was isolated from

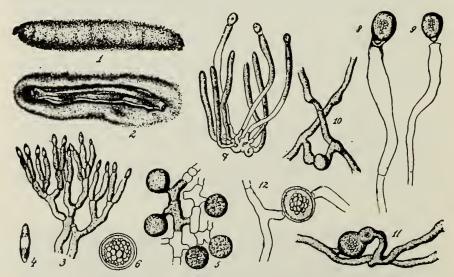


Fig. 46. Entomopthoraceae. 1-6. Empusa sphacrosperma. 1. Larvae of Cabbage Butterfly. 2. Sectional view. 3. Conidiophores and conidia x 300. 4. Conidium x 600. 5. Mycelium with zygospores x 350. 6. Single zygospore x 600. 7-12. Conidiobolus utriculosus; found on the gelatinous fungi like the Jew's Far (Auricularia). 7. Conidiophore x 80. 8-9. Same, much enlarged. 9. Discharging conidium .10-12, Fertilization and forming zygospore. After Brefeld.

the sputum of a tubercular patient. It is pathogenic for rabbits and guinea pigs when inoculated. Lucet and Costantin think that cases of mucormycosis are more frequent than generally supposed. Meyer seems to have made the first observation of a Mucor in animals having observed it in the lungs of a jay. Heisinger in 1821 found a Mucor in the lungs of a goose.

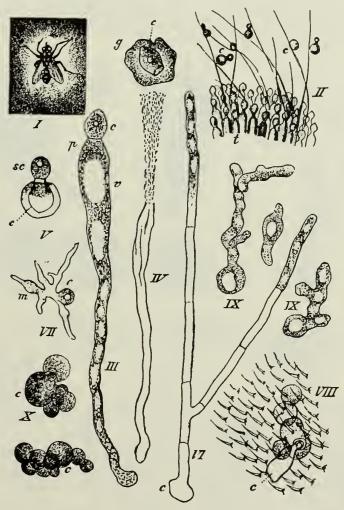


Fig. 47. Fly Fungus. Empuso Muscae. 1. Empuso on fly surrounded by a halo. 2. Part of body of fly; general fructifying part (t); conidia (c) and secondary conidia x 80. 3. Fully formed conidiophore with conidium (c) and vacuole (t) x 300. 4. Tubular conidiophore projecting a conidium (c) surrounded by part of the plasma (g) of the conidiophore. 5. Conidium (c) with a secondary conidium (sc). 6. Conidium (c) forming a mycelium tube 300. 7. Secondary conidium (c) germinating x 300. 8. Part of chitinous integument of fly with conidium (c) penetrating the integument x 500. 9. Fatty bodies of fly containing mycelium of parasite x 300. 10. Yeast-like spronting cells (c) from the fatty bodies of a fly x 500. After Brefeld.

ENTOMOPHTHORACEAE

Mycelium abundant, generally parasitic on living insects; multi-nucleate, non-septate or may become septate; asexual reproduction by means of conidia which are cut off from the end of the sporophore; conidium with one or many nuclei; conidia forcibly ejected; sexual reproduction by means of zygospore; azygospores without fertilization also frequent. One of the most common species of this family is the House Fly Fungus Empusa Muscae. Empusa sphaerosperma is found on the larvae of Cabbage butterfly; E. Grylli is on the Rocky Mountain Locust and the Macrospora cicadina is found on the Cicada. Basidiobolus ranarum occurs on frog excrement.

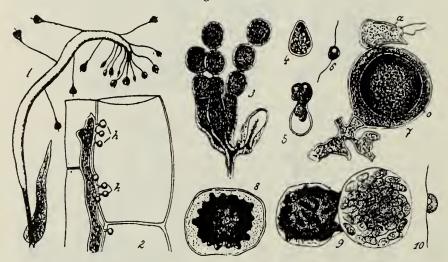


Fig. 48. Peronosporaceac. White Rust. Albugo canida. 1. Inflorescence of Shepherd's Purse with fungus. 2. Mycelium with haustoria (h) x 390. 3. Conidiophores and conidia (spores) in chains x 400. 4 and 5. Formation of zoospores in conidia x 400. 7. Oogonium (o) and antheridium (a) attached, mycelium shown below. 8. Oospore with thick wall. 9. Germinating oospore forming a zoosporangium. 10. Zoospore. 7-10x400. 6. Germinating Zöospore. After DeBary.

OOMYCETES

Mycelium occasionally sparingly developed, tubular, asexual; reproduction by swarm spores or conidia; sexual by the formation of oospores in the *Peronosporaceae* and *Saprolegniaceae*.

Synchitrium has a much reduced mycelium. Sexual reproduction found only in some of the genera of the family Chytridiaceae. The non-septate mycelium is reduced to a single sac shaped cell forming a kind of gall in the host plant. One species of Synchytrium, the S. decipiens, occurs on the Hog-pea (Amphicarpaea monoica).

The family Pythiaceae contains a destructive parasite of seedlings, the Pythium DeBaryanum and the P. proliferum upon dead insects in water.

ALBUGINACEAE AND PERONOSPORACEAE

Mycelium generally well developed. Reproduction sexual and asexual; in sexual reproduction oogonia and antheridia; asexual spores, conidia, or zoospores.

Many members of this group are destructive parasites to cultivated plants like the potato rot fungus (Phytopthora infestans), the onion mildew (Peronospora Schleideviana), the lettuce mildew (Bremia Lactucae), the mildew of the sunflower (Plasmopara Halstedii), the Clover mildew (Peronospora trifoliorum) which may be injurious to animals, the millet mildew (Sclerospora graminicola) which may also be injurious. As a type of this family the downy mildew of the grape (Plasmopara viticola) may be taken. It appears during the early summer and continues till frost. Leaves, berries and stem are affected. The upper surface of the leaf shows yellow patches, underneath a white frosty mould. A section through the leaf will show the mycelium vegetating between the cells. The mycelium gives rise to the fruiting branches of the fungus, the conidiophores, which pass out through the stomata. The conidiophores are dichotomously branched, and at their ends bear the conidia. When these conidia are placed in water they begin to change, at the end of an hour, they swell and the contents divide. According to Dr. Farlow "at the expiration of an hour and a quarter the segments had resolved themselves into a number of oval bodies" which before long succeeded in rupturing the cellwall and making their escape from the mother cell. Each of these zoospores is provided with two cilia. In some, zoospores are not produced, but the whole mass passes out, which soon produces a tube. The zoospores produce germ tubes which probably pierce the leaf of the grape. The temperature most favorable for germination is between 25° and 35° C. Inoculation experiments with the grape vine mildew show that on the second day the disease appears. Sexual method of reproduction takes place later in the season and occurs in the leaf. A slight swelling appears at the ends of the branches of the mycelium, which is spherical in shape, the cell-wall being thick and pale yellow in color. The whole rounded mass is called the oogonium. The central part is the oosphere. A small body is developed from another (or the same thread) which lies along-

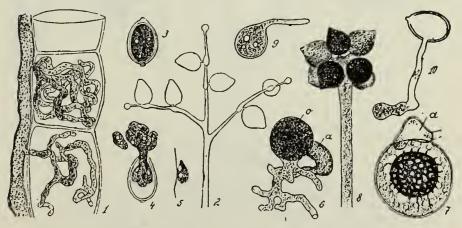


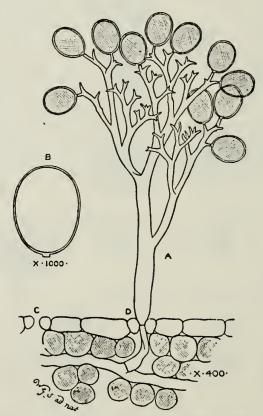
Fig. 49. 1. Downy Mildew. Peronospora calotheca. Mycelium between the cells sending haustoria into the cells, x 390. 2. Potato Rot Fungus (Phytophthora infestans), conidiophores, conidia borne on the branches. 3. Single conidium forming, zoosporangium and the zoospores. 4. Discharge of zoospores. 5. Single ciliated zoospore. 6. Oogonium (0) and antheridium (a). 7. Oospore and antheridium (a) of Peronospora alsinearum x 390. c. Conidiophore and conidia of Basidiophora entospora found on leaves of Erigeron x 200. 9. Germinating conidia of Bremia Lactucae (the Lettuce Downy Mildew). 10. Conidium of Peronospora leptosperma germinating x 300. 1-7 9-10 after DeBary, 8 after Cornu.

side of the oogonium; the antheridium. This pierces the oogonium and the protoplasm of the antheridium passes into the oosphere.

In the species that have been studied like Peronospora parasitica, and Albuyo candida, the oosphere or egg cell contains a single nucleus, situated about at the center, the remaining nuclei having passed into the peripheral layer of the protoplasm of the periplasm.

A single male nucleus passes from the antheridium into the egg cell and fuses with the nucleus of the egg cell. Numerous investigations in this line have been made by Stevens,* Berlese,† and Wager.‡ It is probable that the course of reproduction is similar for other species.

In fertilization karyokinetic changes occur. The protoplasm surrounding the oosphere is used to build up the wall of the oospore. Germination of oospore probably takes place in the spring. In Albugo or Cystopus the conidia are borne in a moniliform chain.



Downy Mildew of Clover (Peronospora Fig. 50. trifoliorum). a. Conidiophore. c. Stoma. b. Conidium; common in Europe on Red Clover, may be injurious to animals. After Smith. jurious to animals.

^{*} Bot. Gaz. 28:149; 23:77; 34:420. † Jahrb. f. wiss. Bot. 31:159. ‡ Annals of Bot. 4:127; 14:263.



Fig. 51. At the left, leaf of Green Foxtail (Setaria viridis), containing the oospores of Sclerospora graminicola, a single spore at a. After Trelease. At the right, spike affected by the same fungus; b spikelet enlarged. The figure at the right, oospores from Ilungarian grass; oog—Oogonium, oos—oosphere; oo—Oospore. The middle figure, Halsted; the right hand, Charlotte M. King.

In some forms the oospores are rare, and in the potato rot fungus they have apparently not been found. In some members of this group oospores are formed without fertilization, (Parthenogenesis).

Dr. G. P. Clinton,* who has made a careful study of the Lima Bean Mildew (*Phytopthora Phaseoli*) and the potato rot fungus (*Ph. infestans*) has been unable to find that the mycelial thread of the antheridium had the same origin as the one which bears the oogonium. It is possible that the fertilization is accomplished in a manner similar to that given for some of the Mucors. He says in a discussion of the potato rot fungus, "All of these facts are now in favor, rather than against distinct mycelial strains (heterothallic forms) except the last, which might indicate a homothallic form, one which contains both antheridia on the same mycelium."

SAPROLEGNIACEAE

Hyphae, long branched, undivided; zoosporangia cylindrical oospores produced from sexual organs, terminal cells are cut off and converted into either

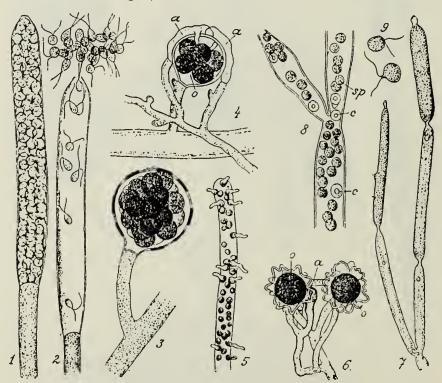


Fig. 52. Saprolegniaceae. Water Mould. 1-3. Saprolegnia Thureti x 200. 1. Zoosporangium before the discharge of spores. 2. Same with biciliated spores heing discharged. 3. The large spherical body, an oogonium and many oospores. 4. Dictyuchus clavatus, o-oogonium, a-antheridium. 5. Aphanos Braunii, zoosporangium with germinating zoospores. 6. Aphanomyces stellatus; o-oogonium and a-antheridium x 390. 7.9. Leptomitus lacteus. 7. Young zoosporangium x 200. 8. Part of older zoosporangium with zoospores (sp) and cellulose grains (c) x 300. 9. Zoospores x 430. Fig. 1-3 after Thuret. 4-6 after DeBary. 7-9 after Pringsheim.

^{*} Rep. Con. Agrl. Exp. Sta. 1905: 304.

oogonia or antheridia. The oogonia may give rise to one or many oospheres or egg cells. The antheridia are tubular and spring from the hyphae below the oogonia. They apply themselves to the oogonia and send out fertilization tubes to the egg cells. The latter then develop into oospores. The asexual method occurs as follows: An examination of the young threads of Saprolegnia will show long filaments which in places are filled with granular protoplasm. Some of these threads are separated from the rest of thread by a cell-wall. Soon the protoplasm arranges itself into polygonal areas. When mature the sporangium breaks and the zoospores are discharged into the water. When emptied a new sporangium is formed by the filament growing up into the old onc, or in some cases a branch buds out below the oogonium.

Recent investigations indicate that the egg cell contains numerous nuclei, but as a general thing they are all degenerate but one. The antheridia also contain many nuclei. According to the investigations of some, one male nucleus enters the egg cell and fuses with its nucleus.

Trow,* Davis,† and Kauffman ‡ have thrown light upon the development of the reproductive body.

Saprolegnia. Nees von Esenbeck. Water mould

Delicate branching hyphae, zoosporangia open from a terminal pore, zoospores pear-shaped with 2 terminal cilia. About 11 species common on decaying objects in water.

Saprolegnia monoica (Pringsheim) De Bary

Zoosporangia cylindrical; antheridia usually in close proximity to the cogonia frequently originating from the same branch; oogonia from short lateral branches; oospore spherical 16 to 22 \mu in diameter, germ tube formed in germination.

Distribution. Widely distributed in North America and Europe.

Pathogenic properties. Occurs on dead insects thrown into the water, parasitic on living fish and crayfish. Frequently troublesome in aquaria. The S. Thureti, DeBary and Achlya prolifera are found on sick fish and crayfish. Hoffmann in 1867 stated that fish in aquaria died under the influence of Mucor mucedo and Saprolegnia,

It is doubtful, however whether the Mucor produced death.

BASIDIOMYCETES

Conidiophores arise from a many-celled, well developed mycelium, hypliae either separate or forming masses; texture soft, powdery or leathery; the spores various, in the most common type, the basidiospore is borne on special structures known as basidia, from which arise little bodies called sterigmata into which some of the protoplasm of the basidium passes. In one group the mycelium consists of septate, branched threads, at maturity nearly disappearing because of gelatinization; mycclium gives rise to chlamydospores formed endogenously; reproduction sexual and asexual, usually the latter; comprises the sub-classes, Hemibasidii and the Eubasidii.

^{*} Annals of Botany. 18:541. † Bot. Gaz. 35:233. ‡ Ann. of Bot. 22:361.

Hemibasidii

Mycelium, local or wide-spread, hyaline, septate, branched, becoming compact and giving rise to endogenous spores; the chlamydospores; color varies, in germination the spores produce a promycelium of terminal or lateral sporidia. These may propagate by budding like the yeast plant. The families are *Ustila-ginaceae* and *Tilletiaceae*.

HEMIBASIDIALES

This includes two families mentioned in the Hemibasidii.

USTILAGINACEAE, Schröt

Usually parasitic fungi in the tissues of living plants; sori usually exposed, forming dusty masses; spores germinate by means of the septate promycelium which gives rise to terminal or lateral sporidia. In some cases, these multiply like the yeast plant, or else produce infection threads. The order contains about 300 species, with the following genera in North America: Ustilago, Sphacelotheca, Melanopsichium, Cintractia, Schizonella, Mykosyrinx, Sorosporium, Thecaphora, Tolyposporium, Tolyposporella, Testicularia. Many plants of the order are destructive parasites occurring upon economic plants like millet, timothy, pink, etc., and one species, the Ustilago esculenta on Zizania latifolia is edible, being used by the Japanese for food.

Ustilago, Pers. Smuts

Mycelium septate, branched, gelatinous, sori on various parts of the host, at maturity dusty, usually dark colored; spores single produced in the fertile threads of the mycelium, the latter entirely disappearing at maturity; promycelium septate, sporidia terminal or lateral, producing infection threads; secondary spores formed in the manner of yeast in nutrient solutions. The largest genus of smuts. About 250 or 260 species. Many of them are destructive parasites on cultivated and wild plants.

The *Ustilago minima* occurs upon the porcupine grass, (Stipa spartea), and the *Ustilago bromivora* upon the brome grass.

Ustilago Zeae. (Beck). Ung. Corn Smut

Sori in the female or staminate inflorescence, leaves and nodes usually forming irregular swellings of variable size; at first covered by a membrane consisting of the gelatinized threads and tissues of the plant; soon rupturing, which exposes the blackish or brownish spores; spores sub-globose or spherical or irregular; echinulate 8-11 μ or sometimes 15 μ long; spores germinate readily under favorable conditions; spore consists of an outer wall, which is spiny, and an inner more delicate, the endospore; the germ tube or promycelium as it is called, normally bears lateral bodies, the sporidia, but under more favorable conditions of food these may branch and bear secondary conidia. If the nutrient material is not exhausted this process of budding may be continued for a long time. These spores may propagate in a decoction of manure. It will then be seen that these budding conidia may be a center of infection.

The conidia as well as the secondary conidia are blown about by the wind and under proper conditions cause the infection of the corn plant. Several years ago Mr. F. C. Stewart made some extended studies of the germination of corn smut in which it was shown that the thermal death point of smut

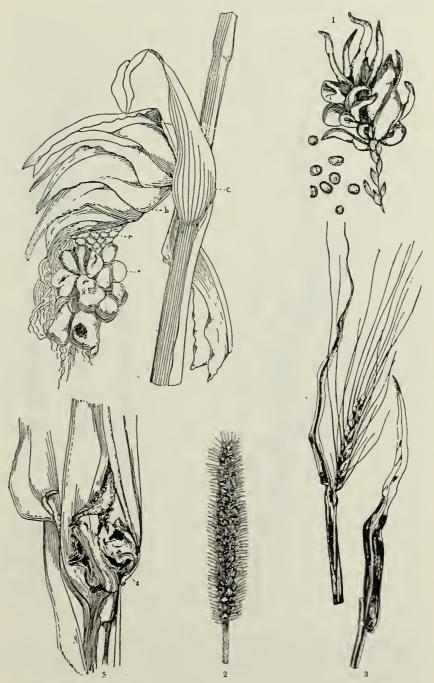


Fig. 53. Maize smut (Ustilogo zeoe). Ear affected. c. Bracts. e. Smut boils. r. Kernels. l. In staminate flowers. Fig. 54. Smut boil making its appearance at the nodes. (C. M. King). 2. Foxtail Smut (U. neglecta). 3. Covered Smut of Barley (U. Hordei).

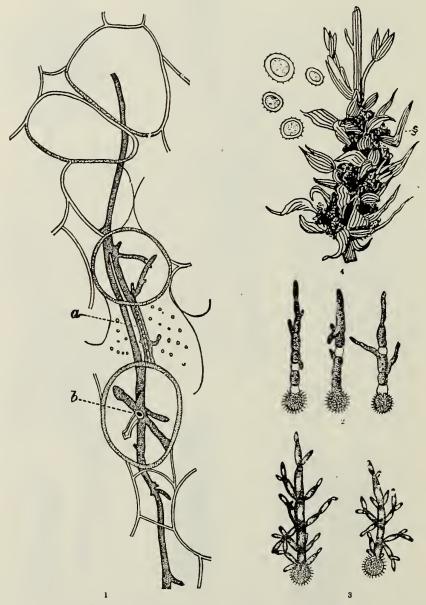


Fig. 55. 1. Maize Smut (Ustilago zcae). Cells showing thread of mycelium passing from cell to cell; (a) thread shows through; (b) section of cut sheath and thread. 2. Corn Smut, Spores in process of germination; each spore is sending out a tube with small lateral bodies. 3. The same, spores germinating in nutrient solution sprouting like yeast. From U. S. Dept. Agr. 4. Kernel Smut of Sorghum (Sphacetotheca Sorghi) on Sorghum.

spores is 15 min. 105°-106° C. in dry oven, and 52° C. when immersed in water; and that corn is unable to come through an inch of soil after 15 minutes treatment with water at 70.5° C., and in dry oven at 78° C. Brefeld found that smut spores produced an abundance of secondary conidia when they were germinated in sugar solution, but with us this has never been a very satisfactory method of propagating them as the cultures soon became infected with bacteria which materially check the progress of the germination of spores.

Distribution and Hosts: Corn smut is found from the Atlantic to the Pacific wherever corn is cultivated, also in other parts of the world. In addition to occurring upon corn, it is found upon teosinte.

Poisonous properties. It has been held by many that corn smut is injurious to cattle. This has been a common belief in some quarters. In some kinds of smut a small amount of ergotin is found. Kedsie reports the following composition: Moisture, 8.30 per cent; albuminoids, 13.06 per cent; carbohydrates, 25.60 per cent; cellulose, 24.69 per cent; sugar, 4 per cent; fat, 1.35 per cent; ash, much sand, 25.5 per cent. Professor Kedsie was unable to find any poisonous alkaloids. In 1868, the United States department of agriculture employed Professor Gamgee to ascertain the cause of the cornstalk disease Professor Gamgee records his experiences as follows and concludes that smut is not injurious:

One cow was fed thrice daily one and one-half pounds of cornmeal and three ounces of smut, mixed with as much cut hay as she would eat. The second had the same allowance, but wet. The amount of smut given in each case was increased to six ounces. The cow fed on dry food lost flesh. Eight days later the dose of smut was increased to twelve ounces three times a day. The cow on the wet food gained in condition, the other one lost. In three weeks the two cows consumed the forty-two pounds of smut. They had a voracious appetite the whole time, and the only indication of a peculiar diet was a very black color of the excrement and the loss of flesh by one animal, although liberally fed on nutritious diet, which, however, was given in a dry state. It is evident that sinut is not a very active poison in combination with wholesome food, and especially if the animal is allowed moist food and plenty of water to drink.

Prof. W. A. Henry, in his work on "Feeds and Feeding" speaking of work done by the Bureau of Animal Industry, Clinton D. Smith and Gamgee, says:

In experiments by the Bureau of Animal Industry, U. S. Department of Agriculture, Washington, corn smut was fed to heifers without ill effects. With all the trials but one ending without disaster, it seems reasonable to conclude that corn smut is at least not a virulent poison, if, indeed, it is one in any sense of the word. It is probable that in the Wisconsin cases, where one cow died and the other was indisposed, the animals suffered because of eating too much highly nitrogenous material rather than anything poisonous. Worse results might have followed the feeding of the same volume of corn meal or cotton seed meal. It would seem that there is little or no danger from corn smut unless cattle consume a large quantity. This is possible where they are allowed to roam through stock fields and gather what they will. There may be cases where animals seek out the smut and eat inordinately of it.

A few years ago Prof. Smith of the Michigan Agricultural College gave the results of some experiments with corn smut. Varying amounts of smut were fed to three grade Shorthorn cows and one grade Jersey. Two of the cows were started with two ounces a day and increased to cleven pounds. Two others were started with two ounces and increased to a pound. The test lasted forty-nine days. They appeared to relish the smut. It produced no signs of abortion in pregnant cows, the milk yield was normal. Prof. Smith concludes that the smut in corn fields is not likely to prove injurious.

Beal states that under certain conditions smut is likely to be injurious to-

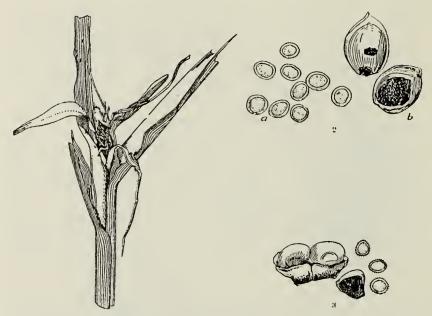


Fig. 55a. 1. Sprouting Grass Smut (Ustilago panici-miliacei) showing large swelling in upper part of plant. C. M. King. 2. Millet smut (Ustilago Crameri). a, spores; b, glumes of millet grains filled with a powdery mass of spores. C. M. King. 3. Kernel of smut corn (Ustilago Fischeri) on maize. Spores at right. Below a sectional view of an affected kernel. Pannuel and King.

cattle. The experiments made by Moore also indicate, as do those of Smith, that smut is not injurious.

Beginning on the morning of January 17, 1894, and continuing until noon of February 2 (sixteen and one-half days), the heifers were fed morning and evening from two to three quarts of a mixture of equal parts by weight of cut hay and a mixture of corn meal, middlings and wheat bran, and sixteen quarts of smut. No injurious affects were observed by Moore. It seems reasonable to conclude from these experiments that under proper conditions corn smut is not injurious. In our experience no cases have ever been reported to us where cattle were supposed to have died from eating corn smut.

Professors Veranus A. Moore and Theobald Smith after making an exhaustive investigation of the so-called corn stalk disease, came to the conclusion that "corn smut is probably not very poisonous, but when fed in considerable quantity no doubt produces injurious symptoms." Miquel in an old work on poisonous plants published in 1838 in Dutch regarded the smuts as poisonous.

Dr. Peters of the University of Nebraska, makes the following comments on the subject of corn smut:

At a Farmers' Institute at David City a gentleman stated that he had often heard his neighbors say, and he had also read the same in agricultural papers, that cornstalk disease was caused by corn smut. He had the opportunity to make the test for himself. He was compelled to clear the farm he rented of the smut. His son gathered the smutty stalks into a yard where two cows ate considerable of the smutty leaves. No bad results followed, as witnessed by the gentleman himself and the owner of the place.

Mr. J. J. Jolliffe in the Drovers' Journal of February 7, 1902, says:

I have never had any had results from smut. I have watched the stock eat ears that seemed 75 percent smut, and they devoured them apparently with as much avidity as sound ears, never affecting the health or appetite in the least.

We have seen cattle die in fields where there was no smut whatever on the stalks and also in some fields that had previously been cleaned as good as one could clean them from this smut; the cattle died in spite of this precaution. Often the owner of the neighboring fields, in which large quantities of smut were found, did not believe this theory and allowed his cattle to run in the stalks promiseuously without suffering any losses.

At the Illinois Experiment Station about sixty pounds of corn smut were fed to a grade Jersey steer, with grain and hay as follows:

From December 13, 1889, to January 2, 1890, 120 pounds of grain, 105 pounds of hay, 20 pounds of smut, and 371 pounds of water were given. From January 2 to January 24, 1890, 176 pounds of grain, 135 pounds of hay, and 39 pounds of smut were fed and 366 pounds of water given. When the experiment was begun, December 13, 1889, the steer weighed 560 pounds. January 2, 1890, he weighed 551 pounds, and January 24, 555 pounds. No evidence of disease was discovered.

Dr. Kilborne records two experiments to test the effects of corn smut. In the first case the smut used came from a field in which several animals had died within five days after they had been turned into it. Three two-year-old steers were fed exclusively on smut-laden stalks and free smut mixed with a small quantity of a mixture of corn meal and wheat bran, for seven days without ill effects. He concludes: "It is safe to say that these animals consumed a much greater quantity of smut than the animals which died in the fields." In the second case, two heifers were fed in addition to corn and hay, sixteen quarts of smut morning and evening for sixteen and a half days. This feeding continued for several months. The animals appeared healthy at the termination of the experiment.

Dr. N. S. Mayo records the experience of a farmer near Manhattan, whogathered the smut from the field and placed it within an enclosure. The cattle broke into the enclosure one night where the smutty corn was thrown and ate all they wished, but no injurious effects were observed.

Ustilago avenae (Pers.) Jens. Oat Smut

Sori found in the spikelets forming a dusty olive brown mass, usually destroying the whole of the inflorescence or only a part; the spore mass at first covered by a membrane which later breaks, thus allowing for the scattering of the spores; spores olive brown, lighter colored on one side, spherical to subspherical or somewhat angular, minutely roughened, 6-9 μ in length. Spores germinate readily in water; infection takes place at the time of germination of the oats.

Distribution and hosts. Found wherever oats (Avena sativa) is cultivated also on wild oats, (A. fatua).

Poisonous properties. Probably not any more injurious than corn smut. When present in large quantities it may produce a sore throat, because of irritation. This fact is mentioned by White. The following note from Dr. White refers to another species found upon grass:

The inflammation affects almost exclusively the face and genitals. It begins upon the former with a violent itching in about twenty-four hours after contact with the reeds, which is followed by a uniform redness, especially marked about the orifices, and swelling of the cyclids. The appearance of the patient strongly resembles that of erysipelas. Later small vesicles develop, terminating in persistent excertations.

Upon the male genitals it begins also with itching, followed by general swelling, with intense redness of the scrotnm, and later by vesicles filled with a yellow serum, terminating.



Fig. 56. A. Oat Smut (Ustilago avenae). From U. S. Dept. of Agrl. B. Tall Meadow Oat Smut (Ustilago perennans).

Fig. 56a. Porcupine Grass Smut (Ustilago hypodites) affecting parts of inflorescence and culm; a, spores, said by Dr. White to be an irritant. C. M. King.

in persistent and very painful erosions. The penis is sometimes affected, producing an inflammatory phimosis.

Delicate skins are most easily affected, and a moist condition favors the action of the parasite. Similar effects were produced upon the skin of the rabbit, by applying to it after being shaved some of the fungus removed from the reeds.

"After reading the above, I wrote to Prof. W. G. Farlow of Cambridge, our authority in crytogamic botany, with regard to the occurrence of this species in America, and received the following reply:"

Your information about the poisonous character of Ustilago hypodites is something quite new to me. I do not know of any reference to the subject in botanical books. The spores of Ustilagineoe are known to be at times irritants when they reach the air-passages, but they are not poisonous to handle. U. hypodites, a species whose characters are not very well marked, I may say, has been found in two places in this country. I found it at Wood's Holl, Mass., on Phragmites (reed). It was found by Curtis in North Carolina on Arundinoria, the cane, and what is probably the same species occurs in Iowa on a species of Stipa. The fungus may be much more common in this country than is now supposed, as few persons have collected fungi of this order.

Ustilago maydis, the corn-smut, grows upon our maize, and U. segetum attacks several of our grains, wheat, oats, barley, and our grasses; but I have never heard of their producing any irritative action upon the skin."

Ustilago nuda (Jens.) Kell. & Sw. Barley Smut

Mycelium found in spikelets forming a dusty olive brown spore mass, protected by a thin membrane which soon becomes ruptured and allows for the dispersal of the spores. Spores lighter colored on one side, minutely roughened, spherical, subspherical or elongated; 5-9 μ in length. Infection probably takes place at the time of flowering of barley.

Poisonous properties. Like the preceding. *Ustilago Hordei*, found upon barley, differs from the species described because of an adhering purple black spore mass covered permanently by the lower parts of the glumes.

Distribution and hosts. Found wherever barley is cultivated in Europe and North America.

Ustilago Tritici (Pers.) Jens. Wheat Smut

Spore masses found in the spikelets of the inflorescence forming black or olive brown mass; usually destroying the entire floral parts, and later spores are scattered by the wind; spores usually spherical or nearly spherical or somewhat elongated, minutely roughened, 5-9 μ in length.

Distribution and hosts. Common upon wheat wherever cultivated in Europe, Australia and North America and South America.

Poisonous properties. Probably injurious like the preceding smuts.

Ustilago Panici-glauci (Wallr.) Wint.

Sori in spikelets infecting all the spikes; spore masses enclosed by glumes, with a rather firm membrane; soon ruptured, permitting the scattering of the spores; spores dark brown, usually spherical or ovoid, occasionally elongated, rather prominently echinulate, 10-14 μ in length.

Distribution. Widely distributed in Europe and North America. One of the most common smuts wherever foxtail grows.

Poisonous properties. It is supposed by some farmers to cause abortion but there is nothing to support this view. According to Professor Power it contains a small amount of ergotin. Possibly injurious like the other species of smut.

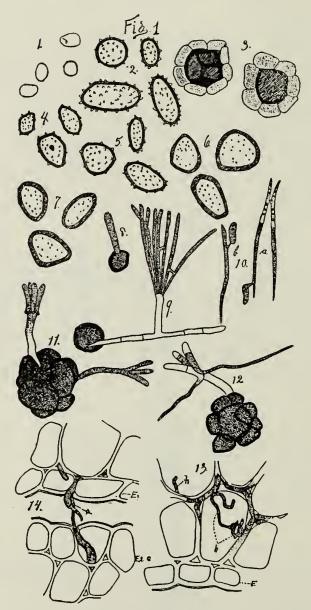


Fig. 57. Smut spores of various kinds. 1. Tall meadow oat smut (Ustilago perenuans). 2. Timothy smut (U. striae-formis). 3. Rye smut (Urocystis occulta). 4. Cheat smut (Ustilago bromicora). 5. Foxtail smut (U. neglecta). 6. Millet smut (U. panici miliacei). 7. Sandbur smut (U. Cesatii). 8, 9. Tilletia gerninating. 10. Secondary spores from conidia. 11, 12. Urocystis spores germinating. 13, 14. Mycelium of same. h. Suckers or laustoria. h. Hyphae passing from one leaf to another. 8-14 after Wolff.

Mr. W. A. Kelty informs me that the smartweed smut (Ustilago utriculosa) often produces severe irritation of the hands when corn is husked.

TILLETIACEAE, Schröt

Mycelium becoming gelatinized in the tissues; the promycelium gives rise to the terminal cluster of elongated sporidia which fuse or do not fuse in pairs, producing secondary sporidia which may be alike or unlike, or the sporidia germinate directly into infection threads. About 150 species, of which Tilletia is the most important genus; aside from the two species described below one species, T. secalis is found upon rye, T. hordei upon barley,, etc., Neovossia Iowensis on Phragmites communis, Urocystis occulta on rye, Urocystis agropyri upon quack grass, and Entyloma ranunculi upon anemone.

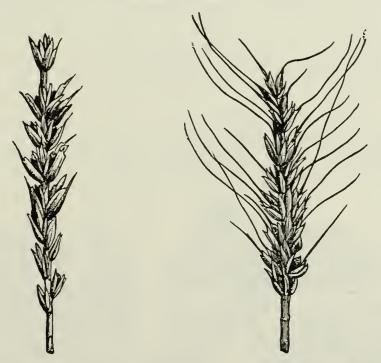


Fig. 58. Wheat Bunt (Tilletia foetens). At the left, a beardless variety with bunt kernels. At the right, a bearded variety with bunt kernels. From U. S. Dept. Agrl.

Tilletia, Tul. Bunt

Sori in various parts of the plant, usually in the ovaries, forming a dusty spore mass; spores 1-celled, formed singly in the ends of the mycelial threads, promycelium of germinating spore short with a terminal cluster of elongated sporidia. 53 species are reported for the genus.

Tilletia foetens (B. & C.) Trel. Stinking Smut or Bunt

Sori in the ovaries, wheat glumes more or less spreading; spores light to dark brown, oblong or sub-spherical, or spherical or sometimes irregular,

16-20 μ in diameter; bad odor just before maturity and retaining the odor even in stored gram.

Distribution and hosts. Common in eastern North America, also in Canada and Manitoba.

Poisonous properties. It produces a bad odor when it occurs in flour and also gives the same a dark color and makes it unsalable.

Tilletia Tritici (Bjerk.) Wint. Wheat Bunt

Sori in the ovaries of wheat ovate or oblong, glumes spreading; spores chiefly spherical or sub-spherical; $16-22 \mu$ in diameter, light to dark brown with winged reticulations. Infection of this and the preceding smut occurs at the time of germination of wheat, hence all of the stalks growing from the single wheat kernel become infected, mycelium growing upward with the growth of the plant.

Distribution and hosts. Common upon wheat wherever cultivated. Reported as destructive and abundant in Michigan, Montana, and Kansas.

Poisonous properties. Same as in the preceding species.

Eubasidii

Conidiophores with true basidia; reproduction generally asexual, sexual in some cases through the fusion of nuclei; spores cut off from the ends of the threads or borne on little sterigmata. The group is divided into two divisions according to the form of the basidia: *Protobasidiomycetes*, the rusts and gelatinous fungi; *Autobasidiomycetes*, toad stools, mushrooms, and puff balls.

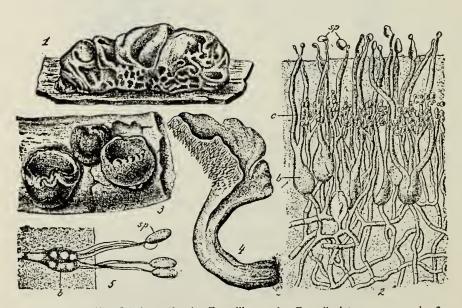


Fig. 59. The Gelatinous Fungi. Tremellineae. 1. Tremella lutescens on wood. 2. Cross section through hymenium, b—Basidia, c—Conidia, sp—Basidiospores, x 450. 3. Eridia, truncata. 4. Tremellodon gelatinosum. 5. Basidia of the same x 560. 1-3 after Brefeld. 4-5 after Mueller.

PROTOBASIDIOMYCETES

Parasitic or saprophytic plants; basidia with longitudinal or cross septa; mycelium septate, branched, either in the interior of the plant as parasites or ramifying the substratum; spores various. The following types occur: spermatia, aecidiospores, uredospores, teleutospores, and sporidia; the spermatia are always accompanied by other spores, generally with the aecidiospores; the aecidiospores are 1-celled and occur in cups; the uredospores are 1-celled, occur in a powdery mass, and germinate immediately; teleutospores arise from the same mycelium that produces the uredospores, one or more cells, on germination they produce a promycelium that bears the sporidia. This group contains the following families: Endophyllaceae with a fungus parasitic on the leaves of spurge, stonecrop and house-leek; Melampsoraceae including several important economic fungi; Pucciniaceae containing a large number of genera; the Auriculariaceae, gelatinous fungi common on decaying wood.

MELAMPSORACEAE

Teleutosori forming incrustations on the surface of leaves; uredosori powdery; aecidia without pseudoperidium (Caeoma) or with well developed

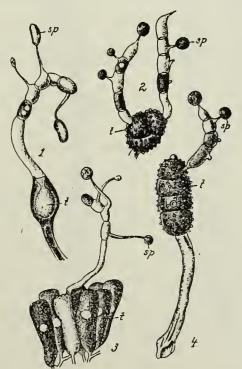


Fig. 60. Uredineae. Teleutospores of different genera germinating. By germination originate the promycelia which divide into cells, each of which produces a conidium. 1. Uromyces Fabae x 460. 2. Triphragmium Ulmariae x 370. 3. Melampsora betulina x 370. 4. Phragmidium Rubi x 370. t—Teleutospore; sp—Conidium. After Tulasne.

pseudoperidium; uredospores 1-celled occurring singly or in groups with or without pseudoperidium; paraphyses present, teleutospores 1-4-celled, closely or loosely united in the plant underneath the epidermis; Calyptospora Goeppertiana occurs upon the huckleberry and blueberry, (Vaccinium) connected with the Aecidium columnare, a very troublesome parasite upon Abies. The Melampsora populina occurs on the cotton-wood, the cotton-wood rust forming red sori on the leaf of the cotton-wood, with waxy incrustations. The other troublesome parasite, the Chrysomyxa Rhododendri occurs upon Rhododendrum. There are about 100 species in the family.

Coleosporium. Lev.

Teleutosori forming flat waxy masses in the leaf; teleutospores composed of several vertical cells enclosed in a thick transparent membrane; each cell germinates by a single undivided promycelium which produces at the end a single sporidium; uredosorus reddish or orange, powdery; spores spherical or sub-spherical, ovate, elliptical, oblong or cylindrical, produced in basipetal chains. A small genus of 30 species.

Coleosporium Solidaginis (Schw.) Thüm. Golden Rod Rust

Uredosori rounded, soon pulverulent and scattered, orange spores in short chains, spherical, oblong, or sub-cylindrical spiny, 20-35 x 15-20 μ ; the teleutosori at first orange, becoming red, flat often confluent forming waxy crusts; Teleutospores cylindrical or somewhat clavate generally 4-celled 60-70x15x25 μ , occasionally longer.

Distribution and hosts. Found in various Compositae, notably Solidago canadensis, S. serotina, etc., Vernonia noveboracensis and Sonchus. Occurs in both Europe and America.

Poisonous properties. Suspected of being injurious to horses; possibly produces stomatitis. Referred to at length under Golden Rod.

PUCCINIACEAE

Teleutospores with a short or long pedicel; spores single or in groups; spores one or more celled; frequently interspersed with paraphyses; spore mass powdery or gelatinous; sporidia arising either from the promycelium or from a similar sterigma after segmentation of the spore contents; aecidia with or without pseudoperidia; uredospores 1-celled, arising from the conidiophores. Includes the Gymnosporangium macropus which produces its aecidium stage on the apple and the teleuto stage on the red cedar, the cedar apple gall with its long gelatinous horn being characteristic; and various aecidia connected with various rusts. Many species have a well marked alternation of generation, an aecidium on one host and the uredo and teleutospores on another host. The Hemileia vastatrix produces the coffee leaf disease of Asia, Phragmidium subcorticum, the rose rust. This large family contains 1500 species.

Recent investigations on the subject of the fertilization and reproduction in the rusts have been made by Profs. Olive, Blackman, Christman, Holden and Harper, and others. These studies seem to indicate that fertilization occurs, but that this fertilization is not, as was thought by the older writers, to be compared with that which occurs in some of the Ascomycetes. The older view was that the spermogonia were male organs and form a strictly morphological standpoint comparable to the structures of like character found in that group of



Fig. 61. Uredosori of Tickle Grass Rust. (Puccinia emaculata).

Fig. 61a. Teleutosori of Puccinia coronata on leaf of oats. Sheath affected with Puccinia graminis.

plants. Clements arranges the rusts with the Ascomycetes. He considers that the apothecium is reduced.**

Puccinia, Pers. Rust

Teleutosori flat, usually powdery masses; teleutospores separate with pedicels, usually consisting of 2 cells, occasionally 1-celled, or sometimes more than 2 cells; the germ pore of the upper cell, at the apex, the lower cell with the germ pore placed laterally below the septum; the promycelium septate with several sporidia. The teleutospores germinate immediately in some species, in others after a period of rest. About 700 species.

Puccinia graminis, Pers. Common Grass Rust

The aecidiospores generally circular, thick swollen with reddish spots surrounding the infected area, yellow below; peridia cylindrical with whitish torn edges; spores subglobose smooth orange yellow, 15-25 μ in diameter; spermogonia on the upper surface consisting of small black dots, uredosori orange red, linear but often confluent, forming long lines, powdery masses; spore elliptical, ovate or pyriform, echinulate, orange yellow 25-38 x 15-20 µ; germ pores 2 above the center on each side; teleutosori persistent; open, generally forming lines on the sheaths, stems and inflorescence; teleutospores fusiform clavate constricted in the middle generally smaller below the apex, thickened, rounded or pointed smooth chestnut brown 15-20 x 35-65 μ; pedicels long and persistent.

The life history of common grass rust is as follows: The common rust produces three stages. One stage occurs in the barberry and is known as the cluster cup fungus. This stage makes its appearance in the northwest some time during the month of June, in the latitude of Ames, a little before the middle of the month. An examination of an affected leaf will show small black specks on the upper surface, surrounded by a yellow spot; this is known as the

in a manner similar to that of higher plants.

^{*} The Genera of Fungi. 5. 98.

** Christman in his studies (Bot. Gaz. 39:267; Trans. Wis. Acad. Sci. 15:517; Bot. Gaz. 44:18.) of the common rose rust (Phragmidium speciosum) finds that the ends of the hyphae produce a terminal sterile cell and a lower fertile one; the fertile cells fuse in pairs with one another, the cell walls breaking down. The nuclei lie side by side and divide, two of the daughter nuclei remaining in the lower part and two passing to the upper part of the dividing cell. Then the upper portion becomes separated by a transverse wall and becomes the first spore mother cell. These fusing cells are approximately equal. Blackman (Ann. Bot. 18:323, See also Ann. Bot. 20:35.) in his studies of another species of the same genus, states that the fertile or female cell contains a larger nucleus and that the male cell is reduced; that the hyphae which gave rise to an aecidium first cut off a sterile cell, and the cell below which at first only contains a single nucleus becomes hinucleated because of the passage of the nucleus from an adjoining cell. Successive divisions of the nuclei occur and finally we have a chain of spore mother cells; each having a pair of nuclei.

Dr. Olive (Annals of Bot. 22:331. Bull S. Dak. Agrl. Ex. 81:119) who has made

each having a pair of nuclei.

Dr. Olive (Annals of Bot. 22:331. Bull S. Dak. Agrl. Ex. 81:119) who has made a close careful study of Triphragmium ulmariae comes to the conclusion that two fusing gamets as well as their nuclei are approximately equal, and that the two gamets differ somewhat in time of development. "That the apparently normal and regular occurence at the base of certain young aecidia of one to many multinucleated cells, points to the necessity of a broader conception as to the mode of development of the aecidium-cup than that beld by either Blackman or Christman. While the part which these multinucleated cells take in the development of the aecidium is as yet somewhat obscure, the evidence appears to point to the conclusion that they are sporophytic structures and that they result from the stimulated growth which followed sexual cell fusions. Should this prove true, it is obvious that the 'fusion cell' does not at once function as a 'basal cell', at the bottom of each spore-row, as maintained by Christman for this type of Rust. Further, the occurrence of occasional instances suggesting 'nuclear migrations,' undoubtedly of a pathological nature, between the multinucleated cells of Puccinia Cirsti-lanceolati, throws doubt on the idea as to the normal origin of the binucleated condition in the aecidium-cup by this means."

It may be of interest further to state that Holden and Harper (Trans. Wis. Acad. Sci. 14:63.) have studied a species of Coleosporium and find that the fusion nucleus divides in a manner similar to that of higher plants.

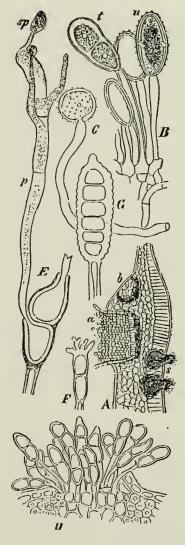


Fig. 62. Some Rusts. A, B. D. E. Puccinia graminis. A. Accidium of Cluster-cup on Barberry; a accidium, s spermogonia on upper surface. B. Uredo spores; u one-celled, teleutospores wo-celled. C. Germination of uredospores showing long tube. D. Connection of stem showing two-celled teleutospores. E. A telcutospore germinating with promycelial tube (p) and sporidia (sp). F. Two-celled teleutospores of Puccinia coronata on oat leaf. G. Teleutospores of Phragmidium incrassatum. All much enlarged. A and G after Luerssen. B-D after DeBary. E after Tulasne.

spermogonial stage; the flask-shaped bodies are called spermogonia and contain the spermatia which do not germinate; their function is not known. A sweetish fluid, which attracts insects, is frequently found in connection with these. Directly opposite the flask-shaped bodies are small globular affairs, "cups," (aecidia), slightly irregular on the margins. Owing to their upward growth they rupture the epidermal cells, and finally the lining layer of cells of the cups also breaks, thus exposing a large number of 1-celled spores borne in chains. These spores arise from short stalks contained at the base of these cups; the cluster cup spores are known as aecidiospores and are transported by the wind and other agencies, and have the power to germinate soon after maturity. When the proper host—a grass, such as bent grass, oats or wheat—appears, the germ

tube enters by way of the stomata, or the so-called breathing pores. The germ tubes produced by the spore of Aecidium berberidis are simple or branched, and in fourteen days usually give rise to the uredo spores, which occur in definite spots called sori. These spots occur in great numbers along the veins of the leaves. Before breaking open, the tissues of the leaf are somewhat paler at those places. The nourishment afforded by the host causes a vigorous mycelium to form, which soon collects in places, pushes the epidermis out, and an orange-colored pustule is formed which is known as the uredosorus.

A section through a diseased sorus shows that an abundance of the vegetative mycelium grows between the cells of the plant, and in some cases haustoria penetrate them. This pustule contains a large number of 1-celled, round or elliptical, spiny, orange colored spores, the uredospores. The spores have two membranes, the outer exospore being provided with wart-like projections, while the inner endospore is provided with several pores through which the germ tube appears. These spores germinate in from three to four hours and can thus start a general infection. These spores, carried by the wind, rain or insects to another part of the same or another plant, germinate, the germ tubes branch and spread over the surface, but the tube cannot enter the host—a grass of some kind, such as wheat, oats or barley—unless it reaches the opening of the stoma, since it cannot bore through the epidermal cells. A single sorus contains hundreds of spores, and as a single plant may contain hundreds of pustules, it can readily be seen that rust must become quite general.

The red rust stage is followed by the black rust stage, known as the teleuto stage. The sori are brownish-black in color, and frequently occupy the same place that the uredo stage did. The spores are dark brown in color, two-celled and smooth, having attached to them a persistent stalk known as the pedicel. The teleutospores do not germinate till the following spring, when each cell produces a germ tube, the promycelium bearing lateral spores, sporidia. These sporidia, when in contact with the barberry leaf, enter by boring their way through the epidermal cells.

The barberry cluster cup fungus, and its connection with common grass rust. It is not absolutely necessary for the common grass rust to have its first stage on the barberry, yet experiment has shown beyond doubt that it does occur on that plant. The theory has been advanced that appearing in one of its stages on the barberry gives the parasite new vigor. It is not improbable that in some places the mycelium or vegetative part of the fungus may be perennial in the tissues of grasses, as it is with many other fungi, probably this is true in southern localities. Beyond question this rust produces spores during the entire year in our southern states, and on the approach of early spring gradually moves northward. It may also be mentioned that in the west this rust certainly does not appear before the cluster cup fungus on the barberry appears. It is usually eight or ten days later, and then appears to a limited extent only. Rust often appears where barberry does not occur within hundreds of miles. This was especially noticeable during the early history of grain culture in the northwest. Rust follows a general infection.

Distribution and hosts. This fungus has been found not only upon wheat but also upon several species of *Bromus, Trisetum* and *Triticum spelta*. Its distribution cannot be given because in most cases the *P. rubigo-vera* included this as well as the *P. glumarum*. It has been intimated above that the uredospores make their appearance on young germinating plants in the fall, but it

appears that the uredo spores are not common the following spring. The investigations of the authors quoted here indicate that not in a single case was it possible to produce uredospores in the spring from those of the autumn.

H. L. Bolley, of Fargo, N. D., remarks in regard to several cluster cup fungi which occur on members of the Borage family:

Several accidia of unknown life history have been studied with reference to their relations to the red rust of *Puccinia rubigo-vero*, many infection tests being made upon young wheat and oat plants, all with negative results.

In this region Onosomodium Corolinianum bears very profusely an aecidium, which, because of its date of appearance, was worthy of suspicion; but tests enough were made to remove this notion.

P. rubigo-vera as well as the common grass rust, is very destructive in England and Australia; but according to Wolf, is not so common in Germany. A few years ago Professor Arthur investigated the subject of wheat rust in Indiana and found that this species was much more destructive to wheat in that state than common grass rust. The same year, 1889, the writer found that this rust was much more common on wheat in Iowa. Carleton says he is confident that the orange-leaf rust (P. rubigo-vera) does very little if any damage to the grain in this country; that in all cases of serious damage to the grain by rust the black-stem rust (P. graminis) is the real cause. In 1907, the leaf rust was very destructive to spring wheat in Iowa.

Puccinia glumarum, Schmidt

Accidium unknown; the uredosori occur along the veins. The diseased leaf is frequently of irregular contour, color orange yellow, spores spherical, or short, elliptical, spiny. Teleutosori, grayish, covered by the epidermis on the stalks and leaves, less frequently on the flowers. Sori divided into chambers,

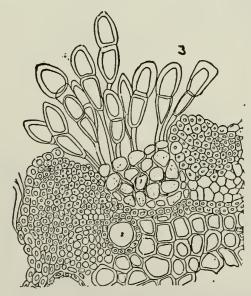


Fig. 63. Covered Rust of Wheat (Puccinia rubigo-vero) from wild Barley, perhaps the same as P. glumarum.

surrounded by paraphyses. Spores with short pedicels, mostly club-shaped, unsymmetrical; apex somewhat truncate, or with one or two projections.

Distribution. Common in Europe and probably also in this country; has usually been referred to as *Puccinia rubigo-vera*. In European mycological works, the aecidium of this fungus is said to be very common on common speedwell (*Lithospermum arvense*), *Echium vulgare*, and *Anchusa officinalis*.

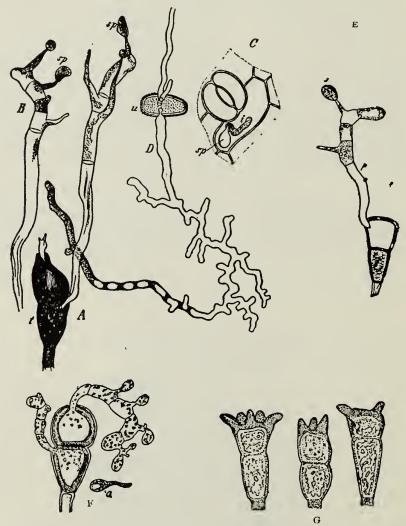


Fig. 64. Forms of rust on cereals. A. Common wheat rust, Puccinia graminis on sheath of wheat, winter spores germinating. B. The same, sporidia sp. C. Epidermis under surface of leaf with sporidium. sp. and germ tube. i. penetrating the epidermis. D. Uredospore germinating after being in water 14 hours in E. Puccinia rubigo-vera germinating. P. Puccinia graminis. Both cells have germinated. a, sporidium germinating, magnified x 600. G. Crowned rust (P. coronata) from oat leaf. G after Bolley; the remainder, DeBary.

Common speedwell is a very common weed in St. Louis and other parts of Missouri and southern Illinois, but so far as known, the aecidium has not been found on these weeds.

Puccinia dispersa. Eriks, & Hen.

This species of rust is apparently very common in Europe. There are three different stages. The aecidium stage produces circular or elongated, somewhat swollen, spots on the leaves, petioles and stem of several members of the borage family. The spores are between 20 to 30 μ or 20 to 30 μ x 19 to 22 μ in diameter. The teleuto spores long remain covered by the epidermis. The sori are chambered, surrounded by numerous brown paraphyses; spores are mostly club-shaped, unsymmetrical and 40 to 50 μ long.

Puccinia coronata, Cda, Crowned Rust

The aecidium produces round or elongated spots with elongated, conspicuous aecidia; the spores from 18 to 25 μ x 14 to 19 μ ; the uredosori are long, confluent, mostly on the upper surface of the leaf; they are orange-colored, and are soon exposed, each pustule containing a large number of 1-celled subglobose, roughened spores which are spherical or short-elliptical; the uredospores are yellow, 20-32 μ in diameter by 28-32 x 20-24 μ . The teleutospores remain covered by the epidermis, and in this respect they resemble the covered rust of wheat (*Puccinia glumarum*). They usually occur on both sides of the leaf. The spores are short-stalked, cuneate and more or less truncate above, crowned with several projecting horns.

Distribution and hosts. Common wherever oats is cultivated and in several of its forms it occurs upon cultivated grasses. This is a well known destructive rust of oats and several other grasses and has received considerable attention from early mycologists. Klebahn has recently described this rust under several distinct forms. The P. coronata dactylidis in a narrow sense includes the rust upon Dactylis glomerata or orchard grass, Festuca sylvatica with aecidia on Rhamnus frangula and P. coronifera.

Ericksson and Henning distribute these forms into P. coronata I, and P. coronata II. Historically this rust is of considerable importance, since Gmelin was familiar with this disease in 1791, and described it as Aecidium Rhamni on Rhamnus. The aecidium stage occurs on species of buckthorn (Rhamnus) especially (R. cathartica and R. Frangula). In Iowa an aecidium is frequently found on a native buckthorn (R. lanceolata), but its connection with this host has not been studied. The aecidium attacks not only the leaves, but occurs on mid-vein, petiole, pedicels and flowers. As a result of the attacks, distorted leaves and flowers are produced.

Puccinia sorghi. Schw. Maize Rust

Uredo and teleutosori upon the leaves and bracts; the former small, light brown sori, soon rupturing the epidermis; teleutosori dark brown; the uredospores are 1-celled, round or more elongated and spiny; the stalk is detached; the spores measure 23-38 x 20-26; teleutosori are elongated dark brown or black being broadly elliptical and 2-celled, 30-52 x 16-24; the apex may be thickened and somewhat pointed. These spores preserve their vitality for some time; but are dormant through the winter. In the spring each cell may germinate by producing a tube, known as the promycelium, which bears lateral bodies

known as sporidia. Acording to Dr. J. C. Arthur, it is undoubtedly connected with an aecidium on Oxalis corniculata.

Distribution and hosts. Common wherever corn is cultivated and according or Carelton, also upon teosinte.



Fig. 66. Corn rust (Puccinia Sorghi) on corn. Winter spores.

Uromyces. Link. Clover Rust

Aecidiospores in cup-like bodies with an evident pseudoperidium; uredosori powdery; uredospores 1-celled with several evident germ pores; teleutosori powdery; teleutospores 1-celled, separate, pedicellate, apex with a single germ pore; sporidia flattened on one side. About 250 species widely distributed. Many of the species produce serious diseases of cultivated plants, as *Uromyces pisi* upon the pea, the alfalfa rust, (*U. striatus*,) and the bean rust, (*U. appendiculatus* (Pers. Lev.) There are many other species found upon our wild plants. Some of these, when they occur upon forage plants, may cause mycotic stomatitis.

Uromyces Trifolii. (Hedw.) Lev.

Aecidia in circular areas of pale colored spots; pseudoperidia short, cylindrical, flattish; edges, whitish, torn; spores sub-globose or irregular, finely roughened, pale orange; $14-23~\mu$ in diameter; uredosori pale brown, round, scattered, surrounded by the torn epidermis; spores round or ovate, roughened; $20-26x18-20~\mu$ with 3 or 4 germ pores; color brown; teleutosori small round almost black; long covered by the epidermis; spores globose, elliptical or sub-pyriform occasionally with wart-like swellings on the summit $15-20x22-30~\mu$; small dark brown in color; pedicels long.

Distribution. Widely distributed upon various clovers, especially red clover and the white clover. So abundant is this fungus at times that the plants are covered with the brown dusty material. Miss Howell reports it as very severe in the state of New York at times. The writer commenting on this fungus some years ago, said:

The fungus did not occur until August and only on the "rowen" or "aftermath." Later it was found quite abundantly on the campus and College Farm. So severely did it attack some of the plants, especially the stems and leaves, that in touching the plants, the hands became covered with brown spores.

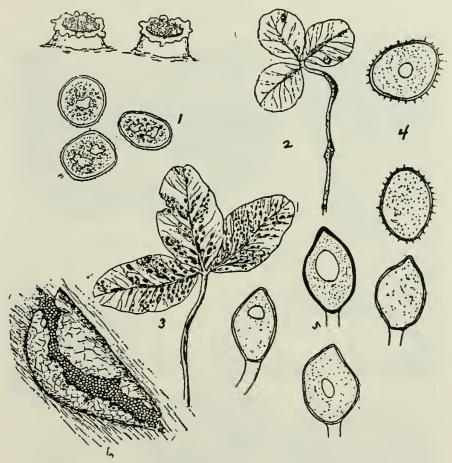


Fig. 67. Clover Rust. Uromyces Trifolii. (Hedw.) Lev. 1. Aecidium spores; above, two cluster cups in which the aecidiospores are found. 2. White clover leaf showing the distortions produced by the aecidium stage. 3. Red clover leaf showing clusters of uredo spores. 4. Uredo spores. 5. Teleuto spores. 6. An uredo cluster more magnified than in 3. Figs. 1, 2, and 3 after Miss Howell. Remainder by Miss King.

How long the fungus has affected clover plants in this country and especially in Iowa is not known.

Poisonous properties. Clover rust has been suspected of being injurious to cattle. Dr. John R. Mohler of the Bureau of Animal Industry, writes as follows with reference to mycotic stomatitis:

Several attempts have been made by the writer to determine the exact cause and alsoto transmit the disease to other animals by direct inoculation, but with negative results.
Suspicion, however, has been directed by various observers to the Uromyces and the red
and black rusts that occur in clovers. These fungi cause very severe irritation of the
lining membrane of the mouth, producing sometimes a catarrhal, at other times an aphthous,
and occasionally an ulcerous stomatitis.

Considerable irritation of the nose and throat is experienced when rusty oats and wheat are threshed. Virchow records a case of severe inflammation of the nose of an old lady in which he found a great deal of *Puccinia graminis*.



Fig. 68. Toadstools, Coral Fungi, &c. Hymenomycetes. 1, Clavaria aurea. 2. Daedalea quercina. 3. Marasmins tenerrimus. 4. Dry-rot Fungus (Merulius lacrimans), 5. Clavaria argillacea. 6. Poisonous Toadstool (Agaricus caesareus) a Ring; v Vellum. 7. Prickle Fungus (Hydnum imbricatum). 8. Polyporus perennis. 8. Corticium amorphum on wood. 1-4, 6-9 after Wettstein. 5 after Harper.

EUTHALLEPHYTA—EUMYCETES—TOADSTOOLS, ETC. 233

AUTOBASIDIOMYCETES

The basidia of the hymenium more or less club-shaped, undivided; sterigmata usually 4, occasionally 2, 6, or 8, coming from the apex of the basidium. The *Dacryomycetineae* with long club-shaped basidia and two long sterigmata. Basidiospores large; spores divided before germination; includes the group *Dacryomycetineae*, and an unimportant group, the *Exobasidiineae*, or small gall parasites containing the *Exobasidium* which occurs upon the cranberry and blueberry. The third group, *Hymenomycetineae*, contains a number of poisonous plants and will be treated more in detail.

HYMENOMYCETINEAE

Mycelium of septate hyphae, loose or delicate in texture or made up into strands or hard masses; hymenium at the time of spore formation free; the basidia form a definite layer or hymenium which may cover the whole surface of the fruiting body, or may be restricted to a definite portion; the fruit is made up of more or less closely compacted threads, hyphae, grown together, or it may be delicate and somewhat ephemeral; the hymenium may be free or gymnocarpous or covered from the beginning; the covering is called the veil, which consists of a layer of threads extending from the margin of the cap to the stem, or the veil may envelop the entire plant; the volva is an envelope which in the young stage completely covers the plant; at maturity it is left in the form of a cup at the base of the stem or distributed from the cap to the base of the stem; the annulus is a ring around the stem formed by

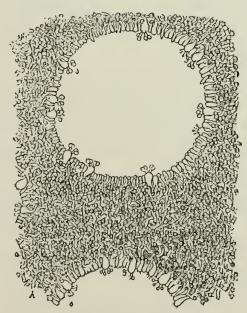


Fig. 69. Cross section of Bracket Fungus. Polyporus igniarius. h. Fungus threads, hyphae between the pores. s. Hymenium surrounding the pores; a number of basidia with spores. After Luerssen.



Fig. 69A. Mushroom (Agaricus campestris). To the left a matured plant and to the right a young plant. (Strasburger, Noll, Schenck and Schimper).

the inner or partial veil. Occasionally cystidia form in Coprinus; chlamydospores are soldom produced.

POLYPORACEAE

Hymenium usually below, porous, tubular, honey-combed, reticulate or of concentric plates; spores produced on the inner surface of the pores. A family consisting of 2300 species, of wide distribution. Some are edible like Bolctus (Boletus edulis B. scaber) and others of this genus, as the Fistulina (Fistulina hepatica) also known as the vegetable beefsteak, and the Sulphur Polyporus (Polyporus sulphureus) when young. Several members are destructive wound parasites of trees. Among these are Polysticius versicolor; the common Bracket fungus (Fomes applanatus), and Trametes radiciperda found on the roots of conifers, and producing death. The dry rot fungus, Merulius lacrymans, is widely distributed and destructive to buildings.

Boletus, Dill. Boletus

Soft or fleshy, the stratum of the tubes on the lower surface of the cap easily separated. They are nearly all found growing on the ground and have the stem attached centrally to the cap. Quite a number of species are edible, some are bitter and some are poisonous. A small genus of 200 species found both in Europe and North America. The Boletus edulis, according to European authority, is one of the most desirable of edible fungi. Professor Atkinson lists this as one of the edible North American species. The B. scaber, also a North American species, is according to Professor Peck, first class, but several species are poisonous and bitter. The B. luridus is regarded as poisonous. The fact that a species turns blue when the plant is cut, should not be regarded as indicative of its poisonous qualities, for this is due to the oxidation process of the fat in contact with the air.

Boletus felleus. Bull

Pileus fleshy, convex above, glabrous or nearly so, grayish-brown, buff-brown, reddishbrown or tawny, flesh, white, taste bitter; tubes long, convex in the mass in mature plants, at first whitish, becoming pale flesh color; stem equal or tapering upwards, usually reticulated at the top only, rarely wholly reticulated, commonly a little paler than the pileus; spores oblong-fusiform, pinkish, .0005 to .0007 inch long.

Distribution. Widely distributed in woods and open places; found upon decayed stumps.

Poisonous properties. Prof Peck says:

The Bitter boletus takes its name from the bitter flavor which its flesh persistently maintains. It is a common species, and one easily recognized by its reticulated stem and flesh-colored tubes taken in connection with its bitter taste.

The cap is rather thick, dry and smooth, but quite variable in color. This is generally some shade of brown tinged with red or yellow. The flesh is white, but when cut or broken and exposed to the air it sometimes assumes a pinkish tint.

The mass of tubes is generally somewhat convex in the mature plant, though it may be plane in the young plant. This also sometimes assumes a pinkish stain when bruised.

The stem varies greatly in length and thickness, and is sometimes crooked and deformed. It is usually reticulated at the top only.

The taste of the flesh in this Boletus, as well as in many species of Lactarius and Russula, is an important aid in the specific identification. In tasting fungi for this purpose care should be taken to select only fresh, sound specimens, and the part tasted should not be swallowed.

Mr. Hurd states that this species is not poisonous. No amount of cooking according to this author, will destroy the bitter flavor.





Honey colored Armillaria (Armillaria mellea). An edible species. (E. A. White in Conn. St. Geol. and Nat. Hist. Survey).

Boletus satanus. Lenz

Pileus large, yellowish-brown on its upper surface; lower surface bloodred at first, later becoming orange red; stalk yellow to reddish-purple with red reticulate markings; spores brownish ovate. Rank and unpleasant taste.

Distribution. In Europe and North America.

Poisonous properties. Said to be extremely poisonous. The B. luridus along with several poisonous species is eaten in Northern Russia. Ford states that these species may occasionally be the cause of transient disturbances in man and may occasionally cause fatal intoxication.

AGARICACEAE

Pileus generally expanded, stipe generally with central attachment, or nearly so, lateral, or sessile; gills simple or branched or anastomosing usually on the lower surface; lamellae folded or veined, radiating from the point of attachment; lamellae bear the basidia which in turn bear the four spores or rarely two, cystidia often present. A large order separated chiefly by the color of the spores. The Melanosporeae have their spores brown, purplish brown or black; in the Ochrosporeae spores are yellowish brown or rusty brown; in the Rhodosporeae, spores are rosy pink; in the Leucosporeae, spores are white, whitish or pale yellow. Many species of the family, like the cultivated mushroom, (Agaricus campestris), the field mushroom, (Agaricus arvensis), the shaggy-mane (Coprinus comatus, Fr.), Lepiota procera, and others, are edible. The Rozites gongylophora of Southern Brazil, is cultivated by the leaf cutting ants for food. No invariable rule can be laid down for the poisonous species. Many of the Leucosporeae are edible, but many are deadly poisonous. A few of the poisonous species are described later.

Amanita, Pers. Amanita

The young plants covered by a membrane which in the button stage is more or less free with the surface of the pileus; later when the stem elongates

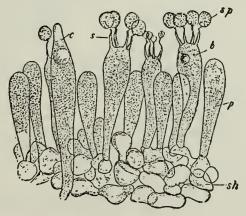


Fig. 70. Part of the hymenium of one of the Agaricaceae. sh. Sub hymenial layer. b. Basidium. s. Sterigmata. sp. Spores from basidium. p. Paraphyses. c. Cyslid. After Bonn text book.

the volva is ruptured; stipe fleshy; volva and annulus present.

In some species the remains of the ruptured volva persist, forming a kind of cup or sheath. In others they occur in the form of small scales or warts on the cap.

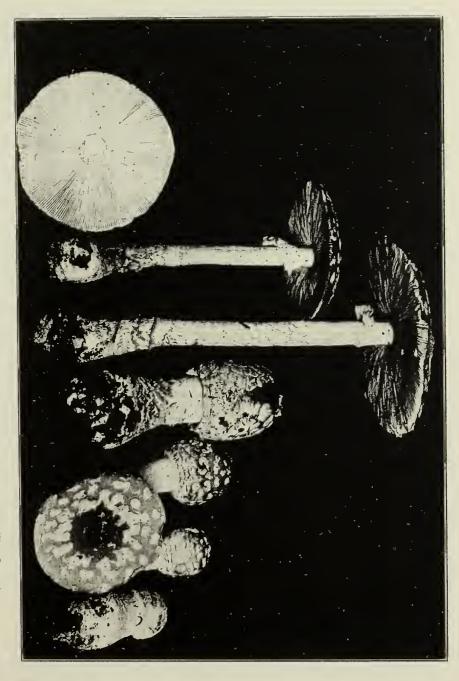


Fig. 71. Fly Agaric or Fly Amanita (Amanita muscaria). a. Mature plant. b. Top view of cap with scales. From U. S. Dept. of Agrl.

Amanita muscaria. L. Fly amanita. Fly agaric

Pileus nearly flat at maturity, warty, slightly striate on the margins, yellow to orange red, cap 3-8 inches broad; gills white or nearly so; stem 4-6 inches long, ½ inch in thickness, cylindrical, hollow, bulbous thickneed at the base, which is more or less scaly from the fragments of the ruptured volva; spore broadly elliptical, white. Dr. Farlow gives the following excellent description of this fungus:

The fly agaric (Amanita muscaria), so called because decoctions of it are used for killing flies, is in most places, at least in the northern and eastern parts of the country, a common species—often a good deal more abundant than the common mushroom. It is found during the summer along roadsides, on the borders of fields, and especially in groves of coniferous trees. It prefers a poor soil, of gravelly or sandy character, and occurs only exceptionally in the grassy pastures preferred by the common mushroom. It grows singly and not in groups, and attains a large size, being one of the most striking toadstools. It



Fly Amanita (Amanita muscaria). Very poisonous. (E. A. White in Conn. St. Geol. and Nat. Hist. Survey).



differs from the common mushroom in having gills which are always white, never pink or purple, and in having a hollow stem which is hulbous at the base and clothed with irregular, fringy scales on all the lower part. The pileus varies in color from a hrilliant yellow to orange and a deep red, the yellow and orange heing more frequent than the red. The surface is polished and has scattered over it a larger or smaller number of prominent, angular, warty scales, which can he easily scraped off. The gills and stalk are white, and there is a large membranous collar, which hangs down from the upper part of the stem. The general appearance together with the color of the pileus and gills noted above, are such that it is difficult to conceive how anyone who has ever seen a common mushroom or read a description of one could mistake this fly agaric for the mushroom. Nevertheless, in the writer's experience, no fungus is so often collected by mistake on the supposition that it is the common mushroom, and it is to the fly agaric that recent cases of poisoning in Washington, D. C., were due.

Distribution. Widely distributed in Europe and North America. Professor Coville, in speaking of this species after the death of Count Achilles de Vecchi, and Chung Yu Ting, says:

The fly amanita is one of the largest, handsomest, and most dangerous of our mushrooms, and is the one whose character has been the most fully studied of all the poisoning
species. It is ahundant ahout Washington in the fall, growing in pine woods, a favorite
situation in these woods being the vicinity of abandoned hog heds. The specimens that
caused the death of Count de Vecchj came from a pine wood ahout a mile west of Fort
Myer, between Balls Crossroads, and Columbia Pike.

Poisonous properties. The chief active poisonous principle of the fly amanita is an alkaloid called muscarin, but other poisonous substances, the chemical nature of which is not yet fully known, also occur in the plant.

Professor Atkinson, in discussing the Toxicology of the species, says as follows:

The substance, Cholin, is of wide occurrence in the animal and vegetable kingdoms. It has heen isolated from Amanita muscaria, A. rantherina, Baletus luridus, and Helvella esculenta. It is not very toxic, but on uniting with oxygen it passes over to muscarin. According to Kobert the substance formed from cholin on the decay of the mushrooms containing it is not muscarin but a very closely related alkaloid, neurin. This transformation of a comparatively harmless alkaloid to an extremely deadly one simply by the partial decay of the plant in which the former is normally found, emphasizes very much the wisdom of rejecting for table use all specimens which are not entirely fresh. This advice applies to all kinds of mushrooms, and to worm-eaten and otherwise injured, as well as decayed ones. Neurin is almost identical in its physiological effects with muscarin which is described helow.

Muscarin is the most important hecause the most dangerous alkaloid found in the mushrooms. It is most abundant in Amanita muscaria, it is also found in considerable quantity in Amanita pantherina, and to lesser, but still very dangerous extent in Baletus luridus and Russula emetica. It is quite probably identical with bulbocin, isolated from Amanita phalloides by Baudier. Muscarin is an extremely violent poison, 003 to .005 of a gram (.06 grain) heing a very dangerous dose for a man. Like other constituents of mushrooms, the amount of muscarin present varies very greatly with varying conditions of soil and climate. This, indeed, may account for the fact that Boletus luridus is regarded as an edible mushroom in certain parts of Europe, the environment heing such that little or no muscarin is developed.

Cases of Mushroom poisoning are frequent in some countries. Gaillard estimated the number of deaths in France at about 100 cases. Among the Americans deaths are not so numerous, although Palmer of Boston, has found 33 cases with 4 deaths.

Inoko of Japan, reports 481 cases in 8 years. The peasants of the Caucasus prepare an intoxicating beverage from *Amanita muscaria* from which many individuals die.

Muscarin acts on the nerve centers, but cases seldom terminate fatally.

Helvella esculenta owes its toxicty to helvellic acid $C_{19}H_{20}O_7$. Very few deaths have been reported in recent years.

According to Kobert, Amanita muscaria contains, besides cholin and muscarin, a third alkaloid, fungus atropin, (pilz-atropin); this alkaloid, like ordinary atropin, neutralizes to a greater or less extent the muscarin. The amount of pilz-atropin present varies, as other constituents of mushrooms vary, with varying conditions of soil, climate, etc., and it may be that in those localities where the Amanita muscaria is used for food the conditions are favorable for a large production of pilz-atropin, which neutralizes the muscarin, thus making the plant harmless. Be this as it may, Amanita muscaria is deadly as ordinarily found. It is undoubtedly used quite largely as food in parts of France and Russia, and it has been eaten repeatedly in certain localities in these countries without harm.

Ford suggests, on clinical grounds, that it may not be the only poison present because even when this drug is completely neutralized by its physiological anti-dote, atropin, the patient, who has eaten *Amanita muscaria*, sometimes dies.*

The alkaloid muscarin, a tasteless alkaline substance with a tobacco-like odor, causes the contraction of pupils; amanitin $\rm C_5H_{18}NO_9$ is an isomer of cholin, and yields muscarin with nitric acid and cetraric acid, $\rm C_{20}H_{18}O_{19}$ Muscarin has been obtained synthetically from cholin. It does not, however, produce quite the same symptoms.

Amanita Frostiana. Peck

Pileus convex to expanded, bright orange or yellow, warty, sometimes nearly or quite smooth, striate on the margin; lamellae white or tinged with yellow; stem white or yellowish, stuffed, bearing a slight, sometimes evanescent annulus, bulbous at the base, the bulb slightly margined by the volva; spores globose; 7.5-10 in diameter. From the character of the poisons it is quite distinct from the A. muscaria.

Distribution. New York to North Carolina.

Poisonous properties. Professors Peck and Atkinson both list it as poisonous. Ford found an hemolysin of low grade intensity. Heated extracts were without action upon animals. Schmiedeberg found a poison.

Amanita phalloides. Fr. Death Cup

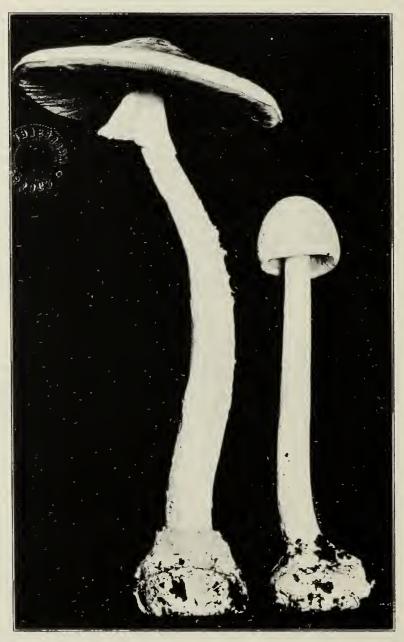
Pileus smooth, fleshy, viscid, greenish, brown or olive to amber; cap 3-5 inches broad, frequently free from remnant of volva; lamellae white; stem 3-6 inches long, annulate; spores globose, white. Prof. Atkinson says: "The presence or absence of these scales on the cap depends entirely on the way in which the volva ruptures. When there is a clean rupture at the apex, the pileus is free from scales, but if portions of the apex of the volva are torn away they are apt to remain on the cap.

Dr. Farlow gives the following excellent description of this fungus:

It is rather common and grows singly in woods and on the borders of fields, rarely appearing in lawns, and is not preeminently an inhabitant of grassy pastures, like the mushroom. It prefers a damper and less sandy soil than that chosen by the fly agaric. The pileus is often a shining white, but may be of any shade, from a pale dull yellow to olive, and when wet is more slimy than the mushroom or the fly agaric. It has no distinct scales and only occasionally a few membranous patches on the pileus. The gills and stalk are

^{*} Sci. 30: 97-108





Deadly Amanita (Amanita phalloides). A very poisonous species of toad stool, (E. A. White in Conn. St. Geol. and Nat. Hist. Survey, page 239).



Fig. 72. Death Cup (Amanita phalloides) one-balf natural size. From U. S. Dept. Agrl.



Fig. 73. Stinkhorn, Phallus impudicus. Common in some vineyards and fields.

white, and the latter has a large ring like the fly agaric, and is hollow, or, when young, is loosely filled with cottony threads, which soon disappear. The base of the stalk differs from that of the fly agaric in being more bulbous and in having the upper part of the bulb bordered by a sac-like membrane, called the volva. The volva is often of considerable size, but more frequently it is reduced to a membranous rim. In this species the stalk is longer and slenderer in proportion to the diameter of the pileus than in either the fly agarie or the common mushroom, and is buried rather deep in the soil or dead leaves, so that it often happens that the bulb is broken off and left behind when the fungus is gathered.

The following differences between the edible and two poisonous species are noted by Dr. Farlow:

(1) The common mushroom has a pileus which is not covered with wart-like seales; gills which are brownish purple when mature; a nearly cylindrical stalk, which is not hol-

low, with a ring near the middle, and without a bulbous base sheathed by a membrane or by scales.

- (2) The fly agaric has a pileus marked with prominent warts; gills always white; a stalk, with a large ring around the upper part, and hollow or cottony inside, but solid at the base, where it is bulbous and scaly.
- (3) The deadly agaric has a pileus without distinct warts; gills which are always white, and a hollow stalk, with a large ring, and a prominent bulb at the base, whose upper margin is membranous or bag-like.
- (4) Other minor points of difference are the different places in which these species grow, and also the colors, which, although they vary in each case, are brilliant yellow or red in the fly agaric, white varying to pale olive in the deadly agaric, and white usually tinged with a little brown in the musbroom.
- (5) A word should be said as to the size and proportions of the pileus and stalk in these three species. In the mushroom the pileus averages from 3 to 4 inches in breadth, and the stalk is generally shorter than the breadth of the pileus and comparatively stout. The pileus remains convex for a long time, and does not become quite flat-topped until old. The substance is firm and solid. In the fly agaric the pileus, at first oval and convex, soon becomes flat and attains a breadth of 6 to 8 inches and sometimes more. The stalk has a length equal to or slightly exceeding the breadth of the pileus, and is comparatively slenderer than is the common mushroom, but nevertheless rather stout. The substance is less firm than in the common musbroom.
- (6) The pileus of the deadly agaric is thinner than that of the common mushroom, and from being rather bell-shaped when young, becomes gradually flat-topped with the center a little raised. In breadth it is intermediate between the two preceding species. The stalk usually is longer than the breadth of the pileus, and the habit is slenderer than in the two preceding species. All three species are pleasant to the taste, which shows that one cannot infer that a species is not poisonous because the taste is agreeable. The fly agaric has scarcely any odor. The two other species have certain odors of their own, but they can not be described.

Distribution. Widely distributed in Europe and North America in woods, groves and pastures.

Poisonous properties. Professor Peck says:

The Poison amanita is very variable in the color of the cap, and yet is so definite in its structural characters that only the most careless observer would be likely to confuse it with any other species. There is, however, a sort of deceptive character about it. It is very neat and attractive in its appearance and looks as if it might be good enough to eat. This appearance is fortified by the absence of any decidedly unpleasant odor or taste, but let him who would eat it beware, for probably there is not a more poisonous or dangerous species in our mycological flora. To eat it is to invite death.

Professor Atkinson says:

Since the Amanita phalloides occurs usually in woods, or along borders of woods, there is little danger of confounding it with edible mushrooms collected in lawns distant from the woods and in open fields. However, it does occur in lawns bordering on woods, and in the summer of 1899 I found several of the white forms of this species in a lawn distant from the woods. This should cause beginners and those not thoroughly familiar with the appearance of the plant to be extremely cautious against eating mushrooms simply because they were not collected in or near the woods. Furthermore, sometimes the white form of the deadly amanita possesses a faint tinge of pink in the gills, which might lead the novice to mistake it for common mushroom. The bulb of the deadly amanita is usually inserted quite deep in the soil or leaf mold, and specimens are often picked leaving the very important character of the volva in the ground, and then the plant might easily be taken for the common mushroom, or more likely for the smooth Lepiota (Lepiota nausina), which is entirely white, the gills only in age showing a faint pink tinge. It is very important therefore, that, until one has such familiarity with these plants that they are easily recognized in the absence of some of these characters, the stem should be carefully dug from the soil. In the case of the specimens of the deadly amanita growing in the lawn on the campus of Cornell University, the stems were sunk to three to four inches in the quite hard ground.

The exact chemical nature of phallin, an extremely toxic substance, is not certainly known, but it is generally conceded to be of an albuminous nature. That it is an extremely

deadly poison is shown by the fact that .0015 grain per 2 lbs, weight of the animal is a fatal dose for cats and dogs. It is the active principle of the most deadly of all musbrooms, the Amanita phalloides, or death-cup fungus. We quote again from Mr. Chesnut's account of phallin and its treatment: "The fundamental injury is not due, as in the case of muscarin, to a paralysis of the nerves controlling the action of the heart, but to a direct effect on the blood corpuscles. These are quickly dissolved by phallin, the blood serum escaping from the blood vessels into the alimentary canal, and the whole system being rapidly drained of its vitality. No bad taste warns the victim, nor do the preliminary symptoms begin until nine to fourteen hours after the poisonous mushrooms are eaten. There is then considerable abdominal pain and there may be cramps in the legs and other nervous phenomena, such as convulsions, and even lockjaw or other kinds of tetanic spasms. The pulse is weak, the abdominal pain is ripadly followed by nausea, vomiting and extreme diarrhoea, the intestinal discharges assuming the 'rice-water' condition characteristic of cholera. The latter symptoms are persistently maintained, generally without loss of consciousness, until death ensues, which happens in from two to four days. There is no known antidote by which the effects of phallin can be counteracted. The undigested material, if not already vomited, should, however, be removed from the stomach and intestines by methods similar to those given for cases of poisoning by Amanita muscaria."

Prof. Chesnut, writing in regard to the poisonous effect of this species, says that:

The *phallin* spoken of is one of the toxalbumins, an extremely virulent poison found in poisonous animals especially the rattlesnake. These toxalbumins are allied to those found in diphtheria and other discases produced by bacteria.

Other species reported as poisonous or probably poisonous are A. flocco-cephala, and A. cothurnata. Ford has shown that A. spreta, and A. virosa, A. strobiliformis, A. chlorinosma, A. radicata, A. porphyria, and A. rubescens are poisonous. The A. verna, a small spring form of A. phalloides, is also very poisonous.

Ford reports nearly 200 deaths since 1900 from this fungus in France, Germany, Italy, and England. Ford gives the pathological changes described by Maschka to be as follows:

- 1. Lack of post mortem rigidity.
- 2. Widening of the pupils.
- 3. Failure of blood to coagulate and a cherry-red color.
- 4. Ecchymoses and hemorrhages in the serous membranes and parenchyniatous organs.
 - 5. Dilation of the bladder with urine.

Studor, Sabli and Schören found extensive necrotic and fatty changes in liver, kidney, heart and voluntary muscles. The amount of fat in the liver is nearly as great as in phosphorous poisoning.

Clinical symptoms. Often latent period of from 6 to 12 hours during which the victims remain quite well. They are suddenly seized with terrible abdominal pain, excessive vomiting and thirst. Diarrhoea may set in with mucous bloody stools, or there may be constipation. The paroxysm of pain may be so severe as to result in a peculiar hipprocratic facies. The patients rapidly lose strength. In 3 to 4 days in children and 6 to 8 days in adults, coma develops, from which the patients cannot be aroused. Cyanosis and lowered temperature precedes the fatal exit. Ocular symptoms and convulsions do not ordinarily occur, but convulsions may be present on a terminal event. The mortality varies from 60 to 100 per cent.

Kobert obtained from A. phalloides a substance with marked hemolytic action, the dried extract dissolving ox blood 1 to 125,000. To this extract he gave the name phallin, which he considered a toxalbumin.

Later this author * found what he thought was a poisonous alkaloid, "that all typical forms contained an alcohol soluble poison;" that phallin was occasionally absent.

Ford found that the extract of the fungus is a powerful hemolytic agent and quickly destroys the erythrocytes of guinea pig, fowl, pigeon, dog, goat, and man. This takes place at 37 degrees C., slower at lower temperatures. The corpuscles of sheep, beef, and swine are resistent. Raw and boiled milk act as an antidote - they are antihemolysins. Animals may be immunized by using non-lethal doses.

Since the above has been written there have come to hand several recent papers by Dr. W. W. Ford, who states that he found muscarin in several "yellow Amanitas" found in New York and Massachusetts. The aqueous extract of Amanita muscaria first agglutinated and then slowly dissolved blood corpuscles. The agglutinin was heat resistant. The extracts produced hemolysis. The agglutinin is a glucoside. The Amanita solitaria also contains an agglutinin. The Amanita frostiana 2 contains a moderately hemolytic substance and free from resistant toxin and muscarin. By the same author the poisonous nature of a number of species is reported as follows. The A. phalloides produces a chronic intoxication in guinea pig, the animal dying in twenty-five days. The lesion is typical for amanita toxin. It is hemolytic for rabbit's corpuscles, in a dilution of 1-20. The poison from A. virosa has a hemolytic strength of 1-200 in two hours and in dilution 1-100 at the end of 24 hours, and when heated to 60° C. Kills guinea pigs in twenty-four hours, with signs of acute intoxication. The A. spreta contains hemolysin and toxin but in rather a low degree. It should be classed with the deadly poisonous mushrooms. The A. porphyria, A. strobiliformis, A. radicata and A. chlorinosma are all poisonous and contain a heat resistant substance which induces in animals a chronic intoxication; the A. vittadini and A. rubescens should also be included according to Kobert.3

Dr. Ford in speaking of the poison in A, phalloides says:

"In a series of investigations published from the John Hopkins University it has now been shown that Amanita phalloides contain two poisons which for the sake of clearness we speak of as the amanita-toxin.4 The hemolysin is probably the same hemolytic substance which Kobert had in his preparation of phallin and the toxin is possibly identical with Kobert's second poison. The hemolysin was found in every specimen of Amanita phalloides which has thus far been examined, and when obtained from the fresh plant is the most powerful hemolysin of vegetable origin known. Drs. Abel and Ford 5 have shown that all coagulable proteid can be removed from this substance by uranyl acetate in alkaline solutions and by freshly prepared metaphosphoric acid, and when thus freed from proteid it continues to act upon blood corpuscles and gives the reaction of a glucoside containing a pentose. We have recently 6 developed a method for the isolation and purification of this glucoside which has an activity of 1-300,000 in the pure state. Since its sensitiveness to heat and the digestive ferments the hemolysin is precluded from playing any important role in human intoxication. We are inclined to believe that the amanita-toxin is the active principle, and Schlesinger and I7 have shown that this poison can be isolated by certain well-defined methods. It also is one of the powerful organic poisons, four-tenths of a milligram killing

^{*} Sitzungsb. Naturforschenden Gesellsch Rostock 1899:26. Statement from Prof. Ford.

1 The distribution of Poisons in Amanitas. The Jour. of Pharma. and Ept. Therapeutics.

1:275-284.

Notes on the Amanita-Toxin. Ford and Prouty. The Jour. of Pharm. and Expt. Therapcutics. 1:389.

2 Jour. Inf. Dis. 4:437.

3 Lehrbuch des. Intoxikationen. Ed. 2. 617.

4 Ford. Jour. Expt. Med. 8:437, May 26. 1906.

5 Abel and Ford: Arch. f. Exp. Path. u. Pharmakol. Supplement-Band Schmeideberg Festschrift, 1908. 7 Schlesinger and Ford: Jour. Biol. Chem. 3:279, Sept. 1907.

a guinea-pig in twenty-four hours. The amanita-toxin contains no proteid, does not respond to any alkaloidal reagents, and on fusion with potassium hydrate gives off idol and pyrol. At first thought to be a congugate sulphate, I have recently found in association with Mr. Prouty that this opinion is incorrect. We hope to ascertain the more exact characterization of this poison shortly."

Lepiota. Fr. Lepiota Toadstool

Plant with fleshy stem which can easily be separated from the cap; gills usually free from the stem; in some species the top of the cap breaks from the scales which adhere; volva absent. A small genus widely distributed. Some species are edible; L. procera is said to be excellent as food.

L. Morgani. Pk.

A large fleshy plant, sometimes a foot across the cap, with a thick stout stem and a ring removed a little distance from the gills; the pileus, when fully expanded, whitish, with dark scales; the spores and gills greenish.

Distribution. From Ohio southward and westward in grassy places, sometimes forming large fairy rings.

Poisonous properties. This plant is quite harmless to some people, but to others it causes very unpleasant symptoms. It should be eaten with caution.

Russula. Pers. Russula

Cap red, purple, violet, pink, blue, yellow, or green; pileus fleshy, convex. readily expanded and at length depressed; stem brittle, stout and smooth, spongy within and confluent with the cap.

Russula emetica. Fr.

Pileus fleshy, quite viscid, expanded, polished, shining, oval or bell-shaped when young, rose-red to yellow or even purple; margin furrowed, flesh white; gills free, equal, broad, distinct and white; stems stout, solid, or occasionally spongy; spores spherical.

Distribution. Widely distributed in North America. Found in pastures and under trees. Readily distinguished by viscid cap and color. Mr. Hand states that it is easily recognized by its acrid taste and free gills.

Poisonous properties. Mr. McIlvaine says that he has repeatedly eaten them and referred to a number of others who have also eaten them without any bad results, but Hand thinks that their acrid taste is against their use or rather cautions their use. Prof. Ford states that they cause profound gastro-intestinal disturbances, such as attacks of vomiting and diarrhoea, recovery only after thorough emptying of the stomach.

Volvaria. Fr. Volvaria

Universal veil forming a perfect volva, separate from outer part of the pileus; stem readily separated from pileus; gills free, at first white, then pink, and then reddish, and soft.

Volvaria bombycina. (Pers.) Fr.

This plant has a silky lustre; pileus is from 6 to 8 inches broad, globose, becoming bell-shaped, convex and somewhat umbonate; flesh white; gills crowd-

ed and flesh colored; stem is 6 to 8 inches long, tapering upward; spores rosy, smooth in masses and elliptical; volva large and somewhat membranaceous.

Poisonous properties. According to many authorities, this plant is edible, and it is likely that this and many other species can be eaten without serious trouble, although Gillot,* states that several species of this genus have caused death when eaten, though nothing is known of poisonous principle.

Inocybe Fr.

In the genus *Inocybe* there is a universal veil which is fibrillose in character, and more or less closely joined with the cuticle of the pileus, and the surface of the pileus is therefore marked with fibrils or is more or less scaly. Sometimes the margin of the pileus possesses remnants of a veil which is quite prominent in a few species. The gills are adnate, or sinuate, rarely decurrent, and in one species they are free. It is thus seen that the species vary widely, and there may be, after a careful study of the species, grounds for the separation of the species into several genera. One of the most remarkable species is *Inocybe echinata* Roth. This plant is covered with a universal veil of a sooty color and powdery in nature. The gills are reddish purple, and the stem is of the same color, the spores on white paper of a faint purplish red color.

Inocybe infida.

This is slightly larger than Panaeolus papilionaceus, with semiorbicular cap surmounted by a prominent nipple, which is dark reddish-brown, while the rest of the upper surface is light tawny-brown. The upper surface also differs from that of the non-poisonous kind in being silky-scaly and shining. The lower surface differs in being much lighter, pale yellowish instead of brownish-black, and the spore-print is about the color of oak wood.

Poisonous properties. Dr. William A. Murrill has recently contributed an account on the poisoning from Inocybe infida, a plant which closely resembles the Panaeolus papilionaceus. It appears that Dr. Deming of West Chester, who poisoned himself and other members of the family, describes the following symptoms: The fungi were gathered in the morning just before dinner. They were stewed and served on toast at one o'clock; he ate about half a slice of toast with mushrooms, drank some tea, and ate one-half a stuffed egg, with lettuce and mayonnaise dressing and after dinner smoked one-half a cigarette. Soon after he began to feel "queer," then there followed a fullness in the head and a rapid heart action as if he had taken nitroglycerin, this was followed by a sweat, his clothing becoming wet, and at the same time there was no nausea or prostration; his mind became a little bit confused. He then washed out the stomach, took castor oil and before the oil operated there was pressure and almost pain in the lower bowel. By evening he was as well as ever except somewhat exhausted. It appears that four other persons were affected with disagreeable symptoms from the eating of the mushroom.

Dr. Deming says: "In my case the beating of the heart, fullness of the head and sweating were very marked, though I ate about half as much as the others."

Dr. Murrill says that there is nothing to suggest an irritating poison and that it is probably not narcotic.

^{*} Etude medicale sur l'empoisonement par les champignons. Lyon, 1900.

PHALLINEAE

Mycelium consists of branched strands matted together; from this is produced an oval body consisting of an outer wall, the peridium, and an inner peridium; between the two is a layer of gelatinous material; the outer portion of the oval body forms the volva; the central portion pushes through the peridium with a long cellular stalk, the upper one bearing the cap-shaped gleba; the spores are brown on club-shaped basidia, surrounded by a mucilaginous material giving off an offensive odor. This sub-order contains the Clathraceae and Phallaceae. The Phallus impudicus and the Mutinus caninus have been regarded as suspicious.

CLATHRACEAE

Receptacle latticed or irregularly branched; gleba enclosed by the receptacle. The following genera of this order are known to occur in the United States, chiefly in the southern states: Clathrus, Phallogaster, Simblum, and Anthurus. Dr. Farlow* is authority for the report from Gerald MacCarthy to the effect that in North Carolina hogs had been killed by eating Clathrus columnatus which a correspondent, Mr. G. W. Lawrence found growing in oak woods near Fayetteville. The animals died within twelve or fifteen hours after eating the fungus. According to Gillot, hogs are poisoned by these and by Phalloideae.

PHALLACEAE.

Receptacle tubular or cylindrical with an external gleba. The common Stinkhorn Phallus impudicus has a thick hollow stalk of whitish color perforated with pores; the upper part is honey-combed, resembling the morel. During the early stages, an egg-shaped body may be seen coming from a mass of white mycelium. The egg-shaped body is more or less mucilaginous and contains the stalk and gleba, the latter becomes exposed later. Flies, attracted by the carrion-like odor and mucilaginous material of the gleba, scatter the spores and, apparently, are not poisoned. The fungus, however, is usually regarded as poisonous as are several related genera and species such as Mutinus caninus. The common Stinkhorn (P. impudicus) was formerly used as a salve in gout.

HYMENOGASTRINEAE

This contains the family Hymenogastraceae. The sub-order Lycoperdineae includes two families, Tylostomataceae and Lycoperdaceae.

LYCOPERDACEAE

Fruiting bodies globular, oval or pear-shaped, solid and fleshy, often of great size; before maturity, a dense white mass of homogenous hyphae occurs; the fruit is surrounded by a peridium, in some cases double; the interior is made up of branched threads called the capillitium, containing the spores; fruiting bodies break open in various ways at maturity. This group contains several interesting families. Many of the Lycoperdaceae are well known; among these are the Earth-star (Geaster), the Lycoperdon giganteum and

^{*} Farlow, W. G. Poisonous Nature of Clathrus columnatus. Bot. Gaz. 15:45-56. See also Halsted, B. D.., Rept. N. J. Agrl. Exp. Sta., 1894:417.

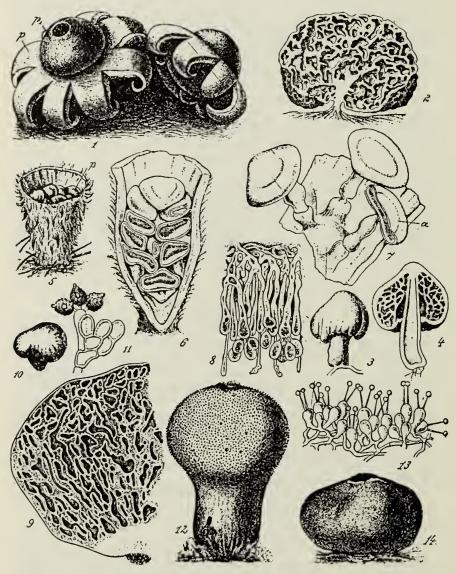


Fig. 74. Puff balls and their Allies. Gasteromyeetes. 1. Geaster fimbriatus, p Outer peridium, p1 Inner peridium. 2. Gautiera morchallaeformis, sectional view of fruiting body. 3. Secotium erythrocephalum. 4. Sectional view of the No. 3. 5. Bird's Nest Fungus (Cyathus striatus), p Peridia of spore bearing body, the outer peridium open on top showing attachment of fruiting bodies. 7. The same showing three fruiting bodies attached to wall. 8. Crucibulum vulgare showing hymenium and spores. 9. Hymenogaster tener, sectional view of fruiting body x 3. 10. Same, natural size. 11. Basidia with spores of No. 9. x 450. 12. Puff-ball (Lycoperdon sp.), natural size. 13. Part of hymenium of L. excipuliforme with basidia and spherical spores. 14. Common Lead-color Puff-ball (Bovista plumbea), natural size. 1 after Kerner, 2 after Vittadini, 3, 4, 6, 7, 9,-11 after Tulasne, 8 after Sachs, 12-14 after Wettstein.

Dr. Miquel lists the Lycoperdon Bovista as poisonous; this and Lycoperdon cyathiforme are edible when fresh, but poisonous when the plants are mature.

NIDULARIACEAE

This is allied to the above and contains the Bird's Nest fungus. (Crucibulum vulgare), which occurs upon wood and manure, and the Cyathus striatus. The false truffle (Scleroderma vulgare) belongs to an allied order.

ASCOMYCETES

Mycelium many celled, branched; reproduction both sexual and asexual; spores known as ascospores, limited in number. Arranged in two divisions, the *Hemiasci* and the *Euasci*.

Hemiasci

Parasites or saphrophytes; reproduction generally asexual, in fertilization, the contents of the antheridium and the oögonium fuse.

HEMIASCALES

An unimportant group with three orders, Ascoideaceae, Protomycetaceae, which contains some plants that are parasitic, Protomyces macrosporus, upon the numbers of the carrot family.

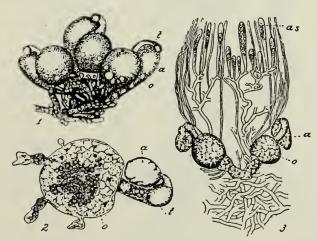


Fig. 75. Fertilization of Pyronema confluens. 1. Three oogonia (o) with fertilizing processes (t) a—antheridia. 2. Oogonium after fertilization, with numerous nuclei. 3. Part of fruiting body, the ascogonium forming hyphae (as), (a) antheridium, (o) oogonium. 1-3 greatly magnified. After Harper.

The family Monascaceae contains one fungus which has been found in mouldy corn and silage in Iowa, the Monascus purpurens Went. It is related to the M. heterosporus (Harz) Shröter, which was found by Harz in a soap factory. The coloring matter from M. purpurens, known as "ang-quac," is used in Eastern Asia as a pigment, being produced by the growth of the fungus on rice. The fungus consists of a mass of septate hyphae, producing conidia and perithecia with numerous asci; the ascospores are from $5-6.5 \,\mu$, in diameter. The

details of the structure of this fungus have been given by Olive, Barker, and Ikeno 3 and in a paper to be published by Dr. Buchanan. Dr. Buchanan found this species in spoiled corn silage, which was responsible for the death of several horses in Iowa. This species possibly has been the cause of the disease, this fungus occurring only where air had access to the silage. The fungus found by Harz produces a mycelium similar to the preceding with thick-walled swellings and color white or carmine red; conidia ellipsoidal, spherical, oboyate, of two kinds, the smaller 2.5-3 μ . to 7-8 μ , occurring in chains or singly, the larger occurring singly 9-11 \(\mu\) in diameter, and arising from lateral branches; sporangia from short lateral branches are spherical 40-53 μ in diameter, many spored; the sporangia are surrounded by branched hyphae, ascospores spherical or oval, colorless 4-5 \(\mu\) in diameter; conidia and hyphae contain a carmine red pigment physomycin.

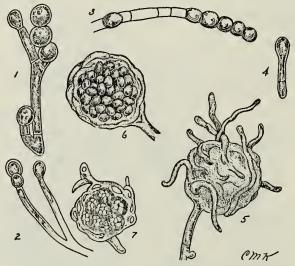


Fig. 75A. Corn Silage fungus (Monascus purpureus.) 1, 2, 3, Conidiophores with conidia; 4, germinating conidium; 5, sterile hypha covering of perithecium sending out branches, these are sometimes tipped with conidia; 6, optical section of mature perithecium, spores still within asci. Found in corn silage by Dr. Buchanan.

Euasci

Asci with definite number of spores, usually 2, 4, 8, 16, 32; seldom, but occasionally 1-celled.

EUASCALES

Contains the yeast plant, peach curl, plum pocket, ergot, blue mould, powdery mildews, etc.

PROTOASCINEAE

Asci single, in one group, without distinctive development of the mycelium; in the other with a distinctive mycelium bearing the asci with their spores.

¹ Annals of Bot. 17:167, pl. 12 &13.

² Bot. Gazette. 39:56. 3 Ber. deutsch Bot. Gesellsch. 12:259.

SACCHAROM YCETACEAE.

Vegetative cells single or in small groups; mycelium usually not evident, reproduction, by budding; ascospores, usually 4, produced in the cell; occasionally 8, 3, 5, or seldom 1.

The Saccharomycetes are fungi important in the process of fermentation. It is only in recent years that any parasitic species has been recognized. Metchnikoff, in 1884, found a parasitic yeast Monospora bicuspidata in Daphnids. Raum and Neumayer in 1891 declared yeasts were pathogenic. Busse, 1894, demonstrated that certain yeasts were pathogenic. Tokishige about the same time observed a yeast pathogenic for horses. Sanfelice isolated from the cancerous-like growth of an ox a Saccharomyces which was pathogenic for guinea pigs. The same author found another species in pigeons. Lydia Rabinowitsch studied 50 species of yeasts, of which 7 proved to be pathogenic. In 1895 Prof. Curtis found the second case of Saccharomyces in a young man; clinically the disease resembled a myxosarcoma. It is doubtful whether these forms are true yeasts. Some of these appear to be Hyphomycetes rather than Saccharomycetes. I have therefore discussed these under the form genus Oidium.

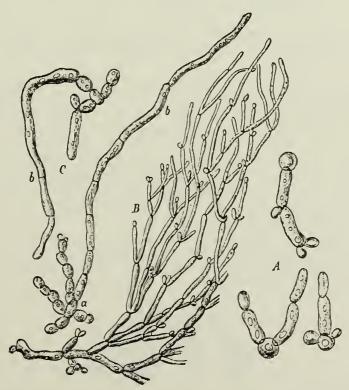


Fig. 76. Yeast. Saccharomyces mycoderma. A. Process of germination. B. Mycelium budding in a weak nutrient solution. C. (a) Yeast-like form budding; (b) long cells.

Saccharomyces. Meyen. Yeast

Vegetative cells spherical, ellipsoidal, oval or pear-shaped, occasionally elongated mycelial like; asci spherical, ellipsoidal or cylindrical with 1-8 ascospores 1-celled spherical or ellipsoidal. About 40 species. The S. apiculatus, Rees, is important in the fermentation of fruit. The S. ellipsoideus causes the fermentation of wine. The S. mycoderna, Rees, forms a white mass on cider, wine, cucumbers, etc., and prepares the way for the acetic acid fermentation. The S. kefyr, Beyerinck, along with Bacillus acidi-lactici and other bacteria is found in Keyfr grains. S. ghitinus Fres., the pink yeast, is found growing on nutrient media in laboratories.

Saccharomyces cerevisiae, Meyen. Common yeast

Vegetative cells, spherical or oval, 8-10x8-12 μ singly, or in several, budding chains with one or more vacuoles; asci spherical or short elliptical 11-14, generally with 4 ascospores tetradform. It produces a white growth on gelatine and potato, does not liquify the gelatine; causes fermentation of grape sugar, maltose and cane sugar. The biology of the fermentation of beer is as follows: Barley, which is ordinarily used for this purpose, is allowed to germinate; during the process of germination the starch, by means of diastase, is converted into sugar, the sugar being afterwards removed with the water; this sugary fluid is then placed in large vats in dark rooms at a comparatively low temperature; the yeast plant is added and fermentation starts.

The fermentation of sugar is due to an enzyme found in the yeast plant, to which Buchner has given the name of Zymase. This enzyme breaks the sugar into alcohol and carbon dioxid. It is not necessary to have the living organism present to produce this fermentation, as a quantity of the yeast extract mixed with the solution of fermentible sugar will produce at the end of some days a small amount of alcohol. The enzyme decomposes very rapidly. Reynolds Green, in his book on fermentation, says:

From these researches it appears certain that the production of alcohol whether in the presence or absence of oxygen is brought about by the activity of an enzyme. Its secretion by the cells of yeast attends the ordinary nutritive processes as well as the abnormal decompositions set up by incipient asphyxiation. The latter condition induces its formation in other parts of plants. The absence of oxygen stimulates the protoplasm of the cells to secrete it, the ultimate effect of its appearance being the liberation of energy as already stated.

Distribution. Widely distributed.

Poisonous properties. The chemical composition of alcohol is C₂ H₅ OH.

Different alcoholic drinks contain different percentages of alcohol. Ale and beers contain from 4 to 8 per cent together with bitters and malt extract; cider from 5 to 9 per cent; sherry from 15 to 20 per cent.



Fig. 77. Yeast. Saccharomyces cerevisiae. Ascospores in cells. Spores at f. Magn. 1000. After Hansen.

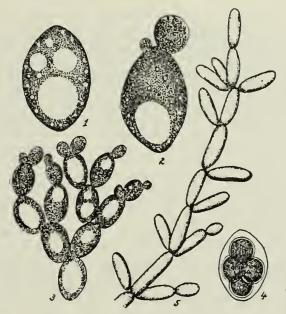


Fig. 78. Yeast. Saccharomyces cerevisiae. 1. Single cell with vacuoles. 2. Cells budding x 1500. 3. Numcrous daughter cells x 1000. 4. Cell with ascospores x 1200. 5. S. ellipsoideus. 1-4 Modified from Luerssen and Rees; 5 after Hansen.

Alcohol in its action is a germicide and when applied to the raw surface or wounds it is a stimulant and local anaesthetic, while in concentrated form it is an irritant and even caustic. When placed upon surfaces of the broken skin it causes cooling and contraction of the superficial blood vessels. When absorbed it hardens the tissues. Internally it causes a secretion of saliva and the heart is stimulated by the irritative action of alcohol.. In large amounts it destroys the peptic ferment. Dr. Winslow says:

Alcohol is essentially a heart stimulant and the most valuable one we possess. It makes the beart beat more forcibly and rapidly, and also increases blood pressure, despite the fact that, normally, alcohol causes dilation of the arterioles. In weakened bodily conditions, with vascular relaxation, alcohol may increase vascular tonicity. The heart and blood vessels are paralyzed by poisonous doses of alcohol and blood tension falls tremendously.

The local effect of alcohol upon the peripheral nerves resembles the action after absorption upon the system generally. The nervous system is affected in nearly the same order and manner as by anaesthetics, and the same stages may be observed. The stages include the stimulant, depressant and paralytic. The law of dissolution is demonstrated by alcohol, as the more highly organized centres and those more recently developed in the process of evolution are first to succumb, and in following out this order, the medulla, the first of the higher centres to be developed, is the last to be influenced by the drug. In accordance with this law the cerebrum is first acted upon. The period of excitement is brief and is due in a considerable degree to the increased cerebral circulation and flushing of the brain. It is essential to emphasize the fact that by far the most apparent and decided action of alcohol is one of depression upon the nervous system as a whole. The stimulating influence of alcohol upon the spinal centres is more marked in the lower animals than in man because the brain is proportionately small and poorly developed in the former. The primary stimulating effect of alcohol is shown in man by increased mental activity and apparent brilliancy, but acute reasoning and judgment are not enhanced, and in many cases there is almost immediate mental confusion and drowsiness induced.

EXOASCACEAE

This small order contains parasitic and saprophytic species. The asci are without perithecium, except in Gymnoascus and Ctenomyces where there is a rudimentary perithecium. The Taphrinae are undoubtedly related to the yeasts and by some are placed in one order known as Gymnoasceae, being represented by Gymnoascus. The Gymnoasceae exclusive of Taphrinae are sometimes placed with the Plectascineae, a fungus occurring on the dung of horses and producing simple-fruiting organs, which consist of short-branched filaments arising either from a single hypha in which a cell is cut off, or several, one being spirally wound about the other. This becomes the ascus, which contains the ascospores. In Eremascus the ascus-producing part resembles certain zygospores. Ctenomyces the ascus is surrounded by simple torulose hyphae, representing a rudimentary perithecium. The Eidamella spinosa described by Matruchot and Dassonville is allied to Gymnoascus. It produces numerous ovoid short stalked asci with 8 ovate colorless ascospores. Parasitic on dog. The life history of parasitic members of this order may be represented by Exoascus pruni. This fungus grows on the fruit of various species of the genus Prunus, producing in plums what is known as plum pockets or bladder plums. The fungus, when fully developed, consists mainly of a single layer of palisade-like asci, which produce their branching mycelium in the parenchyma of the affected part, and later develop between the outer walls of the epidermal cells and cuticle. Here

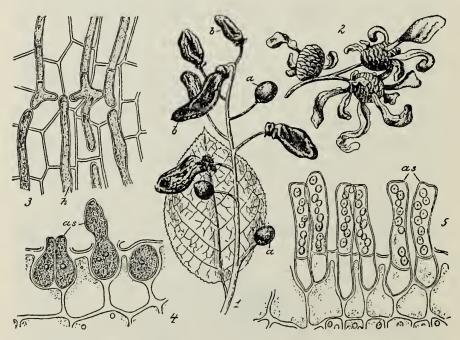


Fig. 79. Exosceae. 1. Plum pocket (Exoascus Pruni), on Prunus Padus. a. Normal fruit; b, abnormal fruit. 2. E. Alni-incaniae on alder (Alnus incana); scales enlarged. 3-5. E. alnitorquus; 3. Surface view, alder leaves showing hyphae (h) between cuticle and remainder of epidermal cell. 4. Formation of asci (as). 5. Ripe asci with ascospores x 100. 1-2 after Wettstein. 3-5 after Sadebeck.

they grow and spread out to the surface, forming a single layer of cells, each cell swells, the cuticle becomes ruptured and a palisade-like layer of asci is formed. There are eight ascospores in each ascus which escape by means of an opening at the tip. The ascospores of Taphrina frequently germinate in the ascus, budding like yeast and in this budding condition they produce a small amount of alcohol. Another troublesome species is the Peach Curl (Exoascus deformans) which occurs on the young leaves of peaches. The E. Cerasi is another destructive species producing the "Witches Brooms" of the cherry Prunus Cerasus. The E. Betulinus produces the "Witches Broom" in the Birches.

PEZIZINEAE, HELVELLINEAE, TUBERINEAE, PHACIDIINEAE

Mycelium well developed; asci borne upon large fruiting bodies and a continuous hymenium consisting of the asci, sterile threads, the paraphyses. Contains a number of common cup fungi like the Peziza, Morchella esculenta, and Hellvella. A Sclerotinia produces a disease on red mangolds, beans, and hemp; Sclerotinia also occurs upon clover and the common brown plum rot, (Sclerotinia). Helvella suspecta with a reddish brown pileus and a dirty flesh colored stalk is suspected of being poisonous. It has a nauseous, sweetish taste, and produces hellvellic acid, a hemolytic, or blood destroying substance. The Gyromitra esculenta also produces helvellic acid and is regarded as poisonous. It owes its toxicity to the blood making properties. The Hellvellas, Morels, and Sclerotinia belong to the family, Helvellaceae. Tuber produces tuber like bodies found in the soil. The organism is parasitic on trees. The fruiting bodies are enclosed by a peridium which consists of corrugated, smooth, or wart-like excrescences. The hyphae are compact. The ascospores occur in winding passages in the interior. Some of the Hellvellas and Morels are edible. The truffle (Tuber aestivum) of the family Eutuberaceae are edible.

The family *Phacidiaceae*, contains one important parasite of the alfalfa, the *Phacidium Medicaginis*. The diseased leaves turn yellow and soon fall. The yellow leaf, or in some cases the green leaves, contain the small blackish or brownish specks usually upon the upper side of the leaf, the injury extend-

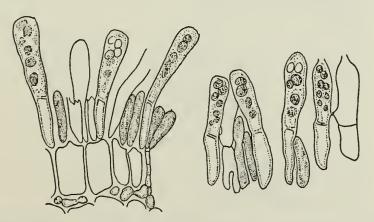


Fig. 80. Enlarged plum branches. Exoascus communis on Prunus maritima, projecting beyond the mass are the asci, some of which contain the spherical ascospores. After Atkinson,

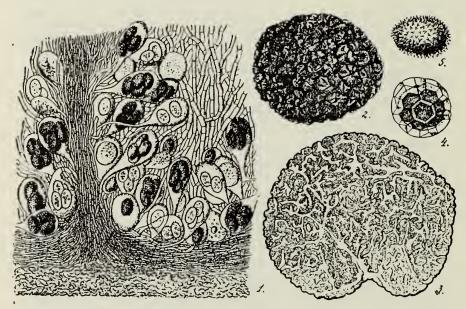


Fig. 81. Tuberaceae. Truffles. 1. Tuber rubrum, Part of interior of a truffle, showing hyphae, asci, and ascospores, greatly magnified. 2. T. aestivum, fruiting hody. 3. T. brumale, section of truffle. 4. Ascospore of T. Magnatum. 1, 3, 5, after Tulasne. 2 after Wettstein.

ing to the lower surface. The spot contains a small pustule called the apothecium, which is cup-shaped. This cup-shaped body contains the asci (sacs) in which 8 small spores occur, the ascospores with the ascus, the two slender threads are known as paraphyses. This fungus is a serious parasitic disease of the alfalfa. The fungus does much injury to the fodder and it is not improbable that at times may be injurious to animals consuming the fodder.

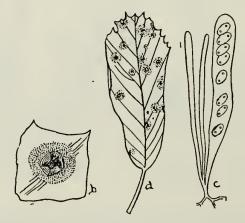


Fig. 82. Enlarged leaf showing spots. b. Single spot enlarged: c. Ascus with ascospores, paraphyses coming from mycelium.

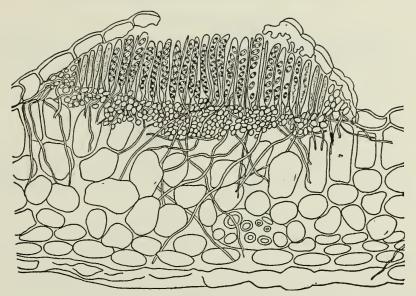


Fig. 83. Section through apothecium found on leaf; the asci, ascospores and mycelium. Combs.

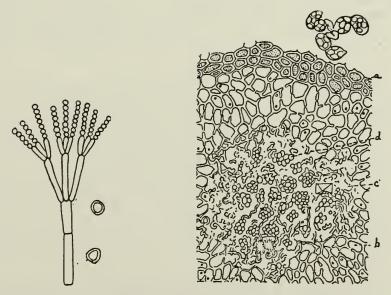


Fig. 84. Common Blue Mould (Penicilium glaucum). 1. Conidiophore, spores in chains. 2. Sclerotium or hard compact mass of fungus (threads hyphae) with asci and ascospores. Asci and ascospores shown above. Brefeld.

PLECTASCINEAE

Generally saprophytic fungi with a well developed mycelium, either buried in the sub-stratum or superficial. Reproduction sexual or asexual; asci either borne directly on the mycelium or in closed fruiting bodies, called perithecia.

ASPERGILLACEAE

Peridium thick; perithecia small; the sexual reproduction may be seen from the development as it occurs in the Blue Mould (*Penicillium*).

Penicillium. Link

A branched septate mycelium; conidiophores with septa, numerous branches near the apex; contains small flask-shaped sterigmata; spores borne in chains; conidiophores sometimes in bundles, as in the old Coremium; asci develop in poorly-lighted places in a sclerotium-like body.

Penicillium glaucum. Link. Blue Mould

At first a white mycelium spreads over the surface or through the substratum; the mycelium, through an enzyme action, undoubtedly, dissolves the starch; raised masses are formed on the surface, which consist of masses of mycelium thread strands; the strands send out lateral branches from the end of which a whorl of short branches appears, which give rise to one or more whorls; from the ultimate branches a chain of small spores is produced, the last one on the chain being the oldest.

The ascospores have not been found in corn, but occur in poorly lighted places and are produced in the absence of oxygen. The spores produced in chains germinate when the required amount of moisture and heat is present, so that unlimited numbers of generations may proceed from a single spore. These spores also preserve their vitality for a considerable length of time.

Brefeld has shown that they will germinate though kept in a dry place for several years. The organism grows at various temperatures, from near the freezing point to a considerable heat. It also resists antiseptics. It is one of the most troublesome fungi in stored fruit.

Penicillium glaucum is an organism which contains diastase, maltase, emulsin and a ferment which inverts cane sugar. Calcium oxalate is deposited in the perithecia. Under certain conditions mannite is said to be produced. When the Penicillium glaucum occurs in grape must it delays fermentation.

Distribution. The common blue mould is widely distributed in nature and is contained in a large number of the spores which drop in on fruits and decaying bodies and there germinate and produce fruiting bodies.

Poisonous properties. This fungus certainly is not pathogenic. It is widely distributed on decaying fruit; it has been suspected, in several instances, of being poisonous, but there is no evidence to support the supposition that this is the case or that it produces toxic substances. Under certain conditions it may, possibly, produce mycotic stomatitis. It has been found in sputum, nasal secretions, and in the stomach, but these cases are without special significance.

Penicillium minimum. Siebenmann

Mycelium at first white, flocculent, changing to blackish green when spores are formed; conidiophores slender, branching, bearing a chain of spores from 2-3 μ in diameter.

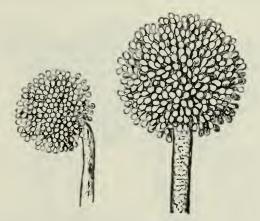


Fig. 85. Green Mould. (Aspergillus glaucus) on the left, A. repens on the right, both with conidia in chains, and conidiophores. After Siebenmann.

Distribution. Found in Europe.

Pathogenic properties. Found by Siebenmann in the ear.

Aspergillus. Micheli

Parasites or saprophytes with branched septate mycelium; reproduction sexual or asexual; in the asexual, conidiophores enlarged at the end, the enlarged portion bearing numerous small sterigmata, or these bearing smaller sterigmata; the conidia borne in chains; occasionally sclerotia form; perithecia small with asci and ascospores. The ascigerous stage of a few only is known. The life history of the common herbarium mould was first worked out by DeBary. A little known A. sulphureus is said to cause muscular contractions, and tubercular bodies.

Aspergillus glaucus. (L.) Link

Mycelium on or in the substratum forming a bluish green growth; conidia spherical or somewhat elliptical, slightly roughened, 6-15 μ in diameter, borne in chains attached to a short simple sterigmata; perithecia form little yellow masses; each ascus has 8 colorless biconvex ascospores 8-10 μ in diameter.

The life history of this fungus is as follows: This species is common in stored grain and hay. The mycelium spreads over the surface and through the substratum; it enters the kernel because of the dissolving action of an enzyme produced by the mycelium. From this mycelium erect threads (conidiophores or sporophores) arise which are enlarged at the end. From the enlarged portion of the conidiophores numerous small and radiating stalks (sterigmata) are produced, each bearing a chain of spores, the end spores of the chain being the older. These spores germinate under favorable conditions of moisture and heat, and again give rise to the same stage. In addition to this, the conidial stage, a second kind of reproductive body occurs. This is produced by the coiling of a branch of the mycelium having several turns. Two or three slender branches grow from the base. One of these grows more rapidly and connects with the top of the spiral coil formed first. The contents of those last formed

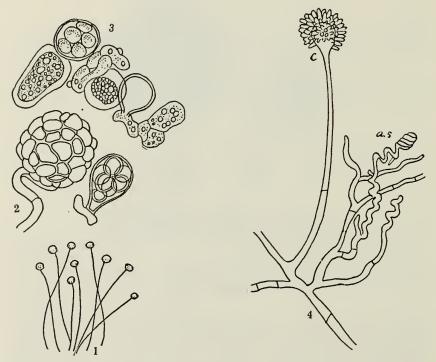


Fig. 86. Common Aspergillus. 1. General appearance showing long conidiophore and sterigmata on end. 2. Perithecia with one ascus and ascospores. 3. Contents from an unripe perithecium. 4. A small part of a mycelium with conidiophore c and spore-bearing sterigmata, young ascogonium a, s. All after DeBary except 1.

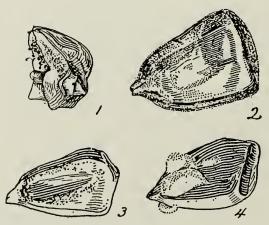


Fig. 87. Mouldy maize kernels, 1. Aspergillus (Sterigmatocystis). 2. Aspergillus. 3. Rhizopus, 4. Pencillium. Charlotte M. King.



Fig. 87a. Mycotic stomatitis caused from eating mouldy hay and parasitic fungi on forage plants. (U. S. Dept. Agrl.)

unite with the spiral known as the ascogonium. After fertilization a perithecium is produced, which contains the asci, each ascus being surrounded by a delicate wall and containing eight biconvex ascospores.

Asperigillus forms diastase and is capable of changing starch into dextrin and maltose.

Distribution. Widely distributed in nature on mouldy hay, corn and other grains.

Poisonous properties. The organism is not pathogenic but probably develops a poisonous substance which may produce disturbance. Dr. Law mentions a serious case, epizootic cerebro-spinal meningitis, in Pennsylvania, due to the feeding of mouldy timothy hay, which was badly fermented. In Cairo, Egypt, 6,000 horses and mules perished from the same cause. Michener attributes this disease to foods undergoing fermentation due to toxic fungi. Williams, of Idaho, thought also that the fermentation of alfalfa, timothy and wild grass hay produced the disease. Dr. Law says:

In all probability as we learn more of the true pathology of the disease, we shall come to recognize not one, but several toxic principles, and several different affections, each with its characteristic phenomena in the somewhat indefinite affection still known as cerebrospinal meningitis.

It occurs in horses, sheep, oxen, goats, and dogs, preferably attacking the young which have not become immuned to the toxic substance. It occurs most commonly in winter and spring when animals shed the coat. Dr. Mayo, who investigated this trouble in Kansas, says that a disease known as "staggers," "mad staggers," or, as he has termed it, enzootic cerebritis, is caused by feeding corn which is attacked by Aspergillus glaucus. The spores of the fungus gain entrance to the circulation, and find lodgment in the kidneys and liver. He supports his conclusions by experiments made by him on a guinea pig and a

young colt. He also quotes Kaufmann, who was successful in producing a disease with *Penicillium glaucum* and *Aspergillus glaucus*. There is considerable loss in many states from cerebro-spinal meningitis. In many parts of the country this is attributed, as I have said before, to mouldy corn. Dr. Bitting, of the Indiana Agricultural Experiment Station, made an investigation of this question and concludes that mouldy corn is not responsible for this disease. Upon an examination of mouldy corn he found several moulds and a bacterium. To test the poisonous properties of these, two horses were injected under the skin with five cubic centimeters. Later, larger amounts were given, and each animal was induced to eat as much as five pounds of the infected meal per day. One of the moulds as well as the bacterium gave negative results; the *Fusarium* produced a redness of the gums and some salivation. In no case did cerebrospinal meningitis result.

The results of the experiment show that inoculations with culture of the bacteria and moulds were ineffective. Eating of the mushes containing pure culture showed that only in the case of a growth of a species of Fusarium did any intestinal disturbance follow, and that in one case the feeding of the rotted grain produced considerable intestinal disturbance and some nervous symptoms, but that the disturbance was light in the other.

Grawitz succeeded in producing infection by adapting the digestive tract of the animals to an alkaline medium.

Roberts and Bitting say in regard to this trouble in Indiana:

It affects horses, cattle and sheep, but the cause is not known. This disease is reported in stables in the fall and winter. The reports indicate that about an equal number of horses and cattle become affected, but that they rarely become affected at the same time. The horses and cattle kept in the same barn and fed the same kind of food will not become diseased at the same time. Most of the cases occur while feeding ensilage or shredded fodder and thus it has come to be called ensilage disease and shredded fodder disease. The character of the food, however, is only an incident, for cases may occur when other spoiled or fermented foods are present, or when only the best foods are used. The disease is often ascribed to mouldy and rat-eaten corn, but our experiments with such foods and pure cultures of moulds from such foods were negative. Bad sanitation is also ascribed as a cause.

In regard to Mycotic Stomatitis of cattle which they attributed to moulds:

The particular organism causing the disease, if there be one, has not been described. It seems probable that the disease is due to more than one form of fungi which may be present on the pasture. The animals affected are cattle of all ages above 4 months. The disease is not contagious, but usually affects a number of animals in a given herd, and always while in pasture. The fact of a number of animals being affected is due to similar exposure and not to infection spreading from one animal to another. Attempts at direct inoculation have not been successful. The disease occurs in some localities every year, and in others seemingly under special climatic influences. I know a locality where it may he developed at any time by permitting cattle to graze along the roadside. The disease is much more prevalent on permanent blue grass than upon timothy pasture, and is of rare occurrence upon pastures used in a crop rotation. The disease develops in pastures allowed to grow for some little time without being used. It is particularly liable to develop a few days after a good rainfall succeeding a dry period.

The symptoms are inability to graze, saliva dribbling from the mouth, and frequent visitations to the watering trough, holding the mouth in the water as though it were burned. The animals appear to be hungry but cannot eat. The mouth is red and lips, guns and tongue swell. Blisters form and these soon give way to ulcers that may remain distinct or run together. In some of the aggravated forms the ulcers may unite so that when the crust comes off, it will make a cast of a lip or the whole end of the tongue. The crusts are usually from the size of a dime to that of a quarter. The tongue may swell to such an extent as to protrude from the mouth and the animal be unable to draw it inside. The muzzle may be increased one-half in size.

Dr. Craig, of the same state, reports somewhat similar experience.

Captain F. Smith, in his manual of Veterinary Hygiene, refers to the injuries from moulds, especially *Penicillium* and *Aspergillus*, calling attention to the brittleness of hay caused by fungi, and that the spores produce irritation to the respiratory passages. He states further that oats and bran have produced diabetes, paralysis, and subsequent death in horses. He refers to the case mentioned by Professor Varnell in which the horse died in three days from eating moldy oats. Professor Gamgee calls attention to the disease in France and Scotland in the years 1854 and 1856, due to horses feeding on grass which had become wet and musty. The animals suffered cerebral derangement, producing stomach staggers, so-called by English writers.

Aspergillus Oryzae. Alılburg

Rabbits inoculated showed convulsive symptoms; tubular foci occurred in the intestines. The Aspergillus Oryzac forms maltose and diastase and in Eastern Asia plays an important part in the manufacture of "sake" or rice beer, which has been a national drink of the Chinese for centuries.

Aspergillus malignum. (Lindt.)

Mycelium bluish gray, conidiophores short, the end pear-shaped, 22-24 μ wide; sterigmata branching, conidia in chains 3-4 μ in diameter; perithecia 40-60 μ , ascospores 6-8.

Distribution. Found in Europe.

Pathogenic properties. Grows best at the higher temperatures. Found by Lindt in the human ear.

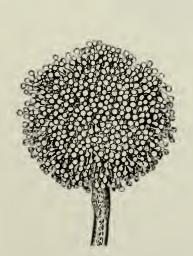


Fig. 88b. Pale Mould (Aspergillus flavus). Showing conidiophore and spores attached in chains. After Siebenmann.

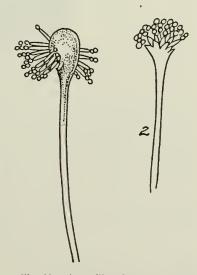


Fig. 88. Aspergillus Oryzae on rice.
1. Conidiophore, sterigmata and conidia
2. Young conidiophore. Modified by
Charlotte M. King after drawing by
Wehmer.

Aspergillus flavus. Link

Hyphae arachnoid, white; the fertile erect, slightly cespitose; conidia 5-7 μ in diameter, small, globose, vari-colored, slightly wart-like, collected about the white sub-globose, wart-like apex; apex finally becoming yellowish; sclerotium very small, dark.

Aspergillus fumigatus. Fresenius

Forms greenish or bluish gray masses on the surface of the substratum, conidiophores short with a semi-spherical mass $8-20\,\mu$ in diameter. Sterigmata bear the spherical conidia 2.5-3 μ in diameter, which are at first bluish green and later brown. Sclerotia unknown. Grows best at a temperature of $37-40^{\circ}$ C.

Distribution. Widely distributed.

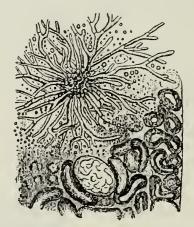


Fig. 89. Section of kidney of rabbit showing mycelium of an Aspergillus. After Grawitz.

Pathogenic properties. It has been known for some time that several species of Aspergillus are pathogenic for animals. In 1815 Mayer and Emmert found the fungus in the lungs of a jay. In 1826 it was reported in the long bones of a white stork by Heusinger, and numerous other cases in birds like the flamingo, duck, chicken, ostrich, and turkey, have been reported, especially in Europe. Kühn, in 1893, furnished quite conclusive evidence that certain species of Aspergillus can produce necrosis and disease. Chantemesse, at the tenth International Congress in Berlin, called attention to a disease of pigeons resembling tuberculosis which he said was produced by an Aspergillus. Saxer attributed mycosis to an Aspergillus, and, according to Sticker, the disease may appear sporadic and endemic, the latter to persons who feed pigeons and to the hair combers in Paris. It is spontaneous in horses, cattle, dogs, and birds, and is sometimes quite epidemic in birds. The form of the disease when it occurs in the lung is callel Bronchopneumomycosis; it appears that various species of Aspergilli also occur in connection with otomycosis, and occasionally in the nose or the eye. A very complete history is given by Drs.

Mohler and Buckley in the report of the Bureau of Animal Industry. Hughes Bennett reported a case in the sputum of a tubercular patient, and, in 1847, Sluyter reported definitely on the Aspergillus in the lungs of a human being. Virchow in 1856 reported several cases. In 1879, Leber first described a purulent keratitis due to aspergillus infection. Drs. Mohler and Buckley, in referring to the observations on pneumomycosis, say:

Dieulafoy, Chantemesse, and Widal, reported their observations and studies of pneumonomycosis as it occurs in a certain class of men in Paris. These men feed thousands of young pigeons daily hy taking into their mouths a mixture of grain and water which they force into the mouths of the birds much in the same way that the old pigeons feed their young. It had been a matter of common observation that these men were sufferers from a severe pulmonary disorder; but when their sputum was examined, instead of finding tubercle bacilli, only the threads of mycelia were detected. This observation was subsequently confirmed by Renon and other investigators. Until this time it had been held that the presence of fungi in the lung tissue was of secondary importance, but these observations dispelled further argument. Experiments on animals in which they were made to inhale the spores, were successful in producing the disease; thus it was that the natural infection was proved.

Renon, who made an exhaustive study of the subject, concludes as follows concerning aspergillosis:

- 1. That aspergillosis is a spontaneous disease affecting the bronchi and lungs of birds and animals, and creating in the animals a generalized affection similar to hemorrhagic septicemia; that it develops in eggs in incubation and may contaminate the embryos contained therein.
- 2. The discase may be transmitted experimentally. The botanical and cultural characters of the fungus and the lesions it provokes are truly specific. In its pathogenic action it bears a strong resemblance to tubercle bacillus.
- 3. In man it develops upon the cornea or skin, but has its particular evolution in the respiratory apparatus, creating pulmonary mycosis, resembling tuberculosis, and pulmonary gangrene, but without the fetid odor. It may coexist with tuberculosis. Occasionally it is fatal after the formation of cavities in the lungs. It may invade the bronchial apparatus alone, causing membranous bronchitis of special form and of long duration.
- 4. In all its manifestations Aspergillus fumigatus may play a primary or secondary role in both man and animals. It is not, therefore, a simple saprophyte, but a true parasite.

Renon points out the relation of the occupation of man to his contracting the disease. When animals and men are kept where the mould is common, as in hair assorting establishments where rye is used to disentagle the hair, they become affected with the disease. The handling of dusty grain and feeds may lead to infection from Aspergillus. Saxer also went into historical details giving his experiments with mycosis in man. In 1857 Aspergillus was observed by Rivolta in the pharyngeal abscess of a horse. Gotti observed it in an auricular catarrh of a dog. Pech observed mycotic pneumonia in seven horses, where they had been fed mouldy hay. Several cases where the Aspergillus occurred in the trachea of cows have also been reported. Pearson and Ravenel record a case of pneumonycosis of the lung of a cow.

Infection takes place generally by the inhalation of the spores. The spores germinate in the bronchial branches, develop a mycelium and produce conidiophores and spores on the surface. Drs. Mohler and Buckley, calling attention to the various aspergilli which have been found, say:

Numerous experiments have been tried with the various fungi, especially in relation to the best temperatures for their development and fructification, and it has been found that, although a few are able to germinate in the bronchioles, the Aspergillus fumigatus is about the only one which develops a vigorous growth there and fructifies, the temperature of the human body seeming to be quite suitable for this species. Most of the other molds develop at a much lower temperature and are therefore usually harmless even if introduced

into the lungs. But for the Aspergillus fumigatus the lungs act as a veritable propagating house, furnishing a moist, nutrient soil upon which to grow and a congenial, warm, moist atmosphere with a sufficient amount of oxygen for its demand to come to complete maturity and for fructification to take place. When the fungous growth is localized in the bronchial mucous membrane, the condition is known as bronchomycosis. It may be that the tissues are able to forestall entrance into their substance and finally the fungi die and recovery takes place. In birds the growth may extend to the air sacs; this condition is then called cytomycosis. Cases of cytomycosis are very rare; and when it does occur, emaciation of the birds is the predominating symptom. When the lung tissue itself is the seat of invasion, the term pneumonomycosis is applied. Invasion of the lung tissue by the mycelium is the occasion for an intense inflammatory disturbance with positive chemotaxis. However, this tissue reaction seems to offer the most trifling barrier to the parasitic encroachment in such weakly subjects as birds. Generali states that delicate breeds of pigeons are noticeably susceptible to this disease.

In regard to the symptoms in birds, he says:

The birds become listless, mope, and do not follow the rest of the flock. When made to run they soon become exhausted and fall and have great difficulty in breathing. Even when disturbed they appear very weak and gasp for breath, extending their heads and making movements as if choking. There is a great thirst, but a diminution or complete loss of appetite. The birds become rapidly emaciated, the wings are pendant, the eyelids droop, comb and wattles become quite pale, and a general dejected appearance follows. Usually there is an intense diarrhea which weakens the bird very much. In the experimental disease the diarrhoea is an accompaniment just as in that of a spontaneous development, The plumage is said to appear ruffled, and the respirations become croupy, even when the disease has not advanced very far; later they are more rapid and a rattling noise can be heard. In the final stages suffocation is threatened.

When the air sacs are affected very few symptoms manifest themselves, though emaciation is marked. As in any similar condition of the lungs, fever is high, and symptoms that would be manifested in pneumonia of fowls would, of course, show here. There is more or less catarrh of the trachea and bronchi, and if these alone were diseased there would probably be nothing to attract notice other than symptoms of bronchitis. Bleeding from the nostrils has been observed in man and in animals, and it may be that this would also occasionally be seen in birds. If the air spaces in the bones become affected, lameness with swelling of the joints may result. The duration of the disease is quite variable and death may take place in from one to eight weeks from asphyxia or marasmus. Duration depends a great deal upon the portion of the respiratory apparatus that is affected; if the aspergillar nodules were localized in the mouth, as it is sometimes in pigeons, or in the bones or air sacs, the duration of the disease would, of course, be much longer than if in the bronchi or lung substance.

The pathological lesions are as follows:

The actinomycotic masses are noteworthy. The fungus may frequently become localized in kidneys, and muscles of heart.

The microscopic examination of these organs disclosed a picture simulating the gross appearance of an advanced case of pulmonary tuberculosis, with the exception that the bronchial tubes were almost completely plugged with a greenish velvety membranous lining.

In the bronchial divisions not wholly occluded by the croupous exudate are seen the characteristic aspergillar fruitheads in various stages of development, from that of a slight bulging end of the hypha to those giving off their spores. Included within this alveolar exudate are quite a few leucocytes and red blood cells, but their presence is by no means constant. The bronchial mucosa is often eroded and the lining epithelium replaced by a fibrinous coagula or by a membranous material composed of matted mycelial threads from which hyphae extend into the air space, forming spore-bearing fruitheads, owing to the presence of oxygen.

In animals in which the disease was experimentally induced by the injection of the spores into the blood vessels or into the lung substance, miliary lesions resembling tubercular formations were quite noticeable in the lung tissues, and in these an occasional giant cell was discovered. In the lungs of a chicken which was inoculated directly into the lung substance, an acute miliary pseudo-tuberculosis was produced, accompanied by intense hemorrhages into the interstitial tissues, as was also the case in intravenous inoculations. In these tubercular nodules penetrating filaments could be made out, but the spores could

not be surely demonstrated, or at least differentiated from other cellular elements. Often the bronchial ramifications were the seat of hemorrhage, in which a noteworthy increase in the number of leucocytes could be observed.

In large rabbits the pathological lesions appeared to be as follows:

Rabbit No. 1008 failed to show any marked symptoms for the first two weeks after inoculation. It then began to lose weight, and on the twenty-fifth day was chloroformed. The postmortem examination showed an involvement of the liver, spleen, kidneys, and abdominal serous membranes, as in the preceding rabbit, but to a less extent. The organs of the thoracic cavity were apparently normal.

The optimum temperature of growth for the fungus is from 35°-40° C. Ceni and Besta in their investigations isolated a toxin from two species of Aspergillus, the A. flavus and A. fumigatus. Dogs inoculated intra-abdominally with large doses died within a few hours, showing tetanic symptoms and general hyperemia of all the organs. This work has not, however, been confirmed. Drs. Mohler and Buckley did not succeed in producing serious symptoms with the filtered product when injected into rabbits.

The Aspergilli also produce disease of the eye but, according to Plaut, this disease is not of frequent occurrence; he discusses several cases under the head of keratomycosis. One case described by Leber is as follows: A farmer forty-five years of age, while threshing had the misfortune to have some chaff of oats thrown into his eye. The sclerotic coat became inflamed, followed by healing and total leucoma (leucom). Another case is cited where a pear was thrown against the eye of a farmer, and another case of a fifty-three year old patient, a miller by profession, who had a slight fever, his right eye becoming inflamed. The conjunctiva had the appearance of trachoma. The sclerotic coat was clouded and the surface of the eye brittle, consisting of threads of fungi. Fuchs, who investigated this case, determined that the fungus was Aspergillus. Aspergillus fumigatus has also been observed in the nasal cavities where it produces necrosis and a disagreeable odor.

In a review of a paper by E. Bodin and L. Gautier * the following statements are made with reference to the Toxin found in Aspergillus fumigatus.

From a study of this fungus in cultures and in experimental animals it was found that Aspergillus fumigatus produces a toxin which may be rightfully compared with the toxins of bacteria. For the formation of this toxin in cultures it is necessary to bave a mixture of protein, especially of the peptone type, and some carbohydrate, especially glucose saccharose, maltose, or dextrin. The reaction of the toxin must be either neutral or alkaline. The effects of the toxin are chicfly observed in the nervous system and arc produced more or less rapidly by the method of inoculation. The symptoms of poisoning from the toxin are muscular convulsions resembling tetanus and leading to death within a few hours if the animal does not recover. The rabbit and dog are very susceptible to the toxin, while the guinea pig, cat, mouse, and white rat are more refractory. The dog and cat are naturally immune to the spores of A. fumigatus, but are quite susceptible to the toxin produced by the fungus.

Treatment: To prevent the disease, do not feed mouldy grain or fodder. Separate at once all diseased animals from the healthy. Use only thoroughly clean dishes; the troughs and boxes should be cleaned with formaldehyde.

Very little can be done in the way of treatment in the case of birds. Mohler and Buckley say that if a large number of birds are affected at one time, or if those affected are very valuable, treatment may be tried in the form of medicated vapors, such as those generated from wood tar or sulphur. A small quantity of wood tar is put in a pint of water and stirred with a redhot iron. The person doing the fumigating should remain in the room and immediately remove any

^{*} The Ann. Inst. Pasteur, 20 (106) No. 3, of the Experiment Station Record.

birds that are overcome by the vapors. Burning sulphur or vapors of formalin may be tried in like manner. Hydrogen peroxid, solutions of potassium iodid, or hyposulphite of soda may be used as intratracheal injections, and in case of local nodules in the mouth or nostrils the tincture of iodine may be applied to them with beneficial results.

It appears from the investigations with reference to kerato-mycosis, that infection generally occurs through the medium of feed, straw, or something that is thrown forcibly into the eye. According to Plaut the simplest and surest method of dealing with the disease is to use a 2% solution of salicylic acid, three times daily, but inhalation of an atmosphere containing iodine is recommended by some of the German investigators, or the inhalation of etheral oils. Immunity cannot be obtained by beginning with the injection of small quantities of spores and increasing the dose. Dogs are not immune against aspergilli. Mice are immune.

Aspergillus niger. Van Tieghem

An abundant mycelium in the substratum and on the surface becoming blackish; conidiophores long; sterigmata branched; conidia 3½-4½ µ in diameter, roughened; spherical or cylindrical sclerotia. The fungus contains diastase, invertase, and emulsin; it breaks up tannin into gallic acid and glucose, and converts sugar into oxalic acid.

Pathogenic properties. This fungus has been found both in the lungs and the ear, although less pathogenic than the preceding species.

Aspergillus subfuscus. Olsen-Gade

Mycelium olive yellow or brownish when mature, in and on the substratum; conidiophores short, club-shaped; spores spherical, colorless.

Distribution. Found in Europe; closely resembles A. fumigatus.

Pathogenic properties. Pathogenic, but less so than the A. fumigatus or A. niger.

Aspergillus nidulans. (Eidam.)

The mycelium forms greenish masses; later the mass assumes a reddish color; conidiophores 0.6—8 millimeters long and 8-10 μ across, colorless, branched; sterigmata consist of a basal branching cell and two or more branches, each branch containing from 20 to 30 conidia; perithecia yellowish, 0.2—3 millimeters in diameter; ascospores 8.

Distribution. Found in Europe.

Pathogenic properties. The disease appears on the second day after inoculation in guinea pigs and death occurs in 60 hours. Kidneys are enlarged and show small white dots. White masses also occur in the peritoneum. It is pathogenic for cattle and man, and is occasionally found in the human ear.

PYRENOMYCETINEAE

An important division of the fungi, containing about 10,000 species, many of which are troublesome parasites on cultivated plants. The mycelium is composed of delicate distinct hyphae or of closely coherent threads, frequently forming a pseudo-parenchymatous tissue; hymenium enclosed in a subglobose envelope called a perithecium, or with an opening at the apex, which is often

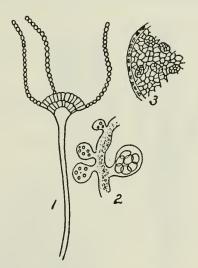


Fig. 89a. Aspergillus nidulans. 1. Conidiophore. 2. Branch of mycellum with asci and ascospores, magnified. 2. Asci. 3. Cross section. Ascus. All greatly magnified. (After Eidam.).

prolonged to form a short tube or beak; numerous transparent asci arise from the base of the perithecium, these contain the ascospores; between the asci slender filiform bodies, called the paraphyses. Polymorphic fungi with conidia, spermogonia, and pycnidia, supposed to be connected with the ascigerous stage. The formation of the ascospores is in some cases presented by the development of sexual organs in which genuine fertilization occurs. The reproduction can be illustrated by the manner in which it occurs in the powdery mildew of the lilac, Microsphaera Alni.

The mycelium spreads over the surface of the lilac leaf; the fungus draws its nourishment from its host by means of haustoria which penetrate the epidermal cells; the mycelium produces erect branches which bear these spores in a moniliform chain, the end spore being the oldest; these summer spores germinate immediately and propagate the fungus; later two hyphae cross and there arises an oval cell, the oogonium, which is separated from the hypha by a cell-wall at the base; from the same hypha springs a longer and thinner cell, also cut off by a cell-wall; this cell is above the oogonium, and is known as the antheridium; from the base of the oogonium other cells arise which soon enclose it; finally a brown perithecium is formed which bears dichotomously branched appendages; the perithecium contains the asci, in which are found the ascospores, which germinate, probably, in the spring. The accompanying figure after Harper illustrates the development.

Another type of one of the Sphaeriaceae, the Gibbellina cerealis, is common on stems of wheat where it produces at first a grayish brown circular spot, the mycelium frequently encircling the stem. The conidia are oval, the perithecia are immersed.

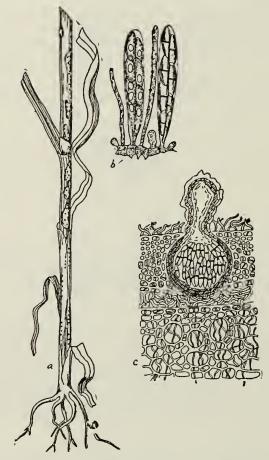


Fig 90. Stem Blight (Gibbellina cerealis), one of the Sphaeriaceae. a, general appearance. b, asci with ascospores and paraphyses. c, stroma, mycelium, and perithecium. After Cavara.

PERISPORIALES

Perithecia spherical, closed, or with the ostiolum obscure, coriaceous or brittle carbonaceous, opening irregularly, generally without stroma, but mostly seated on a well developed, superficial mycelium. This division includes the order *Erysibaceae*.

ERYSIBACEAE.

Superficial mycelium, branching, septate, closely adhering to the surface by means of the haustoria; asci arising from the base of the perithecium, delicate, thin-walled, colorless, oblong, obovate or suborbicular, stalked, usually containing from 2-8 ascospores; perithecium spherical with appendages, without ostiolum; conidia (Oidium) simple, colorless, cylindrical, oval or ovate, borne one above the other on septate, colorless hyphae. Contains many important

parasitic fungi, like the powdery mildew of the grape (Uncinula spiralis), mildew of lilac (Microsphaera Alni), mildew of sunflower (Erysiphe Cichoracearum), mildew of cherry (Podosphaera tridactyla).

Erysiphe. (Hedw.)

Perithecium containing several asci, appendages with simple threads, similar to and frequently interwoven with the mycelium. A small genus of 20 species of wide distribution.

Erysiphe communis. (Wallr.)

Amphigenous, mycelium abundant, persistent, or sometimes evanescent; perithecia variable in size and reticulate, appendages variable in length, often long; asci 4-8 or more, ascospores 4-8.

Distribution. Found on a large variety of different hosts but common on plants of the order of Leguminosae, especially the forage plants like the pea (Vicia sativa), bean, clover and other members of the clovers.

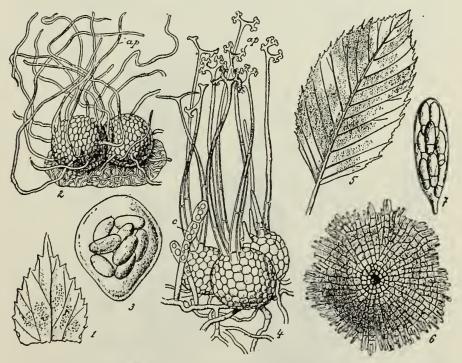


Fig. 91. Powdery Mildews. 1-3. Sphaerotheca Castagnei on Hop. 1. Part of leaf of hop with perithecia shown in the form of dots. 2. Perithecia with tortuous appendages (ap) x 175. 3. Ascus with spores within the ascospores x 380. 4. Powdery Mildew on Cherry (Podosphaera tridactyla), conidiophore bearing conidia (c). 5-7. Microthyrium microscopicum. 5. On leaf. 6. Perithecium, greatly magnified. 7. Ascus and ascospores. 1 after Wettstein. 2-4 after Tulasne. 5 after Lindau. 6-7 after Winter.

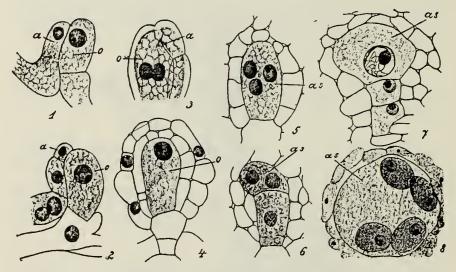


Fig. 92. Powdery Mildew. Sphaerotheca Castagnei. 1. Oogonium (0) and antheridium (a). 2. Separation of antheridium cell. 3. Fertilization and formation of additional cells. 5-8. Further development of cells. All greatly magnified. After Harper.

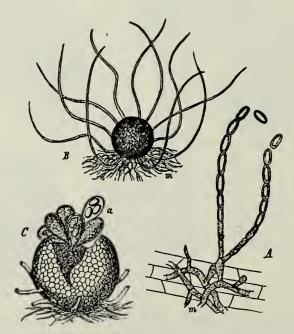


Fig. 93. Powdery Mildew of Grass (Erysiphe graminis).
A. Oidium stage and mycelium m. B. Perithecium with appendages and mycelium m. C. Perithecium with asci and ascospores. After Frank.

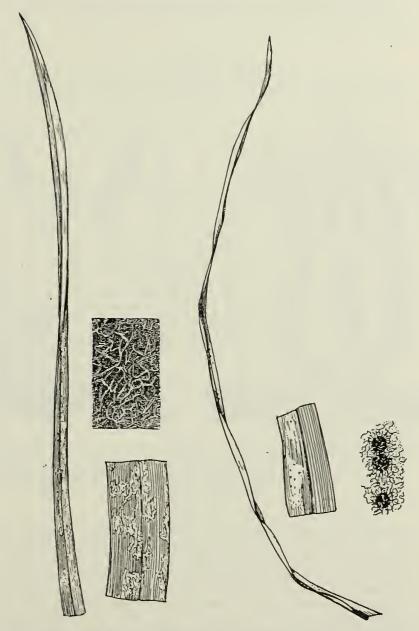


Fig. 94. Powdery Mildew of Bluegrass (*Erysiphe graminis*). Oldium stage; leaves at the right magnified, the one above more highly, showing the powdery substance. (Charlotte M. King).

Poisonous properties. The Veterinarians of Europe ascribe to these mildews a form of stomatitis.

Erysiphe graminis D C.

Amphigenous, often epiphyllous, mycelium dense, felt-like, persistent, white or gray, sometimes tinted hrown; perithecium immersed in the mycelium, few and scattered, large, about 225 μ , in diameter; asci 16-25 μ oblong or oval, stalked, ascospores 8 or rarely 4; appendages rather short.

Distribution and Hosts. Found on many different grasses like blue grass (Poa pratensis), fowl meadow grass (Poa serotina), occasionally also on wheat or orchard grass. The following rather popular account treats of this disease as it is common in the west.

Every one who has had occasion to walk through a blue grass meadow after a rain, especially in damp and shaded places close to the ground, must have noticed a white mealy covering on the blades of many of the leaves. The Germans have called this mehlthau (literally translated meal dew), which is certainly very expressive of its appearance. An examination with a microscope will show that this white substance is composed of spores and a mycelium. The mycelium is cobwebby and spreads over the surface, but does not penetrate the leaf. In numerous places erect branches are produced, these bear numerous spores. This stage was formerly called *Oidium monilioides*, being named Oidium because the spores resemble an egg, although the resemblance is not marked in all cases of Oidium; the species was called *monilioides* because it was necklace like, referring to the manner in which the spores are borne. Worthington G. Smith states that the spores are so small that it would take about a million to cover a square inch.

In a powdery mildew occurring on the squirrel-tail grass, and supposed to be the same fungus, these spores are also capable of immediate germination. On blue grass the fungus frequently does not produce perithecia but ends its existence with the formation of conidia. It produces perithecia abundantly on wheat in Iowa.

These conidia or summer spores germinate, under favorable conditions, in from ten to sixteen hours. The temperature most favorable for germination is from 17-26° C. In a powdery mildew occurring on the squirrel-tail grass, and supposed to be the same fungus, these spores are also capable of immediate germination.

Under favorable conditions, especially moisture and damp weather, the fungus spreads rapidly. The leaf of grass affected by this fungus soon dries, and when the affected plants are disturbed, small clouds of dust arise, especially in shady places. The perfect stage of the fungus is not of common occurrence, though if careful search is made in the fall, small black specks may be seen; these are the perithecia and contain the asci and ascospores. It is the resting stage or winter condition of the fungus. The writer found the perfect fungus abundant on Poa Wolfii in Colorado, and Carver found it abundant on blue grass near Ames one season. The spores of the Oidium stage do not retain their power of germination very long, but the ascospores contained in the perithecium germinate the following spring, and when the tube comes in contact with the proper host the mycelium spreads over the surface of the leaf and causes the mealy appearance.

Poisonous properties. This species is abundant and often causes serious trouble; it certainly renders the hay nearly worthless to be fed to animals. It often, no doubt, gives rise to a stomatitis such as is described for other fungi.

HYPOCREALES

Perithecia spherical or ellipsoidal, with an ostiolum; stroma when present variously colored, reddish, yellow, never black or hard.

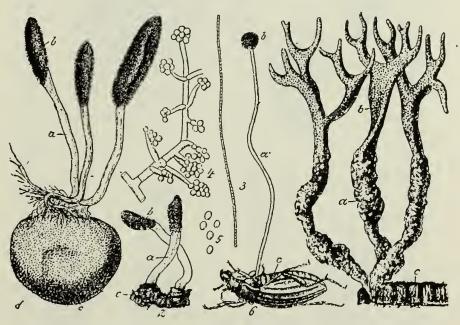


Fig. 95. Various species of Cordyceps. 1. C. ophioglossoides. 2. C. militoris, a Stroma on a caterpillar (c). 3. Stroma on a fruiting form of Elophomyces gronulotus. 3. Ascospore x 200. 4. Conidiophore x 350. 5. Conidia of C. ophioglossoides. 6. C. cinereo on a beetle (c). 7. C. Toylori on a caterpillar (c). o in all figures sterile, b fertile part of the Stroma. 1 and 6 after Lindau. 3-5 after Brefeld.

HYPOCREACEAE

Simple or compound; perithecia somewhat coriaccous, never black; bright colored, opening by a subcentral ostiolum, stroma soft, waxy, or occasionally cottony. A very numerous family containing many species. Contains the genera Nectria, of 250 species, some being parasitic upon trees; the Gibberella and the Hypocrea upon barks of trees, etc., Cordyceps, parasitic upon various insects, C. militaris being found upon Lepidotera, the conidial stage of which is Isaria farinosa, the C. Ravenelii upon the larvae of the June beetle; Polystigma rubrum, parasitic upon the plum; Epichloe typhina, the so-called Cat-tail fungus found upon various species of grass, especially timothy and orchard grass. Contains also the Gibberella Saubinetii, a parasite on wheat, which is a stage of Fusarium roseum described later in this work.

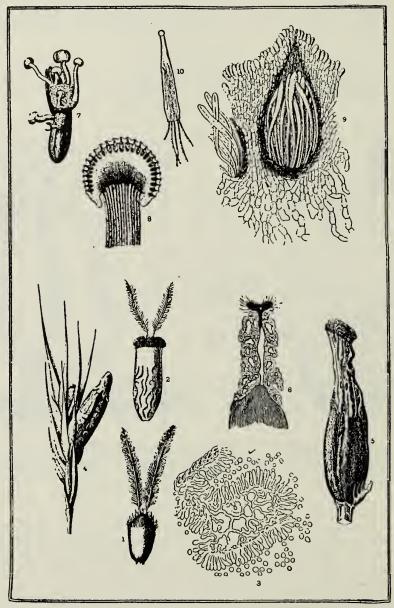


Fig. 96.—Normal ovary of rye. Fig. 2—Same invaded by Claviceps. Fig. 3—Cross-section of ovary showing mycelium and spores of sphacelial stage. The round bodies are summer spores. Fig. 4—Sclerotium stage, Fig. 5—Sclerotium stage, Fig. 6—General view in sphacelial stage, Fig. 7—Development of ergot in spring, Fig. 8—Cross-section of globular head showing flask shaped perithecia, Fig. 9—Asci. A single perithecium showing elongated bodies in the center. Fig. 10—A single ascus with filiform ascospores protruding. These spores (reproductive bodies) germinate and infect the young ovary of rye. After Tulasne. U. S. Dept. Agrl.

Claviceps, Tul. Ergot.

Stroma erect, consisting of a sterile stem; subglobose, fertile head from a subcylindrical, black, hard sclerotium; perithecium immersed in the stroma, flask shaped; asci, clavate-cylindrical; ascospores, filiform, colorless.

Claviceps purpurea, (Fr.) Tul.

Sclerotium variable in length from $\frac{1}{2}$ 8 to 1 inch long or more; long cylindrical; generally somewhat curved, wrinkled, purplish on the outside, white within; usually several fruiting bodies from the same sclerotium; heads spherical, tuberculose, borne on short flexuose stems; asci narrow, linear, 8-spored, ascospores filiform, continuous, attenuated toward the end, 50-76 μ long.

Ergot is a stage of a minute parasitic fungus; although its true nature was not known by early writers, it is mentioned by many of them. Lonicer, about the middle of the sixteenth century, mentions its specific use. Thalius applied the name of "ad sistendum sanguineum."

Bauhin used the name of Secale luxurians. De Candolle called it Sclerotium clavus. Although other names have been applied to it, the credit of working out the life history belongs to Tulasne, one of the most eminent of French mycologists.

There are still many persons who believe that ergot is a degenerate kernel of rye or wheat, but the researches of Tulasne and other mycologists have laid at rest many of the vague theories concerning it. The black, purple, or dark gray spurs found in the flowers of rye, wheat, and other grasses are simply one stage of a parasitic fungus, known as Claviceps purpurea. These spurs consist of a compact mass of threads known as the sclerotium stage; it was formerly called Sclerotium clavus.

No changes occur in ergot while it remains in the head, but the following spring, when laid on damp earth, it produces at different points small, roundish patches which are somewhat elevated. Soon a small white head appears which elongates, becoming stalked, and bearing a globular head at the tip. These heads change from a grayish yellow to a pinkish color. A cross section shows that the central portion is made up of closely woven hyphae or fungus threads, while the edge contains a number of flask-shaped bodies, the perithecia, in which are found elongated bodies known as asci; each ascus contains eight filiform spores, the ascospores. The ascospores germinate and when coming in contact with a very young ovary the mycelium penetrates the delicate walls of the ovary and gradually displaces it. It is quite easy to trace out its life history by placing the ergot in damp sand and allowing it to remain over winter.

The first indication of ergot in the summer is the formation of the socalled honey-dew, a sweetish and rather disagreeable fluid, which is eagerly sought by flies and other insects which feed upon it. This fluid contains a large number of small spores so that insects can readily carry the fungus from a diseased ovary to one not diseased. These spores germinate immediately. This stage is called the sphacelia, and formerly was held to be a distinct fungus. In this stage the mass which has replaced the ovary is soft, but as it becomes older it hardens; ultimately a hard and compact mass, the ergot, is formed. Distribution and hosts. Found on a large number of host plants. Rye is more subject to it than any of the other cultivated cereals. The largest specimens are usually produced on isolated specimens of rye coming up in fields. It seldon happens that all of the ovaries are affected. Wheat, especially winter wheat, is subject to the disease. The officinal ergot is usually obtained from rye. In Europe it has been reported on oats. Mr. C. W. Warburton found it on the same host in Iowa, in 1909.

Of our native wild grasses, wild ryes (Elymus robustus, E. virginicus, E. striatus, E. canadensis, Asprella hystrix) are most subject to the disease. Most cases of ergotism in the United States undoubtedly result from the ergot on various species of Elymus; in Iowa on the Elymus robustus, which is a common plant everywhere. Agropryon occidentale, a grass not uncommon in northwestern Iowa, and Quack Grass (Agropyron repens), are also much subject to its attacks. Scarcely a head of the Western Wheat Grass cultivated on the college farm could be found which did not have some ergot. This may be for the same reason that it occurs most abundantly on rye, namely, that the grasses occurred in isolated places. In some pastures, timothy (Phleum pratense), is much subject to the attack of Claviceps purpurea. Thus in an old pasture in Wisconsin I observed a large percentage of timothy which contained many heads which were ergotized. Blue grass (Poa pratensis), Poa annua, Calamagrostis canadensis, Agrostis alba, Glyceria fluitans, and many others, in some seasons and localities, are diseased. Unusually large spemimens sometimes occur on Wild Rice (Zizania) in Iowa.

It may be possible that some of the forms of ergot on grass may be referred to other species. Halsted states, however, that ergot on Elymus robustus is Claviceps purpurea. The Hordeum jubatum contained apparently the same species, with some minor differences but these were due to the nature of the host. Claviceps microcephala (Wallr.) Tul., occurs on Phragmites, C. setulosa (Quel.) Sacc. with yellow stroma on Poa, and C. pusilla Ces on Andropogon Ischaemum.

Poisonous properties. The subject of ergot and ergotism is one of considerable importance to stockmen in many parts of the country. Scarcely a year passes without some complaints being received by the state veterinarians of the injurious effects of ergot. The writer receives several complaints of this kind every year. But the cases of ergotism today are not nearly so frequent as they were 40 or 50 years ago. We will, therefore, append here a short history of the disease.

Epidemics of ergotism have, without doubt, been correctly referred, before the tenth century. Wood states that epidemics of ergotism or chronic ergot poisoning have been recorded from time to time since the days of Galen (130-200 A. D.) and of Caesar (B. C. 190-44). From the ninth to the thirteenth century epidemics were frequent in France, and in the twelfth in Spain. They were first called plagues but later received special names. In 1596 Hesse and adjoining provinces were visited by this plague which was attributed to the presence of ergot in grain. In the epidemic in Silesia in 1722, the king of Prussia ordered an exchange of sound rye for the affected grain. Freiburg was visited in 1702, Switzerland in 1715-16, Saxony in 1716, and other districts of Germany in 1717, 1736, 1741-42. France was visited in 1650, 1670, and 1674. From 1765 to 1769 it was abundant in Sweden in rye

and barley. Linnaeus attributed it to the grain of Raphanus raphanistrum, which occurred in France in 1816, in Lorraine and Burgundy; it was especially fatal to the poorer inhabitants.

It has been observed that these epidemics follow a rainy season. Fleming states that in 1041, when the weather was so unpropitious, tempests, rains, and inundations occurring, many cattle perished from the disease. "In 1098, after inundations and heavy fogs, there was a general epizootic among cattle in Germany. In the same year ergotism appeared in the human species."

Dr. Randall, in 1849, called attention to a discase in New York, in which the involved parts were finally invariably affected with dry gangrene. He states that in the severe climate of New York farmers allow their cattle to winter in fields on blue grass (Poa pratensis) which is rich in ergot. A disease known as "hoof-ail" was correctly ascribed to ergot by James Mease, of Philadelphia, prior to 1838. The disease was quite severe in Orange county, New York, in 1820. It was minutely described by Arnell. In 1857, the disease was quite severe in Portage county, Ohio. A committee appointed by the Farmers' Association of Edinburg reported that the disease was due to ergot contained in the hay eaten by cattle. In recent years, epizootics of ergotism have been reported by Law in New York, Stalker in Iowa, and Faville in Colorado. In 1884, a very serious outbreak occurred in Kansas which was at first diagnosed as "foot-and-mouth disease." Dr. Salmon found, upon examining samples of hay from various localities in the state, that these contained considerable quantities of wild rye (Elymus virginicus, var. submuticus) which in turn contained a large amount of ergot, in one case, 12 per cent and in another 10 per cent being found. From this he estimated that 5-6 per cent of the entire weight of the plant must have been ergot and that a twenty-pound ration of hay would contain four ounces of ergot.

Dr. Harshberger has called attention to an outbreak of ergotism from the use of ergotized red top, the fungus being common on red top throughout the United States and being one of the most common impurities in red top seed.

The ergot contains the substance leucin and the non-nitrogenous substance ergotine, which according to the earlier investigations was regarded as the active principle and as an alkaloid. According to Wenzell ergot contains the two alkaloids, ecbolin and ergotin C50H52N2O3 an amorphous, alkaline, feebly bitter substance. But according to the later investigations these substances are Tanret isolated the crystallizable alkaloid ergotinin C35H40N4O6; this is a crystalline, slightly bitter substance, subsequently Kobert found that this substance would not produce the action accredited to it and attributed its action to ergotinic acid and the alkaloid cornutin. The more recent investigation of Jacobi attributes the poisonous action to chrysotoxin, an amorphous glucosidal acid. Secalinotoxin is a compound of sphacelotoxin, and secalin C29H55N6O4; accompanied by the harmless substance, ergochrysin. According to Kobert cornutin is an alkaloid having a specific action on the uterus, causing it to contract; sphacelic acid, a non-crystallizable and non-nitrogenous substance which causes the poisoning and gangrene; ergotinic acid, a nitrogenous glucoside without action on the uterus and narcotic in its effects. Besides these substances it contains others, prominent among them being a sugar called mycose, which is also present in other fungi. Ergot stimulates the involuntary muscles of the stomach and the intestines, it causes a constriction of the arterioles and veins throughout the body with an increase of blood pressure. In toxic doses it paralyzes both the vasomotor centers and the heart muscle.

It appears from the experiments of Dale 1 and Barger and Carr 2 that cornutin does not occur as such in ergot but is an artificial decomposition. Tanret discovered the first well defined crystalline alkaloid which he called ergotinin. The secalin of Jacobi 3 is identical with ergotinin. Barger and Carr separate a second alkaloid which can be recognized chemically; to this they gave the name of ergotoxin, $C_{35}H_{41}O_6N_5$. This substance is of great physiological potency. According to Dale 4 ergotoxin produces in doses of a few milligrams "not only the characteristic reaction of ergot described by him, but also gangrene of the Cock's-comb, and other ergot effects described by Kobert and others to sphacelic acid."

According to Cronyn and Henderson 5 ergotoxin is a highly active alkaloid and has the properties of ergot most desired in medicine. It brings on long enduring vaso-constriction, increases uterine movements when injected in-



Fig. 98.—Effects of Ergotism: Hoofs of cattle showing flesh sloughing away. (Salmon.)

travenously and the same to a less extent when injected subcutaneously, but when given per os has very little action.

The toxicology of ergot is well described by Dr. Winslow as follows:

Enormous single doses are required to poison animals or man. When as much as two drachms of ergot to the pound, live weight, are given to dogs, death is not constant. Three ounces, however, have proved fatal to small dogs. Acute poisoning is characterized by vomiting (in dogs), profuse salivation, dilation of the pupils, rapid breathing and frequent pulse. The animal cries out, has convulsive twitchings, staggering gait, paraplegia, intense thirst, and coma, terminating in death. Horses, cattle, and sheep are unaffected by any ordinary quantity of the drug.

Chronic poisoning or ergotism rarely occurs in animals owing to continuous ingestion of ergotized grains. It is characterized by gastro-intestinal indigestion, with nausea, vomiting, colic, diarrhoea or constipation, and abortion ensues in pregnant animals. In addition to gastro-intestinal irritation the symptoms naturally assume two forms: 1. The gangrenous form; 2. the spasmodic form. In the first variety of ergotism there are coldness and anesthesia of the extremities, including the feet, ears, and tail of quadrupeds; the comb, tongue, and beak of birds, — followed by the appearance of passive congestion, blebs, and dry gangrene in the vicinity of these parts. The hoofs and beaks often drop off.

¹ Jour. Phys. 34:163, 1906.

² The alkaloids of Ergot. Jour. Chem. Soc. 91:337, 1907. These writers give a full literature on the subject.

³ Arch. Expt. Path. Par 39:104.

⁴Jour. Phys. 34:163.

⁵ Jour. Pharma, and Expt. Therapeutics. Aug. 1909.

Death ensues from general exhaustion. In the spasmodic form are seen tonic contraction of the flexor tendons of the limbs and anaesthesia of the extremities; muscular trembling and general tetanic spasm, with opisthotonos, convulsions and delirium. Death also occurs from asthenia.

Grünfeld fed various animals with sphacelic acid in food. In the cocks, gangrene soon appeared affecting the comb; next the wattles, tongue linings, and crop. In hogs, the ears became gangrenous and fell off. Horses and cows fed upon grains containing ergot lose their hoofs, ears, and tails. The cornutin, according to Kobert, acts through the nerve centers. Microscopic examination of the abdominal and thoracic regions shows a toxic polyneuritis.

Dr. McNeil in describing the disease says:

Ergot stimulates the nerve centers that cause the contraction of the small blood vessels supplying the different parts of the body and cause one of the two forms of ergotism, namely, a nervous form, and a gangrenous form.

Nervous Ergotism: In this form the contraction of the blood vessels of the brain produces dullness and depression. The animal also suffers from gastro-intestinal catarrh, refuses food, and gradually passes into a condition of general wasting. The nervous form, however, may assume an entirely different aspect and the animal dies suddenly in delirium or spasms, or gradually from paralysis.

Gangrenous Ergotism: In this common form the checking of the blood, resulting from the contraction of the small blood vessels, causes a loss of a part or of all the limb below the knee or hock, the tail, or the ears. This form of the disease may manifest itself by the formation of ulcers at the top of the hoof or between the toes, and a toe may be lost or the entire hoof shed. The affected part dries, a small furrow or line of separation appears, completely surrounding the limb, dividing the living from the dead mummified tissue.

DOTHIDEALES

Perithecia reduced, asci arising from the stroma and not separable from it, stroma present, not fleshy; black or dark colored ostiolum present.

DOTHIDEACEAE

Stroma pulvinate, elongated, black or nearly black, coriaceous; perithecia inseparable from the stroma, asci 4-8 spored; hyaline, yellowish or brown.

Phyllachora, Nitschke.

Stroma variable, elliptical, oblong or lanceolate, covered by the epidermis, black, roughened, ascospores ovate, elliptical, or oblong, mostly hyaline. About 200 species.

Phyllachora Trifolii, (Pers.) Fckl.

Stroma on the lower surface of the leaf, gregarious, collected in small, elongated groups extending along the nerves of the leaf, black, subglobose, prominent, often confluent; ascospores elliptical, hyaline, continuous, $10-20~\mu$ In the early part of the season small whitish or pale brown spots appear on the leaf, which contains the mycelium of the fungus. Dr. Trelease says:

This fruits on the lower surface, producing numerous tufts of necklace-shaped threads, each of which ends in a 2-celled, egg-shaped conidia-spore. These tufts of threads, which, like the spores, are of a deep brown color, are packed so closely together as to completely cover the spots, though under a hand lens they can be distinguished as separate panules. To the naked eye they appear dead-black. Later in the season similar spots are occupied by small, coal-black fruits that contain stylospores. Winter spores, produced in asci, are not known. The conidial form of this fungus is especially common on white clover, though both forms are at times found abundantly on red clover and other species.

The *Polythrincium* is common on red clover and is one of the numerous species which may be injurious to cattle.

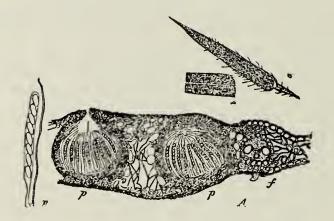


Fig. 99. Black Spot of Grasses (Phyllachora graminis). A. Cross section of leaf through a black mass of the fungus. P. Perithecia. B. An ascus with ascospores. a, b. Spots on grass leaf caused by the fungus. After Frank and Trelease.

Phyllachora graminis (Pers.) Fckl.

Stroma scattered or confluent, penetrating the leaf and more or less prominent on both sides, covered by a black and shining epidermis, roughened; ostiola obscure; asci short, stalked, cylindrical, 75-80 x 7-8 μ , ascospores 8, paraphyses present.

Phyllachora graminis, occurs on many cultivated and wild grasses; other species occur on clover and other leguminous plants. This parasitic fungus disease causes blackish spots on the lower or both surfaces of the leaf. The fungus causing these black spots on grasses has been called the black spot disease.

During August, and especially later, the coal black spots along the veins are especially prominent; they are considerably less than one-eighth of an inch in length and width and occur on both surfaces of the leaf, but are more abundant on the upper. These black spots are composed of dense mycelium, which in the green leaves bears numerous small spores which serve to progapate the fungus in the summer. In dead leaves, small perithecia are found, which contain numerous elongated bodies, the asci, within which are found eight small, colorless spores, known as ascospores; these latter carry the fungus over winter.

Distribution and Hosts. Widely distributed in both Europe and North America, very common upon Quack Grass, Wild Rye, Bottle Grass, Panic Grass, etc.

Poisonous properties. The genus Phyllachora is abundant at times and is associated with stomatitis.

SPHAERIALES

Perithecia generally with a distinct ostiolum, of various consistency, not reddish or membranous, brown or blackish; stroma when present dark colored outside and whitish within. Contains the families: Sordariaceae, found upon decaying plants and substances; Chaetomiaceae, with superficial perithecia, gen-

erally with short ostiolum and an apical tuft of hairs or bristles; one species Chaetomium chartarum common on paper. Sphaeriaceae, with membranaceous perithecia, apex perforated with a simple pore, contains a large number of parasitic fungi like the strawberry rust or spot disease (Sphaerella Fragariae), and the spot disease of the currant (Cercospora angulata).

FUNGI IMPERFECTI

The fungi included in this group are simply form genera, many of the species belong to the *Pyrenomycetes*, some belong to the *Phychomycetes*, and some to *Hymenomycetes*. In this connection we shall describe a few only which may cause trouble in forage.

Helminthosporium gramineum, Rabh. Yellow Leaf Disease of Barley

Spots in parallel rows, causing the leaves to become marked with yellow lines of pale green color; mycelium of the tissue colorless; conidiophores brownish on the surface, spores large 3-6-celled.

Distribution. Widely distributed in Europe and North America on barley.

Helminthosporium turcicum, Pass. Leaf Browning of corn

Spots sharply limited, conidiophores brownish elongated, bearing several brown spores. Widely distributed in Europe on corn, and also in North America.

Helminthosporium inconspicuum, E. & E.

Leaves dead and discolored, discoloration sometimes interrupted by spots of various sizes; conidiophores brown with several-celled conidia.

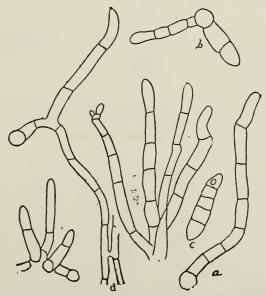


Fig. 100. Yellow Leaf Disease of Barley (Helminthosporium gramineum). a. Hypha arising from short cells. C. Conidium and to the left a cluster of conidiophores. d. Mycelium.

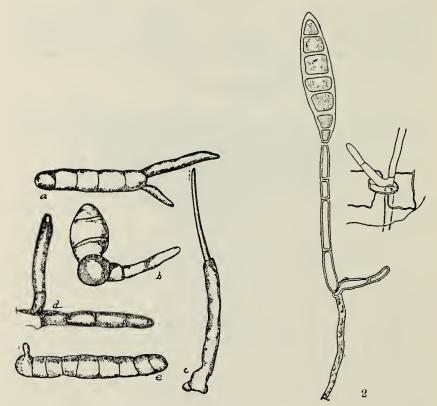


Fig. 101. Spores of Yellow Leaf Disease of Barley (Helminthosporium gramineum). a. and e. Spores germinating. d. Conidiophore. 2. Leaf browning of Corn (Helminthosporum turcicum). Spore and conidiophore to the left. To the right, conidiophore pushing through stoma.

Distribution and hosts. On corn, widely distributed in North America. *Poisonous properties*. All of these fungi may be regarded as injurious, possibly producing stomatitis.

Scolecotrichum graminis, Fuckel

Elongated brown or purplish-brown spots, the centers of which are gray or whitish and contain minute black dots; these small dark spots contain the tufts of brown fungus threads, which make their way out through the stomata; the hyphae are somtimes septate and the spores are usually borne at the end or occasionally in a lateral position; these fruiting hyphae bear small, smokybrown, two-celled spores; the cells of the leaf become much altered, because the colorless threads of the fungus permeate them. On barley the disease is marked by brown or purplish-brown spots which appear on the leaf transversely.

Distribution. Widely distributed in Europe and North America. Poisonous properties. May possibly produce mycotic stomatitis.

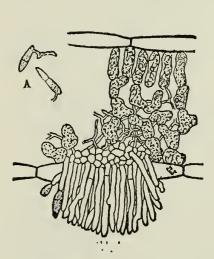


Fig. 101a. Spot Disease of Orchard Grass (Scolectotrichum graminis). Cross-section of leaf, general fruiting layer of fungus with conidiophores and conidia. A, spores germinating. After Trelease.



General appearance of fungus on leaf of Orchard grass. After Trelease.

Polydesmus Mont. Rape Fungus

Sterile hyphae repent; fertile erect, simple or branched septate colorless conidia, interstitial filiform, concatenate, fusiform or clavate; many septate and opaque.

Polydesmus exitiosus Kühn. Rape Fungus

Forming minute, punctate, elongated dark brown spots, conidia elongated or somewhat clavate, narrowed upwardly, 18-12 septate; the septa but slightly constricted, olive-brown in color. The conidia are 120-140 by 14-16 μ ; conidiophores short, straight or slightly irregular, septate, making their way through the stomata.

This fungus is widely distributed on rape and cabbage and has been referred to as Alternaria brassicae. It is, however, thought to be a distinct fungus.

Poisonous properties. In Europe this fungus has long been associated with mycotic-stomatitis of cattle, but mycotic-stomatitis may be produced as indicated elsewhere, by other molds and fungi. This disease is characterized by inflammation and ulceration of the mucous membranes of the mouth. Salivation is a prominent symptom; the feet become swollen and sore. Dr. Mohler says:

"Superficial erosions of the skin, particularly of the muzzle, and of the teats and udders of cows, may also be present, with some elevation of temperature and emanciation."

The disease is not serious and in many cases recovery occurs. But where treatment is not resorted to the disease may prove fatal, death occurring in from 6-8 days. Dr. Mohler states that in serious outbreaks it is about 0.5%.

The treatment should consist of first removing the herd from the infected pasture or inclosure containing the fungus. They should be fed on good wholesome soft nutritious food, plenty of cold water should be given. Dr. Mohler recommends dissolving 2 heaping tablespoonfuls of borax or 1 tablespoonful of potassium chlorate in each of the first two buckets of water taken during the day. If the animals permit the mouth should be swabbed out with some antiseptic wash, such as weak carbolic acid or creolin solution, or permanganate of potash, or hydrogen peroxid. Mohler recommends that range cattle can be treated by the use of medicated salt.

"This salt may be prepared by pouring 4 ounces of crude carbolic acid upon 12 quarts of ordinary barrel salt, after which they are thoroughly mixed. The lesions of the feet should be treated with a 2 per cent solution of carbolic acid or of creolin, while the fissures and other lesions of the skin will be benefited by the application of carbolized vaseline or zinc ointment. If the animals are treated in this manner and carefully fed the disease will rapidly disappear."

Cladosporium herbarum (Pers.) Link

This fungus and its allies are very common upon oats, sometimes very destructive. It attacks all parts of the plant, but is especially common in the heads. The mycelium of the fungus grows not only on the surface of the plant but also in the interior; the conidiophores and spores are olive green, the former pass through the opening of the stomata or break through the epidermis; the spores are 1- to 2-celled, borne on the end or on short lateral branches and are extremely variable in shape and size.

The general effect of the disease is to cause the kernels to shrivel. The disease, as recorded by Cobb, occurs rather destructively on oats. Professor Peck records the occurrence of a Cladosporium on oats, which he describes as a new species, the *Fusicladium destruens*. He says in regard to oats:

"The foliage of the plants presented a singular admixture of green, deadbrown and reddish hues, strongly suggestive of that of a 'rust-struck' field."

Peck thinks this fungus inhabits the leaves of some of our northern grasses and has escaped from them to oat fields. Giltay reports that plants are infected in the same way as in some of the grain smuts, the spores being carried over with the seed, and that the disease can be prevented by treatment with hot water. A species of Cladosporium commonly affects the kernels of maize and is at times quite troublesome.

Septoria Fr.

Perithecia imbedded in the tissues of the plant, appearing as small blackish or brownish spots; conidia generally multicellular and colorless; produced from short conidiophores. A genus containing numerous species of wide distribution. Many of them like the Septoria on the black currant and gooseberry, and the blackberry leaf spot, Septoria rubi, are troublesome parasitic fungi of cultivated plants. All of these fungi irritate the mucous membranes when found in abundance in the leaf.

Septoria graminum, Dem.

Spots at first yellow, then reddish-brown and finally whitish; perithecia blackish or brownish-black; spores 50 to 60 μ long and 1.5 to 2 μ wide, numerous, usually 2-celled.

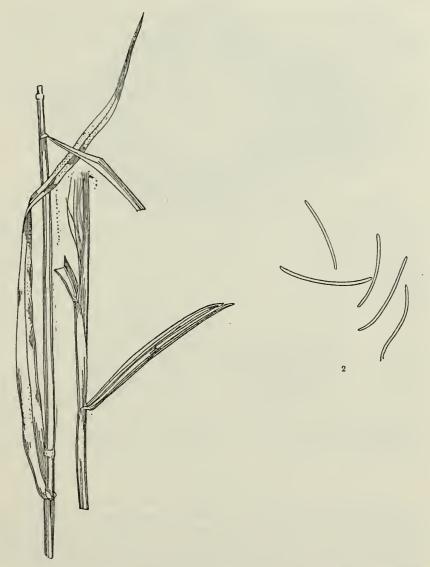


Fig. 102. 1. Leaf of Cheat, showing numerous small specks, the perithecia of Septoria Bromi, the spores in the perithecia shown at 2.

In a somewhat extended account of this disease Cobb states that the entire plant is not always involved. The fungus is variable, its character depending upon the host which it attacks. On Poa annua the leaf is mainly involved and in many cases is totally destroyed. Cavara states that the spots on the leaves are small, elliptical, red or yellow, or the latter may be entirely absent. The injury it does to young plants is very great; in some cases their total destruction has been observed.

Janczewski who has studied the life history of Septoria graminum states that this represents the pycnidial stage of Leptosphaeria tritici and that the conidial form is the Cladosporium herbarum. We have not found the Septoria in Iowa though the Cladosporium is common.

The Septoria tritici Desm. is closely related to the above and should perhaps be regarded as nothing more than a variable form of S. graminum. The spots it produces are at first yellow, then reddish-brown, and finally whitish. The spores are $50-60 \,\mu$ long and 1-5 to $2 \,\mu$ wide and usually divided. A Septoria on the glumes of wheat in Ohio has been reported by Selby.

Several other species of Septoria are allied to the above species, one, the Septoria bromi Sacc. is common in Iowa on Bromus secalinus.

Diplodia, Fr.

Perithecia bursting out sub-cutaneously, sub-carbonaceous, papillate according to type; spores ellipsoidal, ovoid or oblong, 1-celled, fuscous, perforated; basidia rod-like, simple hyaline. From the original genus have been separated five genera as follows: Species with superficial perithecia Diplodiella; with hirsute perithecia Chaetodiplodia; with clustered perithecia Botry-odiplodia; with mucilaginous spores Macrodiplodia; with hyaline spores Diplodina.

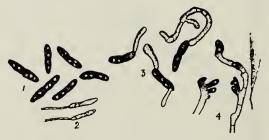


Fig. 103. Spores of *Diplodia Zeae*. 2. Young spores on the conidiophores with spore-phores attached. 3. Germinating spores. 4. Dark swollen hyphae of Diplodia.

Diplodia Zeae (Schw.) Lév.

Pycnidia black and spherical to pyriform, those forming on the husk or stalk developing within the tissues and breaking through at maturity, the greater number of pycnidia, however, occur between the kernels and are situated in a stroma. Conidia dark brown, cylindrical to elliptical, obtuse, straight or usually slightly curved and 1-septate; one to several oil drops in each cell; 5μ in diameter; spores germinate in 18-24 hours in 3 per cent glucose agar at 26° C; in somewhat longer time when grown on corn agar; germ tube arises from near distal end at each spore.*

Distribution. A serious parasitic disease generally found where corn is cultivated, particularly in Illinois,* Iowa, and Nebraska.

Poisonous properties. This fungus is widely distributed in ears of corn and may be responsible for forage poisoning.

^{*}Heald, F. D.; Wilcox, E. M.; and Pool, V. W. The Life-history and Parasitism of Diplodia Zeae (Schw.) Lév.

*Burrill, T. J., and Barrett, J. T. Ear Rots of Corn. Bull. Ill. Agr. Exp. Sta. 133:65-109. 11 pl.

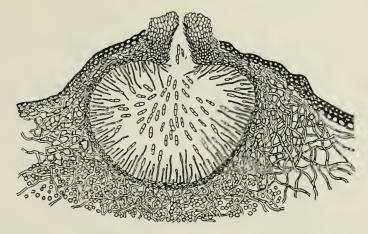


Fig. 103A. Cross-section of the pycnidium of Diplodia Zeae on a corn kernel showing sporophores, conidia and mycelium. After Burrill and Barrett.

Although the fungus has been cultivated, the toxic substance has not been isolated. The writer fed mouldy corn meal to cats and rabbits. It produced injurious effects in kittens; three of these animals fed with mouldy corn meal and milk died from the effects. Unfortunately, in this case, different moulds, Aspergillus glaucus, Fusarium, and Diplodia, were used.

Dr. Erwin F. Smith and Florence Hedges write as follows of this fungus: "It is also worthy of inquiry whether this fungus may not be the cause of the so-called 'cornstalk disease' prevalent among cattle in the west. It is also possible that to Diplodia should be referred the great numbers of deaths of negroes in the south during past three years (1906-1909) from the so-called pellagra, following the consumption of mouldy cornmeal and mouldy hominy. This fungus is also the cause of mouldy corn in Italy. The only other fungi we have reason for suspecting in this connection are species of Aspergillus." *

There are striking similarities between the so-called forage poisoning of cattle and the Pellagra disease in Italy and they are probably referable to some of the fungi found in corn. Dr. Miquel* in 1838 suggested that a Mucor was the cause of Pellagra.

Fusarium, Lk.

Mycelium spreading, more or less effuse; conidia spindle-shaped or sickle-like, many-celled at maturity, conidiophores branching, conidia borne at the apex. A genus of numerous species, many of which are of uncertain affinity, usually found on dead organic matter but several are known to produce diseases of cultivated plants, like F. Lycopersici, Sacc., which produces the "Sleeping Disease" of tomatoes, the mycelium occurring in the vessels of the roots and causing a wilting. The Fusarium limonis, Briosi, produces a maldi-gomma, or foot-rot, of orange and lemon trees. The Fusarium vasinfectum, Atks., produces a disease of cotton, known as "frenching." The cotton wilt is caused by a species of Fusarium and the perfect form of this fungus according to E. F. Smith is Necosmospora.

^{*} Diplodia Disease of Maize (Suspected cause of Pellagra). Science 30:60-61.

^{*} Die Noord-Nederlandsche vergiftige Gewassen. 43 Amsterdam 1839.

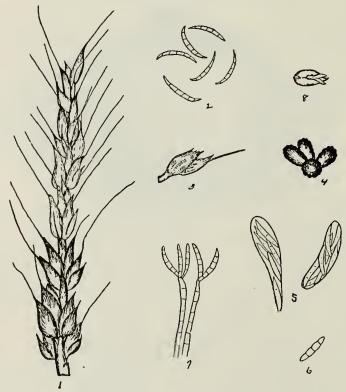


Fig. 104. Wheat Scab (Gibberella Saubinetii), perfect form of Fusarium roseum I.

1. Wheat affected with wheat scab, upper portion destroyed. 2. Glumes covered with perithecia. 4. Perithecia. 5. Asci from perithecia with ascospores, one of these enlarged at 6. 7. Conidiophore and spores grown in agar. After Selby.

Fusarium roseum, Link

Mycelium whitish or varying from yellow to orange, appearing at the time when the grain begins to turn; the head, or part of it, has a whitish appearance and the chaff is glued together; conidiophores branched, spores terminal or lateral, crescent shaped at first, 1-celled, finally 2 or more celled; color of the conidia white or in masses orange or pink. According to Saccardo the ascigerous stage is the Gibberella Saubinettii (Mont.) Sacc. with gregarious perithecia, coriaceous, or somewhat membranaceous; somewhat blackish in color, asci oblong, lanceolate, ascospores fusiform, 3-celled. Definite cultural experiments have not been made in this country to determine the relation of this fungus to the F. heterosporum.

According to Burrill and Barrett* several forms of Fusarium occur on corn. Saccardo in a letter to the writer identified the common Iowa Fusarium on corn as F. heterosporum.

The Fusarium heterosporum Nees, is common in parts of Germany, and Tubeuf quotes Frank as stating that the destruction of ryc is total in some

^{*} Bull. Ill. Agrl. Exp. Sta. 133.

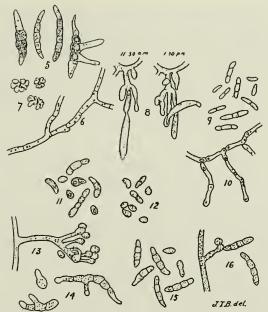


Fig. 105. Fusarium. 5. Macroconidia of Fusarium with the felty mass of mycelium. Produces a deep pink color. 6. Mycelium. 7. Corroded starch grains. 8. Conidiophores or sporophores. 9. Microconidia and macroconidia of another corn Fusarium frequently infecting isolated grains. 10. Mycelium of the same. 11. Microconidia and macroconidia of another Fusarium on corn, which produces a dense felty mass extending between the kernels to the cob. 13. A spore producing lyphae in prune juice culture. 14. Germinating spores of one of the species. 16. Hyphal branches of the same, with microconidia and macroconidia. After Burrill and Barrett.

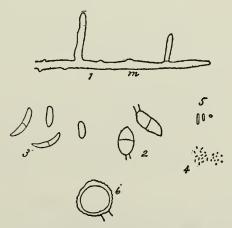


Fig. 106. Moulds and bacteria from corn. 1 and 3. Fusarium heterosporum I Mycelium. 3. Conida. 2 and 6. Other moulds. 4 and 5. Bacteria. After Pammel and King.

places, the fungus investing the whole kernel. Rostrup mentions it as destructive to germinating barley. It also occurs upon ergotized rye and is regarded by some mycologists as distinct from Fusarium culmorum. It is probable that the various species of Fusarium infesting cereals should be referred to one species.

Poisonous properties. Whether this fungus is responsible for the disease referred to by Dr. Mayo and other veterinarians, has not been definitely determined. It is true that experiments made by Dr. Bitting, this writer, and others, show that no doubt the Fusarium fed in considerable quantities to cats and dogs has had an injurious effect. Cats did not relish milk in which this material had been placed. If nothing more, Fusarium may be looked upon as producing stomatatis. Prof. Sheldon refers this fungus to Fusarium moniliforme. In the diseased horses reported by Dr. Peters the horses would lose their hair and hoofs and were said to be alkalied. Cattle and hogs were likewise said to lose their hair. Feeding experiments conducted on hogs with this corn as well as with pure cultures reproduced the symptoms in experimental animals. In this connection this statement of Dr. Law's is of interest:

Fodders affected with cryptogams or bacterial ferments are undoubtedly at times the cause of encephalitis. Veterinary records furnish many instances of wide spread attacks of stomach staggers, abdominal vertigo, and cerebro-spinal meningitis in wet seasons, when the fodders have been harvested in poor condition or when from inundation or accidental exposure they have become permeated by cryptogams and microbes. Among comparatively recent accounts of this are those of Martin and Varnell (nusty oats), Lombroso, Depre, Erbe, Pellizi, and Tireli (smuts), Bouley and Barthelemy (musty fodder), and Ray (fermented potatoes). One of the most extended local outbreaks of cerebro-spinal congestion I have ever seen, occurred in the pit mules of the Wilkesbarre coal mines, while fed on Canadian hay which had been soaked with rain in transit and had undergone extensive fermentation. It should be noted that there were the attendant factors of overwork, in anticipation of a strike, and a Sunday's holiday above ground in a bright summer sunshine.

The experimental administration of moulds, smuts and microbes, have in the great majority of cases led to little or no evil result (Gamgee, Mayo, Dinwiddie, etc.) and there is a strong tendency to discredit the pathogenic action of these agents in reported outerals. The safer conclusion perhaps would be, to recognize the fact that they are not equally pathogenic under all conditions of their growth and administration. The oftrecurring epizootics of brain disease in connection with wide spread spoiling of the fodders in remote and recent times, probably imply that cryptogams or microbes and their products, plus some condition not yet fully understood, are efficient concurrent factors. If we can discover this as yet unknown factor and demonstrate that it operates with equal power in the absence of cryptogams and ferments, as in their presence, it will be logical to pronounce these latter as non-pathogenic under all circumstances. Until then cryptogams and bacteria must be held as probable factors.

In recognizing how much cryptogams and bacteria vary under different conditions of life, and what various products they elaborate at different stages of their growth, we can theoretically explain the absence of the disease at one time and its presence at another under what seem to be identical circumstances, as also the variety of symptoms shown in different outbreaks. While this causation cannot be said to be absolutely proved. it is not antagonistic to the facts in many of the best observed outbreaks, and may serve as a hypothetical working theory until actual demonstration can be furnished. The affection suggests a narcotic poison introduced from without, rather than a disease due to a germ propagated in the system.

In all probability as we learn more of the true pathology of the disease, we shall come to recognize not one, but several toxic principles, and several different affections each with its characteristic phenomena in the somewhat indefinite affection still known as cerebro-spinal meningitis.

The malady has been described in horses, oxen, sheep, goats and dogs, attacking by preference the young, which are not yet inured to the unknown poison, and by preference in winter and spring, the periods of close stabling, dry feeding and shedding of the coat.

Dr. R. A. Craig of Indiana reports as follows in regard to interesting experiments made at the Indiana Station:

In January, four sacks of spoiled, mouldy corn were gathered from a stalk field adjoining a field in which cattle had developed cornstalk disease. A healthy heifer weighing three hundred and fifty pounds was fed four to five pounds (twelve to seventeen ears) of this corn twice a day. In addition stover was fed. On the afternoon of the sixth day of the test the heifer appeared weak, went down in the stall and was helped up twice in the afternoon. When down she struggled some, and when helped up "shivered" as if cold. In the evening she was still trembling and appeared weak. The weakness disappeared the following day. A few days later a slight twitching of the body muscles was noticed. The feeding test extended over a period of sixteen days. Her appetite remained good throughout the test.

Dr. Craig adds the following:

During the fall and early winter of 1898-'99, Bitting reported losses in horses and cattle, supposed to have been due to feeding on spoiled corn. By feeding corn meal that was inoculated with a pure culture of a mold (Fusarium sp.) made from the spoiled corn, he produced salivation and redness of the gums of the two horses used in the experiment. Later spoiled corn was fed. On the fifth day one horse showed a slight salivation, colicky pains and diarrhoea. On the seventh day, noticeable incoordination in moving about and stupor. For two days the animal stood with the head pressed against the wall. A quick recovery followed and the nervous disease from which horses were reported as dying did not develop. The second horse showed nothing more than a slight irritation to the mouth.

Because of the close resemblance between toxic poisoning from sorghum and the symptoms of corn stalk disease, Price deemed it advisable to examine cornstalks for the substances which produce prussic acid in plants. Samples of stalks from fields in which cattle had died were obtained. In these samples he discovered an enzyme which had the property of decomposing a glucoside (amygdalin) and thereby poison as a result of enzyme was found. However, no glucoside capable of forming this poison as a result of enzyme action was found. The results were not regarded as conclusive, as only a few samples were examined, and the failure to discover a suitable glucoside did not prove its absence in the corn plant, or in other plants in the field.

Dr. Peters says in regard to the feeding of moldy corn to horses as follows:

Numerous reports have been received from stock owners of a disease which they call cornstalk disease or spinal meningitis which affects horses in the stalks and also some which have not been in the stalks.

This disease is very rapid in its course. For this reason it is sometimes difficult to see animals alive or in the beginning stages of the disease. In the later stages the animals are usually in such violent excitement that the symptoms have to be studied from a distance. One peculiar feature about the disease is that it comes on without warning, often attacking an animal while at work. One of the first symptoms noticed is the refusal of feed. Some have observed an excessive thirst and a difficulty in swallowing. The head is drooped in a very peculiar manner, denoting dullness. The eyes become very dull and later almost totally blind. This is usually followed by delirium and death. When a horse becomes affected in the stall it sometimes presses its head against the manger or wall and as this symptom increases in violence it is not uncommon to find the stall and manger demolished. Another peculiarity of the disease is that just before the animal becomes violent, one can cross its legs and the animal will remain in the position semi-conscious for some time.

This disease has been attributed to many causes. It is practically conceded at this time that it is due to a fungus found on the food administered. Feeding experiments with mouldy corn at our Station and other Stations, have proven that mouldy corn is capable of producing this disease. In March, 1902, a quantity of mouldy corn, which was taken from cribs of a farmer at Graf, Nebr., who had lost a number of horses with this disease, was fed to four horses. These horses were fed exclusively on this corn with a small quantity of good hay and on April 2 two of the horses were found to be affected, the symptoms being

the same as those in animals that died in various parts of the state. The black horse, John, was found in the afternoon of April 2 to be swaying in his gait. He refused feed, had the peculiar dullness of the eyes, and when his limbs were crossed he remained in that position entirely motionless for some time. The next morning the animal was very much worse and at noon he was killed so that an autopsy could be held. The post-mortem examination showed that all organs were practically normal except the brain which was much softer than normal. I will quote the description of the post mortem as given by Dr. Butler, which is as follows: "On removal of the brain the superior surface of the right cerebral hemisphere was noticed to be slightly flattened over the anterior half. Palpation revealed a soft spot at this place. An incision through the apparently sound gray matter revealed what Mayo described as a sereous abscess in which floated flocculi of broken down brain substance, which presented the appearance, as one stockman said, of a mixture of vinegar and curdled milk. This portion of softened and broken-down white brain substance is in no sense a serous abscess. The line of demarcation between the broken-down and the healthy brain substance was not clearly marked, but surrounding the completely broken-down portion of a zone probably half an inch thick that was softer than normal and of a slightly yellow color. The liquid in the cavity, and in which floated portions of soft and partially broken-down brain substance, was slightly yellow, but in no instance was clotted blood or any other microscopic evidence of a hemorrhage to be found."

Dr. Butler and Dr. Mayo conducted an experiment with some mouldy corn from a farmer who lost four registered Percheron horses at Wakefield, Kansas. Four hundred pounds of the worst of this corn and fifty pounds of the chaff and screenings were sent to the Agricultural College in Manhattan, Kansas, and a feeding experiment was started with two colts, twenty-three months old. The experiment began on July 16, when each colt received 1½ kilos twice daily. On July 22, they were fed 1½ kilos of corn and cob meal twice daily. On the 26th of July 1¾ kilos of the damaged corn, well ground, cob and all. This was continued until August 19. One colt died August 21. Another experiment was conducted with a two year old colt, but fed with mouldy corn and good prairie hay. The temperature of the animal varied from 101-102° F. This colt died on July 26, the feeding experiment having

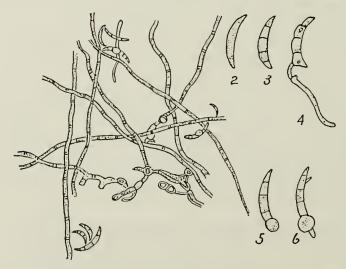


Fig. 107. Dermal mycosis associated with Sarcoptic mange caused by Fusarium equinum, conidia and mycelium. 2-6. Conidia (macroconidia) in various stages of development.

4. Germinating. After Melvin and Mohler.

begun on June 30. It was observed that after three weeks there were no notable changes except a gain in flesh.

It seems to me that there can be no question from these experiments that mouldy corn is dangerous to feed to animals.

Oxen, sheep, and dogs are also affected with a form of meningitis, due to mouldy conditions. Of course, it should be stated that in the above description by Dr. Law, no special mould fungus is referred to.

Fusarium equinum, Norgaard. Itch Disease of Horses

Mycclium immersed, septate and branched; conidia in cultures sickle-shaped, segmented; in hair sacs and sebaceous glands spindle-shaped or crescent-shaped bodies.

Drs. Melvin and Mohler supplement the above characters as follows:

The Fusarium possesses three forms of spores, the microconidia small and oval, non-septate or two celled; the marcroconidia, large falcate, with sharp lanceolate ends, 3-5 septate, forming many aerial threads; 25-55 μ long $2\frac{1}{2}$ 4 $\frac{1}{2}$ 4 wide; the chlamydospores oval or oblong, thin walled, densely granular, 8-15 μ in diameter. The macroconidia occur during the later stage of growth. On culture media there is a white growth which becomes slightly colored. The most favorable medium is potato and sterilized bread, but it grows well in agar, glucose, or saccharine agar.

Distribution. North America, California to Idaho.

Pathogenic properties. In December, 1901, Victor A. Norgaard contributed to Science an account of a disease affecting horses, said to be produced by a fungus to which he suggested that, pending investigation, the name Fusarium equinum, nov. spec. be given. The following is an abstract of the article in question:

An epidemic skin disease appeared among the horses on the Umatilla Indian Reservation, Pendleton, Oregon, upwards of sixty percent out of six thousand horses having been affected. The disease manifested itself through severe itching and loss of hair over almost the entire body. Many of the animals died of starvation. An examination of samples of the skin was made in the Pathological Division of the Bureau of Animal Industry and the presence of Sarcoptes equi observed. However these parasites were not present in sufficient numbers to account for the almost complete alopecia, and examination of samples almost entirely denuded of hair failed to show their presence. Microscopic examination of sections of the skin stained with borax blue showed the presence of large half-moon, spindle shaped bodies, deeply stained, in the hair sacs and sebaceous glands. Further culture produced from one to five circular colonies of a fungus which grew rapidly and assumed a salmon pink color. Cover-glass preparations made from these colonies contained numerous sickle-shaped segmented spores, characteristic of Fusarium. Of the twenty-five known varieties of this fungus, according to Dr. Erwin F. Smith, hitherto none has been known to be pathogenic to animals.

Drs. A. D. Melvin and J. R. Mohler have given a somewhat more extended account of this form of dermatomycosis. They found present with the disease the Sarcoptes scabei. In 1901 the disease appeared in a very aggravated form, some 2,500 animals were diseased out of 6,000 animals on the Umatilla Indian reservation. It is supposed that this disease was introduced from California in 1902 from trailed horses. It appears that the fungus apparently enters the hair follicles, penetrates between the cells of the epidermis or abrasion of the skin and involves the surrounding cuticle, causing irritation, followed by pruritis, the animal attempting to rub itself against anything with which it comes in contact. When the scurf is rubbed off by the finger nail there is left in its place a red moist denuded surface. It affects almost the entire body except the knees and hocks. The crusts are of gray color at first but turn darker. When the tissue

is examined microscopically, the spores are found to be abundant in the hair follicle, the fungus causing the hair to drop out. The inflammatory process spreads in the sebaceous glands causing a suppression of the excretion with the formation of crusts anl scab. Occasionally Sarcoptes were found but Drs. Melvin and Mohler do not believe that they were the principal cause of the disease, although when present the animal parasite may aggravate the trouble. All ages and breeds of horses are susceptible as are both sexes. The animal stands around the rubbing posts all day and finally dies.

Rabbits, dogs, guinea pigs are immune. Experiments with horses were not successful, which these writers think may be because the right stage of the fungus was not used for inoculation experiments.

Treatment. The authors recommend dynamo oil and sulphur in the proportion of one pound of the latter to a gallon of oil. Coal tar sheep dips have also been used.

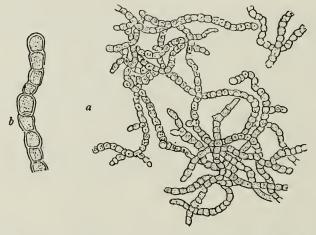


Fig. 108. Favus of mouse (Oospora porriginis) from a culture. a. Mycelial threads. b. Λ single thread more highly magnified. After Flügge.

Oospora, Wallr.

Fungus with small tufts spreading or pulvinate, mucedinous, loose, or somewhat compact; fertile hyphae, short, with few branches; conidia transparent, usually in chains, globose or ovoid, hyaline or slightly colored.

Oospora porriginis (Mont, and Berk.) Sacc. Achorion Schönleinii, Remak.

Favus, Tinea favosa. Honeycomb Ringworm

Mycelium flexuose, simple, branched, or forked, continuous conidia, ovoid, triangular or somewhat cubical, varying 3-6 μ in diameter; mycelium in masses with granular protoplasm occasionally branched at the end, the ends swollen, club-shaped, branches of the mycelium with lateral branches; spores oval, round or angular, 3-8 μ long and 3-4 μ wide, single or in chains. The threads of the fungus are readily detected in the bulbs and the shafts of hairs when sodium nitrate or potassium hydroxid is added, but at a distance of two inches the fungus cannot be detected. Sections of the nail stained also show threads

of the fungus. This fungus has been cultivated in ordinary agar or by Kral's method. It grows well at higher temperatures, and in 24-48 hours the fungus threads appear. The spores germinate at 35° C. in 14 hours, and in 24 hours a fine mycelium appears in the air. In nutrient media like potato, gelatin and agar, chlamydospores and yellowish bodies appear. The organism requires higher temperatures for its best development, the optimum is 35° C. The organism from some of the lower animals, however, grows at lower temperatures.

According to Walsch the best development of the fungus in the hair is near the upper end of the root, from here it extends upward or downward; the mycelium may be exfollicular or on the surface of the hair.

The favus of man does not differ especially from that of animals except in color, and in the shorter duration of the disease in animals.

Distribution. Widely distributed in Europe and North America, but, according to Hyde and Montgomery, less common in the United States, Austria, and England, than in France, Scotland and Poland.

Pathogenic properties. This form of dermatomycosis known as Favus was discovered by Schönlein in 1839. In the middle ages, it was known as Tinea, meaning a moth or worm. Previous to the discovery of the organism by Schönlein, various troubles were classed as favus.

Heusinger suspected the fungus nature of the disease as early as 1826. Remak, in 1845, cultivated the organism upon apple and transmitted it to his arm. He named the fungus Achorion Schönleinii and in medical literature it is frequently referred to by this name.

The favus organism of mice was discovered in 1850 by Bennett and recognized by subsequent investigators like Schroeder and Simon. Favus of cats, guinea pigs, and dogs was recognized by Saint Cyr. Gerlach found it in birds, and Gruby, three years after the discovery of the organism by Schönlein, found it in hairs on the heads of children and the hairs of the beard. In recent times, various views have been expressed with reference to the nature of favus and trichophytosis, it being held that these diseases are produced by different fungi. Pick in 1887, Walsch in 1896, and others considered the fungus to be polymorphic. Quincke distinguished three varieties but pathologists are not agreed on this point. Plaut, in his discussion of the parasitic fungi, divides them into the following groups: the Favus and the Trichophytic groups. According to this view, the favus of man and that of animals are regarded as distinct fungi.

The disease in man generally appears where hairs occur, but may appear also in other parts of the body as the eyes, nails, etc., seldom becoming general. Favus of the nails is called Favus onychomycosis. In lower animals the disease may occur on the head, nose, ears, back and, more frequently, is generalized. Formerly this disease was common among the poorer classes. Today, it is not common in France, Holland, Scandinavia, Germany, England, Switzerland, Japan or America; on the other hand, it is rather frequent in Russia, Scotland, Italy, Spain, Asia, Austria and Egypt, young individuals being more susceptible to it than are older persons, probably acquiring the disease by contact. The disease is spread through spores of the fungus. The mycelium itself, according to Grawitz, is not capable of spreading the disease of animals. In man it is recognized by the development of minute yellowish or reddish points; delicate vescicles may surround these spots. Later, the fungus may develop a

crust; the odor is very disagreeable, being compared to the odor of mice or the urine of cats.

Mice are most susceptible to the disease, cats coming next since they come in contact with mice which have favus, then dogs by means of the cats and finally guinea pigs.

According to the recent investigations of Frank (1891) three different types of fungi were isolated from mice, and Unna and Neebe in 1893, concluded that no fewer than nine species existed, three aerophilous and six aerophobic, as follows: Achorion eutythrix, A. didikroon, A. atkaton, A. radians, A. akromegalicum, A. demergens, A. cysticum, A. moniliforme, and A. tarsiferon, but these may probably be regarded as one widely-polymorphic species.



Fig. 109. Epidermis invaded by Sporotrichum. a—inferior portion of the stratum corneum; b—superior portion of the rete. Both exhibit long mycelial threads, with a few ramifications and a small number of spores. After Kaposi.

Sporotrichum, Link

Hyphae, branching irregularly and repeatedly, septate or continuous, usually equally procumbent; conidia, acrogenous at the apices of the main and lateral branches, usually solitary beneath, ovoid or subglobose. This genus differs from *Botrytis*, especially in all the hyphae being procumbent and the conidia subsolitary; from *Trichosporum*, in never being dark colored. Very many species, imperfectly described by older writers, show mere forms, or inycelia.

Sporotrichum Furfur. Rob

Pale yellow or yellowish brown to dark brown or brownish-red spots, varying in size from that of a lentil to that of a hand, either smooth or shining or dull exfoliating. Found on the breast, stomach, or back. Never upon the hands, feet, seldom on the face. Slender hyphae 3-4 μ wide, 7-13 μ long, variable as to length and thickness. The spores are clustered resembling oil drops. On potato a characteristic growth of yellowish, orange red brown, blackish or greenish color. Old culture is grayish, brownish or violet color. In 3-4 days a whitish gelatinous mass forms, which in 3-4 weeks covers the whole surface. Conidia are oidium-like, surrounded by thick hyphae, occur in scales, 4-7 μ , spherical. In cultures budding occurs.

Distribution. Common in some localities in Europe and America.

Pathogenic properties. Fehr in 1840 observed that most of the inhabitants in a Swiss village were infected through cattle. Bazin in 1853 observed that many cavalry men were infected through horses. Papa in 1840 observed that this disease was frequently transmitted to men. In cities it chiefly occurs in cats and dogs, and through these it is conveyed to men.

It is especially common in people with tender skins and in tubercular patients, and is more common with women than men. In 1846, Eichstedt dis-

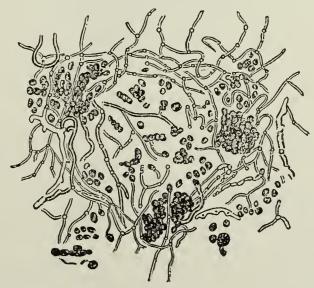


Fig. 110. Sporotrichum Furfur. After Kaposi.

covered the cause while Robin named the fungus Microsporon Furfur. Köbner, in 1866, made the first inoculation experiments.

Grawitz, in 1876, first cultivated the organism. His work was followed by that of such other investigators as Schlen in 1890 and Koltjar in 1892, who succeeded in transmitting the cultivated fungus to guinea pigs and gave it the name Oidium minimum. It has also been named Oidium Furfur. Vuillemin also cultivated the fungus to which he gave the name Malassezia Furfur. Pure cultures of the organism grown on potato were transmitted to man.

Treatment. The best treatment, as recommended by Hyde and Montgomery, is a hot bath, the skin being rubbed with soap; following this the skin is bathed with clean water and sponged with a solution of sodium hyposulfite, 1 drachm (4) to the ounce (30).

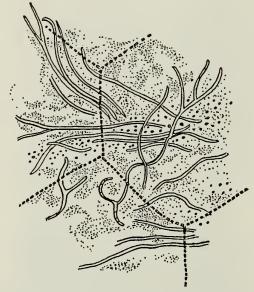


Fig. 111. Erythrasma Fungus. Sporotrichum minutissimum. After Hyde and Montgomery.

Sporotrichum minutissimum (Burckhardt.) Pammel. Erythrasma

It begins as small brown or reddish patches which become confluent; these spots may become as large as the palm of a hand and occur in the axial region; the scales contain Leptothrox-like threads which are branched and septate; conidia small, round or angular. It grows well on agar agar, glycerine agar, gelatine, potatoes, and in blood serum. In nutrient media, branching, septate hyphae $0.8-1.3\,\mu$ in thickness and $5-15\,\mu$ in length are seen; the short hyphae break up into numerous spores.

This disease was first observed by Burckhardt in 1869 and since has been observed by others. It occurs in the form of roundish or punctiform patches, sharply contrasted with the adjacent tissues. The younger areas are livid red while the older are yellowish or brownish.

A so-called Dhobie itch of the Philippine Islands is in part caused by this fungus. Though Hyde and Montgomery state that there are two other types of infection known by this name, one is trichophyton and one of bacterial origin.

Sporotrichum giganteum, (Unna). Pammel

Spores are free or in chains in nutrient medium 1.5-7 μ in diameter, the oval 4-5x5-6 μ , yeast-like budding resembling Oidium lactis also occurs; ectospores as well as chlamydospores present, the latter 8-12 μ in diameter with strongly refringent bodies: in cultures ray like, the rays consisting of hyphae and spores, in liquid media only hyphae with ectospores. In the hair, knot-like bodies are formed with spores and hyphae embedded in mucilage.

The Colombia disease was described as *Trichosporon giganteum*, Unna. It is a polymorphic fungus and the T. ovoides, Behrend is included but by some is regarded as different. The knotty masses of hair are less thick, and the spores are oval in shape; gelatine not liquefied. The superficial colonies resemble Oidium lactis. Chlamydospore $4-12 \mu$ in diameter.

Other species of *Microsporon* have been described like *M. canis* in dogs. *M. tigris*, the *M. equi*, in horses and colts and another species in cattle. In calves a similar form occurs. The sheep are said to have the disease on the neck and breast; it also occurs in hogs, goats, and birds. These forms are said to differ slightly clinically. Pus is formed in follieles, and the hair is especially prone to drop out. A bad smelling liquid of a reddish color occurs. In lesions large spores, the ectospores, occur. The Oidium chain like spores occur chiefly in the roots of the hairs.

Pathogenic properties. The disease is especially common in animals in which the skin is naturally fine, thin, and dry, and covered with hair sparsely, more common in the Arabian Barb, English racer, and American trotter of nervous organization than in heavier draft breeds. Old horses are more subjeet than young ones. Extended desquamation, excessive production of epidermal seales without any elevation of the skin, seurfy products may be found in patches seattered over the body; generalized or circumscribed as to the head, ears, crest and tail; the hair may be pulled out with great ease. In eattle it occurs on neck and develops in connection with anaemia, spoiled fodder, and constitutional predisposition. Affects especially the head, neck, and back of dogs gorged with dainties and those becoming aged. The affected parts are covered with a floury or bran-like product lying upon a dry surface, the affection being usually limited to certain areas more or less destitute of hair. In the cat it may affect the whole dorsal aspect of the body, being associated with extreme electrical susceptibility, the hair when touched, collecting in tufts. The scaly product is abundant.

Animals are said to spread trichophytic fungi, which supposition is important from a hygenic point of view. Since the disease sometimes occurs in school children, separate hooks for clothing and separate towels when bathing the hands and faces are recommended. A one per cent solution of bicloride of mercury will kill the fungus.

The disease was first described by Osorio in 1846, and was then thought to be confined to Colombia where it was called Diedra (Stone), but later was found in Europe, and Vuillemin reported it from Paris in 1902. Desenne of Paris discovered the hyphae of the fungus and Malcolm Morris the spore-like bodies. Behrend in 1890 succeeded in cultivating the fungus.

Sporotrichum tonsurans. Barber's Itch

Hyphae slender 1.5-2 \mu in diameter, straight, undulated, dichotomous, septate or non-septate, penetrate the hair follicles forming a matted mycelium, small pustules and scabs; in places devoid of hairs it forms red, scaly spots, discs and circles. The fungus is found between the uppermost layers of the epidermis just beneath the corneous stratum; the conidia are small, spherical or elliptical, sharply defined 2-3 μ in diameter; the spore masses surround the root of the hair and are frequently densely and closely arranged like beads; in culture media like agar, a many-rayed fungus occurs, the color varies with the medium, yellow, Bismark brown, cherry red, violet, rose, brown, blackish brown; gelatine liquefied; spores swell after a few hours and produce 1-2 germ tubes from a single spore; mycelium with occasional swellings, ectospores formed in 60-96 hours, also small, branched air hyphae; small conidia 1.5-3 \(\mu_i \) in diameter borne on short, lateral branches. The Botrytis-like spores rise on the long, thin, curved air hyphae. Oidium budding does not occur in nutrient media. The spores retain their vitality for six months but exposure to 45° C. for a few hours will kill the organism; it is sensitive to sunlight and common disinfectants. The fungus is polymorphic, one form having been classed, by Sabourne, with Botrytis. The large-spored trichophyte found on the scalp germinates at 37° C, in a few hours; but at room temperature, a much longer time is required.

Conidia 5 μ in diameter, an abundant mycelium with dust-like growths, and, in three days, oidium-like spores, as well as ectospores, are produced on the potato, the disease being known as *Tinea Sycosis*.

The *T. circumscripta* produces, in animals, cherry patches each with a raised border and scales, and is also found on the head, arms, and ncck, of man. *T. disseminata* produces small red pustules.

To T. tonsurans, also, is attributed Eczema marginatum which Kobner, while making a study of trichophytic fungi in 1864, recognized as a trichophyte. He also determined that the fungus on the nails, described in 1853 and 1855 by Baum and Meissner, was a trichophyte.

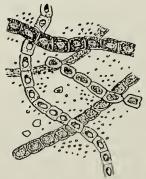


Fig. 112. Barber's Itch. (Sporotrichum tonsurans) Filaments and spores. After Hyde and Montgomery.

Gerlach, in 1857-1859, demonstrated a trichophyte in bovine animals, and other investigators, later, recognized the case as Herpes tonsurans. S. tonsurans has also been described under Trichophyton tonsurans and as Oidium tonsurans. Unna, in 1897, from twenty cultures described four species, T. oidiophora, T. eretmorphoron, T. atractophoron, and T. pterygodes. It is probable that Sporotrichum tonsurans is a very variable species. Lindau places it under the genus Oospora, but it seems preferable to call it Sporotrichum.

Distribution. Occurs in Europe, North and South America, Asia and Africa. Pathogenic properties. In cattle, small, round, sharply defined spots occur which are covered with scabs and scales which project more or less above the skin and vary in size, some being as large as the palm of the hand. Underneath the scales, is a purulent fluid with hollows that represent the empty follicle. In man as well as in animals, the hairs can be pulled out very readily. The eruption lasts from six to twelve weeks, outbreaks occurring from rubbing or scratching as a relief from the itching sensation that accompanies the eruption. In sucking calves it occurs chiefly about the mouth and is called "doughy mange" and, according to Hahn, is produced by the fungus T. tonsurans.

Most of the varieties can be transmitted to guinea pigs, cats, and dogs and have even been transmitted to man during the process of sheep shearing. Healing takes place when the animal is inoculated subcutaneously. The large-spored form, occurring on the scalp, forms pus and resembles moist eczema. Children take the disease from calves and by playing with cats and dogs.

Mycosis of the beard exists in two forms; non-infectious and infectious. In Sycosis parasitaria, the disease is accompanied by a severe inflammation of the hairy parts of the skin leading to infiltration and suppuration. Sabouraud classifies the parasite into a dry and a pus favus.

The Eczema marginatum supposedly caused by the same fungus was first described by Devergic in 1854-1855; Berensprung having discovered the same fungus in 1855. It is slightly contagious, and more frequent in men than in women. Another form of the disease occurs in the mouth. In sheep the wool is felted and beneath it are bran-like, scabby parts, the fleece becoming very ragged in appearance. In poultry, it shows itself by the loss of feathers. In horses, it occurs most often on the seat of the saddle. The spots vary in size, and the surrounding hair can be pulled out easily. In dogs, it affects the head and extremities. Usually the spots are round at first and sharply defined, later becoming hairless patches; occasionally they are dirty gray scabs.

Ringworm of the body or *Tinea circinata*, is characterized by the occurrence of one or more pea-shaped or large circular reddish patches which are on about the same level as the integument and rarely 5-6 inches in diameter. In some forms there is itching. This Trichophyton was discovered by Gruby in 1844. This fungus can be readily recognized by making microscopic mounts. The mycelium is less branched and the threads are more slender than in the form previously described. The spores are like strings of beads.

The ringworm of the scalp, Tinea tonsurans, is a disease, chiefly, of children, especially of those in schools. It differs from the preceding form in the fact that the fungus makes its way into the hair follicles. The patches are, at first, circumscribed, about the size of a small coin, covered or partly covered with roundish patches of slate gray color or a dirty yellow. The fungus is called Microsporon adomini.

Hyde and Montgomery state that there are at least two distinct and unrelated forms capable of producing ringworm, the *Microsporon adouini*, a small-spored fungus, and the *Trichophyton*, or large-spored fungus. The Microsporon appears under the nicroscope in the form of a large number of round spores, irregularly grouped or massed about the follicular portion of the hair. The mycelial threads are all within the hair proper while the spores terminate fine threads on the other surface of the shaft. The spores of Tricophyton vary greatly in size and are much larger than those of Microsporon. They are cuboidal, oval, or irregularly rounded. They occur in chains, up and down the hair or shaft. The mycelium is found without, never within, the hairs. The spores may be within (endothrix) or without (ectothrix).

Oidium albicans. (Robin.) Rees. Thrush

Forms a mould-like growth in the mouth of man and lower animals. Vegetative cells, yeast-like, spherical, elliptical, oval or cylindrical, 5-6 μ long, 4 μ wide, the elongated hyphae-like bodies variable in length; conidia elliptical in chains; grows well in nutrient media where it produces superficial, spherical, white, wax-like, granular colonies, varying color from reddish to white; chlamydospores in nutrient media and occasionally in the epithelial layer; it does not ferment lactose and saccharose, but ferments levulose and dextrose. According to Brebeck and Fischer there are two morphological forms of the organisms, a small oval and a large-spored form; however, this distinction is not generally recognized.

Distribution. Widely distributed in the United States, also in other countries, Germany, Austria, France, Italy, and Great Britain.

Pathogenic properties. John recognized the disease in 1816, while Buchner gave a somewhat detailed description in 1841. Langbeck and Berg dicsovered the fungus in 1839. It was thought by them that it was the cause of typhoid fever. Langenbeck demonstrated that the Fungus could be carried from a child, sick with the disease, to a healthy individual. Gruby, in 1847, described the fungus under the name of Apthaphyta, placing it near the fungus Sporotrichum, while Robin, a French author on parasitic diseases, considered the fungus to be an Oidium, naming it Oidium albicans, a name frequently used by authors. Rees, however, placed it with the yeasts. Monilia candida is regarded by Plaut and Lindau as a synonym. Grawitz, in 1877, made pure culture of the fungus and succeeded in producing the disease in guinea pigs. Klemperer found that when the fungus was inoculated into the circulatory system of guinea pigs, general mycosis resulted. Limossier and Roux (1889-1890) in their monograph, state that the mycelium occurs in the blood vessels of inoculated animals.

The fungus is very common in some sick chambers in regions where the disease is prevalent. It is most abundant in sucklings.

It occurs frequently in children of premature birth, and in weak children; the fungus is also found in aged persons, suffering from disability; it occurs chiefly upon the mucous membrane of the mouth, pharynx, and oesophagus; more rarely, upon that of the stomach, intestine and vagina, and upon the nipples of nursing women and bovine animals. It has also been found in the liver, kidneys and lungs; it penetrates the epithelium and even into the underlying, connective tissue; it is spontaneous in such animals as calves, birds, and

foals. The disease is fatal in many cases, some authors estimating the death rate as high as 22 per cent. Inoculated guinea pigs show a rise in temperature at first a lowering, accompanied by albuminaria, loss of flesh, and diarrhoea, death occurring in from 3-7 days. Immunity may be obtained in guinea pigs by beginning with small doses, and increasing these gradually to three times the strength. The product produced by the fungus is poisonous; 20-40 cc. of the whole substance will kill a guinea pig weighing one kilogram.

Dr. Stuhr has contributed the following upon this subject:

Thrush is a mycosis of the mouth affecting children, calves, foals, and poultry, and is characterized by the formation of white patches upon the mucous membrane, which vary in size from points to large areas. It may involve the pharyngeal and laryngeal mucosae by extension. The disease is transmissable from man to animals. Young age, a weak constitution, gastric indigestion, uncleanliness, milky and starchy diet predispose. Decaying food in the mouth offers a suitable place for the growth of the fungus.

Etiology. The specific cause of thrush is a vegetable parasite, Oidium albicans, first described by Berg in 1840. It is one of the branching fungiclosely related to the yeasts and grows readily on sour milk, in saccharine substances, on decayed wood, and on fresh cow manure. Calves fed milk from wooden pails which are not kept perfectly clean are particularly liable to contract the disease. The fungus descends into the epithelium and sometimes into the subjacent connective tissue, causing inflammatory infiltration and superficial necrosis.

Symptoms. The mucosa is diffusely red, swollen and tender, and shows adherent white patches, varying in size, surrounded by a red inflammatory zone. When these white spots are rubbed off, shallow red ulcers are exposed. When the inflammation in the mouth is severe, or when the disease spreads to the pharynx and interferes with deglutition the prognosis may become serious. Usually, however, the disease is benign and yields readily to treatment.

Lesions. These are usually superficially located and rarely extend deeply into tissues. They begin with diffuse reddening of the mucous membrane and the formation of a somewhat shining, slimy, adhesive layer of grayish-white matter which is said to have an acid reaction. Later whitish dots appear upon the surface and gradually spread, sometimes coalescing. These whitish patches are false membranes composed of detached epithelial cells with a ramifying network of parasitic threads. The white color of the false membranes is markedly in contrast with the congested surrounding tissue. While the lesions are ordinarily restricted to the mouth they may involve the pharynx, oesophagus, (in chickens), larynx and even the stomach and intestines. Metastasis may occur and the fungus be carried to various parts of the body.

Treatment. This is aimed at the destruction of the fungus and for this purpose many substances have been recommended. The mouth should be cleansed at frequent intervals with solutions of borax, sodium hyposulphite, permanganate of potash, or chlorate of potash, etc. The system should be built up by feeding soft nutritious food, and the sanitary conditions should be improved.

Oidium hominis. (Busse.) Pammel. Blastomycosis

Cells spherical or ovoid, variable in culture, 8 μ in diameter with strongly refringent bodies; in young cultures, nearly homogeneous and with oil drops; in old cultures, large cells with a thin membrane; culture, at first white, then grayish or yellowish, or yellowish-brown in plum cultures, in plum decoction

black, gas-producing in sugar medium, grows well in ordinary media but preferably in acid; grows best at high temperature. (An organism isolated by Curtis produced white colonies with oval, or club-shaped cells, frequently producing capsules.) Grows readily in nutrient media, does not form a pellicle on the surface in liquid media.

Descriptions of several so-called species of Saccharomyces causing Blastomycosis will be added to the above. In their development, blastomycotic fungi resemble true hyphal fungi rather than yeasts. It is convenient to discuss them here until their true relationship has been determined.

Distribution. Found in Europe and America.

Pathogenic properties. It was first isolated from the left tibia of a woman thirty-one years of age, the disease having first manifested itself by a purplish-red spot and swelling. An operation was performed, but it failed to relieve the trouble, new foci making their appearance after the operation and finally becoming general, being accompanied with pus formation. The patient died in 13 months, the lungs, kidneys, and spleen having become involved. There was no oedema, the organisms found in the lungs and kidneys being marked by small, nodular swellings.

Large amounts of culture, when inoculated into guinea pigs, dogs, and rabbits, produce the disease followed by death. In another case described by Curtis in 1895, the disease occurred in a young man. The organism isolated is pathogenic for rats, mice and dogs.

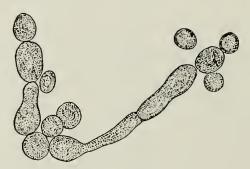


Fig. 113. Blastomycosis of the skin showing the elongated cells and budding forms. After Hyde and Montgomery.

Ziegler, in his General Pathology, summarizes our knowledge of the pathogenic properties of yeast as follows:

As parasites no importance has been attached to them until very recently, but the investigations of Busse, Buschke, Sanfelice, Curtis, and others have established the fact that there are also species of Saccharomycetes of pathogenic importance. According to these observations, the pathogenic yeasts can multiply in different tissues, in the skin, periosteun lungs, and glandular organs, and can excite either purulent inflammations or proliferations of granulation tissue, which run a course similar to that of an infection with actinomycosis or tuberculosis. In inflammatory foci, the yeast cells are for the most part provided with a capsule. They may give rise to tumor-like swellings. Through degenerate changes, crescentic forms may develop from the oval yeast-cells.

In solutions containing sugar the blastomycetes form oval cells. Reproduction takes place through budding and constriction; on any portion of the parent cell there may develop an excrescence, which is constricted off after it reaches the size of the mother cell. Under

certain conditions the cells may grow out into threads, but in these threads no subsequent segmentation occurs; jointed threads arise through budding (Cienkowsky, Grawitz). A dilute culture-medium favors the formation of threads.

Oidium granulomatogenes. (Sanfelice). Pammel. Blastomycosis

Forms nodular masses; grows in ordinary media; ferments sugar; uniform clouding of media; colonies white; the nodules consist of the fungus, giant epithelioid cells; causes a cheesy degeneration.

Distribution. Found in Europe.

Pathogenic properties. Pathogenic for hogs; occurs in the lungs, where it produces nodular masses.

Oidium lithogenes. (Sanfelice.) Pammel. Blastomycosis

The fungus occurs in the cancerous-like growth of the lymphatics; is frequently surrounded by lime, on agar and gelatin forms white colonies; in stick culture the growth is needle-like; sugar is changed into alcohol and carbon dioxid. Fungus consists of spherical bodies.

Distribution. In Europe.

Pathogenic properties. Pathogenic for guinea pigs, white rats, sheep, and cattle, producing nodular enlargements, frequently surrounded by a calcareous capsule.

- W. W. Hamburger, in a recent number of the Journal of Infectious Diseases, refers to a morphological and biological study of blastomycosis as follows:
- 1. The strains of organisms appear nearly identical, so far as growth in test-tubes goes. A few minor differences are summed up below.
- 2. The organisms grow vigorously on the usual laboratory media, with perhaps a slightly more abundant growth on faintly acid glucose-agar.
- 3. Temperature is perhaps the most important factor in varying the gross and microscopic morphology; room temperature favors production of mycelia and aerial hyphae; incubator temperature inhibits production of hyphae and favors coherent, waxy, yeastlike colonies (hudding forms).
- 4. Those cultures which produce yeastlike growths at incubator temperature develop hyphae within 24 hours when withdrawn and placed at room temperature. Likewise the majority of yeast-like colonies will finally (in 17 to 30 days) show evidence of beginning hypha formation even if kept at 37 degrees C.
- 5. Glucose-agar stabs, and broth form the most serviceable culture media if a limited variety is at hand. Duplicates should always be made to control differences in morphology at room and incubator temperature.
- 1. Four strains of organisms isolated from four cases of generalized blastomycosis appear identical.
- 2. Pronounced variations in the gross and microscopic morphology of the organisms are produced by variations in temperature. As a routine for purposes of study cultures should be grown at both room and incubator temperatures.

Distribution. Found both in Europe and North America.

Pathogenic properties. Dr. Harris gives the following:

Towards the lower animals pathogenic properties vary very much with the culture, recently isolated cultures as a rule proving more virulent than older ones. Mice, guinca pigs, and dogs are most susceptible, succumbing often to subcultaneous and intraperitoneal inoculations, whilst the white rat, rabbit, sheep, and horse are more refractory; in all, the lesions may be localized in the form of abscesses, or general infection may ensue where subcutaneous inoculation is practised.

Dr. E. R. Le Count and J. Myers discuss the case of systemic blastomycosis of a Polish laborer.

The first noticeable departure consisted in a feeling of discomfort involving the chest on the right side extending through from front to back, later cutaneous lesions appeared, located below the left ankle and extended down to the heel. He was obliged to stop work in December. The patient was emaciated, pale, anemic, and weak. Marked oedema was present in the ankles, feet, face and arms. His nails were clubbed; inguinal adenopathy was noted. From the lesions blastomycotic fungus was isolated, the sputum also containing the organism.

Eisendrath and Ormsby described the cultures as follows:

On March 22nd pus was removed from a subcutaneous abscess on the left forearm, which was inoculated on various media. Six days later growth was plainly visible, and after this time the cultures grew rapidly. These proved to be pure cultures of blastomycetes. In the pus they occurred as circular forms and budding forms, having a double contour and the usual refractile capsule. On media the growth varied. It presented a moist, pasty surface on glycerin-agar, with at times a wormy appearance or else presenting large folds and depressions. Microscopically, these cultures showed many oval and circular organisms, some budding ones, and much mycelial formaticn, the latter being both coarse and fine containing sporules. Lateral conidia occurred. On glucose-agar the growth was more dry, white, and presented aerial hyphae; and microscopically there were fewer circular and budding organisms and more fine mycelia. On both glucose and glycerin-agar the media were penetrated to a considerable depth in a semi-circular manner.

Drs. Le Count and J. Myers say, as follows:

The body was examined a few hours after death and the following anatomical diagnosis made: Blastomycotic bronchopneumonia; blastomycosis of the peribronchial lymph nodes, of the pleura, the subpleural, and retropharyngeal tissue, the liver, the kidneys, the colon, the spinal column (dorsal vertebrae), the external spinal dura, the cerebellum, the left elbow, both knee and ankle joints, and of the skin and subcutaneous tissue with ulcerations, fistulae, and scars. Fibrous pleuritis. Passive hyperemia of liver and spleen. Serous atrophy of adipose tissue. Emaciation. Adenoma of thyroid and accessory spleen.

One notable feature of this case is the large conglomerate blastomycotic nodule in the cerebellum. In only one other case of systemic blastomycosis, that of Curtis', is there any record of changes in the nervous system, and the statements in that instance are solely clinical, death being due to meningitis. The reproduction by a process of sporulation demonstrable in the cerebellar lesion is likewise a new feature of the changes encountered in the lesions of this disease. The idea that in the nervous tissue the fungus may have found favorable or different conditions of nutrition, as an explanation for this method of multiplication, is opposed by the facts that the regions in which it was found were very minute, that it was not generally present in the cerebellar process, and the budding was commonly observed in the "abcesses" in the partitions between necrotic regions.

Ilighly interesting is the relationship between this case of blastomycosis and one of coccidioidal disease described by Ophuls. Up to the present two of the chief differences between blastomycosis and coccidioidal granuloma have been the endosporulation observed in the tissues in the latter disease and its tendency to spread by the Impyh channels. Although no widespread extension by the Impyhatics was demonstrated in the case reported here, the extension to the tracheobronchial glands and in peribronchial lymph channels is unmistakable; the endosporulation on the cerebellum in part also resembles the methods of production described for the organism of coccidioidal granuloma. Taken together, these features in this instance of systemic blastomycosis are in accord with the belief expressed by Ophuls of a close relationship of the organisms in the two diseases.

Ricketts, in an interesting monograph on "Oidiomycosis (Blastomycosis) of the Skin and its Fungi," gives the clinical history, cultural characters and histopathology of a large number of cases. The fungi are divided into three groups, (1) Blastomycetoid or yeast-like. (2) Oidium-like. (3) Hyphomycetoid. He says:

There are two histological forms of the disease in the skin, the eosinophilous and the non-eosinophilous, the former being associated with the mould type of the organism. Aside from the infections considered in this communication, certain cases which have been described in the literature from time to time indicate that oidium-like organisms may cause other severe pathological conditions in man.

LICHENS

Symbiotic organisms consisting of higher fungi, chiefly of the class Ascomycetes, or rarely Basidiomycetes; the thallus consisting of algal cells enveloped by the mycelium of the fungus forming a felted mass. The algae are called gonidia and belong to the Cyanophyceae or Chlorophyceae. The reproductive bodies consist of spermogonia, which contain the spermatia. The asci contain the ascospores, and occur in apothecia. In the Basideal lichens, spores are borne on basidia. Lichens are sometimes divided into fruticose, crustaceous, and foliaceous; but a more natural classification arranges the lichens into the Basidio-lichenes and Asco-lichenes, with various families, such as the Roccellaceae, that contains the Litmus, Roccella tinctoria; the Lecanoraceae, containing the Lecanora; and the Cladoniaceae that contains the well known Reindeer Lichen, Cladonia rangiferina.

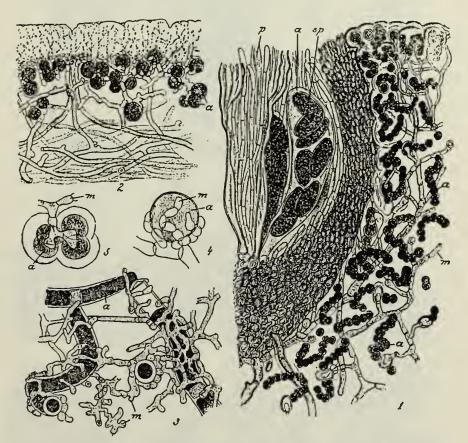


Fig. 115. Lichens, structure of Thallus and Apothecia. 1. Plectospora minutula, section through a part of the apothecium; a (at the right, below) Gonidia, a (above) ascus sp ascospores, p paraphyses; x500. 2. Section of thallus of Cladonia furcata x 330. 3. Portion of thallus of Stereocaulon rumulosum; a gonidia, m hypha. 4. Isolated gonidium (a) with attached hyphae (m) x 950. 5. Synalissa ramulosa, isolated gonidia (a) with attached hypha (m). After Bornet.

EMBRYOPHYTA ZOIDIOGAMA

BRYOPHYTA

Seldom thalloid, generally with stem and leaves with well marked alteration of generations. They contain antheridia and archegonia similar to those of the ferns. The antheridia are stalked, ellipsoidal, spherical, or club-shaped; the sperm cells are biciliated, the archegonia flask-shaped, the ventral portion with a large center cell, the lower portion divided into an egg cell and ventral canal cell. At maturity the new canal cells become mucilaginous and disorganized.

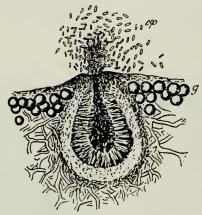


Fig. 114. Spermogonia of Lichen. g. Gonicdia, fungus threads below. sp. Spermatia. Greatly magnified. After Tulacine.

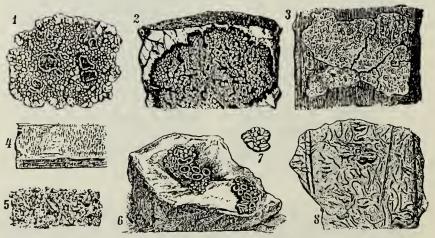


Fig. 115b. Lichens. 1. Ochrolechia tartarea. 2. Rhizocarpon geographicum. 3. Lecanora subfusca, on bark of tree. 4. Calicum. 5. Bacomyccs roseus. 6. Lecanora esculenta. 7. No. 6 removed from substratum. 8. Graphis scripta. 1-8 after Wettstein.

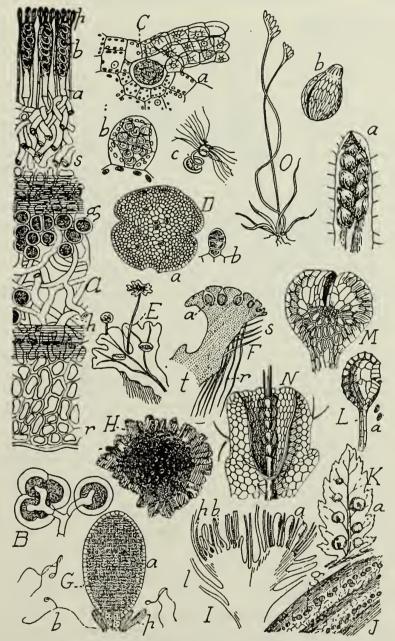


Fig. 116. A. Lichen—Iceland Moss (Cetraris islandica). p. Paraphyses. a. Asci. b. Ascospores. s. Subhymenial layer. g. Gonidia or alga. h. Hyphae. r. "Cortical" portion. B. Lichen. (Synalissa sympharca) sending its hyphae into an alga Gloeocapsa. C. Archegonium of fern (Poiypadium vulgare) with egg cell. b. Antheridium with sperm cells. c. Single coiled sperm cell. D. Liverwort (Marchantia polymarpha). a. Large genmule or bud. b. Same section of thalloid structure bearing scales, s. G. Antheridium with sperm cells shown at b. H. Foliaceous lichen Physicia pulverulenta. I. Section of male plant of moss. Phascum cuspidatum. a. Antheridium. b. Archegonium. l. Icaves. p. Paraphyses. K. Shield Fern (Aspidium Felix Mas), pinnule bearing sori. a. Indusium, undernearth the sporangia. L. Sporangium of the same with ring and stalk with the spores at a. M. Sporangium of the Royal Fern (Osmunda regalis). No ring or mere traces. N. Filmy fern (Trichamanes alatum). O. Schizaea pusilla. a. Fertile pinnule with sporangia. b. Single sporangium with ring at small end.

the mucilaginous material acting as a servant to attract the sperm cells. After fertilization occurs, spores develop. In addition to the sexual method of reproduction an asexual reproduction also occurs. The gametophytic and sporophytic stage are sharply differentiated...

In the development of the sexual generation the spore germinates giving rise to a tube which develops into a new plant, and is called protonema. There are two divisions, the common mosses, and the liverworts. The parts of the fruiting moss plant are as follows: The calyptra or membranous cap which covers the capsule and soon falls off, exposing the operculum, which is a kind of lid, that is also thrown off. The peristome is developed within the operculum and contains teeth, between which the spores are discharged. The elaters in Marchantia are for the dissemination of the spores. The peristome differs in different genera; this affords a convenient means of classification. The spores are found in the capsule and running through the center is a slightly differentiated tissue, the columella. In Funaria the reproductive organs occur on different plants. The sexual organs are borne much like those of liver-worts at the apex of the stem. The antheridia occur in a small rosette of leaves and are club-shaped, the upper part consisting of a single layer of large chlorophyll bearing cells in which small cubical masses occur, the biciliated sperm cells. The archegonia occur in young plants and closely resemble the archegonia of liverworts, except that they have a larger neck. The spores germinate by producing a protonema which early produces a rhizoid.

The liverworts and mosses are much more highly differentiated than any of the Thallophytes, being characterized by more or less differentiation into tissues. Their life history presents a well marked alteration of generations. The gametophyte is more conspicuous than the sporophyte; the germinating spore produces the protonema, which consists of a branched filament, the cells containing the chloroplastids. The protonema is usually short-lived in the Hepaticae but in the true mosses is longer-lived and may persist from year to year. The moss plant is attached to the soil by small unicellular root hairs, or by many curled filaments which, in mosses, are called rhizoids. The shoots of mosses bear lateral organs known as leaves. In Polytrichum and Mnium the leaf consists, essentially, of a single layer of cells except on the midrib. In the leafy-stemmed liverworts like Frullonia two rows of lateral leaves occur. In Marchantia the leaves are rudimentary and occur on the under surface of the thalloid structure in the form of small scales. The small dots on the surface represent the stomata which are dome shaped structures consisting of a number of cells on each side. The stomata communicate with the photosynthetic system of the plant.

Bryophytes are divided into two classes, the liverworts — Hepaticae — and the Mosses — Musci, the latter represented by spagnum moss — Polytrichum, Bryum, etc.

The mosses are distinguished from the thallophytes by their sexual reproduction, the antheridis or male organs are stalked, ellipsoidal, or club-shaped, and enclose small cubical cells, in which the ciliated sperm cells occur. These are ejected, float about in the water till the female reproductive organ, the archegonium, is reached. This is a flask-shaped body containing a neck and an egg cell. At maturity the upper part of the canal cells become mucilaginous, the sperm cells pass down through the canal to the egg cell, where fertilization

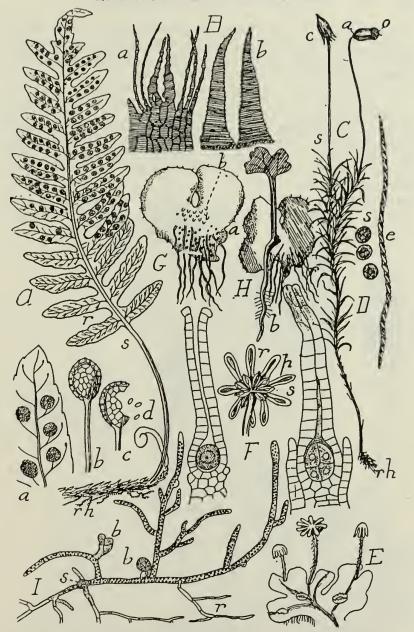


Fig. 116a. A. Common Polypody (Polypodium vulgare). rh. Rhizome. s. Stipe. f. Frond. r. Raeliis. a. Part of frond with sori. c. Sporangium. d. Spores. B. Moss (Mnium hormum). a. Inner peristome. b. Outer peristome, two teeth. C. Juniper Moss (Polytrichum commune). rh. Rhizoid. s. Seta or stalk. c. Calyptra or cap. o. Operculum. D. Common liverwort (Marchantia polymorpha). s. Spores. e. Elater. E. Same as D—thalloid body with female fruiting body and eupules. F. Same more magnified. r. Rays with the spore eases containing the spores and elaters. p. Perigynium, to the left and right archegonia, different stages. G. Prothallus of fern with archegonia, b. and antheridia, a. H. Fern prothallus with young fern and root at r. I. Protonema of Moss (Funaria hygrometrica). r. Rhizoids. b. Buds. s. Spores.

is brought about. After fertilization, the egg cell divides and gives rise to an embryo. The mosses differ from the ferns and their allies in a less differentiation of tissues and a slight development of shoot and root system. The vascular system and leafy shoots and roots are marked in the ferns.

PTERIDOPHYTA

Spores alike or unlike microspores and megaspores developing into flat or irregular prothallia; these bear the reproductive organs, (antheridia and archegonia); flowers and seeds absent; usually a well developed vascular system.

This sub-division includes the class Filicales or ferns proper. The class Equisetales or horsetails; the Lycopodiales represented by the common club moss, (Lycopodium).

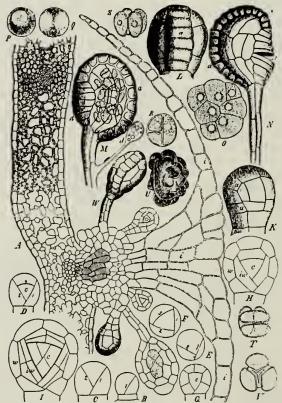


Fig. 116b. Fern. A. Section through frond of Aspidium Filix mas, a leaf-like body with parenchyma cells, an epidermis and vascular bundles x 100. B-J. Development of the sporangia of Aspidium trifoliatum x. 350. K. Young sporangium of Nephrolepis exaluta in the act of differentiating the annulus x 350. L. Immature sporangium of Blechnum occidentale, seen from the back. M. Sporangium with spores. N. Sporangium deliscence at st, a jointed ring x 120. O. Group of spore mother-cells x 350. P.R. Single spore mother-cells in different stages of partition x 425. S.T. Bilateral tetrads of Aspidium Filix mas. U. Mature spores of Aspidium Filix mas x 500. V. Tetrads. After Luerssen.

FILICALES

Leafy plants, fronds usually raised on a stipe; coming from a rootstock; leaves usually rolled up in the bud, circinate; spores all of one kind and size, produced in sporangia which occur on the back of the frond, these at maturity break open and discharge the minute spores, which develop prothalli that bear the antheridia and archegonia. The following sub-orders occur in the United States: the Ophioglossaceae represented by the common adder's tongue, Ophioglossum vulgatum, found in moist meadows, the Moonwort, Botrychium Lunaria and B. Virginianum; the Osmundaceae, large ferns with straight erect rootstocks, pinnate leaves; large globose sporangia with mere traces of a ring; the Royal fern, Osmunda regalis, Clayton's fern (O. Claytonia), the most common species in damp woods, and the Cinnamon Fern (O. cinnamomea), occurring in wet places, marshes, etc.; the Filmy ferns, Hymenophyllaceae, represented by the Bristle Fern (Trichomanes radicans); Cyatheaceae with such tree ferns as Dicksonia; Polypodiaceae, Common Brake, Maidenhair fern;

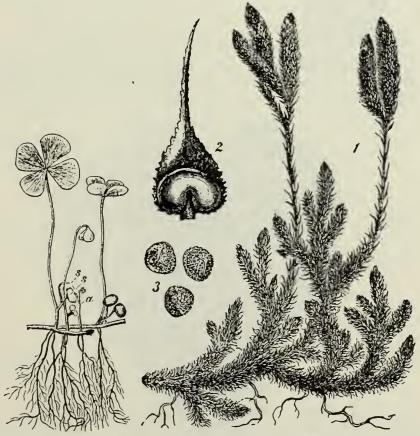


Fig. 117. Marsilia (M. quadrifolia), a young leaf. s. Fruiting body. Bischoff.

Fig. 117a. Club moss (Lycopodium clavatum). 1. Plant fertile shoots. 2. Scale and the sporangium. 3. Spores. After Wossidlo.



Antheridium of fern with sperm cells. After Luerssen.

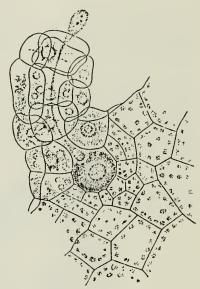


Fig. 119. Pteris serrulata. A single archegonium; canal and neck cells; mucilage protruding x 350. After Strasburger.

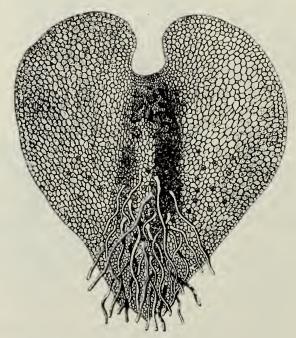


Fig. 120. Prothallus of Fern. Archegonia above, antheridia below among the hairs. After Luerssen.

Gleicheniaceae, some tropical ferns of few species; the Schizeaceae represented by the Small Curly Grass (Schizaea pusilla), Climbing Fern, (Lygodium palmatum), sporangia ovoid or sessile provided with an apical ring, a family containing about 100 species; the Polypodiaceae the largest sub-order with 200 genera and 3000 species; Marsiliaceae containing Marsilia a common aquatic or semiaquatic plant; represented by Salvinia, Azolla, also aquatic. The order Marattiales, contains the Marattia, tropical.

It is not at all strange that the ferns should be poisonous since Greshoff and others have reported the presence of hydrocyanic acid in these plants. Greshoff says the odor of oil of bitter almonds is especially intense in the young leaves of Cystopteris fragilis Bernh; and there is also a trace of HCN in the spores. He also calls attention to the presence of the same substance in the common brake (Pteris aquilina), and states that several tropical ferns namely Davallia braziliensis, and other species are cyanogenetic, and that one fern, the D. pentaphylla, forms a large amount of this substance, especially the cultivated form elegantissima. Several species of the Gleichenia contain saponin.

POLYPODIACEAE

Perennial with horizontal erect, short or elongated rootstocks; leaves various, entire, pinnate, pinnatifid, or decompound, vernation circinate (coiled); sori on the margins of the leaf or on the lower side, generally without an indusium (covering); sporangia with a vertical many celled incomplete ring, which on straightening out ruptures and discharges the spores. A few of the ferns, as Male Shield fern Aspidium Filix-mas, and the A. marginale, are used in medicine.

KEY FOR THE GENERA

Indusium absent

| industum absent |
|--|
| Indusium present, evident. |
| Sori marginal. |
| Indusium with margin of frond rolled over. |
| Sporangia borne on a continuous marginal vein-like receptacle. |
| Stipe light coloredPteris |
| Sporangia on the ends of the veins. Stipe blackAdiantum |
| Sori on back with special indusium covering the same. |
| Sori linear or oblongAsplenium |
| Sori roundish on the back or rarely the apex of the vein. |
| Stipe not articulated. |
| Indusium flat or slightly convex or round reniform, fixed by |
| the center, opening all round the marginAspidium |
| Indusium convex, fixed by a broad base, commonly reflexed |
| as the sporangia ripen |
| Indusium obscure, leaves closely rolled together with necklace-like seg- |
| ments Onoclea |
| |

Polypodium. L. Polypody

Simple or pinnate fronds from horizontal rootstocks; stipes articulated to the rootstocks; sori (fruit dots) round, naked on the back of the frond in one or more rows each side of the midrib or scattered; indusium wanting. About 350 species, mostly tropical. The species in the Northern United States

are P. vulgare, and P. incanum, the P. vulgare being more common northward. The P. aureum found in Florida is a large fern.

Polypodium vulgare. L. Common Polypody

Creeping rootstocks covered with cinnamon-colored scales; stipes light colored; fronds 4-10 inches high, simple and deeply pinnatifid, the divisions linear oblong, obtuse or somewhat acute obscurely toothed; sori large.

Distribution. Throughout North America, also Europe and Asia.

Poisonous properties. Used in catarrh and asthma. Supposed by some writers to be poisonous.

Adiantum L.

Sori marginal, borne on the under side of a transversely oblong, crescent-shaped or roundish, margin of the frond; the sporangia attached to the tips of the forking branched veins; stipe black, polished; leaves divided. About 80 species of wide distribution. The A. Capillus-Veneris in tropical and subtropical regions.

Adiantum pedatum. L. Maidenhair Fern

Root-stock slender, chaffy; stipe black, shining, dichotomously forked at the summit; pinnae arising from the upper sides of two branches of the stipe; pinnules short-stalked, numerous.



Fig. 121. Cultivated. Maiden hair fern (Adiantum).

Distribution. In moist woods from Nova Scotia to British Columbia and Alaska, California, Utah, Arkansas and Georgia, also found in Asia.

Medicinal properties. The Maidenhair Fern has a bitterish aromatic taste and was formerly much used as a demulcent; it is probably poisonous. The European A. Capillus-Veneris was used in catarrhal affections.

Pteris

Fronds once to twice pinnate, coming from a stout root-stock, usually large plants; sporangia in a continuous slender line occupying the entire margin of the fern frond and covered by the narrow edge which forms a continuous membranaceous indusium. Pteris and Pteridium are usually separated; about 100 species in the genus *Pteris*.

Pteris aquilina, L. Common Brake

Frond dull green, from 2-3 feet high, ternate at the summit of an erect stout stalk; variable in height from 1-6 feet.; stipe coming from a black root-stock; the spreading branches twice pinnate, branches oblong-lanceolate.



Fig. 121a. Maiden fern (Adiantum pedatum). (Am. Agriculturist).

Fig. 121b. Brake (Pteris aquilina). The roots and stems of this plant are poisonous. The rootstocks contain a starch which is sometimes used as food. (Ada Hayden).

Distribution. Widely distributed in North America, from the Atlantic to the Pacific coast, also found in Europe. Probably the most common fern in the north, especially on the Pacific coast.

Poisonous properties. This plant has been reported as being an anthelmintic and also an astringent; it is suspected of being poisonous.

Asplenium, L. Spleenwort

Large or small ferns with entire, lobed, pinnate, leaves, 2-3 times pinnate or pinnatifid, veins free; sori oblong or linear, oblique, straight or rarely curved; indusium straight or curved. About 200 species of wide distribution, several species cosmopolitan like A. Trichomanes.

Asplenium Filix-foemina. (L.) Bernh. Common Spleenwort

Fronds 1-3 feet high, ovate-oblong or broadly lanceolate, twice pinnate; pinnules confluent on the secondary rachis, oblong and doubly serrate or pinnately incised; sori short.

Distribution. Common in the north and especially in the dense woods, as far south as Missouri.

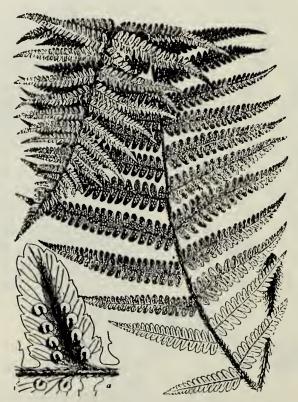


Fig. 122. Aspidium Filix mas. Spore bearing leaf 1/6 natural size. a. A single segment showing the under side x 10. After Luerssen.

Poisonous properties. The rhizome of the root is used in medicine although it is not officinal. It is supposel to possess properties the same as the Male Shield Fern.

Aspidium. Swartz. Wood Fern

Fronds with 1-3 pinnate leaves, free veins; sori borne on the back or rarely at the apex of the veins; indusium covering the sporangia, flat or flattish, scarious, orbicular and peltate at the center, or round-kidney-shaped, opening all round the margin. About 200 species of wide distribution, common in the northern states.

Aspidium fragrans. (L.) Swartz

Fronds 4-12 inches high, glandular and aromatic, fragrant; rootstock stout, nearly erect, densely chaffy, as are the crowded stipes and rachis. Species found in Asia and Europe.

Aspidium Filix-mas. (L.) Swartz

Fronds large, 1-3 feet high; pinnae linear-lanceolate, tapering from base to the apex; pinnules very obtuse, serrate at the apex and obscurely so at the sides, the basal incisely lobed; sporangia nearer the midvein than the margin and usually confined to the lower half of each fertile pinnule.

Distribution. Native to Europe and found in rocky woods from Labrador to Alaska, northern Michigan, British Columbia, Greenland, Europe, Asia, and the Andes of South America.

Poisonous properties. It is suspected of being poisonous. The rootstocks have long been used in medicine as a vermifuge and contain the following substances: a fatty, green oil, traces of a volatile oil, resin, tannin, filicic acid, $C_{35}H_{42}O_{13}$, filicin, aspidin, $C_{23}H_{27}O_7$, a fixed poisonous oil.

Dr. Winslow says:

Large quantities of the drug cause hemorrhagic gastro-enteritis, tremors, weakness, stupor, coma, acute nephritis and cystitis. Six drachms of the oleoresin have proved fatal in man and sheep; five drachms in a medium-sized dog; and three ounces in a cow. Aspidium should never be given with oil which aids its absorption.

Aspidium marginale. (L.) Swartz

Much like the preceding, with evergreen fronds, small, thickish, ovate or oblong in outline, and from 1-3 feet high; pinnae lanceolate, acuminate; pinnules oblong or oblong-scythe-shaped, obtuse or pointed, entire or crenatelytoothed; sori close to the margin.

Distribution. From Canada to Minnesota, Iowa, Missouri, Arkansas, and Georgia.

Poisonous properties. Probably has the same effect as the preceding species. Dr. Johnson says:

Oleo-resin of male-fern is one of the best known remedies for tapeworm, and also one of the most efficient. Doubtless much of the disappointment experienced with it is attributable to inefficient preparations. Since, however, it has been demonstrated that A. marginale is quite as efficient, and as this species is very abundant, there is now no good reason why reliable preparations should not be the rule rather than, as heretofore, the exception.

Cystopteris. Bernhardi. Bladder Fern

Fronds growing in tufts, 2-3 times pinnate, the lobes cut toothed; stipe slender, 2-4 pinnate leaves; sori roundish, borne on the back of the veins.



Fig. 123. Shield Fern (Aspidium marignale). Said to contain filicic acid. Known to be poisonous. (Ada Hayden).

Indusium delicate arched or level-like, attached by a broad base on the inner, partly under the sorus, opening free at the other side; veins free. A small genus of 5 species, 2 common in the United States. C. bulbifera, long slender fronds bearing bulblets which propagate the plant. C. fragilis, with brittle stalk, the pinnae and pinnules ovate, lanceolate, irregularly pinnatifid or cuttoothed.

Onoclea. L. Sensitive Fern

Coarse ferns, creeping root stocks, fertile fronds erect, rigid with contracted pod-like or berry-like divisions, rolled up; sori roundish, imperfectly covered by a very delicate hood-shaped indusium attached to the base of the receptacle; when dry opening, allowing the spores to escape; sterile fronds foliaceous. A small genus of a few species.

Onoclea sensibilis. L. Sensitive Fern

Slender root stock with scattered fronds, sterile long stalked 2-15 inches high, triangular ovate, fertile fronds, contracted closely, bipinnate, pinnules rolled up into berry-like bodies.



Fig.- 124. Flowering Fern (Osmunda Claytoniana). Reported as poisonous. (Ada Hayden).

Distribution. Moist meadows and thickets from Newfoundland to Florida and Minnesota.

Poisonous properties. Very abundant in hay from low meadows. May be injurious.

Onoclea Struthiopteris. (L.) Hoffman. Ostrich Fern

Fronds growing in a crown; root stocks stoloniferous; sterile, short stalked, 2-10 feet high, broadly lanceolate; pinnae pinnatifid, veins free, the veinlets simple; fertile frond shorter, pinnate with pod-like or somewhat necklace-shaped pinnae.

Distribution. Nova Scotia to Illinois, Wisconsin, Iowa and British Columbia, common also in Europe and Asia.

OSMUNDACEAE

Large ferns, root stocks frequently stout and erect; leaves 1-2 pinnate, coiled in vernation; veins free, mostly forked, running to the margins of the pinnules or lobes; sporangia large, globose with mere traces of a ring, or none, borne on contracted pinnae, on the lower surface of the pinnules. Three genera.

Osmunda. L. Flowering Fern

Tall ferns growing in swamps or low ground. Fertile fronds much contracted and bearing on the margins the short pedicelled, naked sporangia on the margins of their radii like divisions without chlorphyll; sporangia thin, reticulated, opening by a longitudinal cleft into two halves, with a few thickened cells the rudiment of the ring.

There are six species in the North temperate regions, three species common in the North, the Cinnamon Fern (O. cinnamomea), clothed with rusty wool; the Royal Fern (O. regalis), a smooth pale green fern, 2-5 feet high, with 13-25 sterile pinnules; Clayton's Fern (O. Claytoniana), clothed with loose wool, but soon smooth; pinnae oblong lanceolate; some of the middle pinnae fertile. The O. regalis is used as a tonic and styptic. By some these ferns are regarded as injurious to stock.

EQUISETALES. Horsetails

Rush-like perennial plants, epidermis impregnated with silica; creeping rootstocks, stem generally hollow jointed, simple or branched, striated or grooved, provided with a double series of cavities and usually a large central one, branches verticillate; leaves reduced to a sheath which is divided into teeth corresponding to the principal ridges of the stem; stomata in furrows;

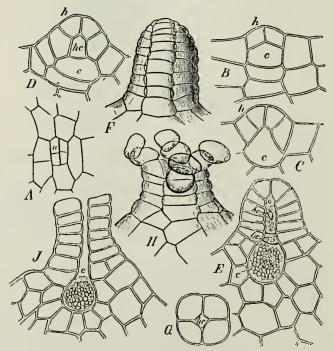


Fig. 125. Formation of Archegonia of Osmunda. A. Early development seen from the surface. B. The same in vertical section. C-E. Farther development. F. Opened and closed neck. G. Neck in oblique section. J. Fertilization of the mature archegonium. a. archegonium, h. neck of the same, c. central cell, e. egg, bc. spermatozoids, hc. neck canal cells. B-J x 240. A. greatly magnified. After Luerssen.

sporangia 1-celled clustered underneath the shield-shaped scale of the cone; spores all alike, two thread-like elastic filaments (elaters) are attached to the base of the spore which roll around it when moist and spreading when ripe; prothallus green formed upon damp ground, usually dioecious. One order, Equisetaceae, and one genus, consisting of 40 species. Fossil horsetails numerous.

EQUISETACEAE

Equisetum. L. Horsetail

Perennial jointed plants with creeping root-stocks, dull and blackish in color, often bearing tubers, roots in whorls from the nodes, stems usually erect, simple or branched, jointed cylindrical, the surface striated, the stomata occur in grooves either in rows or in bands, the nodes bearing a whorl of reduced leaves joined by their cdges into cylindrical sheaths, the tips consist of presistent or deciduous teeth; branches when present in the form of whorls from the nodes; fruit consisting of a terminal cone containing the sporangia in which occur the greenish spores; cach spore provided with four hygroscopic bands, the elaters; spores produce two kind of prothalli, one male the other female; the male containing the antheridia, the female the archegonia. A small genus commonly called rushes or horsetails. Some ten species in castern North America.

Equisetum arvense. L. Common Horsetail

Perennial with annual stems, stomata scattered; fertile stems unbranched, destitute of chlorophyll, 4-10 inches high, soon perishing; sheaths distant, 8-12 toothed; the sterile slender 1-2 feet high, 10-14 furrowed producing simple or sparingly branched, 4-angular teeth, herbaceous, triangular lanceolate.

Distribution. Abundant in sandy fields along roadsides and railroads, especially northward from Newfoundland to Virginia, California and Alaska. Also occurs in Europe and Asia.

Equisctum hyemale. L. Scouring Rush

Stems all alike, slender, rather stiff, evergreen, from 1½-4 feet high, 8-34 grooved. Stem rarely producing branches which are usually short and sometimes fertile; stomata arranged in rows, rough ridges with 2 indistinct lines of tubercles, the central cavity large, sheath rather long, cylindrical, marked with a black girdle, their ridge obscurely carinate; spikes persistent.

Equisetum hyemale. L. var. robustum, (A. Br.) A. A. Eaton

Stem perennial, tall and stout, 8-10 fect high, sometimes an inch thick, occasionally branched; 20-48-grooved, the ridges roughened with lines of transversely-oblong tubercles; sheaths rather short with a thick girdle at the base and a black limb; ridges of sheaths carinate.

Distribution. In wet places, from Ohio, Iowa, to Louisiana, Mexico, California, and British Columbia, also in Asia.

Poisonous properties. The rushes have long been recognized in Europe as being injurious to horses, and there are records of their poisonous properties in American Agricultural Literature.

A writer in the American Agriculturist, many years ago, described accurately a disease which might be called equisetosis, and which was produced by poisoning from these rushes.

Mr. H. Lawrence of Spencer, Iowa, recently sent me a specimen of the last species mentioned above, writing that:

"The cattle staggered and had the scours. One man lost 10 head of young animals."

Friedberger and Fröhner state that the symptoms of poisoning by this rush are as follows:

At first, excitement and anxiety; the sensorium remaining unaffected; later, uncertainty of movement, reeling and staggering; at least, paralysis of hinder limbs, tumbling down, general paralysis, insensibility to external irritants, unconsciousness and coma. Pulse accelerated, appetite at first normal, but in course of time great disturbance of nutrition; sugar in the urine. Course sometimes very acute, death occurring in a few hours, but sometimes protracted (two to eight days), and at times chronic (one to several weeks). In cattle, after excessive eating continuous diarrhoea becomes a prominent characteristic along with the paralytic symptoms; while, if the food be persisted with, cachexia and hydraemlea, combined with weakness bordering on paralysis, make their appearance, Autopsy reveals: hyperaemia, oedema, dropsical effusions on the brain and spinal cord, especially on cerebellum; in cases of longer duration, hydracmia. Sometimes inflammatory changes in the mucous membrane of stomach and bowels. Therapeutics: change of fodder, purgatives and stimulants, especially camphor; blisters along the spine.

Mr. P. J. O'Gara says of this plant:

It has been found growing along roadsides and railroad tracks, but its occurrence in serious amounts is apparently confined to low moist meadows which are more or less sandy. As before indicated, it is confined mostly to the Missouri bottom. A thorough examination of several meadows in this region has shown that this plant often constitutes one-sixth or more of the bulk of the hay. There is no doubt that this plant causes a great deal of trouble, but to what extent is not known as many of the haystacks which were carefully examined contained the Rattle-box in considerable amount. Horses eating this hay suffered the combined effects of both poisonous plants.

Prof. Jones and Dr. Rich state:

The first evidence of the trouble is more or less unthriftiness, the horse appearing thin and the muscles wasted. In from two to five weeks, according to the age of the horse and the manner of feeding, the animal begins to lose control of its muscles, sways and staggers like a drunken man, although its eye looks bright, it eats well, and may even try to caper and play. After muscular symptoms become pronounced many cases refuse to lie down, standing until thrown down by disorderly muscular contractions. If it continues to cat the plant the horse in any case soon loses power to stand and goes down, after which it becomes very nervous and struggles violently to get up, the legs become more or less rigid, and at times all the muscles of the body seem convulsed. Even in this condition one well nursed patient lived two weeks. The horses are generally willing to cat, although unable to rise, but become sore and tired from struggling, finally dying from exhaustion. Life is much prolonged by turning from side to side three or four times in twenty-four hours; thus preventing gravitation congestion of the lungs and kidneys. The pulse becomes slow until toward the end when it is rapid and weak. Temperature is below normal until the animal goes down, after which some fever develops in consequence of the nervous excitement and violent struggling. The extremities are usually cold, and in the winter horses suffering from Equisetum poisoning suffer severely from the cold, presumedly because of diminished oxidation and consequent low body temperature. The visible lining membranes of mouth, nose, eye, etc., become pale.

Conditions Influencing the Effect of the Poison.

Age of the horses.—Young animals develop symptoms much more quickly and succumb to the Equisetum poisoning sooner than older ones. In one case under observation a mare eating Equisetum hay did not show symptoms until after four weeks, while her colt by her side developed typical symptoms of horsetail poisoning and died in ten days. Nine out of fourteen horses on one farm, all fed alike upon good hay, were bedded with swale hay containing large quantities of this weed. They ate this bedding freely and within three weeks all nine showed symptoms of poisoning, the remaining five bedded with straw kept perfectly well. The youngest, a three-year-old, was down and died a few days later. The oldest, an old brood mare, showed but slight symptoms, while the other seven, of internediate ages, all staggered and reeled, although they recovered.

Feed.—Grain-fed horses resist the action of the poison much longer than those not grained. Horses seem to develop a depraved appetite for the weed. In the last mentioned

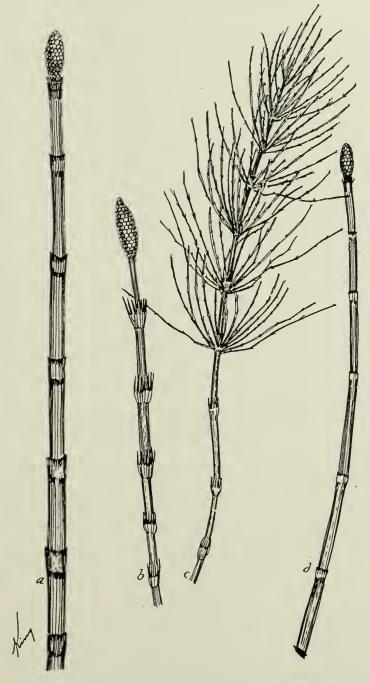


Fig. 125a. a. Scouring Rush (Equisetum hyemale var robustum); b. Horsetail (E. arvense), futile branch; c. Sterile branch of E. arvense; d. E. hyemale. Said to be poisonous to horses. (C. M. King).



case, though all were fed good, clean timothy hay, they seemed to prefer the horsetail bedding, and even left their grain to eat it.

Condition of the plant.—We have no evidence that horses grazing upon the green plant are poisoned thereby. It may be that the plant is less poisonous in the early stages of its growth than when mature, or the laxative effect of the grass eaten with it may prevent the cumulative action of the poison. Moveover the plant rarely occurs in as large quantities in pastures as in meadows and apparently rarely need cause apprehension. If, however, it is abundant, close watch should be kept upon horses pasturing where it occurs that the animals may be removed at the earliest symptoms of trouble.

Treatment.—In the way of treatment, the first and most important thing is to stop inmediately the feeding of the Equisetum hay. Our practice further than this has been to give a purgative pill consisting of one ounce of Barbadoes aloes, one or two drachms of ginger, and sufficient English crown soap—soft soap—to make a ball or pill. This is put down the horse's throat, at one dose, and following this we have usually given bran mashes night and morning until the digestive tract is entirely cleared of the poisonous plant. In case aloes cannot be easily obtained a quart of raw linsced oil will be very well. After the physic has operated, a teaspoonful of powdered nux vomica is added to each grain feed, three times a day. This tends to relieve the muscular incoordination. When poison symptoms are severe and especially when staggering is very profound, slings should be used to support the animal for when once down it is very difficult to make it stand again even with the aid of slings. If, however, the above treatment is begun before the horse loses the power to stand and it can be kept on its feet, its life can be saved in practically all cases.

Stehler & Schröter in their work on the weeds of meadows mention this same plant and several other species as being injurious to stock, not only to horses, about which there is a difference of opinions, but to cattle. In the latter it produces diarrhoea. Cows become poor and the milk flow ceases or is checked.

That this disease is similar to one produced by mouldy corn is shown by the following quotation from Dr. Peters:

It is also known that certain weeds commonly called horsetail have a faculty of producing a disease almost identical with this one. The experiments conducted by Dr. Rich of the Vermont Station show that that weed is capable of producing similar symptoms.

EMBRYOPHYTA, (SIPHONOGAMA, OR SPERMATOPHYTA)

Plants producing seeds which contain an embryo with 1 or more cotyledons, a stem caulicle, a radicle, and a plumule, these parts, occasionally not differentiated before germination; microspores, equivalent to pollen grains borne in microsporangia; ovules (macrosporangia) borne on a modified leaf called the carpel, containing 1 macrospore, equivalent to the embryo sac which develops the minute female prothallium, an archegonium; the egg cell in the embryo sac is fertilized by means of a sperm cell in the pollen tube; the male prothallium generally but slightly developed. The Spermatophyta contain two main divisions based upon the character of the ovules.

GYMNOSPERMAE

Ovules naked, not enclosed in an ovary, attached to scales or wanting; pollen grains develop into the pollen tube; the male prothallium contains the sperm cell and fertilizes the egg cell in the ovule. The Gymnosperms are divided into six classes.

1. Cycadales. These include the Cycas circinnalis well known in cultivation, an important plant of the tropics. The C. media of Australia produces rickets, a Macrozamia causes the same disease. Dr. Stafford states that C.

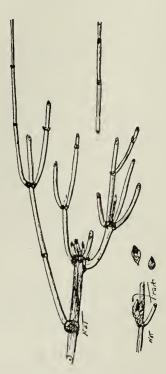


Fig. 126. (Ephedra nevadense). A shrub in Southwestern United tSates.

circinnalis, known as the "Fadong" in the island of Guam, is poisonous, but the poisonous properties of the seeds are removed by soaking and repeatedly changing the water. He says also that the seeds when fresh are so poisonous that the water in which they are steeped is fatal to chickens. The group also includes the Dioon the seeds of which furnish a starch which is an article of food. The species of Zamia, a member of this group, are native to tropical Florida.

- 2. Bennettiales, A fossil group.
- 3. Cordaitales, A fossil group.
- 4. Gingkoales. These include the Gingkoaceae, of which the Ginkgo biloba is well known and is frequently cultivated as an ornamental plant in the United States. Long avenues of these trees are planted in Washington. The fruit of the Gingko has a very disagreeable odor. The tree was common in the tertiary age.
 - 5. The Coniferae.
- 6. Gnetales. This group is represented in the United States by Ephedra, shrubs with horse-tail like branches, small leaves and buckwheat-like seeds.

The Welwitschia of the above group is found upon stony ground in the tropical Old World.



Fig. 127. Hemlock (Tsuga canadensis). A common forest tree of Northern North America. Contains resin and the usual principles found in these resins. Said to be injurious.

CONIFERAE

Resinous trees or shrubs generally evergreen leaves, entire or scale-like; wood consists mostly of tracheids marked with large depressed disks; tracheae only present near the pith and in the leaves; perianth none; flowers monoecious; stamens several, together, subtended by a scale; anthers 2-7 celled; pollen grains frequently of three cells, one fertile and two inflated; ovules with two coats, borne solitary or together on the surface of a scale, straight or partly inverted; fruit a cone, usually papery, but in some instances fleshy, sometimes berry-like; seeds winged or wingless; endosperm abundant, fleshy or starehy; embryo straight and slender; cotyledons 2 or more. About 25 genera and between 275 and 300 species. They include the Podocarpus of the tropical regions, the Taxus or Yew, the Norfolk Pine (Araucaria excelsa and A. brasiliana), frequently cultivated, the White Cedar (Cupressus Lawsoniana) of California, the Cupressus nootkatensis of the northwest coast, the White Cedar (C. thyoides) which occurs in swamps, in the East; the genus Picea consisting of the spruces, Norway Spruce (P. Engelmannii), Tideland Spruce (P. sitchensis), one of the largest trees in Oregon and Washington; the Douglas Fir (Pseudotsuga Douglasii), one of the most valuable of the North American conifers, found in Washington, Oregon and California and in the Rocky Mountains; the Hemlock (Tsuga canadensis), abundant in North America, and the source of Canada or Hemlock Pitch, the bark containing an abundance of tannin; the leaves are said to be abortive; the Tsuga heterophylla of the Pacific coast which produces a valuable lumber; the Balsam Fir (Abies balsamea), which furnishes a kind of balsam that contains four acid resins and a volatile oil; the Black Fir (Abies concolor), a large forest tree of the Pacific coast and the Rocky Mountains; the Sandarac tree (Callistris quadrivalvis), which furnishes not only the sandarac gum used in making varnish, but also a dark-colored, fragrant wood capable of high polish and used in ornamental work; the Arbor Vitae, or White Cedar (Thuja occidentalis), which contains fenshoe, thujonc, thujin, and the bitter glucoside pinicrin, its leaves being irritating to the skin, sometimes producing blisters; the Norway spruce (Picea excelsa), which contains resins and volatile oils and is the source of Burgundy pitch; the Black Spruce (P. mariana), from the young branches of which an essence is prepared that is used in the preparation of spruce beer; the White Spruce (P. canadensis), which, with the preceding species furnishes much of the wood pulp used in the manufacture of paper; and the Pines.

Taxus. (Tourn.) L. Yew

Flowers generally dioecious or occasionally monoecious, axillary from scaly buds; sessile or nearly sessile, from small staminate catkins of a few scaly bracts; 5-8 stamens; anthers 4-celled; fertile flowers solitary, erect, subtended by a fleshy cup-shaped disk; fruit consisting of a fleshy disk which becomes cup-shaped and red and encloses the bony seed.

Distribution. About 6 species native of the north temperate regions. One upon the Pacific coast, *Taxus brevifolia*, is a tree. The European Yew (*Taxus baccata*), a well known poisonous plant, is frequently used for ornamental purposes in this country.

Taxus canadensis. Willd. American Yew

A low shrub, straggling over bushes, with linear leaves, green on both sides. Distribution. In the woods from Newfoundland to New Jersey and Virginia, west to Wisconsin, Iowa and Minnesota.

Poisonous properties. One species is known to contain the alkaloid taxin, $C_{87}H_{52}NO_{10}$. Dr. Johnson says, concerning the poisonous properties of the American Yew:

This plant, a variety, only, of the European yew, cannot be said to have, as yet, a place among medicines. It is believed, however, to possess poisonous properties, and is perhaps worthy of investigation. Regarding the poisonous properties of the berries, the author can state that he has eaten them without deleterious effect, but whether because the quantity was insufficient or not, is an open question. Cases of fatal poisoning from eating the berries of the European yew are on record, and therefore our variety is certainly open to suspicion.

Chesnut refers to the poisonous nature of the yew as follows:

The common yew, or ground yew of the northeastern United States is called poison hemlock in some places. The leaves of this shrub are probably poisonous to stock, as are those of the European yew. This species is more accessible to stock than are those of the western yew (Taxus brevifolia), which grows only in deep canyons.

Dr. Otto Lehmann* in his treatise on poisonous plants, states that older naturalists regarded the yew as one of the most powerful of poisonous plants. Modern testimony is conflicting, but he regards the branches and leaves as poisonous for animals. Friedberger and Fröhner give the symptoms of poisoning from yew as follows: "Death may be sudden, resembling apoplexy; it may be preceded by staggering and convulsions; cases of long standing show gastro-enteritis. Give purgatives as remedies."

^{*} Giftpflanzen. 121. Hamburg. 1882.

Pinus. (Tourn.) L.

Evergreen trees with short scale-like leaves and longer leaves in bundles; the ordinary foliage leaves linear, in bundles varying from 2-5, rarely 1; stamens in catkins, borne at bases of shoots; filaments short; anthers longitudinally dehiscent; pistillate, bearing aments, solitary or clustered on the twigs of the preceding season consisting of numerous imbricated bracts, each with an ovule-bearing scale; fruit a large cone; seeds 2 at the base of each scale; winged above. About 75 species of wide distribution. Of these the more important are: White Pine (P. Strobus), one of the most valuable of North American Pines; Sugar Pine (P. Lambertiana) of California and Oregon; Austrian Pine (P. Laricio), furnishing Austrian turpentine; the Long-leaved Pine (P. palustris) of the South, the most important source of turpentine, which

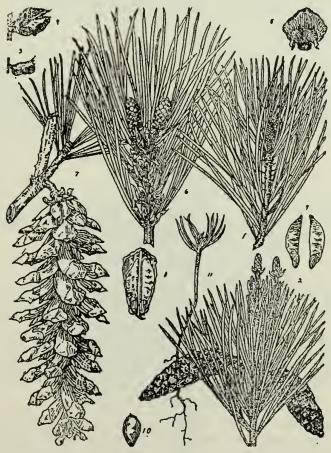


Fig. 128. White Pine (Pinus Strobus). 1. Branch bearing staminate flowers. 2. Branch bearing pistillate flowers and young cones. 3. Anther, enlarged. 4,5. Scales of pistillate flower, enlarged. 6. Autumn branch bearing young cones. 7. Fruiting branch with young cone. 8. Scale of cone with seeds attached. 9. Seeds with wings attached. 10. Seeds, enlarged. 11. Seedling plant. 1, 2, 6, 7, 8, 9 one-half natural size. (S. B. Green).

in this tree amounts to 70-80 per cent., resin, 15-30 per cent, volatile oil and some pinene $C_{10}H_{10}$, a very important constituent; $P.\ sylvestris$, the source of Russian turpentine; $P.\ pinaster$, supplying the French turpentine; $P.\ heterophylla$ and $P.\ echinata$, also turpentine trees; Loblolly Pine $(P.\ Taeda)$ also containing pinene; the Western Yellow Pine $(P.\ ponderosa)$ occurring from mountains of Colorado westward, a large tree 120 or more feet high, 4 or 5 feet in diameter, branching widely, spreading or drooping, bark light red, leaves in 3's or rarely 2's, cones stout, dense, heavy, ovoid-conical, each scale with short recurved prickle; the variety scopulorum of the last named species, found in the front Rockies, a smaller tree with shorter leaves; and Lodge Pole Pine (Pinus contorta), a tall straight tree, 80-120 feet high, and from 12 inches to 3 feet in diameter, with conical head, thin, light grayish-brown bark, leaves 1-3 inches long, light green, rigid, often persistent cones. This last named species occurs from Colorado, Wyoming and South Dakota to the Pacific coast and is closely related to the Jack Pine $(P.\ Banksiana)$.

Phenol and creosote oil are obtained from Pinus palustris and P. Taeda. Poisonous properties. Cattle and sheep do not usually graze upon the leaves of conifers, but when forced to do so because of scarcity of fodder, sheep will eat the leaves, which may produce injurious symptoms. According to Friedberger and Fröhner, plants containing turpentine are poisonous. Chobert, in 1787, observed gastro-enteritis complicated by nephritis as a result of grazing on leaves of conifers. The first named authors find symptoms of haematuria, constipation, evacuation dry, and irritation of the kidneys.

Juniperus, (Tourn.) L. Juniper

Flowers dioecious or monoecious, in lateral catkins, staminate catkins small, fertile catkins consisting of 3-6 fleshy scales; fruit appearing like a berry; color of fruit bluish-black or blackish, frequently with white bloom; seeds 1-3, wingless and bony. The J. Oxycedrus of the Mediterranean region produces "oil of cedar."

Juniperus communis. L. Common Juniper

A shrub or small tree with spreading or pendulous branches; leaves rigid, spreading; berries dark blue. The variety alpina, Gaud. is a low, decumbent, or prostrate shrub with shorter, less-spreading leaves. It contains pinene and cadinene. The oil and fruits are used in the manufacture of gin.

Distribution. From Nova Scotia to British Columbia, to Pennsylvania, Wisconsin, Michigan, Nebraska, and in the Rocky Mountains to New Mexico; occurs also in Europe and Asia. The variety alpina is common in the mountains of New Mexico northward, not, however, as widely distributed as the species.

Juniperus virginiana. L. Red Cedar

A shrub or tree extremely valuable, frequently from 60-90 feet high; pyramidal in form; leaves scale-like, obtuse or acutish, dimorphic, the leaves of young plants being more or less flattened, spiny, and awl-shaped, while those of the stem are scale-like and appressed; catkins terminal; berries on straight peduncles; cones light blue or glaucous. The Platte Cedar (*J. scopulorum*) differs from the other in the development of the seeds.

Distribution. The Red Cedar occurs from New Brunswick to British

Columbia, south to Florida, Texas, New Mexico and Arizona, also the West Indies. The Platte Cedar occurs from Nebraska westward and is common in the foothills of the Rockies.

Poisonous properties. According to Dr. Halsted it poisons goats which orowse on it.



Fig. 129. Red Cedar (Juniperus virginiana). To the left a branch from an old tree; to the right juvenile shoots, spiny. The plant is poisonous and injurious.

Juniperus occidentalis. Hooker

A shrub or small tree, with bark in shreds; leaves pale in color, closely appressed, obtuse or acutish; berries 4-5 lines in diameter.

Distribution. Northwest along Pacific Coast.

The variety monosperma, Eng., shows stunted trees, frequently 2 or more feet in diameter, attaining an age of 1200-1300 years; leaves scale-like; berries

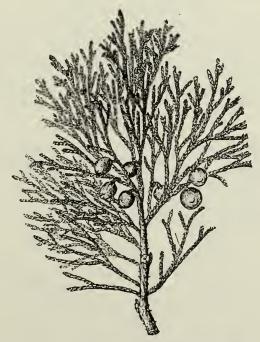


Fig. 130. Savin, Juniper (Juniperus Sabina). Fruiting branch. Known to be poisonous. (After Strasburger, Noll, Schenek and Schimper).

smaller than the above, frequently copper-colored; generally with one seed or sometimes more.

Distribution. Found from Colorado to Western Texas, Arizona, California, and Wyoming.

Several allied species have been described, among them J. Knightii by Nelson, and another by Sudworth, which occurs in the southwest.

Juniperus Sabina, L. Savin, Juniper, Swedish Juniper

A prostrate shrub with appressed leaves in pairs; margin slightly or indistinctly denticulate; berries on short recurved peduncles; 3-4 lines in diameter, 1-3 seeds. It contains the substance sabinol. It is officinal.

Distribution. Along the Atlantic coast, from Massachusetts westward to New York, Minnesota, Montana, and British Columbia, also in Europe and Asia.

Poisonous properties. The wood of Red Cedar is extensively used in the manufacture of lead pencils and was formerly also employed in making cigar boxes. The fruit of the common Low Juniper (J. communis) is used for flavoring gin. Red Cedar contains a fragrant volatile oil consisting of cedrol and cedrene. Cases of poisoning from this genus have been reported.

ANGIOSPERMAE

Ovules enclosed in an ovary.

MONOCOTYLEDONEAE

Embryo with a single cotyledon, first leaves of germinating plantlet alternate; stems endogenous, consisting of an outer part, an inner mass of cells the parenchyma, and the bundles distributed through the mass; no distinction into pith, wood, and bark; leaves generally parallel veined, usually alternate and sheathing at the base; flowers generally on the plan of 3. This group of plants includes the palms, grasses, lilies, duckweeds, etc.

PANDANALES

Marsh plants, herbs or trees with linear leaves; flowers in spikes or heads; perianth of bristles or of chaffy scales; ovary 1, 1-2 celled; endosperm mealy or fleshy. This order includes the Cat-tail (Typha latifolia), Screw pine (Pandanus), and the Bur-reed (Sparganium). The ripe fruit of the Pandanus fragrans is used as a relish in the Philippine Islands.

The Cat-tail is reported as poisonous. It is common across the continent and is found in swamps.

HELOBIAE

Aquatic or marsh herbs, leaves various; flowers perfect, monoecious or dioecious; perianth present or absent; stamens 1-numerous; carpels 1 or more, mostly distinct; endosperm none or little. This order includes the Pond Weeds (Potamogeton), of which there are many species, which float in the water and often give trouble in ponds of parks; fresh water eel grass (Vallisneria spiralis), water weed (Elodea canadensis), a troublesome weed in the canals of England and Europe. All of these plants are abundant in our fresh waters and afford food for crustaceans, which in turn are used as food for fish.

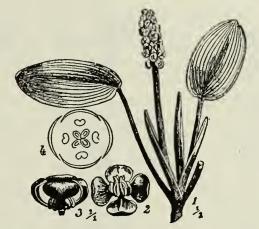


Fig. 131. Pond-weed (Potamogeton natans). 1. Apex of flowering shoot. 2. Flower viewed from above. 3. Flower viewed from side. 4. Diagram of flower. (After Wossidlo).

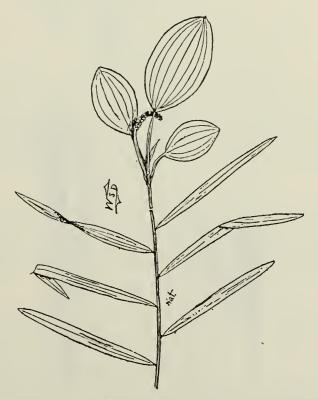


Fig. 132. Pond weed (Potamogeton). Common in fresh water ponds.

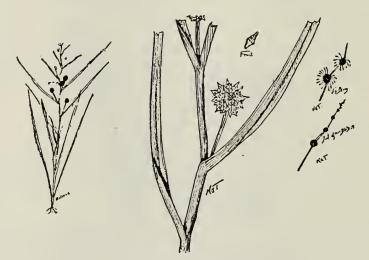


Fig. 133. Bur-reed (Sparganium). .

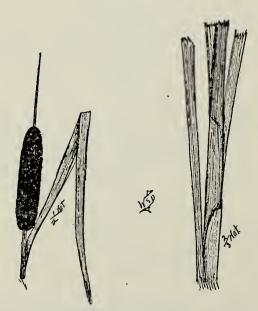


Fig. 134. Cat-tail (Typha latifolia). A common weed along shores of lakes and streams.

ALISMACEAE. D. C. Water-Plantain Family

Aquatic or marsh herbs, generally with smooth, sheathing leaves; flowers perfect, monoecious or dioecious; sepals 3, persistent; petals 3, the larger, deciduous, imbricated in the bud; stamens 6 or more; anthers 2-celled, extrorse;

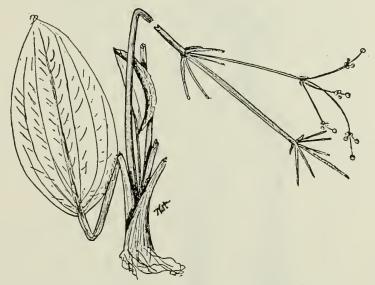


Fig. 135. Water Plantain (Alisma Plantago). A common marsh plant.

pistils numerous or few, usually with a single ovule in each cell; fruit an achenc; seeds small, crcct. About 70 species of wide distribution in swamps. The Water Plantain (Alisma Plantago) of Europe and North America is common in the northern states. Several species of Arrowheads (Sagittaria) are used as food by the Indians and Chinese.

Sagittaria L. Arrowhead

Perennial with tuber-bearing root stocks and milky juice; basal leaves long-petioled, scape sheathed at the base; flowers monoecious or dioecious, borne near the ground in whorls; sepals persistent in pistillate flowers, reflexed or spreading; petals 3, white, deciduous; stamens indefinite; pistillate flowers with distinct ovaries; ovule solitary; fruit an achene in dense clusters; seed erect, curved.

Sagittaria Engelmanniana, J. G. Sm.

Perennial with stolonifcrous roots; leaves very variable; scape 1-4 feet high, angled; lower whorl fertile; pedicel of fertile flowers, at least half the length of the sterile one; filaments smooth; achenes obovate with a long curved or horizontal beak.

Distribution. Across the continent and in Europe.

Poisonous properties. The tuberous stolons are eaten; if there is any poison contained in the raw state it is probably removed by methods of preparation for food.

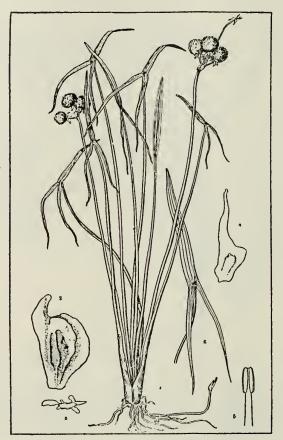


Fig. 136. Arrowhead (Sagittaria Engelmanniana). Common in low grounds. Some species supposed to be poisonous. (After Miss G. E. Johnson, Rep. Mo. Bot. Garden).

GLUMIFLORAE

Endogenous plants mostly herbaceous; stems (culms) narrow or without leaves; leaves usually narrow and clongated; entire or scrulate; flowers small, generally perfect, in the axils of dry chaffy scales, called glumes; arranged in spikes or in panicles consisting of spikelets.. 2 families, *Gramineae* and *Cyperaceae*.

I. GRAMINEAE. Grass Family

Fibrous-rooted annuals or perennials, rarely woody, generally with hollow stems; alternate 2-ranked leaves, sheaths split or open on the side opposite the blade; flowers consisting of 2-ranked glumes, forming a 1-many-flowered spikelet; flowering glumes enclosing a small bract called the palet; stamens 1-6, usually 3; anthers versatile, 2-celled, stigma hairy.

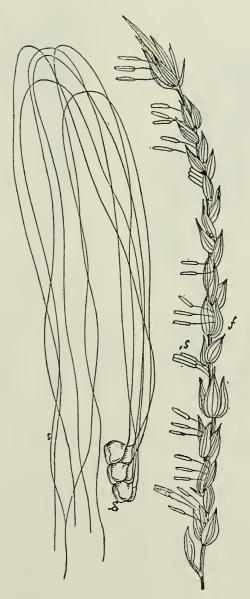


Fig. 137. Corn (Zea mays). To the left pistillate flowers; g, ovary; s, stigma; to the right staminate flowers; s, stamens.

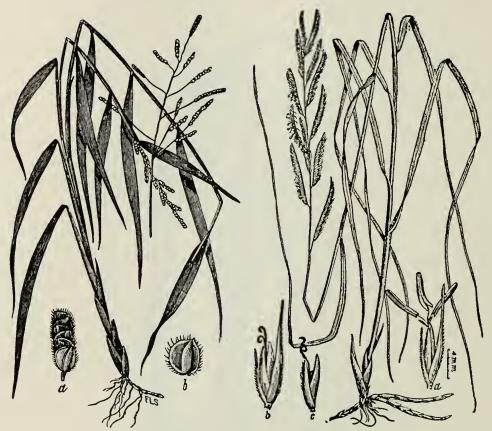


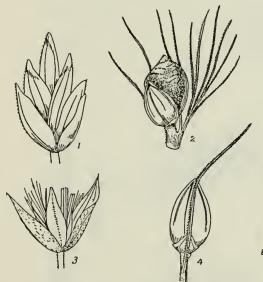
Fig. 137a. Rice-cut-grass (Leersia lenticularis). The sharp edges of the leaf of this grass often cut the flesh of animals.

Fig. 137b. Porcupine grass (Spartina cynosuroides). The sharp edges of this leaf cut like a knife, often wounding animals.

A large order of about 3500 species, many of which are very important to man. Among them are the wheat, oats, rye, corn, wild rice, sorghum, and sugar cane, the two latter being the sources of some of the sugar of commerce. Many grasses, also, are important forage plants, among which may be named blue grass, timothy, brome grass, and red top. Some grasses are used in medicine.

The Bamboo, native of the tropics, is valuable, being used not only for building purposes, but also in the manufacture of household furniture and in other ways.

Very few of the grasses have deleterious properties. A few, such as sleepy grass and millet, the latter of which is injurious to horses, are known to be poisonous. Some grasses, because of their stiff awns, penetrate the skin and even perforate the intestines, inflicting dangerous wounds. Needle grass and squirrel tail grass, or wild barley, are known to inflict injuries by lodging between the teeth, thus causing pus infection.



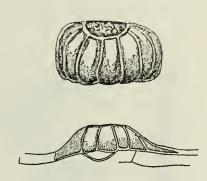


Fig. 137d. Scent glands of stink grass (Eragrostis major).

Fig. 137c. Spikelets of various grasses. 1. Common blue grass (Poa pratensis). 2. Setaria glauca. 3. Spikelet of blue-stem grass. 4. Bristly fox-tail (Setaria verticillata).

A great many grasses, because of their sharp edges on the leaf, inflict injuries by cutting the flesh. Of these we may mention the rice-cut-grass, (Leersia), and porcupine grass, (Spartina cynosuroides).

Holy grass (Hierochloe odorata) is sweet scented and contains coumarin. Indians use it to weave in baskets, mats, etc. Job's Tears, Coix lachryma, is used for rosaries.

KEY FOR GENERA OF GRAMINEAE

| Spikelets jointed upon the rachilla below the glumes, 1-2 flowered. |
|---|
| Rachis bearded, spikelets spicate in pairs |
| Rachis not bearded. |
| Pedicels bristle bearing |
| Pedicels not bristle bearing. |
| Spikelets enclosed by a bur |
| Spikelets plano-convex, not enclosed by a bur3. Paspalum |
| Spikelets in pistillate flowers, borne on a cob |
| Spikelets not usually jointed above the persistent lower glumes. |
| Spikelets 1-flowered. |
| Awn simple twisted |
| Awn 3-pointed |
| Spikelets more than 1-flowered. |
| Spikelets 2-several flowered, rachis often bearded, flowering glume |
| with a twisted awn |
| Spikelets 1 or more-flowered with a zigzag jointed rachis, channeled. |
| Spikelets solitary at the notches. |

| Flowering glumes with the backs turned to the rachis |
|--|
| 10. Lollium |
| Flowering glumes with their sides turned to the rachis |
| 11. Agropyron |
| Spikelcts 2-6 at each joint of the rachis |
| Rachie not channeled 9 Branue |

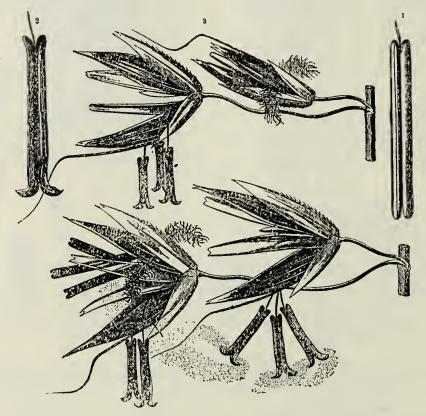


Fig. 138. Spikelets of tall-meadow-catgrass (Arrhenatherum elatius), 1 & 2. Stamens, 3. The lower flower with protruding styles; upper flower with protruding stamens. The lower scales are called sterile glumes. Each flower consists of a palet and flowering glume, stamens and pistil.

1. Zea. Mays. L.

Spikelets unisexual, monoecious; the staminate 2-flowered, in pairs, one sessile, the other pedicellate, arranged in terminal branches of a terminal panicle; the pistillate 1-flowered, sessile crowded in several rows, along the much thickened continuous axis arising from the lower leaf-axil and closely enveloped by numerous large foliaceous bracts; glumes 4, awnless; those of the staminate spikelet acute; those of the pistillate very broad and obtuse or emarginate; grain hard, only partially enclosed by the fruiting glumes. This well-known, tall, and striking annual grass has erect stems and broad leaves;

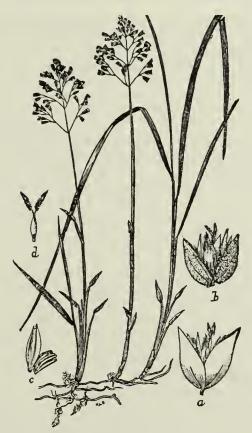


Fig. 139. Vanilla grass (Hierochloe odorata). a. Spikelet with nearly equal lower glumes; b, with lower glumes removed, showing third and fourth scabrous glumes; c, palea with stamens; d, pistil. (Div. of Agros. U. S. Dept. of Agric.).

the terminal, staminate inflorescence forms the "spindle" and the long, projecting styles of the pistillate flowers constitute the "silk"; the cob is formed by the union of the axes of several female spikes into a much thickened body.

The 1 or 2 species are of American origin, presenting many varieties in cultivation known as corn, Indian corn or maize (Zea Mays). Dr. Sturtevant has arranged cultivated corn into the following groups:

Pod-corn, Zea tunicata. Pop-corn, Zea everta. Flint-corn, Zea indurata. Dent-corn, Zea indentata. Soft corn, Zea amylacea.

Sweet corn, Zea saccharata.

Starchy sweet corn, Zea amyleasaccharata.

The so-called species and groups of Dr. Sturtevant are hardly to be regarded as varieties. Some of the forms under conditions of culture and climate, revert to the original type.

A plant cultivated for so long a time by the Indians and civilized man has naturally given rise to diverse forms which we regard as nothing more than races of the very polymorphic species Zea Mays.

Some years ago Dr. Watson obtained from Moro Leon, through Prof. Duges, some corn which he considered a new species, calling it *Zea canina*. He says:

The natural supposition was that we had here at least the original wild state of our cultivated maize. A careful comparison of the two, as thorough as the material at hand of the cultivated forms would permit, has led me first to doubt the probability of this, and now to consider the form in question a distinct species. The differences upon which this conclusion is based are in the habit of growth, the arrangement of the staminate spikelets, and the nervation of their glumes, the form of the glumes of the pisttillate flowers, and the ready disarticulation of the ripened ear.

Dr. Harshberger, who is certainly a most careful observer, and who carried on some most interesting experiments on hybrids, considers our maize of hybrid origin and Zea canina is a hybrid of corn and Euchlaena. He says:

Maize relates itself botanically to a native Mexican grass, teosinte (Euchlaena mexicana); and the fertile bybrids of this grass and maize are known, producing a plant described by Watson as Zea canina. From the peculiar behavior of these hybrids, the writer has suggested that our cultivated maize is of hybrid origin, probably starting as a sport of teosinte, which then crossed itself with the normal ancestor, producing our cultivated corn. This is speculative, but there cannot be any doubt that the close relationship of maize and teosinte points the way to the determination of the botanical charactera of the original wild corn. Recently, Montgomery has suggested a theory as to the mature of the maize ear, in which, in conclusion, he states that corn and teosinte may have had a common origin, and that in the process of evolution the cluster of pistillate spikes in teosinte were developed from the lateral branches of a tassel like structure, while the corn ear developed from the central spike. It is probable that the progenitor of these plants was a large, much-branched grass, each branch being terminated by a tassel-like structure bearing bermaphrodite flowers.

Corn holds the first place in the list of crops produced in this country, and North America produces four times as much as the remainder of the world. According to C. P. Hartley, Europe stands second, South America third, and Africa fourth. As a corn-producing country the United States has no rival; Argentina stands second, Hungary third, and Italy fourth. The average corn yields in four central states for five years, 1902-1906, were as follows:

| State. | Bushels. |
|----------|-------------|
| Illinois | 342,115,835 |
| Iowa | |
| Nebraska | |
| Missouri | |

Maize is one of the most important cereals of North America, being used as a food for man and stock, in the manufacture of starch and glucose, and in medicine, the corn silk being used as a mild stimulant and diuretic. The oil from the embryo is a yellow viscid transparent liquid having a peculiar odor of corn meal. The silk contains maizenic acid.

Injurious properties. In many sections of the country where corn is grown and cattle allowed to feed on corn stalks, a disease occurs which has been called the corn stalk disease. This has been attributed to various causes such as corn smut, a bacterial disease, nitrate poisoning, bacterial poisoning, and impaction of the stomach. Corn stalks are not easily digested and it is not to be wondered at that impaction should occur when cattle do not have access to plenty of water.

This same disease goes under other names, and it may be that there are several distinct types of diseases due to the feeding of corn stalks. Dr. Bitting describes a Septicaemia hemorrhagica which is caused by an organism, the cocco-bacillus. The symptoms of this disease are as follows:

The symptoms depend upon the point of attack. If the respiratory system be attacked, there will be a rapid rise of temperature, difficult and rapid breathing, standing with the feet wide apart as in pneumonia, short coughing, the tongue protruded, and eyes prominent and congested. The animal will move only when urged to do so. The attack lasts for only a few hours. If the pneumonia be of less severe type the kidneys and bowels may show some affection before death.

If the bowels be the seat of attack, there will be bloating colic, noisy intestinal movement, straining and diarrhea. The bowel movements are soft, fluid, and foul smelling, and may be blood stained. The urine will also be blood stained. If the infection take place from a superficial abrasion, the part will swell rapidly, become very large, be hot and painful, does not pit upon pressure, and does not crepitate. The swelling extends rapidly and if in the region of the neck, will cause suffocation. The course is short and generally fatal.

Dr. Bitting states that this disease must be differentiated from the corn stalk disease due to poisoning and that post mortem must be the means of separation in some instances. The only remedy is a change of pasture, as little can be done otherwise.

Recently much interest has been attached to the disease known as Pellagra, which has been treated elsewhere in this volume, but in this case it may be of interest to know that the disease has made its appearance in several of the southern states, notably Alabama and South Carolina; cases have also been reported in Maryland and Massachusetts, and a number of them in the Insane Hospital in Illinois. It is believed by the experts who have investigated this question that it is in some way associated with corn. For instance, Dr. Lavinder who investigated this disease with Assistant Surgeon-general Wyman, cites the case of the disease on the Island of Corfu, where an epidemic followed when the people began to use an inferior imported quality of maize. Previous to this they had used their own maize which was carefully selected and prepared. There can be no question according to Lavinder that the introduction of maize collected in Spain, France, and Italy, with unsanitary conditions and the use of poor maize greatly influence the spread of this disease in those countries. It is practically unknown in those countries where maize is not a staple article of food.

Dr. Arlsberg, of the U. S. Department of Agriculture, in a discussion of this disease before the American Society of Tropical Medicine, states that corn is one-fifth of the food of the Tennessee and Georgia mountaineers and one-third of the food of the negroes. Under the present conditions this corn is collected before maturity and often is shipped to distant points in poorly ventilated cars which makes it possible for moulds of different types to develop. Furthermore, the same person is authority for the statement that, in ten generations the fat content of corn has increased from less than 5% to 7 1/3% and that toxins are found to be related in quantity to the oil produced in the seed. Then, too, the weather conditions in this southern corn region have been extremely favorable in the last ten years for producing corn which would be immature and subject to moulds when transported.

Now it is a well known fact that for many years throughout the south they have had trouble with the so-called forage poisoning affecting live stock which had been fed corn, especially when mouldy. It seems to the writer that there is some relation existing between Pellagra in man and forage poisoning in horses and cattle. Both are essentially produced by some toxic substance.

In this connection, the bulletin on the Grand Traverse or Lake Shore Disease, as investigated by C. D. Smith,* C. E. Marshall and Dr. Ward Giltner, is interesting.

2. Andropogon. (Royen.) L. Beard Grass

Tall annual, or perennial grasses with spikelets in pairs upon each joint of the slender rachis; usually narrow leaves; terminal and axial racemes, one of them sterile, the other sessile, 1-flowered, and fertile; lower glume the larger, coriaceous and nerved, the second acute; stamens 1-3, grain free.

About 150 species widely distributed in tropical and temperate regions. Some of the species of the Andropogon L. are excellent grasses for forage purposes. Quite a number of them produce valuable oils like Pamorusa oil, obtained from Andropogon Schoenanthus, lemon grass oil from Andropogon



Fig. 140. Johnson-grass (Andropogon halepensis).
a, spikelet; c and d, glumes; e, f, g, parts of the flower

^{*} Sp. Bull, Mich. Agr. Exp. Sta. 50: 10.

citratus, and citronella oil from A. Nardus. The fibrous roots of the aromatic Cuscus grass of India (A. muricatus), produce a substance used mainly as sachet powder; the fibres of the plant are used extensively by the natives in making mats.

Andropogon halepensis. Brot. Johnson Grass

A stout perennial with smooth, erect, simple culms, 3-5 feet hight; and strong creeping rootstocks; panicle open, 6-12 inches long; the 3-5 flowered racemes clustered toward their extremities; outer glume coriaceous, second glume equaling the first and convex below, the third glume shorter than the outer ones, membranaceous, palet broadly oval; fourth glume ciliate awned; palet shorter than the glumes; nerves ciliate.

A troublesome weed throughout the Southern States. First introduced as a forage plant.

Andropogon Sorghum. Brot. Sorghum

An annual with long, broad, flat leaves and ample terminal panicle; spikelets in pairs at the nodes, larger and rounder than in the preceding; rachis



Fig. 141. Sorghum (Andropogon Sorghum). 1. Kaffir corn; 2, Jerusalem corn; 3, Ambersorghum. (Kansas State Board of Agrl.).

not articulate; sessile spikelet with 4 scales, the outer hard and shining, the inner hyaline; the fourth scale on and subtending a small palet and perfect flower, or occasionally the palet wanting.

Sorghum is contained in a number of cultivated plants which are classified by Mr. C. R. Ball under (1) Broom Corn, (2) Shallu, (3) Durra, (4) Sorghum and (5) Kaffir. Broom Corn used for the manufacture of brooms, is grown chiefly in the central Mississippi Valley, Kansas, Oklahoma, and the Panhandle of Texas. The Shallu, also known as Egyptian wheat, is culti-

vated extensively in India. The Durras have been cultivated for centuries in Egypt, and other countries of Africa, and in India for human and animal food. They are now cultivated in the United States, chiefly from Kansas to Texas. The Kaffir, native to eastern Africa from Abyssinia to Natal, was introduced into this country in 1875, and is grown chiefly in the semi-arid regions for forage. Sorghum is grown largely for fodder. The pithy juice contains cane sugar in variable amounts, and is used both for sugar and in the making of syrup, although the growing of sorghum for syrup seems to be on the decline. According to the census of 1890 the production was 24,000,000 gallons; in 1900, 17,000,000.

Poisonous properties. Sorghum has long been recognized as poisonous. Mr. C. W. Warburton in Bailey's Encyclopedia of Agriculture, says:

Sorghum makes excellent pasture for hogs, but in many sections it must be pastured sparingly, if at all, by sheep and cattle. After periods of extreme drought, or when growth is stunted from other causes, the leaves of the sorghums often contain a large amount of prussic acid. A small quantity of this poison is fatal to stock, and death frequently results soon after the sorghum is eaten. Normal growth seldom contains prussic acid in appreciable quantities, and it largely disappears in curing, so that cured sorghum may be fed with little danger. There is also some danger from bloating; cattle and sheep should not be turned on sorghum pasture when hungry or when the plants are wet. With the exercise of care, however, the crop can usually be pastured with safety. It should be at least two feet high before stock are turned on it; for cattle, sheep and horses it may be much more matured than for hogs

Frosted cane is said to be especially injurious. Dr. R. H. True, of the U. S. Dept. of Agriculture, in commenting on the poisons from sorghum, says:

This office has from time to time received communications from stockmen, especially in the lower part of California, Arizona, and adjacent territory, expressing a suspicion that the eating of the Johnson grass had caused the death of stock with rather sudden and violent symptoms. There has seemed to be little ground in poisonous-plant literature to support such an explanation. Last summer, however, convincing observations were reported from California by a stockman who had lost heavily, and a supply of the grass in question was obtained. The result of the study of this material was positive.

Mr. A. C. Crawford, who investigated some cases, says:

It has been noted that deaths in cattle frequently occur when, on account of the failure of rain, the plants which have reached a certain size become stunted and withered. The texic principle appears simultaneously over a wide area, but soon disappears if a rainfall occurs. The deaths of cattle have been attributed by some to an insect living upon the plant, and in Australia it is the belief that Sorghum vulgare which also yields hydrocyanic acid, becomes more poisonous when attacked by an insect during a drought. A similar observation has been made with Sorghum vulgare in the Sudan. Balfour found that one specimen of the plant which harbored aphids yielded more hydrocyanic acid than a second one without parasites. Pease has lately claimed that the deaths from Johnson grass in India were really cases of nitrate poisoning, as he found 25 per cent of nitrate of potassium in the stem of the plant and was able to produce somewhat similar symptoms in animals by feeding them this salt. Johnson grass is being introduced into Australia as a fodder plant, but as yet no reports of its poisonous action there have been noted by the writer.

Dr. George H. Glover of Colorado, also reports large loss of cattle in that state from eating Kaffir corn. Twenty-one head out of thirty-two cows died within an hour after first being placed upon the feed. Dr. A. T. Peters of Nebraska, investigated the subject of poisoning from sorghum and Kaffir corn. He states that in most cases where death of animals occurred, the animals did not regularly pasture upon sorghum, but broke into the fields from adjacent pastures or as they were being driven past fields of sorghum. The investigations proved that the animals did not die from bloat as had been re-

ported. Dr. Peters speaks with authority as he examined the cases at first hand:

In response to one of these calls, I reached the farm of Mr. Bert Foss, near Aurora, at 7:30 A. M., on August 3, 1901. Two days previous, fifteen head of his cattle had broken into a sorghum field, where they had remained twenty minutes. They were then driven into another field and were not seen again for several hours. When seen, three were sick, all of which died within a few hours. The symptoms were drowsiness, running at the eyes, twitching of the muscles, numbness of the limbs, staggering gait, inability to stand, involuntary passing of urine. On August 2d, two more cows broke through the fence and were on the sorghum field five minutes. One hour later, one of these animals, a four-year-old cow was very sick, but finally recovered.

We turned a small yearling steer on the sorghum at 8:30 Å M., August 3d, but he refused to eat any sorghum, and after thirty-five minutes, two more were turned into the sorghum, where they remained until 10:00 Å. M., when only one, a small, red steer, had taken any sorghum, and he had eaten only a few leaves. They were then turned hack with the herd. At 10:35 Å. M. the small, red steer acted somewhat drowsy, but soon recovered.

At 11:00 A. M. we turned one red heifer and one yearling steer on the sorghum. The heifer was the only animal that are any quantity, and, as subsequent examination showed, she are only one and one-half pounds of green sorghum. At 11:10 this animal dropped to the ground. Upon examination it was found that she had stopped chewing her cud and there was a peculiar twitching of the muscles of the nose and head and also of the body. The animal was very dull. At 11:15 A. M. she was taken out of the sorghim field and allowed to lie in a stubble field. When lying down her head was turned toward the abdomen, presenting the symptoms shown by a horse having the colic. The eyes seemed dull and gave off a water discharge. There was a partial paralysis of the tongue and great quantities of saliva ran from the mouth. The limbs and ears were cold. The pupils of the eyes dilated, pulse not perceptible, mucous membrane of the rectum protruding, involuntary discharge of urine and faeces. Upon pricking the animal with a knife on the lower limbs it showed no feeling. The animal was closely watched in the field by Mr. Foss and myself and we observed that she did not take any weeds, but simply a small amount of sorghum, eating only the tops of the leaves. At 1:30 P. M. the animal was still lying on its right side; all the muscles of the head were contracted and showed involuntary twitching. The limbs were paralyzed and the animal was unconscious; the mucous membrane of the mouth was of a salmon color. At 2:35 P. M. the animal was in great pain, and it was apparent that she would not recover. At the suggestion of Mr. Foss the animal was killed in order to hold a postmortem examination.

Post-Mortem Examination.—Animal still warm. The bowels were opened and contents of paunch carefully noted; there was in all one pound and a half of sorghum leaves to be found in the paunch. No sourness of the contents. The same was immediately put up in Mason fruit jars with clean water and brought to the laboratory. The mucous membrane of the intestines normal, all other conditions of the animal normal.

In regard to the Colorado disease, the following statement is made:

The cattle died on August 20th last. We lost 21 head out of 32 head which had been turned on the corn. Eleven head lived, but 4 of the 11 head had violent spasms, but recovered. The other 7 head were not affected. They were only on the corn 5 or 6 minutes. The first cow died in 15 minutes; nearly all within an hour. One yearling lived over 6 hours. I gave it several doses of aconite, thinking possibly one poison would counteract the other, but it died in great agony. The cattle seemed to all go crazy at once, then stagger like a person intoxicated, fall in all directions, and die where they fell. I stuck all of them with a knife, the same as in alfalfa bloat, but there wasn't any gas in them. The Kaffir corn was planted on sod ground above irrigation. It was from 6 to 15 inches high and was burnt brown from the drouth."

Shortly after the poisoning, Dr. Glover visited the field and collected samples, which he generously placed at our disposal. These samples yielded prussic acid in greater amounts than any yet examined in Nebraska.

It appears more than probable that the sorghum plant under different eli-

matic conditions and different conditions of growth may produce varying amounts of prussic acid. I was told in Texas that the sorghum most poisonous to live stock is the second growth.

In regard to the chemistry of the subject, Dr. S. Avery says:

In 1886 Berthelot and Andre ascribed the cause to excessive amounts of potassium nitrate (salt peter). Williams of the U. S. Department of Agriculture also suggests saltpeter as a cause of the trouble. Hiltner has shown that the amount of nitrate in Nebraska fields was too small to produce fatal results. This writer suggests that the plant under certain conditions develops a highly poisonous chemical compound. Slade in the Annual Report of the Station for the present year (1902) put forward the theory that such a compound might be produced by the action of an enzyme upon a glucoside found in the plant through a process of abnormal growth. On June 27th of the present year the Chemical News of London contained an article on Cyanogenesis in Plants by Dunstan and Henry. This article, which finally confirms Slade's prediction, was not known to Mr. Slade or to the writer till October 10th. In brief, the English Chemists isolated from Egyptian Sorghum vulgare a glucoside capable of liberating prussic acid. In the meantime Mr. Slade had detected Prussic acid in fatal sorghum grown in western Nebraska, determined the per cent, and secured strong evidence in favor of the glucoside theory.

During the first two weeks in September, the writer discovered that Prussic acid could be obtained from leaves of healthy sorghum in the fields about the Station. As the past season was abnormally wet, nearly all of the fields had made a vigorous growth. By distilling water from a sufficient quantity of leaves, determinable amounts of Prussic acid were evolved in all cases, though the amount was well below the danger line. Of the common Nebraska forage plants, sorghum and Kaffir corn alone yield Prussic acid

The substance dhurrin C₁₄H₁₇NO₇ occurs according to Dunstan and Henry in young plants of A. Sorghum. A glucoside resembling that found in almonds also occurs; it differs however but is capable of being converted into hydro cyanic acid, HCN. The investigations made at this station show that the prussic acid is not present as such, but that it is liberated from a glucoside, (1) by an enzyme in the plant as in the case of sorghum poisoning, and (2) by the action of boiling water on the plant. Glucosides of this sort are in themselves harmless and are dangerous only when they liberate prussic acid. The experiments mentioned above also showed that even dried plants may contain a very large amount of combined prussic acid. We should expect that such a fodder would be as fatal to stock after curing as when standing in the field. Experience, however, seems to prove the contrary. Enzymes rapidly become inactive when dried in the presence of protein substances, according to Dr. A. F. Woods, Chief of Division of Plant Pathology.

Antidotes. Prussic acid has a tendency to unite with certain carbohydrates, forming additional products. These compounds are much less poisonous than the free acid. Both glucose and milk sugar unite with Prussic acid to some extent even in dilute solutions. Aside from this action these carbohydrates retard the action of the enzyme in liberating Prussic acid. These facts suggest that, in case the animal is not in such a condition as to render medical treatment out of the question, the following may be affective:

A strong solution of glucose, which nearly every farmer has at hand in the form of "corn syrup" or molasses, may be administered.

Large quantities of milk have in a number of instances been administered apparently with good effect.

In all cases the animal should have as much fresh air as possible.

3. Paspalum, L. Paspalum

Spikelets spiked or sometimes racemed, in 2 to 4 rows on one side of the flattened or filiform rachis, awnless, 1-flowered; glumes 3, rarely only 2, 1 glume flowering; flower coriaceous, orbicular or ovate; stamens 3; spikes 1 or more at or toward the summit of an elongated peduncle.

. Species about 160, chiefly in warm temperate regions in both hemispheres. In South America they constitute an important part of the plants of the Pampas. One species is used in medicine and several species are excellent forage plants for the South. One species is troublesome as a weed in the Southern States. The Koda Millet (*P. scrobiculatum*) known to be poisonous and injurious to animals and man in India is used during times of scarcity of food and causes poisoning. The seed, especially the testa and pericarp, contains a narcotic poison which causes delirium and vomiting. Cattle should not be allowed to feed on it when it is ripening.



Fig. 142. Corean Foxtail (Setaria italica.)

Fig. 143. Corean Foxtail Millet (Setaria). U. S. Dept. Agr.

4. Setaria, Beauv.

Spikelets jointed upon the pedicels, paniele densely racemed or spiked, surrounded at the base by a few or many persistent awn-like bristles, which rise below the articulation of the spikelet.

Species about 10, in temperate and tropical regions. Some species are used as food, especially in China, Japan and India. Several are important forage plants, like the broom corn millet, and Hungarian grass. Three species are weedy in eastern North America.

Setaria italica, Beauv Italian Millet or Hungarian Grass

A stout, erect, somewhat glaueous annual, 3-8 feet high, with broad leaves; large, dense, compound, spiciform panieles 3-8 inches in length; nodes bearded, with short, appressed hairs; leaf-blades laneeolate, narrowed at the base, long-aeuminate, 8-16 inches long, ½ to 1½ inches wide, seabrous; panieles dense, eylindrical, ½ to 1½ inches in diameter; rachis densely villous; setae 1-3, green or purplish, retrorsely seabrous; spikelets elliptical, strongly convex, 1½ to 2 lines long, obtuse; second and third glumes about equaling the flowering glume, 5-7-nerved; flowering glume glossy, nearly smooth. Widely cultivated. Quebec to Minnesota, south to Florida and Texas.

Setaria germanica, Beauv. German Millet

A eacspitose annual, from 1-3 feet high, with narrow panicles, about ½ inch in diameter, and long, usually purple setae; some forms approaching Setaria viridis.

This form is usually regarded as only a variety of the Italian Millet, and is found in cultivation only or perhaps springing up from seed on land cultivated the preceding season. The German Millet differs from the Italian in having a more dense or compact, and usually erect paniele or "head." Widely cultivated in most parts of the world.

Poisonous Properties. Numerous complaints have been made from time to time with reference to poisoning from millet.

Dr. Hinebauch states in regard to this trouble that in the winter of 1891 and 1892 a disease commonly called millet disease was prevalent to a considerable extent in North Dakota and that this disease was attended by a death rate of 7-10 per cent. It received the name of millet disease from the fact that from 95 to 98 per cent of the animals that were affected had been fed on millet. He says:

"When millet is fed in considerable quantities it stimulates the kidney to increased action. The urine is light colored and the bladder evacuated every two or three hours, large quantities of water being passed at each time. At the time the first symptoms of lameness were noticed, the kidneys had almost ceased to act."

And then he goes on to say:

"When the eause was kept up a sufficient length of time for the reaction to set in, the material which would under normal conditions be secreted by the kidneys was allowed to remain in the system and produce deleterious effects."

Apparently the condition of the millet had little to do with this action. In a later bulletin on the same subject Dr. Hinebauch reports a more extended investigation, giving considerable experimental data as well as urinary analyses.

The post mortem examinations revealed some interesting facts. The cartilages on the ends of the long bones show deep furrows running in a direction parallel with the motion during flexion and extension.

Both grooves of the astragalus were partially denuded of cartilage, so that the corresponding elevations of the tibia which articulate in the grooves did not have cartilage interposed between them. The whole general appearance, instead of being of a white, glistening color, was of a dark, dull color bordering on brown. The fluid which escaped from the joint when opened, instead of being a yellow, amber color, was brown and contained red blood corpuscles, indicating that inflammation was present. The joint fluid was brownish black in color and contained red blood corpuscles.

In conclusion we would say that our experiments here have thoroughly demonstrated that millet, when used entirely as a coarse food, is injurious to horses. (1) In producing an increased action of the kidneys. (2) In causing lameness and swelling of the joints. (3) In producing infusion of the blood into the joints. (4) In destroying the texture of the bone, rendering it softer and less tenacious, so that contraction causes the ligaments and muscles to be torn loose. The experience of many farmers with whom I have talked confirms the above conclusion, and we could multiply case after case showing that the above conditions are the results of feeding millet.



Fig. 143. Sandbur (Cenchrus tribuloides). spiny bur enclosing spikelets; b, section of the same; c, lateral view of a spikelet. U. S. Dept. Agrl.

The North Dakota Station has published the results of further experiments on the subject of feeding millet. Two tests were made. In the first trial two geldings in good health were fed hay and grain for about two weeks. Millet was then substituted for hay for about ten days. These experiments confirmed those made previously.

Ladd has isolated a glucoside from the aqueous extract of millet hay, which, when fed in small quantities, gave the characteristic symptoms.

From the experiments made by Dr. Hinebauch and others, it would appear that feeding millets alone as coarse fodder is injurious to horses. It produces an increased action of the kidneys and causes lameness and swelling of the joints. It causes an infusion of blood into the joints and destroys the texture of the bone, rendering it soft and less tenacious, so that the ligaments and muscles are easily torn loose.

5. Cenchrus, L. Sand Bur

Annual or perennial grasses; flat leaves; spikelets surrounded by a spiny involucre which becomes coriaceous and forms a deciduous, hard, rigid bur which falls away at maturity; glumes 4, the 2nd and 3rd membranaceous, the 4th hard; the palea enclosing the perfect flower; stamens 3; styles united below.

Species about 12 in tropical and warmer temperate regions. One widely distributed from Maine to New York, Florida, Texas, California and the Dakotas.

Cenchrus tribuloides, L. Sand Bur

An annual with erect culms a foot or more high; flat leaves about 6 inches long; burrs of the involucre with strong, barbed spines; 2-flowered.

Distribution. Common in sandy fields and waste places; a weed along rail-roads and in sandy soil.

Injurious Properties. This plant frequently inflicts mechanical injuries, entering the flesh and thus causing serious inflammation. This applies to man as well as to lower animals.

Aristida, L. Triple Awned Grass

Perennial or annual grasses; narrow, often involute leaves; spikelets narrow, 1-flowered; outer glumes unequal, often bristle pointed; flowering glume tipped with 3 awns; palet small, 2-nerved; stamens 3; styles distinct; grain free, linear, enclosed in the scale; callus variable, often sharp-pointed and rigid. About 100 species in warmer regions of both hemispheres but of very little economic value, the majority being found in dry sterile soil; several species, like the Purple Aristida, however, are common in dry soils of the West. The latter is of little value for forage purposes. The awns of Aristida hygrometrica of Queensland bore into the skin of animals and occasionally reach the intestines, thus causing death.

None of our species produces serious trouble except, possibly, the Long-awned Poverty Grass.

Aristida tuberculosa, Nutt. Long Awned Poverty Grass

A rigid, much-branched annual, 12-18 inches high, with nearly simple panicles, 4-7 inches long; branches erect, rather distant, the lower in pairs, one short and few-flowered, the other elongated and many-flowered; empty glumes



Fig. 144a, Long-awned Poverty Grass (Aristida tuberculosa), a, Spikelet with lower glume; b, flowering glume with divergent long awns. (U. S. Dept. Agrl.).

Fig. 144b. Short-awned Poverty Grass. (A. basiramea). Occurs in sandy and gravelly soils. (U. S. Dept. Agrl.).

nearly equal, 12 lines long, awn-pointed; flowering glume about 10 lines long, twisted above to the division of the awns, and with a densely barbate sharp-pointed callus; awns nearly equal, divergent or reflexed, 1½-2 inches long, distinctly articulated with the glume.

Injurious properties... The sharp pointed callus slightly injurious in the same manner as Stipa.

7. Stipa, L.

Perennial grasses with 1-flowered spikelets, flower falling away at maturity from the membranous, persistent, lower glumes, fertile glumes coriaceous, cylindrical, involute, and embracing the smaller palet and cylindrical grain; a long twisted or spiral awn jointed with the apex, the base consisting of a beard and sharp pointed callus; stamens generally 3; stigmas plumose.

About 100 species found in temperate and tropical regions. The Stipas often produce injurious effects upon animals.

Injurious properties. It has long been known that Stipa capillata, L., indigenous to Russia, and the Stipa spartea, Trin., and S. avenacea, L., native to North America, as well as Aristida hygrometrica Br., native of Queensland,

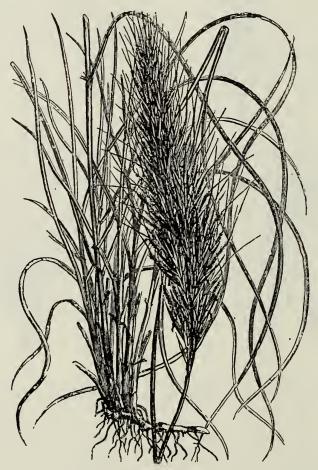


Fig. 145. Esparto Grass (Stipa tenacissima). Used for making paper, ropes and mats. It is not known whether this species, like the St. inebrians and the S. sibrica, acts like a narcotic on animals. (Baillon Dict.).

and Heteropogon contortus, L., native of New Caledonia, frequently bore into the skin and intestines of lower animals where they cause fatal inflammation and peritonitis. Prof. Blanchard in a recent number in "Archives de Parasitologie" calls attention to injurious properties of Stipa Neesiana which is found in Uruguay and other countries of South America. In this case the needles injure the eyes producing in intense keratitis often followed by inflammation of the cornea. Sheep become blind and thus are unable to get food, hence

die from hunger and thirst. An instance is also recorded of a case where so many of these needles had accumulated among the feathers of an American Ostrich as to cause extensive ulceration which finally resulted in the death of the bird. The old world *S. inebrians* acts very much like our sleepy grass.

Stipa comata, Trin. and Rupr. Western Stipa. Needle Grass

A rather stout, erect, caespitose perennial, 11/2-4 feet high, with mostly involute leaves; loosely-flowered panicles, 8 to 12 inches long; spikelets with nearly equal, long-attenuate-pointed, empty glumes about 12 lines long, and thinly

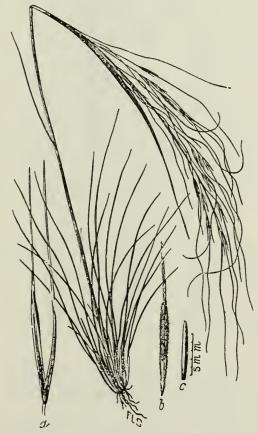


Fig. 146. Western Stipa or Needle Grass (Stipa comata).

pubcscent flowering glumes about 6 lines long; awn slender, 2½-3 inches long, strongly flexuose or variously curled and twisted. Distributed in western Iowa, Nebraska, Utah, Oregon, California and Arizona.

Stipa spartea. Trin. Porcupine Grass

A stout, erect perennial, with simple culms 3 to 5 feet high; long, narrow leaves and contracted, few-flowered panicles, 4 to 8 inches long; spikelets

larger; empty glumes subulate-pointed, 12 to 18 lines long, slightly unequal; flowering glume 8 to 10 lines long, including the barbed and very sharp-pointed stipe or callus, sparsely pubescent below and crowned with a few short hairs;

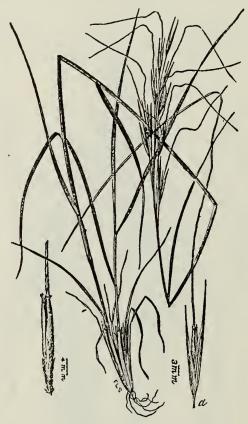


Fig. 147. Needle or Porcupine grass (Stipa spartea). a, a single spikelet; b, floret more highly magnified, with sharp pointed bearded callus. (Div. Agros. U. S. Dept. Agrl.).

palea nearly as long as the glume; awn stout, 3 to 6 inches long, twisted below and twice geniculate above. June to August. Common on dry, gravelly roads and high prairies.

Distribution. North America. From Wisconsin, Illinois to Missouri, Kansas, Nebraska, Dakotas and Minnesota to New Mexico, Manitoba to British Columbia.

Injurious properties. Dr. M. Stalker says the fruits of the porcupine grass are a frequent source of inconvenience and injury to living animals.

In many of the northwestern counties of Iowa this grass grows in the greatest profusion, and during the latter part of June, the season for maturing and consequent falling of these spines, they are the occasion of much annoyance and in some instances the death of domestic animals. Only such animals as are covered with wool or a

thick growth of long hair are seriously inconvenienced. Sheep suffer most. The spines readily find a lodgment in the wool, and after burrowing through it frequently penetrate the skin and bury themselves in the flesh. A large number of these barbs thus entering the tissues of the body produce an amount of irritation that is sometimes followed by death. I have seen large numbers of these imbedded in the skin and muscular tissues of shepherd dogs that were covered with a thick growth of soft hair. These sagacious animals frequently exhibit the greatest dread at being sent into the grass during the season of danger.

Professor Bessey in his inquiries into the structure and nature of this plant received several responses, one of which, from Professor King, formerly of the University of Wisconsin, was as follows:

In connection with the two notes relating to the fruit of the porcupine grass, it may not be without interest to say that while engaged in geological work in Dakota, north of the Northern Pacific railroad, we were much annoyed by the fruit of this grass. Indeed, I found the only way to walk with comfort through this grass was to roll my pants above my knees and my socks down over my shoes. I also observed, on several occasions, these seeds planted two inches deep in the soil with the awns protruding from the ground. It is plain that with the point of one of these fruits once entered below the soil, the swelling and shrinking, due to varying amounts of moisture, would work the seeds directly into the ground.

The Stipa comata, or needle grass of the west, which is common throughout the Dakotas, and throughout west Dakota, Nebraska, Wyoming and Colorado, is common in prairie hay, and Prof. Thomas A. Williams mentions that, though a forage plant, and not cut until the needles have fallen so that the stock may not be injured, the fruit of this plant often injures stock to a considerable extent. During the past summer in Alberta, Canada, the writer suffered some inconvenience from the penetration of the fruit through the clothes.

Stipa robusta, Vasey. Sleepy Grass

A large grass from 4-6 feet high growing in dense clumps; leaves involute, setaceous, large, flattened, 1-2 feet long; panicle 1-1½ ft. long; spikelets 4-5 lines long, on short pedicels; empty glume nearly equal 3-5 lines long; variable in length up to 1½ inches, slender flexuous; palet about ½ length of glume.

Distribution. From Colorado to Texas and Mexico.

Poisonous properties. This is the grass which is properly called Sleepy Grass and is poisonous. Dr. Vasey says the variety in parts of Texas and Mexico is known as Sleepy Grass, so called for its intoxicating and narcotic effect upon horses or cattle which feed upon it.

In the west this species of grass has received the common appellation of sleepy grass. It has long been regarded by range people as poisonous. Dr. Palmer, who found this grass in Coahuila, observed that it was poisonous to cattle, horses, and sheep, causing them temporary sleepiness. Later Dr. Havard states that in 1888 he received from Dr. M. E. Taylor, of Stanton, N. M., a grass with the following statement:

Hereabouts grows a grass—the eating of which by horses will, within a few hours, produce profound sleepiness or stupor, lasting twenty-four or forty-eight hours, when the animals rally and give no evidence of bad effects. It is known among cowboys as "sleepy grass" and dreaded by them on their "round ups" as their horses are liable to eat it and cannot then be kept up with the herds. The tradition is that horses that have once eaten of it will not touch it again.

To quote from Dr. Havard:

From the same gentleman I received a letter in 1890, in which he says: "Since I corresponded with Dr. Taylor it has been brought to my notice that cattle are affected in a similar way to horses, and that the curious properties which so affect animals are contained in the blades Quite a number of our horses have been ill this spring

after having eaten it. It usually takes them about a week to recover, during which time they are unfit for work, and especially so during the first three days."

Captain Kingsbury, of the Sixth United States cavalry, under date of March, 1890, wrote me from Fort Stanton that the sleepy grass affected nearly all his horses at two camping places, it was hard work to make them walk.

The similarity of symptoms, whether observed in Coahuila or in New Mexico, is certainly remarkable, and furnishes strong evidence of the substantial accuracy of the observations as reported. It would seem, then, reasonably established that this plant possesses narcotic or sedative properties, affecting principally horses, but also cattle and probably other animals; that animals are not fond of it but eat it inadvertently or when under stress of hunger; that cases of poisoning occur especially in the spring, when the radicle and lower blades first come up, and that the active principle resides in these blades, and perhaps only during that season.

8. Avena, L., Oats

Annual or perennial grasses, usually with flat leaves and panicled spikelets; spikelets 2, many-flowered, or rarely 1-flowered; lower flowers perfect, the upper staminate or imperfect; empty glume unequal, membranaceous and per-



Fig. 148. Wild Oats (Avena fatua). a. empty glumes; b, flowering glumes. (U. S. Dept. Agrl.).

sistent; flowering glume deciduous, generally bearing a twisted awn on the back between the two acute teeth at the apex; rachis and base of flower often bearded; stamens 3, style short and distinct; grain oblong, linear, grooved on one side invested by the palet. About 50 species in temperate regions. The cultivated Oats (Avena sativa) is the best known representative of the genus and has long been used for food for man and animals. Several native species produce good forage.

Avena fatua L. Wild Oats

An erect, glabrous annual, 3-5 feet high, with flat leaves and spreading panicles of large, nodding spikelets; spikelets 2 to 4-flowered, with empty glumes 3/4-1 inch long and pubescent; flowering glumes 6 to 9 lines long; awns nearly twice as long as the spikelets. Wild oats is highly esteemed as a forage plant on the Pacific Coast, especially California.

Distribution: Native to Europe but now abundant in grain fields of the Rocky Mountains, the Dakotas, Minnesota, and the Pacific Coast.

Injurious properties. Bezoars are sometimes produced by the common oat and Dr. Harz thinks it is a dangerous food material because it favors the development of these "hair balls." The barbed and awned seed of the wild oat may probably sometimes also lodge in the mouth and produce inflammation or other results of mechanical injuries.

Avena sativa, L. Common Oats

A well known erect annual, 2-4 feet high, with flat leaves and expanded panicles of rather large, pendulous, and, usually, 2-flowered spikelets. Lower florets sometimes awned.

Distribution. Widely cultivated in Europe, North America, Asia, and in all temperate regions. Commonly cultivated in Northern United States, Canada, and the Pacific Coast. The species is native to eastern temperate Europe, and western Asia, although the wild form has not been found. According to some authors, cultivated oats originated from wild oats *Avena fatua*. This is very doubtful.

Injurious properties. Harz reports the occurrence of phytobezoars in horses which had been fed oats straw. These bezoars in their origin and structure are similar to those occurring from feeding on cacti and the crimson clover referred to elsewhere.

9. Bromus, L.

Spikelets 5 to many-flowered, panicled; glumes unequal, membranaceous; lower glume 1-5 nerved; flowering glume either convex on the back or compressed-keeled, 5-9-nerved, awned or bristle-pointed from below to the groove of the oblong or linear grain; stamens 3; styles attached below at the apex of the ovary. Coarse grass with large spikelets at length drooping on pedicels thickened at the apex. About 40 species, of which Beal lists 27 as either native or introduced into the U. S.

Bromus tectorum, L. Awned Brome Grass

A slender, crect, leafy annual, 7-25 inches high, with narrow, softly pubescent leaves and open, nodding panicles, 3-7 inches long; spikelets each 5-8 flowered, with unequal, acuminate-pointed, hairy, empty glumes, and rough or



Fig. 149. Common Oats (Avena sativa), sometimes the cause of phytobezoars in animals. (Ia. Seed Co.)

hairy glumes 4-6 lines long; awns 6-8 lines long; blooming period from June to August. First introduced into the United States from Europe, it is without forage value, and, while not greatly troublesome in Iowa or eastward, has become a serious pest farther west, in Utah and Colorado.

Poisonous properties. This plant produces injuries similar to those caused by Squirrel-tail grass, the awned glumes working in under the teeth causing inflammation and suppuration. Animals eating this grass may lose their teeth as a consequence.



Fig. 150. Awned Bromegrass (Bromus tectorum). a. Sterile or outer glumes. b. Spikelet. U. S. Dept. Agr.

10. Lolium, L. Darnel and Rye Grasses

Annual or perennial grasses with flat leaves and terminal spike; spikelets many-flowered, solitary on each joint of the continuous rachis placed edgewise; empty glumes except in the terminal spikelets; only one flowering glume, rounded on the back, 5-7 nerved, palet 2-keeled; stamens 3; grain adherent to the palet, 6 species, 2 more or less naturalized in the eastern states. Natives of the Old World. Two species, the Italian rye grass and the common rye grass, are valuable forage plants. Darnel is a troublesome, poisonous grass.

Lolium temulentum, L. Darnel, Poison Darnel

An annual with smooth stout culm, 2-3 feet high; leaves with scabrous sheaths and short ligule; spike 6-12 inches long; spikelets 5-7 flowered; empty glumes sharp pointed, as long as the spikelets; flowering glume awned or awnless. Commonly found in grain fields.



Fig. 151. Darnel (Lolium temulentum). b. Spikelet. a. Empty glume. U. S. Dept. Agr.

Distribution. Naturalized in eastern North America and abundantly so on the Pacific Coast.

Poisonous properties. It is a well known fact that a number of grasses are poisonous. It was well recognized by the ancients that darnel (Lolium temulentum) was poisonous, for it is written: "But while men slept, his enemies came and sowed tares among the wheat."

Darnel, when ground up with wheat and made into flour, is said to produce poisonous effects on the system, such as headache and drowsiness. This poisonous property is said to reside in a narcotic principle, loliin, a dirty white, amorphous, bitter substance yielding sugar and volatile acids, which, according to Hackel, "causes eruptions, treinhling and confusion of sight in man and flesh-eating animals, and very strongly in rabbits, but it does not effect swine, horned cattle or ducks." Lindley states that the grain is of evil repute for intoxication in man, beast and birds, and brings on fatal convulsions. Haller

speaks of it as communicating these intoxicating properties to beer. It acts as a narcotic, acrid poison. Darnel meal was formerly recommended as a sedative poultice. In Taylor's work on poisons, the statement is made that the seeds, whether in powder or in decoctions, have a local action on the alimentary canal and a remote action on the brain and nervous system. He states further that no instance is reported of its causing fatal injuries to man, and as much as three ounces of a paste of the seeds have been given to a dog without causing death. Then he goes on to cite the experience of Dr. Kingsley, in which several families, including about thirty persons, suffered severely from the effects of bread containing the flour of darnel seed. These persons had staggered about as if intoxicated. It is claimed by some investigators, however, that this plant is not poisonous. One writer claims to have made bread from flour said

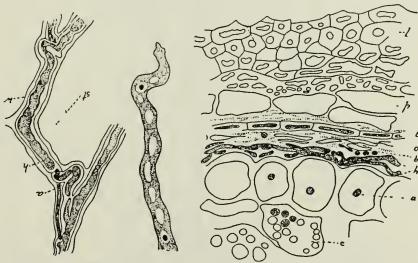


Fig. 152. At left, a hypha from leaf base of seedling of "Darnel" (Lolium temulentum). At right, hyphae in the starch endosperm of a seed. h. hyphal layer of grain nucellus, st, starch cell, w, wall of starch cell, a, knot formation in an intercellular space. After Freeman, redrawn by Charlotte M. King.

Fig. 153. "Darnel" (Lolium temulentum). Section of outer part of a grain which has been in a germinating chamber 24 hrs. l, pale. b. nericarp, i, crushed integuments, o, outer row of nucellus cells, b, cavities with nucellus (probably id cell lumina), h, hyphae, a, aleurone, c, starebendosperm. After Freeman, redrawn by Charlotte M. King.

to contain considerable darnel and experienced no injurious effects. When mixed with flour and water the dough is foamy and narcotic in its action. There are other grasses which produce similar narcotic effects. Quite recently it has been claimed by several European investigators that the fruit of Lolium temulentum contains a poisonous fungus. Guerin states that the hyphae of a fungus constantly occur in the nucleus of the seed and the layer of the caryopsis lying between the aleurone layer and the hyaline portion of the grain or nuccllus. He also thinks that the toxic action of the Loliums is due to this particular fungus hypha. The threads were also found in L. arvense and L. linicolum; but, as yet, have not been found in L. italicum and but once in L.

perenne; the fungus is allied to Endoconidium temulentum, which has been found on rye.

The Lolium fungus, according to Guerin, lives symbiotically in the maturing grain and is therefore not a parasite, but Freeman has observed that occasionally it is injurious although it is generally stimulating. Nestler, who made an examination of L. perenne, L. multiflorum, L. remotum, and L. festucaceum, found nothing comparable to the fungus mycelium which occurred in L. temulentum. He also succeeded in demonstrating the presence of the mycelium of the fungus as indicated by Guerin. According to Nestler, the Fusarium roseum is identical with the fungus occurring in L. temulentum found by Guerin, but this has not been confirmed and seems very improbable. Hanausek considered the fungus to be related to the smuts, but Freeman found no evidence of spore formation; the septa are infrequent and the intercellular course different from that for smuts. The subject has, in recent years, been investigated by Prof. Freeman who, in a general way, confirms the reports of previous investigations and says:

. The probabilities of relationship with the ergot of L. temulentum are very interesting. The frequency of occurrence of ergots of Lolium in England is strangely coincident with that of the fungus in the grain, e. g., most abundant in I. temulentum, less so in L. perenne and exceedingly rare in L. italicum.

It is certainly not one of the rusts and the Ustilagineae are the closest affinity, perhaps, the fungus is carried from one generation to another by the sterile mycelium; when the embryo of the grain pushes out during germination, the hyphae, being in the "seed" keep pace with its growth and can be detected in the growing point throughout the life of the plant. Prof. Freeman says:

The hyphae sometimes penetrate the aleurone layer at any point and invade the starch endosperm. There exists in the nucellus, at the base of the scutellum and at the lower end of the inner groove of the grain, a layer of hyphae which lies directly against the embryo, constituting an infective layer.

· 11. Agropyron, Gaertn. Quack or Wheat Grass

Annual or perennial grasses, with flat, or involute leaves: spikelets 3-many-flowered, compressed, 2-ranked, alternate on opposite sides of the solitary, terminal spike, 1 at each joint, or, occasionally, all, or the lower in pairs, sessile, with the side against the axis; glumes transverse, nearly equal and opposite, lanceolate; flowering glumes rigid, rounded at the back, 5-7 nerved, pointed or awned from the tip; palet flattened, bristly, ciliate on the nerves, adherent to the grain.

About 40 species, in temperate regions. The root of quack grass is used in medicine; several species, like western wheat grass (Agropyron occidentale) and slender wheat grass (A. tenerum) are valuable for forage purposes. Quack and western wheat grasses are also good soil binders.

Agropyron repens, (L) Beauv. Quack Grass

Perennial, 1-3 ft. high, from a creeping, jointed rootstock; sheaths usually smooth, scabrous, or pubescent above; spikes 3-10 inches long, erect; spikelets 4-8 flowered; empty glumes strongly 5-7 nerved near the apex, awnless or sometimes short awned.

Distribution. Widely naturalized, a good forage plant and also a bad weed. In eastern North America, it occurs in cultivated fields and by roadsides and is a troublesome weed.



Fig. 154. Quackgrass (Agropyron repens). The "roots" (rootstocks) used in medicine. The roots contain considerable of a nutritious carbohydrate. (C. M. King.)

Medicinal properties. Quack grass is not known to be poisonous. The ancients since the time of Pliny have used the drug in medicine and it was also used by the Germans in the 10th century. The root stock is officinal. Gerard ascribed to the root diuretic, lithontriptic virtues or properties. The root contains considerable sugar and a substance called triticin, an amorphous, gummy substance easily transformed into sugar. It is found useful in the mucous discharge from the bladder. Quack grass and Western Wheat Grass frequently contain ergot.

12. Hordeum, L. Barley.

Annual or perennial grasses with flat leaves; cylindrical spikes; spikelets 1-flowered, with an awl-shaped rudiment on the inner side, 3 at each joint of the rachis of a terminal spike, the lateral ones usually imperfect or abortive and with a short stalk, empty glumes side by side in front of the spikelets, forming a kind of involucre; flowering glume and palet herbaceous, the former long and awned from the apex; stamens 3; styles very short; grain usually oblong and adherent to the palet; spike often separating into joints.

About 20 species widely distributed in both hemispheres. Of the Barleys, the 2-rowed barley (*Hordeum distichum*) and the 4-rowed barley (*H. vulgare*) are well known in cultivation, being used for malting purposes and occasionally in medicine. The awns of cultivated barleys produce mechanical injuries to stock. Several members of the genus are very troublesome weeds.

Four-Rowed Barley. Annual, 2-3 feet high, smooth; leaves linear-lanceolate, keeled, nearly smooth; sheaths striate, smooth, auricled at the throat; ligule very short; spikes 3-4 inches long, somewhat 4-sided; rachis flattened, pubescent on the margins; spikelets with 1 perfect floret; empty glumes, narrowly linear, pubescent, terminating in a slender awn; flowering glume 5-nerved, scabrous near the apex, long-awned; awn flattened, keeled, somewhat 3-nerved, serrulate on the margins.

The cereal is without doubt one of the most ancient of cultivated plants. It is supposed to have originated from *H. spontaneum* Koch, which grows wild in Asia Minor and Caucasian countries to Persia and Beloochistan as well as in Syria and Palestine.

Hordeum jubatum, I. Squirreltail Grass

An annual or winter annual from 6 inches to 2 feet high producing fibrous roots, forming solid and compact bunches; leaves not unlike those of blue grass, but paler in color, from 2-4 inches in length, margins scabrous; flowers in dense spike from 2-4 inches long, pale green or purplish in color, consisting of a number of 1-flowered spikelets, 3 occurring at each joint, 1 being perfect (bearing stamens and pistil), 2 others awl-shaped, and borne on short stalks, 1 sterile spikelet occurring on each side of the perfect flower which bears a long awn; at each joint will be found 6 empty, long-awned glumes, spreading at maturity which give to the plant its bristly appearance; when mature, the spike breaks up into joints consisting of the rudimentary spikelets and a perfect flower, so that each joint has 1 "seed," the number of seeds in the spike varying from 35 to 60. A single cluster of plants may therefore produce from 300 to 2000 mature seeds. The plant has a wonderful capacity for "stooling." From a single plant as many as forty spikes may be produced and the number often no doubt exceeds this.

Distribution. It is found in marshes, in moist sand along the sea shore, and near the northern lakes. Its present distribution is from Nova Scotia to New Brunswick, along the Atlantic coast, Maine to Maryland and westward to the region of the Great Lakes, Minnesota, Saskatchewan and the Mackenzie river, the Dakotas, Iowa, Nebraska and the Rocky Mountain region, south to Texas, California and southern Mexico. It is also reported from Europe and Siberia.

Originally it was chiefly distributed in the Rocky Mountain region occur-

ring in the saline soils of the plains, the great lakes and along the seacoast extending far northward. Its extension eastward and westward has taken place in the more recent times.

Hordeum secalinum, Schreb. Little Barley

An erect annual from 4-10 inches high, more or less geniculate at the lower end; sheaths smooth or upper often inflated; leaf blade 1-3 inches long; spikes narrow; empty glumes rigid, those of the central spikelet scarcely lanceolate, all awn pointed; flowering glume of the central spikelet awned or nearly so.

Distribution. Common and troublesome especially as far east as Missouri, Nebraska, British Columbia, and California.

Hordeum murinum, L. Wild Barley

An annual from 1-2 feet tall; erect or geniculate at the base; leaves rough; spikes from 2½-5 inches long; spikelets usually in 3's; scales awned, the enipty glumes awnlike and scabrous, the second scale of the lateral spikelets not ciliate, the flowering glumes scabrous at the apex, bearing an awn about 1½-1¼ inches long.

Common on the Pacific coast and the dry regions of Utah, New Mexico, Arizona, and occurring on ballast in the Eastern states.

Mechanical injuries. It has long been known that the barbed awns of barley, wild barley and other plants act injuriously in a mechanical way. In the west this is especially true of wild barley (Hordcum jubatum).

Dr. S. H. Johnson, of Carroll, states in the Carroll Herald, that this grass, when found in hay and allowed to ripen, if in any quantity, is very injurious to horses' mouths. He says:

The small awns seem to work in and cause deep ulcerating sores, which form under the tongue and lips. The writer has seen a large number affected and made a careful examination, and found the awns deep in the flesh, where they had remained for three months or more. I have seen lips eaten completely through and tongues caten almost off hy the grass. As to cattle, I have seen some affected, but not to any extent, because the mucuous membranes are much thicker. The sooner the grass is eradicated the better.

Professor Nelson, who has carefully studied this question, says on the injury to stock:

The awned heads, when taken into the mouth ,break up into numerous sections, scatter within the mouth and everywhere adhere to the mucous membrane, which soon becomes pierced with the long stiff awns. As the animal continues to feed, more awns are added, and those already present are pushed deeper into the flesh. Inflammation soon results and leaves the gums of the animal in condition to be more easily penetrated. The awns are particularly liable to be pushed down and alongside and between the teeth. As the swelling and festering progress the awns are packed in tighter and pushed deeper and cause suppuration of the gums as well as ulceration of the jaw bones and the teeth. Through the absorption of the ulcerated sockets and roots the teeth become loosened and even drop out, but the animal, impelled by hunger, still endeavors to eat such hay as may be offered.

The above statements apply largely to H, jubatum, but are equally true of all other species given above.

2. CYPERACEAE. Sedge Family

Grass-like, or rush-like herbs. Culms slender, solid or rarely hollow, frequently triangular, terete, quadrangular or flattened; roots fibrous and, frequently, creeping rhizomes, leaves narrow, sheathes closed; flowers perfect or imperfect,

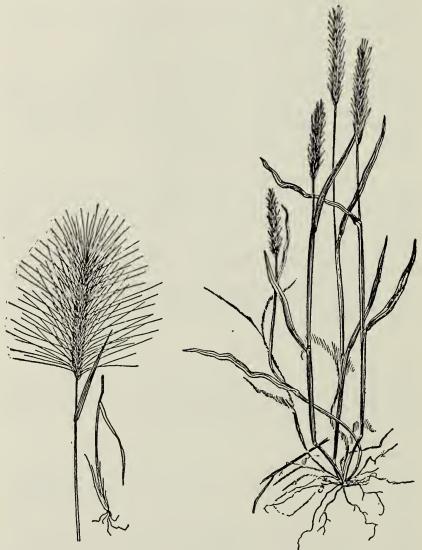


Fig. 155. Wild Barley (Hordeum jubatum.) Produces mechanical injuries to animals. (U. S. Dept. Agrl.)

Fig. 156. Common Little Barley (Hordeum secalinum). Causes mechanical injuries. (Charlotte M. King.)

arranged in spikelets, 1 or 2 in the axil of each glume, spikelets, 1- many-flowered; scales 2-ranked, or spirally imbricated, persistent, or deciduous; perianth free, composed of bristles, scales or rarely wanting; anthers 2-celled; ovary 1-celled, ovule 1, erect style 2 or 3-cleft; endosperms mealy; embryo minute. A large family of comparatively few genera (65) and 3,000 species of wide distribution. Carex is found in colder regions, while Cyperus is in warmer

regions. About 600 species of Cyperus, 200 of Scirpus, 200 of Rynchospora and 1,000 of Carex. The Papyrus (Cyperus Papyrus) of Africa and Sicily was used by the ancients as writing material. Common rush (Scirpus lacustris), a cosmopolitan plant found in water and marshes, is used for making mats and baskets. The rhizome of Carex arenaria is used in medicine.



Fig. 157. Sedge (Carex archaria). 1. Flowering plant. 2. Staminate flower with glume. 3. Pistillate flower. 4. Pistil. 5. Bract of pistillate flower. 6, 7. Staminate and pistillate flowers of C. hirta. (After Wossidlo.)

PRINCIPES

Woody or herbaceous plants with endogenous stems; flowers in spikes, generally on the plan of 3, free, regular or slightly irregular; stamens 3-9 or numerous but generally 6; ovaries, free, 1-7-celled usually; fruit dry or a

fleshy drupe. Contains the family Palmae, a large family of 1,000 species, of which the most important palms are as follows: The date palm (Phoenix dactylifera) of Asia and North Africa, now cultivated in warmer regions of Europe, California and Arizona, and an important article of commerce in North Africa; the Corypha which furnishes sago, fiber, and a seed which is used as a substitute for coffee; and the Washingtonia of Southern California. frequently cultivated. Vegetable wax is derived from Copernicia cerifera. The wine palms (Raphia vinifera and R. pedunculata of eastern Africa), furnish raphia fiber. The Metroxylon Rumphii of the South Sunda Islands furnishes sago. The betel-nut palm (Areca Catechu) is much used as a narcotic, the poison derived from this being known as arecain, half a grain of which is sufficient to kill a rabbit in a few minutes. It acts upon the heart and influences respiration causing tetanic convulsions; it also causes a contraction of the pupil of the eyc. It is used to some extent as a vermifuge and in India and the Islands of the Pacific it is applied as an external remedy. The nut contains the alkaloids, arecolin, arecain, arecaidin, and guvacin, which are used as vermifuges for dogs. The orange colored fruit is about the size of a hen's egg. When the nut is wrapped in quicklime and used, it imparts a red color to the saliva; it injures the teeth, and eventually destroys them. The resinous exudation from dragon's blood (Daemonorops Draco) of the East Indies is used in the manufacture of paints and varnishes. The oil from the oil palm (Elaeis guineensis) of West Africa and eastern South America is an important article of commerce. The cocoa-nut palm (Cocos nucifera) in tropical countries, especially the Islands of the Pacific, is an important article of food. The milk is the endosperm. The juice in the nut before maturity is unwholesome, being strongly diuretic and likely to cause serious results when taken into the system. A fermented drink is made from the juice of the plant which causes obesity and premature old age. A fiber known as ceir is made from the husks. Vegetable ivory (Phytelephas macrocarpa) of tropical countries, is a well known article of commerce. "Tuba" or Philippine toddy is made from the sap of the flowering spadix of Nipa fruticans. Toddy is also made from the juice of Arenga pinnatus, a plant which also furnishes an almost imperishable fiber. The "Royal Palm" is the "Yagua" (Roystonea borinquena) of Porto Rico, the sheathing bases of the leaves of which are used in thatching and siding the houses of the poor. An oil is produced from the husk and nut-like seeds of the Acrocomia or corozo palm which is distributed through tropical America from Mexico and Cuba to Paraguay.

SPATHIFLORAE

Mostly fleshy herbs with endogenous stems, or thalloid floating plants; flowers generally in a fleshy spadix subtended by a spathe or naked, or a few solitary flowers on the margin or back of the thalloid structure.

ARACEAE. Arum Family

Herbs with pungent juice; leaves with long, slender petioles and abounding in raphides; flowers borne in densely-flowered fleshy spadix, subtended or enclosed by the spathe; rootstock tuberous; floral envelopes none or of 4-6 sepais; stamens 4-10; filaments short; anthers 2-celled; ovary 1-several-celled; ovules 1-several in each cell; fruit a berry; seeds various, with 2 coats, the outer fleshy; endosperm abundant or none. About 900 species of wide distribution.

Many of the plants, as the skunk cabbage, Symplocarpus foetidus, possess acrid and noxious qualities. This is a native herb which is acrid and has a disagreeable odor. The fleshy spadix of Monstera deliciosa of the Mexican

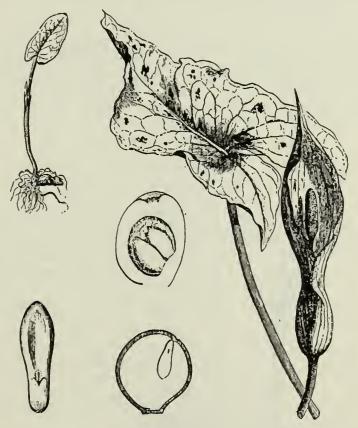


Fig. 158. Common European Arum, Cuckoo-pint, or Wake-robin (Arum maculatum). Leaf; spadix; longitudinal section of ovary; germination; longitudinal section of seed; embryo. (After Faguet.)

Cordilleras is edible. The vegetable calomel (Acorus Calamus) is used in medicine and contains the bitter principle acorin and an alkaloid. The sweet calomel is poisonous, under some conditions, causing disturbed digestion, and, in severe cases, gastro-enteritis, persistent constipation, followed by diarrhoea and passage of blood in the feces.

The Calla palustris, a marsh plant, has acrid properties and is used in Lapland with bread. The bulbs of Amorphophallus are rich in starch and are edible. The Richardia africana is frequently cultivated and used as food, a starch being also made from it. The poisonous substances contained in it are removed on roasting and boiling. In some of the fruits of aroids, like Arum italicum, saponin has been found, also needle-like crystals of oxalate of lime. A. maculatum is poisonous and causes severe dermatitis, paralysis, and, in the

case of children, even death. The *Thomsonia napalensis* of India, according to Major Kirtikar, is an acrid poison but its deleterious properties may be removed by roasting. The arrow arum (*Peltandra virginica*) of eastern North America is an irritant.

Arisaema. Mart.

Perennial herbs with tuberous rootstock or corm, having acrid properties; leaves simple or compound, scape simple; spathe convolute, generally arched above; spadix with flowers near the base; floral envelopes none; flowers monoecious or dioecious; stamens 4; anthers 2-4-celled; pistillate flowers with a 2-celled ovary containing many ovules; fruit a globose, red berry; seeds with copious endosperm. About 50 species found in temperate climate.

Arisaema triphyllum. (L.) Schott. Indian Turnip

Corm turnip-shaped, farinaceous; leaves generally 2, divided into 3-foliate



Fig. 159. Indian Turnip (Arisaema triphyllum). A common plant of our woods. Corm contains an acrid poison.

leaflets, ovate; spadix mostly dioecious, club-shaped, much shorter than the arched spathe, which is green and purple striped: ovules 5-6; berries shining, forming a dense head. The dragon head (A. Dracontium) with solitary leaf pedately divided into 7-11 oblong lanceolate leaflets, and spadix tapering to a long slender point, is common in rich woods from Minnesota and Iowa, eastward.

Distribution. The Indian Turnip occurs in moist woods from Kansas and Minnesota to Nova Scotia and Florida.

Poisonous properties. The corm of Indian Turnip is so extremely acrid that a decoction made from it has been used to kill insects.

The family Lemnaceae is allied to the Araceae. It contains the Duckweeds, (Lemna).

FARINOSAE

Herbs with endogenous stems and mostly narrow leaves; flowers usually complete, parts usually in 3's or 6's; corolla reg-

ular or nearly so; ovary compound, superior; endosperm of the seed mealy. This series contains the Xyridaceae, of which the yellow-cycl grass is an example; the Eriocaulaceae, of which the pipewort (Eriocaulon septangulare) of the Atlantic seashore is a good illustration; the pine-apple family (Bromeliaceae) of 350 species, in tropical and warmer regions, represented in the south by the Spanish moss (Tillandsia usneoides) which hangs in long festoons

from trees, and by the pine-apple (Ananas sativus), a well known fruit now cultivated extensively in Florida; from which has been isolated the enzyme, bromelin, a powerful ferment capable of rapidly digesting vegetable and animal albumen. It acts in the presence of either acid or alkaline carbonates and is related to trypsin and pepsin. In the same family is the pinguin (Bromelia Pinguin) or wild pine-apple, the slightly acrid pulp of which is edible and the fiber valuable. The plant is armed with stout spines which made the passage of troops difficult in the late Spanish war.

In the same order are the Spiderworts belonging to the family Commelinaceae. The common blue spiderwort (Tradescantia virginiana) of sandy and

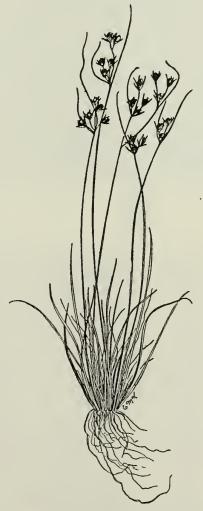


Fig. 161. Common Rush (Juncus temus). A weed with tough stems, along heaten paths and roadsides. (Charlotte M. King.)

gravelly soils, has mucilaginous stems, blue ephemeral flowers, and is common everywhere in eastern North America. Several species of *Tradescantia*, like the wandering Jew (*T. Zebrina*), are commonly cultivated. Another family of the order is the *Pontederiaceae*, containing the pickerel weed (*Pontederia cordata*) and the *Eichornia speciosa*, which is frequently cultivated in greenhoses and has become a very troublesome weed in the rivers of Florida and elsewhere in warm countries.

LILIIFLORAE

Herbs or occasionally shrubs with endogenous stems and monocotyledonous seeds; perianth generally well developed; flowers generally regular and complete, their parts in 3's and 6's; ovary superior, or inferior compound; endosperm horny or fleshy. This series contains the family Juncaceae, called

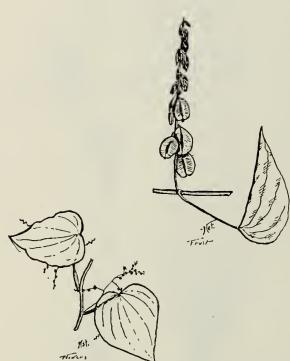


Fig. 160. Flowers, fruit and leaves of Yam (Dioscorea villosa). A common plant in thickets.

rushes, some of which, like wire-grass (Juncus tenuis), are troublesome weeds. Luzula is common at high altitudes and in northern states.

The Dioscoreaceae, or Yam Family, contains but few species in the United States. To this belong the wild vam root (Dioscorea villosa) of our woods, the Japanese yam (D. divaricata) and the air potato (D. bulbifera) of Asia, sometimes cultivated in the Gulf States for its large tubers. Yam starch is obtained from several species, the most important of which are D. alata, D. sativa. D. japonica, and D. aculeata.

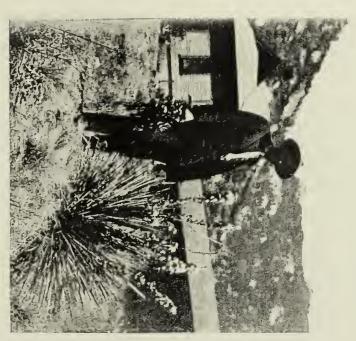
The family Taccaceae contains Tacca pinnatiafida, the roots of which are the source of the Tacca starch of Tahiti

and the neighboring islands. The plant is grown also in Brazil and India.

FAMILIES OF LILIIFLORAE

Ovary mostly superior.

Perianth segments distinct or partly united, the inner, petal-like; fruit a capsule or berry......Liliaceae



A. Yucca (Yucca angustifalia). An ornamental plant thought to contain a poisonous alkaloid. (L. H. Pammel),



B. Greasewood and Tetradymia. (L. H. Pammel).



Ovary inferior, at least in part.

I. LILIACEAE. Lily Family

Herbs or rarely woody plants with regular, symmetrical flowers; perianth not glumaceous; 3 sepals; 3 petals; 6 stamens; ovary 3-celled; fruit a pod or berry; embryo enclosed in the hard albumen. A family of about 1,600 species, including among others, several ornamental plants like the lily, lily of the valley, and yucca; some medicinal plants like squill, aloe, and false hellebore; and several poisonous plants like death camas and colchicum, the latter, native to Europe and Africa. The fatal poisonous nature of *Colchicum* was familiar to the ancients, it being known to contain several poisons, such as the alkaloid colchicin $C_{22}H_{25}NO_6$, an amorphous, yellowish white gum, chiefly an alkaline bitter substance, which, on boiling with acids, yields colchicein $C_{21}H_{23}NO_6$, and a yellowish green resin.

Animals that eat the plant suffer with acute gastro-enteritis, coma, staggering, weak pulse, and increased urination. The family also includes several economic plants like the onion (Allium Porrum); garlic (Allium sativum); chives (A. Schoenoprasum); shallot (A. ascalonicum); hyacinth (Hyacinthus orientalis); New Zealand flax (Phormium tenax), native to New Zealand where it occupies much of the country, and is now used in large quantities for making ropes and mats; (Yucca filamentosa) and (Y. augustifolia), the former a well known plant of the South and the latter a well known plant of the West, both species frequently cultivated for ornamental purposes, a large number of other species of the genus Yucca being also found in the Southwest. The day lily (Funkia subcordata), several species of the tulip (Tulipa), and several species of Lilium are cultivated. Perhaps the most common in the old gardens is the tiger lily, (L. tigrinum). Several species of the aloes are common in cultivation in greenhouses. They are also medicinal, containing the substance aloinum, a neutral principle, which yields barbatorin C17H20O. Aloes are cathartic. The California or Mariposa lily belongs to the genus Calochortus. The asparagus (Asparagus officinalis) is cultivated and is a well known vegetable. The cultivated smilax (Asparagus medeoloides) is a native to the Cape of Good Hope. The dragon-tree (Cordyline terminalis) is frequently cultivated. Some of the species of the latter like "Ti" of the Sandwich Islands are of economic importance. The roots of "Ti" contain a saccharine matter, from which the natives extract sugar; they also bake the roots and eat them. The remarkable dragon-tree of the Canaries is noted for its large circumference and comparatively low height. The Botany Bay resin (Xanthorrhoca hastilis) is chiefly used as a shellac for making colored varnishes.

Yucca leaves contain salicylic acid. Several investigators have reported saponin in the roots of Yucca filamentosa, Y. augustifolia, and Y. imperialis contain the alkaloid imperialin C₃₅H₆₀NO₄. In the former, Dr. Helen Abbot Michael reports the presence of several resins, the amount varying from 8-10 per cent in the root. She regarded the saponin as a constructive glucoside which served to unite what are known as the Saponin groups. Saponin occurs in many different plants, especially in the Sapotaceae.



Fig. 162. Colchicum (Colchicum autumnale). Flowering plant and longitudinal section of stem and bulb. Faguet.

Fig. 163. Camas plant or Death Camas (Zygadenus venenosus). A western plant, extremely poisonous to sheep.

The fly poison (Amianthium muscaetoxicum) is a smooth plant with simple stems from base; broadly linear leaves; white flowers in simple racemes; widely spreading perianth without claws or glands. Occurs from Long Island to Florida to Arkansas. It is a well known fly poison of the south. It is related to Veratrum and Melanthium.

The Bulbine bulbosa, of Australia, is poisonous to cattle, sheep and horses, which, after eating it, display such symptoms as lying down, rolling continually, having scours and a mucous discharge from the nose. The tuberous herb, Gloriosa superba, of India, according to Major Kirtikar, is a violent emetic;

the roots of this, when eaten, produce death in four hours. It is said to contain the bitter principle superbin ($C_{32}H_{60}N_2O_{17}$), perhaps identical with scillotoxin. The leaves and roots of Paris quadrifolia of Europe have a bitter taste. The berries are said to poison chickens and to produce gastro-enteritis in man. The Aloe succotrina contains from 4-10 per cent of a bitter principle aloin, also some emodin. Representatives of the genus Scilla and Urginea yield scillipicrin, scillin, and scillotoxin, the latter of which resembles digitoxin; the first of these acts upon the heart; where used as an emetic, it has proved fatal because of its irritant action on the intestines. The seeds of Sabadilla officinalis are used as a parasiticide. They contain cevadin $C_{32}H_{40}NO_{11}$, cevadillin $C_{34}H_{53}NO_8$, and are the principal source of veratrin $C_{32}H_{65}NO_{11}$, and the glucoside scillain or scillitin. According to Friedberger and Fröhner animals poisoned with "rat poison" (squill) had cerebral convulsions and crysipelas. The rhizome of Solomon's Scal (Polygonatum giganteum) has an acrid bitter taste.

Chickerinchee (Ornithogalum thyrsoides) is reported by Dr. Liautard (1) to have been the cause of acute gastro-cnteritis in horses in South Africa. The species O. Muscari may possibly be poisonous since it is allied to the above which an African veterinarian reports to be poisonous. The Star-of-Bethlehem (O. umbellatum) which is a pretty cultivated garden plant in the northern United States has become an escape in Kentucky and is regarded as a rather troublesome weed. The Tulip and Fritallaria are also poisonous.

GENERA OF LILIACEAE

| GENERA OF LILIACEAE |
|---|
| Perianth bell-shaped, gamophyllous. |
| Fruit a berry |
| Pcrianth cleft or divided. |
| Fruit a berry |
| Fruit a capsule. |
| With scarious bracts4. Allium |
| Without scarious bracts. |
| Roots bulbous |
| Roots not bulbous. |
| Sepals with claws, free from the ovary2. Melanthium |
| Sepals without claws |
| |

1. Zygadenus. Michx. Camas

Smooth, erect, perennial herbs from bulbs or rootstocks; leaves linear; greenish or white flowers in panicles; stamens free from perianth segments; capsule 3-lobed and 3-celled. A small genus of about 8 species, native to North America and Mexico.

Zygadenus venenosus. Wats. Death Camas

A pale green, slender perennial, 6 inches to $1\frac{1}{2}$ feet high, from small coated bulb; leaves rough, somewhat shorter than the stem; flowers borne in a raceme, yellowish or yellow, polygamous; segments of the perianth ovate or elliptical, free from ovary, bearing a roundish gland; capsule much larger than the perianth.

⁽¹⁾ Am. Vet. Rev. 30:298.

Distribution. From South Dakota to Nebraska, Colorado, Utah, California, Montana and British Columbia.

Zygadenus chloranthus. Pursh. Smooth Zygadenus

A glaucus perennial 1-3 feet high, coming from an elongated bulb; leaves flat; flowers borne in racemes, few flowered, greenish; segments of the perianth oval or obovate, united below and adnate to the base of the ovary; capsule longer than the perianth.

Distribution. Common especially northward in Iowa and Minnesota to Alaska, in the Rocky Mountains to New Mexico and east to Missouri. It may also be poisonous.

Poisonous properties. Mention of the poisonous nature of the various species of Zygadenus has frequently been made, especially by the early explorers, the poisonous bulbs encountered by them being referred to as poison camas or poison sago, so called to distinguish them from the edible Quamasa, which is commonly called kamas. These species bear essentially the same name today, except that in some places they are also called Lobelia. The bulbs are apparently much more poisonous than the leaves, but if the ground is very dry, sheep are less likely to pull them up than when the ground is moist. After rains, however, or early in the spring it is possible that some of the bulbs may be pulled up and thus eaten by sheep. In Montana, according to Chesnut and Wilcox, large numbers of sheep are killed by eating death camas. These authors state that in one band two thousand were poisoned and one hundred of these died. In another band two hundred were poisoned and ninety died.

Prof. Hillman reports that the wild sago (Z. paniculatus) is probably responsible for the death of a considerable number of cattle in certain alkaline districts in Nevada. Dr. S. B. Nelson, in experimenting with this species had wholly negative results. He fed one pound of the plant in blossom and fruit to sheep. Dr. Wilcox and Prof. Chesnut made tests on rabbits and sheep with extracts and fresh plants, and in every instance obtained positive evidence of poisoning. In these instances the plants were not in flower. Prof. Chesnut says stock is poisoned while pasturing by eating the bulbs along with the leaves or the leaves alone, or by the seeds when present in hay, as they sometimes are. Stock, especially sheep, are usually killed by eating the plant before it has blossomed in the spring. Cases of poisoning are so common in Oregon and Nevada that the term "lobeliaed" has been used to indicate the result from this kind of poisoning.

According to Chesnut and Wilcox the symptoms of poisoning are remarkably uniform:

The first signs of poisoning are a certain uneasiness and irregularity in the movements of the sheep. These irregularities rapidly become more and more pronounced, accompanied by incoordination of the muscular movements, spasms and rapid breathing. Although sheep are highly excited under the influence of Zygadenus poisoning, the cerebral symptoms seldom constitute a condition of frenzy. It was readily observed that until a few minutes before death ewes were able to recognize their lambs, and indicate in other ways that they were not in any sense crazed. The later symptoms were those of complete motor paralysis, combined with an exceedingly rapid and sharp breathing and a frequent weak pulse. The duration of these different stages of poisoning varies to a considerable extent, and depends entirely upon the amount of death Camas which the sheep had eaten.



Death Camas (Zygadenus venenosus). A poisonous plant of the western United States. (Nev. Agr. Exp. Sta.)



Postmortem examinations made show that in every instance the lungs were congested with blood, being a hepatized condition. There were no lesions in the membranes of the brain. In cases of adult sheep the effect upon the digestive organs was not marked. There were usually to be noticed an increased salivation and continued regurgitation through the mouth and nostrils. "Symptoms produced experimentally by feeding the death camas to sheep were the same as those characterizing natural poisoning by this plant.

The toxic substance has not been isolated. Chesnut and Wilcox observed that the ground material macerated with luke warm distilled water produced a substance that had a decided soapy feeling, and that the pure juice was distinctly irritating when left on the hands for several minutes. The physiological action of the Veratrum is somewhat similarly caused by the active poisonous principle in camas. It is probable that many of the Melanthaceae have similar properties. Dr. Wilcox recommends, in case of poisoning by death camas, the hypodermic injection of strychnin in 1/20, 1/10 and 1/5 grain doses, the hypodermic injection of atropin in 1/60 and 1/30 grain doses, and solutions of potassium permanganate and aluminum sulphate. From 5 to 10 grains of each of these compounds are dissolved in water and given as a drink to adult sheep. Hogs take the same doses as sheep, horses from 15 to 20 grains, and cattle from 30 to 50 grains. Occasionally the material is injected directly into the stomach, but ordinarily the more convenient method is to allow the animals to drink it. The substances veratalbin, sabadin and sabadinin have been obtained from Z. venenosus.

2. Melanthium, L. Bunchflower

Perennial tall leafy herbs with a thick rootstock; leaves linear to oblanceolate; flowers on large panicles, monoecious or polygamous, greenish yellow; perianth of 6 widely spreading segments raised on slender claws free from the ovary; stamens shorter than the perianth; pistil with 3 styles; capsule 3-lobed and 3-celled. A small genus of 4 species, in eastern North America.

Melanthium virginicum. L. Common Bunchflower

Tall leafy stemmed plants 3-5 feet high; leaves linear, the lower sheathing, the upper similar and sessile; flowers in an ample panicle, fragrant; perianth of flat segments, greenish yellow; styles persistent, capsule 3-celled; 8-10 seeds in each cavity.

Distribution. In low meadows and prairies from New England to Iowa river basin to Minnesota to Texas and Florida.

Poisoneus properties. Several correspondents in Iowa have attributed poisoning of horses to this plant. Several related plants of the Melanthaceae like Zygadenus and Veratrum are known to be poisonous.

Mr. J. R. Campbell, of Blockton, Iowa, writes us the following:

The specimens I sent you, and which you have identified as Melanthium virginicum, have been the reputed cause of a number of cases of poisoning here this summer. The veterinarian here pronounced it aconite poisoning as the symptoms are similar, but he decided this weed caused it as it has been found present in every case. In the first cases that he noticed here nearly all the horses in the livery barn were attacked after partaking of hay which contained an abundance of the matured seed pods of this plant. None of the animals died. The liveryman then had bis men pick out all the weed, and he has not been troubled since. Several cases have occured at different places since then, all traceable to this weed.



Fig. 164. Bunch flower (Melanthium virginicum). Common in low meadows Eastern Iowa and southward. Often mixed with hay and causes poisoning of horses. (Charlotte M. King.)

At the place where I obtained these specimens the owner said he had cut the meadow and fed hay off it for fifteen years and never had any trouble until this year. Hay cut last fall seems to contain the poison; seed heads were fully mature; meadow is low and wet.

The following are the symptoms as described by the veterinarian: Heart fast and very weak; respiration shallow and labored; great muscular weakness; retching, considerable slobbering, some sweating; temperature was normal. The effect lasted three or four hours, and the animal was stupid and lacked appetite for one or two days afterwards.

The disease stopped when a ration of hay containing none of this weed was fed. Since writing the above, Dr. Blanche, a veterinarian in Belle Plaine, this state, found that horses fed with hay containing this plant "became ill and acted as if they were crazy. The symptoms were much like those from aconite poisoning." These bunchflowers have long been used to poison flies, and Hyams, of North Carolina, says that they are poisonous to crows. The M. latifolium and M. pariflorum have similar properties. According to Chesnut the Indians of Mendocino County, California, use the soaproot or Yuki (Chlorogalum pomeridianum) to stupify fish. This plant is closely related to Melanthium.





Wild Indian Corn: Swamp Hellebore (Veratrum californicum). Reported to be poisonous to cattle and horses. (Bull. Nev. Agr. Exp. Sta. 51).

3. Veratrum (Tourn.) L.

Perennial herbs; leaves broad, clasping, veined, and plaited; flowers in large panicles, greenish, 'polygamous or monoecious; perianth in 6 parts, spreading, greenish or brownish, without glands or nearly so, and not clawed; stamens short and free, ovary with 3 persistent styles, capsule 3-lobed, 3-celled and several-seeded. A small genus of 10 or 11 species distributed in north temperate regions. One species used in medicine; all are poisonous.

Veratrum viride, Ait. American White Hellebore

A stout, leafy perennial from 2-7 feet high, with fleshy root, 1-3 inches long; flowers in ample, dense, spike-like racemes; it blooms from May to July.

Distribution. Common in swamps and wet woods, especially in eastern North America, west to Wisconsin, south to the mountains of Georgia, and north to Alaska.

Veratrum californicum, Durand. California Hellebore

A stout perennial from 2-8 feet high, fleshy root, flowers in a large loose terminal panicle; perianth segments whitish with long and narrow floral leaves.

Distribution. Common in the mountains of California and the Rocky Mountains as far north as British Columbia, south to New Mexico.

Poisonous Properties. Prof. Chesnut says:

Cases arise mainly from overdoses in medicine, but instances of accidental poisoning are reported for man and for various animals and birds. In one case all the members of a household were poisoned by eating the young leaves, which were mistaken for those of marsh marigold (Caltha palustris) and prepared for food. Animals do not relish the plant, which is acrid and burning in the fresh condition, but young aimals sometimes eat it with fatal results. The roots are not often mistaken for those of edible plants, but being fleshy and especially rich in alkaloids, they are somewhat dangerous. The seeds have been eaten by chickens with fatal results. The general effect is very much like that of aconite (Aconitum Napellus), being directed chiefly against the action of the heart and spinal cord, both of which tends to paralyze.

The symptoms of the poison are burning in the throat with increased salivation, producing a weak pulse, labored respiration and profound prostration. The root was used by the Indians in making snuff. Dr. Halsted attributed deaths of human beings as well as of cattle in New Jersey to this plant.

The number of poisonous substances found in hellebore is quite large. Of these so-called *veratrin*, $C_{32}H_{19}NO_{11}$, of earlier writers, has an alkaline reaction, and a burning taste; it produces violent sneezing and dilates the pupil. However, later investigators have separated this into the following bases: the very toxic *cevadin*, $C_{23}H_{49}$ -09, *veratridin*, $C_{37}H_{63}NO_{11}$ and *sabadillin* $C_{34}H_{63}NO_{8}$.

Veratrum album, L., V. lobelianum Bernh., V. viride, Ait., also contain in addition to the bases named above, two other bases, sabadin, $C_{29}H_{51}NO_{8}$, and sabadinin, $C_{27}H_{45}NO_{8}$, and also the following substances: jervin $C_{28}H_{37}NO_{3}$, a pure alkaloid rubijervin $C_{26}H_{43}NO_{2}$, pseudojervin $C_{29}H_{43}NO_{7}$, protoveratrin

C₃₂H₅₁NO₁₁, protoveratridin C₂₈H₄₅NO₈, and the bitter glucoside veratamarin. Jervin is a powerful depressant to the heart muscles and vaso motor centers; large doses therefore weaken the pulse. It depresses respiration and death occurs from asphyxia. Dr. Winslow, in speaking of the toxicology, says:

The symptoms exhibited in *Veratrum viride* poisoning are: salivation, vomiting, or attempts at vomiting, purging, ahdominal pain, muscular weakness, difficulty in progression, loss of power and general paralysis, muscular tremors and spasms, and, occasionally, convulsions. The pulse is unaltered in rate at first, but later becomes infrequent and compressible and finally rapid, threadlike and running. The respiration is shallow, the temperature is reduced, the skin is cold and clammy; there is semi-consciousness, loss of sight, and death from asphyxia. Treatment should he pursued with cardiac and respiratory stimulants, as amyl nitrite (hy inhalation), alcohol, strychnin and atropin; tannic acid as a chemical antidote; opium to subdue pain, and demulcents to relieve local irritation of the digestive tract. Warm water should be given the smaller animals to wash out the stomach and to asist vomition, and quietude should be enforced. In man, fatal poisoning is rare, since the drug is spontaneously vomited. The same would probably apply to dogs. Recovery has ensued in horses after injection of two ounces of veratrum album root.



Fig. 1642 American White Hellcbore (Veratrum viride). A poisonous plant of Eastern North America.

4. Allium (Tourn.) L. Onion, Garlic and Leek

Perennial bulbous plants, bulb solitary or clustered, leaves generally linear, a few lanceolate or oblong; stem simple or erect; flowers in umbels subtended by bracts; perianth white, purple or pink, the parts distinct or united at the base often becoming dry; the 6 filaments awl shaped, ovary 3-celled or incompletely so; capsule with 1-2 black seeds in each cell. About 275 species of wide distribution; contains a number of important economic plants, among them garlic (A. sativum), garden leek (A. Porrum), chives (A. Schoenoprasum), shallot (A. ascalonicum), onion (A. Cepa), and the golden garlic (A. Moly), cultivated for ornamental purposes.

Allium vineale, L. Field Garlic

A slender scape, naked from an ovoid membranaceous bulb; 1-3 feet high; terete and hollow leaves, channeled above, frequently densely bulbiferous; flowers greenish or purple.

Distribution. Common in meadows and wheat fields of Virginia, and eastern states from Connecticut to Virginia and Missouri. Naturalized from Europe.

Allium tricoccum Ait. Wild Leek

Scape naked, 4-12 inches high from an ovoid bulb; leaves fibrous articulated; leaves oblong, lanceolate or elliptical, few, appearing long before the flowers in spring; flowers in umbels numerous, greenish white, one ovule in each cavity; capsules strongly 3-lobed; seeds black and smooth.

Distribution. Common in the woods from western New England to Minnesota and Eastern and northern Iowa, especially in the low damp grounds.

Allium canadense L. Wild Garlic

Scape 1-2 feet high, coming from an ovoid bulb, the outer coats fibrous reticulated; leaves narrow linear; flowers in an umbel frequently with small bulbs; flowers pink or white.

Distribution. Common in meadows or low grounds in New England to Minnesota and Iowa, south to the Gulf.

Injurious Properties. In parts of the country where these onions grow there is frequent complaint of milk taking the flavor of onions where cattle feed upon them. Chesnut and Wilcox do not mention any species of the genus Allium, except some of the species found in Montana, which may impart to milk a disagreeable flavor. Friedberger and Fröhner state that onions produce slavering.

Prof. A. Liautard * has prepared an abstract of a report by Dr. W. W. Goldsmith in the Journal of Comparative Pathology and Therapeutics upon onion poisoning in cattle. Briefly it is as follows:

Loads of onions partly started to shoot and partly decayed, were unloaded in a meadow where nine head of cattle were grazing. After a week the cattle seemed sick

^{*} Amer. Vet. Review. 36:63

and one died, displaying the following symptoms: Intense onion odor, tucking up of flanks; constipation in some; purging freely in others; one vomited abundantly; another very ill, grunted, was much constipated, staggered in walking, was very tender in loins, temperature 103°, urine dark and smelling of onions. Treatment: Feeding with soft food and hay. Large doses of linseed oil. One animal that was very ill got also extract of belladonna and carbonate of soda. All but one of the animals recovered. At the autopsy of the dead one, the rumen was found inflated and also the bowels. Liver enlarged and of light color. Kidneys dark green and with offensive edor. Rumen contained large quantity of onions and grass. The whole carcass and organs smell of onions.

5. Convallaria L. Lily of the Valley

A low smooth herb with horizontal root-stocks; flowers white in a one sided raceme; stamen 6; ovary 3-celled; berry globose. A genus with one species.

Convallaria majalis, L. Lily of the Valley

A smooth perennial herb with horizontal root-stocks and 2 or sometimes 3 oblong leaves; flowers in racemes; perianth bell shaped, white, 6-lobed, stamens 6, inserted on the base of the perianth; ovary 3-celled, 4-6 ovules in each cell;



Fig. 165. Lily-of-the-Valley (Convallaria majalis). A well known cultivated plant possessing poisonous properties similar to those of Foxglove. U. S. Dept. Agr.



Fig. 166. False Aloe (Agave virginica). A plant of the Southern United States.

berry roundish, red and few-seeded. The species is native to Europe, Asia, and the Alleghanies, and is widely cultivated as an ornamental plant.

Poisonous properties. The plant contains two glucosides; one, convallamarin $C_{23}H_{41}O_{12}$, an extremely poisonous crystalline compound with a bitter swect taste, with a physiological action on the heart like digitalin, a substance found in the common foxglove, and convallarin $C_{34}H_{31}O_{11}$, which is crystalline, has a sharp taste and is purgative in its action. Chesnut says: "The amegative and purgative actions of the lily of the valley are quite marked." The action of the heart is infrequent and irregular, and finally death occurs from paralysis.

Trillium L. Birthwort

Herbs, with naked stem from a short, horizontal root stock, netted veined, simply whorled leaves, in 1 or 2 whorls; colored flowers, 3 green persistent sepals; 3 colored petals which wither with fruit; stamens 6, hypogynous; linear, adnate anthers on short filaments; sessile stigmas 3; ovary 3-celled; fruit a berry.

The principal species of the United States are: the wake-robin (T. nivale), which flowers very early in the spring, is from 2-4 inches high and is common northward and eastward; the sessile-flowered wake-robin (T. sessile) which bears sessile dull purple flowers with narrow sepals and petals, and leaves that are often blotched and occurs from eastern Iowa southward; prairie wake-robin (T. recurvatum) of the west, which has dull purple petals but differs from the preceding in having narrow leaves; large white-flowered wake-robin (T. grandiflorum) which bears a large white flower raised on a peduncle later recurving from the erect, the flowers becoming purplish, and rounded, ovate, sessile leaves; and birthwort (T. erectum) much like T. grandiflorum except that the flowers are not so large and are unpleasantly scented. Both of the two last named are found in the eastern and central states.

Poisonous properties. Trilliums have long been considered poisonous. All species are emetic. Lindley states that the roots have a violent emetic action. The fruit should be regarded with suspicion.

HAEMODORACEAE. Bloodwort Family

Prerennial herbs with fiborous roots; leaves, narrow, lanceolate and somewhat erect; small perfect flowers which are woolly or scurfy on the outside; flowers in panicles; perianth 6-parted or 6-lobed adnate to the ovary; stamens 3, opposite the 3 inner segments of the perianth; stigmas 3; fruit a 3-valved capsule, seeds few or numerous. A small family of 9 genera and 35 species mostly native to Africa, Australia and tropic America.

Lachnanthes. L. Red-Root

A stout herb with short rootstock; red, fibrous, perennial root; leaves, equitant and sword shaped, crossed at the base and scattered on the stem; flowers, numerous, borne in a woolly, cymose paniele; perianth, 6-parted, the outside segments smaller than the inner; stamens, 3, opposite the 3 inner divisions; pistil with 3-celled ovary few ovules in each cavity; seeds few, flattened nearly orbicular, fixed by the middle. A species of a single genus native to southeastern North America and western India. (Gyrotheca).

Lachnanthes tinctoria, (Walt.) Ellis.

A stout, tall herb with numerous yellow flowers, 6-parted perianth and few seeds.

Distribution. In salty swamps near the coast in southeastern Massachusetts, Rhode Island and New Jersey to Florida. This plant is commonly called the pink-root of the Atlantic coast.

Poisonous properties. Prof. Chesnut says that throughout the South, white hogs are supposed to be particlaraly subject to the poison contained in this plant. Dr. Halsted says "Throughout the southern states, this plant abounds and the preponderance of black over white-skinned hogs is claimed to be due to this paint-root. White hogs with free access to the plant are soon killed off, while black ones are not.

This is not the only case of the color of animals seeming to have an influence upon their distribution. Thus, white horses in Prussia, it is claimed, are injured by eating milkweed, while dark horses are not. In Sicily, there are black sheep, only, as white ones are killed off by a species of St. John's wort (Hypericum)." While the claim of the immunity of black pigs from the effects of paint-root seems to be a common belief, further investigation should be made before this should be assured definitely as a fact.

Family Amaryllidaceae. Amaryllis Family

Mostly perennial herbs with bulbs, rootstocks or corms; scapose flowers regular or nearly so; perianth 6-parted or 6-lobed, the lobes or segments distinct, united below into a tube, adnate to the ovary; stamens 6; style single; capsules several, many seeded. About 800 species, chiefly native of tropical or warm regions, a few in temperate regions. Some well known representatives are daffodil, (Narcissus Pseudo-Narcissus); Polyanthus, (N. Tazetta); poet's Narcissus, (N. poeticus) producing intense gastro-enteritis; Jonquil, (N. Jonquilla); snowdrop, (Galanthus nivalis); amaryllis, (Amaryllis Belladonna); tuberrose, (Polianthes tuberosa), the latter widely cultivated; the American aloe or agave, the most common species in cultivation being the century plant (Agave americana) native to Mexico and Central America, the Mexican drink, pulque, being made from the sweet liquid obtained from this plant at the time of flowering. Several species are used for the manufacture of fibre, the best known being the sisal, (Agave rigida). The mauritius hemp, (Furcraea gigantea), is native to Mexico and has been introduced into Zanzibar. Many members of the family have acrid properties and some of them are poisonous. Buphane disticha is used by the Hottentots to poison their arrows. Poet's narcissus contains pseudo-narcissin; Amaryllis Belladonna contains belladonin; and Sprekelia formossissima contains amaryllin, a belladonna-like alkaloid. The Lycoris species contain lycorin, an alkaloid with the formula C32H32N2O8, and a second alkaloid kisanin, C₈₄H₃₆N₂O₂. Agave heteracantha contains agavesaponin. Dr. MacDougal states that the sharp pointed leaves of Agave Schottii often penetrate leggins and leather shoes inflicting painful injuries.

Zephyranthes. Herb

Smooth herb with coated bulbs; narrow leaves; flowers scapose, large erect, pink, white or purple; perianth funnel-form from a tubular base; the 6 divided petals are united below into a tube subtended by an entire or 2-cleft



Fig. 167. Saffron (Crocus sativus). The flowers furnish the saffron of commerce, (Faguet).

bract; ovary 3-celled; style long, filiform, 2-cleft at the summit; ovules numerous; capsules membranceous; seeds flattened, blackish; small genus of 30 species, native to America.

Zephyranthes Atamasco (L.) Herb. Atamasco Lily

Leaves bright green and shiny from an ovoid bulb; scapes erect; bracts 2-cleft; perianth white, pinkish or light purple; segments shorter than the two stamens.

Distribution. In moist places from eastern Virginia to Florida and Alabama.

Poisonous properties. Prof. Chesnut, in speaking of this plant, says that the Atamasco of the southeastern United States is supposed by some persons to cause the disease in horses known as "staggers."



Fig. 169. Blue Flag (Iris versicolor). The rootstock poisonous. The plant grows in low grounds. (After Johnson.)

Family IRIDACEAE. Iris Family

Perennial herbs, frequently with bulbs, corms or tubers; leaves equitant, erect, 2-ranked; perianth of 6 segments or 6-lobed, its tube adnate to the ovary; stamens 3, adnate to the ovary; anthers facing outward; ovary inferior, mostly 3-celled; style 1 or 3-cleft, stigmas 3, opposite the three stamens; ovules generally numerous in each cell; embryo small; endosperm, fleshy, or horny.

About 1000 species, of wide distribution. Common native plants of the family are the blue flag (*Iris versicolor*), growing in low grounds of the North;

the Carolina blue flag (I. carolina) of the South, blue eyed grass (Sisyrinchium angustifolium). Many species of the exotic blue flags, Iris like the dwarf garden iris (I. pumila) and the common flower-de-luce (I. germanica), the common crocus or saffron (Crocus vernus) used for coloring, freesia (Freesia refracta), tritonia and gladiolus are cultivated for ornamental purposes. The orris root (Iris florentina, I. pallida and I. germanica) is an article of commerce used for perfume and tooth powders. It contains myristic acid. A substitute for saffron is obtained from the flowers of the South American saffron (Crocosmia aurea).



Fig. 169a. The petaloid bilobed stigma and stamen of Iris. (Kerner).

Iris (Tourn) L.

Herbs with creeping or horizontal root stocks, and erect stems with equitant leaves; flowers large, regular, panicled; perianth of 6 segments united below into a tube, the outer dilated, spreading or reflexed; the 3 inner, smaller; stamens inserted at the base of the outer perianth; ovary 3-celled; fruit a capsule; seeds numerous. About 100 species in the North Temperate regions. The Iris florentina contains the glucoside irigenin $C_{18}H_{16}O_8$. This is derived from irisin.

Iris versicolor, L.

Root stock fleshy; stem roundish; leaves erect, leaves shorter than the stem; flowers bluish, perianth deeply 6-parted, the 3 outer divisions reflexed, the 3 inner smaller, erect; stamens distinct, covered by the petaloid stigmas.

Distribution. In marshes, thickets, and wet meadows from Newfoundland to Manitoba, south to Florida and Arkansas.

Poisonous properties and uses. The root contains the substance irisin, or iridin. The acrid resinous substance, irisin, acts powerfully upon the gastro-intestinal tract, liver and pancreas, causing a burning sensation and congestion. That the root is poisonous may be seen from the following statement made by Dr. Rusby:

Another rhizome whose acrid taste is likely to prevent ingestion in poisonous quantity, is that of the common *Iris versicolor*, L. Still, because this is commonly known as the blue flag, there is some danger that it might be eaten in mistake for calamus, which is commonly known as sweet-flag. If so, it would prove seriously, if not fatally poisonous

as its well-known emetic-cathartic properties, even when toned by drying and keeping, are powerful, and in a fresh state would be decidedly violent.

Dr. Johnson says:

Iris, in full doses, is an active emeto-cathartic, operating with violence, and producing considerable prostration. Its effects upon the liver appear to be analogous to those of podophyllum. In sick headache dependent upon indigestion, small doses, frequently repeated, often act most happily. It has been largely used by eclectic practitioners, and is highly esteemed by them as a hydragogue cathartic, an alterative, sialagogue, vermifuge, and diuretic.

One case of poisoning has been recorded in this state. Other species of Iris of which we have quite a number in the U. S. must be looked upon with suspicion. Mention may be made here of the Iris missouriensis and I. carolina. The root stocks of our cultivated species like I. pumali and I. sibirica must be looked upon with suspicion. The South African Homeria collina naturalized in Australia, according to Maiden, is poisonous to cattle browsing on the plant.

SCITAMINEAE.

Large herbs with endogenous stems and monocotyledonous seeds; flowers very irregular; ovary inferior, composed of several united carpels; seeds

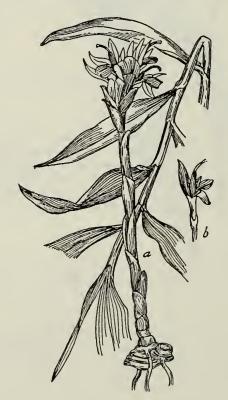


Fig. 170. Ginger (Zingiber officinale).
a. Entire plant. b. flower. (Charlotte M. King, after Strasburger, Schenck, Noll and Schimper.)

with endosperm. This order contains the important family Musaceae in which is found the banana (Musa sapientum), well known as an article of commerce. It is extensively cultivated in the tropics and one of the most important food plants in all warm countries. The fruit is eaten fresh when ripe; a kind of flour is also made from it. M. textilis is an important fiber plant being the source of Manilla hemp, large quantities of which are imported from the Philippines. The ravenala or traveler's palm also belongs to this family. It has an oily, edible, arillus which is bright blue. The family Cannaceae contains the Indian shot (Canna indica), frequently cultivated for ornamental purposes in this country; in tropical regions, however, a starch is made from the rhizome of this species and from C. edulis. C. flaccida is a native of the southern United States and has a pretty blue flower. The family Marantaceae contains the West Indian arrowroot (Maranta arundiacea). The family Zingiberaceae includes ginger (Zingiber afficinale) which contains gingerol and is used as a condiment and stimulant. The ginger of commerce is derived from the fleshy rootstock, the plants grown in Jamaica being considered most valuable. These are cultivated in regions having an altitude of 2000 Malabar cardamon (Elettaria Cardamomum) round cardamon (Alpinia striata), bastard cardamon (Amamum xanthioides), Bengal cardamon (A. subulatum) and Java cardamon (A. maximum) also belong to this order. The Kaempferia rotunda of India, is a bulbous or tuberous rooted biennial which according to Major Kirkitar, causes profuse salivation and vomiting when administered internally. The rhizome of K. Galanga furnishes a perfume.

Arrowroot comes from Curcuma leucarhiza, and turmeric from Curcuma langa. The tuber of the latter, when powdered, is used as a yellow dye-



Fig. 171. Canna (Canna flaccida). A native American Canna.

Fig. 172. Banana Fruit (Musa sapientum). A well known tropical fruit. W. S. Dudgeon.

stuff, in making turmeric paper, as a condiment, especially in curry powder, and as an aromatic stimulant. The zedoary, (C. Zedoraia), is used in Himala-yan India, where it is native, in place of turmeric. Galangal is the root of Alpinia officinarum which grows on the Chinese coast. Another species, A. Galanga is used on the island of Java.

MICROSPERMAE

Herbs with endogenous stems, flowers very irregular or in a few cases regular, generally complete and perfect, and parts in 3's or 6's; ovary inferior compound; seeds small, numerous, without endosperm.

Family Orchidaceae. Orchid Family

Perennial herbs with corms, bulbs or tuberous roots; perfect and irregular flowers; perianth of 6 divisions in 2 sets, the 3 outer similar in texture to the 3 inner petals, one of the 3 inner, different in form and is called the lip; in front of the lip is a column composed of a single stamen, or in *Cypripedium* of two stamens, and a rudiment of the third; pollen in 2 or 8 pear shaped sacs called pollinia which are united by little threads. Stamens variously united with the thick, fleshy style into a column; ovary 1-celled with many ovules on a three parietal placentae; capsule 1-celled, 3-valved, seeds numerous. A large order of about 5000 species of wide distribution, most abundant in the tropics. Many of the plants like the *Cypripedium*, *Angrecum* and the *Catasetum* are cultivated for ornamental purposes.

The salep of commerce is obtained from the Orchis masculata. The flavoring material, vanilla, is obtained from Vanilla planifolia, native to Mexico and widely distributed by cultivation; this plant contains from $1\frac{1}{2}$ to 3 percent of vanillin $C_8H_8O_3$. Other species of the genus Vanilla also furnish vanilla but in smaller quantities; these are V. Pompona, V. guianensis, and V. palmarum. Vanillin is also made from coniferin and eugenol, and occurs in other orchids as Spiranthes and such plants as Spiraea Ulmaria and Lupinus albus. It is used for medicine.

Orchids contain some alkaloids; for example, *Phalanopsis amabilis* contains a tonic alkaloid, according to Boorsnis, which is closely related to *coniferiu*, $C_{18}H_{22}O_8$.

Cypripedium L.

Tufted roots; pcrennial, glandular, pubescent herbs; leaves large, many nerved; flowers solitary or few; sepals shiny, sprcading, 3 distinct or 2 of them united into one, under the lip; petals spreading, resembling the sepals; lip of large inflated sac, column declined with a fertile stamen on each side; a sterile petaloid stamen above, which covers the summit of the style; pollen granular, stigma broad, obscurely 3-lobed, moist and roughish. About 40 species, mostly tropical.

Crypridium parviflorum, Salisb var. pubescens (Willd) Knight. Yellow Lady Slipper

Perennial, with leafy stem, 2 feet high, pubescent; leaves oval, or elliptical, acute; sepals ovate, lanceolate, usually larger than the lip, yellowish or greenish; petals narrower, usually twisted; lip flattened laterally, pale yellow with purple lines.

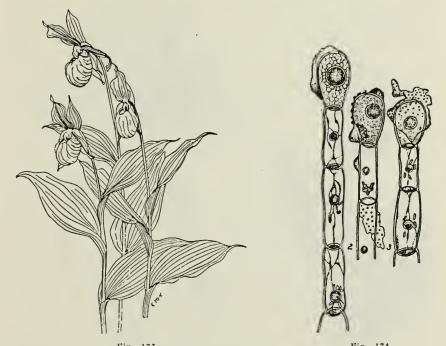


Fig. 173. Fig. 174.

Fig. 173. Smaller Yellow Lady Slipper (Cypripedium parviflorum, var pubesceus). A beautiful flower of early summer, seen in the woods of eastern Iowa. C. M. King.

Fig. 174. Glands of several species of Moccasin flower, which are said to contain the toxic substances. 1. Hair gland of Cypripedium pubescens. 2. Hair gland of Cypripedium hirsutum in water. 3. Hair gland of Cypripedium Calceolus in water. (Charlotte M. King, after Nestler.)

Distribution. In woods and thickets, chiefly east of central Iowa, and Minnesota to Nova Scotia; occasionally in Colorado, Nebraska and Alabama.

Cypripedium candidum Muhl. Small White Lady Slipper

A slightly pubescent perennial; leaves lance-oblong, acute; petals and sepals greenish, purple spotted; sepals ovate-lanceolate, lips white striped with purple inside, flattened laterally, convex above.

Distribution. In bogs and meadows from New York to Minnesota, Iowa, Nebraska and Missouri.

Cypripedium hirsutum Mill. Showy Lady Slipper

A rather stout, downy perennial 2 feet or more high; leaves ovate pointed; sepals round ovate, or orbicular, longer than the petals, which are obovate; lip inflated, white, pink purple stripes.

Distribution. In woods and swamps from Nova Scotia, Ontario and Georgia west to Minnesota and Iowa.

Poisonous Properties. Dr. Babcock, many years ago, found that the several species of Lady's Slipper produced dermatitis. Years ago the writer heard of a case of poisoning where a young man carried a large bunch of



Fig. 176. Flower of another orchid (Habenaria).

Showy Lady Slipper and became poisoned very much as if it were by poison Ivy. Prof. Chesnut in referring to the poisoning from these plants says:

The poisonous character of these plants was not suspected prior to 1875, when Prof. II. H. Babcock, of Chicago, who had annually been suffering, supposedly from recurrent attacks of ivy (Rhus) poisoning, discovered that the affection was most probably caused not by the ivy, but by the two species of Lady's Stipper named above (C. parviflorum, var. pubescens and C. hirsulum) instances were afterward reported, but the facts were not positively ascertained until 1894, when an investigation was made by Prof. D. T. MacDougal of the University of Minnesota. It was discovered that these plants are provided with glandular hairs which cover the surface of the stem and leaves and contain a poisonous oil which is especially abundant at the fruiting season. Its action on the skin is very similar to that of toxicodendrol, the active constituent of poison Ivy (Rhus Toxicodendron), but its exact chemical nature could not be ascertained on account of the small quantity obtainable. Experiments with the stem and leaves upon individuals showed that over half of them were affected by the first two species, and that the last was also poisonous, but in a minor degree. No accidental cases have been recorded against it. No specific antidote has been suggested.

Dr. MacDougal * made a personal experiment with a mature specimen of C. hirsutum on which there were newly formed seed pods. This plant was broken off near the base of the stem and the leaves brushed lightly over the arm.

A slight tingling sensation was felt at the time, and, fourteen hours later, the arm was greatly swollen from the shoulder to the finger tips. The portion covered by the plant - covering an area of 50 sq. cm. - was violently inflamed and covered with macules, accompanied by the usual symptoms of dermatitis and constitutional disturbances. By treatment of the most approved kind, the arm was reduced to its normal size in ten days, but the effects were perceptible a month later.

Nestler discovered that the secretion contained in these hairs was a fatty acid readily soluble in alcohol and benzol and producing a mildly acid reaction. He also states that his results with C. pubescens were negative but that with C. spectabile (C. hirsutum Mill.) he secured positive results, producing a dermatitis, the action, however, not being so pronounced as that reported by Mac-Dougal. He also found that, as stated above, the maximum poisonous effect was during the formation of seed capsules and that the poison was in the hairs of the plant as is the case in the Primrose. Nestler did not succeed in producing dermatitis with C. parviflorum, C: acaule, C. macranthum, C. montanum, or Calceolus. As some of these species produce an abundance of raphides in the stem, it is evident that dermatitis is not caused by these crystals, but rather by a substance found in the stem. Dr.MacDougal suggests that the raphides may serve the plant as a protection from animals.

Nestler also asserts that the Cypripedium may contain an additional substance myelin which Senf has found in Ginkgo seed, and Nestler 1 himself observed in the fruit of Capsicum annuum. It is not a cardol.

From the root of Cypripedium a substance is obtained which is sometimes administered to children as a substitute for opium. It contains a bitter glucosidal principle.

Class, DICOTYLEDONEAE

Stem usually oxogenous with pith, wood and bark (endogenous in a few plants); the woods traversed by medullary rays; leaves usually pinnately or palmately netted-veined; embryo of the seed with 2 cotyledons or occasionally 1; parts of the flower usually in 5's, rarely in 3's or 6's.

Archichlamydeae

Petals separate and distinct from each other or wanting. Includes many plants classed as Apetalae and Polypetalae. In some orders, as Leguminosae, the lower petals are more or less united and joined at the base.

VERTICILLATAE

Contains a single family Casuarinaceae of 20 species, mostly Australian, with monoecious flowers. The Casuarina equisetifolia of the tropical Old World furnishes a hard wood known as iron wood and in Egypt the trees are used as a shelter belt for bananas.

^{*} Minn. Bot. Studies. 1894:32-36. 1 Das Sekret der Drusenhaare der gattung Cypripedium mit besonderer Berücksichtgung seiner hautreizenden Wirkung. Nestler. Ber. der Dent. Bot. Gesell 25:554-567.

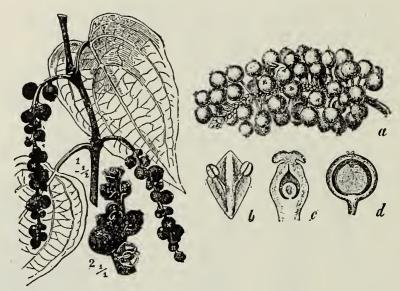


Fig. 177. Black Pepper Plant (Piper-nigrum). 1. Part of shoot with young fruit. 2. Tip of fruit spike. (After Wossidlo.)

PIPERALES

Herbs with exogenous stem, with neither petals nor sepals; flowers in spikes, bracteolate. Largely tropical and includes the family Saururaceae or Lizard's tail; the peppers, Piperaceae, including black pepper (Piper nigrum) a well known condiment of the tropics containing the alkaloid piperin C₁₇H₁₀NO₃ and a volatile oil C₁₀H₁₆, cubebs (P. Cubeba) containing cubebin C₁₀H₁₀O₃ and the oil of cubebs, kava-kava (P. methysticum) native to the Pacific Islands, containing methysticin C₁₅H₁₄O₆, which is used to make stimulating drinks, P. longum of India, P. chaba of India and the Philippines, the Betcl Pepper (P. Betle) of the Malay Islands, the berries of which are chewed with the Betel Nut, and the Matico, or the Soldier's Herb (P. angustifolium) of South America, the hairy leaves of which are used as a styptic. The South American Peperomias are well known greenhouse plants. Other species of peppers are used in medicine. The so-called "caisimon" (P. peltutum), according to Mr. Combs, is a powerful diuretic. "Matico de Peru" (P. angustifolium) is an acrid, bitter plant containing a green volatile oil.

SALICALES

Trees or shrubs with simple flowers, imperfect catkins; perianth wanting; fruit a many-seeded capsule; seeds with a tuft of hair at one end. This series contains only one family the Salicaceae.

Salicaceae. Willow Family

Dioecious trees or shrubs, alternate stipulate leaves, the stipules often minute and soon falling; staminate and pistillate flowers borne in catkins, one to each bract, without calyx or corolla; staminate flowers with 1-numerous



Fig. 178. Peach-leaved Willow (Salir amygdaloides). 1. Flowering branch of staminate tree. 2. Same of pistillate tree. 3. Staminate flower, with scale, enlarged. 4. Pistillate flower enlarged. 5. Fruiting branch. 6. Summer branch. 7. Bud and leaf scar. 1, 2, 5, 6, one-half natural size. M. M. Cheney.

stamens, subtended by a cup-shaped disk; pistillate flowers with a 1-celled ovary, stigmas 2-4, simple or 2-4-cleft; fruit a 1-celled and 2-4 valved pod bearing numerous seeds provided with long silky hairs. There are only two genera and about 200 species, found in temperate and Arctic regions. The bark of some species of the family is used in medicine because of its astringent properties. The willow contains the glucoside salicin $C_{13}H_{18}O_{9}$. Poplar contains populin $C_{20}H_{22}O_{8}$. The Balm of Gilead (Populus candicans) may cause blistering, and the European P. balsamifera causes colic.

Myricaceae. Sweet Gale Family

Monoecious or dioecious shrubs with alternate, coriaceous, aromatic leaves; flowers in short scaly catkins; staminate flowers with 2-16 but usually 4-8' stamens; ovary with 2-8 scales and 2 linear stigmas; fruit a small 1-celled

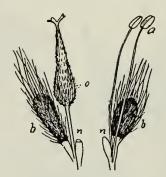


Fig. 179. Pistillate and staminate flower of willow. n. nectar gland. b. scale.

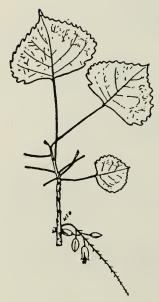


Fig. 180. Leaves of Cottonwood (Populus deltoides). A well known native tree growing on the borders of streams throughout the U. S. east of the Rockies. (W. S. Dudgeon.)



Fig. 180a. Sweet Gale (Nmrica). Common in the east.

drupe, the outer part frequently covered with wax. About 35 species of wide distribution. The sweet fern (Myrica asplenifolia) is sometimes weedy in sandy fields in the North; it contains an oil of strong, spicy, cinnamon-like odor; bayberry wax is derived from M. cerifera, common along the Atlantic coast and the Gulf of Mexico. The bark of Myrica Nagi contains myricetin $C_{18}H_{10}O_8$. The leaves of M. acris are used in the preparation of bay rum.

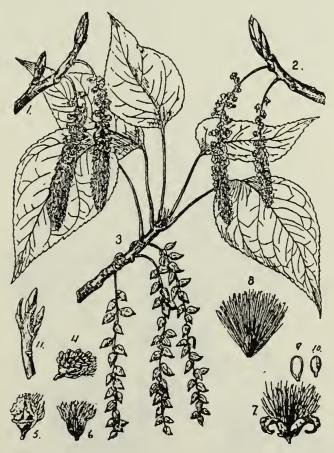


Fig. 181. Balm of Gilead (Populus candicans). 1. Flowering branch of staminate tree. 2. Same of pistillate tree. 3. Fruiting branch. 4. Scale of staminate catkin, enlarged. 5. Scale of pistillate catkin, enlarged. 6. Scale without flower displayed, enlarged. 7. Mature fruit. 8. Seed, enlarged. 9. Longitudinal section of seed, enlarged. 10. Embryo, enlarged. 11. Winter branch, showing buds. 1, 2, 3, 11, one-half natural sizc. M. M. Cheney.

BALANOPSIDALES

Contain a single family, the *Balanopsidaceae* of New Caledonia, which in turn contains a single genus *Balanops* of 7 species.

LEITNERIALES

Shrubs or trees with entire, petioled, simple leaves; flowers in catkins; staminate flowers subtended by what appears to be a perianth; sepals 3-4; ovary 1-celled; style slender; endosperm thin. Only one family, the Leitneriaceae, which consists of a single genus Leitneria, with perhaps 2 species. L. floridana occurs in swamps in southern Missouri to Texas and Florida and produces a wood lighter than cork, probably the lightest wood known.

JUGLANDALES

Trees with alternate, pinnately-compound leaves; flowers monoecious, bracteolate; the staminate in long drooping catkins; pistillate solitary or several together; staminate flowers of 3-many stamens with or without a perianth; peri-



Fig. 182. Black Walnut (Juglans nigra). 1. Flowering branch. 2. Staminate tlower before authesis, enarged. 3. Staminate flower, enlarged. 4. Perianth of staminate flower, enlarged. 5. Stamen, enlarged. 6. Pistillate flower, natural size. 7. Longitudinal section of pistillate flower, natural size. 8. Leaf, reduced. 9. Winter branchlet. 10. Mature fruit. 11. Walnut with husk removed. 1, 9, 10, 11, one-half natural size. M. M. Cheney.

anth adnate to the ovary; anthers erect, 2-celled; pistillate flowers usually 2-bracteolate, calyx 4-lobed or with petals; ovary inferior, 1-celled or incompletely 2-4 celled; ovule solitary, erect; styles 2; fruit generally a drupe, dehiscent or indehiscent; the involucre regarded by some as the calyx, encloses the nut, which is incompletely 2-4 celled; endosperm none, cotyledons corrugated, oily. One family Juglandaceae. The English Walnut (Juglans regia) from the Mediterranean region to the Himalayas is extensively cultivated in California, Spain, France, Italy, and other warm temperate countries; butternut (Juglans cinerea), from New Brunswick to North Dakota and Nebraska, produces a valuable wood which is, however, inferior to the black walnut (Juglans nigra), distributed from Massachusetts to Minnesota, Kansas and Texas, but

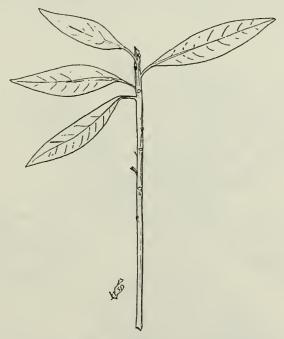


Fig. 183. Cork Wood (Leitneria floridana). The wood of this plant is extremely light. It grows in swamps in Southern Missouri and Florida. (W. S. Dudgeon.)

the timber is becoming scarce. There is a popular impression that the black walnut is poisonous to vegetation growing under the trees. California Walnut is J. californica. The Japanese walnut $(J.\ Sieboldianv)$ produces a large, thick-shelled nut. The bark of Juglans contains juglandic acid $C_{10}H_6O_8$. The Pterocarya caucasion is a native to the Trans-Caucasus.

The genus Carya is native to North America and yields valuable timber and nuts. The nut of the pecan (Carya illinoensis) is an important article of commerce in Texas and other southern states. The wood is also used. The shellbark hickory (Carya ovata) and the Missouri hickory (C. laciniata) supply valuable woods which are used in the manufacture of ax handles and for parts of wagons. The nut of the latter is large but like the preceeding one and the

C. tomontosa has a hard shell. The bitternut (C. cordiformis) and pignut (C. glabra have soft shells. The bark of hickories is used to flavor and color glucose to imitate maple sugar.

FAGALES

Monoecious or rarely dioecious trees or shrubs, with simple alternate leaves; stipules deciduous; calyx usually present; corolla usually wanting; staminate flowers in catkins; stamens 4-20; pistillate flowers solitary, clustered in scaly catkins; ovary more or less 2-7-celled with 1-2 pendulous, straight ovules, all the ovules but one disappearing in fruit; involucre becoming a burr or cup; embryo large; endosperm none. This is an important order, including the chesnut, oak, birches and alders. The bark of a few species of *Quercus*, because of its astringent qualities is used in medicine. The European filbert (*Corylus Avcllana*)

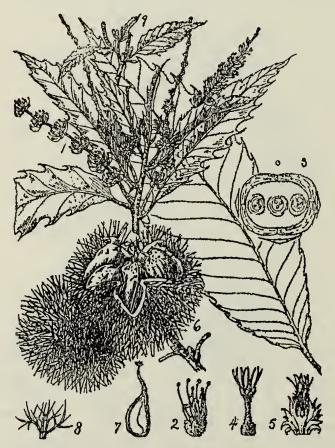


Fig. 183. American Chestnut (Castanca americana). 1. Flowering branch. 2. Staminate flower, enlarged. 3. Diagram of pistillate flower, eluster. 4. Pistillate flower, enlarged. 5. Longitudinal section of involucre of pistillate flowers. 6. Portion of fruiting branch. 7. Longitudinal section of fruit. 8. Involucral spine. 9. End of young branchlet. 1, 6, 7, one-half natural size. M. M. Cheney.

and our hazel nut (C. americana) are articles of commerce. Both the American beech (Fagus ferruginea), and the European (F. sylvatica), are prized for their nuts, which contain a valuable oil. The chestnuts are well known in commerce. The American chestnut (Castanea dentata) is common in the United States. The European species (C. sativa), has long been cultivated in North America; the Japanese (C. japonica) is also cultivated. In southern Europe, starch and flour are made from the nuts of the Spanish chestnut (C. sativa). The wood of many species of oak is used for interior finishing and furniture, among these are white oak (Quercus alba), red oak (Q. rubra), pin oak (Q. palustris), English oak (Q. Robur) and the live oak (Q. virginiana) of the south. The bark of several species like the scarlet oak (Q. coccinea), English oak, and chestnut oak (C. Muhlenbergii) is used for tanning. Cork is obtained from the bark of Q. Pseudo-suber. The galls of Q. lusitanica are used for dyeing purposes. They are known as the Alcppo galls and contain from 60-70 per cent of tannic acid. The bark of the White Oak is used in medicine and is recognized as medicinal in the United States Pharmacopocia. The main constituent of oak bark is tannin which is used like other astringents.

The acorns of the Belotes, the evergreen oak of Europe (Q. Ilex) are used for food. The oaks are well represented in eastern North America, several handsome species also occurring on the Pacific coast.

Formerly, and perhaps even now, in some regions the acorns of the White Oak and the Bur Oak were dried, roasted, ground, and used very much as the coffee berry.

In Europe, various species of oaks cause sickness and death in hogs and cattle. Dr. Chesnut suggests that this might possibly be caused by the tannin or bitter principle contained therein.

In sections of the country, where oaks are common, hogs are allowed to run in the forests, the farmers considering that the acorns are fattening. In some parts of the south, it is believed that the mast of oaks makes excellent feed for hogs but is poisonous to cows, a small amount merely decreasing the flow of milk while a greater quantity causes death. It is also claimed that the "sweet mast," that of the white and bur oaks, is less poisonous than the "bitter mast" of mast of black, pin, red and cow oaks. Mr. E. B. Watson made some inquiries upon this subject for me, among southern farmers and obtained evidence of four men, which differed slightly in detail but agreed in the conclusions that mast is poisonous to cattle but rather beneficial to horses and hogs. Some say that the coarse hulls or cups clog the digestive tract and cause unthriftiness; others that there is actually poison in the mast. That cattle are affected more seriously than hogs or horses may possibly be explained by the difference in the structure of the digestive organs.

In some localities tympanites is said to be produced in cattle that browse on the leaves and bark which are very strongly astringent. The white oak contains about 10 per cent of tannin. Q. lusitanica contains in addition to tannic acid mentioned before, from 2-4 per cent of gallic acid. That other plants of the order are injurious has been indicated by Freidberger and Fröhner who state that the European Beech produces violent colic, tetanus, mania and fits of madness resembling those produced by strychnin; that the autopsy shows lesions resembling strychnin poisoning. They recommend giving tannin, morphin and chloral hydrate.

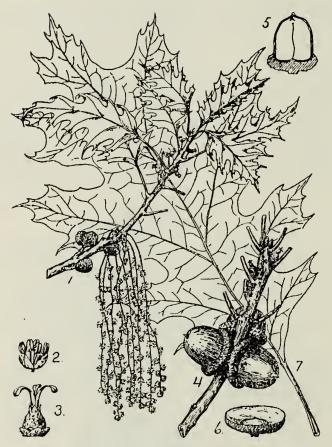


Fig. 184. Red Oak (*Quercus rubra*). 1. Flowering branch. 2. Staminate flower, enlarged. 3. Pistillate flower cluster, enlarged. 4. Fruiting branch, enlarged. 5. Acorn. 6. Cupule. 1, 5, 6, one-half natural size. M. M. Cheney.

The alders (Alnus glutinosa) of Europe are used as ornamental trees. Several trees of the genus, on the Pacific coast, are fair sized and produce good timber. The birches are valuable both as ornamental trees and for the excellent quality of their timber. The white birch (Betula alba) of Europe is frequently cultivated and birch tar is obtained from it. Cherry birch (B. lenta) and yellow birch (B. lutea) produce most valuable wood which is used for interior finishing. Paper birch (B. alba var. papyrifera) is used for making spools and canoes. The black birch (B. nigra) is common along our streams. The common source of oil of wintergreen is the cherry birch. This oil resembles that obtained from Gaultheria procumbens.

URTICALES

Trees, shrubs, or herbs; flowers never borne in catkins; monoecious, dioecious, or polygamous; ovary 1-celled, superior. Urticales are divided into the

Ulmaceae, the elms; Moraceac, the mulberries, and Urticaceae, the nettles. The Castlloa elastica of Mexico, the bread nut tree of Central America (Brosimum Alicastrum) and the Cecropia of tropical America furnish rubber.

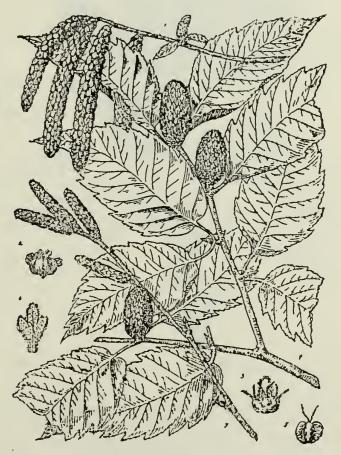


Fig. 185. Yellow Birch (Beinla lutea). Flowering branch. 2. Stanninate flower, enlarged. 3. Pistillate flower, enlarged. 4. Fruiting branch. 5. Nut, enlarged. 6. Scale of fruiting catkin, enlarged. 7. Winter branch, showing staminate catkin, 1, 4, 7, one-half natural size. (After M. M. Cheney, In Green's Forestry of Minnesota.)

The family Moraceae contains the bread fruit (Artocarpus incisa) an important article of food for the natives of the Pacific Islands, and the jack fruit (A. integrifolia) the fleshy envelopes of which are, however, somewhat poisonous. Canoe-gum, a very good substitute for rubber, is obtained from this genus. The figs belong to this family also; the sycamore fig tree (Ficus Sycamorus) produces small fruit which is used in Egypt for food. The common fig (Ficus Carica) is the most valuable; it includes the common and Smyrna fig of commerce, containing 60-70 per cent of grape sugar. The India rubber tree (F. clastica) is the source of some of the India rubber. The ban-



Fig. 186. Rubber-Tree (Castilloa elastica). Staminate flowering branch. A commercial rubber tree of Central America. (After Faguet.)

yan trees (F. Benghalensis, F. religiosa and F. altissima) of the East Indies furnish shellac. The mulberry (Morus nigra) is largely cultivated both for its fruit and for its wood, the latter being very durable for posts. The white mulberry (M. alba) is extensively planted, the leaves being used as food for the silk worm. The Osage orange (Machira pomifera) of Arkansas, Indian Territory and Texas produces a very durable wood used for posts and pulley blocks. The fruit is said to be poisonous. The wood of fustic (M. tinctoria) of the West Indies is used for many purposes. The bark of the paper mulberry (Broussonetia papyrifera) is made into paper, and in Japan is also made into cloth. The paper mulberry is cultivated in the South.

The Upas tree (Antiaris toxicaria) contains antiarin $C_{27}H_{42}O_{10}+H_2O$. This tree furnishes an arrow poison which the natives prepare from the plant. It is a semi-liquid greenish black substance. The poison acts on the brain and respiratory nerves, causing vomiting and loss of sensation.

Ramie grass cloth or China grass (Boehmeria nivea) produces a fine fibre but it is difficult to separate it from the bark and wood. The hop (Humulus Lupulus) is cultivated and is the source of the oil of hops which imparts an aromatic, bitter flavor to beer. It contains lupulin which is a tonic and slightly nar-cotic. The elms are commonly cultivated as shade trees. The best is the American

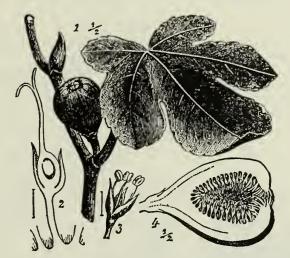


Fig. 187. Fig-tree (Ficus Carica). 1. Flowering branch. 2. Pistillate flower cut through longitudinally. 3. Staminate flower. 4. Fig in longitudinal section. (After Wossidlo.)

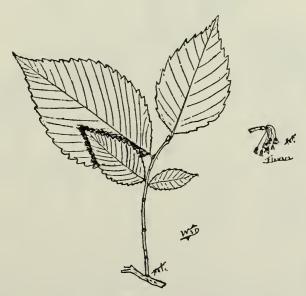


Fig. 188. Flowers and leaves of the American Elm (Ulmus americana). A familiar tree furnishing an important commercial wood. (W. S. Dudgeon.)



Fig. 189. Deadly Upas Tree (Antiaris toxicaria). Flowering branch; portion of staminate flower; longitudinal section of pistillate flower. (After Faguet.)

ehn (Ulmus americana) which supplies a wood used largely in the manufacture of chairs. The partly ornamental elms are rock elm (U. racemosa) and slippery elm (U. fulva). The bark of the latter is used in medicine. The rock elm is used in the manufacture of bicycles; the wood taking a pretty finish. One of the common elms of Europe, cultivated in the United States, is the U. campestris. The U. montana is the Scotch elm. The hackberry (Celtis occidentalis) is a valuable shade tree and furnishes pretty wood which is difficult to work. The Chlorophora excelsa is one of the best timber trees of West Africa.

URTICACEAE. Nettle Family

Herbs, trees, or shrubs with stipules; flowers monoecious or dioecious, rarely perfect; calyx from the 1-2-celled ovary which forms a 1-seeded fruit; stamens as many as the lobes of the calyx or sometimes fewer; opposite, ex-albuminous or albuminous; when albuminous the radical points upward; cotyledons broad. A small family of 500 species some of which, like Boehmeria, produce valuable fibers. The leaves of Pilea pumila are demulcent and are said to be valuable in Rhus poisoning.



Fig. 190. Slippery Elm (Ulmus fulva). 1. Flowering branch.
2. Fruiting branch. 3. Winter branch with buds. 4. Same showing flower buds beginning to enlarge. 5. Summer branch. 6. Flower enlarged. 7. Longitudinal section of flower, enlarged. 8. Longitudinal section of pistil, enlarged. 9. Stamen, enlarged. 10. Cross section of ovary, enlarged. 11. Longitudinal section of fruit. 12. Seed, enlarged. 13. Longitudinal section of seed, enlarged. 14. Embryo, enlarged. 1, 2, 3, 4, 5, 11, one-half natural size. (After M. M. Cheney, in Green's Forestry in Minnesota.)

Genera of Urticaceae

| Herbs with stinging hairs. |
|--|
| Leaves opposite; flowers 4-parted |
| Leaves alternate; staminate flowers 5-parted |
| Herbs or trees without stinging hairs. |
| Herbs; pistillate flowers spiked |
| Trees; staminate flowers racemose4. Maclura. |
| 1 Cannabis Tourn Hemp |

1. Cannabis, Tourn. Hemp

Dioecious herbs with tough fiber to the inner bark; greenish flowers; sepals 5 in the staminate, 1 in the fertile flower; achene, crustaceous.

Cannabis sativa, L.

Stem from 4-8 feet high with broad, divided leaves, the linear-lanceolate segments sharply and closely serrate; greenish flowers with narrow staminate panicles and erect pistillate spikes, the sterile with 5 sepals and 5 stamens, fertile flowered spiked, with 1 sepal; fruit hard ovoid, achene oblong.

Distribution. Native to Europe and Asia and in waste places from New Brunswick to Tennessee, Kansas and Minnesota.

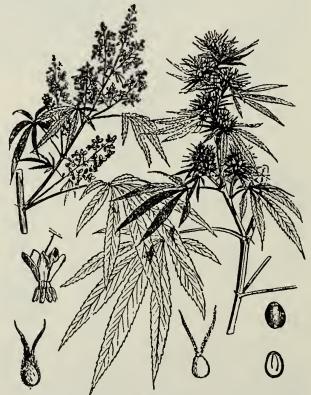


Fig. 191. Hemp (Cannabis sativa). Staminate and pistillate flowering branches; fruit; longitudinal section of fruit. (After Faguet.)

Poisonous Properties. The resinous secretions of this plant possess very powerful medicinal properties which, however, are said not to be produced by the plant when grown in temperate climates.

Indian Hemp (Cannabis indica) is probably not essentially different from the common homp and has been used in medicine for a long time. According to Dr. Houghton and Mr. Hamilton the American grown product is equal to the Indian Hemp.

The use of hemp seems to have spread through India, Persia and Arabia during the early middle ages. The Hashishin, a sect of the Moravians, killed a large number of the Crusaders during the 11th and 12th centuries by the use of hemp as an intoxicant. The drug is largely grown in India and Turkestan. The form of hemp commonly reached by commerce is called Bhang or Hashish and consists of dried leaves and small stalks frequently mixed. with fruits. This is smoked in India with or without tobacco. Ganjah is obtained from the flowering shoots of the female plant or stalk, a stiff woody stem several inches long which is pruned to produce flowering branches. The tops of these are collected then pressed by being trodden by the feet. From this mass comes the drug known as ganjah. It grows in an altitude of six thousand feet. The other forms of the plant consumed in India are Bhang and Charras. Subjec or Bhang is used for smoking. The narcotic ingredient found in majun and charras is undried resin which is obtained by the natives who, when passing among plants wear rubber aprons to which the resin adheres, after which the product is scraped together. The principal constitutents of hemp are resin and a volatile oil. The oil or amber colored substance has an oppressive hemp-like smell, and furnishes a resinous substance, cannabin which crystalizes in needles and acts like strychnin. Cannabinol, with intoxicating properties, is obtained from cannabin and is a product from the glands of Cannabis. Cannabin hybrid (C18H22) is a substance with the coniinlike odor; it is antispasmodic and soporific, and anodyne and a nerve stimulant. Dr. C. F. Millspaugh referring to the products of plants affording this oil concluded from experiments made, that this drug causes depression, epilepsy, vertigo, congestion, followed by cephalalgia, ear-ache, tooth-ache, dryness of mouth, throat, lips and lids; it produces nausea, vomiting after coffee, palpitation of the heart, weakness of the limbs and dreaminess during sleep. It produces the same symptoms in animals.

The stem of hemp is used by the Mohammedans who smoke it in combination with other substances. They also smoke the sun-dried leaves. It is intoxicating and restful to the smoker and alleviates pain, increases the appetite, causes sleep, and induces cheerfulness. It also produces violent coughing and nose bleed.

Hemp is most important in China, and other Asiatic countries, for the manufacture of cordage. The growing of hemp for the same purpose is also carried on to some extent in Nebraska and Kentucky. The seeds of the plant furnish food to birds.

2. Urtica (Tourn.) L. Nettle

Herbs with stinging hairs; flowers greenish, monoecious or rarely dioecious, clustered; staminate, with 4 stamens; fertile, with 4 sepals in pairs; fruit an erect, ovate, flattened acheme. A small genus of 30 species.

Urtica gracilis Ait.

A perennial from 2-6 feet high, sparingly bristly; leaves ovate, lanceolate with slender petioles; long, accuminate, sharply serrate, 3-5-nerved, the slender petioles sparingly bristly; flowers dioecious or with staminate and pistillate clusters. The stinging hairs of this and other species of the genus contain formic acid. A common weed in dry or moist ground along fence rows from Canada to British Columbia, Kansas and North Carolina.

Poisonous properties. The nettle and some other plants produce what is commonly called "urticaria" or nettle rash. It is an inflammatory disorder with a burning and itching sensation. It may come out in large or small patches, remaining for a few minutes or several hours and may disappear as abruptly. It usually leaves no trace behind. The nettle is supposed to contain an irritant toxic principle, formic acid, but recent studies seem to indicate that the urticaria is probably caused by one of the toxins.

The following species of the genus have urticating properties: Urtica membranaceae, U. spatulata and U. pilulifera.



Fig. 192. Stinging Nettle (Urtica urens). (From Darlington's Weeds and Useful Plants.)

Urtica urens L. Small Stinging Nettle

An annual from 1-2 feet high; stem 4-angled, tough, branching with a few stinging virulent hairs; leaves elliptical or ovate, serrate or incised, with scattered stinging hairs; flowers loose or in racemose spikes; sepals 4 petals 4; fruit straight, ovate, flattened achene.

Distribution. From New Foundland to Florida and also on Pacific Coast. *Poisonous properties.* This nettle has been used in medicine but it is not officinal. Formerly it was used for flagellation of the skin.

Urtica holosericea Nutt

A tall perennial with stinging hairs; leaves thick, oblong, ovate or ovatelanceolate; flowers in open panicles.

Urtica dioica L. Stinging Nettle

An erect perennial; leaves and stems beset with stinging hairs; leaves thin, ovate, long petioled, acute or acuminate at the apex, cordate at the base, sharply serrate; flowers in large clusters, cymose-paniculate, often dioecious.

Distribution. Native to Europe but largely naturalized in North America from Atlantic coast to Minnesota and Missouri.

Poisonous properties. Poisonous like the preceding

3. Laportea Gaudichaud. Wood Nettle.

Perennial herbs with stinging hairs; flowers monoecious or dioecious in loose eymes, the lower mostly sterile; staminate flowers with 5 imbricated sepals; 5 stamens and a rudimentary ovary; pistillate flowers with 4 unequal sepals; stigma elongate, awl-shaped; achene ovate flat; endosperm scant or obscure. About 25 species in warm countries.



Fig. 193. Common Nettle (*Urtica dioica*). Sometimes causes urticaria. (From Johnson's Med. Bot. of N. A.)

Laportea canadensis Gaudichaud. Wood Nettle.

Perennial stem 2-3 feet high; leaves ovate, pointed thin, long-petioled, sharply serrate; fertile cymes divergent; achene smooth, as long as the calyx.

Distribution. In rich woods from Nova Scotia to Minnesota and Kansas and south to Florida.

Poisonous properties. It acts similarly to nettle, the poisonous action being even more pronounced. L. crenulata, L. gigas, and L. stimulosa also possess similar properties.

Maclura Nutt. Osage Orange,

Tree with milky juice; leaves alternate, pinnately veined; stipules caducous; stout, axillary spines; flowers dioecious, staminate in loose, short racemes with 4-parted calyx and 4-stamens; pistillate, capitate with a 4-cleft calyx enclosing the sessile ovary and long exserted style; fruit an achene surrounded by a fleshy calyx; endosperm none; embryo curved; it contains a single species named *Toxylon* by Rafinesque.

Maclura pomifera (Raf.) Schneider. Osage Orange

A tree 30-50 feet high; leaves ovate to oblong, lanceolate, pointed, mostly rounded at the base, green and shining; the syncarpous fruit is globose, yellowish green 2-4 inches in diameter; the wood is hard and tough and is used in the manufacture of wagons for paving, fencepost, etc. The tree is extensively planted as a hedge plant.

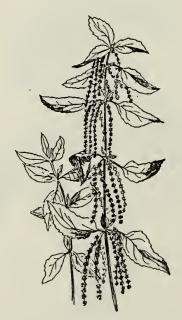


Fig. 194. Wood Nettle (Laportea canadensis). A common wood plant causing urticaria. (C. M. King.)

Distribution. In rich woods from Missouri to Kansas to Texas; widely cultivated in the north from southern Nebraska to southern Iowa, Illinois and eastward.

Poisonous properties. This species is listed as poisonous by Professor Bessey in Nebraska. Dr. Halsted notes that a friend of his while working in Osage Orange hedges suffered considerably because of inflamation following the piercing of the thorn. The writer had a similar experience.

Dr. Bessey says: "The Osage Orange (Maclura pomifera) which has been shown by Dr. Halsted to be more or less harmful as an external poison, is very commonly grown in the southern portion of the state, and it thus adds another to the plants to be avoided by some people. Although I am quite sensitive to some of the external poisons, I have myself never experienced any bad effects from handling the leaves or fruit of the Osage Orange."

PROTEALES

The proteates include one family, the *Proteaceae*, with nearly 1000 species, native to the tropics, mostly of the southern hemisphere.

SANTALALES

Herbs or shrubs generally parasitic; flowers solitary or clustered without corolla; calyx present, imperfect or perfect; pistil 1. Of the two families in the United States, the Loranthaceae contains the Southern mistletoe (Phorandendron) parasitic upon various decidious trees like the oak and elm; the Arceuthobium of Europe, and the Rocky Mountains; species parasitic upon confers, one also occuring on spruce trees in eastern North America. Hyams is authority for the statement that the berries of Pheradendron flavescens are poisonous to children. Several deaths have been attributed to them. Santalaceae contains the fragrant sandalwood (Santalum album) of the Indian-Malayan region which contains an oil used in medicine for venereal diseases and for



Fig. 195. Wild Ginger (Asarum canadense). Wild ginger is used in medicine. The roots are spicy fragrant; some plants related to it are poisonous. (W. S. Dudgeon.)

perfumes. The bastard toadflax (Comandra umbellata) of our northern woods is parasitic upon the roots of flowering plants. The family Balanophoraceae consists of chlorophylless parasitic plants with twining or acaulescent stems, and is native in tropical woods and savannas of Java, India and Australia.

ARISTOLOCHIALES

Plants with twining or acaulescent stems; leaves cordate or reniform; flowers perfect; calyx inferior, the tube adnate to the ovary or partly so; corolla none; ovary generally 6-celled. There are only three families, one of which occurs in North America. The Aristolochiaceae includes the wild ginger of the North (Asarum canadense) which is more or less purgative and probably also to be regarded as suspicious; its rhizome furnishing the substance asarin and a volatile oil which is used in perfumery; the A. europaeum, listed by Lehmann as poisonous because of its purgative action and blistering proper-

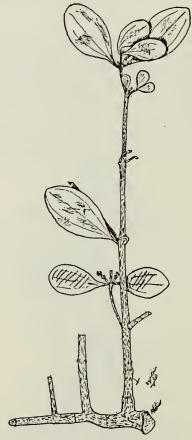


Fig. 196. Southern Mistletoe (Phoradendron flavescens). The berries of this plant are said to be poisonous. (W. S. Dudgeon.)

ties; the Dutchman's pipe (Aristolochia macrophylla) frequently cultivated and hardly as far north as Minnesota and Wisconsin; the gooseplant (Aristolochia grandiflora) of Brazil whose flowers emit an offensive odor, but in spite of this fact the plant is cultivated in greenhouses; Virginia snakeroot (Aristolochia Serpentaria) the root stock of which is used as a tonic and contains a volatile oil borneol, a bitter poisonous principle aristolochin $C_{32}H_{22}N_2O_{13}$ and the alkaloid aristolochinin. The European (A. Clematitis) produces colic and other gastric disturbances and is listed among the pungent narcotic poisons. The substance asarin when heated is irritating.

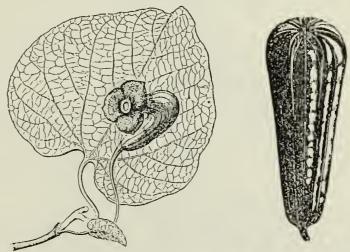


Fig. 197. Dutchman's Pipe (Aristolochia macrophylla). Leaf and flower, dehiscent fruit. (After A. Faguet.)

Several species of the genus Aristolochia are used as antidotes against snake-bites and this use is clearly indicated in some of the specific names, as in Virginia snakeroot (A. Serpentaria). Other plants of the genus are said to be poisonous, A.grandiflora being an example of this. The Arabs use A. sempervirens and A. indica as snake poison antidotes. According to R. B. White, the Guaco (A. mexicana) is a cure for snake-bites. Many other are used for the same purpose, several composites well-known remedies. Among the latter are Liatris squarrosa, Cacalia tuberosa, and Prenanthes alba. Other plants having the same qualities belong to the families Ranunculaceae, Orchidaceae, Violaceae, Polygalaceae, Liliaceae, Umbelliferae, Filices, and Palmae. One has only to look through such works as the Robinson and Gray's Manual, or Britton's Manual, or various old medical works for the common names of plants with the word snake attached to them, to understand how prevalent was the belief that these plants were antidotes against the bite of venemous snakes.

POLYGONALES

Herbs, shrubs, or trees, often climbing vines; leaves alternate or occasionally opposite; jointed stems; flowers small, regular, dioecious, monoecious or polygamous; calyx 2-6 cleft or parted, inferior; stamens 2-9, inserted near the



Pig. 198. European Aristolochia (Aristolochia Clematitis). Flowering stem. Listed as a narcotic poison. (After Faguet.)

base; pistil 1 with superior ovary; fruit an achene; endosperm mealy. Contains a single family (Polygonaceae).

POLYGONACEAE. Buckwheat Family

Herbs, shrubs, or trees, often climbing; jointed stems; stipules in the form of sheaths; juice often acrid or acid; leaves alternate or occasionally opposite; flowers small, regular, mostly perfect; calyx more or less persistent; ovary 1-celled, bearing 2-3 styles or stigmas and a single erect ovule; fruit an achene, 3-4-angled or winged, invested by the calyx; embryo curved or nearly straight; endospern mealy, copious. About 800 species. Of economic importance are the pie plant (Rhenm Rhaponticum); and rhubarb (R. officinale) of Thibet, the root of which contains cathartic acid and is a powerful cathartic; it also contains chrysophan $C_{27}H_{30}O_{14}$, emodin $C_{15}H_7O_2(OH)_3$, rhein $C_{15}H_6O_2(OH)_4$ and chrysophanic acid $C_{15}H_8O_2(OH)_2$. It is purgative and astringent. The canaigre (Rumex hymenoscepalus) produces a thick root valuable for tanning leather. It is a native of the southwest. The tannin is the same

as that found in rhubarb, and rheotannic acid. The patience dock R. Patientia), pale dock (R. altissimus) and curled dock (R. crispus) are troublesome weeds; French sorrel (R. scutatus) is cultivated in Europe and used as a salad. The presence of the silver plant of the west (Eriogonum umbellatum) is said to be indicative of gold and silver. Muchlenbeckia platyclados of the Samoan Islands is frequently cultivated in greenhouses. The mountain sorrel (Oxyria digyna) is used as a salad plant.

Genera of Polygonaccae

Achenes triangular or lenticular.

Embryo slender curved around one side of the endosperm 3 Polygonum. Broad cotyledons of embryo twisted and plaited........... Fagopyrum.

1. Fagopyrum (Tourn.) L. Buckwheat

Annual or perennial; somewhat fleshy, smooth, leafy herbs with erect stems; leaves petioled and alternate; hastate or deltoid flowers, small, white, or greenish, paniculately-racemose, perfect; calyx 5-parted, persistent, the divisions like petals; stamen 8; ovary 1-celled, 1 ovule, style with 3 divisions; fruit an achene, 3-angled; endosperm mealy; cotyledons broad. About 6 species native to the old world.

Fagopyrum esculentum Moench. Buckwheat

Smoothish plants; leaves hastate, abruptly narrowed above the middle; sheath half-cylindrical; racemes somewhat panicled, many flowered; sepals white, fragrant, with 8 honey-bearing yellow glands situated between the stamens.

Distribution. A common escape in eastern North America. Native of Eastern Europe and Western Asia.

Fagopyrum tataricum (L.) Gaertn. India-wheat

Annual, similar to the above species; leaves deltoid, hastate; flowers smaller; pedicel short.

Distribution. In waste places from eastern Canada to New England. Native to Asia.

Poisonous properties. Fagopyrum contains the glucoside indican C₂₆H₃₁NO₁₇ found also in Nerium and other plants. The plant produces bloat especially if consumed before bloom.

Several years ago the writer received a complaint from a farmer stating that the feeding of buckwheat had produced a rash upon his hogs. Feeding of buckwheat and the cruptions or urticaria following are well known to veterinarians.

Dr. Millspaugh says of buckwheat:

Many individuals cannot partake of pancakes made from the flour of the seeds without experiencing a severe itching especially observed about the large joints. A peculiarity of this itching is that it occurs after the removing of the clothing and when first retiring at night. The eruption incident to and following this itching takes the form of vesicles which degenerate into dry, dark colored scabs. Another symptom arising is a glutinous condition of otherwise natural feces, making expulsion quive difficult. Increased urinary discharge is also present in many cases.

2. Rume.r. L. Dock

Coarse herbs, with small, mostly green flowers, which are crowded on generally whorled, panicled racemes; petioles partly sheathing at base; 6 sepals; 3 outer herbaceous, sometimes united at base, spreading in fruit; 3 inner larger, slightly colored, enlarged after flowering and convergent on 3-angled achene, veined, often bearing a grain-like tubercle on the back; stamens 6; styles 3; stigmas tufted; embryo lying along one side of the albumen, slender, and slightly curved.

It has been claimed by some that the seeds of R. Acetosella poison horses and sheep.

Rumex altissimus Wood. Pale Dock.

A tall perennial from 2-6 feet high, glabrous with erect stem, simple or branched above; leaves ovate or oblong; lanceolate, long, acute, pale green, veins obscure; raceines spike-like or somewhat interrupted below, spreading



Fig. 199. Two weeds of the smartweed family. A. Sourdock (Rumex crispus). B. Sheep sorrel (Rumex Acetosella). Both have been suspected. They contain a great deal of oxalate of lime. (U. S. Dept. of Agr.)





Pennsylvania Smartweed or Persicaria (Polygonum pennsylvanicum). Common in the North. Smut found on this plant is very irritating. (Ia. Geol. Sur., p. 421).

in fruit; pedicels nodding, shorter than the fruting calyx; valves broadly ovate with a conspicuous ovoid tubercle.

Distribution. Common throughout the northern part of the United States.

Rumex erispus L. Curled Dock

A smooth perennial from 3-4 feet high; leaves with strongly wavy and curled margins, lanceolate and acute; in the lower leaves bases are somewhat truncate or inclined to be ear-shaped; flowers collected in dense whorls, extended or prolonged into racemes, entirely leafless above, but below with small leaves; flower consists of 6 sepals, fruiting pedicels as long as the calyx wings; wings heart-shaped, erose dentate, each showing a tubercle; achene 3-angled, smooth.

Poisonous properties. The docks contain rumicin C₁₄H₁₀O₄, which is a tasteless, golden-yellow substance, slightly soluble in hot water. It acts as a rubefacient and discutient and is used for destroying parasites of the skin.

Rumex has found a place in Pharmacopoea and is also used in medical practise. It causes nausca, watery brown faeces, copious urination, a dry spasmodic cough, and perspiration. The Rumex orbieulatus, Great Water Dock, according to Dr. Johnson, is tonic, astringent, and slightly laxative.

Medical properties. It is used as a stimulant and diuretic.

3. Polygonum L. Smartweed

Annual or perennial herbs, occasionally woody; stem erect, climbing or floating; leaves alternate, entire, ochreae cylindrical, often fringed; flowers mostly perfect, green white, pink, or purple; calyx 4-5 parted or cleft; stemens 5-9, filaments filiform or dilated to the base; style 2 or 3 parted or cleft; achene lenticular or 3-angled, rarely 4-angled; endosperm present. About 200 species of wide distribution. The P. tinetorium of China furnishes the Chinese indigo. The Saghalen knotweed (P. saehalinense) was widely advertised as a forage plant a few years ago and is used in Japan and Manchuria as we use asparagus. The prince's feather (P. orientale) is cultivated for ornamental purposes. The tanweed (P. Muhlenbergii), smartweed (P. Persicaria), knotgrass (P. aviculare) and black bindweed (P. Convolvulus) are troublesome. The Pennsylvania persicaria (P. pennsylvanicum) is a valuable honey plant and its seed is a common impurity in clover.

Polygonum Persicaria L. Lady's Thumb

A nearly smooth and glabrous annual from 12-18 inches high; leaves lanceolate or linear, marked with a lunar blotch near the middle, acuminate ochreae somewhat bristly; ciliate spikes ovoid or oblong, erect; stamens mostly 6; style 2-3 parted; achene lenticular.

Distribution. Across the continent in moist places; naturalized from Europe.

Poisonous properties. None of the species is relished by stock; the P. aere and P. Hydropiper are very acrid and produce gastro-enteritis and erythema, like that caused by buckwheat. The following species produce similar troubles.

Polygonum acre H.B.K. Water Smartweed

A nearly smooth perennial; stems rooting at the decumbent base; leaves linear-lanceolate; ochreae strigose, fringed with long bristles; spikes erect,

panicled; flowers whitish or flesh colored; stamen 8; achenes 3-angled or 4-angled, smooth and shining.

Distribution. Common southward from Missouri to Louisiana, Texas and Mexico.



l'ig. 200. Ladies' Thumb (Polygonum Persicaria). Common in moint places. (Charlotte M. King.)

Polygonum hydropiperoides Michx. Wild Water Pepper

A smooth, branching perennial, slightly or not at all acrid; 1-3 feet high; the narrow sheaths hairy, leaves narrowly lanceolate or oblong-lanceolate; spikes erect, slender, sometimes filiform; flowers small, flesh colored, or nearly white; stamens 8, style 3-parted to below the middle; achene 3-angled, ovoid or oblong, smooth and shining.

Distribution. In swamps or wet soils across the continent from New Brunswick to California, Florida and Mexico.

Polygonum Hydropiper L. Smartweed. Water Pepper

Smooth, crect annual, 1-2 feet high; stem often reddish; leaves linear-lanceolate, or lanceolate; spikes, nodding, usually short or interrupted; flowers



Fig. 201, Water Pepper (Polygonum Hydropiper). Troublesome to sheep. (After Hochstein.)

mostly greenish; stamens 4 or sometimes 6; ochreae cylindrical, fringed with short bristles; style short, 2-3 parted; achene lenticular or 3-angled, dull, granular.

Poisonous properties. This species and the door yard knot weed (P. aviculare) are said to be troublesome to sheep

CENTROSPERMAE

Herbs mostly with perfect flowers; calyx present; corolla, when present, polypetalous; ovary superior; perisperm present; embryo coiled, curled or annular; fruit not an achene. Generally fleshy plants, many found in saline soils. It includes the families Chenopodiaceae, Amaranthaceae, Nyctaginaceae, Phytolaccaceae, Caryophyllaceae, Portulacaceae and others. The last named contains the garden pussley (Portulaca oleracea), the moss pink (P. grandiflora), and the bitter root (Lewisia rediviva). The family Aizoaceae also of this order, contains the New Zcaland spinach (Tetragonia expansa), and the ice plant

(Mesembryanthemum crystallinum). The family Basellaceae another of this same order contains the Madeira vine (Boussingaultia baselloides) commonly cultivated. The seeds of some species are edible.

Families of Centrospermae

Fruit an utricle.

Flowers bractless or occasionally with bracts.

Flowers bracted.

CHENOPODIACEAE. Goosefoot Family

Annuals or perennials, frequently succulent herbs, or rarely shrubs; alternate leaves without stipules; flowers small; greenish; petals absent; calyx free, stamens as many as the lobes of the calyx or fewer and inserted opposite them on their base; ovary 1-celled; fruit a 1-seeded, thin utricle or rarely an achene; endosperm mealy or wanting; embryo coiled.

About 500 species of wide distribution, common in arid regions. Some of the economic plants of this family are sugar beet (Beta vulgaris), a maritime plant of Europe, and spinach (Spinacia oleracea) from the orient. The beet is one of the most important plants of the family, being largely cultivated in Europe as a source of cane sugar although as late as 1800, its use in that capacity was of little extent. It is also an important plant for stock food and for human food. Spinach is used extensively for greens but, in Utah, is somewhat of a weed. The Australian saltbushes are well known forage plants. Indigo is derived from A. hortensis, a native of Tartary. The strawberry blite (Chenopodium capitatum) is cultivated in Europe for its leaves. The shrubby saltwort (Suaeda fruiticosa) is burned in the south of Europe for Barilla. The Russian thistle (Salsola Kali. var. tenuifolia) is used in much the same way. The Spanish wormseed (S. Webbii) contains an oil much like that found in Chenopodium ambrosioides. The tumble-weed (Cycloloma atriplicifolium) is common on the plains. The white sage (Eurotia ceratoides) is an excellent forage plant of the west.

Poisonous properties. The use of the beet leaves for fodder has sometimes caused bloat. It has been known for some time that the feeding of roots to animals causes the formation of renal calculi. These calculi consist of a combination of uric and phosphoric acid with lime. An experiment conducted by Prof. W. J. Kennedy and Mr. E. J. Robbins at the Iowa Experiment Station in coöperation with Prof. L. G. Michael indicated that sugar beets fed to rams will produce renal calculi. Prof. Michael says* in regard to the effect of mangels and sugar beets on the kidney:

Both roots seem to affect the kidney similarly.

A small calculus was found in one kidney of Ram VI. This ram was fed sugar beets. The membrane about the calculus and extending down into the urethra was pigmented, a decided black.

^{*} Biennial Rept. Iowa State College of Agriculture and Mechanic Arts 23:142.



Fig. 202. Salt Bush (Atriplex concscens). A common plant of saline soil in the west. (U. S. Dept. of Agrl.)

In the kidney of Ram III the same kind of pigmentation occurred as in Ram VI. Ram III was fed Mangels. No calculus was present.

Miss S. Hartzell, who investigated the chemistry, reports as follows:

A post mortem examination of several valuable animals which the Experiment Station lost showed that renal calculi were present. This resulted in the metabolism experiment which was conducted by the Animal Husbandry Section in co-operation with the Chemical Section of the Experiment Station.

Thus far 42 rams have been used in the experiment, of which 11 were fed hay and corn; 9 were fed hay, corn and ensilage; 11 were fed hay, corn and mangels; 11 were fed hay, corn and sugar beets. In the case of those which were fed hay and corn, and also of those which were fed hay, corn and ensilage, the bladders and kidneys were normal while the results were the reverse in the case of those which were fed sugar beets or mangels along with the hay and corn. The sugar beets and mangels had the same effect The kidneys were larger than normal, the gall bladders were distended, the bladders were enlarged, often very much so, and in several cases the heart was enlarged. The kidneys in all cases were of a pale color. Three animals died during the experiment. The bladder of one was highly inflamed while in the other two the bladders contained calculi which were too large to pass thru the urethra. The kidneys of two of the animals were broken

down while the third was much enlarged, and calculi were present. In one of the animals which was slaughtered the following conditions existed: bladder enlarged; ulcerated between nreters; kidney surface mottled; kidneys enlarged; cortex discolored; calyces full of calculi; gall bladder much enlarged; other organs normal.

According to Greshoff the leaf of Kochia scoparia contains saponin, as do the seeds of this species and of K. arenaria. A species of Atriplex native to China causes a skin disease known as "atriplicimus."

Genera of Chenopodiaceae

Embryo coiled into a spiral; calyx horizontally winged...........3. Salsola.



Fig. 203. Common Pigweed (Chenopodium album). Young shoots sometimes used in place of spinach. (Charlotte M. King.)

Chenopodium L. Pigwccd. Goosefoot. Lamb's quarter

Annual or perennial herbs usually covered with a white mealy substance; flowers inconspicuous, in sessile, small clusters, collected in spikes or panicles, perfect; calyx 5-, rarely 4-parted or lobed; stamens generally 5; styles 2, rarely 3; ovary 1-celled, becoming a 1-seeded, thin utricle; embryo coiled around the mealy endosperm.

A small genus of about 60 species of wide distribution in saline soil, around dwellings and in manured soil. Several species like the common pigweed (C. album), the Australian spinach (C. auricomum), and the English Good King Henry (C. Bonus-Henricus), are used as a substitute for spinach. The quinoa (C. Quinoa) is an annual, native to Peru, which produces its flowers in dense, erect panicles. It is cultivated in Chili and Peru for its seeds, which are said to be very strengthening. It was the principal meal food of the Peruvians be-

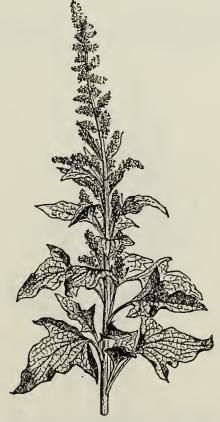


Fig. 204. Good King-Henry (Chenopodium Bonus-Henricus). Used as a substitute for spinach. (From The American Agriculturist.)

fore the conquest by Spain. An oil is obtained from the wormseed (C. ambrosioides). Several species of the genus like Chenopodium album and C. hybridum are weedy.

Chenopodium Botrys L. Jerusalem Oak

A more or less glandular, pubescent, aromatic annual; leaves with slender petioles, oblong, obtuse, sinuate, pinnatifid, flowers in leafless racemes; calyx 2-3 parted, dry in fruit, only partially enclosed.

Distribution. Naturalized from tropical America. Common in eastern North America to Oregon.

Chenopodium ambrosioides L. Mexican Tea. Wormseed

A smooth annual; leaves slightly petioled, aromatic, oblong, lanceolate, toothed or nearly entire; flowers in spikes, leafy or intermixed with leaves; calyx 2-3 parted; fruit dry, enclosed by the calyx.

Distribution. Naturalized from tropical America. Common in eastern North America to California. The fruit is officinal in the U. S. Pharmocopoeia.

Chenopodium ambrosioides L. var. anthelminticum (L.) Gray. Wormseed

An annual or perennial weed, glandular pubescent; leaves lanceolate, or oyate-lanceolate, acuminate at the apex and narrowed at the base, the lower



Fig. 205. Wormseed (Chenopodium ambrosioides). Fruit officinal. (From Darlington's Weeds and Useful Plants.)

laciniate pinnatifid; flowers in spikes without bracts, or the lower spikes leafy bracted.

Distribition. Naturalized from Europe in waste places, from Massachusetts to Ontario and from Wisconsin to Mexico.

Poisonous properties. Several species of the genus contain volatile oils. The C. ambrosioides, var. contains the volatile oil of wormseed. This oil has a peculiar, strong, offensive odor and a pungent disagreeable, but aromatic taste. It is said to contain chenopodin, C₆H₁₈NO₂. In the case of a man who took about one half an ounce of a soluble oil of wormseed, Dr. Millspaugh says that the symptoms were those from a narcotic, acrid poison, affecting the brain, spinal cord and stomach. The patient was insensible, convulsed and foamed at the mouth. In another case a man who had taken a considerable quantity displayed hilarity and made futile attempts at talking like a drunken man. Death followed later. C. mexicanum contains saponin.

2. Sarcobatus Nees. Grease-wood

An erect, branched shrub with spiny branches; leaves alternate; linear, fleshy; flowers dioecious or monoecious; the staminate in terminal clusters without a calyx; the pistillate solitary in the axils with compressed calyx, adnate to the base of the papillose stigmas; in fruit a membranous horizontal wing; seed vertical; embryo green, coiled into a flat spiral. Species 1.

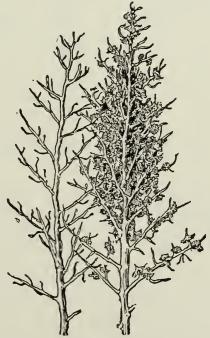


Fig. 206. Grease wood (Sarcobatus Maximiliana). A plant growing in alkaline soils in the Western United States. Poisonous to sheep. The sharp spines cause mechanical injury. (U. S. Dept. Agrl.)

Sarcobatus Maximiliani Nees, Torr. Grease-wood

A glabrous perennial with succulent foliage and spiny branches. Wood hard.

Distribution. In dry, alkaline soil from western Nebraska to New Mexico, Nevada, and Montana; most abundant west of the front Rockies.

Poisonous properties. Prof. Chesnut says:

A correspondent in New Mexico states that on one occasion he counted as many as one hundred sheep that had been killed by eating the leaves of this plant. It is claimed that cows are not affected by eating it at any time and that sheep can eat it quite freely in winter. Death is perhaps due more to the bloating effect than to any poisonous substance which the plant contains.

It might be noted also in this connection that the sharp spines on the plant often inflict serious injuries to persons who come in contact with it and also to animals, setting up inflammation and causing the formation of pus. It is used as a forage plant.

3. Salsola L. Saltwort

Bushy branched herbs, succulent when young, but rigid at maturity; leaves terete, prickly-pointed and sessile; flowers sessile and axillary; calyx 5-parted, persistent, enclosing the depressed fruit, the divisions horizontal, winged on the back, enclosing the utricle; stamens 5; ovary depressed; style 2; embryo coiled in a conical spiral. About 40 species of wide distribution, saline soils.

Salsola Kali L., var. tenuifolia G. T. W. Meyer. Russian Thistle

An herbaceous, smooth or slightly pubescent annual, diffusely branched from the base; from $1\frac{1}{2}$ to 3 feet high, spherical in the mature form; leaves fleshy, alternate, succulent, linear, subterete, 1-2 inches long, pointed in the older specimens; upper leaves in the mature plant persistent, each subtending 2 leaf-like bracts and a flower; stem and branches red; apetalous flowers solitary and sessile; calyx consisting of 5 persistent lobes, enclosing the dry fruit which is usually rosc colored, about 1-12 of an inch long; 5 stamens, nearly as long as the calyx; pistils simple with 2 slender styles producing a single obconical depressed seed, dull gray or green, without albumen; embryo spirally coiled. The plant flowers in July or August.

Distribution. Common from Minnesota to Kansas, west across the continent, Illinois and Kansas to New Jersey.

Injurious properties. The Russian thistle not only clogs the harvesters and harrow, injures horses legs so that boots have to be put on them but is equally disagreeable to come in contact with, to man. On this point, Prof. Dewey says:

The sharp spines on the plants not only irritate and worry both horses and men, but often, by breaking under the skin, cause festering sores on the horses' legs, so that in many localities it has been found necessary to protect them with high boots or leggings. In handling grain or flax, in the processes of hauling and threshing, the sharp spines cause considerable irritation and consequent loss of time.

AMARANTHACEAE. Amaranth Family

Herbs, or in some cases, shrubs; leaves simple, mostly entire; flowers small, green or white with bractlets, usually in terminal spikes or heads; petals none; calyx herbaceous or membranous, 2-5 parted; segments distinct or united;



Fig. 207. Russian Thistle (Salsola Kali, var. tenuifolia). Causes mechanical injuries to man and stock. (Charlotte M. King.)

stamens 1-5, mostly opposite the calyx-segments; ovary 1-celled; ovules solitary; fruit an utricle; circumseissile, irregular or indehiseent; seed generally smooth; endosperm usually copious and mealy. About 425 species in tropical countries mostly. Several like *Celosia cristata* are cultivated for ornamental purposes and several are weedy. Among the latter are the tumble-weed (*Amaranthus graecizans*), pig-weed (*A. retroflexus*), and prostrate pig-weed (*A. blitoides*). The leaves of several species are used as food.

Amaranthus (Tourn.) L. Pig-weed. Tumble-weed

Annual, branching or creet herbs, smooth or pubescent leaves, simple; small flowers, monoecious, dioecious, or polygamous, green or purplish, generally with 3 bractlets; in spikes or axillary clusters; sepals 3-5, distinct; stamens 2-5; styles 2-3; fruit oblong, utricle. About 50 species of wide distribution, mostly of southern states.

Amaranthus retroflexus L. Pig-weed. Red Root

Roughish, slightly pubescent, annual with stout stems 2-4 feet high; leaves ovate or rhombic ovate; upper lanceolate, acute or acuminate at apex; flowers in dense spikes; bractlets about twice as long as the 5 scarious mucronate-tipped sepals; stamens 5; seed black.

Distribution. Naturalized from tropical America; found throughout the United States, especially on waste ground far northward. Also naturalized in Europe.

Amaranthus hybridus L. Slender Pig-weed

Similar to the preceding but with darker green or purple foliage; stcm more slender, erect; leaves ovate or rhombic ovate, smaller than the preceding;

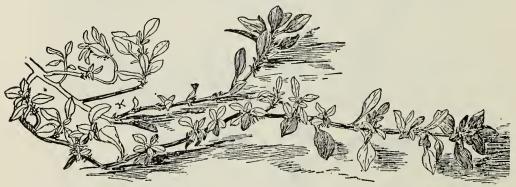


Fig. 208. Prostrate Pigweed (Amaranthus blitoides). Λ common weed. May cause bloat. (Charlotte M. King.)

spikes linear-cylindrical, forming dense terminal panicles; bracts subulate, twice as long as the acute or cuspidate sepals; stamens 5; utricle but slightly wrinkled.

Distribution. Species naturalized from tropical America but rare or local in places; common southward.

Amaranthus spinosus L. Spiny Amaranth

Stout, branched stem, leaves ovate, rhombic-ovate or lanceolate, acute at both ends with a pair of rigid stipular spines; scpals mucronate-tipped 1-nerved; utricle scarcely circumscissile.

Distribution. In waste or cultivated ground as far north as Massachusetts, Illinois and common in Missouri and Southward. Naturalized from tropical America.

Poisonous properties. The spiny amaranth sometimes produces mechanical injuries. Mr. O'Gara calls attention to the injurious properties of the first species in Nebraska. He says that it doubtless causes a great deal of trouble in some parts of that state. Mr. C. C. Palmer near North Platte lost 5 head of cattle in his pasture. In all cases they were very much bloated and a postmortem examination revealed a good deal of pig-weed in the stomachs. The animals in question had been accustomed to prairie grass pasture and broke into a field containing considerable of this pig-weed, some Russian Thistle and



Fig. 209. Pigweed (Amaranthus retroflexus). A cause of bloat in cattle. (From Darlington's Weeds and Useful Plants.)

Fig. 210. Spiny Araranth (Amaranthus spinosus), Sometimes produces mechanical injuries. (From Darlington's Weeds and Useful Plants.)

lamb's quarter. After the death of the animals the fence was replaced and no further trouble was noticed. An experiment with animals carried on by Mr. O'Gara proved negative. Many families in the vicinity of North Platte regard the weed as a bad bloater, by some considered as serious as green clover and alfalfa.

PHYTOLACCACEAE. Pokeweed Family

Generally herbs, a few tropical species, trees or shrubs; leaves alternate, entire, without stipules; flowers regular, perfect, polygamous or monoecious; calyx petal-like, of 4 or 5 sepals, or 4-5-parted; stamens 5-30 alternate with the segments of the calyx or with the sepals, of the same number or more numerous; ovary several-celled; ovules solitary.

A small family of about 85 species, mostly tropical. The juice of the berries of the Umbra tree (Phytolacca dioica) of South America, now naturalized in Europe and other warm countries, is used to color wines. Other plants of the family, like bloodberry are frequently cultivated. The latter produces small spikes of white flowers, followed by red berries. Strong drastic substances occur in P. littoralis and Anisomeria drastica, natives of Chili. P. abvssinica. Villamilla peruviana and our native pokeberry contain saponin and red coloring matter.

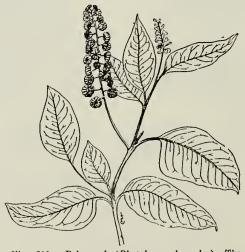
Phytolacca (Tourn.) L. Pokeweed

Tall, stout, perennial herbs with large petioled leaves; flowers borne in racemes; calyx of 5 petal-like sepals; stamens 5-30, ovary of 5-12 carpels united to form a ring, 5-12 celled, with a single seed in each cell; embryo around the endosperm.

Phytolacca decandra L. Pokeweed, Garget

A tall, glabrous, perennial herb, 6-9 feet tall, with strong odor; large poisonous root; leaves oblong-lanceolate, acute, or acuminate; flowers perfect; calyx white; stamens 10, shorter than the sepals; ovary green, 10-celled; berry dark purple, filled with crimson juice.

Poisonous properties. The young shoots of this plant may be boiled and eaten, the acrid property being dissipated in boiling. The leaves are eaten by the natives of the island of Guam. A tincture of the plant is used for rheumatism. The root is alterative, emetic, cathartic, and narcotic. Prof. Chesnut, in speaking of its poisonous nature, says:



Pokeweed (Phytolacca decandra). root of the plant is very poisonous. (C. M. King.)

Most instances of poisoning arise from an overdose when the plant has been used as a medicine, but there are also accidental cases due to eating of the root, which has been variously mistaken for that of the parsnip, artichoke and horseradish. A few fatal cases of poisonoing of children have been attributed to the fruit, but whether death was really due to the seed or the pulp is somewhat uncertain. The evidence is chiefly against the

seed, for it is known to contain a poisonous substance. Pokeweed is a violent but slow-acting emetic, vomiting beginning only after about two hours. It also effects the nerves and muscles, producing retching, spasms, severe purging and sometimes convulsions. Death is frequently due to the paralysis of the respiratory organs.

Dr. Guttenberg makes a similar report in regard to effects of poisoning by pokeweed, adding that death often is a result.

The roots of pokeweed are often mistaken for other fleshy roots, such as horse-radish. The leaves, as has been said, are harmless when boiled, somewhat resembling spinach, but the root is very poisonous. The poke root was used by the Indians in medecine. Dr. Millspaugh, who values the plant not only as an emetic, but also as an efficient remedy, says:

In certain forms of rheumatism, the root with lard was found to be an excellent ointment as a cure for many forms of skin diseases; psoriasis, eczema, capitis, and tinea circinata, also in syphilitic ulcers.

Dr. Millspaugh says:

The fresh root, gathered late in autumn or early in spring, is chopped and pounded to a pulp and weighed. Two parts hy weight of alcohol are taken, and after thoroughly mixing the pulp with one-sixth part of it, the rest of the alcohol is added. After having stirred the whole, pour it into a well-stoppered bottle, and let it stand eight days in a dark, cool place. The tincture is then separated by decanting, straining, and filtering.

Thus prepared it has a light straw-color by transmitted light, at first a stinging, soon followed by a decided bitter taste, and a very slight acid reaction.

He adds:

I noted in my readings several years ago that the berries had been used for pies by frugal housewives, and often since have half determined to try poke-berry pastry; discretion has, however, always overruled valor, and the much-thought-of pie is still unmade and uneaten. The young shoots, however, make an excellent substitute for asparagus, and I much prefer them, if gathered early and discriminately.

The aerid alkaloid phytolaccin, according to Dr. Edmond Preston, occurs in the root of this plant; also phytolaccic acid and an amorphous yellowish brown, transparent substance, very soluble in water and alcohol. Nagi reports a toxic substance phytolaccotoxin C₂₄H₃₈O₈. The berries have been used for coloring, but this is not entirely successful, because no mordant will fix the color. The juice of the berry is a delicate test for acids when lime water is added to it.

Dr. Johnson says:

All parts of the plant possess acrid and somewhat narcotic properties. The juice of the fresh plant, or a strong decoction of the root, applied locally, may strongly irritate the skin, especially if tender or abraded. Taken internally it causes nausea, vomiting, and purging, and, in overdoses, acro-narcotic poisoning. It bas been employed with more or less satisfactory results in a great variety of cutaneous affections, and in rheumatism, especially when chronic or of a syphilitic origin. There is little doubt that, in view of the uncertainty which at present exists regarding it, this plant would well repay further careful experimentation.

Nagi reports that *phytolaccotoxin* resembles *picrotoxin* and *cicutoxin*. A glucoside has also been found in common poke; saponin also occurs.

CARYOPHYLLACEAE. Pink Family

Herbs with opposite entire leaves, frequently swollen at the nodes; flowers perfect or rarely dioecious; sepals 4 or 5, persistent separate or united with the calyx tube; petals of equal number; styles 2-5, or rarely united into 1; ovary usually 1-celled, occasionally 3-5-celled; ovules attached to a central column; seeds several or many; small coiled or curved embryo, with a mealy albumen.

A large family of about 70 genera and 1500 species, widely distributed, most abundant in the northern hemisphere. Many of the plants of this family

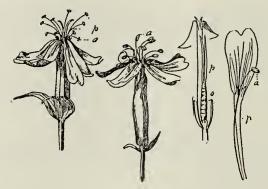


Fig. 211a.

Fig. 211a. Flowers of Soapwort or Bouncing Betty (Saponaria officinalis), Calyx, Corolla, Stamens and Pistil. (C. M. King).

are cultivated for ornamental purposes. Of these we may mention the hardy pink (Dianthus barbatus) and carnation (D. Caryophyllus). The spurrey (Spergula arvensis) is occasionally cultivated as a forage plant in Europe and sometimes in this country, but is a weed of grain fields in Europe. A few species like Saponaria officinalis and the catchfly are medicinal. A red dye is obtained from a species of Coccus found on Scleranthus perennis. The leaves of Paronychia argentea are used as a substitute for tea. The stitchwort (Alsine crassifolia) of Europe and some parts of the United States is poisonous to horses. The European sandwort (Arenaria serpyllifolia) common eastward in sandy waste places is said to cause salivation in horses. Several species of the family like Saponaria officinalis, Gypsophila Struthium of Spain, Agrostemma, Lychnis, and Herniaria contain saponin.

Genera of Caryophyllaceae

Sepals united into a tube or cup.

Calyx ovoid or sub-cylindrical, 5 angled; not prominently nerved..5. Saponaria. Calyx 5-toothed, prominently nerved.

| Styles 3 | 2. | Silene. |
|--|------|-----------|
| Styles 5 or 4, alternate with petals | .3. | Lychnis. |
| Styles 5 or 4, opposite petals, silky plants4. | Agro | stemma. |
| Styles 21. | Gy | psophila. |
| distinct | 5 9 | Stallaria |

1. Gypsophila L. Gypsophyl

Glabrous and glaucous herbs; leaves narrow; flowers small, in paniculate, axillary clusters; calyx cylindrical, 5-toothed, 5-nerved without bractlets; petals 5 claws, narrow; stamens 10; styles 2. About 50 species native to Europe; 2 species introduced to North America.

Gypsophila paniculata L. Tall Gypsophyl

A glabrous or pubescent perennial, from a simple fusiform root; leaves lanceolate, narrowed at the base; flowers in panicled cymes; calyx campanulate;

segments with scarious margins; petals white or pink, slightly emarginate, larger than the calyx.

Distribution. Native to Europe and Asia. From Manitoba to Nebraska.

Poisonous properties. Used in medicine as a detergent. An allied G. Struthium contains Sapotoxin and the glucoside saponin. It is an acrid poison.

2. Silene L. Catchfly

Herbs with pink or white flowers, solitary or borne in cymes; calyx more or less inflated and five-toothed; petals 5, narrow and clawed; stamens 10; styles 3, rarely 4 or 5; ovary 1-celled or incompletely 2- to 4-celled; pod 1-celled, dehiscent by 6, apical teeth; seeds roughened.

About 250 species of wide distribution. Several like sweet William (S. Armeria) are cultivated for ornamental purposes. The starry campion (S. stellata) of our prairies and thickets might well be cultivated more than it is.

Silene latifolia (Mill.) Britten & Rendle. Bladder Campion

A branched perennial, a foot or more high, with opposite glaucus ovate lanceolate leaves; flowers in loose cymose panicles; calyx bladdery, inflated; petals 2-cleft, white; seed roughened.



Fig. 212. Deptford Pink (Dianthus Armeria). Frequently cultivated for ornamental purpose. (Charlotte M. King.)



Fig. 213. Night flowering Catchfly (Silene noctiflora). (After Fitch.)

Distribution. Native to Europe, Common in fields and along roadsides from New England to Illinois and Iowa.

Silene antirrhina L. Sleepy Catclifly

A puberulent annual with glutinous nodes and slender stem; lower leaves spatulate or oblanceolate, petioled; upper leaves linear to subulate; flowers small in cymose panicles; calyx not inflated, but expanded by the opening pod, ovoid; petals pink, obcordate, minutely crowned, seeds small, roughened.

Distribution. Common in sandy fields, gravelly soils, and in waste places from New England and Florida to Mexico, north to British Columbia, and east to Ontario.

Silene noctiflora L. Night-flowering Catchfly

A viscid hairy annual, from 1-3 feet high; lower leaves obovate or oblanceolate; the upper sessile and lanceolate; flowers few, in a loose panicle, white or



Fig. 214. Sleepy Catchfly (Silene antirrhina). A weed of sandy fields. (Charlotte M. King.)

Fig. 215. Night-flowering Catchfly (Silene noctiflora). Possibly poisonous. (Charlotte M. King.)

pinkish, fragrant, opening at night, calyx tube elongated and enlarged by the ripening pod; petals 2-cleft and crowned; seeds small blackish, roughened, kidney shaped.

Distribution. Native to Europe. Common in waste places from New Brunswick to Florida, Kansas and Iowa to Manitoba.

Poisonous properties. According to Stebler and Schröter, the leaves of Silene latifolia are eaten by stock and it is regarded as of some value for forage purposes; but Prof. Schaffner, in his "Poisonous and Other Injurious Plants of Ohio," suggests that the sleepy catchfly may be poisonous.

3. Lychnis (Tourn.) L. Campion

Erect herbs, with ovoid tubular oblong or inflated calyx 5-toothed, 10-nerved, occasionally with leaf-like lobes; petals 5, or rarely 4; styles 5, rarely 4, alternate with the often appendaged petals; seeds numerous, globular or kidney-shaped pod opening by as many, or twice as many valves. A small genus of about 40 species native to the cooler regious. Several species cultivated for ornamental purposes. The scarlet lychnis (*L. chalcedonica*) is frequently cultivated in old gardens.

Lychnis Flos-cuculi L. Ragged robin

A downy, branching, pubescent annual, or viscid above, from 2-3 feet high; leaves lanceolate or linear lanceolate; flowers in loose panicles, red, bluish, or whitish; calyx glabrous, short, petals cleft into 4 lobes; capsule globose.

Poisonous properties. It contains a form of saponin called lychnidin.

Lychnis dioica L. Evening Lychnis

Bicnnial, usually dioccious, viscid, pubescent; leaves ovate-oblong or ovate-lanceolate; flowers few, loosely paniculate, white or pinkish, opening at evening; calyx tubular, becoming swollen with the ripening fruit; styles 5.

Distribution. Native to Europe, common in eastern and middle states. In the West it is not uncommon in clover fields, where it is introduced with clover sced.

4. Agrostemma Linn. Corn Cockle

Calyx ovoid, 10-ribbed; teeth elongated, longer than petals; stamens 10; styles 5, opposite unappendaged petals; leaves linear. Tall annual or biennial plant.

Agrostemma Githago L. Corn Cockle

A hairy annual weed; leaves linear-lanceolate, acute or long-acuminate; flowers perfect, long-peduncled, calyx lobes long, linear, surpassing the purple red petals, capsules 1-celled; large with numerous large seeds which are roughened and black.

Distribution. This plant is widely distributed from Nova Scotia to Quebec, south from New England to the southern states, and westward and northward, generally in wheat growing regions. Difficulty in screening wheat by ordinary methods has caused this weed to be generally scattered in wheat growing regions. These screenings are much used in feeding stock in some places. The farmer often sows cockle with his wheat.



Fig. 216. Meadow Lychnis (Lychnis Flos-cuculi). Contains saponin. (After Fitch.)

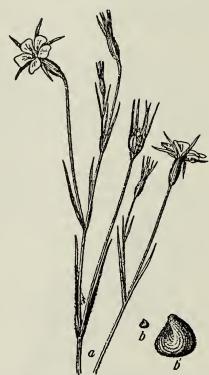


Fig. 217. Corn cockle (Agrostemma Githago). a, sprays showing flowers and seed capsule, one-third natural size; b, seed, natural size; b', seed, four times natural size. (U. S. Dept. Agr.)

Poisonous properties. According to Kruskal, the seeds contain githagin 2(C₁₇H₂₈O₁₁). The ripe dried seeds are broken into a coarsc powder and used in medicine. Dr. Millspaugh gives the proportions as follows: "Five parts by weight of alcohol are poured upon the powder, and the whole allowed to stand cight days in a well stoppered bottle in a dark cool place, shaking thoroughly twice a day." The tincture is somewhat acrid. The seeds of the cockle are frequently used to adulterate cheaper grades of flour in Europe. Dr. Millspaugh gives a case in which death followed where two 141/2 oz. lots of wheat flour containing respectively 30% and 45% of these seeds were fed to two calves. This amount of cockle caused severe cramps of the stomach within an hour, followed by diarrhoea and finally death. Where ducks and geese ate the seeds, death followed when sufficient was taken, and the post-mortem showed inflammation of the bowels. Prof. Pierce states that this is especially true when the secds are crushed. A large amount of screenings are sold for chicken feed, and frequently complaints are made of poison, or at least that chickens will not cat the screenings.

In describing symptoms indicative of poisoning by corn cockle, which, Dr. Allen says, place the seeds among the cerebro-spinal irritants, he agrees essentially with Dr. Chesnut.

Dr. Chesnut says:

The poisonous constituent, saponin, is a non-crystalline powder, very freely soluble in water, and possessing a sharp, hurning taste. It has no odor, but when inhaled in the smallest quantity it produces violent sneezing. When briskly sbaken with water it froths like soap. The poison is found in nearly all parts of the plant, but mainly in the kernel of the seed. Cases of poisoning have been noted among all sorts of poultry and household animals, but are rarely due to any portion of the plant as found growing in the field. The poisoning is generally produced by a poor grade of flour made from wheat containing cockle seeds. Machinery is used to remove these seeds from the wheat, but the difficulty of separating them is so great that the result is not entirely accomplished. The quantity remaining determines the grade of flour in this particular regard. It sometimes amounts to 30 or 40 per cent, but this quality is sent out only by ignorant or unscrupulous dealers or is intended for consumption by animals only. Flour containing a smaller amount has often heen made into bread and eaten, sometimes with fatal results, the haking not always heing sufficient to decompose the poison. The effect may he acute, or, if a small quantity of the meal is eaten regularly, it may be chronic. In the latter case it is sometimes known as a disease under the name of "githagism." The general symptoms of acute poisoning are the following: Intense irritation of the whole digestive tract, vomiting, headache, nausea, diarrhoea, hot skin, difficult locomotion, and depressed breathing. Coma is sometimes present, and may be followed by death. Chronic poisoning has not been closely studied in man, hut experiments upon animals show chronic diarrhoea and gradual depression, the animal losing vigor in breathing and in muscular movements until death ensues. The action is antagonized by the use of digitalin, or of the simple extract of digitalis (Digitalis purpurea) a dangerous poison, which should be given only by a physician.

The more prominent symptoms as recorded by Friedberger and Fröhner are, briefly, colic, vomiting, slavering, paralysis, stupor, hyperaemia of brain and spinal cord.

Dr. Chesnut also adds:

Corn cockle meal is easily detected in second and third class flour by the presence of the hlack, roughened scales of the seed coat. These are sure to occur if the flour has not heen well bolted. Its presence is otherwise detected by the peculiar odor produced when the meal is moistened and by chemical tests with iodine. Wheat containing corn cockle seeds should be rejected for planting.

It has been asserted in Europe that corn cockle is injurious in flour and bread stuffs. Dr. Chesnut says:

A person eating 1200 grains of bread made from flour containing only one-half per cent of corn cockle seed would consume six grains of cockle seed, an amount which the author believes beyond a doubt to he poisonous in its effects.

The poison in corn cockle is $sapotoxin\ C_{17}H_{20}O_{10}$ and is partially decomposed while baking, but nevertheless some of it remains and the use of flour which contains corn cockle should be forbidden. It has long been suspected of being poisonous. Mr. John Smith in his Domestic Botany, says:

It heing difficult to separate the seeds from the grain, the value of the latter is deteriorated, and the flour is rendered unwholesome.

5. Saponaria Linn. Soapwort

Calyx ovoid to sub-cylindrical, 5-toothed, obscurely nerved, terete or 5-angled, smooth; stamens 10; styles 2; pod 1-celled, or sometimes 2-4-valved, and 4-toothed to apex. Coarse annual or perennial with mucilaginous juice, hence common name of soapwort because of the property of forming a lather with water.

Saponaria officinalis L. Bouncing Bet

Percnnial herbs with large flowers in cymose clusters; calyx narrowly ovoid or oblong, five toothed; petals clawed or unappendaged, stamens 10, styles 2, pod 1-celled or incompletely 2 to 4-celled and 4-toothed at the apex. About

40 species in Europe, Asia, and Northern Africa. Saponaria officinalis is frequently cultivated in old gardens. The mucilaginous juice forms a lather with water and is valuable for taking grease spots out of wollen cloth.

Saponaria Vaccaria L. Cow herb

A glabrous annual from 1-2 feet high with opposite ovate lanceolate leaves; flowers in corymbed cymes; calyx 5-angled, enlarged and angled in fruit; petals pale red. Cow herb is another important constituent of "cockle" in wheat screenings, and like the preceding weed has been largely spread by means of wheat culture.

Distribution. Common in Europe; found in wheat fields of the east and as far west as Missouri, Kansas, the Rocky Mountain region, and Pacific Coast, and wheat regions of the northwest.

According to Sohn, it contains the substance saponin, $C_{22}H_{54}O_{18}$ a neutral sharp, amorphous substance, having a burning taste and producing a violent sensation. The toxic substance is partially removed by baking.



Fig. 218. Bouncing Betty (Saponaria officinalis). A branch with flowers. The double flowered form is sometimes cultivated for ornamental purposes. (Charlotte M. King.)

6. Stellaria L. Chickweed

Tufted herbs with white flowers in cymose clusters; sepals 4-5, deeply 2-cleft, sometimes none; stamens free, 10 or fewer; styles 3, rarely 4 or 5; capsule ovoid 1-celled, several to many seeded.

Stellaria media (L.) Cyrill. Common Chickweed

A nearly smooth annual or winter annual, decumbent or ascending; leaves ovate or oval, the lower on hairy petioles; flowers white in terminal leafy cymes or solitary in the axils; sepals oblong, longer than the 2-parted petals; stamens 2-10.

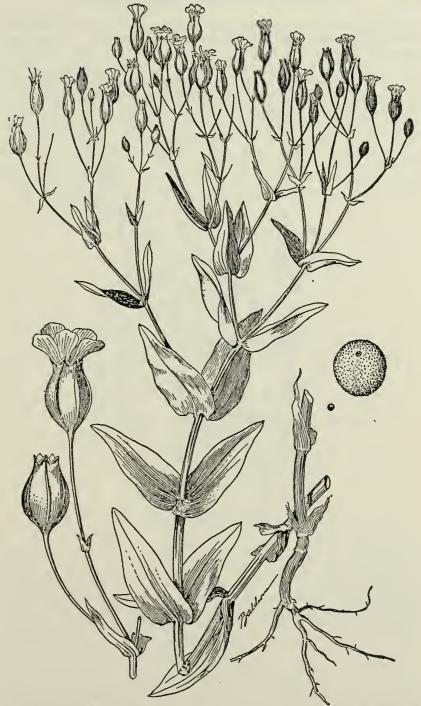


Fig. 219. Cow-herb (Saponaria Vaccaria I.). The seed of this plant is common in wheat screenings and is supposed to be poisonous. (U. S. Dept. Agrl.)

Distribution. A weed in waste places, lawns and fields, very troublesome in lawns. Naturalized from Europe. Extends from New England and Canada across the continent.

Poisonous properties. The seeds of common chickweed are used as food for cage-birds and are also readily eaten by chickens, but, according to Mr. Wm. Carruthers, they cause disorder to the digestive system when eaten by lambs in large quantities.



Fig. 220. Chickweed (Stellaria media). Seeds said to be injurious. (Charlotte M. King.)

RANALES

Herbs, shrubs or trees; calyx usually of separate sepals; corolla usually present and of separate petals; ovary or ovaries superior, free from the calyx; carpels 1-many; stamens mostly hypogynous and more numerous than the sepals. Contains the families Nymphaeaceae, Ceratophyllaceae, Ranunculaceae, Berberidaceae, Menispermaceae, Magnoliaceae, Calycanthaceae, Anonaceae, Myristicaceae, and Lauraceae. The Nymphaeaceae are aquatic perennial herbs. The rhizome of water chinquapin (Nelumbo lutea) of the Mississippi Valley and introduced into Massachusetts by the Indians, was used for food. The sacred bean or lily (N. nucifera) cultivated for ornamental purposes produces an edible seed and rhizome rich in starch. The pods of wokas (Nymphaea polysepala) are

used as food by the Indians in the northwest. The blue flowered Nymphaea stellata of tropical Africa and the Egyptian lotus (N. Lotus) are frequently cultivated, as are the Victoria regia of the Amazon region and Euryale ferox of eastern Asia. The water lilies (Castalia odorata and C. tuberosa) are pretty water plants of North America. The family Ceratophyllaceae contains the water-weed (Ceratophyllum demersum) of North America, troublesome also in Europe. The family Myristicaceae contains the nutmeg (Myristica fragrans) of which mace is the aril; the oil contains myristicin. Prof. Cushny in an address before the Royal Society of Medicine in London, referring to Nutmeg poisoning, says that the symptoms are drowsiness, stupor, and diplopia ('seeing double'). Delirium is frequently present, and sometimes the first symptom is burning pain in the stomach, with anxiety or giddiness. The symptoms generally resemble those resulting from Cannabis indica (hashish). One fatal case oc-

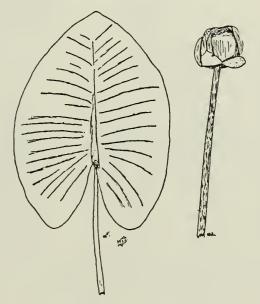


Fig. 221. Yellow Water Lily (Nymphaea polysepala). (W. S. Dudgeon.)

curred in a boy who had eaten two nutmegs. From experimental work Prof. Cushny has come to the conclusion that the symptoms are to be attributed to the action of the oil of nutmeg on the central nervous system. This is depressed; but there are some signs of stimulation in the form of restlessness, slight convulsive movements, and tremor. The oil has also a marked local irritant action, whether given by the mouth or hypodermically. Several other species like M. succedanea and M. fatua, are used by the natives where these plants are indigenous.

Families of Ranales

Stamens numerous sepals distinct, petals absent or present.

Receptacle not hollow; flowers generally perfect.

Fruits cohering over each other, cone-like..............Magnoliaceae Fruits not cohering over each other, separate.

RANUNCULACEAE. Crowfoot Family

Herbs or a few woody plants with acrid juice; flowers polypetalous or apetalous, regular or irregular; calyx free, often colored like the corolla; sepals 3-15; petals 3-15 or absent, stamens numerous; pistils few or numerous, distinct; fruit a dry pod, berries or achene seed-like; embryo minute, albumen present.

A rather large, widely distributed family of plants many of which like aconite, larkspur, and marsh marigold, are poisonous. Many, such as virgin's bower (Clematis virginiana), C. Jackmanni and other species, are cultivated for ornamental purposes; the C. Jackmanni being especially desirable. The columbines, like the European columbine (Aquilegia vnlgaris), the Rocky Mountain columbine (A. caernlea), and our eastern columbine A. canadensis), the paeonies (Paeonia officinalis and P. Moutan), and the larkspurs (Delphinium) are also cultivated for ornamental purposes, the most familiar of the latter being the garden annual, Delphinium Consolida. Several perennial species of Delphinium are also very showy. The seeds of stavesacre (D. Staphisagria), native to southern Europe and the Levant, contain an alkaloid delphinin $C_{22}H_{35}NO_6$ which is a powerful and acrid poison.

Nigella is said to contain an alkaloid, nigellin; N. damascena contains the alkaloid damascenin $\rm C_0H_{11}NO_3$. In 1872, a German chemist found an alkaloid in Isopyrum thalictroides, the so-called isopyrin $\rm C_{28}H_{11}NO_3$. Thalictrum macrocarpum contains the alkaloid thalictrin. Aquilegia is said to be free from alkaloids.

Glucosides have also been found in some of the members of this family, as adonidin $C_{40}H_{40}O_{10}$ in Adonis amurensis.

The European Adonis aestivalis and A. vernalis are recorded as poisonous by Lehmann. The black cohosh (Cimicifuga racemosa) is also somewhat acrid.

Many of the plants of the family contain anemonin $C_{10}H_8O_4$ and some are used for medicinal purposes.

Among these are aconite (Aconitum Napellus), crocus, Pasque flower, pulsatilla Anemone patens var. Wolfgangiana (Bess) Koch. and yellow puccoon (Hydrastis canadensis). The black roots of black hellebore (Helleborns niger) are used in medicine, as a purgative, being poisonous in overdoses. The tuberous roots of one of the crowfoots (Ramnculus Ficaria) resemble grains of wheat and are sometimes boiled and eaten but they have a sharp acrid taste and are known to produce blisters. The water crowfoot (R. aquatilis var. capillaceus) is apparently harmless and is used as forage in England and on the Continent.

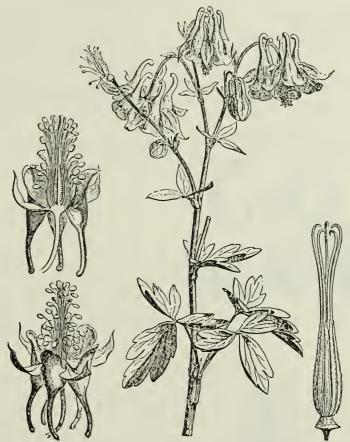


Fig. 222. European Columbine (Aquilegia vulgaris). Flowering branch; flower; longitudinal section of flower; pistil and stamens. (After Faguet.)

According to Greshoff Clematis Fremonti; C. integrifolia; C. lanuginosa, C. orientalis; C. pseudo-flammula contain HCN. He also states that saponin is of widespread occurrence in this genus and that he found it in the leaves of C. Pitcheri, and C. recta and in the leaves of Trollius pumilus, and T. chinensis.

Genera of Ranunculaceae

| Flowers regular. | |
|---------------------------------|--------------|
| Sepals 3-20 apetalous. | |
| Achenes tailed | 5 Clematis |
| Achenes not tailed | 4 Anemone |
| Fruit a follicle; sepals yellow | 1 Caltha |
| Flowers solitary; sepals 3 | 9 Hydrastis |
| Petals and sepals present. | |
| Petals 5, yellow or white | 6 Ranunculus |
| Petals small, tubular 2-lipped | Helleborus |
| Petals small, stamen-like | |

Flowers irregular.

| Upper sepal | spurred; petals 4 | Delphinium |
|-------------|-------------------|------------|
| Upper sepal | hooded3 | Aconitum |

1. Caltha L. Marsh Marigold

Herbs with heart-shaped or kidney-form leaves; flowers yellow, white or pink; sepals large, 5-9, petal-like; petals none; stamens numerous; pistils 5-10; styles nearly wanting; pods follicles, spreading, many seeded; marsh plants of temperate and colder regions. About 8 species; 3 species native to Eastern North America, and 1 species common in the Rocky Mountains at high altitudes.



Fig. 223. Black Hellebore (Helleborus niger). Entire plant, The roots contain a purgative substance that is poisonous in over-doses, (From Vesque's Traité de Botanique.)

Caltha palustris L. Marsh Marigold

A stout, glabrous perennial with a hollow stem from 1-2 feet high; the basal leaves on long petioles, leaves reniform; upper leaves shorter, petioled and sessile; flowers with yellow sepals.

Occurs in swamps and meadows.

Poisonous properties. The marsh marigold or cowslip is regarded as poisonous in Europe. In this country, however, it is frequently used as a pot herb. The flower buds are sometimes pickled. Coville says:

By many it is considered superior to any other plant used in this way. There is no doubt that boiling dissipates the active principles found in the plant.

Stebler and Schröter say that it is poisonous in a green state, and Rusby states that when fed with hay it produces diarrhoea and stoppage of the flow of milk. According to Lloyd, it contains a small quantity of an acrid substance identical with the acrid oil of Ranunculus. Cattle and sheep refuse

to eat the plant. Marsh marigold is known to contain an alkaloid which is said to be identical with *nicotin* but it has not been isolated.

Dr. Millspaugh in speaking of the uses of this plant, states that it is extensively gathered in early spring and cooked for greens, making one of our most excellent pot-herbs. Rafinesque asserts that cattle browsing upon it die in consequence of an inflammation of the stomach produced by it. According to Freidberger and Fröhner it causes haematuria.

2. Helleborus L. Hellebore

Erect perennial herbs, with large, palmately divided leaves; flowers large, white, greenish or yellowish; sepals 5, petal-like; petals small, tubular; stamens numerous; carpels generally few; fruit several-seeded follicles. A small genus of about 10 species, natives of Europe and Western Asia.

Helleborus viridis L. Green Hellebore

Basal leaves smooth, consisting of 7-11 oblong, acute, sharply-serrate segments; flowers large.

Distribution. In waste places from Long Island to Penn, and W. Va.

Poisonous properties. Black Hellebore is said to be a drastic purgative when used for domestic animals. The plant contains the glucosides, helleborin $C_6H_{10}O$, which is a highly narcotic, powerful poison, helleborein $C_{37}H_{66}O_{18}$ which is slightly acid and heleboretin $C_{19}H_{30}O_{5}$.

The symptoms from poisoning are: Stupor followed by death with spasms. H. foetidus is also poisonous.

3. Aconitum L. Aconite

Perennial herbs with palmately lobed or divided leaves; flowers large, irregular, showy, paniculate or racemose; sepals 5, irregular, petal-like, the upper helmet-shaped or hooded, prolonged into a spur; petals 2, small, concealed under helmet, spurred, 3 lower absent or very minute; pistils 3-5 forming follicles, several seeded.

About 60 species. Native of the North Temperate regions. One species, the A. Napellus, used in medicine, is the source of aconite. One western species is poisonous to live stock.

None of the species of this genus is weedy in Eastern North America. The three species, A. noveboracense Gray, A. uncinatum L. and A. reclinatum Gray, occur in Eastern North America but are very local. None of these seems to be very poisonous, but the roots of A. uncinatum are bitter, even in a dry state. The exotic A. Lycoctonum and A. Fischeri are employed to kill wild animals.

Aconite is derived from the European Aconitum Napellus which produces not only poisonous stem and leaves, but also a very poisonous tuberous root which is from 2-4 inches long and sometimes an inch thick. The Indian aconite is obtained from Aconitum ferox, a plant growing from 3-6 feet high and bearing large dull-blue flowers; it is found in the Alpine regions of the Himalayas, and is used as an arrow poison. Among other equally poisonous species mentioned by Flückiger and Hanbury, is A. uncinatum growing in Eastern North America. They also state that the root of another species of Aconitum, A. heterophyllum, with large flowers of dull yellow, and purple, or blue, is poison-

ous. This root contains the chemical substance atisin $C_{22}H_{31}NO_3$, an intensely bitter alkaloid. The European aconite contains aconitin $C_{33}H$ NO $_{12}$, pseudo aconitin $C_{36}H_{49}NO_{12}$, and aconin $H_{39}C_{20}NO_{11}$. The North American species A. septentrionale contains, according to Rosendal, the following alkaloids: lappakonitin, $C_{34}H_{48}N_2O_8$, a crystallized form; septentrionalin $C_{31}H_{48}N_2O_8$, and synaktonin $C_{38}H_{55}N_2O_{13}$.

Blyth who has collected records of poisoning in Europe by aconite states that there have been two cases of murder, seven suicidal, seventy-seven more or less accidental; six were from the action of the alkaloid, ten from the root, and in two cases, children ate the flower, in one case, the leaves of the plant were cooked and eaten by mistake, in seven cases, the tincture was mistaken for brandy, sherry or liquor, in the remainder of the cases the tincture, the liniment or the extract was used. The Indian species are much used, especially A. ferox, which is applied to poison stock and arrows, the latter to destroy the wild animals. It is a common practice to mix a decoction of the root with water or food.

Dunstan and Anderson * summarize the alkaloids obtained from Aconitum as follows:—"The first, a toxic group, of which the type is ordinary aconitin, contains alkaloids which are diacyl esters of a series of poly-hydric bases containing four methoxyl groups, the aconines."

"The members of this group are:—
Aconitin from Aconitum napellus.
Jupaconitin from Aconitum deinorrhysum.
Bikhaconitin from Aconitum spicatus.
Indaconitin from Aconitum chasmanthum."

These are all highly poisonous.

The second group is the atisin group which contains atisin from A. heterophyllum and palmatin from A. palmatum. These are non-poisonous alkaloids.

Aconitum columbianum Nutt. Western Aconite

An erect, stout perennial, 3-6 feet high, more or less pubescent above, with short, spreading or viscid hairs; divisions of the leaves broadly cuneate and toothed, lobed; flowers purple or white, in a loose terminal raceme; hood variable in breadth and length of beak.

Distribution. Grows at an altitude of 5000-10,000 feet in low grounds, near brooks and springs, from Montana, Wyoming and Colorado to the Sierras.

Poisonous properties. The chief effect of aconite results from its influence over the heart and blood vessels. It decreases the force and frequency of the cardiac pulsations. After long continued use, aconite affects the nervous system causing the loss of sensation; bodily temperature is also reduced after medicinal uses of the drug. Dr. Winslow, in his Veterinary Materia Medica and Therapeutics, speaking of its toxicology says:

The minimum fatal dose of aconite is about 3i. for the horse; gr. xx. for medium sized dogs; and gr. v.-vi. for cats. The smallest fatal dose recorded in man is a teaspoonful of tincture of aconite, equivalent to about gr. xxx. of the crude drug. The minimum lethal quantity of aconitin is gr. 1/10 for man, and about the same for cats. For dogs it is from gr. ½ to gr. ½. The writer has found that cats will live from fifteen minutes to half an hour after receiving the smaller deadly doses under the skin, but large doses produce death immediately by paralyzing the heart. Large therapeutic doses cause, in horses, restlessness, pawing the ground, shaking of the head, champing of the jaws, increased secretion of salivary mucus, and attempts at swallowing, probably owing to the peculiar sense of irritation

^{*}Trans. Jour. Chem. Soc. 1905: 1650. See Blyth Poisons: Their Effects and Detection.



Fig. 224. Aconite (Aconitum Napellus). The source of the aconite of commerce. Stem, leaves and root are poisonous. (After Faguet.)

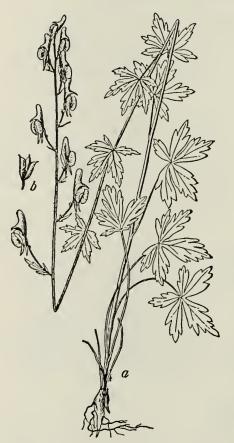


Fig. 225. Aconite (Aconitum columbianum). a, flowering plant; b, seed capsule—both one-third natural size. A poisonous plant of the Western U. S. (U. S. Dept. Agr.)



Fig. 226. Western aconite (Aconitum uncinatum). A poisonous plant in Eastern North America, from Pennsylvania to Iowa. (Ada Hayden.)

produced by the drug in the throat. Nausea and retching are observed in all animals, while vomiting occurs in dogs and cats. The pulse and respiration are weakened and generally retarded. After lethal doses these symptoms are intensified. We observe violent retching, frequent and difficult attacks of swallowing, ejection of frothy mucus from the mouth, in horses copious sweating; pulse first weak and infrequent, later rapid, running and almost imperceptible; respiration slow, interrupted, and shallow, and reduction of temperature. Death is preceded by muscular twitchings, in the horse, and loss of strength so that the subject falls and is unable to rise; or in the case of cats and rabbits, the animals jump vertically into the air, topple over backwards and go into convulsions, lying helpless on their side. The labial muscles are retracted and the lips drawn back, showing the teeth covered with foam. The face is anxious, the eyeballs are retracted or protruded, and the pupils more commonly dilated. Death takes place usually from asphyxia, occasionally from syncope. The post mortem appearances are simply those resulting from asphyxia.

The western aconite is bitter and retains its bitterness even on drying. It also benumbs, according to Lloyd, just as does the European aconite. The Lloyds quote Prof. Power in asserting that it contains some alkaloids, one probably aconitin, and several other poisonous principles. According to Dr.

Bartholow of Jefferson Medical College, it is a paralyzer of mobility, but does not impair the contractility of muscles or the irritability of the motor nerves. Death is caused by paralysis of respiration, the heart continuing its action some time after respiration has ceased. The *Aconitum Napellus*, affects the heart in opposite ways. Prof. Chesnut says in regard to the poison of this plant:

All of the parts of the west American aconite are poisonous, but the seeds and roots are the most dangerous. The active principle is not well known, but chemical and physiological experiments point to the existence of one or more alkaloids which resemble aconitin. The effect of the poison is characteristic. There is first a tingling sensation on the end of the tongue which gives rise shortly to a burning sensation, and is rapidly followed by a very pronounced sense of constriction in the throat. The choking thus produced is made the more alarming by the retarding effect which the poison has upon the respiration. The tingling and prickling over the entire body is also characteristic. Besides these symptoms, there are generally severe headache, abdominal pains, confused vision, vomiting, and diarrhoea. Delirium is usually absent. Death ensues from a stoppage of the respiration in from one to eight hours.

Mr. R. Schimpfky, in his Important Poisonous Plants of Germany, makes this statement upon the authority of a physician: He tried the nectar of the European aconite by chewing the flower. After chewing a little while, the same was thrown away and an hour later, he felt upon the end of the tongue, a dull pain as though he had burned it. This sensation remained for three days. In Europe it is not unusual to mix the leaves of this with other salad plants. Frequently the plant is cultivated to be used in destroying insects.

Dr. Chesnut says:

No specific antidote is recognized, but physicians have used atropin, or digitalis and nitrite of amyl, with good effects. The ordinary emetics and stimulants must be given. Artificial respiration should be maintained for a couple of hours, if necessary, and a recumbent position must be maintained throughout the treatment.

Aconitum uncinatum L. Wild Monkshood

Plant smooth; stem slender and somewhat reclined; root thickened; leaves 3-5 lobed, petioled, lobes ovate-lanceolate, with coarse teeth; large blue or pale flowers with erect helmet; found in rich shady woods along streams.

Distribution. Eastern North America extending into Iowa.

Poisonous properties. Contains the same active principle as the other species of aconite.

4. Anemone

Erect perennial herbs; root leaves lobed, divided or dissected; stem leaves forming an involucre either remote or near the flower; sepals few or many, 4-20 petal-like; or in one section, petals stamen-like; stamens numerous; pistils numerous; achene pointed or tailed, flattened; single seeded. About 80 species in temperate regions. Several as *Pulsatilla* are medicinal.

Anemone patens L. var. Wolfgangiana (Bess.) Koch

A perennial herb, with radical leaves, appearing after the bluish or purplish flowers have blossomed, villous with long silky hairs; flowers erect, coming from a simple stem, which is naked except for the involucre; petals wanting, or abortive, stamen-like; sepals petal-like, about 1½ inches long; leaves ternately divided with the lateral divisions 2-parted; stamens numerous; pistils numerous in a head with long, hairy styles, in fruit forming feathery tails.

Distribution. Illinois and Nebraska, Wisconsin, Minnesota, Missouri, N. W. Territory, Rocky Mountains to Texas, British Columbia. Known as the pasque or sand flower, but very commonly and incorrectly called the crocus.

Poisonous properties. A very poisonous plant. The allied European Anemone Pulsatilla is also regarded as poisonous. The different parts of the plant are extremely acrid and when applied to the skin cause irritation and vesication. The acridity of the plant is due to the presence of a crystalline substance called anemonin $\rm C_{10}H_8O_4$ which when heated with acids, forms anemonic acid $\rm C_{10}H_{12}O_6$. Lloyd states:

All parts of fresh Anemone patens are acrid and very irritating. Dr. W. H. Miller informs us that his hands have heen very hadly hlistered, in consequence of the juice having spattered over them while pressing the plant. The vapors evolved from the fresh juice are of such an acrid nature as to have inflamed the eyes, and have closed them temporarily. For this reason, persons refuse to work with the fresh herh, and hotanists have heen known to severely irritate their hands simply from contact with the recent plant.

The only demand for this plant is by Homeopathic physicians. All portions of the European Anemone patens are very acrid, but the dried plant is merely an astringent. The plant evidently contains a volatile acrid substance, which is given off when heat is applied. Our sand flower was one of the chief medical plants of the Indians of Minnesota. The plant is still used, when in a fresh condition by the Indian. Dr. Millspaugh gives the following method for preparing it:

The whole, fresh, flowering plant is chopped and pounded to a pulp and weighed. Then two parts by weight of alcohol are taken, the pulp thoroughly mixed with one-sixth

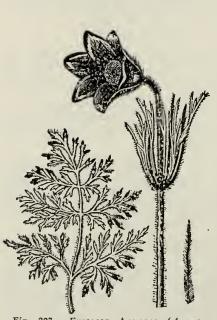


Fig. 227. European Anemone (Anemone Pulsatilla). A poisonous plant with acrid (From Vesque's Traité de Botanique.)



Fig. 228. Pasque Anemone (Anemone patens, var. Wolfgangiana). A plant with well known acrid properties. (Charlotte M. King.)

part of it, and the rest of the alcohol added. After thorough mixture, the whole is allowed to stand eight days in a well-stoppered bottle. The tineture thus prepared, after straining and filtering, should have a light, seal-brown color by transmitted light, an acrid astringent taste, and a decidedly acid reaction.

Dr. White in his Dermatitis Venenata calls attention to the irritating porperties of the common wind-flower A. quinquefolia. The species is widely distributed in woods in Eastern North America. He states that a large wholesale dealer in medical plants regarded it as an externally corrosive poison. It is probable that other species of the genus are more or less acrid. Some of these plants like the white meadow wind-flower A. canadensis are probably looked upon with some suspicion. Dr. Johnson in his Manual of Medical Botany, makes this statement in regard to the common Wind-flower:

Pulsatilla is an aerid irritant which, in large doses, has often produced serious and alarming effects. In safe medicinal doses, however, its effects are by no means so well known. At various times and by numerous authors it has been highly praised as a remedy in diseases of the eye, in rheumatism, amenorrhoea, dismenorrhoea, etc. In this country it has been employed chiefly by homeopathic practitioners, and usually in very minute doses. Many of the results claimed for it under such circumstances are at least doubtful. Certain it is that other practitioners bave not been able to confirm them.

Anemone quinquefolia L. Wind Flower

A low smooth perennial with filiform rootstock, involucre or 3-petioled, trifoliolate, toothed leaves, sepals 4-7, ovate, white, pale blue or purple; carpels 15-20 oblong with a hooked beak.

Distribution. In woods from Nova Scotia to Georgia and the Rocky Mountains, also in Europe.

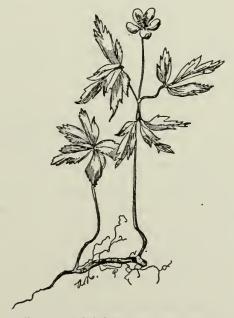


Fig. 229. Wind flower (Anemone quinquefolia). A well known plant with more or less acrid properties. (Ada Hayden.)

Poisonous properties. Leaves, roots, and stems are acrid; same properties as in the Pasque Flower.

Anemone canadensis L. Meadow Anemone

A hairy perennial from 1 ft. to 18 inches high; involcure 3-leaved bearing a long peduncle or a pair with a 2-leaved involucre at the middle; radical leaves long, petioled, 5-7 parted or cleft; sepals white 6-9 lines long; head of fruit globose, achenes flat, tipped with stout style.

Distribution. In low grounds, meadows, especially westward from Labrador to Saskatchewan, Colorado to Maryland and New England.

Poisonous properties. All parts of the plants with acrid properties.

5. Clematis

Climbing vines or perennial herbs, more or less woody; leaves opposite, slender petioled, pinnately compound, lobed, or entire; sepals 4 or rarely more, petal-like; petals none; stamens numerous; ovaries free; the fruit an achene 1-seeded; style long, persistent, plumose, silky, or occasionally naked. About 100 species of wide distribution, abundant in temperate regions. The most common species in Northern States is the Clematis virginiana, in the South and Rocky Mountains, C. ligusticifolia; and in the South, the C. Viorna, and C. Pitcheri. The European C. vitalba contains anemonol.

Clematis virginiana L. Virgin's Bower

Perennial, climbing, with leaflets mostly broadly ovate, acute, cut low; flowers axillary, clusters panicled, polygamo-dioecious, white; the style persistent, plumose. The western *C. ligusticifolia* is nearly like this species.

Distribution. From Canada to Florida, and Kansas, Nebraska, northward. C. ligusticifolia is common throughout the Rocky Mountains from Western Nebraska to the Pacific Coast.

Clematis Pitcheri Torr. & Gray

Perennial herb with pinnately compound leaves, high upper leaves often simple; flowers large, solitary, on long peduncles; usually nodding calyx, bell-shaped; dull purplish sepals, with narrow and slightly margined recurved points; tails of the fruit naked or shortly villous. From Southern Indiana to Central Iowa, and Kansas and Texas.

Poisonous properties. J. U. and C. G. Lloyd, in their Drugs and Medicines of North America, report the medical properties of several species of the genus Clematis as follows:

It imparts a rank taste, which, after prolonged chewing, becomes acrid and irritating, although at first it is only disagreeable.

Dr. Rusby also mentioned the poisonous character of the species of Clematis. In Cuba, one species of the genus is used in case of tooth-ache to blister the face, and this as well as another species is used in the same way for rheumatism. Dr. J. C. White refers to the European *Clematis recta* as producing blisters and often ulcers, and causing the eyes to water and become inflamed.

An infusion of the plant in oil has been used to cure the itch, and violent inflammation of the skin has been produced by friction with it.



Fig. 230. Virgin's Bower (Clematis virginiana). Probably poisonous. (From Johnson's Medical Botany of N. A.)



Fig. 231. Clematis, Virgin's Bower (Clematis Pitcheri). The flowers are dull purplish in color; the plant is more or less acrid. (Ada Hayden.)

6. Ranunulucus (Tourn.) L. Crowfoot Buttercup

More or less acrid, annual or perennial, herbs with alternate, simple, lobed, divided, or dissected leaves; solitary or somewhat corymbed flowers, yellow, white or red; sepals 5; petals 5 with a nectariferous pit or scale at the base; stamens numerous; pistils numerous; achenes numerous in heads, generally flattened or pointed with an erect seed, tipped with the style. About 200 species in temperate and colder regions.

Ranunculus sceleratus L. Ditch Crowfoot

A glabrous annual, 1 foot or more high, stem hollow; root leaves 3-lobed, rounded; stem leaves 3-parted, the lobes cut and toothed; upper leaves nearly entire; flowers pale yellow, petals but slightly longer than the calyx; stamens numerous; pistils numerous, in oblong and cylindrical heads.

Distribution. Wet ditches in the Northern States; also in Europe.

Poisonous nature. The plant is highly acrid, blistering the mouth and skin. In Europe it is used by beggars for making sores. Other species with similar properties are R. acris, R. bulbosus, and R. repens.

Ranunculus septentrionalis, Poir. Creeping Crowfoot

A low, hairy, or nearly smooth, glabrous perennial; ascending, or often producing long runners; leaves with 3-stalked divisions, the terminal one broadly



Fig. 232. Cursed Crowfoot (Ranunculus sceleratus). Common in low grounds. Contains an acrid poison. (Ada Hayden.)

wedge-shaped, 3-cleft or parted; flowers yellow, petals obovate and larger than the sepals; stamens numerous; pistils numerous; style long and attenuate; fruit an achene.

Distribution. Common in moist shady places in the northern states.

Ranunculus acris L. Tall Crowfoot

Hairy, perennial, with fibrous roots, from 2-3 feet high; basal leaves tufted 3-7 divided, divisions cleft in narrow acute lobes; upper leaves short petioled, 3-parted; petals much longer than the spreading calyx; head when in fruit globose.

Distribution. Native to Europe, widely naturalized in the Northern States and Canada.

Poisonous nature. Juice acrid, poison dissipated on drying. Symptoms produced in animals are blistering, slavering, choking, vomiting, in some cases, followed by death resembling that from apoplexy.

Ranunculus abortivus L. Small Flowered Crowfoot

Smooth, branching biennial from 6 inches to 2 feet high; root leaves round, heart-shaped or kidney shaped, crenate, or lobed; stem leaves sessile

or nearly so, divided into oblong linear lobes; flowers small; sepals 5, reflexed; petals 5, yellow; head globose; carpels mucronate with a minute curved beak.

Distribution. In moist woods and meadows; a troublesome weed. Newfoundland to Manitoba, Nebraska, Colorado and Florida.

Poisonous properties. The leaves of the plant have an acrid, peppery taste and cause blistering.



Fig 233. Buttercup (Ranunculus acris). The juice of this plant has acrid properties. (Ada Hayden.)



Fig. 234. Small-flowered (Trowfoot (Ranunculus abortivus). The leaves cause blistering. (Ada Hayden.)

According to Basiner, the oil of Ranunculus acts, in warm-blooded animals, as an acrid narcotic, producing, in small doses, stupor and slow respiration; in larger doses, also, paralysis of the posterior and anterior extremities, and, before death, convulsions of the whole body. The acrid action is shown by a corrosive gastritis and by hyperaemia of the kidneys, more particularly of their cortical substance. Anemonin causes similar symptoms, but is followed by no convulsions, nor does it irritate sufficiently to corrode the organs, as the oil does.

Dr. Millspaugh mentions especially the R. bulbosus as having a peculiarly powerful irritant action upon the skin, whether applied locally or internally.

Murray states that a slice of the fresh root (bulb?) placed in contact with the palmar surface of a finger brought on pain in two minutes; when taken off, the skin was found without signs of extra circulation or irritation, and the itching and heat passed away; in two hours it nevertheless returned again, and in ten hours a serious blister had formed, followed by a bad ulcer, which proved very difficult to heal.

Stebler and Schröter state that *Ranunculus acris* produces diarrhoea, abortion, and loss of flesh, and when eaten in large quantities, death ensues in a few hours. Dr. Johnson in his Mannual of Medical Botany of North America, says:

The ranunculi are too acrid to render their internal use either desirable or safe. Most of them are avoided by domestic animals; one may often see R. acris, for example, growing luxuriantly in pastures where almost every blade of grass is cropped close. Their acrid properties have, however, led to their employment externally as rubefacients or vesicants in cases where other and perhaps better agents were not at hand, or were for any reason contra-indicated. As is well known, cases of idiosyncrasy occur in which cantharides are inadmissible on account of their effect upon the urinary organs. In some such cases ranurculus has been used with good effect. One of the faults of this agent is its extreme violence. The fresh plant, bruised and applied to the skin, may vesicate in an hour or hour and a half, and may possibly produce an ulcer not easy to heal. It is, therefore, far less safe as a rubefacient than mustard, and, as a rule, much less desirable as a vesicant than cantharides. It has been employed to some extent in European countries as an external application in chronic rheumatism, neuralgia, etc., but never sufficiently to have obtained a place in the pharmacopoeias. In this country it is used still less, and is little more than mentioned in works on materia medica. An interesting observation regarding the possible effect of R. acris on pregnant cows was reported to the author by bis brother, Mr. F. M. Johnson. In a herd of cows pastured for years in succession in an old field thickly beset with this weed, abortion was frequent and troublesome. As soon, however, as this pasture was broken up and the herd moved to another part of the farm in which the plant did not grow, abortion disappeared. Now although, as stated above, . domestic animals avoid this plant, yet when feeding where it is very abundant, they must occasionally swallow it accidentally; and though there is no positive proof that the abortions were due to the plant in question, the facts as stated are interesting and significant. It is at least possible that ranunculus exerts an influence upon the reproductive organs like that which is claimed by some for pulsatilla.

Leaves, flowers, and stems of the Ranunculi have a peppery and pungent taste, when eaten, reminding one of mustard. According to Lloyd, boiling water dissipates the acrid principle.

Many other species of the genus Ranunculus such as R. Flammula, and R. arvensis are acrid and poisonous, causing the formation of blisters.

Delphinium Tourn

Perennial or annual herbs of erect, branching habit, with racemose or paniculate showy flowers; leaves palmately lobed or divided; sepals 5, irregular, petal-like, the upper one prolonged into a spur; petals 2-4, irregular, the upper one spurred, and enclosed in the spur of the calyx; stamens numerous; pistils 1-5, forming follicles in fruit, many seeded. Species about 60 in the North temperate regions.

Several species like Stavesacre (D. Staphisagria) are used in medicine, and many are poisonous to live stock. Several species like the field larkspur (D. Consolida), the rocket larkspur (D. Ajacis), the great flowered larkspur (D. grandiflorum) and blue larkspur (D. elatum), are ornamental.

Delphinium Penardi Huth. Prairie Larkspur

A perennial pubescent or hairy herb, more or less glandular above, from 3-5 feet high; leaves 3-5-parted, the divisions 2 or 3 times cleft, the lobes linear; flowers in racenes; white, slightly tinged with blue; spur horizontal, straight or slightly curved upward; follicle pubescent, many seeded. This is closely related to *D. azureum* of more southern distribution, with light blue flowers and downy follicles.

Distribution. The Prairie Larkspur is common on sandy soil, gravelly knolls and prairies of Illinois and Wisconsin to Manitoba, Kansas and Arkansas.

7. Delphinium Geyeri Greene

A hairy perennial 1-2 feet high; leaves dull green, somewhat branched; flowers in dense racemes, azure blue.

Distribution. Common on the high plains of Colorado to Montana.



Fig. 235. Carolina Larkspur (Delphinium Penardi). Like other species of this genus it is poisonous. Common on prairies and gravelly soil. (Ada Hayden.)

Fig. 236. Purple Larkspur (Delphinium bicolor) is found in Montana and Westward. Poisonous. (Chesnut, U. S. Dept. Agr.)

Delphinium Menziesii DC

Glabrous below, at least at the very base, pubescent above with spreading hairs, especially the inflorescence; leaves 5-parted, divisions 2 to 3-cleft; flowers large, deep-blue, in a loosely few to many-flowered simple raceme; upper petals veined with purple; spur long and slender; ovaries somewhat tomentose.

Distribution. Common from San Francisco, California, north to British Columbia, Idaho, Montana and Wyoming.

Poisonous properties. Chesnut and Wilcox say:-

Experiments have been made which show that at one stage of growth the leaves of the species (D. Menziesii) may be safely eaten, to a certain extent, by sheep. Dr. S. B. Nelson, professor of veterinary science in the Washington Agricultural College, in an article entitled Feeding Wild Plants to Sheep, published by the Bureau of Animal Industry of this Department, showed that it is possible to feed as much as 24¾ pounds of the fresh leaves of D. Menziesii to a sheep within a period of five days without causing any apparent ill effects. The stage of growth of the larkspur was not stated, but, judging from the other experiments described in the same report, it was probably in a well-advanced flowering stage.

In regard to the poisoning from this species, opinion seems to differ. Dr. Nelson states that Dr. Wilcox was in error in regard to the plant that he worked with at first. More than likely it was a D. bicolor, which is corroborated by the report. Under D. bicolor, mention is made of D. Menziesii. However, it is more than likely there are certain stages in the development which are more poisonous than others. Dr. S. B. Nelson from his experiments concludes as follows:

The results obtained in these eight trials with Delphinium Menziesii, the feeding of nine and one-half pounds of the bulbs, stems and leaves of the immature plants to sheep No. 6, the feeding of nearly twenty-five and six pounds of the plants gathered while in full bloom, to Nos. 2 and 3 respectively, and the hypodermic injections of the two extracts into Nos. 4 and 5, certainly constitute evidence sufficiently convincing to justify the conclusion that Delphinium Menziesii is not poisonous to sheep and they may be allowed to graze where it grows even in abundance without fear of any loss from it.

Dr. Nelson likewise carried on an experiment with D. simplex with similar results.

Delphinium bicolor Nutt. Purple Larkspur.

A smooth or somewhat pubescent, tuberous rooted, perennial; 1-2 feet high with a cluster of finely divided leaves; the lower orbicular in outline, all deeply cleft or parted; racemes few or several-flowered; flowers dark purple; sepals and spur ½ to 34 inch long; upper petals pale yellow, and white with blue veins, follicles smooth or minutely pubescent when young.

Distribution. Common in dry ground, Eastern Oregon, and Washington, to Utah, British Columbia, Montana, and Colorado. Grows in elevations from 4000 or 5000 feet to 10,000 feet.

Poisonous character. It is regarded as poisonous by stockmen. Experiments reported by Dr. Wilcox in the Montana Agricultural Experiment Station Bulletin show that an extract from less than one ounce of the dried leaves of this species was fatal to a yearling lamb. Chesnut and Wilcox report further experiments as follows:

The following experiments were made with a view of ascertaining whether the permanganate of potash, which it was proposed to use in many cases of plant poisoning, could be used with satisfactory results as a chemical antidote in case of this plant, and also of determining which parts of the plant were most toxic. During some seasons the purple larkspur causes extensive poisoning of sheep and calves. Cattle and horses, on the other hand, seem to cat it less frequently than the tall larkspur. In the season of 1900 conclusive evidence against this plant was obtained in only one locality. This was derived from two cases among calves in the Flathead Valley. The calves were about 5 weeks old, and at the time when the poisoning occurred, were running in a native pasture where the purple larkspur grew sparingly. The symptoms of poisoning in these two cases were similar to those already outlined from poisoning in the tall larkspur in cattle, with the exception that a slight bloating was to be observed in the case of the calves. The respiration and heart beat became exceedingly rapid as the symptoms of poisoning increased in severity. The body temperature was slightly lowered, and this was accompanied by pro-

fuse sweating. The increased perspiration may have been due in part to the violent spasms in which the animals finally died. Death occurred about four hours after the appearance of the first symptoms. No remedy was applied in these cases.

Delphinium scopulorum Gray

A glabrous or finely pubescent perennial with leafy stem 1-6 feet high from fascicled thick roots; leaves numerous orbicular, 5-7, parted, lower cuneate, and the upper consisting of narrow, cleft, and laciniate divisions; racemes many-flowered, sparingly pilose, flowers blue varying to white or pink on short erect pedicels; spur longer than sepals, lower petals deeply notched, and upper whitish, and a little shorter than the oblong sepals; follicles about half an inch long, erect, seeds small, with a loose coat.

Delphinium occidentale Watson. Tall Larkspur

A glandular pubescent perennial from 4-6 feet high; leaves deeply 3-5 cleft, divisions broadly cuneate, somewhat 3-lobed; flowers numerous in a many-flowered sparingly-branched panicle; sepals spatulate, acuminate; dull or dark blue, very variable in size; seeds light colored, and somewhat spongy.

Distribution. At higher altitudes from Colorado to Eastern Oregon, and Nevada.

Delphinium trolliifolium Gray. California Cow Poison, or Poisonous Larkspur

A tall smooth perennial, 2-5 feet high, sparingly villous, hairy; leaves large, long, petioled, 5-7 lobed, lobes lacinately cleft and toothed with acuminate segments; flowers large in loose racemes; color bright blue, 1½ inches broad, spur as long as the sepals; sepals oblong-lanceolate, acuminate, sparingly villous, follicles smooth, 6-8 lines long, seeds turbinate.

Distribution. Common along the Pacific Coast from British Columbia to California.

Poisonous properties. Prof. Chesnut says: "In Humboldt County, Cal., it is known as Cow Poison on account of its fatal effect on cattle. Its toxic character has been questioned. Perhaps it is not equally poisonous throughout all stages of its growth."

Delphinium tricorne Michx

A simple stout perennial 6 inches-2 feet high, with a cluster of roots; leaves slender, petioled, deeply 5-7-cleft, or divided; the divisions linear or ovate; flowers in loose racemes, blue, occasionally white or purple; spur slightly bent. Follicles tipped with a short beak.

Distribution. In open rich woods or clay soil, Western Pennsylvania to Southeastern Iowa to Arkansas and Georgia.

Poisonous properties. The Stavesacre (Delphinium Staphisagria) of Italy and Greece to Asia Minor has long been used in medicine, having been known to the ancients. Pliny mentions the use of the powdered seeds for destroying vermin on the head. It is still largely used for destroying pediculi. The eclectic physicians use it for its specific action on the reproductive organs. The disease produced by Delphinium may be called delphinosis.

According to Prof. Hills, Stavesacre and D. Consolida are used in the treatment of dropsy and spasmodic asthma. The effects produced are due to



coast, and said (Ada Hayden.)

Fig. 238. Tall Larkspur (Del-phinium glaucum) of the Pacific Coast region. It is poisonous. (Chesnut, U. S. Dept. Agrl.).

the alkaloid delphinin C22H36NO6, very poisonous with a bitter sharp taste. Three other alkaloids have been isolated, staphisagrin C22H23NO5, bitter, the poisonous delphinoidin C42H58N2O7, and delphisin C27H43N2O4, an extremely poisonous alkaloid, to which may be added the substance calcitripin. J. C. White in his Dermatitis Venenata states that acute dermatitis resembling eczema may appear from the use of stavesacre seed. Delphinocurarin C₂₃H₃₃NO₇, has been obtained from the root stock of several species of Delphinium, D. bicolor contains 0.27 and D. scopulorum, 1/3 per cent.

In this country, it appears, from Mr. Cheney's observation, that D. consolida is largely sold for the European plant. A tincture of the seed is often mixed with Lobelia inflata and sold as a parasiticide. The Stavesacre seeds are still employed as in old times for the destruction of pediculi in human beings. For this purpose, they are converted into powder and dusted among the hair.

Results of the experiments made by Dr. Crawford of the U. S. Dept. of Agr. show in 1905 in regard to larkspur poisoning. (The first batch of plants was collected April 26th, 1905).

1 c. c. injected into a guinea pig (subcutaneously), weight 730 grams. Caused no disturbance.

3 c. c. in guinea pig, no symptoms.

6. c. c. in guinea pig, killed.

6 c. c. injected into guinea pig, 285 grams, killed in 33 minutes.

4 c. c. injected into guinea pig, 352 grams, no symptoms.

Repeated:

5 c. c. killed guinea pig weighing 196 grams. Died in 55 minutes.

4 c. c. injected into guinea pig, 299 grams. No symptoms.

Evidently lethal dose for this solution lay between 4 to 5 c. c.

Second Stage, Gathered May 16th, 1905

Solution corresponding to 4 c. c. of No. 1 caused no symptoms in guinea pig weighing 445 grams, while 5.3 c. c. killed one of 350 grams, but death was delayed longer than with extract of first stage.

Third Stage, Gathered in June, 1905

Solution corresponding to 4 c. c. caused no symptoms in guinea pig weighing 376 grams.

5.3 c. c. caused no symptoms in guinea pig weighing 500 grams.

6.6 c. c. caused no symptoms in guinea pig weighing 480 grams.

Evidently a lethal dose is much higher and the plant loses much of its activity in development.

This report is very conclusive in proving that the plant contains an active poison, and further in substantiating the claims of experienced observers that the plant loses much of its toxic properties as it approaches the flowering period.

Seven and one-half grams of dried purple larkspur fed to each of three rabbits on April 20th. No results.

Seven and one-half grams of fresh purple larkspur from same patch fed April 25th to each of three rabbits. Two showed slight uneasiness, and one was bloated a little. One, showing less effect than the others, had eaten but three and one-half grams.

On May 1st a like quantity from the same patch was given to the same rabbits under similar conditions. Results, two died, and the other distressed.

On June 15th, plants from the same source, being in full bloom, but the leaves and stems dry, were fed to rabbits. Although very hungry, they at first refused to eat, but later ate large quantities of it without any ill effects. The experiments with tall larkspur were equally as confusing. The fact that the plants at one period of growth gave negative results was no guaranty that they would not be dangerous at another. The tall larkspur growing luxuriantly on the college campus proved to be very active, physiologically, and furnished the best specimens for producing the physiological effects upon animals. In the experiments with antidotes this domesticated species was found to be very poisonous while in bloom in the middle of August.

Lloyd in Drugs and Medicines of North America, quotes from a letter from Wm. C. Cusick, a botanist of the West, who states that in some places the D. decorum F. & M. var. nevadense poisons cattle, but only in the early spring when they are first turned on the crop. It is thought by cattle men that the cattle pull the plants up by the roots and eat them, which really causes, the

poisoning. Many die, but most recover. Prof. Chesnut, in his Principal Poisonous Plants of the United States, says:

The percentage of fatal cases in cattle which have eaten this and other larkspurs is said to be small. A rough estimate by a cattleman places it at about 20 per cent for one species of the group, when the animals are not properly treated, and 5 per cent otherwise. This is probably a low estimate, bowever, for in a case of poisoning from D. Menziesii that occurred in Montana in May, 1897, and was reported by Dr. E. V. Wilcox, nearly 600 sheep were affected, 250 of which died.

Dr. Wilcox says in regard to D. glaucum:

The tall larkspur appears not to be eaten by sheep. All cases of poisoning from this plant observed in Montana during this year and previous years have been among cattle. As already indicated, sheep are not driven to the mountain ranges until about the middle of July, and at this season the tall larkspur is altogether too large and coarse for their consumption. It is well known, on the other hand, that cattle will feed on much coarser forage than slicep, and at the same time they are allowed to run on the high ranges in the early spring. In Montana the light green tufts of leaves of the tall larkspur first become conspicuous about the last of April or the first of May, and the flowers begin to open about the middle of June. This fact is significant for the reason that light falls of snow often occur in the larkspur belt as late as the first week in June, and, since there is then no other verdure in sight, the uncovered portion of the larkspur is in a high degree tempting to stock, all the more because it is succulent. The danger is increased by the fact that at the time of snow falls cattle seek the shelter of creeks where the larkspur is more abundant and most advanced in growth. At this period, moreover, the leaves are, as we can testify from personal observation, very bitter, and they are probably then more poisonous than at any other stage of growth. The older ones, as is also the case with the blue larkspur, are not so bitter. The plant is recognized by cattlemen as dangerous to cattle from May until about the middle of June. Mr. Vard Cockrell informs us that on his range in the lower basin of the Gallatin it is the sole duty of one man during this period to keep his cattle away from the broken mountainous regions where this larkspur abounds.

The symptoms are described as follows:

In general, the animals affected manifest symptoms similar to those produced by overdoses of aconite. The first signs of poisoning are usually a general stiffness and irregularity of gait. There is often a pronounced straddling of the hind legs in walking. These symptoms increase in severity until locomotion becomes difficult or impossible, and the animal finally falls to the ground. It usually falls and gets on its feet again a number of times, the muscular movements becoming more and more irregular and incoördinated. At the same time the skin is very sensitive to touch, and the muscles of the sides and legs soon begin to quiver spasmodically. This a very characteristic symptom, being usually exhibited for several hours. The function of the special senses is seldom impaired, the animal being apparently able to hear and see as well and as correctly as under conditions of health. Although a slight increase in the quantity of saliva is to be noticed in some cases, this symptom is never so pronounced as in cases of poisoning by death camas. During the later stages of poisoning the animal is usually attacked with violent convulsions, in one of which it finally dies. In this respect, also, the symptoms differ distinctly from those of death camas poisoning, which is usually quite without spasms. The digestive functions seem not to be affected by larkspur poisoning The temperature is lowered slightly during the first stages, in one instance having been as low as 97°. During the later stages the pulse becomes very frequent and the breathing rapid and shallow. The cerebral symptoms are simply those of excitement, and the appetite seems not to be lost until shortly before death.

In regard to D. bicolor, the symptoms observed from experiments are described by the same author as follows:

Summing up the results of these experiments, we find that the most prominent, easily observable symptoms were a stimulation of the respiration and a brain symptom manifested by dizziness or a rhythmical movement of the head. As it was not the object of the experiment to determine the symptoms of poisoning, these having been already secured by Dr. Wilcox, no special effort was made to determine the pulse rate, but it was to be noted that in experiment 2, which was as nearly fatal as any, the heart action was extremely rapid and weak. None of the experiments proved fatal and no results were obtained on which

a satisfactory trial of the permanganate of potassium as an antidote could be based; hence no experiments were made in that line.

Dr. B. Kennedy in the Nevada Agricultural Experiment Station, reports in regard to the symptoms of poisoning from D. glaucum, as follows:

This dangerous, poisonous plant was seen quite frequently from Lincoln Valley to Summit Soda Springs. It occurs usually in large patches by the side of creeks in the valleys. Although sheep do not care for it, yet it is sometimes eaten by them. Sheepmen do not consider it dangerous. Cattlemen, however, are afraid of it, and consider it very poisonous. Mr. Ridinger of the Tehauntepec dairy, about eight miles from Webber Lake, told us that cattle die after eating it, with the following symptoms: Trembling and shivering, succeeded by extreme weakness, which makes affected animals stagger and fall or lie down.

That Larkspur is poisonous, at least some species, appears from the experiments carried on by Dr. Geo. H. Glover and C. Dwight Marsh:

There can be no question but that the several species of larkspur growing native in the mountainous districts of Colorado arc a greater source of loss to the stockmen than all other weeds combined. While the larkspur is confined to the mountainous regions, it nevertheless holds true that in the aggregate mortality throughout the state from poisonous plants larkspur takes second place only to loco. We have no statistics at hand whereby we can estimate, with any degree of accuracy, the total loss, but judging from the reports of other western states and from information received from most every section of the state, it would seem that \$40,000 annually is a conservative estimate. There are four species of larkspur found growing abundantly in the middle and western portion of this state, and one found growing sparingly in the eastern plains section. Other species have been found in isolated places, but have not been especially accused of doing any harm, and their toxicity has not been proved. The four species found in the greatest abundance and named in the order of their importance, are purple larkspur, Delphinium Nelsonii. Greene; tall larkspur, Delphinium elongatum, Rydb.; D. Geyeri, Greene, and D. Barbeyi, Huth. These all have the same characteristic flowers, and are found growing in the mountains at altitudes from 5,000 to 11,000 feet. The D. Penardi, Huth, has a white flower and may be seen growing adjacent to streams and in the arroyas on the plains as far east as the state line.

The *Delphinium elongatum* and *D. Nelsonii* are the only ones which contain a sufficient quantity of deleterious substance to produce poisoning. From all accounts it seems probable that the plants are most poisonous in the spring when they are fresh. When the poison has been absorbed into the system atropin is an antidote.

8. Actaea L. Baneberry

Perennial herbs with 2 or 3 ternately-compound leaves; sepals 4-5, petal-like, soon falling; petals 4-10, small, flat, spatulate, on slender claws; stamens numerous, free, with slender white filament; ovary solitary; stigma sessile; fruit a berry; seeds compressed, smooth, horizontal.

Actaea rubra (Ait.) Willd. Red Baneberry

A smooth perennial 1-3 feet high with biternately divided leaves, on long, smooth petioles, leaflets ovate, sharply cut, and toothed. Calyx with 4 greenish sepals; corolla 8-10 petals, white, shorter than the stamens; stamens numerous; berries cherry red.

Distribution. Eastern North America to Hudson Bay, and the Rocky Mountains.

Poisonous properties. Prof. Chesnut states that sheep are occasionally poisoned by eating the leaves of the closely related European species. These plants are seldom, however, eaten. In its medical action Actaea is similar to Cimicifuga. It contains a resinous body which is neither acrid nor bitter, but accord-

ing to Fred Stearns, the rhizome has purgative properties. This, according to Mr. Lloyd, may be somewhat overdrawn. Prof. Sayre states that it is a violent purgative, irritant and emetic. The berries are known to be somewhat poisonous.

9. Hydrastis Ellis. Yellow Puccoon

A low perennial herb with knotted yellow rootstock, and a single root; stem bearing 2 leaves near the summit; flowers large, greenish-white; 3 sepals, petal-like, soon falling; petals none; pistils 12 or more; 2-ovuled; fruit 1 to 2-seeded berry, crimson in color. A genus of 2 species

Hydrastis canadensis L. Golden Seal. Yellow Puccoon

Rootstock 1 and 2 inches long, knotted; berries numerous, small; fibrous root, roots and rootstocks yellow in color; leaves pubescent, palmately 3-5-lobed; calyx petal-like.

Distribution. From New York to Southern Michigan, Southern Wisconsin and Eastern Iowa to Arkansas to Northern Georgia.

Poisonous properties, also medicinal properties. The plant contains the alkaloid berberiu $\rm C_{20}H_{17}NO_4$, and hydrastiu $\rm C_{21}H_{21}NO_6$, a so-called alkaloid, but which, according to Mr. Lloyd, cannot be considered in the pure condition; also canadin $\rm C_{20}H_{21}NO_4$ and xauthopuccin; it also contains a fixed oil of a disagreeable odor and taste, and a black resinous substance. It produces ulceration and catarrhal inflammation of the mucous surfaces. The plant acts very similarly to Cimicifuga. The Lloyds have given an extended account of the anatomy, structure and therapeutical properties of this plant. The alkaloid berberiu $\rm C_{20}H_{17}NO_4$ is identical with the substance found in the barberry, according to Prof. Power.



Fig. 239. Golden Seal (Hydrastis canadensis). The rootstocks of this plant as well as the stem and leaves contain more or less acrid substances. (Charlotte M. King).

BERBERIDACEAE. Barberry Family

Shrubs or herbs, with alternate leaves; stipulate or exstipulate flowers either solitary or in racemes, perfect; stamens as many as petals and opposite them, hypogynous; fruit a berry or capsule. A small family of about 20 genera and 105 species, widely distributed in the North Temperate region, also in Temperate South America and Asia. Some of our North American species are weedy, others woody and several are more or less poisonous. A few like the common



Fig. 240. Common Barberry (Berberis vulgaris). The spines often inflict mechanical injuries while the leaves and wood are more or less poisonous. (Ada Hayden).

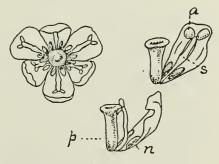


Fig. 240a. Single flower. s, stamen; a, anther; p, pistil; n, nectar gland.

barberry (Berberis vulgaris), blue cohosh or pappoose root (Caulophyllum thalictroides), the twinleaf (Jeffersonia diphylla), and mandrake (Podophyllum peltatum) are used in medicine. The root of Berberis aristata and B. vulgaris sometimes used as a fish poison. Probably some other members of this order are poisonous. The blue cohosh (Caulophyllum) contains saponin. This plant is said to be extremely bitter to the taste, but is not, however, common.

Key for genera

1. Podophyllum L. Mandrake; May-apple

Perennial herbs with simple, smooth, erect, stem; creeping rootstocks and thick fibrous roots; stems bearing 2 leaves with large flowers; flower buds with 3 green bractlets, 6 fugacious sepals; petals 6-9; stamens twice as many

as the petals; pistil 1; stigma large, flat, sessile; fruit a large fleshy berry, 1-celled and many-seeded, each seed enclosed in a pulpy aril.

4 species, one in eastern North America and the others in India and Eastern Asia. The *P. emodi* of Asia contains the same principles as the American species and is poisonous.

Podophyllum peltatum L. Mandrake

Perennial herb, with creeping rootstocks and thick fibrous roots; flowering stems with 2-leaves, 1-flowered, the flower bud with 3 small green bractlets, which fall away early; calyx of 6 unequal sepals, corolla white, of 6-9 petals, about twice the length of the sepals; stamens 12-18, twice the number of petals, inserted below the pistils, with short stamens; anther cells opening longitudinally; creeping rootstock, from 1-5 feet long, fibrous rooted.

Poisonous properties. Its medicinal virtues were well known to the Indians of North America, and an early writer, Catesby, remarked that the root was an excellent emetic. It has a bitter, acrid taste, similar to that of roots of other plants of the family. Its active properties seem to reside in the resinous substance. Prof. Power failed to find an alkaloid. The name podophyllin has been given to the product found in the resinous substance contained in other members of the family. This in turn contains podophyllotoxin $C_{11}H_{14}O_2+2H_2O$, and picropodophyllin $C_{11}H_8O_2+H_2O$, producing a very bitter taste; and intensifying the action of podophyllin. Berberin $C_{20}H_{17}NO_4$, which is feebly toxic to man, and saponin have also been found. Cases of poisoning have occasionally



Fig. 241. Mandrake (Podophyllum peltatum). powerfully purgative. (Ada Hayden).

been reported. It produces fatal prostration. It seems to exert a special influence upon the liver. Dr. Rusby says:

Its taste, especially when fresh, is very repugnant, and yet if eaten in quantities it would unquestionably prove fatal, as shown by the effects of over-dosage in medicine. In the Philadelphia Medical and Surgical Reporter, XIX, 308, a fatal case is recorded in which the evidence is perfectly clear that poisoning resulted from continued large doses administered by an ignorant and careless physician. The poisonous symptoms were all referable to the howels, those of enteritis. It is also very interesting to note the peculiar effects of poisoning of the external skin by the powder and by the resin of this drug. It produces an ulcer of a very peculiar character, closely resembling one of venereal origin. Serious errors of diagnosis, leading to the gravest injustice to the reputation of the patient, have heen known to occur in reference to these cases. A very serious ulcer upon the eye-hall is among these recorded cases. The very greatly elongated rhizome of Podo-phyllum, with its very long smooth internodes, broadened nodes with their very large, low, cup-shaped ears, and sparse roots underneath, is doubtless well known to all pharmacists. The plant is not only very common, but extremely ahundant east of the Mississippi, and is liable to be encountered almost anywhere."

In regard to its action on man, Dr. Millspaugh says:

Here the same action takes place, hut extends to the rectum with sufficient intensity to cause prolapsus and hemorrhoids. The first effect of the drug is an excitation of salivary and hiliary secretions, followed by torpor and icterus. The symptoms of disturbance caused by the drug in doses varying from ¼ to ½ grains of "podophyllin," and in persons working in the dust of the dried root, are substantially as follows: Inflammation of the eyes, soreness and pustulation of the nose; salivation and white-coated tongue; extreme nausea, followed by vomiting severe pains in the transverse colon and abdomen, followed by an urgent call to stool; thin, offensive, copious stools; weak pulse, prostration, drowsiness, and cold extremities.

Among other experiments with this drug upon animals, those of Dr. Anstie seem to be the most characteristic. He found, resulting from his many applications of an alcoholic solution to the peritoneal cavity direct, that no local inflammation arose, although an intense hyperaemia occurred in the duodenum especially, and the whole of the small intestine, even going so far as to cause a hreaking down of the tissues and resulting ulceration, causing discharges of glairy mucus streaked with hlood; this hyperaemia ceased usually at the ileocaecal valve. Post mortem: The mucous-membranes were found inflamed and covered with hloody mucus. Other observers noted that retching, salivation, and emesis, followed hy purging, colic, and intense tenesmus, with low pulse, and rapid exhaustion followed the administration of the drug.

Dr. Schaffner says:

Roots, stems, and leaves, drastic and poisonous, but the ripe fruit less so. Leaves, when eaten hy cows, produce injurious milk. The ripe fruit may he eaten in small quantities.

The root of Mandrake affects the skin. Mr. Lloyd, in White's work, writes:

Our employes experience great trouble in working this, owing to the irritating action
of the skin. We have in numerous instances had our men cease work for several days owing
to its action, which causes very painful inflammation of the skin, especially of the eyes.

Dr. Winslow says:

The action is exerted mainly on the duodenum, which is intensely inflamed and even ulcerated in poisoning. Podophyllin directly increases the secretion of hile in small doses, while purgative quantities hasten its excretion by stimulation of the muscular coat of the gall hladder (except in the horse) and small intestines. It is probable that the intestinal secretions are somewhat augmented. The faecal movements, after medicinal doses of podophyllin, are liquid, often stained with hile, and may he accompanied by some nausea and griping.

Berberis L. Barberry.

Shrubs with yellow wood, simple or compound leaves, often spiny; flowers, yellow in racemes, or rarely axillary; sepals 6-9, like petals; petals 6, imbricated in 2 series; stamens 6; fruit a berry with 1-3 seeds. About 75 species of wide distribution.

Berberis repens Lindl. Trailing Mahonia.

A smooth, trailing shrub, 1-4 ft. high, leaves petioled, pinnate; leaflets 3-7, ovate, acute; flowers several in a raceme, yellow; persistent bracts; fruit globose, bluish purple.

Distribution. From Western Nebraska to Arizona, and British Columbia, Northwest Pacific Coast from Washington to California.

Berberis Aquifolium Pursh. Oregon Grape

A low shrub 2-10 feet high, leaflets 5-9 oblong ovate, spinulose dentate above; flowers yellow in racemes and terminal clusters; fruit globose, dark in color.

Distribution. Idaho to the Rocky Mountains.

Poisonous properties. Both Berberis repens and B. Aquifolium contain the alkaloids berberin, oxyacanthin $C_{19}H_{21}NO_3$, and berbamin $C_{18}H_{19}NO+2H_2O$.

Prof. Schaffner reports that the berries of the trailing Mahonia are injurious to birds. When eaten fresh they are emetic and cathartic.



Fig. 242. Oregon Grape (Berberis Aquifolium). Berries are said to be poisonous to birds. (Ada Hayden).

MENISPERMACEAE. Moonseed Family

Woody plants with alternate lobed or entire leaves, climbing without stipules; flowers small, dioecious, in panicled racemes or cymose clusters; sepals 4-12; petals 6; fewer, or more; stamens of the same number or fewer; fruit a 1-sceded drupc; embryo long, curved endosperm scanty. About 300 species mainly in the tropics.

Moonsced (Menispermum canadense) is a beautiful native climber of the North with black drupes and contains menispin. The Carolina moonseed (Cocculus carolinus) is common in the South. Fish poison made from Anamirta

paniculata is used for destroying vermin and to poison fish. The wood of this species is very bitter to the taste and contains an alkaloid menispermin $C_{38}H_{24}NO_4$ and a toxic substance picrotoxin which is said to be a mixture of two bodies picrotoxin $C_{15}H_{18}O_6$ and picrotin $C_{15}H_{18}O_7$. The Columba Root (Jateorrhiza Columba) is used as a medicine and contains several alkaloids, among them, berberin and columbamin. The false Columba Root (Cascinum fenestratum) produces a yellow dye. The alkaloid pelosin is obtained from Chondrodendrum tomentosum.

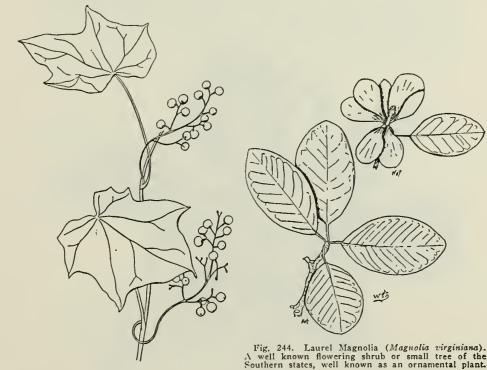


Fig. 243. Moon-seed (Menispermum canadense). The fruit is blue, much the color of Concord grapes. Plants like the fish poisons belonging to the same order are known to be poisonous. This plant has been suspected of being poisonous. (Charlotte M. King).

MAGNOLIACEAE. Magnolia Family

Trees or shrubs, leaves alternate; leaf buds covered by membranous stipules; flowers large solitary; sepals and petals hypogynous, colored alike; stamens numerous, adnate anthers, carpels numerous, separate, or coherent, packed together, ripening into an aggregate fruit; seeds 1-2 in each carpel, achenes or follicles, endosperm fleshy; embryo minute. About 70 species of North America and Asia. The bark of several species used in medicine.

The family Magnoliaceae contains the well known magnolias planted for ornamental purposes. Among these are the great flowered magnolia (Magnolia

grandiflora) which has evergreen leaves and fragrant flowers and is extensively planted in the South; the sweet bay (M. virginiana) a shrub or small tree with fragrant flowers; the cucumber tree (M. acuminata); the umbrella tree (M. tripetala); the Yulan tree (M. Yulan) of China and Japan; the purple magnolia (M. obovata); the tulip tree (Liriodendron Tulipifera) frequently planted as an ornamental tree and producing the most valuable tumber of the family; the star anise (Illicium floridanum) of the south has aromatic bark and pods. The fruit of I. anisatum of Japan, found growing around Buddist temples, furnishes the poisonous sikimin; from the same species in China is made the liquor anisette; Anise is furnished by I. anisatum.



Fig. 245. Great-flowered Magnolia (Magnolia grandiflora). An ornamental tree of the southern United States. (After Faguet). The odor of the flowers said to be injurious to some people.

Poisonous properties. The crystalline substance magnolin, a glucoside, and a volatile oil occur in the large leaved Magnolia (M. macrophylla) of the south. The tulip tree contains the bitter principle liriodendrin, also an alkaloid, and a glucoside. The Talauma macrocarpa of Mexico contains a haemolytic substance capable of dissolving the red corpuscles of the blood. The flowers of Michelia nilagirica are used in perfume. The winter's bark (Drimys Winteri) is used in medicine.

Illicium anisatum of Japan is said to contain a poison belonging to the picrotoxin class, says Blyth. In 1880 five children in Japan were poisoned by eating the seeds of this plant; three died. After considerable experimentation

Dr. Languard concluded that all parts of the plant were poisonous. The poison causes excitation of the central apparatus of the medulla oblongata and elonic convulsions analogous to those produced by picrotoxin, toxiresin and cicutoxin. Small doses kill by paralyzing the respiratory center. Large doses cause heart paralysis. When animals are poisoned by small doses chloral hydrate is an efficient remedy but has no effect when large doses have been taken.

CALYCANTHACEAE

Shrubs with entire short petioled opposite leaves, without stipules; flowers fragrant, large, solitary, on leafy branches, sepals and petals numerous; stamens numerous, the inner short, sterile; pistils numerous; fruit of an ovoid pyriform receptacle, enclosing few to many smooth solitary achenes; seed erect. About 5 species of North America and Asia. Several species are cultivated for ornamental purposes.

Calycanthus L. Carolina Allspice.

Flowers purple or red; sepals and petals numerous, stamens numerous, pistils numerous. A small genus of 4 species, 3 in Eastern North America, and 1 on the Pacific Coast.

Calycanthus floridus L. Strawberry Bush

A branching shrub 3-9 feet high; branches pubescent; leaves oval, soft, downy underneath, roughish above; flowers dark purple, with the odor of strawberries; sepals and petals linear-oblong.



Fig. 246. Strawberry Bush (Calycanthus floridus). Commonly used as an ornamental shrub and is known to be somewhat poisonous. (Charlotte M. King).

Distribution. In rich soil, North Carolina to Georgia, Alabama and Mississippi, cultivated in Missouri, north to Central Iowa.

Poisonous properties. This plant contains several alkaloids among them calycanthin. Chesnut says:

The large oily seeds of the calycanthus, or sweet-scented shrub are strongly reputed to be poisonous to cattle in Tennessee.

Anonaceae. Papaw or Custard Apple Family

Trees or shrubs, generally aromatic; leaves entire, alternate, stipules absent; flowers with calyx of 3 sepals and a corolla of 6 petals in 2 rows; hypogynous; anthers adnate, filaments very short; pistils many, separate, or cohering in a mass, fleshy or pulpy in fruit; seeds large with a hard seed-coat, small embryo and copious endosperm. About 550 species, many in the tropics.

Asimina Adans. North American Papaw

Shrubs or small trees with solitary flowers from the axils of the leaves of the preceding year; sepals ovate, petals 6, imbricated in the bud; pistils few, ripening into 1; large and oblong, pulpy, several seeded fruits; seeds horizontal, flat. A small genus of about 7 species, natives of eastern North America and of Asia.

A. triloba Dunal. American Papaw

Shrubs or small trees with thin, obovate-lanceolate leaves, petals dull purple. Distribution. Along streams from Ontario and New York to McGregor, Iowa, Nebraska and Texas.



Fig. 247. Common Papaw (Asimina triloba). Common in woods of the South. (Charlotte M. King).

Poisonous properties. While the papaw is edible, there are some cases of poisoning on record. Care should therefore be exercised in its use. The leaves contain alkaloids. The papaw (Polyalthia argentea) contains asimin, a tasteless amorphous alkaloid.

LAURACEAE

Aromatic trees and shrubs with alternate or rarely opposite leaves without stipules; flowers small, fragrant, polygamous, dioecious or monoecious; calyx 4-6 parted; corolla absent; stamens in 3 or 4 series on the calyx, some imperfect; fruit a 1-seeded berry or drupe; endosperm none. About 1000 species, mostly tropical.

The spice-bush (Benzoin aestivale) produces fragrant flowers and aromatic leaves. Camphor $C_{10}H_{10}O$ is obtained from Cinnamon Camphora which comes from the islands of Formosa and Japan; cinnamon, a well known spice, comes from C. zeylanicum, which is extensively cultivated in Ceylon; cassia is from C. Cassia. Cassia and cinnamon were well known to the ancient especially to the



Fig. 248. Cinnamon Tree (Cinnamonum zeylanicum). Flowering and fruiting branch. The cinnamon of commerce is the bark of this tree. (After Faguet).

Israelites who used them as incense on their altars. The oils derived from these plants are excellent antiseptics.

Cinnamonum contains a volatile oil camphorin which is found in the roots and leaves; eugenol occurs in the leaves, stems and bark. Clove bark is derived from a small Brazilian tree (Dicypellium caryophyllatum) belonging to this family. The alligator pear or avocado (Persea gratissima), a native of the West Indies and tropical America, is much esteemed as a dessert fruit and in making salads. Rolfs has written of its successful culture in Florida. It is said that the oil is used extensively in America in soap manufacture. Leaves of laurel (Laurus canariensis), native to Canary and Medeira Islands, and bay (L. nobilis) of southern Europe are used in culinary processes. Nectandra Rodioei contains berberin, which is identical with the pelosin $C_{18}H_{21}$ NO₃ of Cissampelos Pancina. The Indian laurel contains laurotetanin $C_{19}H_{23}NO_{c}$. The California laurel (Umbellularia californica) is a strong local anaesthetic said to be irritant and acrid. The leaves, according to the Indians, will drive flies away.

Sassafras Nees

Trees with spicy aromatic bark; small mucilaginous twigs and foliage; flowers greenish yellow, naked in racemes; calyx 6-parted spreading, sterile, with 9 stamens, 3 inner with pair of glands; fertile flowers with rudiments of stamens; ovoid blue drupes.



Fig. 249. Sassafras (Sassafras variifolium). A well known tree of the south which furnishes the sassafras oil and bark of commerce. (W. S. Dudgeon).

Sassafras variifolium (Salisb.) Ktze. Sassafras

Leaves oval and entire, mucilaginous, or 2-6 lobed to about the middle and often as wide as long, membranous, pinnately veined, petioled; stamons about equaling the calyx-segments; fruiting pedicles red, much thickened below the calyx.

Distribution. From E. Mass. to S. E. Iowa, Kansas and southward. *Poisonous properties*. It is said to be poisonous; its bark however is used medicinally as a tonic and its wood is valuable.

RHOEADALES

Mostly herbs with regular and perfect flowers; sepals and petals usually present; polypetalous; stamens free; ovary superior, free from the calyx, compound, composed of 2 or more united carpels. It contains the families Resedaceae,, Cruciferae and Papaveraceae. The mignonette (Reseda odorata) is a well known, cultivated, fragrant plant of the family Resedaceae, which also includes the dyers wccd (R. luteola), the latter contains the substance luteolin.

Families of the Order Rhoeadales

PAPAVERACEAE. Poppy Family

Annual or perennial herbs, with milky or colored juice; leaves alternate, stipules none; perfect, regular, or irregular flowers; sepals 2, occasionally 3, falling when the flower expands; petals 4-12, spreading, soon falling; stamens inserted under the pistils, distinct; pistil 1, many ovuled, chiefly 1-celled; fruit a capsule containing numerous oily seeds. Genera 24-26, species about 200. Widely distributed chiefly in north temperate zone.

Comparatively few of the plants of this family are weedy and quite a number are medicinal and poisonous. The common poppy (Papaver somniferum) is used in medicine. It is found as an escape near buildings, especially in sections where Germans have settled, undoubtedly due to the fact that they cultivate it for its beauty as an ornamental plant, and use the seeds in culinary operations. The poppy is largely cultivated in China, Smyrna, Joppa, and several countries of Europe and India, for the opium. Opium yields a large number of alkaloids. The more important of these are morphin, and codein. A perfectly harmless oil equal to olive oil is obtained from the seed. The seed is also fed to birds. The red poppy (Papaver Rhoeas) is sometimes cultivated. A syrup is made from the petals, and also a coloring matter used in red ink. The California poppy (Eschscholtzia californica) is a valuable soporific, and analgesic "free from the disadvantages of opium." The Indians, according to Chesnut, use it to stupify fish. Celandine (Chelidonium majus) native to Europe, has been naturalized in places in the East and is occasionally somewhat weedy. The juice of this plant is yellow, while that of the common cultivated poppy is white, and that of the blood root (Sanguinaria canadensis) is reddish. The

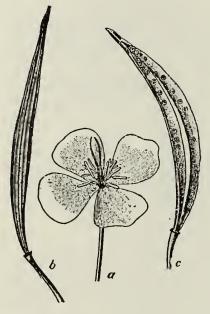


Fig. 250. California Poppy (Eschscholtzia californica). a, flower; b, fruit before, and, c, after dehiscence. The juice of this plant is a valuable soporific. (After Strasburger, Noll, Schenck and Schimper).

rhizome of the blood root is used in medicine and contains an alkaloid sanguinarin and a dye. The corydalin is found in a species of the genus Dicentra The bleeding heart (Dicentra spectabilis), native to China, and the climbing fumitory (Adlumia cirrhosa) are frequently cultivated for ornamental purposes. According to Blyth, the root of the tuberous-rooted corydalis (Corydalis tuberosa) contains eight alkaloides; of which corydalin $C_{22}H_{27}NO_4$ is the most important, since, when taken in large doses it may cause epileptiform convulsions, death taking place from respiratory paralysis. The C. lutea contains corydalin. Schlotterbeck and Watkins found 5 alkaloids in the American celandine (Stylophorum diphyllum) among them chelidonin $C_{20}H_{19}NO_5+H_2O$. The alkaloids stylopin $C_{19}H_{19}NO_5$, protopin $C_{20}H_{19}NO_5$, and sanguinarin, have been in part found in other plants in the family.

Genera of Papaveraceae

| Petals 8-12; pod 1-celled 2-valved. |
|---|
| Petals white; rootstock short red |
| Petals 4; pod 2-valved or more. |
| Flowers yellow4 Chelidonium. |
| Pod 4-20 valved. |
| Ovary incompletely many celled Papaver. |
| Stigmas and placentas 4-6 2 Argemone. |
| 4 D . D |
| 1. Papaver. Poppy |

Plant with milky juice, leaves lobed or dissected, alternate, flowers and buds nodding; sepals 2 or occasionally 3; petals 4-6; stamens numerous, ovules numerous; stigmas united into a persistent disk; capsule globose, obovoid or oblong; seeds small, with minute depressions. About 25 species, natives mostly of

the Old World. P. nudicaule is found in high mountains in the Rockies and in the Alpine regions of Europe and Asia.

Papaver somniferum L. Garden Poppy

An erect glaucous herb; leaves clasping, large, oblong, wavy, lobed or toothed; flowers broad, bluish-white with purple centre; filaments somewhat dilated, capsules smooth.

Distribution. Native to Asia, but widely naturalized in Europe, and extensively cultivated in China, India, and Smyrna. Occasionally found spontaneous around gardens in North America.

Poisonous and medical properties. From the milky exudation that comes from making an incision in the unripe eapsule, opium is obtained which yields not less than 5 per cent of crystallized morphin and occasionally as high as 22 per cent in Turkey opium, the usual yield being between these two extremes. The chief markets for opium are Turkey, Asia Minor, India, and Egypt, that of Smyrna being considered to be the best although good opium has been grown in the United States. This opium has a sharp, narcotic odor, and a bitter taste. Opium has been a fruitful source of a large number of alkaloids. Flückiger and Hanbury enumerate the following: "Hydrocotarnin, morphin, pseudomorphin (C_1 , $H_{18}NO_2$)₂+ H_2O , codein, thebain C_1 , H_2 , H_2 , H_2 , H_3 , H_3 , H_4 , H_3 , H_4 , H_4 , H_4 , H_5 , H_4 , H_5 , H_6 , H_7 , H_8 ,

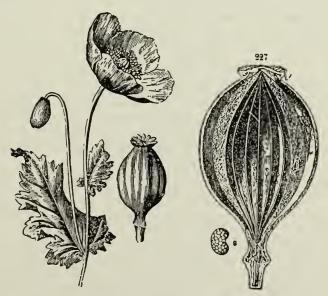


Fig. 252. Garden Poppy (Papaver sominferum). Flower and capsule. Opium is made from the milky juice exuding from the unripe capsule. (Faueg).

ly bitter taste which occurs in amounts varying from 0.5-2 percent. Narcotin is found in quanties varying from 0.75-9 percent.

It may be of interest in this connection to state that the German chemists Wolfgang, Weichardt, and Stadlinger found toxins in opium. These writers expressed the opinion that the complex physiological action is due to these substances.

In regard to the properties of opium, Flückiger and Hanbury speak as follows:

Opium possesses sedative powers which are universally known. In the words of Pereira, "it is the most important and valuable medicine of the whole Materia Medica;" and we may add, the source, by its judicious employment, of more happiness and, by its abuse, of more misery than any other drug employed by mankind.

There are occasionally cases of poisoning from the poppy plant. Certainly cases from overdoses of opium are frequently recorded in the annals of medical jurisprudence.

Opium may be absorbed to a slight extent by the unbroken skin, according to Winslow, and causes a mild, anodyne action. Opium diminishes the two principal activities of the digestive organs, namely, secretion and motion. The action upon the alimentary tract in lessening secretion, is partly a local one and partly constitutional, following the absorption of the drug. The mouth is made dry, thirst is increased and appetite impaired. Opium is absorbed rather slowly from the stomach and bowels, and stimulates the splanchnic nerve centre of the sympathetic system, which inhibits the movements of the stomach and intestines, and thus lessens peristaltic action of these organs. Opium is directly opposed to belladonna in this respect, as the latter drug paralyzes the intestinal inhibitory apparatus (splanchnic endings), and so increases peristalsis.

The most important action of opium is upon the nervous system, and its influence is more powerful upon man than upon lower animals. At first, opium exerts a stimulating influence upon the spinal cord. Ruminants are comparatively insusceptible to opium. Dr. Winslow says: "Ounce doses of the drug cause, in cattle, restlessness, excitement, hoarse bellowing, dry mouth, nausea, indigestion and tympanites. Sheep are affected in much the same manner. One or two drachms of morphine have led to fatality in cattle. Fifteen to thirty grains of the alkaloid comprise a lethal dose for sheep. Swine are variously influenced, sometimes excited, sometimes dull and drowsy.

According to the same authority, its action on horses causes drowsiness, sometimes, and at other times produces no visible effect,

Four to six grains, given in the same way, cause restlessness, a rapid pulse, and moisture of the skin. The animal paws the ground and walks in a rhythmical manner about the stall. The pupils are dilated. Large doses (12 grains) are followed by increased excitement, sweating, muscular rigidity and trembling; while still larger doses (four dracbms of the extract of opium) cause violent trembling, convulsions, insensibility to pain and external irritation, without coma; or (morphine, gr. 36 under the skin) stupor for several hours (3 hours), dilated pupils and blindness, followed by delirium and restlessness, continuing for a longer time (7 hours) and ending in recovery. Horses have recovered from an ounce of opium, but 2½ ounces of the drug, and 100 grains of morphine have proved fatal.

Dr. Winslow is here quoted upon the toxicology of opium:

The symptoms of poisoning have already been sufficiently described in previous sections. The treatment embraces irrigation of the stomach, or the use of emetics, as apomorphine hydrochlorate under the skin, and the subcutaneous injection of strychnine and atropine sulphate in the first stages, and enemata of hot, strong, black coffee; leading the animal about slapping him, or using the faradic current. Dr. Moor, of New York, has apparently found in potassium permanganate the most efficient antidote for opium and morphine. Ten to fifteen grains, dissolved in eight ounces of water, should be given by the mouth, to large dogs. One to two drachms of potassium permanganate may be administered to horses in two or three pints of water. Permanganate solution oxidizes and destroys morphine, and should be acidulated with a little vinegar or diluted sulphuric acid, after the ingestion of morphine salts. The antidote has been recommended to be given subcutaneously after absorption, or hypodermic injection of morphine, but this is not of the slightest use. Hypodermic injections have not infrequently caused poisoning.

Papaver Rhoeas L. Corn Poppy

An erect annual with hispid spreading hairs; lower leaves petioled, the upper, smaller, sessile, pinnatifid, lobes lanceolate, acute, and serrate; flowers scarlet with darker center; filaments dilated; capsule smooth with 10 or more stigmatic rays.

Distribution. In waste places along the Atlantic and Pacific coasts. It is frequently cultivated.

Poisonous properties. Poisonous like other species of this genus.

Freidberger and Fröhner give the symptoms of poisoning from this plant as colic, constipation, tympanites in cattle, raging fit of fury in horses.

In India this species is a troublesome weed but the seeds are collected and a yellow acrid oil obtained therefrom which is used both in medicine and as an illuminant.*

2. Argemone L. Prickly Poppy

Herbs with yellow juice; spiny toothed leaves and stems; flowers large; sepals 2-3; petals 4-6; stamens numerous; styles short; stigma 3-6-radiate; capsule prickly, oblong, opening by 3-6 valves; seeds small, numerous.

A small genus of about 8 species of the southern states, Mexico and Western North America.

Argemone mexicana L. Mexican or Prickly Poppy

A glaucus annual from 1-3 feet high, with spines or without; leaves sessile, clasping by narrow base, glaucus, runcinate-pinnatifid, spiny-toothed; flowers large, whitish or generally yellowish; calyx with 2 sepals, bristly pointed; stamens numerous; stigma sessile, seeds numerous, reticulated.

Distribution. Introduced along the Atlantic coast as far north as the middle states. Native from Florida to Texas. A most common and troublesome weed in Texas. It yields however a valuable painter's oil.

Argemone intermedia Sweet. Prickly Poppy

A spiny, leafy, plant from 2-2½ feet high; setose, hispid; flowers large white, sepals green, hispid; petals obovate; capsule armed with stout spines; horns with a terminal spine; seeds numerous, black sunken meshes.

Distribution. From Central Kansas, Nebraska, South Dakota, to the Rocky Mountains and Texas. Common in dry soil.

Poisonous properties. The small prickles cause somewhat painful injuries when they penetrate the skin. According to Schlotterbeck the A. Mexicana contains fumarin $C_{20}H_{19}NO_5$ and berberin. In Mexico used in the same way as the poppy.

Sanguinaria L. Bloodroot

Percnnial with a horizontal, thick rootstock; juice red; leaves basal, palmately veined and lobed, heart-shaped or reniform; flowers white; sepals 2, soon falling; petals 8-12, arranged in several rows; stamens numerous; placentae 2; capsule oblong, dehiscent to the base; seeds smooth, crested. A single species.

^{*} D. Hooper. Agrl. Ledger 1907:35.

Sanguinaria canadensis L. Bloodroot

Calyx; sepals 2, light green, falling as the bud opens; petals 8-12 or more, ½ to 1 inch long, oblong-spatulate, spreading, white or slightly rose-tinted, increasing in size for two or three days after the bud opens, and then falling away; stamens about 24, in several rows, much shorter than the petals, those in the inner rows longest; anther narrow, opening longitudinally.

Distribution. In rich woods, N. S. to Manitoba, Neb., Fla. and Ark.

Poisonous and Medical Properties. Lloyd in White's book on dermatitis, writes:

There are two native drugs that are very irritant to mucous surfaces, so much so that the dust is very disagreeable, and we presume that they would have a similar irritating action on the skin: Bloodroot, and Caulophyllum thalictroides, blue cohosh or pappooseroot.

Bloodroot has a bitter and acrid taste due to the substance sanguinarin. In small doses, this substance exerts a tonic influence, promoting gastro-intestinal secretion and thus aiding digestion. On its physiological action, Dr. Millspaugh says of sanguinarin $\rm C_{20}H_{16}NO_4$: "This alkaloid is very acrid to the taste, and toxic, and causes violent sneezing." Millspaugh gives its physiological action as follows:

Sanguinaria in toxic doses causes a train of symptoms showing it to be an irritant; it causes nausea, vomiting, sensations of burning in the mucous membranes whenever it comes in contact with them, faintness, vertigo, and insensibility. It reduces the heart's action and muscular strength, and depresses the nerve force, central and peripheral. Death has occurred from overdoses, after the following sequence of symptoms: violent vomiting, followed by terrible thirst and great burning in the stomach and intestines, accompanied by soreness over the region of those organs; heaviness of the upper chest with difficult breathing; dilation of the pupils; great muscular prostration; faintness and coldness of the surface, showing that death follows from cardiac paralysis.

Rusby says:

The effects of Sanguinaria canadensis L., or blood root are distinctly poisonous and Johnson definitely records that fatal results follow overdoses. Yet the rhizome is not at



Fig. 253. Blood root (Sanguinaria canadensis). The colored latex contains poisonous alkaloids.

all liable to he eaten, on acount of its peculiar blood red color, which is forhiddingly suspicious, and more especially hecause of an exceedingly acrid taste which would render the chewing and swallowing of a poisonous quantity an act of heroism. It is exceedingly common throughout the northeastern United States, and in a number of localities within a few miles of this city. The root also contains chelerythrin, homochelidonin and protopin.

4. Chelidonium L

Erect branching herbs, with alternate deeply pinnatifid leaves; yellow juice and flowers; 2 sepals; 4 petals; stamens numerous; distinct styles; capsule linear, dehiscent to the base; seeds smooth, shining, and crested.

Distribution. A genus of one species, native to Europe, but widely naturalized in North America.

Chelidonium majus L. Celandine

Flowers consisting of 2 sepals which are ovate, yellowish, soon falling; corolla 4 petals, contracted at the base; stamens numerous, shorter than the petals.

Poisonous and Medical properties. The alkaloid chelerythrin $C_{21}H_{17}NO_4$ is identical with the sanguinarin of the last plant. Chelidonin, $C_{20}H_{10}NO_5$ H_2O , an alkaloid existing particularly in the root, is colorless and bitter. Homochelidonin, consisting of three basic substances is found in Bocconia, Sanguinaria, Adlumia etc. This plant produces congestion of the lungs and liver; it is also an excessive irritant, and has a narcotic action upon the nervous system, in its action resembling gamboge. On this point Dr. White says:

Mr. Cheney informs me that he has known the plant to poison the skin, if handled so as to crush the leaves or stem. To indicate this extent to which it is used in medicine, it may be stated that a collector in North Carolina offers fifteen hundred pounds of the leaves for sale.

CRUCIFERAE. Mustard Family.

Herbs or rarely woody plants with acrid, watery juice; alternate leaves without stipules; flowers in racemose or corymbose clusters, cruciform of 4 deciduous sepals and 4 petals, placed opposite each other in pairs, spreading and forming a cross; stamens 6, 2 shorter; 1 pistil, consisting of 2 united carpels; fruit a pod either much longer than broad (silique), or short (silicle), or in-



Fig. 254. Common Celandine (Chelidonium majus). Poisonous to the skin. (After Fitch).

dehiscent, separating into joints; seeds without endosperm; seed coat frequently mucilaginous; embryo large. About 1500 species of wide distribution.

The cabbage (Brassica oleracea), native to Europe, has long been cultivated; cauliflower, brocoli, and brussels sprouts also belong to the same species. Rape, a well known forage plant, the turnip, the Swedish turnip, and rutabaga (B. campestris) are native to Europe. The Chinese cabbage (B. Pe-Tsai) is commonly cultivated in China. Black mustard (B. nigra) and white mustard (B. alba) are extensively cultivated for their seeds, which when ground make the commercial mustard. The radish (Raphanus sativus), cultivated for the root, is native to Europe. Water cress (Radicula nasturtium-aquaticum) is much used as a salad plant in colder regions. Horse radish (R. Armoracia) the well known condiment is native to Europe. European pepper grass (Lepidium sativum) is cultivated as a salad plant, while the seeds of our pepper grasses (Lepidium apetalum and L. virginicum) are used as bird food. Seakale (Crambe maritima), native to Europe, has also been introduced into this country as a vegetable. The Pringlea antiscorbutica of Kuerguelen's Land resembles the common cabbage and is used by sailors as a vegetable when they touch that country. The Rose of Jericho (Anastatica hierochuntica) of North America and Syria is regarded as sacred by the natives. Many plants of this family are cultivated for ornamental purposes; among the most familiar are the candytuft (Iberis), stock (Matthiola incana), sweet alyssum (Alyssum maritimum), and wall-flower (Erysimum asperum). Dyer's woad (Isatis tinctoria) of China was formerly cultivated for a dye obtained from the leaves. The characteristic odor of plants of this order when crushed or when mustard seed is ground in water is due to an enzyme myrosin, discovered by Bussey in 1839. This same ferment occurs in the families Cruciferae, Capparidaceae, Resedaceae, Tropaeolaceae, Limnanthaceae, Papaveraceae.

It occurs in special cells known as myrosin cells which give a marked protein reaction. The contents are finely granular, free from starch, chlorophyll, fatty matter, and aleurone grains. These cells become red with Millon's reagent, and when heated become orange red, and a violet red color when treated with copper sulphate and caustic potash. The subject is treated fully by Reynolds Green in his work on "The Soluble Ferments and Fermentation," and in Effront and Prescott's "Enzymes and their Applications."

Some of the European plants of the family like (Erysimum crepidifolium) cause staggering in animals. The Cheiranthus Cheiri contains a glucoside which acts on the heart. Rape under some conditions is poisonous. Several correspondents in Breeders' Gazette (Chicago) have ascribed poisoning where rape was frozen, or when the plants were wet with dew.

Genera of Cruciferae

Pod terete or turgid or 4-angled.

Cotyledons accumbent.

Pod angled or terete; flowers yellow or white.....1. Sisymbrium Cotyledons conduplicate; flowers yellow or white......2. Brassica

Pod short.

| Pod many | or | few | seeded; | obcordate-triangular. |
|----------|----|-----|---------|-----------------------|
| 75 1 | | | 1 1 1 | 1 4 4 2 1 |

| Pod | many seeded, obcordat | e-triangular. | | Capsella |
|-----|-------------------------|---------------|------------|----------|
| Pod | few seeded, orbicular, | obovate or | obcordate7 | Thlaspi |
| Pod | 2-seeded, flat, notched | | 6 L | epidium |

1. Sisymbrunt (Tourn.) L.

Annual or biennial herbs with usually simple spreading pubescent, hairs occasionally forked or stellate; leaves entire, oblanceolate, divided, pinnatifid, or runcinate; flowers in raceines; calyx open, greenish, of 4 sepals; corolla white, yellow or yellowish, or rarely pink, small; pistils terete, flattish or 4-6-sided; small entire stigma; seeds small oblong; cotyledons incumbent.

A small genus of 60 species. Found in temperate regions of both hemispheres. Several species are well known troublesome weeds.

Sisymbrium officinale Scop. Common Hedge Mustard

A slender erect annual or winter annual, $1\frac{1}{2}-2\frac{1}{2}$ feet high; lower leaves divided, runcinate, pinnatifid, upper entire or hastate at base; flowers small, yellow, borne in spike-like racemes; seeds small, brown; cotyledons incumbent.

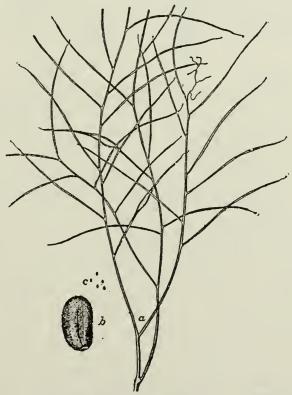


Fig. 255. Tumbling Mustard (Sisymbrium altissimum). Common in Canada and from Minnesota to Washington. (Dewey, U. S. Dept. Agrl.)

Distribution. A very common weed along railroads, door yards, and fields from Canada south to Florida and west to Illinois, Wisconsin, Minnesota, Missouri, Dakotas, Nebraska and Kansas, and Pacific North coast. Common hedge mustard is a naturalized weed from Europe. It occurs throughout Europe, Russia, Germany, Great Britain, France—except Northern Scandinavia.

Sisymbrium atlssimum L. Tumbling Mustard

An erect, much branched annual from 1-4 feet high, lower leaves runcinate pinnatifid, irregular toothed or wavy margined; upper leaves smaller, thread-like; after flowering, leaves drop, leaving the stem and pods; flowers pale yellow, rather large; sepals 4, green; corolla of pale yellow petals; pods narrowly linear, divergent; seeds small, longer than broad, generaly oblong in outline with rather blunt ends; radicle usually very prominent and straight, curved spirally around the cotyledons.

Distribution. This weed has spread with considerable rapidity in the Northwest. Dr. Robinson states that it was once scarcely more than a ballast weed about the large cities of the Atlantic seaboard, and records its occurrence sparingly in southern Missouri (Bush). It is common now, however, from the Mississippi Valley northwest to the Pacific Coast. One of the most common weeds of Montana, Idaho, Eastern Washington, Oregon, and native to British Columbia. Found also along the Atlantic seaboard.

Poisonous properties. This plant has properties somewhat similar to those described for Mustard; therefore may produce deep ulcers which are difficult to heal.

Brassica (Tourn.) L. Mustard, Turnip, Rape

Annual or biennial branching herbs, basal leaves pinnatifid, flowers yellow, racemose, pods elongated nearly terete or 4 sided; seeds spherical, 1 row in each cell; cotyledons conduplicate. About 85 species of Europe, Asia, and North Africa, introduced in North and South America, Australia. The black mustard (B. nigra) and charlock (B. arvensis) common in grain fields and waste places across the continent.

Brassica nigra Koch. Black Mustard

A tall, coarse, much branched annual, 2-5 feet high; hairy or smoothish, somewhat bristly, at least on the veins; leaves variously divided or deeply cut, and sharp toothed;—large terminal lobe; the upper leaves small, simple, usually linear; flowers yellow, smaller than in charlock; pods smooth, about ½ inch long, 4-cornered, tipped with a slender beak; seeds black or reddish brown, smaller than in charlock; cotyledons conduplicate.

Brassica arvensis (L.) Ktze. Mustard or English Charlock

Branching annual from 1-3 feet high, hispid or glabrate; lower leaves petioled with 1 large terminal lobe, and several small lateral lobes, with the divisions unequal; upper leaves barely toothed; flowers yellow, large and very fragrant; pods 1-2 inches long, irregular in outline, appearing somewhat nodose, 3-7 seeded, or more occasionally; the upper part of pod forms the beak; seeds round, brownish black, darker than in *B. nigra*, and more minutely pitted. When moistened, the seeds become mucilaginous.

Distribution. It is a common and troublesome weed in cultivated ground from Mass. to Oregon. The most troublesome weed in grain fields of the Northwest.



Fig 256. Charlock (Brassica arvensis). U. S. Dept. Agrl. Fig. 256a. Common Black Mustard (Brassica nigra). U. S. Dept. Agrl.

Medicinal and Poisonous properties. White mustard contains a glucoside also found in other members of the family sinalbin, $C_{30}H_{42}N_2S_2O_{15}+H_2O$, and in addition, the ferment known as myrosin which converts the sinalbin into an active principle, oxy-benzyl-thiocyanate (a very acrid volatile body) sinapin sulphate and glucose. The following formula represents the change that occurs: $C_{30}H_{44}N_2S_2O_{16}=C_6H_{12}O_6+C_7H_7O-NCS+C_{16}H_{24}NO_5-HSO_4$.

OxySinalbin Glucose benzyl-thiocyanate Sinapin sulphate

Black mustard contains the glucoside *sinigrin*, and a ferment, *myrosin*, which produces the following reaction:

$$C_{10}H_{18}NKS_2O_{10} = C_3H_5CNS + C_6H_{12}O_6 + KHSO_4$$
.
Sinigrin Sulpho-cyanate Glucose Hydrogen of allyl Potassium sulphate (Essential oil of Mustard)

White Mustard sced when reduced to a powder and made into a paste with cold water, acts as a powerful stimulant. Large doses cause vomiting. Intestinal secretion is increased by the use of mustard, which is rarely used, however, as an emetic because of its pungency. It is extremely valuable for relieving pain or congestion. It is a splendid emetic for dogs in the spoonful doses, given in warm water. Dr. Millspaugh states that in the case of black

mustard, no specific toxic symptoms have been noted, but in speaking of white mustard, he states that the essential oil of mustard is a virulent, irritant poison, causing, when ingested, severe burning, followed by increased heart action, and if pushed to extremes, loss of sensibility, paralysis, stupor, rigors, and death. It causes immediate vesication, followed by deep ulceration, which is difficult to heal.

Dr. White in his Dermatitis Venenata, says:

The action of the sinapism is well known. In a few minutes after its application the skin begins to feel warm, and by the end of a half-hour, if the patient bear it so long, this sensation has increased to an intolerable burning. The changes in the cutaneous tissue are, within a few minutes, a considerable degree of hyperaemia, which, after a time, increases to an intense redness, which persists for a day or two, and often leaves behind it a persistent pigmentation, at times of a dark brown color, to mark the seat of the sinapism. On this account one should never be applied upon the upper chest or other part of a woman which the dress will not always conceal. If the action be continued beyond its legitimate rubefacient effect, a period which varies greatly in persons, it may produce vesication, or even deep suppuration, effects at times very intractable under treatment. It is stated that the addition of vinegar to a mustard poultice greatly lessons its activity.

Sarepta mustard (B. Besseriana) much resembles black mustard and is used in the same way. Sarson or Indian colza (B. campestris var. Sarson) is used in India in place of white mustard and oil cakes made from it serve as stock food. A brown variety (B. Napus var. dichotoma) is grown both as an oil seed and as a vegetable. Another variety, Indian mustard (B. juncea) is also known as of economic importance.

The mustards of India are not always easy to distinguish. They have been carefully studied by Col. Prain.

Brassica Rapa causes inflammation of the bowels, tympanites, constipation, diarrhoea, some brain irritation, and haemorrhagic enteritis. The disease is more common in Europe where rape seed is used as stock food.

3. Radicula (Dill.) Hill

Annual, biennial or perennial, usually glabrous herbs of pungent qualities, pinnate, entire, or pinnatifid leaves; flowers yellow or white; calyx with spreading sepals; pistil with short or slender style and 2-lobed, or entire stigmas; pod usually short, varying from oblong-linear to globular-terete; seeds numerous, small in 2 rows in each cell; cotyledons accumbent. About 25 species of wide distribution.

Radicula Armoracia (L.) Robinson. Horse radish.

A stout perennial with long deep roots; leaves large, on thick petioles, oblong, crenate, or pinnatifid, glabrous; stem leaves lanceolate, or oblong cordate; flowers with 4 green sepals and 4 white petals, not common; pods short, globular, but fruit seldom found.

Distribution. It is native to the eastern part of Europe, Turkey, Greece, and the Caspian Sea through Russia, Poland and Finland. In Germany, France, Sicily, Norway, and Great Britain, it has escaped from cultivation. Common in Northern United States.

Radicula palustris (L.) Moench, var. hispida (Desv.) Robinson. Marsh Cress.

An erect annual, or biennial, pubescent herb, from 1-2½ feet high; leaves pinnately cleft or parted, or occasionally the upper laciniate; the lobes toothed;



Fig. 256b. Horse radish. (C. M. King).

upper leaves nearly sessile; pedicels as long as the small flowers, generally longer than the pods; pods ovoid or oblong; styles short.

Distribution. Common in northern portions of United States, to the Gulf and west to the coast; also Canada. Native to Europe.

Medical and poisonous properties. It is certain that horse radish and other members of this genus have properties somewhat similar to those of the mustards mentioned above. Horse radish contains both sinigrin and myrosin. Dr. Rusby mentions that it may produce serious trouble. He says:

The common horse radish, likewise, loses its irritating properties when heated or dried. These are almost identical with those of mustard, and while it would not generally he regarded as a poisonous article, yet used in excess it may become so through its powerful irritation of the urinary organs, by which it is excreted. Johnson gives a case in which this result was extreme and serious. It may therefore be borne in mind that it should not be consumed in inordinate quantity. This result, should it occur, would be found excruciatingly painful.

Dr. Johnson in his Manual of Medical Botany of North America, writes as follows:

The acrid principles of these plants appear, clinically, to be eliminated by the kidneys, and hence, incidentally, they produce a decided diuretic effect. The urine is not only increased in quantity, but partakes also of the acrid character of the plant employed. In one case that came under the author's observation, the individual, though in perfect health, so far as the genito-urinary tract was concerned, suffered extremely from vesical pain and irritation for hours after using horse-radish as a condiment. In animals it produces a violent colic.



Fig. 256c. Shepherd's Purse (Capsella Bursapastoris). U. S. Dept. Agrl.

4. Capsella, Medic. Shepherd's Purse

Annual or winter annual, erect herbs, pubescent with more or less branched hairs; flowers in racemes, small, white; basal leaves tufted; pistils with short styles; pods obcordate, triangular, compressed at right angles to the partition; valves boat-shaped; seeds numerous, small, without margins; cotyledons accumbent. A small genus of 4 species, 2 in North America.

Caspella Bursa-pastoris (L.) Medic. Shepherd's Purse

An annual or winter annual, 1½ feet high, root leaves clustered, lobed, pinnatifid, or merely toothed, stem leaves sessile, lanceolate, auricled; flowers small, white; pods triangular, truncate, or emarginate, many seeded; seeds light brown, clongated with a prominent ridge; seeds mucilaginous when moistened with water; cotyledons incumbent.

Distribution. One of the most common weeds everywhere in eastern North America from Nova Scotia to Florida, west to Texas and the Pacific Coast, from Eastern Canada to Manitoba to Vancouver in B. C. Cosmopolitan, as common in Europe as in the United States. Naturalized from Europe.

Poisonous properties. It produces the same symptoms as other members of the family only somewhat less severe.

5. Camelina, Crantz. False Flax

Erect, annual herbs, sparingly branched; leaves entire, lanceolate, or pinnatifid; flowers racemose, yellow; sepals 4, green; petals 4, yellow small; pistil with stigma entire, style slender; pod obovoid or pear-shaped; flattish; seeds small, numerous; cotyledons incumbent. The 5 species are native to Europe and Asia.

Camelina sativa (L.) Crantz. False Flax

An erect, glabrous annual with simple or sparingly branched stem, 1½ feet long, smooth, or slightly pubescent, hairs stellate; leaves erect, lanceolate or arrow shaped, entire or nearly so; flowers small, yellow, pedicels in fruit spreading; pod obovoid or pyriform, smooth reticulated, margined; seeds light brown 1 line long, minutely pitted, cotyledons incumbent, caulicle prominent, running lengthwise. On the addition of water, the seeds become mucilaginous.

Distribution. It is common in Manitoba, south to Minnesota, Northern Iowa, and the Dakotas, where it is a well known and a troublesome weed, in flax and grain fields from Ontario to the Middle States across the continent.

Poisonous properties. The plant has a disagreeable sharp odor and causes counter-irritation.

6. Lepidium (Tourn) L. Pepper Grass

Ercct or diffuse, annual, biennial or perennial herbs; leaves entire, or pinnatifid; flowers racemose, white; petals small or none; stamens 6 or fewer; pod roundish, flattened contrary to the partition, winged or wingless; seeds solitary in each cell; cotyledons incumbent or rarely accumbent. About 65 species in temperate regions. The European *L. campestre*, native to Europe cultivated for salad purposes, is occasionally spontaneous. The seeds of two of our native species are used for bird food.



Fig. 256c. False Flax (Camelina sativa). Seeds become mucilaginous on addition of water. (After Hochstein).

Fig. 257. Small Pepper-grass (Lepidium apetalum). Causes sinapism. (Charlotte M. King.)

Lepidium virginicum L. Large Pepper Grass

Pod circular or oval with a little notch at the upper end; seeds light brown, elongated, with a prominent ridge on one side, on the addition of water they become mucilaginous; cotyledons accumbent.

Lepidium apetalum Willd. Small Pepper Grass

Seeds light brown, elongated, with a prominent ridge on one side. Seeds become mucilaginous when moistened with water. Cotyledons incumbent.

Distribution. In nearly all parts of the United States.

Poisonous properties. Pepper grass produces counter-irritation.

7. Thlaspi L.: Field Pennycress

Low plants with undivided root leaves, stem leaves arrow-shaped and clasping; flowers small, whitish or purplish; pod orbicular, obovate or obcordate; seeds 2-8 in each cell; cotyledons, accumbent.

Thlaspi arvense, Field pennycress, Frenchweed or Stinkweed

A smooth annual with small white flowers; pod, broadly winged, about ½ inch in diameter, deeply notched at top. Commonly naturalized in some places; becoming more abundant in the Northwest. A common weed in grain fields. It is common in the Canadian Northwest and not infrequent in Iowa and Minnesota, abundant in the Dakotas.

Poisonous properties. Probably causes counter-irritation. In the Canadian Rockies the weed is carefully avoided by stock because of its pungent properties.



Fig. 258. Field Pennycress (Thlaspi arvense). (After Fitch).

CAPPARIDACEAE. Caper Family

Herbs, shrubs or, occasionally, trees; alternate leaves and cruciform flowers; sepals 4-8; petals 4 or none; stainens 6-numerous, not tetradynamous; fruit a 1-celled pod or berry with 2 parietal placentae; seeds similar to those of the *Cruciferae*, but with the embryo coiled. An order of about 35 genera and 400 species. Generally found in warm regions, few in the United States. The plants are often acrid or pungent; the flower-buds of one, the Caper (Capparis spinosa), are pickled. Several of the species like the Rocky Mountain bee plant are cultivated for ornamental purposes. A few of the plants are weedy.

Capparis contains the coloring matter rutin.

Genera of Capparidaceae

| Plants clammy pubescent. | |
|----------------------------------|------|
| Stamens 8 or more | sia |
| Plants not clammy pubescent. | |
| Stamens 6, pod few seeded | ella |
| Stamens 6, pod many sceded1 Cleo | me |

1. Cleome L.

Glabrous annuals; leaves trifoliolate or simple; flowers in leafy bracted racemes; petals entire, with claws; stamens 6; pistil with a 1-celled ovary;

pod linear with a long stalk (stipe) many seeded; the receptacle bearing a gland beyond the stipitate ovary. A small genus of about 75 species, mainly tropical.

Cleome serrulata Pursh. Rocky Mountain Bee Plant

An annual, from 1-3 feet high, with digitate, 3-foliolate leaves and leafy, bracteate racemes; calyx 4-cleft, petals 4, cruciform, short clawed, and rose-colored; pods linear, many seeded.

Distribution. Widely distributed west of Missouri, from the Canadian Rockies to Kansas, Mexico, Arizona and Utah, eastward occasionally from Minnesota to Illinois. The plant is regarded with great favor as a bee plant.

Cleome lutea Hook. Yellow Cleome

This plant is like the preceding, but the leaves are 5-foliolate or the upper 3-foliolate, leaflets oblong or oblong-lanceolate, entire, stalked or sessile; flowers yellow; pod linear, stipe longer than the pedicel.

Distribution. In dry soil from Nebraska to Washington and Arizona. The former species is particularly conspicuous west of the 100th meridian.

Poisonous and Medical properties. These plants are not generally placed with the poisonous plants, although they contain the same pungent principles that members of the Mustard family have. They are seldom eaten by stock.



Fig. 259. Rocky Mountain Bee Plant (Cleome serrulata). A plant with pungent properties. (Ada Hayden).



Fig. 260. Yellow Cleome (Cleome lutea). Common in the West. (Ada Hayden).

2. Polanisia Raf.

Clammy herbs with whitish or yellowish flowers, palmately compound or simple leaves; flowers produced in racemes; sepals 4, deciduous; petals with claws and notched at the apex; receptacle not clongated, bearing a gland at the base of the ovary; stamens 8-numerous, unequal; pod linear or oblong, turgid, many-seeded, seeds reticulated. About 14 species in tropical and temperate regions. Annuals, with glandular hairs; common in sandy soils or on railroad embankments.

Polanisia graveolens Raf. Clammy-weed

The near relative of the Rocky Mountain bee plant is a clammy weed with loose racemes of conspicuous flowers; petals with claws; stamens 8-32; pod linear or oblong, turgid, many-seeded.

Poisonous properties. The same may be said of this as of Cleome. It is a clammy, pubescent weed with very pungent properties.

3. Cleomella DC.

Annual herbs with 3-5 foliolate leaves, calyx of 4 sepals; flowers generally in racemes; petals 4, entire, without claws; receptacle short; stamens 6, inserted on the receptacle; ovary short, long-stalked; pod linear to oblong, many-seeded. About 75 species, found chiefly in southwestern North America and Mexico.

Cleomella angustifolia Torr

A glabrous annual from 1-2 feet high, leaflets 3, linear lanceolate or linear oblong, bracts simple; flowers small, yellow; pod rhomboidal, raised on a slender stipe, but shorter than the pedicel, few seeded.

Distribution. From Nebraska and Kansas to Texas, New Mexico and Colorado. Abundant in waste places.

SARRACENIALES

Carnivorous plants secreting a viscid liquid; radical leaves; scapose flowers; corolla choripetalous; sepals generally distinct; stamens usually free; ovary compound superior. Contains the families Droseraceae, Sarraceniaceae and Nepenthaceae; the genus Sarracenia has 6 species in eastern North America; S. purpurea, found as far west as Minnesota, contains the alkaloid Sarracenin. Darlingtonia californica occurs in California and Oregon. Heliamphora is native to Guiana. The family Nepenthaceae with 40 species is found mostly in the India-Malayan regions; some species being frequently cultivated in greenhouses. The plants of these orders are insectivorous, capable of digesting insects.

Droseraceae

Perennial or biennial glandular pubescent bog herbs or somewhat shrubby plants; leaves mostly from the base with tentacles, which secrete a viscid substance to catch insects; circinnate in the bud; flowers perfect, racemose; calyx persistent, 4-8 parted, or the sepals distinct; petals 5 free; stamens 8-20; ovary free, 1-3-celled; styles 1-5, simple 2-cleft; capsule 1-5-celled. A small order of 125 species of wide distribution. The most important genus is *Drosera*, commonly called sundew, the tentacles of which secrete a viscid fluid which catches

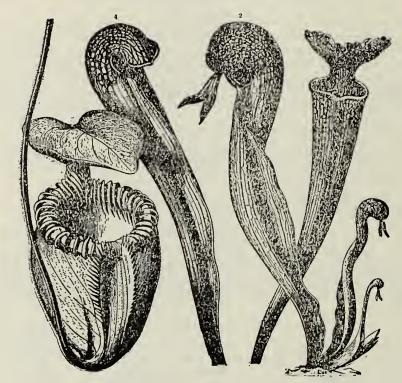


Fig. 260a. Various insectivorous plants: 1, Sarracenia variolaris; 2, Darlingtonia californica; 3, Sarracenia laciniata; 4, Nepenthes villosa.

insects and clings to them. The *D. rotundifolia* is commonly found in our northern bogs. The *Drosophyllum lusitanicum* is found on the sandy hills of Portugal. The Venus fly-trap (*Dionaea muscipula*) of the Carolinas grows on the sandy barrens and feeds on insects. These plants are somewhat rare.

Poisonous properties. According to Dr. Schaffner, the common sundew is poisonous to cattle. From one species of Drosera two pigments have been isolated, the rcd having the formula $C_{11}H_8O_5$ and the yellow, $C_{11}H_8O_4$. Plants of the family in Australia are said to be poisonous to sheep.

According to Greshoff the leaves of *Drosera binata* contain hydrocyanic acid. *D. rotundifolia*, and *D. intermedia*, were also found to contain a little HCN. The leaves of *Dionaea muscipula* contain the same substance.

ROSALES

Herbs, shrubs or trees; flowers usually polypetalous; stamens mostly perigynous or epigynous; sepals chiefly united or confluent with receptacle; carpels 1 or more, distinct or sometimes united into a compound ovary. The order includes the families Podostemonaceae, Crassulaceae, Cephalotaceae, Saxifragaceae, Pittosporaceae, Hamamelidaceae, Platanaceae, Rosaceae, Connaraceae, Leguminosae and other small families. The family Saxifragaceae includes the red currant (Ribes vulgare), black currant (R. nigrum), the Missouri currant,

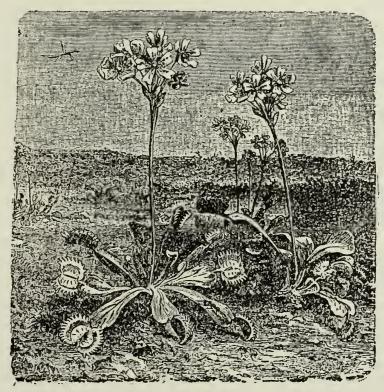


Fig. 260b. The Venus Fly Trap (Dionaea muscipula) showing the rosette arrangement and with some of the traps closed and others opened to catch the insects. (Kerner-Oliver, Dept. Ent. Univ. of Minn.).

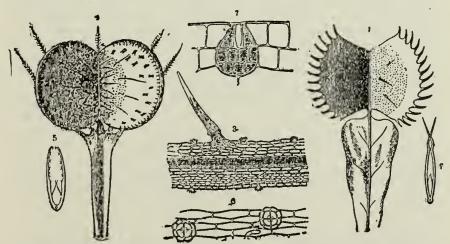


Fig. 260c. Insect Traps. 1, Dionaca; 2, Section through folded leaf; 3, One of the Spines. 4, Leaf of Aldrovanda; 5, Section of closed leaf; 6, Glands upon trap; 7, Glands in the wall of trap of Sarracenia. (Kerner. Dept. Ent. Univ. of Minn.).



Fig. 261. Common Sundew (Drosera rotundiflora). Poisonous to cattle. (After Fitch).



Fig. 262. Missouri Currant (Ribes aureum). The flowers are spicy fragrant. It is a well known shrub. (W. S. Dudgeon).

(R. aureum), the Crandall, a well known cultivated form; the cultivated gooseberry (R. Grossularia), Missouri gooseberry (R. gracile); the cultivated hydrangeas (Hydrangea paniculata); the wild hydrangea (H. arborescens); used as a diuretic; the mock orange (Philadelphus coronarius), and P. grandi florus; Deutzia scabra, the Astilbe japonica and the alum root (Heuchera americana) a powerful astringent. Several members of this family are poisonous; the western Jamesia americana, a pretty shrub of the Rocky Mountains with white flowers, contains an appreciable amount of HCN, according to Greshoff. The same substance also occurs in the garden Hydrangea hortensia, H. arborescens, an American species of the southern region. The H. Thunbergii; H. Lindleyana and H. involucrata all contain HCN. The HCN, however, is in a transitory stage. Greshoff states that the leaves of Philadelphus coronarius, P. Lemoinei and P. microphyllus, and the seeds of P. grandiflorus contain saponin, as do the leaves of Deutzia staminea. The family Hamamelidaceae includes the witch hazel (Hamamelis virginiana) containing a bitter principle; the storax (Liquidambar orientalis) of the Old World which contains two resin alcohols, storesin and ester, and red gum (L. Styraciflua) a common tree of moist woods of the South supplying a well known commercial wood; it is also an ornamental tree, the leaves of which, when bruised, are fragrant. The family Platanaceae contains the sycamore (Platanus occidentalis), a large tree which furnishes valuable wood and is also used for ornamental purposes. According to Greshoff the leaves of Platanus acerifolia, P. cuneata, P. occidentalis and P. orientalis contain HCN. The young leaves are poisonous.

Families of Rosales

| alyx free | from the ovary, wholly superior. | |
|-----------|--|----------------|
| Simple | 1-celled | . Leguminosae. |
| Ovarie | s 2 or more compound; stamens twice as many as the | pistils |
| | | Crassulaceae |

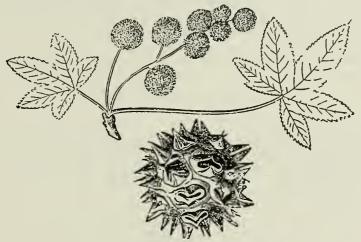


Fig. 251. Red Gum (Liquidamber Styraciflua). Furnishes a commercial wood.

CRASSULACEAE DC. Orpine Family

Succulent herbs; leaves generally sessile without stipules; flowers small, symmetrical, usually cymose; petals and sepals equal in number, from 3-20; stamens as many or twice the number; pistils distinct, fewer than the sepals; receptacle usually with small scales 1 back of each carpel; fruit a dry dehiscent follicle; usually many seeded. This is a small family of about 50 species, many being found in dry soils, rocks, etc. in North Temperate regions. A few of the species are cultivated for ornamental purposes, among these being the Rochea coccinea of the Cape of Good Hope, whose flowers have narcotic properties; poisoning sometimes resulting from smelling them. Some of the Crassulaceae contain crassulacic and malic acids. The Cotyledon ventricosa of South Africa is said to produce the Nenta disease of that region, although this is usually ascribed to some members of the Pulse family and is probably identical with loco disease. The leaves of wild tea (Catha edulis), according to Bull. Miscellaneous Information of Kew Gardens, when chewed are said to have great sustaining powers.

Sedum (Tourn.) L. Stone Crop

Fleshy, smooth herbs, mostly perennials, with alternate leaves; flowers cymose, perfect or dioecious; calyx 4-5-lobed; petals 4-5, distinct; stamens 8-10, perigynous; pistils 4-5, distinct or united at the base; styles short; follicles many-seeded. About 150 species, mostly of temperate and cooler regions of the northern hemisphere. A few are cultivated for ornamental purposes. There are several somewhat weedy species as S. purpureum and mossy stone crop (S. acre), the latter native to Europe but escaped to roadsides in the East. It has acrid properties, which is also true of the live-forever, a plant of the Rocky Mountains.

Sedum purpureum Tausch. Live-forever

A stout perennial 2 feet high with fleshy oval or obtuse, toothed leaves; and flowers in compound eymes; eorolla purple, with oblong-lanceolate,, purple petals; stamens perigynous; pistils with a short style; fruit a follicle with a short pointed style.

Distribution. Native to Europe, frequently escaped from cultivation and found around dwellings and in cemeteries.

Sedum acre L. Mossy Stone Crop

A moss-like plant spreading on the ground; leaves small, alternate, ovate thick and fleshy; flowers yellow, perfect, in spike-like clusters, cymose; the central flower with 5 sepals, 5 petals, and 10 stamens, the others with 4 sepals, 4 petals and 8 stamens; follicle spreading, tipped with a slender style.

Distribution. On rocks and along roadsides from New Brunswick to New York and Pennsylvania. Native to Europe.



Fig. 263. Common Livefor-ever (Sedum purpureum). A plant with acrid properties. (Fitch).



Fig. 264. Live Forever (Sedum stenopetalum). Common in the Rocky Mountains; a plant with acrid properties. (W. S. Dudgeon).

Poisonous properties. Sedum acre, according to Dr. Schaffner, produces inflammation and vesication when applied to the skin; it is used to remove the false membrane in diptheria. Dr. White, in his "Dermatitis Venenata," says with reference to the Sedum acre: "Wood states that the whole plant abounds in an acrid, biting juice. Oesterlen says that it is sharply irritative to the skin. The National Dispensatory states that the juice is capable of blistering the skin, and that it is used upon corns and warts to soften them, and upon swollen glands as a resolvent. Mr. Cheney, a wholesale dealer in vegetable drugs, informs me that the juice of the green plant is poisonous to the skin of many persons." This plant, however, is not common in the United States.

Rosaceae. Rose Family

Herbs, shrubs, or trees; leaves alternate or some opposite, stipulate, frequently falling soon after the leaves appear; flowers regular; stamons generally numerous, distinct, inserted on the calyx; petals as many as the sepals or rarely wanting; pistils 1-many, generally distinct, except in Pomeae, where the pistil is united to the calyx; fruit various, achenes, follicles, drupes or pomes as in the apple; seeds 1-many, without albumen; embryo straight, with large cotyledons. The order contains about 90 genera and 1500 species of wide distribution; in temperate and tropical regions, some boreal. But few of the plants are noxious or have noxious qualities.

The several cherries, like Prunus serotina and P. virginiana are known to cause stock poisoning, and the seeds when eaten likewise produce fatal results in man. The P. serotina or wild black cherry, is used in medicine, under the name of P. virginiana. The choke cherry (P. virginiana) is also used in medicine. The bark of the wild black cherry is officinal. It contains tannic and gallic acids, and a volatile oil resembling the volatile oil of bitter almonds. It is used as a tonic and astringent. The leaves of the laurel cherry arc used for making cherry laurel water which is a sedative narcotic. From Prunus Amygdalus, var. amara, native to Asia, is obtained the amygdalin of bitter almonds, which is converted into hydrocyanic acid. This acid is deadly poison, and is obtained from a great many different plants. The leaves of the laurel cherry also contain the same substance. HCN has been found in Prunus paniculata, P. pendula, P. Pennsylvanica; Pyrus Aria, P. pinnatifida, P. japonica; Crataegus orientalis; Cotoneaster integerrima; Nuttallia cerasiformis; Amelanchier alnifolia; Chamameles japonica. According to Greshoff the leaves of Kageneckia angustifolia eontain hydrocyanic acid. The same writer reports this substance in the mountain mahogany (Cercocarpus parvifolius) of the Rocky Mountains. It must be regarded as poisonous. In laurel cherry, it is largely derived from the decomposition of laurocerasin. This species also contains prulaurocerasin. Several species of other orders also might suitably be mentioned here as containing substances capable of being converted into hydrocyanic acid. In this class are the toadstools (Agaricus oreades), bitter cassava (Manihot utilissima) and sorghum (Andropogon Sorghum).

According to Greshoff Cornus foliolosa, Spiraca japonica and many other plants of the family contain saponin.

The same substance is obtained from other plants of the genus Prunus. Oil of roses is obtained from the Rosa damascena, Miller, var. The mucilaginous seeds of the quince (Pyrus Cydonia) have been used in medicine for a long time.



Fig. 265. Wild Red Cherry (Prunus pennsylvanica). It contains the glucoside amygdalin. (Ada Hayden).

Many plants of the family are cultivated for ornamental purposes and some are economic. In the first class is the laurel cherry (Prunus Lauro-cerasus), a handsome fragrant shrub of the Caucasus to northern Persia, cultivated in the southern states and very common in the Mediterranean regions of Europe. The Mayday tree of Europe (Prunus Padus), as well as numerous species of the genus Spiraea, like Spiraea Douglasii, S. salicifolia, S. japonica, S. Thunbergii; the nine-bark (Physocarpus opulifolius), species of the genus Rosa, such as the prairie rose (Rosa setigera), sweetbrier (Rosa rubiginosa), dog rose (Rosa canina), R. rugosa, R. gallica, and the cinnamon rose (R. cinnamonea) are frequently cultivated. Kerria japonica, Rubus odoratus, Pyrus coronaria, P. japonica, P. Ancuparia, P. americana, Crataegus mollis and C. punctata are also cultivated.

The family contains a large number of valuable fruits; of these we may mention the service berry (Amelanchier canadensis and A. spicata), the apple (Pyrus Malus), the pear (Pyrus communis), the quince (P. Cydonia), straw-

berry (Fragaria vesca, F. virginiana, var. Illinoensis), the F. chiloensis, native to Chili and the Pacific coast (the common garden strawberry is a modified form of the Chilian strawberry), and the Indian strawberry (Duchesnea indica). We may also mention the wild northern plum (Prunus americana), the Chickasaw plum (P. angustifolia), European garden plum (P. domestica), sand cherry (P. pumila and P. Besseyi); the cherries, English cherry (P. avium), naturalized in the southern states, especially in Virginia and Maryland, the sour cherry (P. Cerasus), also naturalized in the East and extensively cultivated, the wild red cherry (P. pennsylvanica), commonly used in the north, the Japanese plum (P. triflora), the apricot (P. armeniaca), peach (P. persica), almond (P. Amygdalus) flowering almond (P. nana), wild red raspberry (Rubus idaeus var. aculeatissimus), black raspberry (R. occidentalis), garden raspberry of Europe (R. Idaeus), salmon berry (R. parviflorus), dewberry (R. cuneifolius) and the wineberry of Japan (R. phoenicolasius). The fruit of the Icaco plum (Chrysobolanus Icaco) of tropical America is edible. The wood of the wild black cherry (P. serotina) takes an excellent finish, and therefore is highly desirable for cabinet making and for interior finishing of houses. The wood of other species of the genus is used in the manufacture of pipes and furniture. Most of the plants of the order contain no injurious substances. Malic acid C₄H₈O₅ occurs in the fruit of the cherry (Prunus Cerasus), plum (P. domestica), the apple (Pyrus Malus), the strawberry (Fragaria virginiana), (F. vesca), etc. Salicylic acid C7H6O3 occurs in the fruit of the strawberry, citric acid in Rubus, the strawberry and Prunus domestica. The Quillaja Saponaria contains saponin, the bark yielding 2 per cent. Kobert distinguishes two substances quillajic acid C19H30O10 and sapotoxin C17H26O10.

The glucoside amygdalin was first obtained in 1830 by Robiquet and Boutron from the seeds of the bitter almond. Liebig and Woehler named the substance which converts the amygdalin into the so-called essence of bitter almonds, emulsin. They found that through the action of emulsin, sugar and prussic acid were formed. The name synaptase was given to emulsin.

$$C_{20}H_{27}NO_{11} + 2H_{2}O = C_{7}H_{6}O + HCN + 2C_{6}H_{12}O_{6}$$

Amygdalin Benzoic Prussic Glucose aldehyde acid

Emulsin can also convert salicin, helicin, phlorizin, and arbutin. The change in arbutin is as follows:

In the cherry leaves emulsin occurs in the leaves and younger branches. Emulsin also occurs in Penicillium glaucum and Aspergillus niger.

The Kooso (Brayera anthelmintica) is a large dioecious ornamental tree from Abyssinia. The drug comes from the pistillate flowers which have a tealike odor but a bitter, nauseous taste and contain cuscotoxin which is a muscle poison, protocosin and cosin which is bitter and acrid. In medicine it is used as a taeniafuge but in large doses produces vomiting and colic. Agrimonia gryposepala, Gillenia stipulacea, and Geum urbanum are used as astringents. The roots of the water avens (Geum rivale) are tonic and powerfully astringent. The soap-bark (Quillaja Saponaria) of Peru and Chili is used as an expectorant and is an irritant poison. According to Schneider who investigated a great many of the saponins which occur injuriously in about fifty families, they act

poisonously by dissolving the blood corpuscles. Cholesterin contained in the body acts as a natural antidote against them.

Several plants of the order produce members of the terpene group; rose oil, contains rhodinol $C_{10}H_{18}O$, supposed to be identical with geranium oil; and a second terpene, roseol, $C_{10}H_{10}O_7$. Many fruits of the order, especially Pomeae, contain mannite and sorbite. The arbutin, $C_{12}H_{10}O_7$ obtained in many plants also occurs in some plants of this order; the glucoside hydrochinon, $C_7H_{12}O$, occurs in the buds of pears. Quercetrin $C_{15}H_{10}O_7$, derived from a glucoside, is found in the flowers of haw (Crataegus), the bark of apple trees and of Prunus instititia. Amygdalin $C_{20}H_{27}NO_{11}$, occurs in the seeds of many plants of the family, especially in Pomeae and Pruneae; also in the bark of Prunus Padus, P. serotina, etc., and in the seeds of Pyrus Aucuparia.



Fig. 266. Kooso (Brayera anthelmintica). Flowering branch. Contains a muscle poison. (After Faguet). Fig. 266a. Common Wild Plum (Prunus americana). (C. M. King).

Genera of Rosaceae

| Ovary inferior or enclosed in the calyx tube. | |
|---|---------|
| Carpels numerous; fruit an achene | Rosa. |
| Carpels few, fruit not an achene. | |
| Carpels cartilaginous; fruit a pome4. | Pyrus. |
| Carpels bony, drupe-like6. Cra | taegus. |
| Ovary superior not enclosed in calyx tube. | |
| Calyx deciduous; fruit a drupe | Prunus. |

| ~ | | | |
|----|------|----------|-------|
| | 2035 | DATE LET | ant. |
| Ca | IVA. | persist | CHIL. |
| | | | |

| Pistils numerous; | fruit | drupelets1. | Rubus. |
|-------------------|-------|-------------|----------|
| Pistils numerous; | fruit | an achene | ragaria. |

1. Rubus. Raspberries and Blackberries

Perennial herbs, shrubs or vines; vine prickly, with alternate leaves, 3-7 foliolate or simple; flowers terminal; axillary or solitary, white reddish or pink, usually perfect; calyx 5-parted, petals 5, deciduous; stamens numerous; achenes usually many, inserted on the receptacle, which is either fleshy or dry; carpels forming drupelets. About 200 species chiefly Northern.

Rubus Idaeus L. var. aculeatissimus (C. A. Mey.) Regel & Tiling. Wild Red Raspberry

Stems biennial, upright shrubs covered with straight, stiff bristles, some hooked, and glandular hairs; leaflets 3-5, oblong, ovate, pointed, whitish, downy underneath; petals as long as the sepals, whitish; fruit light red. Spreads by suckers.

Distribution. The species is native to the Appalachian Mountains as far south as the Carolinas; common at high altitudes in the Rockies. Frequently



Fig 267. Wild Red Raspberry (Rubus Idaeus, varaculeatissimus). The prickles of the red raspberry produce mechanical injury. (Ada Hayden).

Fig. 268. Wild black cap raspberry (Rubus occidentalis). This plant has thorns which are irritating. (Ada Hayden).

troublesome in fields for several years in the north and persists for a long time in gardens. A common native of the north.

Rubus occidentalis L. Black Raspberry, or Black-cap Raspberry

Stems biennial, glaucus, recurved, beset with hooked prickles; rooting at the tip; leaves pinnately 3-foliolate, or rarely 5-foliolate; leaflets ovate, coarsely doubly serrate, whitish underneath; flowers corymbose clusters; petals shorter than the sepals; fruit usually purplish-black, occasionally white.

Distribution. Quebec to Georgia, to Missouri, to Minn. Like the preceding species, often troublesome in fields and gardens.

Rubus villosus Ait. High Bush Blackberry

Shrubs 1-6 feet high, upright or reclining, armed with stout recurved prickles, branchlets and lower surface of leaves glandular; leaflets 3-5, ovate, pointed, terminal one stalked; flowers in corymbose clusters; petals white; fruit not separating from the juicy receptacle, blackish.

Distribution. From Nova Scotia to Georgia, Missouri, Kansas, to Minn. Troublesome like the black-cap Raspberry.

Injurious properties. No species of the genus Rubus is known to be poisonous, but the bristles and spines on various species frequently inflict injuries Numerous cases of inflammation, and later pus formation, are reported from the prickles found on the common red raspberry. This is especially true of the larger prickle of the black raspberry and the dewberry. The bark of the blackberry contains villosin. The leaves are said to cause an irritation of the skin of berry pickers or others who walk among the bushes.



Fig. 269. Common Blackberry (Rubus villosus). The common blackberry produces prickles which act injuriously in a mechanical way. (Ada Hayden).

2. Fragaria L. Strawberry

Perennial stemless herbs; leaves petioled, stipulate, 3-foliolate; flowers white, in clusters, polygamo-dioecious; calyx persistent, with 5 bractlets, deeply 5-lobed; petals 5; stamens numerous; carpels or pistils numerous; the receptacle elongated, which become fleshy in fruit. A small genus of about 25 species containing the common cultivated strawberry (F. chiloensis) of the Pacific coast, our wild strawberry (F. virginiana), the European strawberry (F. vesca) and the Indian strawberry (F. Indica or Duchesnea indica).

Fragaria vesca L. European Wood Strawberry

A perennial with ovate leaves, dentate, thin; flowers white, racemose receptacle, elongate, fruit with achenes, seeds free above the receptacle. It is very different from our common wild strawberry, in which the achenes are sunken in the flesh.

Poisonous properties. Few people would suspect that the seeds of the strawberry are injurious, but I have known people who have found that the eating of the common cultivated strawberry is injurious, and it is known that the European strawberry (F. vesca) produces a rash that sometimes resembles that produced by scarlatina. Dr. Millspaugh, in speaking of the European strawberry, says of a lady coming under his care who had consumed the fruit of the strawberry grown in Florida:

In the afternoon of the same day the skin was hot and swollen, the patient thirsty and restless, and little sleep was gained that night; the next day the eruption began to fade, the appetite returned, and restlessness ceased. On the third day exfoliation began and was very profuse, the skin appearing quite similar to the condition existing after a severe attack of scarlatina.

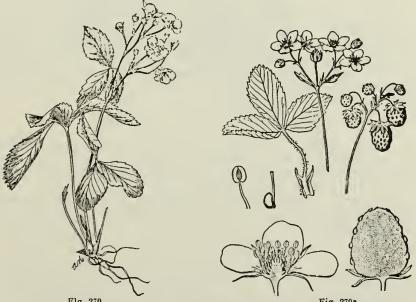


Fig. 270. Wild Strawberry (Fragaria virginiana). (Ada Hayden). Fig. 270a. Wild Strawberry (Fragaria vesca). (Willis).

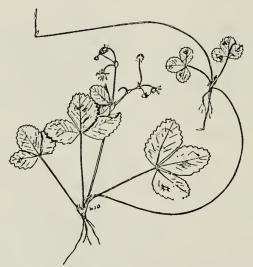


Fig. 271. European Wood Strawberry (Fragaria vesca). This plant occurs in fields and along road-sides. Sometimes causes dermatitis. (W. S. Dudgeon).

Prof. Prentiss * reports the case of a man who, at the age of 14, had become quite ill from eating strawberries and forever afterward could not eat them without becoming ill.

3. Rosa L.

Erect or climbing shrubs, with prickly stems, alternate leaves adnate to the petioles; flowers showy, corymbose, or solitary; calyx urn-shaped; stamens and carpels numerous; achenes, enclosed in a berry-like calyx tube. Several species of the genus Rosa are more or less troublesome in fields. The Rosa centifolia, used for preparing rose water contains a volatile oil. A confection is made from the hips of Rosa canina. R. gallica contains a volatile oil and a yellow crystalline glucoside quercitin.

Rosa pratincola Greene. Prairie Rosc

An erect perennial shrub with densely prickly stems bearing slender bristles; narrow stipules, more or less glandular, toothed; leaflets 7-11, broadly elliptical to oblong-lanceolate, sessile or nearly so; flowers corymbose or rarely solitary, pink; sepals lanceolate, somewhat glabrous; fruit smooth.

Distribution. Common on prairies of Wisconsin, Iowa to Texas, New Mexico and Montana. In Iowa, Missouri and parts of Minnesota and Arkansas, it is most troublesome.

Rosa blanda Ait. Smooth Rose

An erect shrub with few straight prickles or wholly unarmed; from 1-3 feet high; leaflets 5-7, short stalked; oblong-lanceolate; cuneate; stipules dilated, naked or slightly glandular-toothed; flowers usually large, corymbose or solitary.

Distribution. From Newfoundland to Ontario and Illinois.

^{*} Bot. Gazette 13:19.

Rosa Sayi. Schw. Say's Rose

A very prickly shrub with low stem, 1-2 feet high; leaflets 3-7 broadly elliptical or oblong, lanceolate, glandular, ciliate and resinous; stipules dilated; flowers large, solitary or rarely more; outer sepals usually with 1 or 2 narrow lateral lobes.

Distribution. From northern Michigan, Northwest Territory to Colorado.



Fig. | 272. Arkansas Rose (Rosa pratincola). A native rose. (Ada Hayden).

Rosa Woodsii Lindl. Wood's Rose

Stems usually low; 6 inches to 3 feet high with slender, straight or recurved spines and scattered prickles, or unarmed above; leaflets 5-7 obovate to oblong or lanceolate, more or less toothed; flowers corymbose or solitary; scpals naked or hispid; fruit globose.

Distribution. Prairies of Minnesota and from Montana to New Mexico and Texas.

Mechanical injuries. The prickles inflict injury to cattle, and are especially troublesome in grain fields. Before the binder came into use men were

injured by the prickles and spines of the rose which produced inflammation and caused the formation of pus.

4. Pyrus L.

Trees or shrubs; simple leaves; flowers in cymose clusters, white or pink; calyx urn-shaped, 5-lobed; petals 5, short-clawed; stamens usually numerous; styles mostly 5, distinct or united at the base; ovules 2 in each cavity; carpels leathery; fruit a pome. About 37 species of wide distribution, chiefly in the North Temperate region. The following species of the genus are cultivated for their fruits; quince (P. Cydonia), pear (P. communis), Japan or sand pear (P. sinensis), apple (P. Malus), Old World crab apple (P. baccata), mountain ash (P. americana), (P. sitchénsis P. sambucifolia), European mountain ash (P. Aucuparia).

The fresh bark of the wild mountain ash is used in medicine; it is known to produce irritation of the alimentary mucous membranes, and a reflex nervous irritation.

Pyrus coronaria L. Wild Crab Apple

A small tree with petioled or ovate to triangular-ovate leaves, sparingly pubescent beneath; sharply serrate and often lobed; flowers rose-colored, fragrant; calyx slightly pubescent; pome fleshy, fragrant, greenish-yellow, acid. Two other species are found in eastern North America, namely, *P. angustifolia*, with small leaves and few flowers, and *P. ioensis*, with firm leaves, narrowed at the base, and pubescent calyx, chiefly west in the Mississippi Valley, *P. rivularis* Dougl., occurs from California to Alaska.

Distribution. Our Eastern wild crab is found from Ontario to Michigan and South Carolina; in the west it is replaced by the *P. ioensis*.

Poisonous and medicinal properties. All the species of the genus Pyrus contain the glucoside amygdalin, $C_{20}H_{27}NO_{11}$, which is converted by the action of the ferment into hydrocyanic acid. The bark also contains citric acid, $C_0H_8O_7$, and malic acid, $C_4H_0O_5$, both of which appear in the fruit of the apples. There may be occasionally cases of poisoning where animals are allowed to browse upon the wilting leaves of the apple.

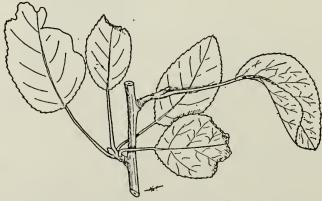


Fig. 273. Common apple (Pyrus Malus). The well known cultivated apple. (W. S. Dudgeon).



Fig. 274. Iowa Crab (Pyrns ioensis). 1. Flowering brancb. 2. Longitudinal section of flower with petals removed, natural size. 3. Fruiting branch. 4. Longitudinal section of fruit. 5. Summer branch. 1, 3, 4, 5, one-half natural size. (M. M. Cheney).

Crataegus L. Hawthorn. White Thorn

Shrubs or small trees, usually spiny; leaves petioled; flowers in corymbose clusters, white or pink; calyx-tubc urn-shaped; limb 5-cleft; petals 5, roundish; stamens numerous or few; styles 1-5; fruit a pome, containing 1-5 bony, 1-seeded stones. About 75 species, although the number is sometimes estimated as high as 125. Some species, like C. punctata, and C. mollis, are ornamental.

Cratacgus mollis Scheele

Shrub or small tree; shoots densely pubescent; leaves large, slender-petioled, cuncate, truncate or cordate at base, usually with acute lobes; more or less densely pubescent beneath; flowers large, 1 inch across; fruit bright scarlet with a light bloom.

Distribution. Common in thickets from Eastern Canada to Iowa and Kansas, and Texas.

Poisonous properties. Large numbers of haw fruits are eaten, and several deaths due to the eating of Crataegus have been reported in Iowa. These were probably largely due to strangulation or indigestibility of the stony "sced." The flesh is said to be indigestible as well.



Fig. 275. Common Red Haw (Crataegus mollis). (C. M. King)

5. Prunus L. Plum and Cherry.

Shrubs or trees with alternate petioled leaves and small stipules; flowers variously clustered, mostly perfect; calyx inferior, free from the ovary, with a bell-shaped or urn-shaped tube and 5 spreading lobes; falling after flowering; petals white or pink; spreading stamens 15-20 or more, distinct, inserted on the throat of the calyx, perigynous; pistil solitary; style simple; stigma capitate; ovary 1-celled, 2-ovuled; fruit a drupe; seed usually one; embryo large, cotyledons fleshy, endosperm absent. Species about 90, of the north temperate regions, tropical America and Asia. The sweet cherry (Prunus avium), sour cherry (P. Cerasus), native plum (P. americana), Chicksaw plum (P. Chicasa), European plum (P. domestica), Japan plum (P. triflora), the flowering almond (P. triloba), peach (P. persica), and apricot (P. armeniaca) are all well known in cultivation.

Prunus virginiana L. Choke Cherry

A tall shrub or small tree, bark gray; leaves thin, oval, oblong or obovate, acuminate at the apex, smooth or slightly pubescent, sharply screate, teeth large; often doubly serrate; flowers white in rather loose racemes, terminating leafy branches; petals roundish, fruit red, turning dark or crimson. Astringent.

Distribution. Forming thickets from New Foundland to Manitoba to Texas and Georgia.





Wild Cherry ($Prunus\ demissa$). Common shrub of the Rocky Mountains. (Photo by Colburn).

Prunus demissa Walp. Western Wild Cherry or Choke Cherry.

A shrub or small tree; leaves thick and oval or obovate, acute or more or less obtuse at the apex; teeth rather short; flowers white in dense racemes, terminating leafy branches; fruit dark or purplish black, less astringent than the preceding.

Distribution. Dry soil, common in thickets and woods from Dakota to Kansas, New Mexico to California and British Columbia.

Prunus serotina Ehrh. Wild Black Cherry

Large tree with reddish brown branches, reddish wood; leaves thick, oblong, or lanceolate-oblong, taper pointed, serrate, with short teeth shining above; flowers in elongated spreading or drooping racemes; petals obovate; fruit purplish black, and slightly astringent.



Fig. 276. Black cherry (Prunus serotina). 1. Flowering branch. 2. Longitudinal section of flower, enlarged. 3. Fruiting branch. 4. Cross section of fruit. 5. Longitudinal section of fruit. 6. Winter branchlet. 1, 3, 6, one-half natural size. 4, 5, natural size. (M. M. Cheney in Green's Forestry in Minnesota.)

Distribution. From New England to Ontario, to Florida and Texas, Kansas, Dakota and Minnesota.

Prunus pennsylvanica L. Wild Red Cherry

A small tree 20-30 feet high, light brown bark; leaves oval or lanceolate, acute or acuminate, finely and sharply serrate, glabrous, with slender petiole; flowers white in corymbose clusters; fruit small, globose, light red hue and sour.

Distribution. In rocky woods. New Foundland to the Rocky Mountains to Georgia.

Poisonous properties. Many cases of poisoning have been recorded from eating the seeds of peach and bitter almonds. They contain a highly poisonous



Fig. 277. Wild Red Cherry (Prunus pennsylvanica). 1. Flowering branch. 2. Longitudinal section of flower. 3. Fruiting branch. 4. Longitudinal section of fruit, slightly enlarged. 5. Cross section of fruit. 6. Embryo enlarged. 7. Axil of leaf, showing stipules. 8. Winter branchlet. 1, 3, 7, 8, one-half natural size. (M. M. Cheney.)

substance from which prussic acid is obtained. Mr. Chesnut says in regard to the black cherry:

The fruit is rather agreeable, being but slightly hitter and astringent in taste. In some localities it is much used to flavor liquor. Poisoning is frequently caused by cattle eating the wilted leaves from hranches thrown carelessly within their reach or ignorantly offered as food. Children occasionally die from eating the kernels of the seed or from swallowing the fruit whole.

Prof. Chas. D. Howard, of the New Hampshire Station, says:

The poisonous property of all species of cherry leaves is due to hydrocyanic acid, popularly known as prussic acid. This compound does not exist as such in the growing leaf, but is derived from a class of substances called glucosides, of which amygdalin is the type peculiar to the cherry. This, or a closely allied hody, is to be found not only in the leaves and bark, but especially in the stones of cherries, peaches and plums, and the seeds of the apple. By the action of moisture and a vegetable ferment called emulsin, which exists in the plant, a complex chemical reaction takes place, that begins in the leaf the moment connection with the circulatory system is cut off. The three products of this reaction are hydrocyanic acid, grape sugar and benzaldehyde, or bitter almond oil.

There is a popular opinion that the leaves of the cherry are poisonous only when cut and in the wilted condition; that cattle may safely nibble them from the growing shrub without danger of injury; and that they are quite harmless when dried. Our observations, however, prove these views to be but partially correct. As a matter of fact, distillations of samples, made within twenty minutes of cutting, show that the freshly cut leaves yield nearly as much acid as the wilted ones do when calculated on the weight of fresh material taken, and when caten fresh, the character of the juices within the animal stomach is such as to render that organ a most favorable place for the conduct of the reaction in which prussic acid is liberated.

On the other hand, while it is true that the thoroughly dried leaves yield a comparatively small amount of acid, still they may always produce some, and as ordinarily dried in the hayfield, they may be capable of generating a considerable quantity of the poison. The desirability of carefully excluding them from all hay is therefore apparent.

One hundred grams of bitter almonds (Prunus amygdalus communis), in the form of pulp, yield 250 milligrams prussic acid; the same amount of kernels from cherry stones yields 170 mgs.; leaves of the cherry laurel (Prunus Laurocerasus), occurring in Europe and Mexico, 39 mgs.; kernels of peach, 164 mgs.; apple seeds, 35 mgs. The stones of all these species must therefore he regarded as dangerous; the fruit, in every case, so far as can he ascertained, is harmless.

The leaves of the wild hlack cherry are the most poisonous of the three species investigated, though all are dangerous.

Both the wilted leaves and fresh leaves are poisonous, while the dried are to he regarded with suspicion.

Vigorous, succulent leaves from young shoots, which are the ones most liable to be eaten by cattle, are far more poisonous than the leaves from a mature tree or stunted shrub.

Leaves wilted in bright sunlight to ahout 75 per cent original weight, or until they hegin to appear slightly limp and to lose their gloss, yield the maximum amount of prussic acid.

I have seen cattle browse on the leaves of *Prunus demissa* in the Rocky Mountains. It is believed by stockmen to be poisonous. Probably the danger is not so great because the leaves are eaten direct from the tree and not wilted. The Bureau of Forestry has shown that much loss occurs which can easily be prevented by changing the trail.

It has been known for a long time that seeds of the various members of the genus Prunus contain poisonous properties. The bark of several of our wild cherries is also known to contain a poisonous principle, a ferment known as *emulsin*, which in the presence of water acts on the glucoside *amygdalin* $C_{20}H_{27}NO_{11}$, and produces *hydrocyanic acid*, a powerful poison; it is a clear, colorless liquid of a characteristic taste and odor, resembling that of a bitter

almond. Externally, hydrocyanic acid produces a paralytic effect. When taken internally, it acts as a sedative upon the mucous membrane. Prof. Winslow gives the following characteristics of poisoning:

In poisoning, the blood becomes first a bright arterial hue, and later assumes a dark, venous color. The first condition is due to the fact that the blood does not give up its oxygen for some reason. Brunton suggests that it is because the blood is hurried so rapidly through the dilated peripheral vessels that it does not have time to yield up its oxygen. The dark color of the blood is probably owing to asphyxia and accumulation of carbonic dioxide, following the paralytic action of prussic acid upon the respiratory centre.

Prussic acid has an essentially depressing action upon the nervous system as a whole. The brain, cord and nerves become paralyzed by large doses.

The spinal cord is paralyzed at a period after coma and convulsions have appeared. The peripheral nerves and muscles are paralyzed directly by toxic doses, and not through the mediation of the central nervous apparatus.

Inhalation of the pure acid will cause death in a confined atmosphere, and even inhalation of the medicinal solution will induce the physiological symptoms of the drug.

He also says of the toxicology of prussic acid:

Prussic acid is one of the most powerful poisons in existence. Death may be instantaneous, or life may be prolonged for over an hour after lethal dose. More commonly the animal survives for a few minutes, and we observe the following symptoms in dogs: The animal falls, froths at the mouth, the respiration is of a gasping character and occurs at infrequent intervals. There is unconsciousness, the pupils become dilated, there are muscular tremblings, and clonic or tonic spasms. Defecation and micturition occur, and erections often ensue in the male. Respiration ceases before the cardiac pulsations.

Three stages may be distinguished in fatal poisoning. First: a very short period elapses before the symptoms appear. There are giddiness, difficult breathing, and slow pulse in this stage. Second: the pupils dilate, vomiting may occur, and the animal utters loud cries. Spasmodic defecation, micturition and crections may be present, with convulsions and unconsciousness. Third: the last stage is characterized by collapse, spasms, general paralysis and death. The subacute form of poisoning may ensue and prove fatal, or, owing to the volatile character of the drug, complete recovery may take place within one-half or three-quarters of an hour. Occasionally dogs continue to be paralyzed for several days and get well. The minimum fatal dose recorded in man is 9/10 of a grain of pure acid, or about 50 drops of the medicinal solution. Four to five drachms of the diluted acid frequently, but not invariably, cause subacute poisoning and death, in horses, within an hour. One to two drachms of the pharmacopoeial preparation usually kill dogs within ten minutes.

Prussic acid is commonly used to destroy the domestic animals. Two to four drachms of the medicinal acid are to be given to dogs and cats of the ordinary size, and certain, painless, and rapid death will occur if a fresh preparation of the drug can be obtained. The unopened, half-ounce vial, kept by druggists, is recommended. Big dogs, horses, and the other larger animals are not killed rapidly, nor sometimes at all, by great quantities of the diluted acid. Hence, shooting is a more humane and preferable mode of death for them. In the experience of the writer, one to two dracbms of prussic acid saturated with potassium cyanide, failed to kill a horse, when injected directly into the jugular vein. The odor of the acid lingers about the animal for a few hours after death; the eyes are fixed and staring; the pupils dilated; the teeth are clinched tight and covered with froth, while the blood is of a very dark color. The treatment embraces emptying the stomach by large doses of promptly acting emetics, or by the stomach tube, or pump; atropin, ether, and brandy subcutaneously, and inhalations of ammonia, together with artificial respiration, and hot and cold douches upon the chest.

Hydrocyanic acid is produced by a number of other plants referred to in another connection.

Wild cherry bark (*Prunus serotina*) and leaves by distillation yield a volatile oil resembling that of bitter almond. The same is true of the *P. virginiana*. Hydrocyanic acid is formed only by the action of a ferment upon amygdalin which is present in all plants of this sub-order. Fresh leaves are generally considered harmless, but Chesnut says that cattle are frequently poisoned from

eating the wilted leaves. He also adds that the seeds of all varieties of cherries and plums, both native and introduced are subject to suspicion. The flesh of none of the species is in any way poisonous. Chesnut also says in another contribution, that no cases are on record where stock have been poisoned by eating the leaves of any species, while still on the tree. It is only after they have been cut off and are partially wilted that they are considered dangerous. The reason for this is that during the process of wilting, prussic acid is formed from non-poisonous constituents which are always present in the leaf and bark. The wilted leaves have the characteristic odor of prussic acid. The amount of amygdalin contained varies from 3.6 to 4.12 per cent and yields from 0.23-0.32 hydrocyanic acid. Recent studies indicate that an amydonitril glucoside occurs in young leaves of both Prunus Padus and P. serotina.



Fig. 278. The European May Day tree (Prunus Padus), poisonous. This is frequently cultivated. (Ada Hayden)

LEGUMINOSAE

Herbs, shrubs, trees, or vines with alternate, mostly compound, stipulate leaves, papilionaceous or sometimes regular flowers; calyx 3-6, or 4-5 cleft; stamens 10, rarely 5 and sometimes many, monadelphous, diadelphous or distinct; pistil, simple, free, becoming a legume in fruit or sometimes a loment; ovules 1-many; seeds generally ex-albuminous or nearly so or, in some, with copious albumen. About 6500 species of wide distribution, but most abundant in the tropics.

Economic plants

The order contains a large number of economic plants, especially food plants. The common bean, (Phaseolus vulgaris), was originally found in the southwestern United States, but is now cultivated in all civilized countries. Common string and golden wax beans are types of the last named. The scarlet runner, (P. multiflorus), generally cultivated for ornamental purposes is also used as food, although the ripe beans are unwholesome and sometimes poisonous. The three-lobed kidney bean, (P. trilobus), is commonly cultivated in India. The lima bean, (P. lunatus), also native to America, supposedly Brazil, is not known in a wild state. The seed of the Adzuki bean, (P. Mungo, var. glaber), is used as food in Japan. The soy bean, (Glycine Soja and G. hispida), of which there are many varieties, is native to China and Japan and is used in large quantities by the Japanese and Chinese for food, but is little used in the United States, being here cultivated as a forage plant.

Soy beans can only be fed in moderate amounts to cattle because of their purgative properties. A loss of a considerable number of cattle occured in England recently where soy bean cake had been used. When fed mixed no trouble was caused, but when fed alone it caused poisoning.

The cow-pea, (Vigna Catjang), native to China, has been cultivated for centuries by the Chinese and extensively used for food. It is also used in many other warm countries, especially the southern states, not only for human food, but also as one of the best forage plants, for which purpose it is now

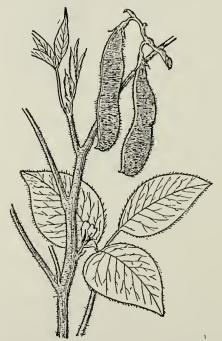


Fig. 279. Soja Bean (Glycine hispida), used both as a food and as a forage plant. (U. S. Dept. Agr.)

cultivated as far north as Minnesota; it is also a soil renovator. The Dolichos Lablab is used for food in the tropics. The yam bean, (Pachyrrhizus angulatus), in some countries, forms a resource as food in case of the failure of the usual crops. The garden pea, (Pisum sativum), probably originated in western Asia along the foothills of the Caucasus. It has, however, long been cultivated in Europe. Some authorities believe that it may have originated from the field pea (Pisum arvense). The pea is extensively cultivated in Europe and Canada as a forage plant. The chick pea, (Cicer arietinum), is a native to Caucasus and the Caspian Sea region and has been cultivated since



Fig. 280. Garden pea (Pisum sativum), a valuable food plant. (U. S. Dept. Agr.)

remote times in northern Africa and other Mediteranean countries. In the United States, it is cultivated chiefly in the arid regions both for stock and for human food. The lentil (Lens esculenta) has been cultivated in the Mediterranean region for centuries but its original home is not known. It is grown and used now from Central Europe south and east to India, as food for both men and stock. The peanut (Arachis hypogaea), probably native to Brazil, was cultivated by the ancient Peruvians but is now widely scattered in all warm countries. The nut (seed) is used as food and a fine oil is extracted from it. A plant allied to the peanut (Voandesia subterranea) is used as an article of food in western and southern Africa. The pigeon pea (Cajanus indicus) is an important article of food in the tropics, especially in India, and is also used as a fodder plant. The sword bean (Canavalia ensiforme) and the Jackpea (C. obtusifolia), cultivated in the tropics are used as food, the skin having been

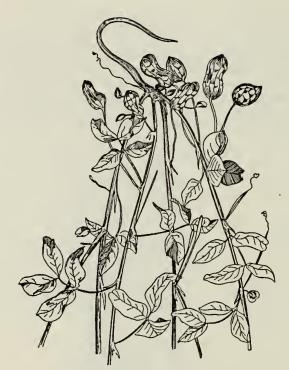


Fig. 281. Peanut (Arachis hypogaca).



Fig. 282. Broad Bean (Vicia Faba). Cultivated both as a food and as a forage plant. (From the American Agriculturist.)

first removed. The broad bean (Vicia Faba) is much cultivated in Europe for both animal and human food. The seeds of Paprika ufricana are used as food by native Africans and in Abyssinia and in the Indian Archipelago are esteemed as a good substitute for coffee.

The Australian wattles (Acacia), of which there are many species, vary greatly in size. The bark of some of these is used for tanning purposes. The wood is valuable and takes a fine polish, A. Gerrurdi being an example of this class. From A. Farnesiana is derived the oil of cassia, much used in perfumery. It is prepared by macerating the flowers in olive oil. Cassia pomade is prepared from fatty substances to which the cassia flowers have been made to impart their perfume. C. occidentalis is used as a substitute for coffee. Many of the legumes produce important gums. From the Algarrobe, or locust tree of Jamaica (Hymenaea Courbaril) is produced a gum said to be superior to shellac; the sweet pulp of the fruit is edible. The gum Kino (Pterocarpus Marsupium) is a native of India and yields a gum that is used both for tanning and dying and as an astringent. Kino contains from 40-80 percent of tannin and kino red. P. tinctorius produces a valuable wood, and a related species (P. Dalbergioides) produces a wood similar to mahogany. The Tonka bean or Tonquin (Dipteryx odorata) of Guinea contains the substance cumarin and is used as a snuff and as a scent in cigars. Cumarin is widely distributed in the plant kingdom, especially in such Leguminosae as Dipteryx, Melilotus, and Myroxylon. It occurs in species of other families, as the palm, vernal

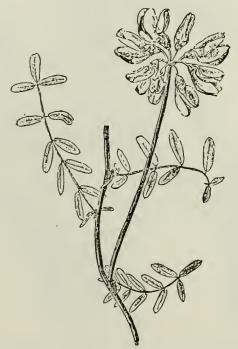


Fig. 283. Axwort (Coronilla varia). A poisonous plant of Europe—adventitions in the U. S. (From Strasburger, Noll, Schenck and Schimper.)

grass, madder, rue, and orchids, in such composites as Trilisa, in the rootstock of Vitis sessilifolia and in Prunus Mahaleb. It is most easily detected when the plant begins to wilt or after it is dead. Pseudocumarin $C_7H_4O_{2n}$ is like the odorous substance found in Coronilla scorpioides. The seeds of Mucuna giganteum and of M. pruriens are used for various purposes as watch charms, or as other ornaments.

Some species of the family which contain a good fiber are applied in weaving cloth. A species of Crotalaria, C. juncea is cultivated in India for its tough fiber, and is used for making ropes and bags. Sesbania aculeata, a branched annual, is also cultivated for its fiber. The stems of Shola (Aeschynomene aspera) native to India are used for making hats. Blue indigo dyes come from Indigofera Anil of the West Indies. The Genista tinctoria or Dyers' Broom



Fig. 284. Hairy Vetch (Vicia villosa). Cultivated as a forage plant. (U. S. Dept. Agr.)

of Europe and Asia, and naturalized in the U. S. contains a yellow coloring principle. The seeds of Entada scandens are used in the Samoan Islands in playing games. The Pithecolobium dulce contains a pulpy pod which is catcn. The plant is good fodder. Guava (Inga vera) is grown as a shade tree and as a substitute for coffee. It is not to be confused with the fruit producing guava (Psidium guajava). Important forage plants not previously mentioned are red clover (Trifolium prateuse), native of Europe and used extensively in northern United States; alsike clover (T. hybridum), white clover (T. repens), well known as a forage plant and a good honey plant, T. alexandrinum the great forage crop of Egypt, known as the Berescm, Japan clover (Lespedeza striata) a well known forage plant of the South, French honeysuckle (Hedysarum coronarium) an ornamental plant, native to Spain, also used as a forage plant of Europe and Western Asia, Desmodium triflorum used as a forage plant in the tropics, Florida beggar weed (Desmodium tortuosum) of India, alfalfa (Medicago sativa) of Europe and Western Asia, the vetches (Vicia villosa and V. sativa), lupines (Lupinus albus) cultivated for forage purposes, besides many valuable native forage plants like the Hosackia Purshiana, the wild pea (Lathyrus venosus) etc.

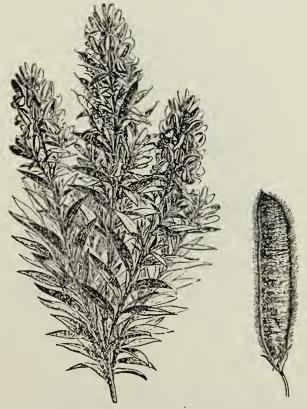


Fig. 285. Dyer's Broom (Genista tinctoria). Contains a yellow coloring matter. (After Faguet.)

Prof. N. E. Hansen through his exploration has brought into prominence the yellow-flowered alfalfas from Siberia, Medicago ruthenica and M. platycarpa, and some of the hardy alfalfas are apparently hybrids between M. sativa M. falcata. C. V. Piper * calls attention to a number of valuable leguminous forage plants that should be cultivated in this country, such as the Lyon bean (Stizolobium Lyoni). The Kudzu (Pueraria Thunbergiana) a woody native of Japan, is much used as a forage plant in that country and has been cultivated in Florida. The Guar (Cyamopsis tetragomoloba) is an East India annual legume and is said to be very drouth resistant. Tangier pea (Lathyrus tingitanus) is a native of Northern Africa. It is excellent and said not to be poisonous like the other species. The moth bean (Phaseolus aconitiis used for food in India folius) and is said to be a splendid The Adzuki bean (Phaseolus angularis) forage plant. native of southern Asia is used for food in China, Japan and India, and is said to possess vaulable qualities as a hay plant. The Kulti (Dolichos biflorus) native to India is said to give promise in the semi-arid regions in Texas as a valuable forage plant. Under the Vetches the more recent introductions that give promise are the scarlet vetch (Vicia fulgens) of Northern Africa, the black purple vetch (V. atropurpurca) of Algeria and the woolly pod



Fig. 286. Carobtree, or St. John's Bread (Ceratonia siliqua). The fruit is edible and is supposed to be the "locust" of Biblical history. (After Faguet.)

^{*} Yearbook U. S. Dept. of Agr. 1908-245.

vetch (V. dasycarpa) of the Mediterranean region, the Carob tree or St. John's bread (Ceratonia siliqua) is a small tree of the Mediterranean region, the pods of which contain much mueilage of a sweet nature from which syrup was made, and is supposed to be the locust which John the Baptist lived upon in the Wilderness and is used as food; and cattle also relish it. Manna (Alhagi maurorum) is a dwarfed, thorny, shrubby plant which produces a kind of manna. The locust trees of the West Indies, or Courabaril (Hymenaea Courbaril) produces a hard timber. The pods contain bean-like seeds embedded in a white spongy mass. The Zamang (Pithecolobium Saman) is a large tree of Venezuela which produces thick, flat pods, containing a sweet pulp commonly used by eattle and horses for food but which are liable to cause internal disorder. The honey locust of eastern North America (Gleditschia triacanthos) produces a hard wood. The pod contains a gummy sweetish substance much relished by stock. The pods of the mesquite tree (Prosopis juliflora), native to Texas, are used by stock. The Kentucky coffee tree (Glymnocladus dioica) is native to the Mississippi Valley and it produces a broad, tough god which contains large, hard seeds. The pod contains a sweetish, disagreeable and nauscating material more or less poisonous. The hard wood is durable. The seed of hairy vetch (Vicia hirsuta) is a common impurity in grain seed. The plant is used for forage. The seeds of Castanospermum australis are used in New South Wales in the production of starch. The seed of the coffee astragalus (A. baeticus) is said to produce, when roasted, the true coffee flavor, and is much used in Sweden.

Many of the species of the order are ornamental, among which may be named the Judas tree or red bud (Cercis Siliquastrum); the Caraganas, shrubs with beautiful yellow flowers; the broom (Cytisus scoparius) of Europe naturalized along the sea coast, also used in medicine; the black locust (Robinia Pseudo-acacia), commonly planted as an ornamental tree. The laburnum (Laburnum anagyroides) is an ornamental tree with poisonous seeds and hard wood, used for turned work. The wisteria (Wisteria speciosa) a hardy species of the southern states and W. Chinensis of China, are cultivated as far north as central Iowa. The seeds of several species of the order are used for making necklaces, among these are the red seeds of the coral tree (Erythrina Corallodendron) of the West Indies, the crab's eye (Abrus precatorius) and the Ormosia dasycarpa. The Jequirity seed (Abrus precatorius) is used as a weight in India, according to Dr. Spafford, each seed weighing approximately I gram.

Medicinal Plants. Of the medicinal plants, in this family, the most important only will be mentioned here. The broom (Cytisus scoparius), native to Europe, is used as a diuretic and purgative. Fenugreek (Trigonella Foenum-graecum) was formerly used in medicine, but the powdered seeds are now used as an ingredient of curry powder and also in the preparation of stock foods; they have a characteristic odor and bitter taste. Licorice root (Glycyrrhiza glabra and var. glandulifera (Russian), native to the warmer regions of Europe and extending eastward into Central Asia, is made into extract of licorice which is used to cover the flavor of nauseous medicines and contains the glucoside glycyrrhizin. Cowhage (Mucuna pruriens) is a lofty climbing plant with dark purple flowers of the size of the sweet pea; pods from 2-4 inches long, covered with rigid, pointed, brown hairs, which if touched, enter the skin and cause

itching. The plant is used for the expulsion of intestinal worms, the hairs being mixed with honey and molasses. The young pods are used as food. The Calabar or ordeal Bean (Physostigma venenosum) is a climbing perennial plant resembling the scarlet runner and is native to tropical western Africa. It was formerly used by the natives to produce poisoning and is also an antidote against strychnin poisoning. The seeds contain several poisons, among them the alkaloids physostigmin $C_{15}H_{21}N_3O_2$ which contracts the pupil of the eye, calabarin a tetanizing principle, and eseridin $C_{15}H_{23}N_3O_2$ a purgative. The wood of Araroba (Andira araroba) is very bitter and is used in ointments. Balsam of Tolu (Myroxylon toluiferum), a native of Veneuzela and New Granada, is used as an ingredient in lozenges and contains a volatile oil tolene $C_{10}H_{10}$.

Logwood (Haematoxylon campechianum) is a spreading tree, native to Central America and Honduras, the bark of which is used for dyeing and staining, also for domestic purposes, such as in chronic diarrhoea, and contains heamatoxylin $C_{16}H_{14}O_6$ and heamatein $C_{16}H_{14}O_6$. Senna leaves are derived from the leaves of Cassia (C. acutifolia and C. angustifolia), the former being found in Nubia and the latter in southern Arobia and India. Senna leaves are used in medicine as a purgative and contain cathartic acid, chrysophan and two bitter principles sennacrol and sennapicrin. Several of our native species of Cas-



Fig. 287. European Licorice (Glycyrrhiza giabra). (After Faguet).

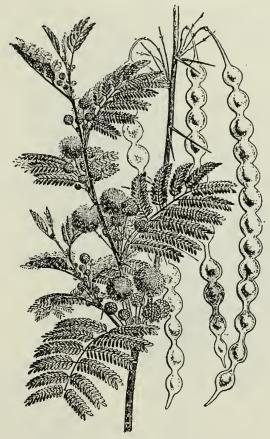


Fig. 288. Acacia (Acacia arabica). Flowering and fruiting branch. The source of gum arabic. (After Faguet.)

sia like the partridge pea (C. Chamaecrista), produce scours in sheep because of their purgative properties. The purging cassia (C. Fistula) is a tree indigenous to India. The pulp of the pod is a mild laxative. Clitoria ternatca of the Pacific Islands is a powerful cathartic. Tamarind (Tamarindus indica) a large, handsome tree indigenous to tropical Africa, is now widely distributed in other tropical countries. The fruit is used in medicine as a mild laxative and also in making a drink. It contains citric, tartaric, and other organic acids. The pulp and seeds are also eaten; the latter, when boiled, make a tenacious glue. The leaves and flowers are used as mordants in dyeing. Copaiba balsam (Copaifera officinalis) is a native to South Africa. The balsam is collected by Indians and used because of its stimulating action on the mucous mebmrane. It contains several acids, among them copaibic acid, $C_{20}H_{30}O_2$. Gum Arabic is obtained from Acacia arabica, the finest product coming from several species of the genus Acacia (A. Senegal), a plant well known to ancients. It possesses no real medicinal value. Catechu (Acacia Catechu), a small tree with thorny

branches, found in tropical eastern Africa, is used in medicine as an astringent. Under another name, Cutch, it is used in medicine because of its astringent properties. The resin from Cutch is made into cakes used for dyeing and tanning. Cutch contains catechol C_aH_aO₂ and catechutannic acid.



Fig. 289. Copaiba Balsam (Copaifera officinalis). Used medicinally. (After Faguet.)

Gum tragacanth (Astragalus gummifer) native to western Asia, is a spiny shrub with yellow flowers and is used to give consistency to lozenges; it contains traganthin, $C_6H_0O_5$ and arabin.

Poisonous and medicinal plants. Most of the substances occurring in the poisonous plants are mentioned under the species described. Many of the Leguminosae contain alkaloids; few of these are, however, found in Mimoseae, although alkaloids have been found in Acacia tenerrima, Albizzia lucida, and Pithecolobium Saman which are Mimosae. Of the Papilionaceae, the Sophoreae, Podalyrieae and Genisteae frequently contain alkaloids. Ulexin, found in seed of the Ulex europaeus, sophorin, in Sophora speciosa, and baptitoxin are identical with cytisin. Matrin C₁₅H₂₄N₂O, an alkaloid, resembling lupanin occurs in the root of Sophora augustifoli, Anagyrin C₁₅H₂₂NO₂, and cytisin, both occur in the seeds of Anagyris foetida, Baptisia and many other genera.

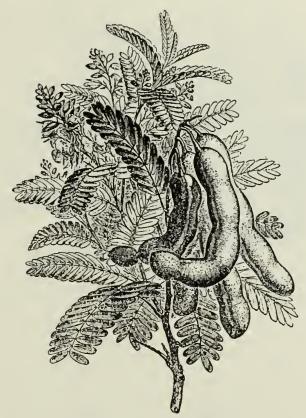


Fig. 290. Tamarind (Tamarindus indica). Fruit, flowers and leaves. Fruit used in making a refreshing drink; seeds furnish a glue. (After Faguet.)

Retainin $C_{15}H_{28}NO_{20}$, is obtained from the young branches of Genista sphaerocarpa. The seed of Trigonella Foenum-Graecum contains trigonellin $C_7H_7NO_2$; the same alkaloid is said to also occur in the pea, hemp and oats. Physostigmin $C_{15}H_{21}N_3O_2$, or eserin occurs in the ripe seed of Physostigma venenosum. The so-called calabrin is a secondary product; Robinia Nicou is said to contain nicoulin. Pancin $C_{27}H_{39}N_5O_5$ is found in the fruit of Pentaclethra macrophylla.

Quite a number of the Leguminosae also contain glucosides. One of the earliest discoveries made in connection with glucosides was of glycyrrhyzin found in some species of Astragalus, Abrus prectorius, and the root stock of Polypodium vulgare and other plants. The root of our wild licorice also contains a glucoside to the extent of 8.53 per cent. Glycyrretin, found in licorice, has the formula $C_{10}H_{24}O_{6}$.

Ononis spinosa contains a glucoside ononid and a second glucoside, ononin, $C_{30}H_{34}O_{13}$. Lupinin, $C_{29}H_{24}O_{16}$, is a glucoside found in the seedlings of Lupinus luteus, which through hydrolysis forms lupigin, $C_{17}H_{12}O_{6}$. Gastrolobin is found in the leaves and young branches of Gastrolobium bilobum; bap-

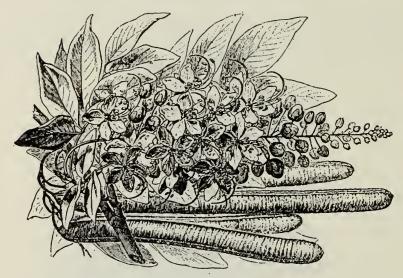


Fig. 291. Purging Cassia (Cassia Fistula). Flowering and fruiting branch. The pulp of the pod is a mild laxative. (After Faguet.)

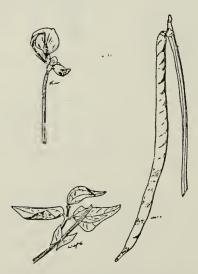


Fig. 292. Cow Pea (Vigna Catjang). A well known forage plant of the South. To the right a legume; to the left a flower and a part of a branch in the lower left hand corner. (W. S. Dudgeon.)

tisin, $C_{20}H_{32}O_{14}$, occurs in Baptisia; and tephrosin, a poisonous substance in Tephrosia toxicaria. A very toxic unnamed glucoside, $C_{33}H_{30}O_{10}$, occurs in the bark and root of Derris elliptica, Mundulea suberosa, and Lonchocarpus violaccus. Power, however, failed to find this glucoside in Derris uliginosa. Turboin, $C_{34}H_{32}O_{10}$, occurs in Tephrosia toxicaria. Gallotannic acid, $C_{14}H_{10}O_{9}+_{2}H_{2}O$ so called, occurs in the pod of Caesalpinia coriaria. The bark of certain species of Acacia contains 30 per cent of tannic acid. The wood of A. Catechu is colored red by catechin, a crystallizable substance.

Lindley in the earlier edition of his Vegetable Kingdom, states, that the plants of this family are on the whole wholesome and nutritious, and later declares that the family must be considered poisonous. The species used as food must be considered an exception.

In Australia there are several members of this family that are poisonous. Dr. Gray in an American Agriculturist of Oct., 1878, says:

What a pity that our cattle are not better acquainted with the corrected rule. In Europe and in the Atlantic States, no harm is known to come to cattle from want of proper discrimination. But when European flocks were taken to Australia and to pastures and forage almost wholly new, thousands of sheep perished in the Swan River Valley Colony in consequence of cropping the leaves of some leguminous plants to which they were attracted. What made the matter worse for the botanists, was that the very plants which did the mischief had been recommended by one of them (Mr. Preiss, a German) as the best thing the Agricultural Society could cultivate, as artificial food for stock. But another botanist, Drummond, a canny Scotchman, made some experiments, that proved that the people were right in charging the damage to these very species (of Gastrolobium) which the German botanist, on general principles, expected to be innocent and useful.

The Australian Gastrolobiums are all more or less poisonous, Baron Müller having long ago reported Gastrolobium grandiflorum as poisonous. calycinum known as the York road poison has a toxic base cygnin, cygnic acid C₁₀H₁₀O₄ which decomposes and forms gastrolobic acid C₂H₁₀O₅H₂O. The following species are recorded as poisonous by Maiden in Australia: G. trilobum, G. polystachyum, G. grandiflorum, poisonous to sheep and goats, the seeds being especially toxic but not to pigeons. The diseased animals have difficulty in breathing, then they stagger and die, death occurring in from 3-6 hours. The poison enters the circulation, stops the action of the lungs and heart. The raw flesh is said to poison cats, and the blood, dogs. The boiled or roasted flesh is, however, eaten by the natives and is not injurious. The blossoms are very poisonous. The Mirbelia racemosa is also poisonous to sheep, cattle, and goats. Two species of Goodia, according to Maiden, are poisonous, the G. lotifolia and G. mcdicaginea. These plants produce what locally goes by the name "black scours." The animals become weak, emaciated, and dic. The Gastrolobium and Crotalaria are stock killers in Australia though used as forage in South Australia. Maiden reports that the bean tree (Castanospermum australe) is poisonous to stock, especially the beans; when cooked, however, they are eaten by the Abyssinians. The box poison (Oxylobium parviflorum) is said to be a very poisonous plant to stock. The Gompholobium uncinatum is very injurious to sheep in New South Wales. The Swainsona Greyana and S. coronillaefolia arc poisonous. Sheep that eat them are called indigo-eaters. Both species act much like the loco weeds of the United States, "sheep go wrong in the head;" horses also act strangely. "The eyes stand out of their heads." This disease is identical with the "Nenta" disease of South Africa and the "Pea eating" disease of Australia. The South African disease is produced by Lessertia. The symptoms

from Astragalus mollissimus, Gompholobium, Sophora secundiflora, Cytisus proliferus, are all cerebral. Mac Owen regards them all as belonging to the same category and that Lathyrism caused by Lathyrus sativus is allied to them. That the well known forage plants Lotus corniculatus, and L. australis, of excellent repute, are often injurious to stock, but perhaps only from causing indigestion, as stated by Maiden, is worthy of note in this connection. Moussu and Desaint report the deaths of a flock of 54 sheep due to poison resulting from eating another plant of this order, Galega officinalis; 80 others in the same flock were badly affected. Ecchymoses were found in the walls of the alimentary tract and in the fatal cases a large amount of serum had collected in the pleural cavity. In subsequent experiments, it was found that 3 kg. of G. officinalis was sufficient to poison a sheep; the plant, however, seems not to be poisonous to rabbits. It is of interest to note that the genera Robinia, Indigofera, Wisteria, Caragana, Colutea, Swainsona, Galega, Lessertia, Astragalus, and Sesbania, all of which contain poisonous species, many of which are exceedingly toxic, belong to the tribe Galegeae of this family. Glychyrrhiza of the same tribe is not poisonous nor are all of the species in genera like Astragalus, Caragana, etc.

The Erythrophlaeum guineense contains an alkayoid erythophlein which acts like digitalin and picrotoxin. The Indigo of Australia (Indigofera australis) is regarded as poisonous in Australia. The Lathyrus sativus, L. cicera, and L. clymneum are poisonous but the active principles have not been isolated. Of other poisonous genera Robinia, Baptisia, Gymnocledus, Thermopsis, may be mentioned, but the treatment will be given more at length in another connection.

According to Dunstan and Henry, Lotus arabicus, when moistened with water and crushed, produces prussic acid. The glucoside lotusin is converted by the enzyme lotase into prussic acid. We may mention here that many of the spiny Acacias (A. palleus) of Australia may be injurious in a mechanical way.

The poisonous substance of Jequirity (Abrus precatorius) is a toxalmumin called abrin (found also in Cassia hispidula of Mexico) which is easily decomposed by heat. Behring has produced an antitoxin against the abrin or A. precatorius. The beans when cooked are eaten in Egypt.

A poisonous resin has been found in Wisteria chinensis, and a glucoside wisterin; Colutea arborescens, a well known southern European ornamental plant is poisonous, the leaves being so strongly purgative that they are frequently substituted for the genuine Senna. European authors list it among the poisonous plants.

The Tephrosia purpurea of Australia is poisonous to stock, and is used to stupefy fish. A large number of other plants of the order are used as fish poisons. Mention may be made of Derris, Abrus and Clitoria; others are mentioned in Part I. Some like Afzelia and Pithecolobiun are used as arrow poisons. The seeds of the jequirity plant (Abrus precatorius) are much used in India for the purpose of poisoning especially in criminal cases of cattle poisoning, less than 2 grams of the powdered seed causing death in 48 hours. The usual method of the "Chamar" or "Skinner" caste is to prepare small spikes, first soaking the seeds in water, then pounding them, and drying them in the sun; they are then sharpened upon a stone attached to a handle, and driven under the skin and left there. Daggers are rendered poisonous by being dipped into the powdered seed.

| According to Greshoff the leaves and also the seeds of Cassia marylandical contain saponin; leaves of Prosopis juliflora, Galega officinalis, Psoralea macrostachya as well as the seeds contain saponin. P. tenuiflora is regarded as poisonous and is avoided by cattle. Corolla not papilionaccous or only slightly so, endosperm copious. Flowers perfect; leaves abruptly pinnate | |
|--|---|
| Flowers polygamous or dioecious | |
| Leaves palmately 3-foliolate. | |
| Pod inflated4. Baptisia. | |
| Pod flat | |
| Leaves pinnate | |
| Stamens monadelphous, diadelphous, or rarely distinct. | |
| Anthers of 2 forms, stamens monadelphous. | |
| Leaves simple. | |
| Pod inflated6. Crotalaria. | |
| Pod flat. | |
| Leaves 1-3-foliolate8. Cytisus. | |
| Leaves usually 7-11 foliolate | , |
| Anthers all alike. | |
| Leaves generally 3-foliolate. | |
| Flowers in racemes; pods coriaceous9. Melilotus. | |
| Flowers in heads; pods membranous10. Trifolium. | |
| Flowers in spikes or heads; pods curved10. Medicago. | |
| Leaves pinnately foliolate. | |
| Leaves not tendril bearing; plants not climbing. | |
| Herbs with glandular dots | |
| Leaves odd pinnate. | |
| Herbs. Flowers large | |
| Trees or shrubs | |
| Leaves evenly pinnate | |
| Pod turgid inflated. | |
| Leaflets not toothed, or only at the apex. | |
| Keel tipped with an erect point17. Oxytropis. | |
| Keel not tipped with an erect point16. Astragalus. | |
| Leaslets toothed all round20. Cicer. | |
| | |

Leaves with tendrils; style bearded at the apex.......18. Vicia.

Style bearded down one side19. Lathyrus.

Herbaceous plants; leaves with tendrils or climbing.

Herbs, shrubs, or in tropical regions, trees, with abruptly pinnate leaves; calyx of 5 sepals united at the base; petals 5; somewhat unequal, spreading, imbricated, and clawed; stamens usually 10, or 5, often unequal, and some imperfect; anthers all alike, or the lower larger, opening by 2 pores at the apex;

ovules numerous. About 270 species, mostly in warm and temperate regions. A species well known in medicine is Senna (C. acutifolia and C. angustifolia) with leaves which are laxative.

Cassia Chamaecrista L. Partridge Pea

An annual, spreading, 1 foot long; leaves with a sessile gland on the petiole; leaflets of 10-15 pairs; flowers large, showy; petals yellow, with a purple spot at the base; anthers 10, elongated, and unequal, 4 yellow, the others purple.

Distribution. In dry or sandy soil from Maine to South Dakota, Texas to Florida.



Fig. 293. Pea Partridge (Cassia Chamaecrista), c. Pod. a. Pistil. b. Stamens. (C. M. King)

Poisonous properties. This plant is common in hay and when the seeds are consumed in large quantities, has a cathartic action. Cases of mild poisoning to sheep have been reported to the writer. C. marilandica, a plant with curved pods that are somewhat hairy at first, possesses similar properties. C. hispidula contains abrin.

2. Gymnocladus, Lam.

Trees with large, bipinnate leaves, and showy, white, dioecious or irregular, polygamous flowers; calyx elongated-tubular below; 5-cleft, the lobes narrow, nearly equal; petals 5 (rarely 4), oblong or oval; stamens 10, distinct, short,



Fig. 294. Wild Senna (Cassia Marylandica). 1. Flower. 2. Pods. A plant growing in the Eastern Atlantic States as far south as North Carolina. Laxative like the Common Partridge pea. (Selby, Ohio Agrl. Exp. Stat.)

inserted on the petals; ovary rudimentary, or none in the staminate flowers, sessile and many-ovuled in the pistillate; pod oblong, thick, large, and coriace-ous.

Gymnocladus dioica (L.) Koch. Kentucky Coffee-tree

A large tree with rough bark; leaves large and ample, 2-3 feet long; 7-15 leaflets, ovate or acute; glabrous or pubescent on the veins beneath; racemes many-flowered; flowers slender-pedicelled; seeds hard, ½ inch across, imbedded in a sweet, but disagreeable, and somewhat mucilaginous, material.

Distribution. From Western New York to Pennsylvania, Eastern Nebraska, and Arkansas.

Poisonous properties. Cases of poisoning are not uncommon. The alkaloid cytisin $C_{11}H_{14}N_2O$, a crystalline, rather bitter, and caustic substance which causes dilation of the pupil, is reported to have been found, according to Chesnut, in the leaves and soft pulp of the fruit of the coffee bean. The pulp has long been used, when mixed with milk, to poison flies. In speaking of the symptoms and treatment, Prof Chesnut says:

Few accidental cases of poisoning arise, but the pulp, in one instance, caused severe illness in a woman who ate a small quantity, mistaking it for that of the honey locust (Gleditsia triacanthos), which is frequently eaten by children. The symptoms were not fully noted at the time, but are described from memory as conspicuously narcotic. The effect began within five minutes and lasted several hours. The treatment should probably be the same as that for laburnum, viz., emetics, stimulants, injections of coffee, and an alternately hot and cold douche to the head and chest.



Fig. 295. Kentucky Coffee Tree (Gymnocladus dioica). Inflorescence from staminate tree. 2. Pistillate flower. 3. Diagram of flower. 4. Longitudinal section of staminate flower. 5. Pistillate flower with a portion removed. 6. Pistil with a section of ovary removed. 7. Portion of branch bearing a single fruit, showing seed and embryo. 8. Cross section of seed. 9. Portion of leaf. 10. Portion of winter branch. 1, 2, 7, 8, 9, 10, one-half natural size. (M. M. Cheney in Green's Forestry in Minnesota.)

3. Thermopsis R. Br.

Perennial with finely oppressed pubescence, 2-3 feet high; leaves rhombicolate leaves and foliaceous stipules; flowers large, yellow or purple, borne in racemes; calyx bell-shaped or short-turbinate, with equal and separate lobes or the upper united; standard nearly orbicular, as long as the oblong wings and the keel; stamens 10, separate and in-curved; pistils sessile or short-stalked, frequently flat, linear, oblong or curved, ovules numerous. A small genus of about 15 species of North America and Asia.

Thermopsis mollis (Michx.) M. A. Curtis. Alleghany Thermopsis

Perennial with finely appressed pubescence, 2-3 feet high; leaves rhombic-lanceolate, 1-3 inches long, entire and nearly sessile; stipules ovate or lanceolate;

racemes chiefly terminal; flowers yellow, pod short-stalked and narrow and somewhat curved.

Distribution. In the mountains of Virginia, North Carolina and Tennessee.

Thermopsis rhombifolia Richards. Prairie Thermopsis.

An erect perennial from 1-2½ feet high, appressed, silky pubcscent; stem angular; leaves with broad conspicuous stipules; leaflets obovate, at length nearly glabrous, bracts oval; flowers yellow, in a rather short raceme of few flowers; pod linear and curved, spreading, several seeded.

Distribution. In sandy soil and foot hills of the mountains from Manitoba to South Dakota, Nebraska and Kansas, west to the Rocky Mts., and in Utah, Wyoming and Montana.

Poisonous nature. This plant is very common in the foot hills, and is supposed to produce poisoning of stock. It is often consumed by sheep. It is said that the seeds of the plant are poisonous and the Canadian Department of Agriculture reports several cases of poisoning to children where the seeds were eaten. T. montana is a species occurring from western Nebraska anl Kansas to the Pacific Coast. Species of Thermopsis are said to contain cytisin.

4. Baptisia Vent

Perennial herbs with palmately 3-foliolate, or rarely simple leaves; basal sheathing scales; flowers large, in racemes; calyx 4-5-toothed; corolla with a

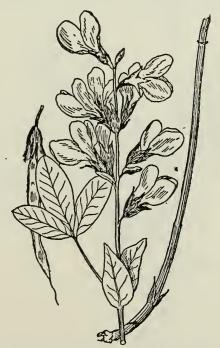


Fig. 296. Yellow-flowered Bitter weed (Thermopsis montana). Plant is exceedingly bitter. (From U. S. Dept. Agr.)

large standard, but not longer than the wings; stamens 10, distinct; pods stalked, roundish, oblong, inflated, and many-seeded; seeds often spreading and rattling. About 16 species in Eastern North America.

Baptisia australis (L.) R. Br. Blue False-Indigo

Tall, smooth, stout perennial 4-5 feet high; leaves short-petioled; leaflets oblong or oblanceolate, obtuse; stipules conspicuous, persistent; racemes terminal, loosely flowered; 1-2 feet long, erect; flowers blue.

Distribution. From Western Penn. to Arkansas and Kansas to Ga.

Baptisia tinctoria (L.) R. Br. Yellow or Indigo Broom

A smooth, slender perennial herb 2-3 feet high; leaves nearly sessile; leaflets obovate or oblanceolate, sessile or nearly so; racemes few-flowered; flowers yellow.

Distribution. In dry soil from Maine to La., west to Minn.

Poisonous properties. Baptitoxin which is probably the same as cytisin occurs in Baptisia tinctoria. The glucoside baptisin $\rm C_{20}H_{32}O_{14}$ occurs in some species of the genus.

Dr. Millspaugh states that disturbances produced by taking considerable quantities of the tincture are:

Vertigo; dull, heavy headache with weakness and weariness of body, and tendency to delirium; soreness and lameness of the eyeballs, with hot, flushed face; tongue coated white, yellow, or yellowish-brown; loss of appetite; nausea, and burning in the stomach;



Fig. 297. Wild Indigo (Baptisia bracteata). Said ot be poisonous. (Ada Hayden.)

dull pains in the region of the liver, especially at the site of the gall-bladder; face sallow, with burning cheeks; constant pain and aching in the abdomen, followed by marked distention, and soreness on pressure.

According to Dr. Hughes, Baptisia excites true primary pyrexia in the human subject. This pyrexia is very much like that of the early stages of typhoid.

Baptisia leucantha T. & G. Large White Wild Indigo

A smooth, erect perennial herb, petioled leaves; leaflets obtuse, rounded, or sometimes slightly emarginate; stipules deciduous; racemes lateral; flowers white or cream color. Prairies and alluvial soils, Ont. to Minn., to Fla. and La.

Poisonous properties. According to Hyams it is a violent emetic and cathartic when taken in large doses and in small doses a mild laxative.

Baptisia bracteata (Muhl.) Ell. Large-bracted Wild Indigo

Perhaps more common in sandy soil in the west than B. leucantha. It is also shorter and flowers earlier in the season. Prairies, Mich, to Minn., La., Tex.

Poisonous properties. Dr. Schaffner states that the blue wild indigo and the yellow wild indigo are emetic, and that the latter species is regarded as

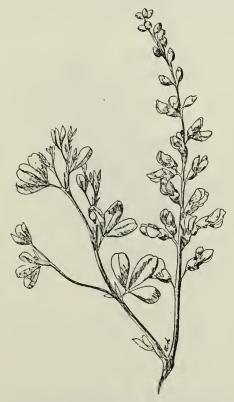


Fig. 298. Wild Indigo (Baptisia leucantha). This plant is a native of prairies and alluvial soils. Said to be poisonous. (Ada Hayden.)

poisonous. The taste of the plant is rather disagreeable, and it is not generally eaten by stock.

5. Sophora L.

Shrubby or herbaceous perennials; leaves odd pinnate with numerous leaflets; flowers white, yellow or violet in terminal racemes or panicles; calyx bell-shaped, with short teeth, standard rounded or obovate; wings obliquely oblong; keel oblong, nearly straight; stamens all distinct or nearly so; pistil short-stalked; style incurved; pod stalked and terete, constricted between the seeds. About 25 species, of warm and tropical regions.

Sophora sericea Nutt. Silky Sophora

A low perennial herb from 6-12 inches high, more or less silky canescent; stipules subulate, deciduous, leaflets about 21, elliptical or cuneate oval; flowers in short terminal racemes; nearly sessile; corolla white, pod dry, leathery, finely pubescent and few seeded.

Distribution. The prairies of Nebraska and Colorado to Texas and Arizona. Poisonous properties. Mr. Chesnut says of this plant:

The silky sophora, of the Southern Great Plains region, has been somewhat vaguely reported as one of the plants that "loco" horses in that region. The seeds contain a very poisonous alkaloid.



Fig. 299. Silky Sophora (Sophora sericea). A plant of the plains and thought to be poisonous. (Charlotte M. King.)

Sophora secundiflora DC. Coral Bean

A stout shrub or small tree, with deep green leaves of about 9 elliptical, oblong, obtuse, coriaceous leaflets; terminal racemes of showy violet, fragrant flowers; and large, woody pods 3-5 inches long, containing 3-4 round red beans as large as small marbles.

Distribution. Common from the Gulf Coast to the Pecos and less abundant in mountain canons to New Mexico. It is mostly shrubby, but becomes a tree 30 feet high and forms groves in the vicinity of Matagorda Bay.

Poisonous properties. Mr. Chesnut says:

The beautiful bright red berries of the Frijolillo or coral bean of southern and western Texas contain a powerful poisonous alkaloid. The plant is said to have poisoned stock in Texas and in northern Mexico.

It contains *sophorin*, an amorphous alkaloid, which, according to Czapek, is probably identical with *cytisin*. The beans are somewhat used by the Indians to produce intoxication.

6. Crotalaria (Dill) L. Rattle-box

Herbs or occasionally somewhat woody plants, with simple or 3-7-foliolate leaves; yellow flowers borne in racemes; calyx 5-cleft, somewhat 2-lipped, standard large, heartshaped; wings oblong or obovate; keel curved, stamens monadelphous, with anthers of 2 forms; pod inflated like the pea, but shorter and many-seeded. About 250 species found chiefly in the tropics.

Crotalaria sagittalis L. Rattle-box

Annual from 3 inches to a foot high, with a small straight root; stem branched, villous, terete or wing margined; leaves oval or oblong-lanceolate,



Fig. 300. Rattlebox (Crotalaria sagutalis). a, whole plant; b, cross section of seed pod—both one-third natural size. The cause of crotalism in horses. (U. S. Dept. Agr.)

from 1/2-1/3 of an inch wide, edge of the leaf entire or somewhat wavy and hairy; stipules united and decurrent on the stem, inversely arrow shaped; peduncles few-flowered; flowers yellow, about ½ of an inch in diameter; calyx 5-cleft, standard of the flower large, heart-shaped; keel scythe-shaped; stamens monadelphous, anthers of 2 sizes, 5 smaller and roundish; pod large, inflated, bears a close resemblance to the garden pea, greenish at first, becoming blackish; seeds from 1/10-1/12 of an inch in diameter, flattish, kidney-shaped, which, when mature, break away from the point of attachment and rattle in the pod, hence the name "rattle-box."

Distribution. This plant is common in sandy soil from Maine to Minnesota, South Dakota, Iowa, Nebraska, and Northern Texas. The plant is extremely common on the sandbars of the Missouri river, where it may be collected by the wagon load.

Poisonous nature. The earliest mention of the poisonous nature of the weed was made by Drs. Stalker and Bessey. Dr. Stalker who performed some experiments with the plant gives the following symptoms:

The disease had been known in this region for three or four years, but had not until the present summer (1884) prevailed to such an extent as to attract generally public attention. But now the loss in horse stock on some farms was not to be counted by hundreds, but by thousands of dollars. The disease proved to be one that had not hitherto come within the range of my experience, nor had I any information of anything exactly identical with it. I spent several days among the farmers on the Iowa side of the Missouri river, taking careful notes of the symptoms, and gathering the history of the progress of the disease. On some farms I found almost all the horses affected, and on others but a few individuals. Deaths were an almost daily occurrence, and the farmer who owned a large stock of horses did not know today whether he would have teams for his farm work a week later. The disease in most cases is very slow in its progress, but proving almost uniformly fatal after a number of weeks or months. There is a general decline of bodily vigor throughout this period, and the only ahnormal symptom in many cases is that of marked emaciation and consequent weakness. Horses that have been kept at pasture through the summer, without work, and where the grass grew in greatest abundance, were so thin in flesh that they walked with the greatest difficulty. A critical examination of many of these patients revealed nothing more than the condition resulting from starvation. This was not uniformly the case. In a number of instances there was marked coma or stupor, the animal often falling asleep while eating. In some instances the animal would remain standing for a whole week, sleeping much of the time with head resting against some object. In a few instances the animal lost consciousness, and broke through fences and other obstructions. A number of the diseased animals were placed at my disposal, and assisted by Dr. Fairchild and Dr. Milnes, I made post mortem examinations of five subjects with the most perfect uniformity as to the lesions presented. In every instance there were marked haemorrhagic effusions into the fourth ventricle, the liver and splcen were abnormally dense, the walls of the intestines were almost destitute of blood, and the stomach enormously distended with undigested food. The stomach with its contents in some instances weighed as much as seventy pounds. These post mortem conditions, together with clinical symptoms, led me to believe the animals were obtaining some poisonous principle with their food. The symptoms in some cases bore such a resemblance to those produced by eating Astragalus mollissimus, or "loco plant" of the Western plains, as to direct my investigations to that family of plants. A careful examination of the meadow and pasture lands was not rewarded by the discovery of a single "loco plant."

It took but little investigation, however, to find a closely related plant growing in great abundance, both in the meadows and pastures. This was the Crotalaria sagittalis, or rattle-box. This is also known as the wild pea, and is accounted hy many farmers as the best of forage plants. Knowing the bad reputation of some of its near relatives, I determined to make some experimental tests with the plant. I employed a boy to collect about thirty pounds of the green plants, which I brought with me on my return to the college. I procured a strong young horse, affected with incurable catarrh, and attempted to induce him to eat the plant. This he persistently refused to do, though I sharpened



Fig. 300a. White Lupine (Lupinus albus). A forage plant introduced from the Mediterranean region. Seeds contain a bitter alkaloid, U. S. Dept. Agr.

his appetite by a protracted fast. It is a matter of common observation that animals eat it with the greatest relish in localities where it grows. Failing to induce the animal to take the plant voluntarily I prepared a strong infusion, and by means of the stomach pump gave the preparation obtained from about ten pounds of the plant. In twenty minutes stupor began to ensue, the eyes were closed, the head was rested against the side of the box, the breathing became stertorous, and all the symptoms developed that were to be seen in the patients previously examined. At the end of six bours the stupor began to disappear, the eye began to regain its brightness and in another bour the horse began to eat. The following day, when he had apparently recovered from its effects, he was given half the quantity of the drug as on the previous day. In this instance the symptoms were developed much more rapidly, the animal became unconscious in a short time and died in an hour and a half. The post mortem revealed the same condition of the brain as in the cases examined in the Western part of the State. I now resolved to make a second experiment, in which the animal should receive a small quantity for a number of days in succession. Having procured another subject for experimentation, and a bushel of mature fruit, or pods of the plant, I commenced on Sept. 5th, to give daily the infusion obtained from about one quart of the pods. On the fifth day of the experiment the characteristic stupor came on. The animal rested its head against the box and slept while standing. The symptoms grew more marked till the thirteenth day of the experiment, when the animal died. The post mortem showed the same as in the other cases. These experiments leave no doubt in my mind that the trouble along the Missouri river is occasioned by the animals' feeding on this little plant. It is from eight inches to a foot in height, with branching stems bearing yellow flowers in July and developing large pods resembling the pea, but containing a number of black, hard seeds. It grows on sandy bottom land, and is very abundant in the meadows and pastures in portions of the Missouri bottom. It is seldom seen among the tame meadow grass in any considerable amount. It thrives best among the wild grasses. Animals, doubtless, eat it much more than formerly, when the wild pasturage was better than at present. Cattle sometimes, though not often, suffer in the same way as borses.

The disease is also known as the Missouri Bottom disease. Hundreds of horses in the Missouri Bottom in Western Iowa and Eastern Nebraska die from eating this weed, it being most common in unbroken fields. Horses should be kept from all suspected fields. Only cultivated grasses and forage plants should be grown. Some doubt has been expressed that this plant is the cause of the trouble. The writer a number of years ago, in collaboration with Dr. Miller, investigated an outbreak near Council Bluffs. This disease occurred only in the bottoms, where the weed was common, and a large number of horses die from it annually. No other injurious plants could be found except some ergot on wild rye. A decoction of the weed found here was fed by Dr. McNeill to a horse but no injurious symptoms followed. A decoction of the seeds was fed to a guinea pig without any serious symptoms. Dr. F. B. Power however found a small amount of an alkaloid in the seeds which caused slight illness in a kitten. From all of these experiments we may conclude that rattle box is injurious under some conditions.

7. Lupinus (Tourn.) L. Lupine.

Herbs or rarely shrubs with generally palmately compound leaves; stipules adherent to the base of the petiole; flowers showy, in long, dense racemes; calyx deeply toothed and 2-lipped; corolla with an orbicular or ovate standard with margins reflexed; wings oblong, or obovate, lightly cohering, and enclosing the keel, which is incurved or beaked; stamens monadelphous, anthers of 2 forms; pistil with an incurved style and sessile ovary; pod flattened, somewhat constricted. About 100 species of temperate regions, or a few in warm regions. The North American species are chiefly west of the 100th meridian. The Lupinus perennis occurs in sandy soil from New England to Minnesota and Louisiana; L. albus, L. luteus and L. angustifolia are cultivated for forage in Europe, the seeds being used as a substitute for coffee.

Lupinus argenteus Pursh. Hairy Lupine

A much branched perennial, slightly shrubby, from 2-3 feet high, silky pubescent hairs appressed, leaves with small stipules; petioles equalling or longer than the leaves; leaflets sessile, narrowed at the base; flowers in rather dense, terminal racemes, purple; pod silky, pubescent, generally 3-5 seeded. A very variable species.

Distribution. Prairies of South Dakota to Western Nebraska to New Mexico, Utah, and from Arizona to Montana. Abundant in the foothills.

Lupinus perennis L. Wild Lupine

Perennial, somewhat hairy; erect stems, 1-2 feet high; leaves compound; 7-11 oblanceolate leaflets; flowers showy, purple-blue, in a long raceme; pods/broad, very hairy, 5-6-seeded.

Distribution. Sandy soil from New England to Minnesota, Missouri, and the Gulf region.

Lupinus plattensis Watson. Nebraska Lupine

Somewhat like the preceding, with appressed silky-villous hairs, and a glaucous hue; leaflets spatulate; flowers in loose and short peduncled racemes; petals pale blue.



Lupine in flower (Lupinus sp.) On the western ranges. (Bull. Nev. Agr. Exp. Sta. 55),





Fig. 301. Wild Lupine (Lupinus Plattensis). Causes lupinosus. Charlotte M. King

Lupinus leucophyllus Dougl.

Leafy, densely silky-tomentose perennial; compound leaves; 7-10 oblong-lanceolate leaflets; flowers in sessile racemes, densely flowered; petals blue or pink.

Distribution. Rocky Mountains, Colorado to Washington, and Northern California.

Lupinus holosericeus Nutt

A perennial, shrubby plant with silvery-canescent leaves; 12-20 inches high; compound leaves; 5-9 lanceolate leaflets; flowers in whorls or scattered; calyx bracteolate, the upper slightly 2-cleft; petals bright blue.

Distribution. Oregon to California.

Lupinus species. There are many other species of *Lupinus* in the Rocky Mountains and along the Pacific Coast. A great many of these have been looked upon with suspicion.

Poisonous properties. European white lupine, Lupinus albus, L. luteus, and others contain the glucoside lupinin $C_{20}H_{24}O_{16}$, a crystalline substance with a bitter taste and a fruity odor; lupinidin $C_8H_{16}N$, a pale yellow, heavy, oil with a pungent, bitter taste; lupinin $C_{10}H_{10}NO$, also bitter with an apple-like odor; Lupinus angustifolius contains lupinin $C_{15}H_{24}N_2O$, an intensely powerful alkaline substance. The substance arginin $C_6H_{14}N_4O_2$, found in the etiolated cotyledons of the lupine and the Soy bean, is a proteid. Prof. Chesnut says in regard to the Lupinus leucophyllus Dougl.:

The above species is very abundant in Montana, where it is said to have caused the death of a very large number of sheep. There is some question whether the animals were killed by a poisonous constituent of the plant or merely by bloat. The seeds of all the lupines are probably deleterious in the raw state. In Europe, bowever, the seeds of Lupinus albus, after the bitter taste has been removed by steeping and boiling, are eaten by luman beings as well as by cattle.

The so-called ictrogen obtained by European chemists from some of the lupines can be extracted by weakly alkaline water and is to be regarded as an active poisonous principle. Some European investigators, however, think that the alkaloids are not the cause of the poison. To the above poisonous species we may add I. linifolius, and I. hirsutus. The disease caused by these has long been known in Europe and has received the name of lupinosis. It is common where lupines are used for forage purposes. According to Friedberger and Fröhner from one-half to three-fourths of the animals perish. According to Arnold and Schneidermuhl the disease can be produced experimentally with lupinotoxin in sheep, horses, goats, and pigs. This substance occurs chiefly in seeds and pods. Dry heat does not destroy it but steam under pressure does. There is a probability that the poison is produced by metabolism.

Chesnut and Wilcox, in their paper on Stock-poisoning Plants of Montana, make the following statement in regard to the Lupine poisoning of Montana:

So far as we have been able to observe, lupines are not very extensively eaten by sheep during the spring and summer. This statement is at least true for normal conditions where sheep are acquainted with the range and are not being trailed or driven. Horses and cattle take kindly to lupines and eat them in large quantities during their immature stages. When sheep are being trailed through strange country, or when they have just been unloaded from cars, and are in a hungry condition, they eat lupines ravenously in any stage of growth. The lupines are not considered valuable as forage plants for sheep until after early fall frosts, or until other forage plants have become dry and uninviting as



Great Basin Lupine (Lupinus holosericeus). Ripened pods of this are probably poisonous. (Bull. Nev. Agr. Exp. Sta. 62).



fodder. In late fall, and especially after early snowstorms, the lupines constitute one of the chief forage plants on some of the mountain ranges. It should be remembered that the leaves of lupines remain green and the plants offer slightly succulent forage after other plants have become dry.

The first case of poisoning from lupines which was brought to our attention occurred in August, 1896. A band of sheep, while being moved from one range to another was driven rapidly, and was constantly in a very hungry condition, when it was allowed to feed in a field of lupine for a short time. Within two hours after beginning to eat the lupine a number of sheep manifested violent symptoms of poisoning, and a few died within one hour after the appearance of the first signs of poisoning. Of the 200 sheep in the band 100 had died before the following morning. The season of 1896 was rather late and at the time when the poisoning occurred the lupine pods were fully formed, but the seeds were not quite ripe. In this case the sheep were driven away from the lupine as soon as the first symptoms of poisoning had been noticed and some of the sheep had eaten only small quantities of the plant. About 150 out of the 200 were affected, and as only 50 of these ultimately recovered it will be seen that the death rate was very high.

The owner of these slicep, during the same season cut a quantity of lupine hay during the second half of July. In the winter of 1897 a band of 150 bucks belonging to the same sheep raiser were kept in a covered corral and were fed on cultivated hay. On one afternoon during the winter these bucks were given a liberal quantity of the lupine hay. About three hours after feeding this hay a noisy disturbance was noticed among the sheep. Upon investigation the owner found the sheep in a frenzied condition, and during the night about 90 of them died. No more lupine hay was fed and no more trouble was experienced.

They state further that the lupine poisoning occurred in various parts of the state, in 1898 about 2,000 having been poisoned. 1,150 sheep died out of a single band of 2,500 sheep. They also state that one sheep raiser in Deerlodge Valley lost 700 sheep from the poisoning of lupine. They report another case which occurred on June 28, 1900, near Livingston, in two bands of sheep, each numbering 3,000, which were being trailed westward from Livingston. The sheep were liberally salted before being started on the trail; the first day they traveled about 5 miles, and camped on opposite sides of a small stream. After watering, one band was driven across the creek and camped on a bench about 30 feet higher than the stream. On the following morning, the sheep which had been driven across the stream manifested symptoms of poisoning; ultimately 1,900 died. This poisoning is referred to as lupinosis, a disease of which in Europe both acute and chronic forms are recognized, but in the United States the chronic form only has been recognized. The marked symptoms of poisoning are acute cerebral congestion, and great mental excitement. The sheep rush about in different directions, often running against the herder or other persons. The first stage of frenzy is followed by a second stage in which there is pronounced irregularity of movements and violent spasms, and falling fits. In many cases death occurs in from 1 to 11/2 hours. The pulse during the attack is strong and regular. Lower animals are attacked by convulsions, and these convulsions resemble those caused by strychnine poisoning. The excretion of the kidneys is increased, and sometimes it is bloody. The post mortem conditions are described by Chesnut and Wilcox as follows:

Post mortem examinations of the sheep poisoned by lupines revealed conditions very similar to those found in the acute cases of loco disease, already described above, with the exception that in loco disease the kidneys were not affected. The lungs were slightly congested, but this condition was not so pronounced as in cases of larkspur poisoning. The cerebral membranes were in all cases congested. In the more violent cases small blood vessels had been ruptured in various parts of the body, which may bave been due either to increase of blood pressure or to the struggles of the animal.

In regard to the treatment, Dr. Wilcox recommends as follows:

No remedies have been tried in cases of stock poisoning from American species of lupine. From our general experience with potassium permanganate it seems reasonable

to suppose that this substance would probably destroy the lupine alkaloids in the stomach if administered promptly after the first signs of poisoning. In the main, however, reliance should be placed upon prevention. With regard to the use of lupine hay, our experience and observations indicate that this is always dangerous for sheep if cut at a time when the seeds are retained in the hay. Since the limit of the period during which lupines are not poisonous can not be determined for the present with any certainty, it seems advisable to abandon entirely the use of lupine hay for sheep, except after a preliminary test in feeding large quantities of the hay to one or two sheep. If it should prove to be non-poisonous, it may then, of course, be fed with safety.

The poisonous principle in all plants which have been fully investigated varies in quantity according to the stage of growth of the plant, and is located more abundantly in one part of the plant than in another. These facts seem to be strikingly true of lupine, since, as already indicated, the plants are sometimes eaten in large quantities with impunity, while at other times the plants cause extensive losses, especially among sheep. The evidence thus far collected regarding this matter indicates that the seeds are the most poisonous part of the plant.

Mr. O'Gara of Nebraska, in speaking of the Lupine says this:

There are three species of Lupines in the western part of the sand-hill region and throughout the foot-hills, which are worthy of attention. So far as can be learned, cattle and horses either do not eat them or are not harmed by them, but sheep men say that they are extremely poisonous to sheep when eaten after the pods have formed and have begun to ripen. Many sheep owners are very careful to avoid patches of Lupine in driving their sheep from one range to another, and never trust the flock to a green herder who is unacquainted with the range.

The three species common to the regions mentioned are the Nebraska Lupine (Lupinus Plattensis S. Wats.), the Silvery Lupine (Lupinus argenteus Pursh.), and the Low Lupine (Lupinus pusillus Pursh.). The last named is a small hairy plant four to eight inches high, much branched near the root, bearing commonly five leaflets at the end of the leaf-stalk. The densely clustered blue flowers are borne on a stalk four to eight inches long. The pod is finely-hairy and is three-fourths to one inch long.

Dr. Nelson conducted some experiments in poisoning from three species of Lupinus: L. ornatus, L. sericeus, L. leucophyllus; from which the following results were obtained with reference to the effect of feeding quantities of these plants. In regard to the first of these species, negative results were obtained in part; to sheep fed as early as May 30th, June 8th, July 14th, July 31st, and August 2nd, partly in the year 1898, and partly in 1901, these experiments being made in 1898 and 1901. In 1904 experiments were made with two sheep fed between November 16th, and December 22nd, receiving 274 pounds of this hay. This hay was eaten fairly well, and the sheep were given no other food except the lupine, and had constant access to water.

Some loss of flesh occurred in both sbeep and one of them became affected, December 28th, with an attack of stomatitis with quite well developed ulcers in the mouth. He practically recovered by January 1st. No other untoward symptoms were manifested during the course of the experiment.

In regard to *Lupinus sericeus*, the results were in part negative, but June 28th, 1899, a sheep was fed 2 pounds of lupine that was in full bloom and partial fruit, gathered a few days previous. On the morning of the 29th, the sheep was drowsy, and kept a recumbent position. In the afternoon the comatose condition was more marked; he walked with an unsteady gait and pressed his head against the fence when he happened to reach it, showing a partial paralysis. The animal died on June 30th, slightly bloated.

The ventricles of the heart were partially and the auricles completely filled with a black coagulated blood. The lungs were congested, the stomach filled with partially digested food, otherwise apparently normal.

The experiments with Lupinus leucophyllus were negative.

8. Cytisus L.

Shrub with trifoliolate or unifoliolate leaves; showy flowers, chiefly in terminal racemes; calyx 2-lipped, with short teeth; standard ovate or orbicular; keels straight or curved; anthers large and small; ovary with many-ovuled, and incurved style; pod flat, oblong or linear. About 40 species, natives of Europe, Western Asia, and Northern Africa.

Cytisus scoparius (L.) Link. Broom

A stiff, nearly glabrous shrub; elongated, straight, angled branches; lower leaves 3-foliolate, obovate; leaflets which are mucronate-tipped; upper leaves sessile, often unifoliolate; flowers bright yellow, in elongated leafy racemes.

Distribution. Along the seacoast of Nova Scotia to Virginia, and very common along the Pacific Coast.

Poisonous properties. The Scotch Broom (Cytisus scoparius) common on the Pacific and Atlantic coast but naturalized from Europe, contains the alkaloid cytisin $C_{11}H_{14}N_2O$ and is poisonous. Blyth records 400 cases of poisoning from this. The symptoms in stock are slavering, vomiting, staggering, and general paralysis. Cytisin, occurring in many of the Genisteae, was found, in 1818, in Laburnum anagyroides and since then, has been found in many other species of the genus Cytisus and in Ulex europaeus, Sophora sp., Thermopsis sp., Baptisia tinctoria, Anagyris foetida, Lotus suaveolens, Colutea cruenta, and Euchresta Horsfieldii. Some species of the genera Genista and Cytisus do not contain cytisin. Cytisus scoparius also contains a volatile alkaloid spartein $C_{15}H_{26}N_2$, a single drop of which, according to Blyth, killed a rabbit that showed symptoms similar to those of nicotin poisoning.

9. Melilotus Tourn

Annual or biennial herbs with trifoliolate leaves; small white or yellow flowers in racemes with the odor of *cumarin*; teeth of the calyx short and nearly equal, shorter than the pod; corolla deciduous with obovate or oblong

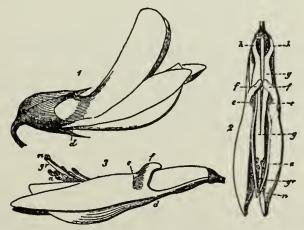


Fig. 302. Flowers of Sweet Clover (Melilotus alba). 1, Standard above. 2, Showing wings and keel. 3, Showing stamens and pistil in keel.

standard, obtuse keel, free from the stamen tube; stamens diadelphous; pod wrinkled, straight, ovoid or globose—1-2-seeded. A small genus of 20 species, native to Europe, Africa and Asia. 3 species naturalized, found in North America, 2 of them quite weedy.

Melilotus alba Der. Sweet Clover

An erect annual or biennial from 2-4 feet high; rather distant, compound leaves, leaflets obovate, oblong, obtuse, serrate, narrowed at the base, truncate, emarginate or rounded at the apex; flowers with white petals, small, fragrant; pod ovoid, reticulated and smooth.

Distribution. Abundant in waste places in the eastern and Atlantic states, also in the southern states and throughout the Mississippi valley, the Rocky Mountain region and the Pacific coast. Sweet clover is one of the most common weeds in pastures, and along roadsides.

Melilotus officinalis (L.) Lam. Wild Yellow Sweet Clover

An upright, yellow flowered herb from 1-4 feet high; leaflets oblong, or oval, the apex more or less obtuse; corolla yellow; pod with irregularly reticulated veins.

Distribution. Common in waste places in the irrigated districts of the west, becoming more or less common in the Mississippi Valley and along the Atlantic Coast.

Melilotus indica (L.) All. Sweet Clover

An upright annual like the preceding, but with much smaller yellow flowers. Distribution. Native to Europe, introduced in ballast along the Atlantic coast and abundant on the Pacific coast.

Poisonous properties. The sweet clovers contain the substance cumarin C₂H₈O₂, which is found in the Tonka bean, sweet vernal grass, vanilla grass, etc. In Europe the sweet clover is suspected of being poisonous. This plant is used as a forage plant in the South, and Mr. Cohagen of Iowa, has had excellent results in feeding this plant to stock. Its protein content is equal to that of alfalfa. It is probable that some forms are entirely inert. Some years ago, the writer conducted an experiment in feeding considerable quantities of sweet clover, but without any injurious symptoms resulting. A tincture prepared by mixing the fresh flowers with alcohol has a vanilla-like odor, and a bitter taste. Dr. Millspaugh states that in large doses, cumarin causes nausea, vomiting, vertigo, great depression of the heart's action, and cold extremities. Dr. Schaffner states that both of the sweet clovers are objectionable in wheat, because of the foul odor the seed imparts to the flowers. According to Friedberger and Fröhner sweet clover causes paralysis of the muscles. Dr. MacOwen states that in New South Wales, the M. indica is said to cause paralysis of horses.

10. Medicago L. Medick, Alfalfa

Herbs with pinnately 3-foliolate leaves; leaflets dentate toothed; flowers small, yellow or violet in axillary racemes or heads; calyx teeth short nearly equal; standard obovate or oblong; stamens diadelphous; ovary 1-ovuled; pod curved or spirally twisted, indehiscent 1-few seeded. About 50 species native to Europe and Asia. Bur clover (Medicago hispida) and hop clover or black

medick (M. lupulina) are used as forage plants on the Pacific coast but east-ward are regarded as troublesome weeds, the former injurious to wool.

Medicago sativa L. Alfalfa

An upright, smooth perennial; leaflets obovate, oblong, toothed, obtuse emaginate or mucronate; stipules entire; flowers in a short raceme violet; pod spirally twisted. A valuable forage plant.

Distribution. Common in the irrigated districts of the West, also frequent eastward, but common southward; spontaneous from New England to Minnesota, Kansas, northward and westward; native to Europe and Asia.

Poisonous properties. A large amount of the green fodder is said to produce tympanites, but alfalfa is, however, one of the best of forage plants.

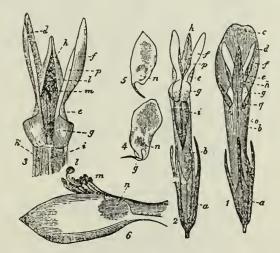


Fig. 303. Flowers of Red Clover. 1, a, Calyx. c, Standard. 2, f, Wings, h, keel, 4, 5, Wings, 6, keel.

11. Trifolium (Tourn.) L. Clover

Herbs; leaves mostly 3-foliolate, palmately or pinnately; stipules united with the petioles; leaflets usually toothed; flowers in dense heads or spikes; calyx persistent; lobes 5, nearly equal, corolla withering or persistent, claws alternate to the stamen tube; stamens diadelphous or the tenth one separated for a part of its length; pods small and membranous, indehiscent or dehiscent, 1-6 sceded.

A large genus of about 250 species mostly in the northern hemisphere. Many are valuable forage plants, among these are red clover (*T. pratense*), alsike clover (*T. hybridum*) and white clover (*T. repens*). Several are weedy as yellow hop clover (*T. agrarium*), low hop clover (*T. procumbens*), and stone clover (*T. arvense*).

The alsike clover (T. hybridum) and red clover (T. pratense) occasionally produce bloat.

Dr. Jacob Moses and A. M. Harcourt have recently described a disease sometimes caused by alsike clover.¹

¹ Bull. Tenn. Agr. Exp. St. 18:28 (1905).



Fig. 304. Alfalfa (Medicago sativa). a, b, seed pod; c, seed. An excellent forage plant, sometimes causes bloat. (U. S. Dept. Agr.)

"The cause of this trouble among horses and mules is not positively understood. Whether the toxic effect is due to the plant itself, which possibly undergoes some change within the digestive tract and subsequently liberates a poison, or whether it is due to the presence of a mould in connection with alsike clover, is still undetermined. The mould has been strongly suspected. To determine this point will require further investigation. It is known, however, that the principle lesions are produced on the skin and mucous membranes.

"The symptoms of this disease vary to some extent, depending upon the location of the lesions and the length of time the animal remains on the alsike pasture after the symptoms begin to develop. The cases which came under observation in Marshall County showed marked similarity of symptoms, involving principally the skin, the mucous membranes of the mouth, and the eyes.

"The prevailing symptoms of the disease are as follows:

On the skin are inflamed areas, appearing at first as more or less rounded vesicular swellings, varying from one-half inch to five or six inches, or more, in diameter. The hair over the affected areas stands erect, and has a dull appearance, indicating loss of vitality. Later the skin becomes hard and puffed out, as the result of the formation of pus underneath. Finally, the deadened skin is cast off, leaving a deep, raw, angry-looking ulcer, which eventually heals, with the formation of a conspicuous scar, covered with more or less white hair. These changes in the skin may occur on any part of the animal,

but especially on the limbs, body and croup. The eye symptoms consist of a marked conjunctivitis, with swelling of the eyelids, sensitiveness to light, and a watery discharge from one or both eyes. The mucous membranes of the mouth become inflamed (stomatitis), ulcers form, and the animal slobbers and refuses to eat. The advanced cases are frequently accompanied by emaciation. The tongue is usually affected, and the inflammation may extend throughout the entire digestive tract. The functions of the liver may be disturbed, and a yellowish (jaundice) coloration of the tissues follows. In such cases symptoms of colic are not uncommon, and the respiratory tract may become involved and pneumonia develop. Some observers in other countries have noticed marked nervous symptoms, such as excitement, convulsive movements, staggering gait, and paralysis of the throat, with inability to swallow, the paralysis at times becoming generalized, the animal getting down and being unable to rise. In the cases observed in this state, the nervous symptoms, except the general depression, were not very noticeable.

"The outcome of the disease depends upon the location and extent of the lesions upon the horse or mule affected. If they are situated on the exterior the animal will readily recover as soon as removed from the alsike pasture. If the vital organs are involved, such as the brain, lungs and liver, the disease may readily produce death. Among those cases occuring in this State, not a single fatality has been heard of at the Station. But even though the death rate is small where the ordinary precautions are taken, the disease



Fig. 305. Red Clover (Trifolium pratense). Occasionally the cause of bloat. (U. S. Dept. Agr.)

has considerable economic importance, since it leaves the animal more or less disfigured by the formation of scars, which materially depreciate his market value.

"The treatment is comparatively simple. As soon as the disease is recognized the animal should be removed from the alsike clover pasture and the wounds subjected to ordinary antiseptic treatment, such as frequent washing with 5 per cent solutions of carbolic acid or creolin, and the application to the ulcers on the skin of drying powders, consisting of boric and tannic acids in equal amounts."

The so-called clover sickness is supposed to be caused by the clover rust which has been described elsewhere. No doubt some of the trouble arising from feeding clover hay is caused by moulds found on the hay. Dr. W. D. Gilchrist says that he has observed several cases of the kind in this state. The animals showed extreme restlessness followed by coma, bloody discharge from faeces followed by diarrhoea, weakness and debility. Change in fodder caused the trouble to cease.

I have recently received a similar complaint from Dr. C. J. Scott of Knox-ville, Iowa, three animals having succumbed.

Trifolium incarnatum L. Crimson Clover

A soft pubescent, slightly branched, annual; leaves long petioled; broad stipules; leaflets nearly sessile, obovate or obcordate cuneate at the base,



Fig. 306. Crimson Clover (Trifolium incarnatum). Sometimes
produces phytobezoars, which may
cause death. (U. S.
Dept. Agr.)

denticulate; tlowers in elongated, oblong or ovoid, heads, sessile; calyx hairy, lobes plumose pointed, corolla crimson.

Distribution. Used as a cover crop and a forage plant in the south and east. Found on ballast from Maine to Pennsylvania. Native to Europe.

Injurious properties. According to Prof. Coville it produces phytobezoars and ocasionally causes death in animals.

Trifolium repens L. White Clover

A smooth perennial with slender creeping and spreading stems; leaflets inversely heart-shaped or notched, obscurely toothed; stipules narrow; peduncles very long, flowers in small loose heads reflexed when old; calyx shorter than the white corolla; pods 4-seeded.

Distribution. In fields and waste places throughout eastern North America, the Northwest and the Rocky Mountains.

Poisonous properties. Said to cause tympanites in cattle and slobbering in horses.

Psoralea L. Psoralea

Perennial herbs, usually sprinkled with glandular dots; leaves generally 3-5 foliolate; flowers spiked or racemed, white or mostly bluish-purple; calyx 5-cleft, persistent; stamens diadelphous or occasionally monadelphous; pods about as long as the calyx, 1-seeded.

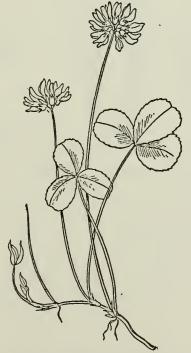


Fig. 307. White Clover (Trifolium repens). Occasionally the cause of tympanites in cattle. (Lamson-Scribner, U. S. Dept. Agr.)

About 100 species of wide distribution, many native to the plains, the roots of some being tuberous and farinaceous. The Indians used the tuberous roots of the *P. esculenta*, known as Pomme Blanche, or Pomme de Prairie, of the voyageurs for food. The roots of *P. hypogaea* and *P. cuspidata* were also used.

Psoralea argophylla Pursh. Silver-leaf Psoralea

Densely silvery, pubescent with appressed hairs; stem zig-zag, divergently branching, from 1-3 feet high; leaflets elliptical-lanceolate; flowers spicate, interrupted, blue; pod oval, membranaceous, plant seldom seeding.

Distribution. From Wisconsin to Kansas and New Mexico to the Northwest territory.

Poisonous properties. This plant was suspected of being the cause of a severe case of poisoning in Story County, Ia., two years ago. This is the first time the writer has known plants of this genus to cause poisoning, but he has had some correspondence with the parties concerned and thinks there can be no doubt that the poisoning was caused by the plant in question. A child was seriously poisoned by eating the seeds of this plant, but she finally recovered. It was thought this poisoning might have been caused by Astragalus caryocarpus but the plant sent me was the above.

13. Tephrosia Pers. Hoary Pea

Herbs or somewhat shrubby plants; odd-pinnate compound leaves; flowers in racemes or short clusters, red or white; stipules small; calyx 5-cleft; petals 5; standard roundish, usually silky outside, turned back, about as long as the coherent wings and keel. About 120 species, native of warm and tropical regions, a few are found in the United States.

Tephrosia virginiana Pers. Goat's Rue. Catgut

Perennial with villous or silky and whitish hairs; stem erect and simple, 1-2 feet high; leaflets 17-25, linear-oblong; terminal oblanceolate, narrowed to cuneate at the base; emarginate at the apex; flowers yellowish purple in long, dense racemes.

Distribution. In dry and sandy soil from Maine to Louisiana, west to Minnesota and Eastern Iowa, to Mexico.

Poisonous properties. This species, along with others, was formerly used to poison fish. The Mexican species, T. toxicaria, gets its name from its supposed toxic properties, and in South America, one species is commonly employed by the natives to poison the fishing streams.

The root is also poisonous to frogs and guinea pigs.

Tephrosia toxicaria contains the glucoside tephrosin. Several active substances have been obtained by Hanriot* from one species, Tephrosia Vogelii. Three substances were isolated; one, tephrosal $C_{10}H_{10}O$ is toxic especially to fish; a second toxic substance is tephrosin $C_{21}H_{20}O_{10}$.

14. Robinia L. Locust Tree

Trees or shrubs; stipules often prickly or spiny; leaves compound, oddpinnate; the oblong leaflets with short stipules; flowers showy in axillary race-

^{*} Compt. Rend. 1907; 498-651. Journal Chemical Society Abs. 92:386, 1907.

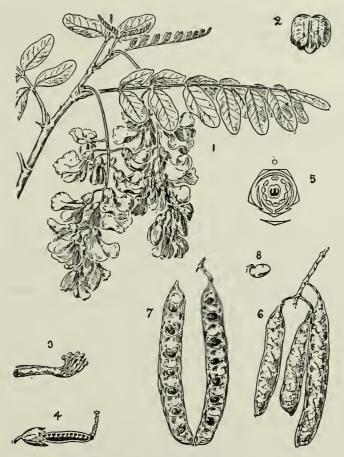


Fig. 308. Black Locust (Robinia Pseud-acacia). 1. Flowering branch. 2. Flower. 3. Tube of stamens. 4. Longitudinal section of pistil. 5. Diagram of flower. 6. Legumes. 7. Pod open, showing seed. 8. Seed. 1, 2, 6, 7, 8, one-half natural size. (M. M. Cheney in Green's Forestry of Minnesota.)

mes; calyx short, 5-toothed, and slightly 2-lobed; standard large, about as long as the wings and equal; stamens diadelphous; pod flat, several seeded and margined. A small genus of 6 species, native of North America and Mexico. Several species are cultivated for ornamental purposes, like the Robinia viscosa, which is native from Virginia to North Carolina and Georgia, and the Robinia hispidia, native to the mountains of Virginia and Georgia. The R. neo-mexicana, with purple flowers, native to S. Colorado and New Mexico, is frequently cultivated.

Robinia Pseudo-acacia L. Locust Tree, False Acacia or Black Locust

A large tree with rough bark, spiny stipules; 9-19 stalked leaflets, obtuse, emarginate, or mucronate; flowers in loose drooping racemes, white, fragrant; pods smooth; standard yellowish at the base.

Distribution. Widely planted as an ornamental tree. It produces valuable timber which is extensively used for posts. This species is, however, badly infested with the borer.

Robinia viscosa Vent. Clammy Locust

A small tree with rough bark; stipules short, occasionally spiny; twigs and petioles glandular; leaves 11-25, stalked; leaflets obtuse and mucronate; nearly smooth; racemes dense; flowers in rather dense racemes, pinkish, not fragrant; pedicels glandular, hispid; pod hispid.

Distribution. Southwestern Virginia to Georgia, occasionally escaped from cultivation northward.

Poisonous properties. The bark and leaves of this species contain a powerful poison which has proved fatal to persons eating them. Children have been poisoned by eating the roots. It is true, however, that the flowers of the plant are often eaten with impunity and that bees collect from them large quantities of nectar. Dr. Rusby states that the occasional poisonous properties of honey are due to its origin in these flowers, though there are gool theoretical reasons for doubting this. The bark of young twigs is sometimes pounded to a pulp, and used to make a tincture which is used in medicine as a tonic and cathartic, while the medicinal use of the flowers is mildly narcotic. It contains the substance robinin C₈₈H₄₀O₁₉ an aromatic glucoside which resembles the glucoside quercetrin, and is found chiefly in flowers, also the substance obigenin C15 H10O8 +H,O. The seeds are also poisonous, and Dr. Millspaugh quotes Dr. Shaw as follows, in regard to the symptoms produced by poisoning from eating the seeds: "Inability to hold the head upright, nausea and attempts to vomit, with a tendency to syncope, when in an upright position; voice, respiration and heart's action feeble, as from exhaustion, a painful paralytic condition of the extremities, which become shrunken on the fifth day. All the symptoms seemed like those produced by a long-continued diarrhoea, though in this case purging was not present." Dr. Johnson states that the symptoms of poisoning are those of Belladonna poisoning, a fact also noted by Dr. Waldron in the case of a horse that had eaten the bark; Friedberger and Fröhner state that the animals have colic, tympanites and paralysis.

Dr. Rusby comments upon the poisonous character of the common black locust as follows:

Of this Dr. Johnson records that by eating the roots children are poisoned with symptoms like those of Belladonia poisoning, and that the bark and leaves are emetic. Prof. F. W. Power has experimented upon himself with the stem bark of this tree, proving the very serious effects which it produces, and he has examined its composition with the result of showing that the poisonous constituent is an albuminous substance, thus confirming the general character of that family, the Leguminosae. The most positive and prominent case recorded in regard to this article is that of Dr. Z. P. Emery. In the latter part of March, 1887, thirty-two boys, inmates of the Brooklyn Orphan Asylum, were poisoned at one time by eating a bark which was being stripped in the vicinity for the making of fence posts. None of the cases terminated fatally. The prominent symptoms, stated in the order of their occurrence, were the vomiting of a ropy mucous, flushing of the face, dilated pupil, dryness of the throat, fceble pulse, extremities cool, face pale, vomiting of blood, cold extremities, heart feeble and intermittent, face deathly pale and stupor. The symptoms as I have named them are seen to be progressive. A rash similar to that of Belladonna poisoning was also present, but very fleeting. In the beginning there was a high fever. Treatment consisted of sinapisms over the stomach, subcarbonate of bismuth, camphor and brandy.

A farmer in Dallas County, this state, informs me that sometimes the leaves are macerated in water and used to kill flies.

A case of poisoning to a horse was recorded in *Breeders' Gazette* in 1909. In this case the horse had eaten some of the bark of a tree. The symptoms were similar to those recorded by Dr. Waldron.

15. Sesbania. Scop. Sesban

Tall, smooth, branching herbs or shrubs with pinnate leaves and yellow flowers in axillary or compound racemes; calyx bell-shaped, obliquely truncate, 5-toothed; standard short, orbicular; wings oblong; keel blunt; stamens diadelphous; style short, incurved at the apex; legume oblong, stalked, compressed, the endocarp membranaceous, at length separating from the coriaceous epicarp and enclosing 2 seeds. A small genus of 15 species of warm or temperate regions.

Sesbania platycarpa Pers.

A tall, smooth, branching annual vine; leaflets 10-35 pairs, mucronate, pale beneath; racemes shorter than the leaves; corolla yellowish purple spotted, with membranaceous sacked pods.

Distribution. From the Carolinas to Florida, Missouri, and Texas.

Poisonous nature. In 1897, Dr. A. P. Anderson sent this to the writer with a letter from some stockmen from South Carolina, who stated that it was suspected of poisoning his cattle. Mr. Chesnut records a similar statement as follows:

In 1897, the United States Department of Agriculture received from South Carolina the seeds of this plant, which were found in the stomachs of cows.

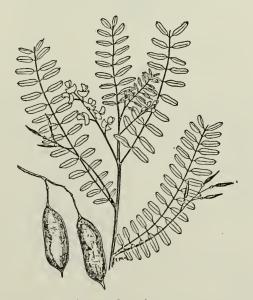


Fig. 309. Sesban (Sesbania platycarpa). This plant is common in the southern states; found along roadsides and woods; known to be poisonous. (Charlotte M. King.)



Fig. 310. Loco Weed (Astragalus mollissimus). Charlotte M. King

16. Astragalus L. Milk Vetch

Chiefly perennial herbs with odd pinnate or occasionally simple leaves, with stipules; flowers spicate or racemose; calyx 5-toothed, tubular; corolla clawed, usually long and narrow; standard erect, wings oblong; keel nearly as long as the wings; stamens diadelphous; pod several to many-seeded, dehiscent or indehiscent; 1-2 celled. A large genus of about 1000 species of wide distribution in the United States, largely western. Several species are used in medicine. One plant is widely known as the loco weed. One species native to gravelly knolls or dry places of the Mississippi Valley is known as the ground plum or hog pea (A. caryocarpus). This species is said to be edible. It should, however, be used with caution.

Astragalus mollissimus Torr. Loco Weed

A stout, short-stemmed perennial with membraneous stipules; leaflets 19-27, ovate oblong; flowers in dense spikes, violet purple, rather large; pod oblong, glabrous somewhat compressed, sessile, furrowed at both sutures, at length curved.

Distribution. Found on plains of Nebraska, Kansas, Colorado to Texas and New Mexico.



Fig. 311. Ground Plum (Astragalus caryocarpus). It is not known whether this species is poisonous or not, although some of the related species are known to be poisonous and belong to the loco weeds. It is said that the pods of this species are edible. (Ada Hayden.)

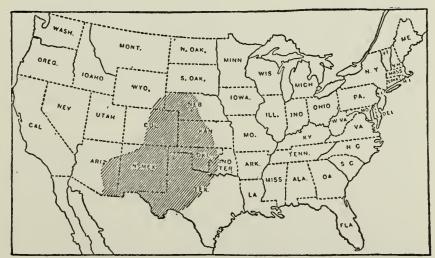


Fig. 311a. Map showing distribution of Loco Weed or Purple Loco Weed (Astrogalus mollissimus). After Marsh, U. S. Dept. Agr.

The term loco weed is applied to a great many plants. In addition to the species mentioned above A. oocarpus, A. Crotalaria and A. lentiginosus must be added; species of Oxytropis are considered in another connection. The Sophora sericea of this family, Malvastrum coccineum of the Mallow family and corydalis (Corydalis aurea var. occidentale) have at times been classed as loco weeds. Thus far none of these plants except those belonging to the pulse family have been found to cause loco poisoning.

Astragalus Bigelovii Gray. Rattle-box

A subacaulescent, soft, silky, villous perennial; long, scape-like peduncles; flowers in dense spikes; violet pod oval-oblong, densely woolly, sulcate.

Distribution. From S. W. Colorado to Texas and Mexico.

Astragalus Pattersoni A. Gray

Perennial, robust, 1-2 feet high, with appressed-pubescence, or sometimes smooth; leaflets oblong, rather thick; peduncles racemosely many-flowered; corolla white, the keel sometimes purplish at the tip; pods smooth, sessile or stipitate; abruptly contracted within the calyx.

Distribution. Southwestern Colorado to Utah.

Astragalus Hornii A. Gray

A glabrous or minutely pubescent perennial; slender stems; leaflets about 21, narrowly oblong; peduncles longer than the leaves; flowers in a dense head, or short spike, numerous, small; calyx teeth subulate; pods ovate, from a broad base, straight, villous pubescent.

Distribution. From Southern Utah to California.

Poisonous properties. It is regarded as a very troublesome weed. Colorado has passed a law for its extermination, the state having paid out nearly \$200,000 in bounties between 1881 and 1885 to check its ravages. Much has been written on the subject of the poisonous properties of the woolly loco weed and other members of the genus. Brewer and Watson, in their Botany of California, state that the last species described is said to be poisonous to sheep, and Prof. Chesnut states that stock are affected by this loco weed in the southern part of California. It has certainly been regarded as a poisonous plant for a long time, and numerous investigations have been carried on in regard to its poisonous properties by Dr. Stalker, Prof. Sayre and others; Dr. Stalker coming to the conclusion that the loco poisoning might be brought about through the action of intestinal worms.

Prof. Sayre reported the death of a jack rabbit with symptoms similar to those recorded for horses and cattle. He was, however, unable to find a poisonous principle. Miss C. M. Watson reported a small amount of an alkaloid from the stemless loco weed, Oxytropis Lamberti. Dr. Mary Gage Day in Dr. Vaughn's laboratory found that when a half grown kitten was fed with milk containing a decoction of the root, stem and leaf, emaciation, convulsive excitement, and, finally death occurred; when an adult cat received 60-70 cc. of a more concentrated solution death occurred on the thirteenth day and subcutaneous injection of a concentrated decoction in frogs and chickens caused death in 1-2 hours from heart paralysis. Dr. J. Ott experimented with Astragalus mollissimus and found that it decreased irritability of the motor nerves

affected the sensory ganglia of the nervous system, preventing them from readily receiving impressions and killed by arresting the heart action.

Dr. B. T. Galloway states that the loss from "loco" poisoning in Colorado alone has reached the sum of one million dollars per annum. It might be said in this connection that Prof. Power and Mr. Gambier investigated the subject, but were unable to locate definitely the kind of alkaloid. These gentlemen state:

One kilogramme of the Astragalus berb gave 0.2 of a gramme of the alkaloid, equivalent to 0.006 per cent. Nothing further was determined concerning its nature, as it does not appear to be especially active. An extract from one kilogramme of the seed of the Crotalaria gave 1.1 grammes of an alkaloid, 0.036 per cent of the weight of the seed. It bad a bitter taste, and seems to be more potent than that obtained from Astragalus.

From these investigations these authors conclude that both the Astragalus and the Crotalaria contain very small amounts of toxic alkaloids, to which the symptoms of poisoning may be reasonably attributed. Prof. Sayre, who has not gone into the details in the paper referred to, however, reiterates what he has stated in several previous ones, that it is a question whether so small amount of alkaloid could produce such grave physiological disturbances.

The symptoms of poisoning are very well given in a paper by Prof. Chesnut: Horses, cattle, and sheep are affected by loco, but the principal damage is done to horses. The effect is not acute, but in its slow progress stimulates diseases caused by bacteria, worms, or other parasites or such as are caused in man by the continued use of alcohol, tobacco, or morphine. Two stages are recognized. The first, which may last several months, is a period of hallucination or mania accompanied by defective eyesight, during which the animal may perform all sorts of antics. After acquiring a taste for the plant it refuses every other kind of food, and the second stage is ushered in. This is a lingering period of emaciation, characterized by sunken eyeballs, lusterless hair, and feeble movements. The animal dies as if from starvation, in periods ranging from a few months to one or two years.

Dr. Carl Ruedi isolated an acid (loco acid) from it to which he attributed the poisonous qualities of the plant.

Astragalus is said to cause considerable trouble in Nebraska. "In regard to the treatment," says Mr. O'Gara,

There is little to be said. All medicines that have been tried seem to have been of doubtful effect. More can be done by keeping animals away from loco than in any other way. As long as there is a plentiful supply of grass, there is little to be feared, but when pastures and ranges run low, stock should be closely watched. At the very first appearance of trouble, affected animals should be removed to some place where they cannot gain access to loco. Good nourishing food should be given. If the disease bas obtained a strong hold on the animal there is little hope of recovery under the best of treatment, hence the need of early treatment and pro upt removal from the source of the trouble.

Dr. Mayo suggests the following treatment:

| Ounc | CES |
|-------------------------------|-----|
| "Sulphate of iron, pulverized | 1 |
| Gentian root, pulverized | 4 |
| Ammonium chloride, pulverized | 1 |
| Potassium nitrate, pulverized | 1 |

Mix thoroughly and give from a heaping teaspoonful to a tablespoonful, according to the size of the animal, in the food three times daily."

Dr. Dwight C. Marsh and Albert C. Crawford under the direction of Dr. R. H. True who has charge of the Poisonous Plant Investigation of the U. S. Department of Agriculture, have arrived at conclusions wholly at variance with previous investigation. They conclude that Oxytropis Lamberti poisons horses, sheep, and cattle and that Astragalus mollissimus poisons horses, but does not

poison cattle because they rarely eat it. Dr. Marsh states that the symptoms described by stockmen were corroborated; these are:

The lowered head, rough coat, slow, staggering gait, movements showing lack of muscular coördination, sometimes more or less paralytic symptoms, a generally diseased nervous system, and in the later stages of the disease extreme emaciation.

The principal pathological changes are pronounced anemia of the whole system, diseased stomach walls, and in acute cases a congested condition of the walls of the stomach; while in chronic cases there are frequently ulcers. Generally speaking, locoed cattle have ulcers in the fourth stomach. There is an excess of fluids in the various cavities of the body. This is especially noticeable in the epidural space of the spinal canal. Here the effusion is more or less organized, presenting the appearance of a gelatinous mass, which is especially abundant in the lumbar region and about the exits of the spinal nerves. In most locoed females the ovaries are found in a diseased condition.

Dr. Crawford from his laboratory work concludes:

The symptoms described in stock on the range can be reproduced in rabbits by feeding extracts of certain loco plants. Those especially referred to here under the term "loco plants" are Astragalus mollissimus and Astragalus Lamberti (Oxytropus Lamberti).

The production of chronic symptoms in rabbits is a crucial test of the pharmacological activity of these plants.

It is the inorganic constituents, especially barium, which are responsible for this poisonous action, at least in the plants collected at Hugo, Colo.; but, perhaps, in the future, loco plants from other portions of the country may be found to have other poisonous principles.

There is a close analogy between the clinical symptoms and pathological findings in barium poisoning and those resulting from feeding extracts of certain of these plants. Small doses of barium salts may be administered to rabbits without apparent effect, but suddenly acute symptoms set in analogous to those reported on the ranges.

The administration of sulphates, especially epsom salts, to form insoluble barium sulphate would be the chemical antidote which would logically be inferred from the laboratory work, but of necessity these sulphates would have to be frequently administered, and their value, after histological changes in the organs have occurred, remains to be settled. But the treatment of acute cases of barium poisoning in man is not always successful, even when sulphates combined with symptomatic treatment are employed. The conditions under which sulphates fail to precipitate barium must be considered.

I,oco plants grown on certain soils are inactive, pharmacologically and contain no barium. In drying certain loco plants the barium apparently is rendered insoluble, so that it is not extracted by water, but can be extracted by digestion with the digestive ferments. To be poisonous the barium must be in such a form that it it can be absorbed by the gastro-intestinal tract.

There are plants with barium salts which are not injurious.

Dr. Marsh found that it is easy to kill the woolly loco weed (A. mollissimus) in fenced pastures because it occurs in small patches. The stemless loco weed is of wider distribution and, when in fenced pastures, can be killed but not so easily when it occurs on the ranges. He recommends treatment as follows:

In regard to the second phase of remedial work, it was found that locoed cattle can in most cases be cured by a course of treatment with strychnin, while locoed horses can generally be cured by a course of treatment with Fowler's solution. The animals under treatment must not be allowed to eat the loco weed and should be given not only nutritions food but, so far as possible, food with laxative properties. To this end, magnesium sulphate was administered to correct the constipation which is almost universal among locoed animals. It should be noted, too, that magnesium sulphate may serve to some extent as an antidote to the poison.

Dr. C. Dwight Marsh 1 in a recent publication in speaking of this plant as well as the White Loco Weed and the experimental work of the Department of Agriculture, states that they are the weeds which produce the disease from Montana to northern New Mexico, Arizona and in western Texas, but there are many locoed animals where these two species do not grow. In California,

[&]quot;The Loco-Weed disease. Farmers' Bull., U. S. Dept. Agr., 380. 1909.

Arizona and New Mexico there occur other leguminous plants which are known as loco weeds. Some of these are poisonous and the symptoms of locoed animals and the pathological findings are similar to those produced by the plants found in the eastern Rockies, especially in Colorado, where the purple loco is abundant. Dr. Crawford finds barium in them which he thinks may be connected with the poisonous effects of the plants. In California the A. diphysus, A. arizonicus, A. Thurberi and A. Bigelovii are called "rattleweeds" and have been suspected. They are being studied by Dr. Crawford of the Bureau of Plant Industry.

In his most recent paper on Loco Poisoning* Dr. Marsh affirms Dr. Crawford's opinion that the purple loco weed (Astragalus mollisimus) is more poisonous than the white species (Oxytropis Lamberti). Dr. Marsh adds this caution, that since such animals are extremely sensitive to strychnin, it has been found necessary to give it in small doses. He says:

The daily doses should not ordinarily exceed three-twentieths or four-twentieths of a grain, or 0.009 to 0.012 of a gram. Large animals may take as much as one-half grain, but this is a maximum dose and often will be found too much. It was also found that sodium cacodylate when given to cattle in hypodermic injections of 6 grains, or 0.4 grains, daily, commonly gave beneficial results. The best results, however, were obtained from the use of strychnin and Fowler's solution as already outlined.

It may be added, in regard to the question of immunity, that loco poisoning comes on in a slow and cumulative manner so that there is no possibility of animals becoming immune.

It does not seem that the above is final in regard to all of the loco weeds. It is hardly likely that Profs. Power, Sayre, Gambier, and others have been entirely wrong in regard to their conclusions. When we find that related plants have strongly toxic properties we may expect to find the same properties also in some of these plants.

17. Oxytropis DC. Stemless Loco Weed

Perennial herbs or sometimes shrubby, generally acaulescent, with numerous tufts of short stems covered with scaly stipules. Flowers in racemes or spikes; calyx teeth nearly equal; petals clawed; standard, erect, keel erect; stamens appendaged; diadelphous; pod more or less 2-celled or 1-celled sessile or stalked. About 125 species of the North Temperate regions. Most of our species are western. Several of our species are known to be poisonous to live stock. The following species of this genus are classed with the loco weeds: O. Lamberti, O. deflexa and O. multiflorus. According to Greshoff the young leaf of Oxytropis lapponica has an extremely bitter taste; there is also an indication of saponin. On analysis the leaf was found to contain hydrocyanic acid. Greshoff also found hydrocyanic acid in the seeds of O. sulphurea.

Oxytropis Lamberti Pursh. Purple or Stemless Loco Weed

Nearly acaulescent perennial herbs or shrubby plants, with tufts of very numerous short stems coming from a hard and thick rootstock containing many scaly stipules; stems and leaves are covered with silky and fine appressed hairs, or smooth; leaves pinnate; leaflets linear; flowers racemose or spicate, rather large and elongated, purple, violet, or sometimes white; stamens diadelphous; keel tipped with a sharp projecting point.

^{*} Farmers Bull. U. S. Dept. Agr. 380:16.

Distribution. Western Minnesota, Western Iowa, and Missouri to Texas, and New Mexico, north to British Columbia, and northwest territory.

Poisonous properties. The stemless loco weed is one of the most characteristic loco weeds of the West. The symptoms of poisoning are similar to those produced by the woolly loco weed described at The elsewhere. poisonous substance has not been isolated. An alkaloid, however, has been reported by Prof. Prescott. Chesnut and Wilcox, in speaking of the history of the loco poisoning in Montana, say that in Colorado the plant which is most commonly known as loco weed is Astragalus mollissimus. In Montana, on the other hand, the plants most generally called loco weeds by the stockmen are species of Aragallus (Orytropis). "The species which is most concerned in causing the loco disease in Montana is the Aragallus spicatus and is closely related to Oxytropis Lamberti," Stockmen are of the opinion that a condition somewhat similar to loco poisoning may be brought about by eating undue quantities of alkali soil.



Fig. 313. Stemless or Purple Loco Weed a, Plant. b, Seed pods. c, Seed. U. S. Dept. Agr.

It should be stated also that the larvae of sheep bot flies, which are frequently found in the frontal sinuses of the head, can not possibly be considered the cause of the nervous symptoms characteristic of the loco disease, for the reason that these larvae are not found in greater abundance in locoed than in healthy sheep. For the same reason the presence of the common tape worm (Taenia serrata) in the small intestines and bile duct of sheep can not be considered as the cause of the locoed condition. These worms are almost

universally present in the intestines of sheep, and under ordinary conditions do not cause any recognizable disturbances. The disease of sheep known as "gid" is not to be mistaken for the loco disease and, furthermore, is not prevalent in this country. No indications were found during the post-mortem examinations that the walls of the stomach were affected to any appreciable extent by the action of loco weeds, although these plants were invariably found in the stomach contents of such sheep. In the majority of cases no apparent changes have been produced in the spleen, liver, or kidneys. In some instances a slight congestion of the intestines was noticed. The cerebral membranes were in all cases somewhat congested. This condition is probably one of the immediate physical causes of the mental excitement exhibited by locoed animals. Post-mortem examinations of locoed horses disclosed the same conditions as those found in the sheep.

They made a number of experiments with a young Belgian hare and other rabbits, using the water extract of the leaves, and it was shown that this was not an acute poison if from 10 to 15 cubic centimeters of the liquid were administered. An acute case of loco disease was observed in an old ewe with a lamb at her side. She had eaten eonsiderable quantities of the white loco weed (Aragalus spicatus-Orytropis spicatus). A slight locomotor ataxia was manifested. The eyelids twitched rapidly and there was a slight champing of the jaws. Each attack lasted from 1 to 2 minutes, and the intervals between the attacks were about 5 minutes.

The lips and eyelids twitched violently and the jaws were moved upon one another with such force that the sound could be heard for a distance of 200 yards.

Similar symptoms were observed in the lamb, which died in the afternoon.

Locoed sheep are exceedingly difficult to herd.

It is the universal experience of sheep raisers that locoed sheep are exceedingly difficult to herd. The sheep may, without a moment's warning, stray away from the band, each one in a different direction, and it is easy to understand how nearly impossible it is to prevent such a band of sheep from becoming separated. Besides giving the herder much trouble in directing the course of the band on the range, locoed sheep alse refuse to enter the corral at night, and under any and all circumstances may suddenly manifest perplexing stubbornness.

These writers did not observe many locoed cattle, but the symptoms are essentially the same as in sheep and horses. In regard to post-mortem conditions, they say:

Numerous autopsies made on locoed sheep and horses revealed conditions which, though fairly uniform, did not constitute a well-defined series. We made a large number of post-mortem examinations upon bodies of locoed sheep which had been killed and bled inunediately before examination. In these cases there was no lesion or marked change in the alimentray tract. A slight congestion of the membranes of the brain was to be observed in all cases. The lungs and heart were apparently not affected. The voluntary muscles were of a paler color than under normal conditions, and the fat tissue was considerably reduced in quantity.

As to remedies, the following suggestions have been made:

Locoed sheep should be removed from the band and fattened for market on alfalfa or other forage plants, as above explained.

The immediate isolation of locoed sheep is advisable in order to prevent the habit from spreading in the band.

It seems desirable to give sheep a regular and abundant supply of salt in order to prevent the development of any perversion of the appetite.

Lococd horses are used to the best advantage as draft animals, but they must be maintained in good condition and prevented from eating loco weeds.

Dr. Marsh recommends to cut the roots below the crown of buds. A man with a spade can destroy a large number of plants in a day. The seeds however retain their vitality for some years, hence the field will have to be gone over again. That this method will effectively destroy the plants has been demonstrated by the U. S. Dept. of Agr. at Hugo, Colorado. The larvae of a moth (Walshia amorphella) feeds on the purple loco weed and this insect, Dr.

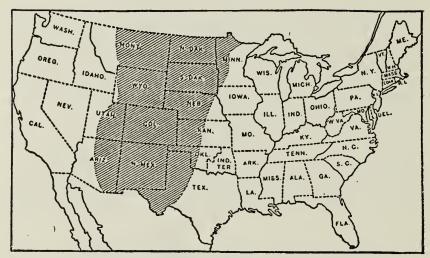


Fig. 313a. Map showing distribution of stemless loco weed (Oxytropis Lamberti). The plant reaches western Iowa on the loess bluffs where it is abundant. The disease however has never been reported so far as I know from this section of the state. Map after Marsh, U. S. Dept. Agr.

Marsh thinks, will help to keep the weed in check if the insect will not lose its efficiency in the course of a few years.

18. Vicia (Tourn.) L. Vetch or Tare

Herbs, generally of trailing or climbing habit, with pinnate tendril bearing leaves; flowers generally racemose; calyx 5-cleft or 5-toothed, divisions nearly equal; corolla with the standard clawed and the wings adherent to the keel; stamens diadelphous or monadelphous; pod flat, 2-valved with several seeds; seeds globular; embryo with thick cotyledons. About 120 species, widely distributed. Some species used for forage, especially in Europe. The hairy vetch (V. villosa) has been widely distributed in the west because of the drouth resisting qualities. Our most common native species is the American vetch (V. americana) which might well be introduced as a forage plant.

Vicia sativa L. Common Vetch

A smooth or slightly pubescent annual from $1-2\frac{1}{2}$ feet high with simple stem; leaflets 5-7 pairs, obovate-oblong to linear, notched or mucronate at the tip; the 1 or 2 nearly sessile flowers are borne in the axils of the leaves; flowers bluish purple; calyx teeth about as long as the tube; pod linear, several seeded, seeds black.

Distribution. From eastern Canada to Northwest Territory, New England to the Carolinas, west to Missouri and northward, generally in the wheat growing sections of the northern and western states. This is another weed commonly found in wheat screenings, abundant in the northwest.

Poisonous properties. In Europe it is the cause of tympanites. Dr. Schaffner, in The Ohio Naturalist, states that caution must be observed in feeding this plant to pigs. It is not injurious to cows. The seeds of this Vetch are often



Fig. 318b. Common Vetch. (Vicia sativa)



Fig. 314. Common Vetch (Lathyrus silvestris). The seed of this plant poisonous to stock. Charlotte M. King

found in screenings and fed in large quantities to cattle. As far as the writer knows, there are no cases of poisoning recorded from eating the screenings of this seed. The substance vicin $C_8H_{15}N_3O_6$ has been found in the seeds of this species. Convincin $C_{10}H_{15}N_3O_8+H_2O$ also occurs in this species and in V. Faba. Citric acid $C_8H_8O_7+H_2O$ is found in V. sativa.

19. Lathyrus (Tourn.) L. Vetchling. Everlasting Pea

Mostly perennial, herbaceous vines although there are a few erect herbs, generally smooth, with pinnate, usually tendril-bearing leaves; flowers in racemes or solitary; calyx oblique or gibbous at the base, upper teeth sometimes shorter than the lower; corolla larger than that of *Vicia*, wings adhering to keel; style dilated and rather flat above, hairy along side next to free stamen; stamens 10 (9 and 1, or monadelphous below); ovules numerous; pod flat, sometimes terete, 2-valved, continuous between the seeds, dehiscent.

About 100 species are distributed throughout North America and a few others are found in South America and the mountains of tropical Africa. One species, L. sylvestris, is considered poisonous, in its native home in the Carpathian Mountains. It contains certain alkaloids which, by the process of cultivation have become eliminated so that in many localities at the present time it is considered a good forage plant and is relished by horses. In the western United States, the prairie vetchlings L. ornatus and L. polymorphus, and the marsh vetchling L. palustris are considered valuable forage plants, the latter forming a very important part of the hay and adding materially to its feeding value. L. venosus and L. ochroleucus occurring in similar localities are much less valuable. A form of intoxication, known as Lathryism, is said to be caused by different species of Lathyrus.

In Dr. Wilson's "American Text-Book of Therapeutics," Victor C. Vaughan translates the following account of Lathyrism from Kobert's work "Intoxikationen:"

By Latbyrism we mean an intoxication that was known to the contemporaries of Hippocrates, and which was caused by the seeds of at least three species of vetch, Lathyrus hirsutus, the red vetch, Lathyrus sativus, the German vetch, and Lathyrus Clymenum, the Spanish vetch. In Spain, France, Italy, and in certain parts of Africa and India there bave repeatedly appeared, from the eating of the seed of the vetch, epidemics of a disease that especially affects males and which induces a transverse myelitis with motor and sensory paraplegia. The paralytic symptoms gradually disappear, but there remains spastic tubes with heightened tendon-reflexes attributed by Pronst to secondary degeneration of the lateral columns, while Strümpell considers the case a typical spinal paralysis. However, the symptoms may wholly disappear and recovery be apparently complete. Men and animals, especially horses, are affected in the same manner. Duvernoy described the disease in 1770; Doir saw it follow the eating of vetch-bread in 1785; Despranches observed it in France in 1829, and Pellicotti in the Abbruzzo mountains in 1847. Reports of the disease were made by Irving, in India, in 1861 and 1869, and by Bourlier in Africa in 1882. In 1883, Marie published in Le Progrès Medical a review of the literature of the subject and more recently Schuchardt has done the same. Hogs are killed quickly by the vetch. Horses suffer from paralysis of the recurrent laryngcal nerve, necessitating tracheotomy. More chronic poisoning causes paralysis of the posterior extremities, and death. Méricourt believes the disease beri-beri is due to a similar intoxication, but this is denied by Marie and others. In horses there is atrophy of the muscles of the larynx, especially of the cricoarytenoideus posticus and lateralis, also of the thyroarytenoideus. The left recurrent laryngeal nerve is much wasted. Microscopic examination shows the muscle greatly atrophied, without striation, and undergoing fatty degeneration. In the central nervous system one finds atrophy of the ganglion-cells in the vagus center and of the multipolar ganglion-cells in the anterior horns of the cord. Attempts to isolate the poison have not succeeded. Teilleux found an acid that induced typical effects upon rabbits. Bourlier found an active alkaloid in the alcohol-ether extract of the seeds, and poisoned birds with it. Astier isolated a volatile alkaloid by the Stas method, and he thus explains the fact that long-continued heating at a high temperature renders the seeds inert.

20. Cicer L. Chick pea

Calyx tube oblique or gibbous posteriorly; lobes nearly equal or the two upper somewhat shorter, conniving; standard ovate or nearly orbicular, narrowed into a broad claw; wings obliquely obovate, free; keel somewhat broader, incurved, dilated; anthers uniform; ovary sessile 2-8 ovuled; style filiform, incurved or bent, beardless; stigmas terminal, legume sessile, ovoid or oblong, turgid, 2-valved; seeds sub-globose or irregularly obovoid; funiculus scarcely dilated, hilum small; cotyledons thick; radicle short, slightly incurved or nearly straight.

Cicer arietinum L. Chick pea

Annual herbs, or perennial often glandular-pubescent; leaves pinnate, petiole terminating in a small tuft of spinescent hairs or in an odd leaflet; leaflets dentate or incised without stipels; stipules foliaceous oblique, often dentate or incised; flowers white, blue or violet; solitary pedunculate, or few pedicelled; bracts small; bractlets 0. About 14 species, especially in the eastern Mediterranean and in Central Asia—extending westward.

Distribution. Cultivated in the Rocky Mountains and in the Southwest. Also extensively in Southern Europe and tropical Asia. Considered an excellent food plant.

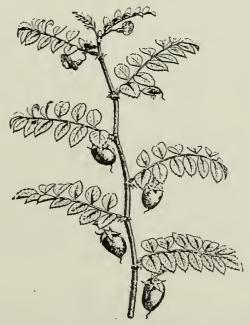


Fig. 315. Chick pea (Cicer arietinum). (After Faguet).

Poisonous properties. Friedberger and Fröhner in Veterinary Pathology give the symptoms from Cicer poisoning:

In horses roaring and difficult breathing, owing to paralysis of the laryngeal muscles; paralysis, weakness in the loins, suffocation. Post-mortem reveals nothing of moment. Therapeutics: Change of fodder; tracheotomy.

21. Phaseolus L. Bean

Usually vines with pinnately 3-foliate leaves, stipules and racemose, howers; calyx 5-toothed or 5-cleft, the upper teeth more or less united; standard orbicular recurved, spreading; keel spirally coiled enclosing the stamens and style; stamens diadelphous, 9 and 1; style bearded; pod linear 2-valved, several seeded; seeds with large embryo. About 170 species mostly of tropical regions; 12 species native to southern states. The common bean (Phaseolus vulgaris), native to tropical America is widely cultivated. The scarlet runner (P. multiflorus) is widely cultivated as an ornamental plant and is said to be poisonous. The P. Mungo is cultivated in the tropics.

Phaseolus lunatus L. Lima Bean

A twining plant with racemose flowers shorter than the leaf; pod broad and curved, scimitar shaped; seeds few, large and flat; some dwarf, some with long twining stems.

Distribution. Widely cultivated, native to South America.

Poisonous properties. Several cases of poisoning from the use of lima beans are reported. L. Guignard, according to an abstract in the Experiment Station Record, has determined this to be due to hydrocyanic acid. He says: "A number of forms that have been described as distinct species are by the author believed to be varieties or cultural forms of P. lunatus. Those principally studied were the white and colored Java beans, Burma or white Indian beans, Sieva beans, Cape beans, which are extensively cultivated in Madagascar, and Lima beans. These different varieties are widely cultivated and extensively used as food, although a number of fatalities have been attributed to their use. Descriptions of the different varieties and detailed reports of the chemical studies are given.

Practically all varieties of *P. lunatus*, whether wild or cultivated were found to contain the principle which when acted upon by an enzyme yields hydrocyanic acid. The proportion of hydrocyanic acid varied from almost inappreciable amounts in some of the more improved forms, like the Lima bean, to as much as 60 to 320 mg .per gm. dry weight in certain varieties of Java beans. It was found impossible by cooking to remove all the cyanogenetic compound in Java beans. Prolonged boiling extracts the greater part, but it is merely withdrawn and not destroyed, and if the water is absorbed it presents the same danger as the beans themselves, since either in the alimentary tract or in the blood are sufficient ferments to act upon the dissolved glucoside, resulting in the liberation of hydrocyanic acid.

GERANIALES

Herbs, shrubs or trees; petals usually present and generally polypetalous; sepals mostly distinct; stamens few, rarely more than twice as many as the sepals, opposite them when as many; compound ovary superior. Contains the

families Geraniaceae, Oxalidaceae, Tropaeolaceae, Linaceae, Erythroxylaceae, Zygophyllaceae, Rutaceae, Simarubaceae, Burseraceae, Meliaceae, Malpighiaceae, Polygalaceae and Euphorbiaceae. The family Tropaeolaceae contains the nasturtiums (Tropaeolum majus and T. minus) frequent in cultivation, the fruits of the species being used for pickles. T. Lobbianum is a showy greenhouse plant. The T. tuberosum of Peru produces a tuberous root used for food in Bolivia, cooking dispelling the unpleasant flavor.

Dr. Halsted states that some persons have suffered from an inflammation on the hand caused by handling the garden nasturtium.

T. majus contains glucotropaeolum, similar to the essential oil of mustard. The family Burseraceae contains Commiphora abyssinica furnishing myrrh, and Almacigo (Bursera simaruba) the most characteristic tree of Porto Rico and one which furnishes a resin known in commerce as "chibon." The family Meliaceae includes mahogany (Swietinia Mahogoni) a valuable timber tree of the Antilles; myrtle (Melia Azedarach), widely cultivated in the South as an ornamental plant, the fruit of which contains mangrovin and is said to be poisonous; the West Indian cedar (Cedrela odorata), which furnishes a valuable wood used for furniture, cigar boxes, shingles, etc.; and Trichilia emetica, which yields an oil and tallow. The carapa oil made from the seeds of Carapa procera is toxic for insects. The family Zygophyllaceae furnishes lignum-vitae (Guaiacum officinale), a heavy wood used in machinery and casting work.

The ratsbane, broken-back or mendis (Chailletia toxicaria) of the family Chailletiaceae is much used in Sierra Leone country of Africa for poisoning; it is placed in water to poison enemies and live stock. According to Dr. Renner "No one in this colony, it would appear, dies from natural causes." Dr. Renner found the cause of this mysterious trouble to be due to poisoning from ratsbane poison. In one case, a laborer was poisoned from "having eaten some fish on which the ground fruit of Chailletia toxicaria had been strewn for the purpose of killing rats." This shrub and an allied species are common in Upper Guinea and Senegambia. Drs. Frederick B. Power and Frank Tutin who made chemical and physiological examination of the fruit of Chailletia toxicaria found that the fruit contained neither an alkaloid nor a cyagenetic glucoside although a glucoside of this character is said to occur in South African C. cymosa. The ratsbane contains a resinous substance which is extremely poisonous, but a toxin could not be isolated. The syrup prepared from the resin when given to a dog caused delirium and epileptiform convulsions soon followed by death. Drs. Powers and Tutin found that the fruit of this plant contains two active principles, one of which causes cerebral depression or narcosin and that the poison which causes convulsions is cumulative in its effect.

To the family Erythroxylaceae belongs coca ($Erythroxylon\ Coca$) which contains a number of alkaloids as follows: $cocain\ C_{17}H_{21}NO_4$, $cinnamylcocain\ C_{19}H_{23}NO_4$, $truxillin\ (a)\ (C_{19}H_{29}NO_4)_2$ $truxillin\ (b)\ (C_{19}H_{23}NO_4)_2$, $benzoylecgonin\ C_{16}H_{19}NO_2$, $tropa-cocain\ C_{15}H_{19}NO_2$, $hygrin\ C_8H_{15}NO$, $cusoylgrin\ C_{13}H_{24}NO_2$. The injurious effects of cocain are well known. Dr. Winslow says:

Solutions of cocain (4.10 per cent), applied to mucous membranes, produce perfect local anaesthesia by paralyzing the sensory nerve endings. Cocain exerts a local anaesthetic action upon the gastric mucous membrane, and in this way lessens the appetite and sometimes stops vomiting. Intestinal peristalsis is increased by moderate doses, but is decreased and destroyed by the paralytic action of large doses. The action of cocain upon the heart and



Fig. 316. Coca-tree (Erythroxylon coca). Furnishes the coca of commerce. (After Faguet).

vessels is not very marked, except in poisoning. The alkaloid is, however, a slight cardiac stimulant in moderate doses, increasing the pulse-rate and tension. The action upon the heart is caused by depression of the cardio-inhibitory centres, and sometimes as well by depression of the cardiac inhibitory ganglia. Vascular tension is increased because of stimulation of the medullary vasomotor centres, smooth muscle of the walls, and because of the increased action of the heart. On the other hand, both minute and large doses may diminish the pulse rate.

Cocain is a respiratory stimulant in medicinal doses, but a paralyzant in toxic amounts. The respiratory centres are first stimulated and the breathing is made deeper and quicker. Depression and paralysis of the respiratory centres follow; cyanosis supervenes, and the respirations are shallow and irregular. Death occurs from asphyxia. In man, an amount of cocaine exceeding gr. 34 should not be employed under the skin, or upon mucous membranes, and death has occurred in susceptible patients from even smaller uoses. The most powerful action follows the use of cocain in very vascular parts, as about the face. Onc half a grain of cocain given subcutaneously to a girl cleven years old, was followed by a fatal result in 40 seconds, and the writer has seen violent convulsions produced by the instillation of a few drops of a 2 per cent. solution into the eye of a man. On the other hand, spontaneous recovery has obtained in the human subject after the ingestions of 22 grs. of the alkaloid. In the horse, the toxic dose of cocain causes restlessness and excitement. dilated pupils and salivation, culminating within an hour in a state of acute mania and intense excitement. These symptoms are followed by gradual recovery after a lapse of a few hours. Three grains of cocain given under the skin, will sometimes induce nervous excitement in susceptible horses. The treatment of dangerous forms of cocain poisoning, with respiratory and heart failure, consists in the use of rapidly acting stimulants,-as nitroglycerin upon the tongue, and strychnin, atropin and brandy subcutaneously.

Families of Geraniales

Flowers regular or nearly so, petals present usually as many as the sepals; flowers perfect; leaves not punctate.

| Herbs. |
|--|
| Capsule splitting into 5 carpels; leaves 3-foliolate or dissected |
| Geraniaceae. |
| Capsule 2-5 celled not splitting into carpels. |
| Stamens 2-3 times as many as the petals. |
| Leaf 3-foliolateOxalidaceae. |
| Stamens as many as the petalsLinaceae. |
| Trees or shrubs with compound leaves; leaves often punctate. |
| Leaves punctateRutaceae. |
| Leaves not punctateSimarubaceae. |
| Flowers irregular; petals 3, stamens diadelphous or monadelphous Polygalaceae. |
| Flowers regular generally apetalous, monoecious; carpels mostly 3; generally |
| herbs with milky juice |

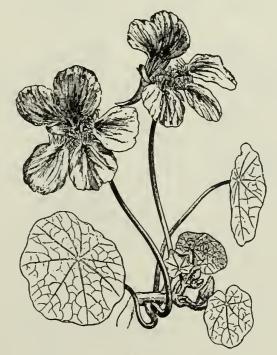


Fig. 317. Nasturtium (Tropaeolum majus). Flowering branch. (After Faguet).

GERANIACEAE

Herbs with alternate or opposite leaves; flowers perfect, regular, 5-nerved, hypogynous; stamens as many or twice as many or more than the petals; ovary 1, usually 5-lobed; ovules 1-2 in each cavity; fruit capsular. About 450 species of wide distribution. Native to the tropics and temperate regions of both hemispheres. Many plants of this order are frequently cultivated; among these are the South African pelargonium, commonly called the geranium, which contains geraniol C₁₀H₁₈O. The sharp points of the fruit of some are injurious.

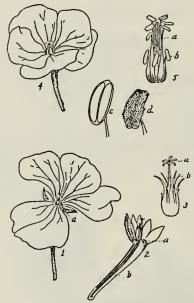


Fig. 318. Cultivated Geranium (Pelargonium zonale). 1, 4, Flower; b, c, d, Stamens; a, Stigmas.

frequently entering the flesh and in some cases this mechanical injury has produced death in sheep, just as in the case of Stipa. The Geranium Robertianum has a disagreeable bitter taste.

Erodium L'Her. Storksbill

Herbs with opposite or alternate stipulate leaves; flowers nearly regular, axillary or umbellate; sepals 5, imbricated; petals 5, hypogynous, the upper slightly smaller; glands of the disk 5, alternate with the petals; stamens 10; anther bearing 5, and as many sterile filaments; ovary deeply 5-lobed and 5-celled, beaked by the united styles, 5 in number; lobes of the capsule 1-seeded; the style when mature breaks away elastically and is coiled spirally; tails of carpels hairy on the inside; seeds not reticulated. The 65 species found in temperate and warm regions. Some species have become widely distributed because the seeds cling to the fleece of animals. Some species are troublesome in western United States.

Erodium cicutarium L'Her. Alfilaria or Storksbill

A hairy, tufted annual with low spreading stems; plant viscid or sticky; leaves pinnate or once to twice pinnatifid; flowers in umbel-like clusters, purple or pink; fruit hairy on the inside and spirally twisted when ripe. The E. moschatum is a stouter plant which occurs occasionally eastward.

Distribution. This plant is common upon the Pacific Coast especially California, occurring in grain fields and waste places. It is also abundant in dry soils in the Salt Lake basin and from Colorado to Texas; occasionally found in the eastern states and Manitoba. Native to the Old World. The weed is commonly scattered by animals. It is injurious to wool.

Injurious properties. Species of Erodium, like those of Stipa, have in some cases sharp pointed calluses which bury themselves in the flesh and inflict injuries to animals. Our common species is but slightly troublesome in this way. The carpels of the *Erodium* get into the fleece of sheep and thus the wool is rendered somewhat less salable.

E. moschatum is injurious.





Fig. 320. Musk Erodium (Erodium moschatum). (After Fitch).

Fig. 319. Hemlock Stork's Bill (*Erodium cicutarium*). This widely distributed plant sometimes causes mechanical injuries in animals. (Charlotte M. King).

OXALIDACEAE

Generally herbs, frequently with bulbs; acid juice; leaves palmate, with obcordate leaflets; flowers regular, 5-merous; stamens 10-15; ovary 5-celled; carpels with few or many ovules, loculicidal. A small order of 250 species chiefly tropical.

Oxalis L. Sorrel, Oxalis

Annual or perennial herbs with sour juice; often bulbous with alternate, digitately compound leaves of 3 leaflets; flowers in umbel-like clusters, solitary or several flowered, regular, often dimorphic or trimophic; sepals 5; petals 5;

stamens 10; pistil 1; ovary 5-celled; ovules several in each cell; 5 separate styles; pod 5-celled, opening loculicidally; seeds 2 or more in each cell, the outer coat dehiscent; embryo large, endosperm present. The 250 species chiefly found in the tropics. The Oxalis violacea with violet corolla is a common plant in woods and prairies. The O. corniculata, a yellow flowered species, occurs from Pennsylvania to Illinois. The fresh juice of this is said to be an antidote against poisoning from the seeds of Jimson weed. Several South American species like O. flava and O. Ortgiesi are cultivated indoors. The O. tetraphylla and O. lasiandra with their crimson flowers are also handsome for indoor cultivation. The O. crenata of Peru is cultivated for its tuberous roots.

Oxalis violacea L. Violet Wood Sorrel

Perennial with brownish bulb and ciliate scales; leaves smooth; leaflets obcordate, the midrib sometimes sparingly hairy; flowers in cymose clusters; sepals 5; petals 5, violet purple; capsule ovoid; seeds flattened, rugose-tuber-culate.

Distribution. New England to Florida and New Mexico.

Poisonous properties. Dr. Schaffner notes a case of poisoning as follows: "A case is recorded of a boy being thrown into violent convulsions by eating a considerable quantity of the leaves."

Prof. Hyams states that children have been known to die from constantly eating the raw herbs of O. grandis.

LINACEAE. Flax Family

Herbs, rarely shrubs; stipules small or none; flowers regular and symmetrical, hypogynous; sepals 5, rarely 4, imbricated and persistent; petals 5, or rarely 4, convolute; stamens 5, monadelphous at the base, alternate with the petals; pistil 1, 2-5-celled; styles 2-5; fruit capsular; seeds 1-2 in each cavity; cotyledons large, flat, without endosperm or with a small amount. A small order of 4 genera and 90 species, mostly in the genus Linum.

Linum (Tourn.) L. Flax

Herbs, sometimes with a woody base with tough fibrous bark; leaves sessile; stipules wanting or a pair of glands; flowers in cymes, racemes, or panicles; sepals 5; petals 5, soon falling; stamens 5; pistil 1; ovary 4-5-celled or becoming divided by false partitions, making 10 cells; seeds shining with a mucilaginous coat; large cotyledons. Several species commonly cultivated for ornamental purposes. The blue-flowered *L. perenne* of the Rocky Mountains, and the red garden flax (*L. grandiflorum*) a hardy annual from North Africa, are cultivated.

Linum usitatissimum L. Flax. Linseed

Annual; stem corymbosely branched at the tip; acuminate sepals; flowers broad; petals large, blue. Widely cultivated in the North and frequently spontaneous.

Poisonous properties and uses. The blue-flowered annual (L. usitatissimum) has been cultivated for centuries. The fiber has been found among the remains of the Swiss Lake dwellers. The ancient Egyptians as well as the Greeks and Romans also used the fiber for the manufacture of cloth. It is extensively cultivated in various European countries, fine fiber being produced in Belfast, Ire-

land; Brussels, Belgium; in Russia, and the Nile region. The seeds are also used extensively for making linseed oil. The chief regions where it is cultivated in North America are the Dakotas, Minnesota, Nebraska and Manitoba. Russia also cultivates the plant for the oil. The seed contains linolleic acid C, H, O, and is rich in oil. The compressed refuse is manufactured into oil cake, which is used for cattle food. The flax oil found in the seed of the plant is about onethird of its weight. Commercially, between 20 per cent and 30 per cent are obtained. When fresh it is without color and has little taste. The commercial oil is yellow and has a repulsive taste. On exposure to the air after having been heated with oxide of lead, it dries up to a transparent varnish consisting chiefly of linoxyn C₃₈H₅₄O₁₁. In medicine the flax seed is used in the form of a poultice, which is made of the pulverized seed. When oil cake or oil meal is fed in concentrated form it produced digestive trouble to hogs, frequently resulting in death. Dr. Schaffner states that it causes death to cattle, probably due to the prussic acid evolved from the plant when wilting. This substance has been reported.

Friedberger and Fröhner state that it causes violent colic, inflation, diarrhoea, staggering, palpitation, death with convulsions; autopsy shows gastro-enteritis and signs of axphyxiation.

Linum rigidum Pursh. Large-flowered Yellow Flax

An herbaceous glaucus or slightly puberulent annual with rigid angled branches from 1-2 feet high; leaves narrow, erect, usually with stipular glands; flowers large ,yellow; sepals acute or awn-pointed, glandular, serrulate; petals cuneate-obovate longer than the sepals; styles separate only at the summit; capsule 5-valved and ovoid.

Distribution. Loess soil of western Iowa to Missouri, Texas, Mexico to Arizona and Manitoba.

Poisonous nature. According to Chesnut the plant is reported as poisonous to sheep in the Pecos Valley, Texas.

RUTACEAE. Rue Family

Trees, shrubs, or herbs with simple, compound, alternate or opposite leaves, glandular, with punctate dots without stipules; flowers mostly in cymose clusters, polygamo-dioecious hypogynous, or perigynous; sepals 4-5; petals 4 or 5; stamens of the same number or twice as many, distinct, inserted on the receptacle; pistils 2-5, distinct or one compound; 2-5 carpels raised on an annular disk; embryo large, curved or straight; endosperm fleshy or none.

About 875 species, mostly in tropical regions of South Africa and Australia. Few representatives in North America. Two species of prickly ash (Zanthoxylum americanum Mill and Z. Clava-Herculis L.) and our hop-tree (Ptelea trifoliata) are common in the United States. The fruit of the hop-tree is used in Russia as a substitute for hops. A bitter alkaloidal principle occurs in Xanthoxylum. The gas plant (Dictamnus albus) a viscid glandular plant with strong aromatic scent is commonly cultivated. The common rue (Ruta graveolens), a native to Europe, is sometimes cultivated in country gardens. It has a strong disagreeable odor, and is so acrid that it will even blister the hands. It contains an acrid narcotic poison. The cork tree (Phellodendron amurense) from the Amur region, is occasionally cultivated. The most important genus of the

order is Citrus. The orange (Citrus Aurantium) is extensively cultivated in California and Florida. C. Aurantium var. vulgaris is the bitter orange which has run wild in Florida and other parts of the world. It is used in the manufacture of candied orange peel. The citron (C. Medica) produces the oil of citron, the thick peel being used to make the citron of commerce. The lemon (C. Limonum), wild in northern India, introduced into Europe by the Crusaders, is now well known in cultivation in California. The lime (C. Limetta) is cultivated in all tropical countries, and with the lemon is used to make lime juice. It is a refreshing drink and on sea voyages is used as an antiscorbutic. The lemon and lime are forms of C. Medica. The mandarine or tangerine (C. Aurantium) having a small flattened fruit with a thin rind and rich fruit, is grown in California and China. It is hardier than the orange, but probably a form of it. The shaddock or grape fruit (C. decumana) with large and somewhat bitter fruit, is native to Polynesia, and in recent years has become much better known in the United States. The kumquat (C. japonica), native to Japan and China, produces a small and pleasantly flavored fruit. The Aegle sepiaria (C. trifoliata) hardy as far north as Washington, is a spiny shrub producing a many-seeded, yellow, austere fruit. Hybrids of the species and C. Aurantium with better and larger fruit, have been produced by Webber. The sour orange or Naranja (C. Bigaradia) of Porto Rico and Florida is used for stocks in all plantings on moist lands because it resists the foot-rot which affects other varieties. The Beal fruit (Aegle Marmelos), native of India, with fruit about the size of an orange, produces a delicious fragrant material used in medicine. Jaborandi (Pilocarpus pennatifolius) native to Brazil contains the alkaloid pilocarpin C11H17N2O2 and is a powerful diaphoretic. The administration of more than 5 grs. of the alkaloid is dangerous to horses when given subcutaneously. Atropin is an antidote. The alkaloid jaborin C₂₂H₃₂N₄O₄ resembles atropin, also the alkaloid pilocarpidin. The bark of angustura (Cusparia febrifuga) native to Venezuela contains cusparin C20H10NO3 three other alkaloids and the bitter principle angusturin. The C. toxicaria of Brazil is poison

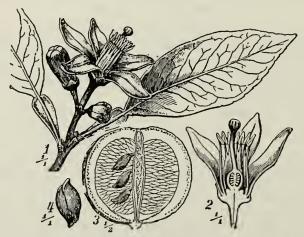


Fig. 321. Orange (Citrus Aurantium). 1. Flowering branch. 2. Longitudinal section of flower. 3. Longitudinal section of fruit. 4. Seed. (After Wossidlo).

ous. Lunsia amara contains a toxic glucoside. Citric acid is found in fruits of lemons, lime and other members of the genus Citrus. The glucoside hesperidin $C_{50}H_{60}O_{27}4H_{2}0$ occurs in ripe and unripe fruits of Citrus; the resinous principle naringin $C_{23}H_{21}O_{12}$ in C. decumana. The essential oil of lemons is one of the terpens $C_{10}H_{10}$; the oil of bergamot similar to the preceding is from Bergamot; limettin $C_{10}H_{14}O_{6}$ is the bitter principle of Citrus Medica—C. Limetta. The leaves of Buchu (Barosma crenulata) act as a mild diuretic. It contains a volatile oil of which 30 per cent is disophenol, also a crystalline glucoside (diosmin). The Commiphora abyssinica contains a volatile oil consisting of cuminol and eugenol. Aurantiamaric acid occurs in several species of Citrus...

SIMARUBACEAE. Ailanthus Family

Trees or shrubs with bitter bark; leaves pinnate, alternate, without punctate dots; stipules minute or none; flowers in axillary panicles or racemose clusters; regular, dioecious or polygamous; calyx 3-5 lobed; petals 3-5; stamens of the same number as the petals or twice as many; pistils 2-5 and 1-5 celled; disk elongated or annular.

A small family of 125 species of warm or tropical regions. The most widely known member of the family in the United States is the tree-of-Heaven or Chinese sumac (Ailanthus glandulosus). The quassia (Q. amara) of Guiana is used in fevers and as a substitute for hops to impart bitter flavor to beer. It contains quassin $C_{32}H_{40}O_{10}$ a bitter principle. The bark of other plants of the order is bitter, like the simaruba bark. The cedron (Simaba Cedron) of Central America is used in the tropics for snake bites. The bitter fruit of Simaba valdivia contain a gulcoside $C_{18}H_{24}O_{10}$.

Ailanthus Desf.

Large trees; leaves compound, odd-pinnate; flowers in panicles, greenish white; calyx short, 5-cleft; 5 spreading petals; disk 10-lobed; 10 stamens of the staminate flowers inserted at the base of the disk; ovary of the pistillate flowers deeply 2-5 cleft, 1-celled; stamens 2-3; winged fruits 2-5. Three species native of China and Eastern Asia.

Ailanthus glandulosus Desf. Tree-of-Heaven

A tall tree with ample leaves, smooth or slightly pubescent; 13-41 stalked leaflets; ovate or ovate-lanceolate flowers, greenish pedicelled, the staminate ones badly scented.

Distribution. Commonly escaped from cultivation, along roadsides from Southern Ontario to Kansas, Southeast Iowa, hardy as far north as Central Iowa.

Poisonous properties. The bark is known to be poisonous. Dr. White, in his Dermatitis Venenata states that he read an account in some medical journal of the suspected poisoning by this tree during its flowering season, and the statement was made that a case of marked dermatitis of the face, had been attributed to the emanations of a tree of this species, growing very near the sleeping-chamber of the patient. He records a case where a lady was poisoned by contact with it. Dr. Halsted states that when the flowers are handled they produce an irritation of the skin.

In the Medical and Surgical Reporter of Philadelphia for 1872, this statement is quoted by Dr. Rusby in regard to the poisoning coming from the roots of this plant.

A case in which four persons were apparently poisoned by this root. They were members of one family and were successively, that is, at intervals of a few days, attacked, with no other possible cause than their drinking water which they took from the well of a neighbor. They all drank water exclusively, except the husband, who was the last to be taken. Others who drank of this water occasionally suffered similarly but to a slight extent. All immediately began to recover as soon as the drinking of this water was stopped. The symptoms, which had been slight for many weeks, appeared in a violent form in November, at which time an Ailanthus tree growing in the vicinity of the well must have shed its leaves, and to a great extent its fruit also, if a pistillate tree, which fact was not stated. On examination the soil all about the well was found to be thickly permeated with the roots of this tree, and these were also supposed to extend into the water, though an investigation regarding this was apparently not made. Inasmuch as the symptoms had existed in a mild form before the fall of the leaves, it is fair to assume that the roots had contributed toward the result, while the violent out-break in November would seem to indicate a sudden increase in the cause due to the accumulation of the leaves in the well. The symptoms were jaundice, a dingy aspect of the face and eyes, countenance fixed and anxious, pulse frequent and soft, yellowish fur on tongue, except on the tip and edges, tenderness over the liver, and most important, a persistent pain over the stomach with paroxysmal vomiting, pain in the back, difficult urination and obstinate constipation. The symptoms were thus apparently to a great extent those of chronic gastritis.

Dr. Schaffner says that cows will not eat grass near the young shoots. Quercetiin occurs in the leaves. They also contain the bitter principle linuttin.

POLYGALACEAE. Milkwort Family

Herbs or rarely shrubs; stipules none; flowers perfect; sepals 5; petals 3 or 5, free; stamens 4-8, monadelphous, or diadelphous; anthers 1-celled, open-

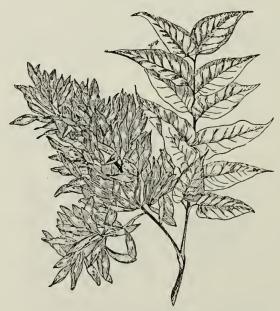


Fig. 322. Tree of Heaven (Allanthus glandulosus). The bark is supposed to cause dermatitis. (Ada Hayden).

ing at the top by a pore; ovary 2-celled; ovules 2; fruit a 2-seeded pod. A small order of about 700 species, found chiefly in the tropical and temperate regions. Some species of the order produce a strong fiber.

Polygala (Tourn.) L. Milkwort

Herbs or shrubs; simple entire, dotted leaves without stipules; flowers perfect, irregular, occasionally cleistogamous; calyx of 5 sepals, the 2 lateral known as wings, large, colored, the other small, greenish; petals 3, free, connected with each other and the stamen tube; stamens 6 or 8, filaments united below or in 2 sets; pistil 1; ovary 2-celled; ovules 1 in each cell; fruit mainly capsular; seeds with a caruncle, anatropous; embryo large; little endosperm. About 250 species, of wide distribution, chiefly of warm regions. A genus of little economic importance.

Polygala Senega L. Seneca Snakeroot

Plants clustered, several from a woody and knotty rootstock, simple 6-12 inches high; leaves lanceolate or oblong-lanceolate with rough margins; dense spike, long peduncle; flowers white, none cleistogamous; wings round-obovate; crest short; seeds hairy.

Distribution. In rocky woods or clay soil. New Brunswick to Minnesota, Central Iowa to the Rockies in Canada.

Poisonous properties. The dried root is gathered when the leaves are dead,



Fig. 323. Seneca Snakeroot (Polygala Senega). Dried root is made into a powder which is very acrid. (From Johnson's Med. Bot. of N. A.).

and made into a powder and a tincture prepared. This tincture has a peculiar acridity. Dr. Millspaugh says:

After tasting the tincture or chewing the rootlets, a very peculiar sensation of acridity and enlargement is felt at the root of the tongue, which, once recognized, will always mentally associate itself with this plant. The root contains polygalic acid $C_{22}H_{18}O_{11}$. It is a white, odorless, acrid, amorphous powder.

This acid forms a frothing, saponaceous solution in boiling water, and breaks up into sapogenin and amorphous sugar, to which the name senegin has been given, which by some has been regarded as identical with saponin. According to the author quoted above, in doses of 10 minims of the tincture to a scruple of the powdered root, it produces:

Anxiousness, with dullness of the head and vertigo; aching and weakness of the eyes, with lachrymation, pressure in the ball, flickerings, dazzling vision, and contracted pupils; sneezing; ptyalism; inflammation of the fauces and oesophagus, with thirst with anorexia; nausca; mucous vomiting; burning in the stomach; cutting colic; roughness and irritation of the larynx, with orgasm of blood to the chest, accompanied by constriction, aching, soreness, and oppression; general debility; restless sleep; and profuse diaphoresis.

Senegin resembles other saponins. Recent investigations indicate that the plant also contains quillagic acid $C_{19}H_{30}O_{10}$ sapotoxin and two senega saponins. The saponin of Polygala virginiana has the formula $C_{32}H_{52}O_{17}$. Other species of Polygalaceae like P. venenosa contain saponin.

EUPHORBIACEAE. Spurge Family

Herbs, shrubs or trees usually with a milky acrid juice, opposite alternate or verticillate leaves; monoccious or dioecious flowers, much reduced, subtended by bracts resembling a calyx or corolla; ovary usually 3-celled; ovules 2 in each cell, pendulous; stigmas as many or twice as many as the cells; styles generally 3; fruit a capsule, separating elastically into a 2-valved capsule; fleshy or oily endosperm; seeds with flat cotyledons.

A large family of 4000 species, chiefly tropical, many of which possess noxious qualities. Some species of the genus Manihot found in tropical Amer-



Fig. 324. Manchineal Tree (Hippomane Mancinella). Furnishes an arrow poison. (From Vesques' Traité de Botanique).

ica are poisonous. The fresh juice of bitter cassava administered to dogs and cats causes death in twenty minutes. The starch from this is used for sizing. Cassava (Manihot utilissima) is extensively cultivated in tropical America and to some extent also in Florida. The sweet cassava roots are used as food for cattle and man. Tapioca is the starch which settling from the water used to wash cassava meal, is afterward dried. An intoxicating drink is made from cassava bread. Rubber plants of the order are the Hevea, Micrandus and Manihot. The manchineal tree (Hippomane Mancinella), the celebrated poison tree of tropical America furnishes an arrow poison. The fruit, though tempting, contains an acrid poison, which causes blisters to form. The poisonous properties are said to rival those of the deadly Upas tree (Antiaris toxicaria).

The following is an extract from "West India Sketches":

The branches contain a milky juice which will certainly blister the skin ,and it has heen a common trick among the negroes to apply it to their hacks in order to excite the compassion of those who might mistake it for the effects of heating.

Kingsley, in his charming "At Last," writes of it:

We learnt to distinguish the poisonous manchineal, and were thankful in serious earnest that we had happily plucked none the night hefore, when we were snatching at every new leaf; for its milky juice by mere dropping on the skin hurns like the poisoned tunic of Nessus, and will even, when the head is injured by it, cause hlindness and death.

Dr. White in his Dermatitis Venenata, speaks of the use of the plant in the West Indies as follows:

This large family of Euphorbiaceae contains some of the most poisonous plants. One of the most virulent is the manchineal, a small tree, bearing fruit resembling an apple, which grows in Southern Florida.

Loudon states that it abounds in a white milk which is highly poisonous, and so very caustic that a single drop placed upon the skin instantly causes the sensation of a hot iron, and in a short space of time raises a blister. It is a common belief that to sleep under it causes death. Whole woods on the seacoast of Martinique have been burned in order to clear the country of such a dangerous pest. The fruit is highly poisonous.

Mr. Combs in his paper on Cuban Medical plants, states that its poisonous effects may be overcome by the use of Tecoma leucoxylon or Jatropha gossypifolia. The uncooked rhizome of Maranta arundinaceae is sometimes used for the same purpose. The latex of the sandbox tree Hura crepitans is also very poisonous and when applied to the skin, causes eruptive pustules resembling those of erysipelas. It also produces injuries to the eyes. It contains a sharp acrid poison. When taken internally it produces vomiting and diarrhoea. The seeds are used as emetic.

The Nigeria species of Mahogany (Ricinodendron africanus) also produces a valuable wood.

The tallow tree (Sapium sebiferum) is cultivated in tropical countries for wax found on the fruit, which is made into candles. The candlenut tree (Aleurites triloba) is cultivated on the islands of the Pacific Ocean for oil found in the seeds, which is made into candles, soap, etc. The seeds of A. moluccana are roasted and eaten. The Kalo Nut (A. Fordii) according to Holmes* is poisonous having produced toxic symptoms in five children. It is the source of tung or Chinese wood oil. The seeds of pinhoen oil (Jatropha Curcas) are eaten. They are nutty and have a pleasant flavor, but when eaten in excess, produce serious trouble and death often results. The drastic principle of Croton

^{*} Pharm. Jour. 4:25, 231, 241. Brit. Yearhook of Pharm. 1908:240.



Fig. 325. Sandbox Tree (Hura crepitans). 1. Flowering and fruiting branch. 2. Part of a large branch. Latex causes dermatitis, (From Vesque's Traité de Botanique.)

oil (Croton Tiglium) has not been definitely determined, but it has been refined to a dark brown oil known as crotonol. The plant contains tiglinic acid C₅H₈O₂. Crotinic acid C₄H₆O₂ and Croton oil are derived from the Croton Tiglium, that is cultivated in southern India. This plant is a drastic purgative, capable, when given in excessive doses, of causing death. The resin produces vesication. The seeds, according to Blyth, are very poisonous. The fixed oil has the

formula C₀H₁₄O₂.

The bark of Cascarilla (Croton Eluteria), native to the Bahamas, is used as a tonic and contains cascarillin C₆H₉O₂. The milky juice of Euphorbia resinifera of Morocco is used as a purgative and is so intensely acrid that people in collecting it are compelled to tie a cloth over their nostrils and mouths. It contains the substance euphorbon C27H44O7, which has a burning taste. The milky juice of agallocha (Excoecaria Agallocha) of tropical Asia is very acrid and blisters the skin. It is said that if the juice drops into the eye as sometimes happens to the woodcutter, blindness may be caused. Excoecaria glandulosa contains excoecarin C13H12O5. The Homalanthus Leschenaultianus is said to be poisonous. The fruit of Hyaena poison (Toxicodendron capense) of South Africa is very poisonous, and is used to destroy beasts of prey. Gum elastic or Para rubber is derived from the South American Hevea brasiliensis. Other plants of the family yield caoutchouc, which contains hydrocarbons that are readily soluble in chloroform. The alkaloid drumin occurs in Euphorbia Drummondii. Several species of Euphorbia like Poinsettia (Euphorbia splendens) and E. heterophylla are cultivated for ornamental purposes. From the



Fig. 326. Croton (Croton Tiglium). Flowering and fruiting branch. The source of croton oil. (After Faguet.)

glands and hairs covering the fruit of kamala (Mallotus philippinensis) a dye is made. The fruit is also used as a vermifuge; it contains rottlerin C22H20O6 and isorottlerin. Many species of the genus are regarded as poisonous. Maiden states that the E. Drummondii is poisonous to stock in New South Wales. known as the milk plant and is especially troublesome to sheep. It causes the head to swell to an enormous size so that the animal cannot support its head. Suppuration frequently follows. E. alsinaeflora is also poisonous to sheep in the same country. E. eremophila is another suspect in that country. E. heptagona is an arrow poison. Some species of this genus are used as fish poisons. Emanations of E. characias at one time were supposed to cause malarial fever which, however, was an erroneous assumption. Lehmann, a German writer on poisonous plants lists the following species as poisonous: E. Lathyris, E. Heliscopia, E. platyphylla, E. Esula, E. Cyparissias, E. palustris, E. Peplus, E. exigua. The E. antiquorum of the East Indies, E. canariensis of the Canary Islands, and E. Reinhardtii of the Transvaal contain a milky acrid poisonous juice.* The resin from Euphorbia produces sneezing, irritation of face and skin, vomiting and diarrhoea and when used in large doses, death. Where the drug is manufactured, workmen must protect themselves; but, even then, headache, dizziness and weakness follow. To poisoning from members of the genus Euphorbia, Friedberger and Fröhner ascribe such symptoms as constipation, severe and bloody diarrhoea, feeble pulse and tympanites.

^{*} Bull. Misc. Inf. Roy. Gard. Kew, 1908: 154.

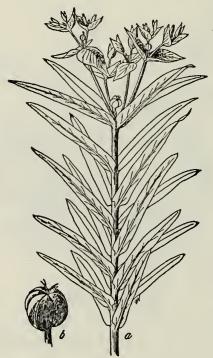


Fig. 326. Caper spurge (Euphorbia Lathyris). a, upper half of plant, one-third natural size; b, seed capsule, natural size. Listed by Lehmann as poisonous. (Chesnut, U. S. Dept. Agr.)

According to Greshoff the leaves of Andrachne cordifolia and other members of the family contain hydrocyanic acid.

Key for genera of Euphorbiaceae:

Flowers without a calyx inclosed in a cup-shaped involucre. 5. Euphorbia. Flowers with a calyx; involucre absent.

Flowers apetalous in panicles: stamens 10.

Calyx corolla-like; plant with stinging hairs. 4. Jatropha. Flowers in terminal racemes or spikes covered with scurfy or stellate hairs, glandular.

Flowers spiked or glomerate; ovary usually 3-celled. 1. Croton. Flowers in axillary spikes or paniculate; stamens 8 or more.

Fertile flowers in the axils of leafy bracts; stamens usually 8.

7. Acalypha,

Flowers in interrupted axillary spikes; stamens 8-20. 6. Mercurialis. Flowers paniculate; stamens very numerous; filaments branched.

3. Ricinus

Flowers apetalous in racemes or spikes; stamens 2 or 3 style simple.

Flowers racemose, hirsute or pubescent. Flowers spicate, glabroid. 2. Tragia.8. Stillingia.

1. Croton L.

Stellate, pubescent herbs or shrubs; leaves generally alternate, occasionally with glands at the base of the blade; flowers spicate or racemose, the staminate above; calyx 4-6 parted; petals usually present, small or rudimentary, alternating with the glands; stamens 5 or more; pistillate flowers with calyx 5-10 parted; petals usually wanting; ovary mostly 3-celled, with a single ovule in each cell.

Croton capitatus Michx. Hogwort

An annual, dense, soft woolly herb, somewhat glandular, from 1-2 feet high, occasionally branched; leaves entire, lanceolate oblong, with long petioles; sterile flowers with 5-parted calyx and as many glands alternating with the obovate lanceolate petals which are fimbriate; fertile flowers several, capitate or crowned; calyx 7-12 parted; 5 petals wanting; styles twice or thrice forked; seeds gray, smooth.

Distribution. A common weedy plant from Missouri to Texas; from New Jersey to Georgia, Iowa and eastern Kansas.

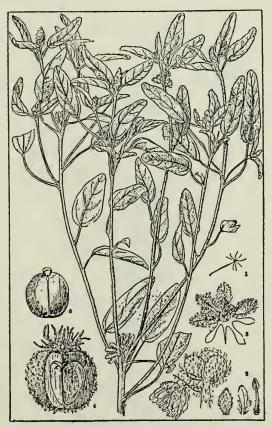


Fig. 327. Hogwort (Croton capitatus). Suspected of being poisonous. (After A. M. Fergusonn, Rep. Mo. Bot. Garden)

Croton texensis Muell

A branching annual from 1-2 feet high, covered with close stellate pubescence; leaves narrowly oblong-lanceolate to linear; dioecious; calyx lobes 5, unequal; petals none; staminate spikes short; stamens 10 or more; style 2 or 3 times dichotomously 2-parted; capsule stellate, tomentose and roughened; seeds ovoid or oval.

Distribution. From South Dakota to Colorado, Texas, Mexico, Missouri and Alabama.

Poisonous properties. Several species of the genus Croton are used in medicine. The Croton Tiglium contains an oil which given internally is a powerful cathartic, but when applied externally, is a rubefacient. Loss of the hair follicles and of hair may occur. When gently rubbed into the skin, it produces, after a short time, a considerable degree of itching, redness, and burning, and within a few hours small red papules may develop. If more of the oil is applied the papules are more abundant and are often surrounded with a bright red halo. They often become pustular and scars fill the pustules.



Fig. 328. Texas Croton (Croton texensis). Sometimes causes irritation of the skin. (After Mrs. M. H. D. Irish, Rep. Mo. Bot. Garden.)

Croton oil contains several fatty acids, such as stearic, palmitic, myristic and lauric acids. The volatile part of the acids contains an acid called tiglinic C₅H₆O₂, which is the same as angelic acid. The drastic principle of Croton oil has not been definitely determined, according to Flückiger and Hanbury. Crotonol C₁₈H₂₄O₄, is a non-purgative body causing irritation of the skin. According to Winslow, in his Veterinary Materia Medica and Therapeutics, "10 drops of croton oil will kill a dog unless vomiting occurs. 30 drops prove fatal to a horse, intravenously. The treatment of poisoning includes the use of emetics or stomach tube, demulcents and opium." None of our native species is mentioned as poisonous by Dr. Schaffner or Prof. Chesnut, but a few years ago I had a query through the Wallace Farmer in Des Moines, from a correspondent in Western Nebraska who suspected that the Texas croton was poisonous. The writer has eaten a few seeds of our southern Croton capitatus with slight uneasiness. On the other hand, a few seeds of the Texas croton produced powerful irritation which lasted for an hour, and then disappeared. It is listed by Bessey and O'Gara as possibly poisonous in Western Nebraska. Prof. Chesnut states in his paper on Plants used by the Indians in Mendocino county, California, that the bruised leaves of Croton setigera are used to stupefy fish. The common name, fish soap-root, indicates its use.

The bark of the cascarilla (Croton Eluteria), native of the Bahama Islands is used as a tonic.

Tragia L. Tragia

Monecious herbs or shrubs, usually armed with stinging hairs; leaves alternate; flowers in racemes with bractlets, apetalous; sterile flowers with a 3-5 cleft calyx; fertile flowers with a 3-8-parted calyx, divisions entire or pinnati-



Fig. 329. Spurge Nettle (Tragia urens). This spurge is common in some places in the South and has hairs that are irritating like those of the common nettle. (Charlotte M. King.)

fid; styles 3; capsule 3-lobed, separating into three 2-valved carpels. A small genus of 50 species.

Tragia urens L. Common Nettle or Tragia

A dull green, pilose plant with pilose or hirsute hairs; erect, branched stems; leaves obovate, or ovate-linear; short, petioled, pistillate flowers, several at the base of the racemes, with a 5-6 lobed calyx; capsule short-pedicelled.

Distribution. From Virginia to Florida and Texas.

Tragia nepetaefolia Cav. Tragia or Nettle.

A somewhat hispid, erect, or slightly twining plant, bearing stinging hairs; leaves ovate, or triangular-lanceolate; base cordate or truncate; short petioled; racemes many-flowered; pistillate flowers with a 5-lobed calyx; seeds chestnut brown.

Distribution. From Kansas to New Mexico.

Poisonous properties. The hairs have the same stinging property as those of the common nettle.

3. Ricinus (Tourn.) L. Castor Oil Bean

A tall, stout herb or tree in tropics; glabrous and glaucus; large, alternate, peltate leaves; flowers in large, panicled clusters; the fertile above, the staminate below; calyx 5-parted; stamens numerous; styles 3, united at the base, each 2 parted, red; capsule subglobose, or oval, separating into 3, 2-valved carpels; cotyledons large; endosperm fleshy and oily. A single species naturalized in warm countries, probably native to Asia.

Ricinus communis L. Castor Oil Plant

A tall, smooth, branching herb with palmately-lobed leaves; seeds oblong, shining, variegated with white.

Distribution. Widely cultivated as an ornamental plant, and an escape from cultivation from New Jersey to Texas.

Poisonous properties. The seeds furnish the well known castor oil, which is a mild and safe purgative. It contains ricinolein, or ricinoleic acid glyccrid, 'C5H5 (C18H34O3)3; an acrid principle; also palmitin, stearin, and myristin. 'The purgative principle found in it is unknown. Castor oil is not poisonous, thut the pulp contains an acrid, albuminous substance, ricin CaHaNaOa. Dr. Winslow, in speaking of the poisonous character, says, the seeds "contain 50 spercent of oil, and an acrid, poisonous substance. Three seeds have caused death in man,, and they are ten times more purgative than the oil." A few seeds eaten entire by a child might produce serious symptoms. According to Chesnut, the seed eaten accidentally by horses has caused death. They are used also to poison sheep, according to the same authority. The oil cake is said not to be poisonous to poultry and cattle. A case is known of a young lady whose eyes became inflamed when in contact with a mere trace of the material in the laboratory. The toxin is very poisonous, but animals may be rendered immune, and the seeds then fed to them. Behring has produced an anti-toxic serum against the ricin or toxin of the castor oil bean.



Fig. 330. Castor Oil Plant (Ricinus communis). Furnishes the well known castor oil of commerce. (After Faguet.)

The symptoms of poisoning are vomiting, gastric pain, bloody diarrhoea and dullness of vision.

Stillmark 1 states that the toxalbumin of castor oil bean, when injected into the circulation is more poisonous than strychnin, prussic acid, or arsenic.

Quite recently Dr. W. N. Bispham² reported on several cases of poisoning in Cuba from eating the seeds of the Castor oil plant. Some persons showed peculiar susceptibility; in one case poisoning occured from eating a single seed, while in another a good many were eaten; in both cases the seeds caused nausea, vomiting, and purging of a violent nature.

Toxic substances similar to ricin occur in the following plants. Abrus precatorius (abrin), Jatropha curcas (curcin), Croton Eluteria (crotin), Robinia Pseudo-Acacia (robin), Brayera anthelmintica (costoxin). According to Ceni and Besta a toxin also occurs in Urtica, Viscum seedlings, Aspergillus flavus, and A. fumigatus.

¹ Dorpat. Arcli. 3: (1889). 2 Am. Jour. Med. Sci. 126: 319-321.

4. Jatropha L. Spurge Nettle or Bull Nettle

Monoecious, or rarely dioecious, perennial herbs, with bristly hairs, entire or lobed leaves; flowers in cymes; calyx colored like petals in sterile flower, mostly salver-shaped, and 5-lobed, enclosing 10-30 stamens; pistillate flowers in the lower forks of the cymes; capsule ovoid or subglobose, separating into 2-valved carpels. A small genus of 4 or 5 species.

Jatropha stimulosa Michx

A branching, perennial plant with a stout root, 6-12 inches high, and stinging hairs; leaves round, heart-shaped, 3-5 lobed or variously cleft; calyx of the staminate flower salver-form, white or pinkish; stamens 10, filaments almost separate; seeds oblong-ovoid, smooth and mottled.

Distribution. In dry sandy soil from Virginia to Texas.

Poisonous properties. Mr. John Smith says that a plant growing at Kew was placed on his wrist, and produced in a few minutes, serious symptoms extending to the upper part of his body; the lips became swollen, and the whole of a livid red, fainting coming on in ten minutes. The writer was told of numerous instances of poisoning in Texas where it is much dreaded.

Jatropha urens, known as the Brazilian stinging nut, is considered to be one of the most poisonous plants known. The Cuban physic nut (Japtropha Curcas) is used as a purgative.

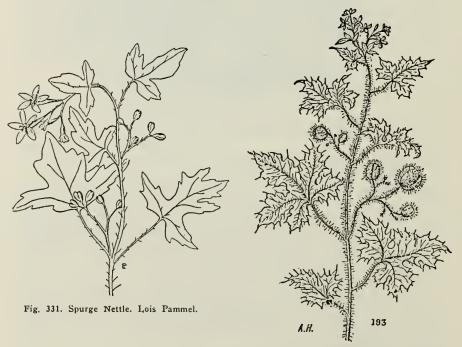


Fig. 332. Spurge nettle (Jatropha stimulosa). The Jatropha has stinging hairs that produce injuries similar to those produced by nettle but much more powerful. (After Hochstein).

5. Euphborbia L. Spurge

Monoecious shrubs or herbs with alternate or opposite, verticillate leaves; flowers involucrate, involucres resembling a calyx or corolla, bearing a large thick gland in the sinuses; staminate flowers consist of a single jointed stamen on a filament-like pedicel; pistillate flower solitary at the bottom of the involucre consisting of a 3-lobed and 3-celled ovary; capsule at maturity breaking into 3-lobed 1-seeded carpels; seeds frequently caruncled, smooth, variously pitted. About 700 species, chiefly in warmer regions. A few are weedy, some poisonous and some planted for ornamental purposes. The milky juice of the Brazilian *E. heterodoxa* produces a ferment which acts much like papain.

Euphorbia Presilii Guss. Large Spotted Spurge

An ascending, erect annual from 1-2 feet high, opposite oblique leaves, which are ovate, oblong or oblong-linear, falcate, serrate, usually with a red spot or red margins; stipules triangular; flowers collected in a loose terminal cyme; appendages entire, white or red; pod smooth, angled; seeds small, blackish, ovate, obtusely angled, wrinkled, and tubercled.

Distribution. Common in eastern North America west to the Rocky Mountains.

Euphorbia maculata L. Spotted Spurge

A prostrate spreading, hairy annual; leaves oblong-linear, pubescent or smooth, oblique at base, serrate above, small brownish spots on leaves; stipules lanceolate, fimbriate; flowers monoecious, included in a 4-5-lobed involucre; glands of the involucre minute; peduncles as long as the petioles, in dense clusters; pods minutely pubescent; seeds sharply 4-angled, having 4 shallow grooves, whitish.

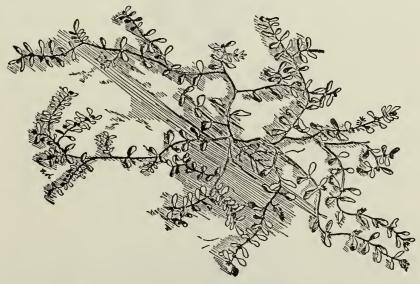


Fig. 333. Spotted Spurge (Eupharbia maculata). Common roadside plant. Probably poisonous. (C. M. King).

Distribution. Common along roadsides, walks, etc., from New England to the Rocky Mountains and the Gulf States.

Euphorbia marginata Pursh. Snow on the Mountain

An erect, stout annual from 2-3 feet high; stem hairy or somewhat smooth; leaves sessile, scattered, ovate or oblong, entire; deciduous stipules; uppermost leaves opposite or whorled with conspicuous white petal-like margins; involucre bell-shaped in umbels; glands of the 5-lobed involucre with broad and white appendages; seeds ovoid, globose, terete, dark ash colored, reticulate.

Distribution. Frequently cultivated in gardens from whence it has escaped. Found in Ohio, Illinois and Indiana. Native from western Minnesota, Iowa to Colorado, and Texas.

Euphorbia corollata L. Milkweed or Flowering Spurge

Perennial with a long, stout rootstock, glabrous or sparingly hairy; leaves ovate, lanceolate, or linear, obtuse, short-petioled, or sessile; inflorescence in umbel-like clusters; involucre long peduncled with white conspicuous appendages; seeds thick, ovoid, slightly pitted, ash-colored.

Distribution. In rocky or sandy soil, Mass. to New York, New Jersey, Florida, Wisconsin, Minnesota, and Texas.



Fig. 334. Common Flowering Spurge (Euphorbia corollata). A plant with milky juice which has irritating properties. common in sandy fields. It has white bracts resembling flowers. (Charlotte M. King.)

Euphorbia Lathyris L. Myrtle Spurge

A glabrous annual or biennial, simple below, branched above, from 2-3 feet high; leaves thick, linear or oblong, scattered, the upper lanceolate

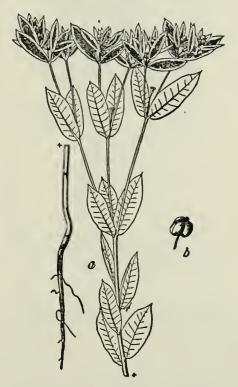


Fig. 335. Snow on the mountain (Euphorbia marginata): a, whole plant, one-third natural size; b, seed capsule, natural size. Cultivated in old gardens. (Chesnut, U. S. Dept. Agr.)

or linear-lanceolate; inflorescence umbel-like, bearing 4 crescent-shaped glands, prolonged into horns; seeds oblong-ovoid, terete, usually wrinkled.

Distribution. In waste places. New Jersey to North Carolina, Iowa, and California. Native to Europe.

Euphorbia Ipecacuanhae L. Wild Ipecac

Perennial from 5-10 inches high; long perpendicular root; entire, smooth leaves, varying from obovate or oblong to narrowly linear, nearly sessile; involucres long peduncled and 5 transversely elliptical or oblong green glands; seeds smooth, ovate, white, pitted, and obscurely 4-sided.

Distribution. In sandy soil from Connecticut to Indiana and Florida.

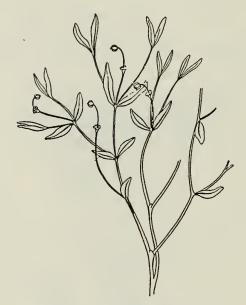


Fig. 336. Wild Ipecae (Euphorbia Ipecacuanhae.) Plant that possesses irritating properties and is also a purgative. (Millspaugh and Charlotte M. King.)

Euphorbia heterophylla L. Cruel Plant

An erect, smooth annual from 1-3 feet high; leaves alternate, petioled, linear lanceolate to orbicular, undulate, entire or toothed; the upper leaves usually fiddle-shaped, with a red base; involucre in terminal clusters, 5-lobed, with a single or a few almost sessile glands; seeds nearly round, transversely wrinkled and tubercled.

Distribution. From Illinois and Missouri to Nebraska.

Euphorbia Cyparissias L. Cypress Spurge

A bright green perennial from 6-12 inches high with running rootstocks; stems clustered, occurring in patches; stem leaves linear, entire, densely crowded, those of the flower heart-shaped and entire; flowers in umbellate clusters, umbel many-rayed, glands crescent-shaped; pods granular; seeds oblong and smooth.

Distribution. Native to Europe, but widely scattered in eastern North America. First introduced as a cultivated plant in North America.

Poisonous properties. All of the species are more or less irritating and in drying give off very disagreeable odors. Many of the species of the genus are used by quacks to remove warts and freckles; the juice produces an erysipelatous-like inflammation, and in one case mentioned by Dr. White, the whole abdominal wall became gangrenous.

The milky juice of the plant causes itching and inflammation. The general effect is very much like that of poisoning from the poison ivy. In Texas, according to Chesnut, the juice of E. marginata is used to brand cattle. The honey



Fig. 337. Yellow Flowering or Cypress Spurge (Euphorbia Cyparissias). A branch with large bracts and small flowers. (Straburger, Noll, Schenck and Schimper.)

obtained by bees from the plant is poisonous and is rendered unfit for use. The acrid properties of this species were described some years ago by Dr. Schneck. The juice of *E. corollata*, according to Dr. Halsted and many other observers, is acrid, and, on the authority of Dr. Bigelow, formerly was used for blistering purposes. The bruised root will vesicate the skin. According to Dr. J. C. White, the dust of this species produces painful swelling and vesicles upon men who handle the plant. It is used as an emetic, and is troublesome to those who collect it.

The Euphorbia pilulifera is used as a sedative in spasmodic conditions of the respiratory apparatus. It produces dermatitis. Dr. White, in his Dermatitis Venenata, has this to say of the species of the genus:

More than one hundred species of Euphorbia, or spurge, grow in the United States, either indigenous or immigrants from Europe. Of every species Loudon says the juice is so acrid as to corrode and ulcerate the body wherever applied; and of E. resinifera, from which the official euphorbium is obtained, Pliny and Dioscorides, according to the Dispensatory, describe the method of collecting juice, so as to prevent irritation of the hands and face. This substance is used as a plaster to prolong suppuration.

Van Hasselt states that the juice of several species is used by quacks to remove warts, freckles, as depilatory, etc.; and that the application of the juice, powder, and extract produces not only crysipelatous, pustular, and phlegmonous inflammation, but even gangrene. In one case mentioned the whole abdominal wall became the seat of gangrene.

Of our native species, Bigelow says that the juice of several was used in his day to destroy warts. and Gray describes them all as containing an acrid, poisonous juice. The most active of them are *E. corollata, E. Ipecacuanhae*, and *E. Lathyris*. The first of these, commonly called snake-milk, according to Bigelow, has been used for blistering purposes, and the Dispensatory states that the bruised root will vesicate the skin.

Mr. Cheney informs me that the juice of E. Ipecacuanhae is quite troublesome to many who collect and handle it; and Bazin states that the dust of E. Lathyris, growing both in Europe and in this country, causes redness, painful swelling, and vesicles upon the workmen employed in handling it.

With reference to the poisonous nature of the juice of the several species, nothing very definite is known. Euphorbon C₂₉H₄₄O₇ has been found in Euphorbia Ipecacuanhae. This euphorbon acts as an irritant to the mucous mem-



Fig. 338. Large Spotted Spurge (Euphorbia Preslii). Supposed to cause "slohhers" in horses. (Charlotte M. King.)

branes throughout the alimentary tract. The caper spurge (Euphorbia Lathyris) is poisonous, and the following physiological actions are described by Dr. Millspaugh:

Brilliant, staring, wide-open eyes, dilated pupils; death-like pallor of the countenance; retching and vomiting; violent purgation, stools frequent, copious, and in some cases bloody; irregular pulse; whole body cold and rigid, followed by heat and perspiration. M. M. F. Sudour and A. Caraven-Cachin state that emesis always precedes purgation, and that the seeds have an irritating action upon the mucous membrane of the intestinal canal, principally in the larger intestines. They divide the effects into three stages: a, the cold stage, including vomit-

ing and diarrhoea; b, the stage of excitation, including nervousness, vertigo, and delirium; y, the state of reaction, including heat and copious sweat.

With reference to the physiological action of the common spurge (Eu-phorbia Preslii), the following statement is made by Dr. True:

Headache with frontal fullness and heat; heat about the eyes; languor and drowsiness; oppression of the stomach; and constipation. The juice applied to the eyes causes severe irritation, with smarting and burning, lachrymation, and momentary blindness; this we have experienced twice while gathering the plant. It is supposed that this species causes the affection in horses called "slobbers."

6. Mercurialis L. Mercury

Annual or perennial herbs; with opposite pinnately veined leaves; flowers dioecious or monoecious in interrupted axillary spikes, apetalous; calyx small, green, 3-parted; capsule 3-lobed.

Mercurialis annua L. Annual Mercury

A leafy stemmed, erect, annual herb; leaves lanceolate or ovate-lanceolate, crenate serrate; carpels hispid; indigenous to Europe, found in waste places from Nova Scotia to Ohio, and South Carolina. The M. perennis differs from M. annua in having a creeping perennial root, and hairy leaves.

Pvisonous properties. Both species are acrid and poisonous.

7. Acalypha L. Three seeded Mercury

Herbs or shrubs, leaves alternate, petioled; flowers stipulate in spikes or spike-like racemes or solitary; calyx of staminate flowers 4-parted; calyx of



Fig. 339. Annual Mercury (Mercurialis annua). Staminate and pistillate branches. An acrid, poisonous plant. (After Faguet.)

the fertile flower 3-5 parted, subtended by a foliaceous bract; petals wanting in both staminate and pistillate flowers; stamens 8-16 united at their bases; capsule consisting of 3 globular 2-valved carpels, each 1-seeded. About 250 species chiefly tropical, 3 species in the central and eastern states. A. gracilens has smaller leaves than A. virginica. A. ostryaefolia has echinate fruit and occurs from New Jersey to Texas and Kansas.

Acalypha virginica L. Three-seeded Mercury

A smoothish or hairy annual from 1-2 feet high often turning purple, especially in the autumn; leaves ovate or oblong ovate, sparingly serrate, long petioled; sterile spike few-flowered; pistillate flowers 1-3 at the base of staminate peduncle; capsule 3-lobed subglobose; seeds ovoid, reddish striate.

Distribution. From Nova Scotia to Florida, Texas, Kansas and Minnesota. Poisonous properties. This has been sent to me several times as supposedly poisonous. It is distasteful to cattle and they refuse to eat it in the pasture.

8. Stillingia L. Queen's Root

Smooth upright herbs or shrubs; leaves alternate or rarely opposite, frequently with 2 glands at the base; flowers in spikes, apetalous; calyx 2-3 cleft or parted; staminate flowers, several together in the axils of the bractlets, stamens 2 or 3 pistillate flowers solitary in the axils of the lower bractlets; capsule 3-celled and 3-seeded. About 15 species of tropical America and the Pacific Islands.

Stillingia sylvatica L. Queen's Delight

A bright green herb 1-3 feet high; leaves nearly sessile lanceolate or elliptical, 2 glandular base; flowers lemon-colored subtended by small bracts with saucer-shaped glands; calyx cup-shaped; capsule depressed; seeds ovoid, light gray, minutely pitted and a flat base.

Distribution. From Virginia to Florida, Texas and Kansas in light sandy soil.

Poisonous properties. This plant is commonly used in medicine. It is said to be an efficient alterative. It contains an acrid resin sylvacal and an acrid fixed oil.

SAPINDALES

Trees, shrubs or herbs; petals usually present and separate; sepals usually distinct; stamens rarely more than twice as many as the sepals or fewer; opposite or alternate; ovary superior, compound; ovule pendulous. Contains many tropical plants, some with milky juice. In the family Buxaceae is the common box (Buxus sempervirens) which is used as a hedge plant and furnishes the best wood for wood engraving. The plant is an acrid poison. It is sometimes substituted for hops in the manufacture of beer and thus becomes the occasion of serious accident. The edible crowberry (Empetrum nigrum) belongs to the family Empetraceae and occurs far northward in America and Europe. The bladdernut (Staphylea trifolia) of Atlantic North America belongs to the family Staphyleaceae. The family Sapindaceae contains the balloon vine (Cardio-



Fig. 340. Three-seeded Mercury (Acalypha virginica). Supposed to be poisonous to cattle. (Ada Hayden.)

spermum Halicacabum), a well known ornamental climber, but a weed in the South. A refreshing drink is made fom the seed of guarana (Paullinia Cupana) of South America; it contains caffein, saponin and an acrid green fixed oil. The fruit of Sapindus Saponaria contains a great deal of saponin and is used as a substitute for soap. The soapberry tree (Sapindus marginatus) is used as a shade tree in the South where it is a native. A shellac is derived from the Indian Schleichera trijuga and marcassa oil is obtained from the seeds of the same plant. The Indians of Brazil use the honey collected by wasps from the flowers of Serjania lethalis to poison their arrows. It is also used as a fish poison and contains a narcotic principle which causes death. Another fish poison is furnished by the black seeds of S. curassavica of Brazil. The natives use the same substance for criminal purposes on man. The nectar obtained from the flowers is also poisonous. Lehmann lists as poisonous S. nodosa, which is used by the natives of Brazil as an arrow poison. The fruit of S. trifoliatus of India contains saponin. The same substance occurs in other



Fig. 341. Common Box (Buxus sempervirens). The plant is acridly poisonous. (After Faguet.)

species, notably in the seeds of the Brazilian Magonia. Narcotic principles occur in the following genera: Serjania, Nephelium, Magonia and Harpullia. The fruit of the litchi (Nephelium Lit-chi), a native of China and the Philippines and cultivated in the tropics, is something like a plum and is eaten fresh or dried. The Blighia sapida of West Africa is cultivated for its edible arillus; the Koelreuteria paniculata of China is cultivated as an ornamental plant.

The family Coriaceae contains the genus Coriaria. The leaves and bark of the C. myrtifolia of southern Europe contain much tannin which is used in dyeing. The C. ruscifolia of New Zealand contains a black dye. The fresh leaves are used in making an intoxicating drink. C. myrtifolia and C. thymifolia of Mexico contain a toxic principle known as coriamyrtin which resembles picrotoxin. Many species of the genus are poisonous. Coriaria sarmentosa, C. arborea, and the tree-toot (C. Tutu) of New Zealand are poisonous. Easterfield & Ashton * have isolated a crystallin glucoside called tutin $C_{17}H_{20}O_7$ which appears to be closely allied to coriamyrtin $C_{15}H_{18}O_5$. Tutu plants are highly toxic to animals that have not become immune by first becoming accustomed to small quantities. Blyth says:

For the native cattle in the Tutu districts apparently consume moderate amounts of the shrubs with impunity, whereas other cattle become seriously ill. Both coriamyrtin and tutin belong pharmacologically to the picrotoxin group of substances. Tutin is somewhat less toxic that coriamyrtin. There is first depression, followed by salivation; the pulse is slowed, the respirations increased in frequency, and finally, clonic convulsions occur: 129 mgrms. killed a kitten weighing 1 kilogramme in 40 minutes; 1 mgrm. induced in a cat, 2 kilogrms. in weight, a convulsive seizure, and the animal did not recover for 24 hours.

Other important families of this order will be described farther on.

Families of Sapindales

ANACARDIACEAE. Cashew Family

Trees or shrubs with acrid properties, milky or resinous juice; alternate or opposite leaves; flowers small, frequently polygamous, regular; calyx 3-7-cleft; petals of the same number; stamens as many or twice as many as the petals, inserted at the base of the disk; ovary 1 or sometimes 4 or 5-celled, and 1 ovule in each cavity; styles 1-3; fruit generally a small drupe; endosperm scanty; cotyledons large.

There are about 500 species in temperate and tropical regions. The cashew (Anacardium occidentale) is much cultivated in the tropics. According to Dr. Cock, the fleshy receptacles of the fruit are used in the West Indies in

^{*} Jour. Chem. Soc. Trans. 1901.

preparing conserves and have an acid flavor which is very palatable; the pericarp, however, contains an irritant substance cardol C₂, H₂₀O₂, which is black, acrid and vesicating, and is used to protect books and furniture from insects; cashew oil, equal to the finest almond oil and superior to olive oil, is also a product of the plant. The juice of the shell of the nut produces poisoning similar to that of poison ivy. The kernels of the cashew may be eaten raw or roasted like chestnuts, but the fumes coming from the roasting nuts are very caustic. The pistachia nut (Pistacia vera) produces a fruit about the size of a plum, which contains a seed much prized for eating. The mango (Mangifera indica) a native of India, is now cultivated in most warm countries for its fine edible fruit. The bark of many species, like the smooth sumac (Rhus glabra) of the North, and the European R. Coriaria, contains a valuable tanning material. The smoke tree (Rhus Cotinus) and the stag horn sumach (Rhus typhina) are frequently cultivated for ornamental purposes. The fruits of Spondias dulcis, S. purpurea and S. lutea are edible, the last of these is called the hog plum, being so named because the hogs are fond of it. The juice of another member of the family (Comocladia) causes an eruption similar to that from poison ivy.

Tannic and pyrogallic acids are derived from the Chinese Indian Rhus semialata. Chios turpentine (Pistacia Terebinthus) well known to the ancients, produces red galls that are used for tanning morocco leather. The mastic (Pistacia Lentiscus) native to the Mediterranean region, was formerly used for making varnishes.

The Querbrachia Lorentzii and Q. Balansae of Argentine and Paraguay produce a very hard red wood which contains a great deal of tannin and gallic acid. The fruit of Dracontomelon mangiferum of the Sunda islands is used much like lemons. The ink tree of India (Semecarpus Anacardium and Holigarna ferruginea) contain cardol.

The pepper tree (Schinus Molle) cultivated in California, is a native of Peru. The saw-dust of sneezewood (Ptaeroxylon ntile) produces sneezing.

Rhus L. Sumach

Trees or shrubs with alternate, simple trifoliolate or odd-pinnate leaves; small polygamous flowers in panicles; calyx deeply 5-parted; petals 5, spreading; stamens 5, inserted below the flattened disk, fruit small, 1-seeded. About 120 species in the temperate regions common in southern Africa. Some species are poisonous. The Japanese Rhus vernicifera and R. succedanea are cultivated in Japan for the lacquer which is taken from incisions made in the trees. Dr. White states that some of the embossed Japanese papers which are used in houses have caused severe inflammation, and according to Dr. H. N. Allen, natives as well as Europeans in the East are often affected with "varnish poisoning"

Rhus Toxicodendron L. Poison Ivy. Three Leaved Ivy

A climbing or trailing shrub, sometimes erect, with 3 leaflets; plant erect or climbing by means of its aerial rootlets; flowers inconspicuous, polygamous in loose and slender axillary panicles; fruit globular, glabrous, whitish and waxy, frequently remaining on the plant until late spring.

Distribution, This plant is distributed from Nova Scotia to Wisconsin, Utah, Arkansas and Florida.

Rhus Vernix L. Poisonous Sumac or Dogwood

A shrub or small tree with pinnately compound leaves; leaflets 7-13, obovate-oblong entire, smooth, or somewhat pubescent; flowers polygamous in loose slender axillary panicles; drupe white, globose, oblong.

Distribution. Found in the swamps from New England to Ontario to Minnesota, Missouri, to Louisiana and Florida.

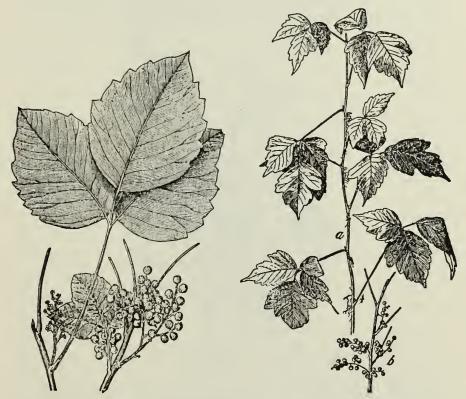


Fig. 342. Poison ivy (Rhus Toxicodendron). a, spray showing aerial rootlets and leaves; b, fruit—both one-fourth natural size. (Chesnut, U. S. Dept. Agr.)

Rhus diversiloba Torrey and Gray. California Poison Ivy

Nearly glabrous, erect or climbing shrub; leaflets 3 or rarely 5, obtuse or deeply pinnately lobed; flowers in loose axillary panicles; drupes subglobose.

Distribution. Common on the Pacific Coast from California to Washington. Poisonous properties. All three species are poisonous to many persons, some persons being much more sensitive to irritation from the plants than others Dr. J. C. White describes the effect of poison oak and poison ivy. He had collected freely of the plant for many years without any disturbance. Specimens were picked on September 28th, Oct. 6th, and Oct. 10th. He felt



Fig. 343. Poison Sumac (Khus Vernix), showing leaves, fruit and leaf-scars, one-fourth natural size. (Chesnut, U. S. Dept. Agr.)

a sensation of irritation about the eyes and throat from the specimens of poison oak collected on the first named date, while working with the plants under an Argand gas burner, but nothing further was noticed. No unpleasant symptoms were observed from the poison oak (R. venenata—R. Vernix) collected Oct. 6th. From that collected Oct. 10, he experienced symptoms similar to those observed Sept. 28th. On Oct. 12th a single vesicle with a peculiar thick cover appeared; the next day another and larger appeared on the wrist; two others came on the fourth day; others continued to appear up to Nov. 3rd, after which data the effervescences gradually subsided and were no longer perceptible. In another case described by Dr. White, the head was greatly swollen and features greatly disturbed. The skin of the face and neck was deeply oedematous and largely covered with vesicles of all sizes "many of which were seated on an erythematous base, others being still in their papular stage

of development." There were also large excoriations from which fluid was exuding freely, which on drying formed small crusts. The hands were also covered. "The subjective symptoms were great retching and burning of the parts affected, with the feeling of local discomfort, consequent upon so great swelling of the features. The eyes were nearly closed. There was a slight general febrile action." Dr. White also reports the death of a child from a severe case of poisoning from poison ivy. The child though healthy was not robust. A recent case was reported from Packwood, Iowa, where a fourteen-year-old girl died after terrible suffering from the effects of coming in contact with the ivy; her face alone showing the eruption from the poison.

Hundreds of persons are poisoned every year from the three species. Dr. White says:

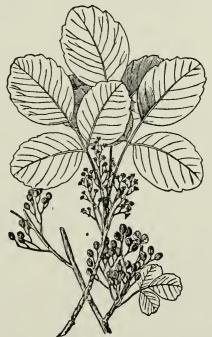


Fig. 344. Poison oak (Rhus diversiloba), showing leaves, flowers, and fruit, one-third natural size. (Chesnut, U. S. Dept. Agr.)

Taking the simple vesicle, with scarcely any erythema surrounding it or any very perceptible infiltration of the underlying tissues, as the type of the eruption, whether occurring singly or in groups, we may have in a small percentage an abortive attempt at vesiculation, and an arrest of the development at the papular stage—a failure, that is, of the free exudation to force apart the layers of epithelial cells; or a considerable infiltration into the papillary layer may elevate a cluster of the vesicles noticeably above the general surface. or they may be surrounded by a well-defined erythema or congestion of the tissue immediately surrounding them, in consequence mainly of the scratching and itching, which are the only subjective symptoms present.

In the severe cases, we have greater areas of simple erythema, a multiplication of the number of vesicles—either single or massed in close contiguity, and covering large surfaces, or by fusion forming blebs—a greater infiltration into the underlying corium, with proportionate distension of the capillaries and external redness, and a free exudation of serum into

the cutis. The overfilling of the vesicles causes a rupture of some of their epidermal coverings, and the discharge of their fluid contents upon the surface, forming moist, excoriated surfaces, covered in part with crusts.

With reference to a sequelae question of duration, there is a diversity of opinion. There is a popular belief that within a year after the first attack there will be a repetition of the original manifestations upon the skin which may be repeated for several seasons. Dermatologists think that a variety of cutaneous affectations are developed in consequence of the action of the Poison Ivy. Dr. White considers that there are good grounds for this belief and in referring to his exhaustive researches on the subject, states that he was unable to find a single instance on record of the poisonous Rhus on the lower animals. After placing a notice in the "Spirit of the Times," a physician wrote him that once or twice while hunting where ivy abounded, his dog's eyes had been closed by swelling which he attributed to the action, but he had never observed any eruptions. The poisoning has been attributed to toxicodendrol $C_{27}H_{42}O_{10}+4H_2O$.

Remedies. The most popular remedy is to wash with sugar of lead (acetate of lead). Prof. Chesnut says:

In practise it is not desirable to use strong alcohol, which is apt to be too irritating to a sensitive surface, but a weaker grade of from 50 to 75 per cent should be preferred. To this the powdered sugar of lead is to be added until no more will easily dissolve. The milky fluid should then be well rubbed into the affected skin, and the operation repeated several times during the course of a few days. The itching is at once relieved and the further spread of the eruption is checked. The remedy has been tried in a large number of cases and has always proved successful. It must be remembered, however, that the lead solution is itself very poisonous if taken internally.

Much has been said in regard to the relative poisonous character of these three plants. It has been generally claimed that the poison sumac is the most poisonous, and after it comes, first, the poison ivy and then the poison oak. These conclusions were arrived at from the occasional experience of individuals who were poisoned by handling one species when supposedly immune to others. Experience teaches, however, that immunity is somewhat variable in the same individual, and therefore these general statements can not be accepted without more careful experimental evidence.

Annie Oakes Huntington in her recent book on Poison Ivy and Swamp Sumach says regarding the treatment:

Soap, water, and a scrubbing-brush seem altogether too simple a method of treatment to advise for the painful eruption brought on by handling these two poisonous plants. Yet, if we begin with this old-fashioned country remedy and study the various methods of treatment from one generation to another, we return at last, through the most recent scientific investigations, to our original starting-point. The only effective measures are preventive ones; the only remedy is a wash which mechanically removes the poisonous oil from the skin. In this lies the sum and substance of the entire method of treatment.

She made an experiment in which it was shown that oily preparations spread the poison and that constant washing with soap and water removes the poisonous oil which causes the trouble. The toxic principle is soluble in alcohol and this may consequently be used. A weak solution, 50 or 75% is advisable, but the treatment must be renewed. One part of hyposulphite of soda to 3 parts of water is another good solution recommended by her.

Syme in Dr. Remsen's laboratory, has come to the conclusion that the active principle of Poison Ivy is a glucoside and not an unknown volatile oil, as stated by Pfaff. The glucoside as determined by Syme, is a compound of rhamnose, gallic acid, and fisetin. It can be precipitated by a lead acetate. Syme tested the toxic action of the various fractions upon himself and was able to determine the chemical nature.

In a recent paper by Dr. Ford there seem to be some evidences for concluding that immunity may be obtained. That such immunity exists may be taken from the clinical symptoms that different persons are sensitive to even small amounts of the poison and in other cases persons who have been poisoned become accustomed to it. Syme in his experiments upon himself found that after four or five months he was no longer susceptible to the poison.

The experiments performed by Dr. Ford are of interest. The experimental material was obtained in the alcoholic fluid extract of the native plant prepared by Parke, Davis & Co.

It had already been shown by Pfaff that the internal administration of his non-volatile oil produced definite lesions in rabbits, the animals dying of an acute nephritis at the end of 14 to 15 days. Occasionally the rabbits died in acute convulsions without any microscopic brain lesions. The subcutaneous administration of the fluid extract of Rhus Toxicodendron produces the same effect upon rabbits as those described by Pfaff. Rarely, the rabbits die in convulsions within 24 to 48 hours, but the majority of inoculated animals succumb in from 8 to 15 days. In addition to the nephritis an extensive necrosis and slough is found at the point where the poison is introduced beneath the skin. Following the inoculation we have a fairly long latent period during which the weight of the animals remains stationary. After seven or eight days in a typical case, the animal loses weight rapidly, the necrosis and slough develop, and the animal dies of the nephritis after the lapse of about two weeks. At times the skin lesions are less marked, the damage to the kidney being the important change; an intraperitoncal inoculation seems able to produce these kidney changes more rapidly than does the subcutaneous method. In addition to rabbits we have found that guinea-pigs are susceptible to the drug, the lesions being produced with greater certainty and regularity. With these animals the necrosis and slough at the point of inoculation are more extensive, while the animals die of the kidney changes in about the same time. The fatal dose of the poison can be estimated for both animals with tolerable accuracy. For guinea-pigs of 250 gram weight, 0.25 c.c. of the alcobolic extract always represents a fatal dose; and a guinea-pig of 350 gram weight practically never survives a dosage of 0.5 c.c.

The fatal dose for rabbits of 800 grams is 1 c.c. Certain rabbits of this weight die from smaller doses, but not regularly and larger animals show greater resistance. Animals of 1,800 to 2,000 gram weight occasionally survive 2 to 3 c.c. doses, but not more than this amount.

Experiments were made to determine whether animals which had withstood some doses of the poison were susceptible to the amounts of poison capable of killing untreated animals. The experiments were made with four guinea pigs, varying in weight from 450 to 900 grams and on rabbits varying in weight from 800 to 2800 grams. The initial doses were small; when the animals regained weight larger doses were given. It was found that the serum of immunized animals contained substances neutralizing the poisonous glucoside when both were injected into susceptible animals. Goats have been immunized and it is probable that immunity may be obtained in other animals.

How to Treat the Poison Ivy Patch. Various methods of treating the poison ivy have been tried. The iron sulphate, 100 pounds to a barrel of water, is only partially successful. Covering with tarred paper creosoted below is effective according to Dr. G. E. Stone. This writer has also shown that sodium arsenate (commercial) at the rate of 2 pounds to 10 gallons of water poured around the roots is effective.

Rhus glabra L. Smooth Sumach

A low shrub from 2-12 feet high, stems with large pith, brownish bark; compound leaves 11-31; leaflets pointed serrate, whitish beneath green above; flowers borne in a large panicle, greenish, polygamous; calyx small 5-parted; petals 5; stamens 5, inserted under the edge of a flattened disk; pistil with a 1-celled and 1-ovuled ovary; styles 3, terminal; fruit red, small 1-seeded.

Distribution. Common in Northern United States to Florida, Mississippi, Arizona, Utah, British Columbia to Nova Scotia. A troublesome weed at times in rocky soils.

Poisonous properties. There is a popular impression that the red drupes of this species are poisonous, but I know of no record where the eating of this fruit has produced poisoning. The fruit is decidedly acid.

AQUIFOLIACEAE. Holly Family.

Trees or shrubs with small, simple, mostly petioled, alternate leaves; flowers in axillary clusters, chiefly polygamo-dioecious; calyx minute, free; petals 4-8 or more; stamens free, as many as the petals; pistil 1, ovary superior, 4-8 celled; ovules 1-2 in each cavity; fruit a small berry-like drupe; seeds with small embryo; endosperm present.

A small order of 160 species. The American holly (Ilex opaca) from Maine to Florida, Missouri and Texas, is much used for Christmas decoration. The European holly (IlexAquifolium) is used for a similar purpose. The berries of this species, though eaten by birds, are said to be poisonous. Whether the seeds of the American species are poisonous is not known although it contains the principle ilicin. The Ilex Cassine, which ocurs from southern Virginia to Florida and Louisiana along the coast, was used by the Indians during their religious ceremonies to make what they called their "black drink," an emetic intended to clear the head and stomach. It contanis caffein. The yerba or maté (Ilex paraguensis), native to southern Brazil and Argentine Republic is used like the Chinese tea and is stimulating. Properly the term "maté" is applied to the drinking cup made from a small gourd. Yerba is an important article of commerce in South American countries. The leaves contain the same active principle, caffein, that is found in the tea. Two other species I. theezans and Symplocos lanceolata also furnish the maté. The wood of the larger trees of the genus, like Ilex opaca and Ilex Aquifolium is white and is used by cabinet makers.

CELASTRACEAE. Staff-tree Family

Shrubs, or trees, with simple leaves; stipules small or absent; flowers regular, usually perfect; calyx 4-5-lobed; petals 4-5; stamens inserted on a flat or lobed disk; pistils with 3 or 5-celled ovary; ovules 2 in each cavity; fruit 2 to 5-celled, fleshy; seeds with an aril, embryo large, and fleshy endosperm. About 350 species of wide distribution.

The burning bush or waahoo (Euonymus atropurpurens) is a well known native, frequently cultivated and is a most desirable shrub. The Catha edulis of Arabia is extensively cultivated and is used as coffee by the Arabs. The leaves are also chewed by the natives, having a stimulating effect similar to that of cocain. It contains the alkaloids cathin and celastrin. The Elaeodendron australe of New South Wales is used for cabinet work. The genus Pachistima is represented in the Rocky Mountains by Pachistima Myrsinites, and in the Alleghany Mountains by Pachistima Cambyi both of which are pretty shrubs.

Celastrus L. Staff-tree. Bitter-sweet

Mostly climbing shrubs; leaves thin; flowers racemose or paniculate; polygamo-dioecious; calyx 5-lobed; petals 5, crenulate, inserted under the disk;

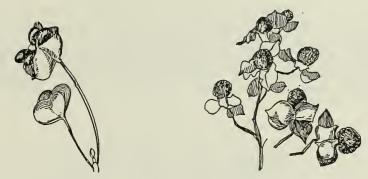


Fig. 345. Waahoo (Enonymus atropurpureus). 345a. Bitter sweet (Celastrus scandens).

pod globose, orange color, 2-4-celled dehiscent into as many valves; seeds enclosed in a scarlet aril; endosperm fleshy. About 30 species.

Celastrus scandens L. Shrubby or Climbing Bittersweet

Chiefly climbing shrubs with alternate leaves; flowers small, polygamo-dioecious; calyx 5-lobed; petals 5; stamens 5, inserted under the 5-lobed disk; capsule globose, orange-colored, 3-celled and 3-valved; seeds 1-2 in each cell, enclosed in a pulpy aril. About 30 species. The *C. articulatus* and several other species are commonly cultivated and are hardy. Several species are natives of the Cape of Good Hope.

Distribution. From Quebec to Manitoba in Canada, and from Kansas to Indian Territory, New Mexico and the Carolinas.

Poisonous properties. The aril of "Bitter-sweet" has a sweetish, somewhat disagreeable taste. The leaves of the plant are said to be poisonous to horses. The plant Euonymous contains the amorphous bitter, odorless substance, euonymin, which acts as a powerful heart poison. The waahoo acts as a drastic purgative. The symptoms are those of deathly nausea, vertigo, prostration and cold sweat.

ACERACEAE. Maple Family

Trees or shrubs with opposite, simple or compound leaves; flowers polygamous or dioccious in cymose or racemose clusters; calyx 5-parted; petals of the same number or none; stamens 4-12, inserted on a fleshy disk; ovary 2-lobed and 2-celled; styles 2, fruit a samara, exalbuminous; cotyledons thin, folded. There are 3 genera and about 100 species most of them in the genus Acer, the maples being widely distributed in temperate regions. The maple, (Acer) is commonly used for the manufacture of furniture and for inferior finishings, floorings, etc. The most highly prized are the hard maples (Acer nigrum and A. saccharum). Sugar maple is also derived from these species. Curly maple is only a form of wood of these and of the A. macrophylhum of the Pacific Coast, which is also much prized for cabinet work. The maples, including the box elder (Negundo aceroides or A. Negundo), are also used for shade trees. The silver maple (A. saccharinum) is widely distributed in the United States. The red maple (A. rubrum) is less commonly used. The bark of A. rubrum was used by the Indians as a remedy for sore eyes.

HIPPOCASTANACEAE. Buckeye Family

Trees or shrubs; leaves opposite, petioled, digitately 3-9-foliolate; flowers in terminal panicles, irregular and polygamous; calyx 5-lobed or 5-cleft; petals 4-5, unequal, clawed; disk entire; stamens 5-8; ovary sessile, 3-celled; ovules 2 in each cavity; style slender; capsule leathery, smooth or spiny, 1-3-celled; seeds large shining; cotyledons very thick. Only 2 genera and 15 species in America and Asia.

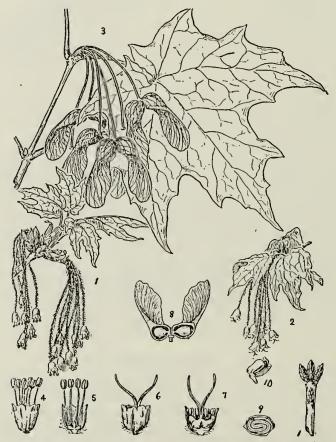


Fig. 346. Sugar maple (Acer saccharum), 1. Branch bearing staminate flowers. 2. Branch bearing pistillate flowers. 3. Fruiting branch. 4. Staminate flower, enlarged. 5. Longitudinal section of staminate flower, enlarged. 6. Pistillate flower, enlarged. 7. Longitudinal section of pistillate flower, enlarged. 8. Longitudinal section of fruit. 9. Longitudinal section of seed, enlarged. 10. Embryo, cnlarged. 11. Winter branchlet. 1, 2, 3, 8, 11, one-half natural size. (M. M. Chency in Green's Forestry of Minn.)

Aesculus L. Buckeye

Trees or shrubs with opposite petioled digitately 3-9-foliolate leaves; flowers in panicles, irregular, polygamous; calyx 5-lobed, lobes unequal; petals 4-5, unequal, clawed, stamens 5-8; filaments long, often unequal; pistil with 3-celled

ovary and two ovules in each cell; capsule leathery; seeds large with shining coat; cotyledons thick and fleshy. A small genus of 15 species native of America and Asia. The horse chesnut (Aesculus Hippocastanum), escaped from cultivation is planted for ornamental purposes, as are others of the genus, like the species described below and A. parviflora, a small shrub. By washing and boiling, the starch in the seed may be utilized, and this is done in France with the horse chesnut. The wood is light and brittle. The wood of the Ohio buckeye is used for making violins.

Aesculus glabra Willd. Ohio Buckeye

Trees with long-petioled leaves; rough and fetid bark; flowers pale yellow, in large panicles, polygamo-monoecious; calyx bell-shaped; stamens exserted, curved; petals unequal; fruit slightly prickly when young, smooth when old.

Distribution. Western Pennsylvania to Central Iowa, Kansas and Indian Territory.

Aesculus Pavia L. Red Buckeye

Shrubs with 5-7 digitate, nearly smooth, leaflets, acute or short acuminate, pubescent when young, becoming smooth; flowers in loose peduncles; calyx tubular, bright red; petals bright red.

Distribution. In fertile valleys from Virginia to Florida, Arkansas and southern Missouri.

Aesculus Californica Nutt. California Buckeye

Usually a shrub from 10-15 feet high, or occasionally a tree from 25-40 feet high, 3 feet in diameter; leaflets 4-7, usually 5, smooth, oblong-lanceolate, acute, petiolate; flowers in a close panicle; calyx 2-lobed; petals somewhat unequal, white or palc rose, ½ inch long; stamens 5-7; ovary densely pubescent; fruit usually 1-seeded.

Distribution. In California.

Poisonous properties. The leaves and fruit of the above species are regarded as poisonous. Many farmers claim that this is true only at certain seasons of the year. The seed produces sneezing and enters into the manufacture of snuff. The California species, according to Chesnut, causes abortion in cows. Dr. Rusby states that in southern states the seeds are crushed and thrown into water to stupefy fish just as the bark and roots of the relatives are in the tropics. Fatal cases of poisoning of children are reported from Texas. Suspicion has been attached to the common horse chestnut. The European chestnut is said to be useful in affording food for live stock, especially sheep and goats. This species contains aesculin $C_{15}H_{16}O_9+H_2O$, a glucoside found in the bark of many trees of the order Sapindacne; also the glucoside aesculetin CoHoO4; and paviin Co2HanO20; the testa of the seed contains quercetrin C27H30O17; argyraescin, an acrid, amorphous glucoside; aphrodaescin, also an acrid, amorphous principle; and saponin C32H54O18, a glucoside which is also found in the roots of Polygala Senega, and other plants. Dr. Millspaugh states that the horse chestnut causes inflammation of the mucous membranes of the respiratory and digestive tracts, and especially of the rectum; constant burning in the stomach and epigastrium, followed by nausea, retching, and violent vomiting with great tenderness and colic throughout the ab-



Fig. 347. Horse Chestnut (Aesculus Hippocastanum). Flowering branch. Entire flower. Stamen. Pistil. Entire seed and longitudinal section. Wood used in making violins. (After Faguet.)

domen, are markedly present. The buckeye is an irritant of the cerebro-spinal system, the more prominent symptoms being confusion of mind, vertigo, stupe-faction and coma.

BALSAMINACEAE. Balsam Family

Succulent herbs; leaves alternate; thin, petioled; flowers axillary showy, irregular; sepals 3, the two lateral small, green; the posterior large and petallike, spurred; petals 3 or 5, some 2-cleft; stamens 5; ovary oblong 5-celled; style short or none; stigma 5-toothed or 5-lobed; ovules several in each cell; fruit a capsule in *Impatiens*, coiled elastically, expelling the seeds; seeds



Fig. 348. Ohio Buckeye (Aesculus glabra). Causes irritation of the cerebrospinal system. (Lois Pammel.)



Fig. 349. Red Buckeye (Aesculus Pavia): a, flowering branch; b, seed; both two-ninths natural size. (Chesnut, U. S. Dept. Agr.)

ridged; embryo straight. About 200 species mostly of tropical Asia. One genus with 2 species is native to eastern North America. The Balsam (Impatiens Balsamina) is frequently cultivated. The sap of some species contains a dye.

Impatiens L. Jewel Weed

Succulent herbs with simple, thin, petioled leaves; sepals 3, the 2 alternate small, green, the posterior one largest, and forming a spurred sac; petals 5, or 3, with 2 of them 2-cleft into dissimilar lobes; stamens 5, short; ovary oblong, 5-celled; fruit an oblong or linear capsule, dehiscent elastically into a coiled valve, scattering the seeds; endosperm none; embryo nearly straight; cotyledons flat.

Impatiens biflora Walt. Spotted Touch-me-not

A glabrous annual from 2-4 feet high; leaves ovate or elliptical, pale and glaucous beneath; flowers orange-color, thickly spotted with brown; peduncles 2-4 flowered.

Distribution. In moist grounds from Eastern Canada to Florida, to Kansas and northward to Oregon and Alaska.

Poisonous properties. Dr. Schaffner states that the plant is emetic and suspected of being poisonous to stock.

RHAMNALES

Shrubs, vines or small trees; leaves generally alternate; flowers small regular; sepals generally more or less united; petals distinct or wanting; stamens as many as the calyx lobes and alternate with them, opposite the petals when present; ovary superior, compound; ovules erect. They contain two families, the Rhamnaceae and Vitaceae. The genus Vitis embraces 40 species found in warm and temperate regions. The European grape (Vitis vinifera) native from Eastern Europe to Central Asia is now widely cultivated in California, Spain, Germany, the Cape region and elsewhere. The Worden and Concord grape (V. Labrusca) of eastern North America are also widely cultivated. Other species are, the small grape (V. aestivalis), the wild blue grape (V. bicolor) native from New York to Wisconsin, the southern fox grape (V. rotundifolia) with musky flavor, cultivated in the South, the cultivated northern fox grape (V. riparia) with very fragrant flowers. Improved forms are the Janesville and Clinton. The fruit of the mustang grape (V. candicans) of Texas is very acrid. The Virginia creeper (Psedera quinquefolia) is a well known ornamental climber. The Boston or Japan Ivy (P. tricuspidata) native of Japan, is a handsome climber scarcely hardy north. The P. heterophylla, another ornamental from China and Japan is a hardy plant with small blue berries. It does not cling. The Vitis inconstans of Japan contains toxicodendrol and is poisonous.

RHAMNACEAE. Buckthorn Family

Shrubs, often climbing; or trees, often thorny, with astringent or bitter qualities; leaves chiefly alternate; stipules small, deciduous; flowers in cymes or panicles, small, regular; calyx perfect or polygamous, 4-5-toothed; petals 4-5, inserted on the calyx, or none; stamens 4-5, inserted on a disk which lines the calyx tube, which is often united with the single 2-5-celled ovary; ovules 1 in each cell; fruit often mucilaginous and drupaceous. A small family of 550 species, of temperate and warm regions.

The supple-jack (Berchemia scandens) is a pretty climber of southern woods. The buckthorn (Rhamnus cathartica) is frequently cultivated for hedges in the north. The juice of the unripe drupe was formerly used for staining maps and the ripe drupe is the sap green of painters. The sap has strong purgative properties. This and R. Frangula are local irritants. This plant contains rhamnetin, $C_{16}H_{12}O_{7}$, the rhamnin of earlier authors, a glucoside found in the berry. Dyes are obtained from R. infectoria and a dye for silks from R. lahurica and R. tinctoria. The R. Frangula contains frangulin and is an ornamental shrub, the charcoal from which is used for making gun powder. Probably all are more or less injurious. Cascara sagrada is obtained from R.

Purshiana, a shrub or small tree native to the Pacific coast which contains purshianin and is an excellent laxative. The Paliurus australis contains oil of wintergreen. The jujube (Zizyphus sativa) is used as food in South Europe and Western Asia. The Chinese Z. Jujuba is extensively cultivated in India and China. The fruit of Z. Lotus is made into a kind of bread, used by the natives of West Africa, but the ripe fruit is said to be injurious. The root of the New Jersey tea (Ceanothus americanus) is used as an astringent and expectorant and is also said to have been used in place of tea during the Revolutionary war. According to Greshoff the seeds of Ceanothus americanus, and Ceanothus ovatus contain a small amount of saponin. It is therefore not strange that some members of the family are poisonous. The genera Zizyphus, Tapura and Gouania furnish fish poisons. Saponin occurs in Gouania tomentosa of Mexico. Colubrina fermenta is used as a substitute for hops. The wood of Colletia spinosa of South America contains a bitter principle.

The brownish-black berries of coyotillo (Karwinskia Humboldtiana) of Texas are said to be very poisonous and Dr. A. Mitchell of San Antonio writes me that the plant is poisonous to goats.

MALVALES

Herbs, shrubs or trees; leaves simple, generally alternate; flowers regular, usually perfect; sepals separate or more or less united; corolla polypetalous or rarely wanting; stamens usually numerous; ovary superior compound; placenta axial. The more important families of this order are Tiliaceae, Malvaceae, Bombaceae and Sterculiaceae. The first family contains the basswood (Tilia americana), a well known timber and ornamental tree of North America, used for making boxes, lumber, excelsior, etc. T. cordata is also frequently cultivated under the name of Linn tree. The inner bark of this produces an elastic fiber. Several species of the genus Grewia are used as fish poisons. Jute is derived from Corchorus capsularis of the East Indies and is a valuable fiber. The broomweed (C. siliquosus) of the West Indies furnishes a substitute for tea. The fruit of Apeiba is edible. Corkwood (Ochroma Lagopus) of the family Bombaceae is used by the fishermen of Trinidad on their nets in place of cork and is one of the lightest of all woods. The monkey-bread trec (Adansonia digitata) produces valuable fiber in its bark. The silk cotton or kapok tree (Eriodendron anfractuosum) furnishes a soft fiber used in upholstery. The seed known as "kapok" seed is used in the Celebes as food and in making oil cake, and according to Reinders and Kobus, is an adulterant of linseed. The Durio zibethinus of the Malayan region produces a large edible fruit. The family Sterculiaceae of tropical countries includes the cacao (Theobroma Cacao) which produces the well known cacao beans. Cacao is a nutrient food and contains the alkaloid theobromin, C, H, N, O, a caffein alkaloid. The cola nut (Cola acuminata) of West Africa is a stimulant. The negroes of Brazil used large quantities because of its stimulating properties. It is also used for similar purposes by the negroes of the south ,and the "Cola habit" is increasing among the negroes of that region. It is probable that the cola is frequently adulterated with injurious ingredients. It is a muscle stimulant used by the Alpine climbers of Europe. Fresh cola nuts do not contain caffein, but a glucoside kolanin which is converted into kolarea C14H13(OH)5. An



Fig. 350. Jute Plant (Corchorus capsularis). A valuable fiber plant from the East Indies. (From American Agriculturist.)

oil is manufactured from Sterculia foetida. The Abroma augusta yields a fine fiber,

To the family Elaeocarpaceae of the same order belongs the Echinocarpus Sigun, a poisonous plant which contains hydrocyanic acid. Several members of this family are economic plants. The maqui fruit is obtained from Aristotelia Macqui, native to Chili. The seeds of Sloanca dentata are caten like chestnuts. The seeds of Muntingia are edible.

MALVACEAE. Mallow Family

Herbs, shrubs, or, in tropical countries, trees, with mucilaginous properties, tough fibrous bark and stems; leaves alternate and small; stipules small, deciduous; flowers regular and generally perfect; sepals 5, usually more or less united; petals 5, hypogynous; stamens numerous, monadelphous, several-celled; pistils several; styles united, projecting beyond the stamens above; ovary several celled; seeds nearly exalbuminous; embryo curved. A family of about 800 species of wide distribution and of great economic importance. The cotton plant is the most important member of the family and comprises several species, the most important in the United States being Gossypium herbaceum, which

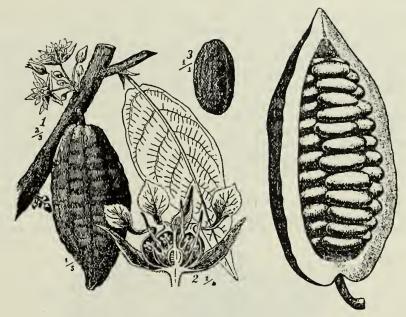


Fig. 351. Cacao Trec (*Theobroma Cacao*). 1. Branch with flowers and fruit. 2. Flower cut through longitudinally. 3. Seed. 4. Fruit with a portion of pericarp removed. The beans are well known articles of commerce. (After Wossidlo).

is cultivated for its fiber. The cotton fibers are plant hairs coming from the seed. Cotton seed is used to manufacture an oil serving in place of olive oil, and cotton seed meal, the latter a valuable stock food. The refuse material is used as a fertilizer. The highly explosive gun cotton is made by soaking cotton in sulphuric and nitric acids. Other cottons are Sea Island cotton (G. barbadense) and tree cotton (G. arboreum). The bark of cotton root has very active principles. Marsh mallow (Altheae officinalis) used in medicine as a stimulant and in confectionery, contains asparagin $C_4H_8N_2O_3H_2O$ and bassorin $C_{12}H_{26}O_{10}$.

Okra or gumbo (Hibiscus esculentus) is extensively cultivated in the south and in Europe for the young mucilaginous pods which are edible. The H. ficulneus is used in a similar way. The fleshy red calyx of the Jamaica sorrel (H. Sabdariffa) is used in making jellies and sauces; the fiber makes good cordage material.

The wood of the Cuba bast (Hibiscus elatus), native to the West Indies, produces a timber of greenish color used in cabinet work. The lace-like inner bark is used for wrapping and is known as lace bark. From the fiber of H. tiliaceus a strong paper can be made very cheaply. This is also used by the natives of the Pacific Islands in making ropes.

Fibers are obtained from Hibiscus tiliacens, H. cannabinus, Arena lobata, Abutilon indicus, Sida retusa and Napaea laevis. A beautiful wood is derived from the magar (Thespesia grandiflora) of Porto Rico, the color being red when fresh, black when dry. The hollyhoek (Althaea rosea) is well known in

cultivation, as are some species of the genus Abutilon and the poppy mallow (Callirboe involucrata and C. triangulata).

Chorisia of eastern South America furnishes a soft fiber. The seeds of Pachira macrocarpa indigenous to Brazil contain a valuable oil resembling that found in cacao; kapok oil is obtained from the seeds of Eriodendron anfractuosum. Musk seed is obtained from Hibiscus abelmoschus of the East Indies and is used in perfumery. The Malva moschata of Europe produces a similar odorous product. The cheeses or dwarf mallow (M. rotundifolia) is a trouble-some weed in gardens, waste places and barnyards. Saponin is found in the roots of Sida jamaicensis and Hibiscus Sabdariffa. The Sida paniculata is used as an anthelmintic. The ripe capsules of Queensland hemp (Sida rhombifolia) causes the death of fowls that feed on it.



Key for Malvaceae

| riowers involucrate. |
|---|
| Flowers small; seed solitary and not covered with cotton2 Malvastrum. |
| Flowers large; seed covered with cotton |
| Flowers not involucrate |

Gossypium L. Cotton

Herbs, shrubs, or rarely trees from 2-10 feet high; leaves alternate, palmately veined, and lobed, stipulate; involucre of 3 heart-shaped leaf-like bracts; flowers large, regular, white or whitish; sepals 5; petals 5; stamens numerous; anthers borne along the outside of the tube of the filaments; ovaries 3-5-celled, as many as the cells of the pod; seeds numerous bearing cotton. The description of one species only is given below, the Sea Island cotton (G.barbadense), cultivated in Florida and along the Gulf Coast. Tree Cotton G. arboreum is cultivated in the tropics. The fiber is long, silky and an inch or more in length. But little is produced, its use being restricted, it is said, to making thread for turbans for the priestly class.

Gossypium herbaceum L. Common Upland Cotton

An annual, 3-6 feet high; leaves with 5 short and roundish lobes; flowers large, pale yellow, turning rose color; seeds covered with cotton. Prof. L. H. Dewey thinks that our upland cotton should be referred to G, hirsutum (G. herbaceum), which is a native of Mexico. 1 The crop in India, according to Dr. H. J. Webber is derived chiefly from G. herbaceum, and in Egypt, the crop is obtained chiefly from G. barbadense. In warm climates, cotton is a perennial. Cotton was cultivated long before the Christian era. It is one of the most important crops of the world. Dr. Webber states that in 1792 the crop was 60,000 bales; in 1820, 6,000 bales; in 1860, the product increased to 4.483,311 bales, reaching 8,547,468 bales in 1892, and in 1904, 13,693,279 bales. In a single century, from 1804 to 1904, the crop increased from 130,000 bales, valued at \$13,-000,014, to 13,693,279 bales valued at \$557,147,306. In the early history of cotton cultivation, the seeds were not valued at all. Growers were troubled to know how to get rid of them. But in 1904 the seeds alone were valued at \$90,258,-227.86, making the total value of that year's crop, unmanufactured, \$647,405,-534.51.

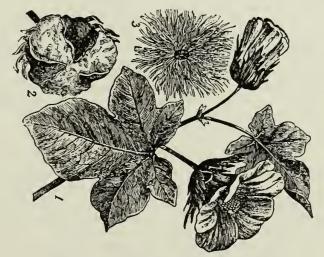


Fig. 353. Cotton (Gossypium herbaceum): 1, a plant with flowers and leaves; 2, a cotton boll; 3, seed. (Strasburger, Noll, Schenck and Schimper.)

Distribution. Commonly cultivated in the Southern States.

Poisonous properties. The root is commonly used in the south and tropical countries to produce abortion. Dr. Johnson says:

Cotton acts as an abortifacient. Its action upon the uterus is similar to that of ergot, and it is used instead of the latter in cases of uterine inertia during parturition, and in amenorrhoea, dysmenorrhoea, and seanty menstruation. Whether its action upon the system at large be similar to that of ergot is unknown but worthy of investigation.

^{*} As to the botanical statues of the different species of the genus Gossypium, the following papers should be consulted: L. H. Dewey Cyclopedia of Agrl. 2: 281; Advance article on cotton by Webber in an earlier part of the same work; G. Volkens, Die Nutzpflanzen Togos Notizblatt Königl, Bot. Garten Berlin, App. 22, No. 2, p. 60; O. F. Cook, Origin of the Hindi Cotton Cir. Bur. Pl. Ind. 42. (This last paper gives some of the literature); Watt. St. George, The Wild and Cultivated Cotton Plants of the World, London, 1907.

The fact that the bark of cotton root should be possessed of such active properties is an interesting and suggestive one, for it affords the only instance of specific virtues attributed to a plant of the order Malvaceac. It will be noted that it is only the bark of the root that possesses these properties, all of the other parts of the plant partaking of the bland character of the order—the seeds especially, which not infrequently possess the most active principles of plants, in this instance yielding a hland oil which is largely used as an adulterant of olive oil. Now in an order of plants of such strongly marked characteristics as the Malvaceae, the discovery of striking properties in any individual should stimulate investigation of other related individuals, for it may reasonably be assumed that investigation which starts with a rational clue will be more fruitful in results than if conducted entirely in a haphazard manner. Cotton seed is often injurious to animals.

In Friedberger and Fröhner's Veterinary Pathology, Capt. Hayes writes as follows:

Only young animals are usually affected. They display disorder of digestive and urinary apparatus: tympanites, diarrhoea (afterwards bloody), haemorrhage from the urinary organs, albuminaria, strangury and paralysis of the bladder. Duration is variable. Autopsy reveals in the acute form haemorrhagic gastro-enteritis, opaque swelling of the liver and kidneys, oedema of the lungs, dark red urine, collection of liquid in the cavities of the body. There is no enlargement of the spleen.

The liver is opaque and swollen. In chronic cases only, one finds general emaciation and dropsical phenomena. In dairy cows it induces garget and mammitis. Cotton seed oil cake contains a poisonous principle *ricin*.

The investigations Prof. M. B. Hardin of South Carolina made in 1892 indicated the presence of meta and pyrophosphoric acid in cotton seed meal. He suggested that these acids are poisonous. Dr. Crawford of the U. S. Dept. of Agrl. has recently investigated the subject and concludes that the pyroprosphoric acid is toxic. The seed from upland cotton is more toxic than the Sea Island. The toxic effect may be increased by heating; when the temperature rises high, this is due to the corversion of the orthophosphoric acid into the pyro form. The investigations of Dr. Crawford are very important.

Malvastrum Gray

Herbs with entire cordate or divided leaves; flowers solitary or racemose, short pedicelled; involucrate, or none; calyx 5-cleft; petals 5, notched at the end or entire; styles 5 or more; stigmas capitate; carpels indehiscent or imperfectly 2-valved; seed kidney-shaped. About 75 species, natives of America and South Africa.

Malvastrum coccineum (Pursh.) Gray. False Mallow

A low hoary, perennial herb, with dense silvery stellate pubescence; lower leaves pedately 3-5 parted; flowers small, red, in dense, short racemes, usually without bractlets; calyx lobes shorter than the pink-red petals; carpels 10 or more, indehiscent, rugose, and usually 1-seeded.

Distribution. From South Dakota to Texas, New Mexico to British Columbit.

Poisonous properties. Has been suspected of being poisonous, but there is no evidence to support this view. Profs. Chesnut and Wilcox regard the plant as non-poisonous. Some ranchers consider that it may be the cause of loco poisoning.

Abutilon (Tourn.) Mill. Indian Mallow

Herbaccous or shrubby plants, or in the tropics, trees with soft, pubescent leaves and stems; involucral bracts none; calyx 5-cleft; petals 5; styles 5 or

more; carpels 2-valved, 2-9 seeded; seeds reniform. About 90 species in tropical or warm temperate regions. Many of the species are under cultivation.

Abutilon Theophrasti Medic. Velvet-leaf

A stout, strongly-scented annual, 2-4 feet high, with tough, fibrous stems; leaves roundish, heart-shaped, tapering to a point, velvety; peduncles shorter than the petioles; flowers yellow; carpels 12-15, pubescent, opening at the apex; each valve beaked by a slender awn.

Distribution. Common in the northern states. Naturalized from India.

Poisonous properties. It is reported as poisonous; the strong odor is so very objectionable that it is not likely that much of the plant will be consumed by stock.

PARIETALES

Herbs, shrubs, or trees; flowers generally complete, perfect and regular or in some cases irregular; scpals distinct or united, imbricated or convolute; petals nearly always present and distinct; stamens mostly numerous; ovary compound superior or inferior in some; placentae mostly parietal. The order includes a number of important families. Caricaceae contains the pawpaw (Carica Papaya) the edible fruit of which is from 6-10 in. long; of yellow color and contains the alkaloid carpain C14H25NO2, acting similarly to digitalis, and a glucoside caricin; it also contains the enzyme papain, which resembles trypsiu in its proteolytic action, converting animal proteins into proteoses forming peptones. This does not occur in vegetables. These changes do not go on in acid and alkaline substances, but act best in a neutral medium. The most active changes occur best when the medium is from 53 to 40 degrees C. A similar ferment occurs in the leaves of Carica quercifolia of Argentina. Caryocaraceae includes gamboge (Garcinia Hanburyi) a cathartic; the mammey apple (Mammea americana) cultivated in the West Indies; the Calophyllum Calaba which furnishes the Calaba balsam, the mangostecn (Garcinia Mangostana) of the Moluccas, widely cultivated in the tropics. The Dipterocarpaceae includes the genus Dipterocarpus, which furnishes resin, and shorea, furnishes Chalia resin. "Piney rcsin" is obtained from Vateria and is used as a substitute for dammar. The Tamaricaccae contains the ornamental Tamarix gallica, an excellent honey plant; the Tamarix mannifera from which a manna-like sugar is derived; the Fouquiera splendens of Mexico frequently cultivated and producing the ocotilla wax; and the Myricaria germanica, the twigs of which are used as a substitute for hops. Cistacae includes the rock rose (Helianthemum canadense), the pinweed (Lechea minor) and the Cistus polymorphus furnishing laudanum.

The Begoniaceae includes the commonly cultivated genus of Begonia with 550 species. The Bixaceae includes Bixa Orellana which furnishes the Annatto used to color butter, silks, etc. In the Canellaceae, is Canella alba, from which is obtained the Canella bark of commerce, often called wild cinnamon and which is used as a condiment in the West Indies and Florida where it is a native.

The family Flacourtiaceae of the tropics includes a number of plants like Gynocardia odorata of India, Hydnocarpus venenata and the Kiggelaria africano of Ceylon, the seeds of which contain hydrocyanic acid. The cocos oil is obtained from Myroxylon and is used in perfumery. The Casearia esculenta of Asia and Australia is a purgative. The family Turneraceae contains a few medicinal

plants. A variety of Turnera diffusa furnishes a laxative. The Datisca cannabina of Southern Europe belongs to the family Datiscaceae and furnishes the yellow coloring matter datiscin used to dye silk; some members of the family Dilleniaceae are cultivated for their beautiful flowers. The Marcgraviaceae are occasionally cultivated and the Marcgravia umbellata is used in medicine; the family also contains the "Bitter-sick tree" Datisca glomerata which is used, according to Chesnut, by the Indians of California to poison trout.

Families of Parietales

| Caryx gamosepaious. |
|--|
| Throat of the calyx with a fringed crown |
| Throat of calyx without a crownLoasaceae. |
| Calyx with sepals; sepals generally distinct and persistent. |
| Flowers irregularViolaceae. |
| Flowers regular. |
| Trees or shrubs; leaves alternate |
| Herbs or shrubs: leaves opposite or whorled |

THEACEAE. Tea Family

Trees or shrubs with alternate simple leaves without stipules; flowers large, showy, regular, hypogynous, mostly axillary; sepals 5, or rarely more, often with 2 bracts; petals 5 or rarely more; stamens more or less monadelphous; anthers 2-celled; ovary 2-5-celled; ovules 2 or more in each cell; fruit a woody capsule; embryo large; endosperm scant. A small order of 160 species, mostly native of warm regions. The tea plant (Camellia Thea) is native to Assam, and extensively cultivated in Japan, China, and India. The Thea sinensis with the varieties viridis and bohea furnish tea. Successful attempts at cultivation have also been carried on in South Carolina. The black and green teas come from the same species. Tea is an important article of commerce in all civilized countries, Russia, England and the United States using large quantities.

The active principle found in tea is caffein or thein C₈H₁₀N₄O₂, a feebly alkaline, bitter alkaloid which is a cerebral and cardiac stimulant. Thea also contains the alkaloid caffeidin C, H, 2N, O; theophyllin C, H, N, O, +H, O; theobromin C, H, N, O,, which is a bitter alkaloid having a physiological action similar to that of thein. The principle alkaloid thein, or caffein, as it is known, is a feebly basic, proximate substance, obtained from the tea plant, from the dried seeds of coffee, and from some other plants. This alkaloid has no particular action upon the digestive tract, unless it is used in large quantities, when it may cause gastro-intestinal irritation. Caffein increases the blood pressure, causing the heart to beat more forcibly and rapidly. It is a certain and direct stimulant. It produces wakefulness and restlessness and stimulates the reasoning and imaginative faculties in man. In the lower animals, according to Dr. Winslow, it often causes "the most intense cerebral excitement and mania when given in large doses, - produces restlessness, increased reflex excitability and convulsions in the lower animals." Dr. Winslow states further that caffein is a spinal and muscle poison to the frog, and tetanic convulsions occur in the batrachian similar to those produced by strychnin, but there is also muscular rigidity. The symptoms of poisoning in dogs, cats, and mammals generally, are restlessness, occasionally vomiting in dogs, rapid breathing, primary reduction followed by rise in temperature, clonic or tonic convulsions, muscular weakness, and

general paresis. Tea contains some saponin, but it is found especially in Thea Sassanqua of China and Japan and a saponin-like substance called assamin occurs in T. assamica; the flowers of the former species are used to flavor tea. The flowers and leaves of T. Kissi are used as an insecticide.

The Camellia (C. Hongkongensis) is a well known evergreen shrub cultivated in green houses. Two east North American genera of the order are Stuartia and Gordonia.

The leaves of many plants are used as substitutes for tea, among them are: Gromwell (Lithospermum officinale), Willow herb (Epilobium angustifolium), Willow (Salix), Ash (Fraxinus sp.), European Mountain Ash (Pyrus Aucuparia), Mulberry (Morus alba and nigra), Coffee (Coffea arabica), Camellia (Camellia hongkongensis), Cherry (Prunus spinosa), Rose (Rosa canina), Strawberry (Fragaria vesca and virginiana), Meadow Sweet (Filipendula ulmmaria), Wistaria (W. chinensis), Hydrangea (H. Hortensia), Boxelder (Negundo aceroides), Oak (Quercus), Akebia (A. quinata), Blueberry (Vaccinium Myrtillus). In Russia the leaves of Vaccinium and Arctostaphylos are made into tea. In North America, in addition to plants previously named, the leaves of Labrador tea (Ledum latifolium), New Jersey tea (Ceanothus americanus), Oswego tea (Monarda didyma) and Mexican tea (Chenopodium ambrosioides), are frequently used.

In South America the following are tea substitutes: Lippia pseudothea, Stachytarpheta jamaicensis, Psoralea glandulosa, Myrtus Ugni, Symplocos Alstonia, Capraria biflora, Angraecum fragrans, and Eritrichium gnaphaloides.

In China Sageretia theezans is used as a tea substitute, and in Australia various species of Myrtaceae are used for the same purpose.

HYPERICACEAE. St. John's-wort Family

Herbs or shrubs or occasionally small trees with opposite entire punctate, dotted leaves, without stipules; flowers solitary or cymose, paniculate, perfect and regular; sepals and petals 4 or 5, sepals persistent; stamens numerous, hypogynous in 3 or 5 sets; ovary 1-7-celled, and as many styles; pod 1-celled with 2-5 parietal placentae; seeds numerous, small; endosperm absent. About 275 species, some cultivated for ornamental purposes. Kalm St. John's-wort (H. Kalmianum) of the lake region, shrubby St. John's-wort (H. prolificum) and the great St. John's-wort (H. Ascyron) are desirable species for ornamental planting. The H. perforatum and other species yield a yellow dye. The Vismia viridiflora of Guiana yields a resin called American gamboge.

Hypericum (Tourn.) L. St. John's-wort

Herbs or shrubs with opposite punctate leaves; flowers borne in cymose clusters; sepals 5; slightly unequal; petals 5, yellow; stamens numerous, distinct or somewhat united in sets; fruit a capsule 1 to 5-celled; seeds numerous. About 200 species, of wide distribution.

Hypericum perforatum L. Common St. John's-wort

A perennial, much branched herb, with numerous sterile shoots at the base; leaves sessile, oblong or linear, black, punctate dots; flowers borne in cymose clusters, yellow sepals shorter than the yellow and black-dotted petals; stamens many in 3-5 clusters.

Distribution. Common in the eastern states, rare west of the Mississippi and in the southern states.

Hypericum punctatum Lam. Spotted St. John's-Wort

Herbaeeous, perennial, 1 or more feet high; copiously marked with black, pellucid dots; leaves sessile, oblong or ovate-lanceolate; cymes terminal, manyflowered; flowers crowded; petals pale yellow, large, longer than the oblong sepals; styles mostly not longer than the pods.

Distribution. In moist soil, Maine and Ontario to Minn., Florida, Kansas and Texas.



Fig. 354. St. Johns-wort (Hypericum perforatum). Flowering branch, fruit, section of fruit, part of leaf. Contains an oil and an acrid resin. (From Vesque's Traité de Botanique.)

Hypericum Ascyron L. Great St. John's-wort

Large stems, from 2-5 feet high, branched, 2-4-angled; leaves oblong, partly clasping; petals narrowly obovate; cymes terminal, few-flowered; flowers large, bright yellow; stamens in 5 sets; styles 5, united below; capsules ovoid, 5-celled; seeds small, numerous.

Distribution. From Vermont, Canada and Manitoba to Kansas, Illinois, eastward, also found in Europe and Asia.

Poisonous properties. It is believed that, where the plant is common, it is poisonous. It is said to cause eruptions on cows' udders and on the feet of white-haired animals. A writer in Breeders' Gazette reports the former species as poisonous. Prof. Chesnut says:

This species and the spotted St. John's-wort (H. masculatum, H. punctatum), were hrought into the Department by Dr. G. W. Bready, from Norwood, Md., who stated that five horses were poisoned in May, 1898, by eating meadow hay which contained nearly 50 per cent of these plants. One horse died from the effects of the poison, and two were killed to prevent their further suffering.

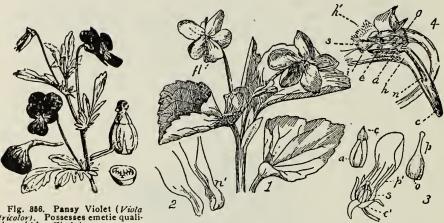
The oil of Hypericum is obtained from the European St. John's-wort (H. perforatum) and is apparently found in the black dots of the petals and fruits. The resin, however, found in the plant, is acrid and slightly bitter. The physiological action of the St. John's-wort is: mental depression and exhaustion; vertigo and confusion of the head; dilation of the pupils, and increased heart action.



Fig. 355. Great St. John's-wort (Hypericum Ascyron). (Charlotte M. King.)

VIOLACEAE. Violet Family

Usually herbs or rarely shrubs or trees, caulescent or acaulescent, with alternate, simple, entire or lobed leaves with stipules; flowers mostly irregular; sepals 5; corolla of 5 petals, 1-spurred, hypogynous; stamens 5, short, filaments broad and flat, often cohering with each other around the pistil; ovary simple, 1-celled, with 2 parietal placentae; fruit a capsule; seeds anatropous. About 300 species, of wide distribution. The best known is the pansy (Viola tricolor) running into numerous varieties, native to the Old World. It is sometimes used in skin diseases. The fragrant violet (V. odorata) is also frequently cultivated. Some of our native species are very handsome. Among these are the bird-foot violet (V. pedata), common in gravelly soils from Maine to Florida, west to Minnesota and Iowa and the V. pedatifida from Illinois to Kansas and Minnesota. Our most common eastern yellow violet is V. pubescens; the V. Nuttallii, also yellow, occurs from central Kansas westward. The white Canadian violet (V. canadensis) is common in the north and in the mountains. V. striata is common from the New England states to Minnesota and Missouri. The white-flowered violet (V. blanda) occurs in swamps. Brazilian ipecac is



tricolor). Possesses

Fig. 356a. 1-2, Yellow violet (Viola pubescens). 2, Spurred petals. 3-4, Pansy. c, Caylx. p, Pistil. a, Another spur.

derived from Hybanthus Ipecacuanha. The sweet pulp of the fruit of the Leonia glycycarpa of Peru is eaten by the natives; the fruit being about the size of a peach. We shall describe only a single species characteristic of the order, viz., the common pansy, which is a common weed in the South.

Viola L.

Acaulescent or leafy-stemmed herbs; annual or perennial; flowers solitary or rarely 2, smaller than the cultivated pansy; sepals 5; petals 5; unequal, the lower spreading at the base; stamens 5; anthers erect, united; in many of the species early blossoms are conspicuous, the later being cleistogamous.

Viola tricolor L. Heart's-ease.

Plant usually smooth; stem angled, branched; leaves roundish, or the lower oval, often heart-shaped; petals variable in color or variegated, yellow, whitish, violet-blue, and purple.

Distribution. Common in dry or sandy soil from New England to Kansas, especially southward. Also common on the Pacific Coast.

Poisonous properties. The substance violin, an acrid, bitter principle, has been extracted from the above species, and also is found in some of our native varieties. It is a pale yellow, bitter powder. The substance violaquercitrin C₁₂H₁₂O₂₄ is a coloring matter which is found in the pansy. The emetic effect of violets is well known and is supposed to be due to the presence of violin. Dr. Millspaugh states:

The most characteristic symptom of its action is an offensive odor of the urine, like that of the cat. The pains caused by this drug are of a stitching character, while its action seems spent almost entirely upon the skin, and the male sexual organs. On the skin it causes burning, stinging, and itching, followed by breaking down of the tissues into either squamous spots, or any grade of incrusted eruptions; the eruption pours out a thin yellow fluid.

Dr. Schaffner states that the sweet violet is somewhat poisonous, the underground parts being emetic and cathartic.

Dr. Rusby says:

Many violets are noted for their ipecae properties, yielding a glucoside called violin long confused with the emetin yielded by ipecac. They may be classed among the emetico-cathartics, and a large quantity might easily be productive of serious results to a child.

According to Spatzier, the seeds of violet contain myrosin and a glucoside.

PASSIFLORACEAE. Passion Flower Family

Shrubs or herbs climbing by axillary tendrils; leaves alternate, simple, generally 3-lobed; flowers perfect, regular, axillary; calyx tube persistent; petals usually 5, inserted on the throat of the calyx tube, which is fringed with a crown of a double or triple row of long, slender fringe; stamens 5, monadelphous, enclosing the stipe of the ovary; pistil 1; ovary with 3-5 parietal placentas; styles 1-5; fruit a berry or capsule, usually many-seeded.

A small family of about 300 species of warm and tropical regions. The common blue passion flower (Passiflora caerulea) of South America is frequently cultivated. The P. edulis, native of the West Indies, about the size of a hen's egg, is eaten; the grandilla (P. quadrangularis) producing a fruit 6 inches long, often weighs 3 pounds. The common maypop of the South (P. incarnata) with a fruit about the size of a hen's egg, is eaten. This species and the P. lutea are sometimes weedy. The Tacsonia yields hydrocyanic acid.

LOASACEAE. Mentzelia Family.

Herbs with rough, often stinging hairs, leaves without stipules; flowers regular, perfect, whitish, yellow or reddish; calyx tube adherent to the ovary, lobes 4-5; petals 4-5, inserted on the calyx; stamens numerous; pistil 1, usually 1-celled, with 2 or 3 parietal placentae; fruit a capsule, 1-celled with the persistent lobes of the calyx; endosperm scant.

About 200 species, nearly all native to North America. The Kissenia, however, being found in Africa. Species of the genus Blumenbachia (B. grandiflora) produce pretty flowers and are cultivated in greenhouses. The Mentzelia gronoviaefolia, from Mexico and Texas, is also cultivated. The leaves of Mentzelia ornata and other species produce hooked hairs which are often annoying to man.

Mentzelia (Plumier) L. Mentzelia

Herbs with erect stems, alternate leaves with barbed hairs; flowers usually showy, terminal, solitary or clustered; calyx tube cylindrical or club-shaped, lobes persistent; petals 5-10, regular, spreading, falling, usually turning black in drying; stamens numerous, inserted on the throat of the calyx tube; styles 3, more or less united; capsule dehiscent at the summit, many-seeded; seeds flat. About 50 species, American, chiefly west of the Mississippi river.

Mentzelia ornata Torr. and Gray. Showy Mentzelia

A rough herb from 1-2 feet high; leaves oblong-lanceolate, deeply toothed or pinnatifid; flowers solitary, much larger than the lanceolate calyx lobes; petals 10, yellowish-white, 2-3 inches long; capsule 1½-2 inches long; seeds numerous, with narrow margin.

Distribution. Northwestern Iowa to Dakotas to Central Kansas and Texas. Injurious properties. The hooked hairs of the plant cause the leaves to stick to sheep, clothing, etc. Prof. Goodale of Cambridge, Mass., is quoted by

Dr. White, in his Dermatitis Venenata as saying: "Mentzelia has grown in our garden, and has always been amazingly irritating to us all. Some species are even said to have stinging hairs." The writer has had considerable experience in collecting these species in the West, and he has learned from experience that the barbed hairs are quite irritating. Dr. Halsted states that M, oligosperma has the same properties. This plant is frequently cultivated.

OPUNTIALES.

Fleshy plants, usually spiny with jointed stems; leaves small; flowers mostly solitary, regular; calyx tube adnate to the ovary with a many lobed limb; stamens numerous, inserted on the throat of the calyx; filaments filiform; ovary 1-celled; ovules numerous; fruit a berry. Contains the important family Cactaceae.



Fig. 357. Showy Mentzelia (Mentzelia ornata). The hispid hairs of this plant produce mechanical injuries. (Charlotte M, King.)

CACTACEAE. Cactus Family.

Fleshy plants, leafless or with small leaves; stems flattened, columnar or globular, generally abundantly spiny; flowers solitary, sessile, perfect; calyx tube adnate to the ovary, limb many lobed; petals numerous, imbricated in several rows, mostly distinct; ovary 1-celled; ovules numerous, anatropous, borne on several parietal placentae; fruit a 1-celled berry or a dry fruit; endosperm wanting or copious.

About 1000 species, chiefly in the regions west of the Missouri river. species occur in the arid regions of the southwest. One of the most interesting is the giant cactus (Cereus giganteus) of Arizona, with fluted columns 50 or 60 feet high. The common night blooming cereus (C. grandiflorus), with white flowers opening at night, is well known in cultivation. The most commonly cultivated species is C. speciosissimus, with crimson red flowers that open during the day. The old man cactus (C. senilis) is cultivated because of its long white hanging hairs. Species of the genus Echinocactus, with stem of many ribs, are often cultivated, the most common being E. texensis of southern Texas and Arizona and the E. Ottonis of Brazil. A very good quality of leather has been produced from E. Wislizeni and Cercus giganteus. Species of Mamillaria with tufted stems covered with nipple-shaped tubercles, are often found in cultivation. The Epiphyllum truncatum, from Brazil, with flattened, leaflike stems and flowers 2-3 inches long, and the Phyllocactus, native to South America and Mexico, are also cultivated. The large genus Opuntia, of over 150 species, is entirely American. The O. vulgaris, Mill, naturalized in southern Europe, extends from southern New England west and south; O. Rafinesquii extends from Michigan west; and O. polyacantha from Wisconsin westward. Several Mexican species are cultivated. The Indian pear or prickly pear (O Figure Indica) of the West Indies and South America, produces an edible fruit. Extensively naturalized in North America, South Europe and Asia and as hedge plants. It grows on the lava slopes of Mount Aetna, converting the lava into soil. The cochineal plant (Nopalea coccinellifera) a native of Mexico, is cultivated as the host of the cochineal insect, from which a scarlet carmine dye is obtained. O. Tuna, O. Dillenii and Pereskia also act as hosts of the same insect. The fleshy stem of some Opuntias, after the spines are removed, are used as stock food. The berries of some species like Opuntia Larreyi which belongs to the O. Ficus-Indica group, O. Striptacantha and Echinocereus stramineus, etc., are eaten. A recent paper by Hare and Griffith described many details of their uses. The Mexicans call them tunas; an alcoholic drink is also made from the fruit. Some species are used as soil binders in Texas and elsewhere.

Injurious properties. The barbed trichomes penetrate the flesh and are difficult to remove. Death of animals has been caused by "hair balls" phyto-bezoars from them being formed in the stomach. Dr. William Trelease, who had occasion to examine the "hair balls" produced in some Mexican animal, gives the following account of this phytobezoar:

"The hair balls were a little over 3½ inches in diameter and weighed 7½ ounces. One ball was probably 4 inches in diameter." It was stated by the physician who sent them to Dr. Trelease that 16 such balls had been taken from the stomach of a bull at the Hacienda de Cruzes; it appears that the chief food of cattle at this time of the year consisted of Opuntias, and that the particular animal in question being allowed to roam at large, sought such food as could be found. These phyto-bezoars were brown in color, "and in appearance somewhat suggest felt or rubbed sole leather, and on examination prove to be composed, aside from the small nucleus at the center, of the barbed hairs with which the pulvini of the Platopuntias are armed. To the barbs with which these hairs are covered is due their power of felting together, and there is every indication that, starting about some small nucleus of vegetable fiber, they have been compacted into the dense, felty texture by the visceral movements of the

animal, to which, causing friction against one another, their perfectly round form is attributable." As is well known, the *Opuntias* produce spines and two kinds of trichomes. In the *Cylindropuntias*, each spine is invested by a deciduous sheath, "which is downwardly barbed, so that a person or animal brushing carelessly against a plant is certain to remove some of the barbed sheaths." In the *Platopuntias*, to which the ordinary flat-stemmed prickly pears belong, the spines, when present, are destitute of such a sheath. The protection to the plant is afforded simply because of their rigidity and pungency. The spines have their origin in pulvini, and in this particular genus of cacti are coated with delicate flexible hairs, divided into partitions. These hairs are lightly attached to the epidermis of the plant, so that when the pulvinus is touched they are almost certain to be removed in considerable numbers. The points of the stiffer hairs penetrate the skin, the barbs with which they are closely beset preventing their ready withdrawal. Dr. Trelease, in summing up the injurious effect of cacti, says:

It is a frequent practice in Texas to cut the branches of cacti which are fed to stock into half-inch lengths. In this way, every one of the obliquely set longer spines of Opuntia Engelmanni (and of some other species which are so used) is almost certain to he cut off, so that the danger from the spines is removed. This treatment, however, does not destroy the barhed the practice, in some places, to roast the fragments as a means of completely removing the spines and barbed hairs, but this is objected to hy some feeders, hecause the roasting has heen asserted to add to the laxative properties of the cactus. Where some such treatment has not

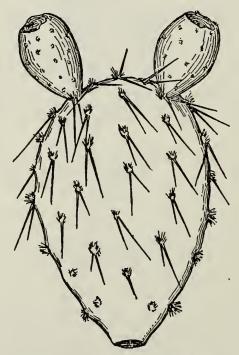


Fig. 358. Prickly Pear (Opuntia Engelmanni), from the harhed trichomes of which phytobezoars are sometimes formed. (U. S. Dept. Agr.)

been resorted to, injury to the animals not infrequently results; and in the bulletin referred to, Dr. Vasey gives a number of instances in which cattle have died from an accumulation of spines in the mouth and stomach, an effect somewhat comparable with that caused by the awns of *Hordeum* when cattle feed upon these.

The eating of the fruit of some species of Opuntia produces diarrhoea.

Under the name of Pellate (Anhalonium sp.), the Indians of the Rio Grande Valley of Mexico have for ages used the tops of this plant which they commonly call "mescal button" or "mescal bean." The use has extended to Indians in Oklahoma and Indian Territory and, it is said, to the Tama Indians of Iowa. The Kiowa Indians use 14-15 grams (4-5 buttons) to produce the peculiar sensations. The so-called mescal beans are 1-11/2 inches long and about 1/4 inch in diameter, brittle when dry, but soft when moistened. They have a bitter, disagreeable taste. Prentiss and Morgan were the first to call attention to the character of the drug dried from Anhalonium. During intoxication, the pupils become dilated, there is muscular relaxation, the pulse is somewhat slower, there is loss of sense of time, partial anaesthesia, weakened heart action; in some nausea and vomiting, and wakefulness. In man the influence has been described as causing an incessant flow of visions of infinite beauty, grandness, and variety of color and form. Intoxication closely resembles that produced by Cannabis indica. Dr. Lewin found that an aqueous extract given to lower animals produced convulsions causing death by respiratory failure.

The A. Lewinii contains the alkaloid anhalonin $C_{10}H_{17}NO_1$, mescalin $C_{11}H_{17}NO_3$ and anhalonidin $C_{12}H_{15}NO_3$. The A. fissuratum contains pellotin. Although this substance, according to some authors, is inactive, it has been used as a calmative on insane patients and in many cases causes sleep to come on. The cactin found in some species is a cardiac stimulant. It appears also that in addition to the above species the same or allied substances occur in A. prismaticum, A. Williamisii and A. Jourdanianum.

Anhalonium is closely related to the genus Cactus. It bears a dense penicilate tuft of long soft hairs which persist above the apical region of the plant as matted wool.

According to several recent investigators, especially Kauter and Heyl, alkaloids scem to be widely present in the family Cactaceae. Pectenin is found in a species of Cereus; pilocerein $C_{30}H_{44}N_2O_{57}$, occurs in Pilocereus Sargentianus; the alkaloid pellotin $C_{11}H_{12}NN(OCH_{32})OH$, is found in species of Anhalonium; and Lophophorin occurs in Anhalonium Lewinii and allied species. The alkaloidal substances appear to the extent of 1.1 per cent in dried material. A. Lewinii is a cardiac and respiratory stimulant. Saponin also is found in several species of the family among them in Cereus gummosus. Quite a number of other species of the family are used in medicine. The night-blooming cereus (Cereus grandiflorus) contains a glucoside which acts much like Digitalis. The Opuntia Karwinskiana contains an astringent principle. Several species such as Rhipsalis and Opuntia have anthelmintic properties.

MYRTIFLORAE.

Mostly shrubs or trees. Leaves simple; flowers incomplete; calyx inferior, 4-5 lobed or entire; corolla usually wanting; stamens twice as many as the calyx lobes or fewer; ovary 1-celled; ovule 1.

Species of the genus Cuphea of the family Lythraceae are cultivated in the South as border plants; the cape myrtle (Lagerstroemia indica) of the East

Indies, of the same family, is a handsome ornamental shrub, the seeds of which contain a narcotic principle; Henna (*Lawsonia inermis*) yields a yellow dye which is used in Egypt and Arabia as a cosmetic for the hands.

Tannin occurs in the root of Lythrum Salicaria. Cuphea viscosa of Mexico contains a substance similar in action to Digitalis. The fresh leaves of Ammannia baccifera of India contain a vesicating substance.

The family Lecythidaceae contains the well known Brazil or Para nut (Bertholletia excelsa, and B. nobilis), the Sapucaya-nut from the monkey-pot tree (Lecythis allaria) and other species.

Narcotic and poisonous principles occur in the fruit of *Planchonia valida*, native to the Molucca Islands, and in the seeds of some species of *Lecythis*. The roots and fruits of Chinese and Japanese species of *Barringtonia* are used as fish poisons. The family *Punicaceae* contains the pomegranate (*Punica granatum*) from the orient, cultivated in green houses in the North and out of doors in California and the South for its acid fruit, which is about the size of a small apple. The bark is used as a vermifuge and is an active irritant, its medicinal properties being due to a number of alkaloids present, of which four

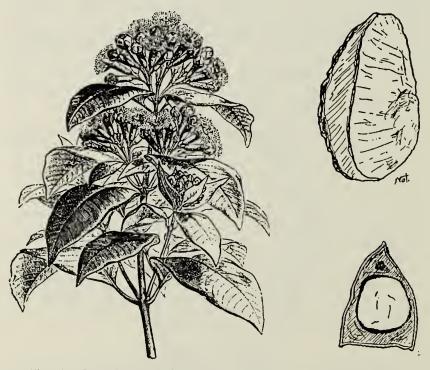


Fig. 359. Clove Tree (Eugenia caryophyllata). A native of the Molucca Isles. (After Faguet.)

Fig. 360. Brazil Nut (Berthollettia excelsa). (W. S. Dudgeon.)

have been isolated. The Mangrove (Rhizophora Mangle) belongs to the family Rhizophoraceae. Its wood is used in boat building. The Indian Almond (Terminalia Catappa) through its bark and leaves furnishes a black dye. The family

Myrtaceae contains the guava (Psidium Guajava), a most delicious fruit from which is made a jelly that is considered a great delicacy; the bay-berry (Amomis caryophyllata), from the dried leaves of which, when placed in water, an essential oil may be distilled which is used in the medication of rum; rose apple (Jambosa malaccensis) which yields an edible fruit and furnishes a material suitable for baskets, hoops, sugar casks, etc.; Jambuse berries (Jambosa vulgaris) cultivated in the tropics; the spices, piment or allspice, (Pimenta officinalis) and cloves, (Eugenia caryophyllata) from the Moluccas, common articles of commerce. Cloves contain an essential oil which is an excellent antiseptic and contains caryophyllin, $C_{10}H_{10}O$, and eugenin, $C_{10}H_{12}O_2$. Cajaput, (Melalenca leucodendron) is an excellent stimulant containing cajaputol, $C_{10}H_{10}H_2O$.



Fig. 361. Buffalo Berry (Shepherdia argentea). Fruit used for jellies, etc. (From American Agriculturist.)

The oil of Myrcia is obtained from the leaves of Pimenta acris of the West Indies and contains engenol and chavical. The volatile oil of Cheken (Engenia Chequen) contains cineol, pinene, a volatile alkaloid, and a glucoside. leaves of Myrtle (Myrtus communis) yield a substance used in perfumery. The Eucalypts, natives to Australia, are important timber plants, producing valuable wood for interior finishing, furniture, parts of vehicles, etc. A few species are extensively cultivated in California, of which Encalyptus globulus is one of the most frequently seen. The oil of eucalyptus and eucalyptol are used in medicine, and have valuable antiseptic properties. The E. rostrata produces Kino. The family Combretaceae contains many plants rich in tannin like the bark of Terminalia Catappa of Asia. The fruit of Myrabalons (T. Chebula) is rich in tannin. The T. fagifolia of Brazil has cathartic properties. of the plants of this family are used in arrow poisons. Several plants of the family Melastomaceae produce edible fruits; some of them like Tococa guiunensis yield a black dye. The only indigenous genus in the Northern States is Rhexia. The family Trapaceae contains the edible Chinese water-nut or water chestnut (Trapa natans) naturalized in New England and New York. The family Halorrhagidaceae contains several water plants of common occurrence like the water milfoil (Myriophyllum spicatum), the mare's-tail (Hippuris vulgaris) and the mermaid-weed (Proserpinaca palustris). Shrubs.

KEY TO FAMILIES OF MYRTIFLORAE

ELAEAGNACEAE. Oleaster Family.

Shrubs or small trees; leaves silvery, scurfy; flowers perfect or dioecious; calyx regular, simple, colored; calyx tube becoming pulpy and berry-like in fruit, strictly enclosing the achene; seed erect, ascending. A small order of 20



Fig. 362. Sea Buckthorn (Hippophae rhamnoides). Produces a berry. (After Fitch.)

species and 3 genera. The buffalo berry is well known. The Shepherdia argentea is a thorny shrub from 5-18 feet high, from Western Iowa and westward, the acid fruit of which was much used in early days for jams, jellies and pies. The Russian oleaster or wild olive (Eleagnus angustifolia), a well known ornamental shrub, adapted especially to the North West, is hardy and handsome. It has spiny branches which bear fragrant flowers. The wood is durable and makes an excellent post. The Goumi (E. multiflora) of Japan, produces edible fruit. The E. hortensis formerly included two species, the E. angustifolia and E. orientalis. Prof. Hansen has introduced the edible form of the shrub into the Northwest. The Arabs dry the berries and make a kind of cake. The Hippophae rhamnoides, an ornamental plant from Europe, is known under the name of sea buckthorn. It is commonly used for fish sauce in Russia. The plant is hardy in South Dakota. Prof. Hansen says that the berries of the tree contain a narcotic poison which is eliminated by boiling.



Fig. 363. Goumi (Elaeagnus multiflora). Cultivated for its edible fruit. (From American Agriculturist.)

THYMELAEACEAE. Mezereum Family.

Shrubs or small trees with aerid, tough, fibrous bark, simple opposite entire leaves; flowers in spikes or umbels, regular; ealyx petal-like, tube, urn-shaped; petals present or absent; stamens twice as many as the lobes of the calyx, and borne on it; ovary free, 1-celled and 1-ovuled; fruit a berry-like drupe; embryo straight; endosperm scanty or none.

About 400 species of wide distribution, most largely represented in Australia and South Africa. The leatherwood or moosewood (Dirca palustris) with a tough fibrous bark, is used by the Indians for thongs. The Mezereum (Daphne Mezereum) with fragrant flowers and bright red berries, of Europe, naturalized from Europe in New England, and the handsome D. Cneorum, with rosepink flowers, are cultivated. The berries and leaves of the Mezereum cause blistering; it is an aerid poison. Bark paper (D. cannabina) native of the Himalayas to Japan, produces a tough bark which is made into paper. Lace-bark (Lagetta lintearia) of Jamaica, with bark that separates into layers, was formerly used for veils, bonnets, etc. The bark of Wikstroemia viridiflora, of the Polynesian Islands, is used for making fishing nets, ropes, etc. The bark of Funifera utilis of Brazil, causes vesication like that produced by the Dirca palustris. Several exotic plants of the family are poisonous like the Pimelia trichostachya of Australia. The fruit and leaves of Gnidia carinata are emetic.

Daphne, L. Laurel

Shrubs, with alternate leaves, and small purple, pink, or white flowers in fascicles, heads or racemes; perianth tubular, with 4 spreading lobes; stamens 8, attached to the calyx tube; filaments very short; disk none; ovary sessile; stigma large; calyx deciduous or persistent. About 40 species, native of Europe and Asia.

Daphne Mezereum, L. Spurge Laurel. Lady Laurel.

A small shrub with young twigs somewhat pubescent; leaves thin, oblonglanceolate, or oblanceolate, petioled; flowers in sessile fascicles, very fragrant; perianth-tube pubescent, rose-purple; drupe red.

Distribution. Escaped from cultivation from Quebec to New York, native to Europe and Asia; frequently cultivated as an ornamental plant.

Poisonous properties. Some of the European species, like Daphne Cneorum contain aerid poisons. The plant produces blisters. The bark is used internally and in the form of an ointment. According to Loudon, in France the bark is applied to the skin for the purposes of a "perpetual blister." The bark, when fresh or when soaked in water, reddens the skin, when applied to it, and at length occasions vesicles followed by ulcers. Oesterlein remarked that all parts of the plant produced, on contact, irritation and inflammation. Schimpfky mentions this among the twenty-six important poisonous plants of Europe and states that the bark and berries are most poisonous, and that the pleasant odor of the flowers produces headache, for which reason, therefore, they should not be placed in a living room. Linnaeus seems to have recorded cases of poisoning from this plant. Daphne contains the glucoside daphnin, C₃₀H₃₄O₁₉, bitter and astringent, an aerid resin mezerein, daphnetin (CaHaO4)H2O, also with an astringent taste, coccognin C20H22Os, and the glucoside aesculin, C15H16Os+ H₂O. Friedberger and Fröhner state that animals poisoned by the Daphne have stomatitis, slavering, colic and a feeble pulse. In Europe the fruit is sometimes used as a substitute for pepper, in some cases with fatal results. Blyth says: "There are a few cases of poisoning on record, and they have been mostly from the berries. Thus, Linne has recorded an instance in which a little girl died after eating twelve berries. The symptoms observed in the recorded cases have been burning in the mouth, gastro-enteritis, vomiting (giddiness, narcosis, and convulsions, ending in death. The lethal dose for a horse is about 30 grms. of powdered bark; for a dog, the oesophagus being tied, 12 gms.; but smaller doses of the fresh leaves may be deadly."



Pig. 364. Mezereum (Daphne Mezereum). An acrid poison. (After Fitch.)

Dirca, L.

A small shrub with tough, fibrous bark; short-petioled leaves; flowers yellowish, in peduncled fascicles of 2-4 scaly buds at the nodes of twigs of the preceding season; stamens 8, borne on the calyx, the alternate ones longer; filaments very slender; perianth bellshaped, or funnelform; disk obsolete; ovary nearly sessile; drupe red, oval, oblong. 2 species known, 1 in Eastern North America, and 1 in California.

Dirca palustris, L. Leather-wood.

A shrub with yellowish green twigs; leaves obtuse; bud-scales 3 or 4, oval, with brown hairs, deciduous; style longer than the stamens.

Distribution. In woods and thickets, Eastern Canada to Minnesota, Central Iowa to Missouri and Florida.

Poisonous properties. The bark is acrid, like that of the Daphne; all parts of the plant having a nauseous, acrid taste. The principle, however, is unknown. The fresh bark applied to the skin causes redness and vesication, the sores thus produced being quite difficult to heal.

ONAGRACEAE. Evening Primrosc Family.

Herbs or rarely shrubs, with alternate or opposite leaves, generally without stipules, or stipules glandular; calyx adnate to the 2-4-celled ovary; petals 2-4; stamens as many as the petals or twice as many; ovules numerous. About 300

species. A few of the plants are medicinal. The great willow herb (*Epilobium angustifolium*) is occasionally used in medicine. The hairs of the seeds of some species are used in the Arctics as lamp wicks. Many species of the family are used for ornamental purposes, especially some of the western species of the genus, *Oenothera*, the *Clarkia elegans* of the gardens and the greenhouse *Fuchsia*. The genus *Oenothera* with many species, some southwestern and some western, contains very pretty plants.



Fig. 365. Leather-wood (Dirca palustris). This plant is well known in northern woods, especially on the banks of streams; occasion-ly found on high land. The bark is very tough and regarded as poisonous. (Charlotte M. King.)

Gaura, L. Gaura.

Annual, biennial, or perennial herbs with alternate sessile leaves; flowers white, pink or red in spikes or racemes; calyx tube narrow, prolonged beyond the ovary, the limb usually 4-lobed, reflexed; petals clawed, unequal; stamens usually 8, with a small scale before the filament, frequently declined; ovary 4-celled; styles declined; fruit hard and nut-like, 3 to 4-ribbed and angled. About 18 species.

Gaura biennis, L. Gaura.

An erect, soft, hairy or downy annual or biennial; leaves lanceolate or oblong-lanceolate, denticulate; flowers in slender spikes, white, turning pink; fruit oval or oblong acute at each end, 4-ribbed.

Distribution. From Ontario to Georgia, Arkansas and Nebraska and Minnesota.



Fig. 366. Willow-herb (Eptlobium augustifolium). Occasionally used in medicine. (After Fitch)

Gaura parviflora, Dougl.

A hairy, branching, soft pubescent annual from 2-5 feet high; leaves lanceolate or ovate lanceolate, acute or acuminate, sessile, repand, denticulate, covered with long soft hairs; the pinkish flowers about ½ inch long, borne in long flexuose spikes 2-3 feet long; fruit contracted at the base, obtusely 4-angled, glabrous.

Distribution. Common in dry soil from South Dakota to Missouri, Louisiana, the Rocky Mountain region and New Mexico and Mexico. A common weed along irrigation ditches.

Gaura coccinea, Pursh. Scarlet Gaura.

An erect or ascending, much branched, smooth or canescent herb; leaves lanceolate, linear-oblong, repand or entire; flowers red, turning scarlet; fruit canescent, terete below, and narrowed above.

Distribution. From Western Minnesota, Iowa, Nebraska, and Texas to Utah, Arizona, and Mexico.

Poisonous properties. The Gauras, or at least one species, the Gaura coccinca, have been suspected of being poisonous to live stock in the West. This is an excellent honey plant.

UMBELLALES

Herbs, shrubs or trees; flowers nearly always with petals; divisions of the calyx and petals usually 5; stamens 4 or 5; ovary compound inferior, adnate to the calyx; epigynous ovule 1 in each cavity.

FAMILIES OF UMBELLALES

| Fruit | a d | rupe | or | berry. |
|-------|-----|------|----|--------|
|-------|-----|------|----|--------|

| Flowers umbellate; stamens 5 |
|--------------------------------------|
| Flowers not umbellate; stamens 4 |
| Fruit dry splitting into 2 mericarps |

ARALIACEAE. Gingseng Family.

Herbs, shrubs or rarely trees; leaves alternate or whorled; flowers in umbels, heads or panicles; calyx tube adherent to the ovary; usually 5 petals inserted on the calyx; stamens as many as the petals, inserted on the disk; ovary 1 or more celled, 1 ovule in each cell; fruit a several-celled drupe.

About 50 genera and 450 species, of wide distribution. Genera common to eastern North America, China and Japan. Some of the species are occasionally cultivated for ornamental purposes. One of the best known of these is the Hercules Club (Fatsia horrida), native from Florida west to Missouri and Texas, and the common European ivy (Hedera Helix) well known in cultivation.

Few of the Araliaceae have injurious properties, however, the prickly spines of Fatsia horrida of the Pacific Coast, are quite irritating.

Several species of the genus Aralia and Panax are used in medicine. The



Fig. 367. Ginseng. (After Faguet.)

most important of these is the Ginseng, (Panax quinquefolium), which is native from eastern Canada to Alabama and in woods from Kentucky to Iowa, Missouri, Nebraska and Minnesota. This species is now widely cultivated, large quantities of Ginseng being exported to China, where the roots are in great demand. The Chinese Gingseng is P. ginseng. Several other species are used in medicine, among them the Spikenard (Aralia racemosa), and Wild Sarsaparilla (Aralia nudicaulis). These plants are not officinal, but they are quite commonly used. They have aromatic and stimulating properties. Barringtonin, $C_{18}H_{23}O_7(OH)_3$, is found in Barringtonia, a Japanese Aralia, Panax, and other genera. Aralian occurs in the roots of Fatsia horrida. The terpene, aralien, $C_{15}H_{24}$, occurs in Aralia nudicaulis. Some members of this order are occasionally weedy, especially the Sarsaparilla. Rice paper is made from Tetrapanax papyrifera, native to Formosa. It is a small tree about 10 feet high. The tree is cut into to obtain the pith, which is divided into thin slices and the paper cut with a sharp knife.

Umbelliferae. Carrot Family.

Herbs with alternate compound or sometimes simple leaves, petioles often dilated at the base, rarely with stipules; flowers 1, small, in compound or simple umbels or heads, frequently polygamous; calyx tube adnate to the ovary, limb obsolete or 5-toothed; petals 5, inserted on the margin of calyx; stamens 5, inserted on the disk; pistils with 2 styles; fruit dry, composed of 2 carpels; generally spreading from each other at maturity.

About 1600 species of wide distribution in tropical and temperate regions. A number of the plants of the family are of economic importance, among them the carrot (Daucus Carota), native to Europe, cultivated before the Christian Era. The thickened roots of the carrot are important as food for man and

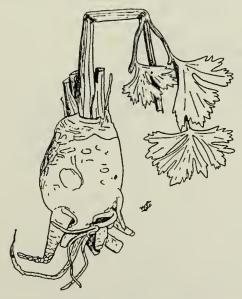


Fig. 368. Turnip rooted Celery or Celeriac (Apium graveolens). (W. S. Dudgeon.)

domestic animals. Parsley (Carum Petroselinum,—Petroselinum hortense), native to the Mediterranean regions of Europe and Asia Minor, is used for garnishing. Celery (Apium graveolens) is indigenous to Great Britain and other European countries, and is found growing in low lands. There are two types, the turnip rooted, cooked and eaten as a salad, and the blanched leaf stalks. Celery was known to the ancient Greeks and Romans. It is said that in a wild state the plant is somewhat poisonous. Some people are known to be sensitive to the cultivated plant. Dill (Anethum graveolens), containing dill oil and caraway (Carum Carvi), were known to the ancients. The essential oil of caraway is obtained from the seed, which is used in Europe to flavor bread and meats and contains carvol C₁₀H₁₄O. Dill (Peucedanum graveolens) is commonly used in flavoring pickles and salads. Cummin seeds from Cuminum sativum resemble those of caraway in odor and taste.

Many members of the order have medicinal properties. Among the more important of these are the Indian pennywort (Centella asiatica), and poison hemlock (Conium maculatum), which contains the alkaloid coniin, which is deadly poisonous. The caraway "seeds," fennel "seeds" (Foeniculum vulgare), the latter indigenous to the Caspian Sea regions, and yielding anethol, also contain fenchone, trigonellin and cholin.

The anise seeds (Pimpinella Anisum), containing the oil of anise, are used in confectionery. Asafoetida (Fernla Narthex) native to Thibet and western Asia, obtained from the milky juice of this plant, is used in medicine, and by the Persians as a condiment. The button snake root (Eryngium yuccaefolium), the cow parsnip (Heracleum lanatum), and the water hemlock (Cicuta maculata), are common plants in the northern states. The leaves and roots of the fennel (Foeniculum vulgare), used in southern Europe as table vegetables, and in Germany to flavor bread and cakes, contain phellandrene C10H16 and chavicol; lovage (Levisticum officinale) is found in salt marshes along the Atlantic coast from Labrador to Connecticut and in Europe. The Arracacia xanthorrhiza of Peru is much used in the Andes region. The same species, known in Venezuela as Arracacha and introduced into Porto Rico, is said by Fairchild to be one of the most important of food plants to the peon. The roots arc large and fusiform. The roots of sea holly (Eryngium maritimum) when candied, boiled or roasted resemble chestnuts in taste. Gum Galbanum is derived from Ferula galbaniflua, and was used by the ancients for incense and perfumery. It contains cadinene and d-pinene. This is referred to in Exodus 25:10. Coriander (Coriandrum sativum) contains coriandrol C₁₀H₁₈O. Musk root or Sumbul (Ferula Sumbul) contains umbelliferone ColloO3 and angelic acid C₅H₈O₂ used as an antispasmodic. The plant occurs in Asia. Cicely (Osmorhiza longistylis) yields an oil similar to anise and contains anethol. In the Umbelliferae the substance peucedone C, H, O, occurs in the roots of Imperatoria Ostruthium, and Peucedanum officinale; athamantin C24 H₃₀O₇, is found in Peucedanum Oreoselinum; laserpitin, C₁₅H₂₄O₄, is from roots of Laserpitium latifolium; pimpinellin is obtained from the roots of Pimpinella Saxifraga. Oenanthe crocata contains oenanthotoxin somewhat similar to cicutoxin; thymol is found in the fruit of a great many of the Umbelliferae; cumin oil is secreted from the fruits of Cuminum Cyminum and other plants of this order, and contains cymene; anise seed or anise fruit contains anise oil which resembles that found in star anise; the Oenanthe Phellandrium



Fig. 369. Coriander (Coriandrum sativum). Flowering stem. (After Faguet)

contains phellandrene; a native lovage, Ligusticum canadense, is used to flavor tobacco.

This family contains a large number of plants with active principles, some of which are entirely harmless, but others must be considered among the deadly poisons. The water drop-wort (Oenanthe crocata), with its parsnip-like roots, and the O. Phellandrium, poisonous European plants, are Umbelliferae. Friedberger and Fröhner state that the former causes stomatitis and paralysis, Blyth

states that the chemistry of the plant has not yet been worked out, but that all parts are poisonous, the root especially deadly. Lehmann states, however, that the first species is not as poisonous as was formerly supposed; sheep and hogs eat it, although it is poisonous to horses. In the latter it is said to produce paralysis of the hind legs. Berula erecta of Europe and North America is poisonous, especially the root. It is a smooth aquatic perennial, with compound simple pinnate leaves; leaflets linear oblong, serrate to cut-toothed; flowers white, and fruit globose.

In Australia according to Maiden the Apium leptophyllum when grown in damp soils is poisonous. The wild parsnip of that country is one of the most poisonous plants of Australia, no antidote to it being known. The Chaerophyllum temulum of Europe causes colic and stupor. The parsley is not ordinarily considered poisonous but is said to be injurious to birds. The gum resin ammoniac found on the stem of Dorema Ammoniacum is acrid. The resin results from the sting of an insect. The genus Ferula from which Asafoetida is derived causes haematuria and bleeding at the nose.



Fig. 370. Creeping Waterparsnip (Berula erecta). Very poisonous. (After Fitch.)

Genera of Umbelliferae

| Flowers yellow |
|--|
| Flowers white or greenish. |
| Fruit bristly, winged |
| Fruit, not bristly, winged. |
| Fruit winged, dorsally flattened. |
| Flowers greenish |
| Flowers white. |
| Leaves pinnate or ternate, clustered, tuberous roots 6 Oxypolis. |
| Leaves ternately-compound, root not tuberous8 Heracleum. |
| Fruit wingless flattened dorsally or laterally |
| Fruit ovoid or oval. |
| Flowers white. |
| Biennial plant1 Conium |
| . Perennial, roots usually fascicled. |

| Oil tubes | solitary |
|-----------|-------------|
| Oil tubes | 1-3 3 Siuma |

Conjum L. Hemlock

Smooth biennial herbs with spotted stems and pinnately compound leaves; flowers small, white, in compound umbels; calyx teeth obsolete; petals small, obcordate or entire; fruit glabrous, somewhat flattened laterally; carpels wavyribbed; oil tubes none, two species, one in Europe and Asia, the other African, deadly poisonous. Plant well known to the ancients.

Conium maculatum, L.

An erect, branching, smooth herb, with spotted stem and pinnately decompound leaves; flowers small, white, in compound umbels; calyx teeth obsolete, petals white; fruit smooth, ovate, flattened, with prominent wavy ribs; oil tubes absent.

Distribution. In waste places, Canada to Indiana, California, Utah and Mexico. Native to Europe.

Poisonous properties. The plant is very poisonous. It was used by the ancients to poison criminals condemned to death, and it is said that Socrates was poisoned by it. The plant is avoided by stock because of its strong odor, but the dried plants are not so poisonous. The alkaloid coniin $C_8H_{17}N$ is derived from it. Coniin is volatile in vapor of alcohol or water, and somewhat volatile at ordinary temperatures. It has an alkaline reaction and burning taste and causes dilation of the pupil. Two other principal alkaloids occur, namely: conicein $C_8H_{15}N$ said to be 18 times more poisonous than coniin; conydrin $C_8H_{17}NO$, pseudoconydrin $C_8H_{17}NO$ and methylaconiin $C_9H_{19}N$ occur in small amounts. The percent of coniin in fresh leaves is 0.095; the ripe seed contains 0.7 percent.

Mr. Chesnut says: Recent cases of poisoning have arisen accidentally from cating the seed for that of anise, the leaves for parsley, or the roots for parsnips; also from blowing whistles made from the hollow stems. It has recently been shown that some of the anise seed in both foreign and domestic markets is contaminated with hemlock seeds, but it is not known whether serious consequences have resulted therefrom.

Symptoms. The symptoms in man are due to a general and gradual weak-ening of muscular power. The power of sight is often lost, but the mind usually remains clear until death ensues, as it soon does from the gradual paralysis of the lungs. The poisoning differs from that of the Water Hemlock (Cicuta maculata) in the absence of convulsions. Many domestic animals have been killed by eating the plant, the prominent symptoms described for cows being the loss of appetite, salivation, bloating, much bodily pain, loss of muscular power and rapid, feeble pulse.

This plant, though called hemlock, should not be confused with the hemlock tree, which belongs to the family Coniferae. It paralyses the ends of the motor nerves, then trunks and lastly the motor center itself. Respiration is quickened and pupils contracted. The fatal dose according to Blyth is 2.3 grains.

2. Cicuta L. Water-Hemlock.

Tall, smooth, erect perennial herbs with pinnate or pinnately compound leaves and serrate leaflets; umbels terminal; flowers white; calyx teeth acute;

fruit ovate or oblong with solitary conspicuous oil tubes; corky ribs, the lateral ones strong; marsh herbs. Eight species, of north temperate regions. The European C. virosa is deadly poisonous. Hundreds of people have been poisoned in Europe. It acts much like our native cowbane, the symptoms being violent gastro-enteritis, dizziness, trembling, suggestive of hydrophobia, prostration, paralysis and convulsions.

Cicuta maculata L. Cowbane

A smooth marsh perennial from 2-5 feet tall, and with fasicled fusiform roots; leaves pinnately compound 2 or 3 times pinnate, long petioled; the coarsely serrate leaflets lanceolate to oblong lanceolate; stalks of the umbellets numerous and unequal; flowers white, fruit broadly ovate to oval, small, about 1½ inches long.

Distribution. Grows in marshes and low grounds in the Dakotas, Nebraska, the Rocky Mountain region of Colorado, Wyoming and Montana to the Uintahs, east of New Brunswick and Florida.

Poisonous properties. The European C. virosa contains contin $C_8H_{17}N$ found also in Conium maculatum, and the bitter principle cicutoxin, an amorphous, resinous substance with a disagreeable taste. The poison resides in the root, stem, and leaves, but more particularly in the root. It seems to occur in an oily aromatic fluid.



Fig. 371. Water Hemlock (Cicuta maculata), showing section of spindle-shaped roots and lower stem, the leaves, flowers, and fruit, one-half natural size; also fruit and cross section of seed, enlarged five times. A very poisonous plant. (U. S. Dept. Agr.)

The resinous cicutoxin, according to Boehm, is an uncrystallizable bitter body. The fatal dose, according to Chesnut, is 50 milligrams for each kilogram of body weight when administered through the mouth and 7 milligrams when injected hypodermically.

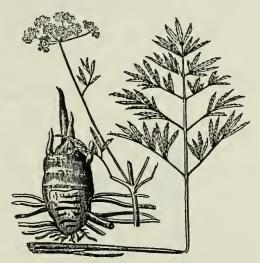


Fig. 372. European Water Hemlock (Cicuta virosa). A poisonous plant containing cicutoxin. (From Vesque's Traité de Botanique).

Mr. Chesnut says:

Its true chemical nature is not definitely known, but it is probable that it contains the alkaloid coniin. and the bitter principle cicutoxin, the latter of which is characteristic of the European water hemlock (Cicuta virosa). Both are powerful poisons, but the latter is the more violent and produces most of the symptoms characteristic of the plant. The American water-hemlock is one of the most poisonous plants native to the United States. Its victims include both men and animals. The underground parts are the most poisonous, and are especially dangerous, because they are often washed or frozen out of the soil and thus exposed to view.

There are quite a number of cases of human poisoning on record in Wisconsin, Iowa, and Minnesota. Stock is also poisoned. People who are poisoned generally mistake the roots for parsnips. In Iowa it is often called wild parsnip. The roots of this plant are fascicled and never conical as in the true parsnip. During one season five children were poisoned in the state of Iowa, three dying, from eating the roots. Several cases of stock poisoning have occurred in Iowa and are referred to by the writer. The following interesting experience is related by Mr. J. A. Minteer, who says:

I have just had a strange experience with my cattle, having lost a four year old cow and a yearling calf. I think that they were poisoned on some kind of weed root found in the slough. I locate it on a spot where a hay stack stood about two years ago. It had been removed except the spoiled hay in the bottom. Last fall being dry, I pitched it up, dried and burned the old hay, sowed rye and timothy secd, ran the disc harrow over it several times and noticed that we turned out lots of roots like small sweet potatoes, except that they were all connected at the top. I thought they were the root of a weed that grows a stalk similar to a seeded parsnip, have a straggly top similar to an elder berry when in bloom. The stalk when mature is hollow. Now I am not certain that I am right about the top as it had been mowed before I discovered the tubers. I never thought of them doing any harm, just thought we bad torn them out so they would die and do me no harm, but as the cattle, 17 in number were

hrought up Sunday evening they appeared to be all right until they came into the barn yard, when a cow fell down and seemed to have a spasm. It only lasted a few minutes when she got up walked ahout 100 feet and fell again, got up and walked ahout thirty rods, fell again and died in ahout thirty minutes. The yearling was all right until turned into the lot. In about 20 minutes she was taken in the same way except a little more severely, rose 2 or 3 times and died in ahout 15 minutes. I was satisfied that they were poisoned, but the cause worried me for a while then I remembered the tubers I saw in the slough, I went next morning before turning the cattle out and found that the cow and yearling had eaten some of the roots. I gathered up nearly one-half bushel of the tubers, turned out the cattle and have had no trouble since. On opening the cows, I found considerable of the tubers in the stomach, and the inside of the stomach was very black.

The plant above ground likewise affects horses and evidently the poison may reside in the leaves for considerable length of time even after they are dried. The following experience of a correspondent in Ruthven, Iowa, calls attention to the danger of using hay that contains cowbane:

I mail herewith a small paper hox which contains some weed, of which I sent you a specimen last summer. This species of hemlock as you call it, I picked out of a manger of a stallion, which took suddenly sick this morning. Sickness lasted but a short spell. Do not know whether this had anything to do with this sickness but am terrihly prejudiced against it. Another instance a few days ago of a colt taking violently sick all at once, apparently no cause, there heing considerable of this weed in the hay, and I had two cows lose their calves a short time ago; the cows had access to this kind of hay. This quite frequently occurs hereahouts. On a neighboring farm where this weed ahounds, they lost nearly all their calves two years ago. Apparently no cause, hut of course there is a cause somewhere. I am satisfied some stock will eat the leaves of this weed.

Dr. Erwin F. Smith, in referring to the poisonous nature of this weed, speaks of a case as follows:

During the warm days which melted the snow and brought hack the birds and gave indication of spring time, some children of a neighborhood on the outskirts of the city gave vent to their feelings by digging and eating some artichokes which grew upon some low ground borderng a brook. Two of these hoys were taken violently ill and one of them eight years old, died, within an hour after he had eaten the root.

Dr. Smith states that upon an examination of the stomach and the root from which he ate, it was proven beyond a doubt that *Cicuta maculata* was the cause of death.

Professor A. A. Crozier calls attention to the poisonous nature of cowbane in this state and refers to a case occurring in northern Iowa as follows:

Hon. Eugene Secor, of Forest City, this state, a member of the Board of Trustees of the Iowa Agricultural College, brought me today a fleshy root of a plant of the Water-hemlock, (Cicuta maculata, L.). The circumstances which brought it to his notice were as follows: A neighbor of his hy the name of Mr. Oleson, a farmer of about fifty years of age, while dragging some potato ground upon bottom land ahout two weeks ago discovered one of the fleshy roots of this plant, and supposing it to he an artichoke, ate it and gave a portion of it to his two sons. He soon began to feel queer or "funny" as he expressed it, and went to the house where he was taken with a spasm, followed hy two or three others, when he became unconscious and within half an hour, before a physician could be summoned from the village, two miles distant, he was dead. The children had prohably eaten less of the root and heing given an emetic, recovered. The plant is very common in the state and the roots are so pleasant to the taste as to make it particularly dangerous. I may add that I ate a piece of the root the size of a filhert with little or no unpleasant effect."

One season the writer had a record of five cases of poisoning in this state. From a press bulletin issued by the writer the following facts were given to the public:

Ira, aged ten, and Ross, aged eight years, children of T. Y. Johnson, died last night from eating the roots of a poisonous water plant that grows in front of their house on the Keg Creek flat east of the Ridgeway lumher yard. A third child, John, the seven year old son of Mrs. Amanda Kingery, also ate the root but it seems was not made so sick as the others.

Shortly hefore six o'clock the children came into the house showing Mrs. Johnson what they had been eating. Not knowing what it was she had them spit it out and throw away

what they had in their hands. They went out to play again, but in about twenty minutes the two Johnson boys fell to the ground as if in a fit, soon passing into convulsions. The Kingery lad was able to walk to his home, but was soon taken with spasms. Dr. Hester was called and by a vomiting process in a few hours had the child out of danger.

Dr. Lyon was summoned to the Johnson home, getting there at 6:30. The little fellows were already in terrible convulsions and nothing could be done for them, one dying at 6:45

and the other at 7:15.

Dr. Millspaugh, in his American Medicinal Plants, Fascicle 4., No. 67, has recorded the following observations concerning the physiological action of the Cicuta maculata:

"Many cases of poisoning from the root of this species have been reported, all showing, by the symptoms, that Cicuta produces great hyperaemia of the brain and spinal cord. The following case reported by letter to Dr. Bigelow* by Dr. R. Hazeltine, (1818), gives all



Fig. 373. Purple Stemmed Poison Hemlock (Cicuta Douglasii) of the Pacific Coast. It is very poisonous. (Chesnut, U. S. Dept. Agr.)

^{*}Bigelow, Amer. Med. Bot. Vol. III., p. 181,

the symptoms noted by other observers in various other cases. A boy had eaten of certain tuberous roots, gathered in a recently plowed field, supposing them to be artichokes but which were identified as the roots of Cicuta maculata. Ilis first symptom was a pain in the bowels, urging him to an ineffectual attempt at stool after which he vomited about a teacupful of what appeared to be the recently nussticated root, and immediately fell back into convulsions which lasted off and on continuously till his deatb. The doctor found him in a profuse sweat and convulsive agitations, consisting of tremors, violent contractions and distortions, with alternate and imperfect relaxions of the whole muscular system, astonishing mobility of the eyeballs and eyelids, with widely dilated pupils, stridor dentium, trismus, frothing at the mouth and nose, mixed with blood and occasionally violent and genuine epilepsy.

The convulsive agitations were so powerful and incessant, that the doctor could not examine the pulse with sufficient constancy to ascertain its character.

At the post-mortem no inflammation was observed, the stomach was fully distended with flatus, and contained "about three gills of muciform and greenish fluid, such as had flowed from the mouth; the mass assumed a dark green color on standing."

Chesnut in his paper on Some Poisonous Plants of Northern Stock Ranges says that Dr. Wilcox and himself observed 105 cases of water hemlock poisoning among sheep of which 50 were fatal, and 36 among cattle of which 30 were fatal. The loss was \$4,000, only a fraction of what occurs in Oregon. The C. vagans and C. Douglasii are poisonous, the latter along the coast.

Mention may be made in this connection of a series of most valuable papers on "The Medicinal Plants of North America" by Dr. T. Holm in Merck's Reports.* In one paper he discusses the anatomy as well as the poisons found in this very poisonous plant. The effect of the poison is similar to that of Cicuta virosa and is due to a resinous substance cicutoxin and to the volatile alkaloid cicutin, which has been obtained from the fruits.

Cicuta vagans Greene. Oregon Water-Hemlock

A smooth perennial with glaucus stem and vertical rootstock divided into horizontal chambers; plant 2-3 feet high, with compound leaves; flowers white.

Distribution. From Idaho to British Columbia and west to northern California.

Poisonous properties. Same properties as the preceding. Professor Hedrick estimates that 100 head of cattle are killed by it every year in Oregon. A piece about the size of a marble of the winter rootstock is believed to be fatal to man. Professor Hedrick says:

It is hard to estimate the number of cattle killed yearly in Oregon by eating Cicuta. One hundred would be a low estimate in my judgment. Animals eat the underground portion of Cicuta in getting the tops which form about the first green herbage in early spring; as they browse the foliage, the roots, being only partly subterranean, and growing in a soft soil, are pulled up and eaten. A piece the size of a walnut ,it is found by experiment, is sufficient to kill a cow. It is probable that the poisonous constituent is found only in the underground stem and the roots.

While the victims of the plant are chiefly cattle, yet they are not exclusively so. The poisonous parts are often mistaken for Parsnips, Artichokes, and Horse-radish, and thus human victims are not infrequent. A number of cases of poisoning from Cicuta are annually reported in the United States. A writer in a local paper a few months ago, reported the case of two cattlemen in Southern Oregon, who, after eating "Wild Parsnip," presumably Cicuta, died in a few hours. Falk reports, that in Europe in thirty-one cases of poisoning from Cicuta, 45 per cent died.

The observations made by Prof. French and reported by Prof. U. P. Hed-

^{*}Medicinal Plants of North America, 24; Cicuta maculata, Merck's Rep. XVIII: 35-38, f.1-12. Feb. 1909.

rick formerly of the Oregon Experiment Station have shown experimentally that this species is poisonous. A bulb was cut in small pieces, mixed with a carrot and fed to a two year old grade heifer. The animal was fed at 8:00 a. m. and at 9:30 it was dead. A post-mortem examination showed that pieces



Fig. 374. Oregon water hemlock (Cicuta vagans): a, plant with leaves, one-sixth natural size; b and b', rootstock and horizontal roots, showing section, half size; c, terminal leaflets, one-sixth natural size; d, flowering spray. (U. S. Dept. Agr.)

of the root occurred in the rumen, and in the reticulum or second stomach. Only a very small portion of the root had been eaten by the animal. Two grams were found in the stomach. The lungs were highly congested.

The following day a one year old calf was fed with the poisonous roots. The temperature of the animal was normal, $102\frac{1}{2}^{\circ}$. Two bulbs the size of an egg were cut and mixed with carrots; this material was eaten under protest. The roots were fed at 9:15 a. m. and at 9:40 the temperature was 103; at 10:00, $103\frac{1}{2}^{\circ}$; at 10:25, 104° and the animal was trembling about the flanks, the eyes watered freely; at 10:35 the animal was excited and the temperature was $104\frac{1}{4}^{\circ}$; at 10:45 the animal fell over on its side in a spasm. The eyes were drawn and the muscles were rigid and contracted violently.

Before death an attempt was made to revive the animal by giving it spirits of turpentine; this caused the calf to recover its feet and walk about. A second

dose of turpentine and milk were given and the animal remained standing until 11:30, then it fell down as before. A full dose of aconite was given, but no results noticed and then a hypodermic injection of nitro-glycerin was given. Death occurred at 11:45 and spasms were almost continuous.

The most interesting discovery made by Prof. Hedrick was in regard to the apparent harmlessness of the roots while growing during the summer.

Prof. Hedrick says concerning the roots fed to an animal early in May:

The bulbs were of the same lot used in the first experiments, but had been growing for a month in a green house. It was expected that growth would remove some of the dangerous properties of the bulbs, but it was a surprise to find that an animal eating many times as much of them as had killed the cows in the previous experiment suffered no ill effects whatever. The eonclusion is obvious; the bulbs are only dangerously poison when in the dormant state, or for a short time after growth begins in the spring. Cattle are likely, then, to be poisoned only from the first of January to the middle of May.

Cicuta Bolanderi Watson.

Leaves bipinnate, leaflets narrowly lanceolate, long acuminate, acutely serrate, lower leaflets petiolate and often deeply lobed; involucre of several linnear leaflets; fruit 2 lines long, nearly orbicular, strikingly ribbed, and broad oil tubes.

Distribution. In salt marshes along the Pacific coast in California.

Poisonous properties. Like those of the preceding species.

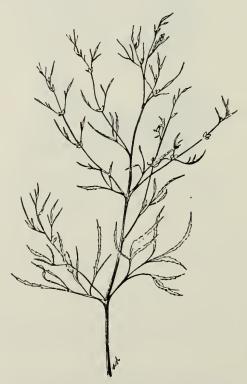


Fig. 375. Water Hemlock (Cicuta bulbifera). A powerfully poisonous plant. (Ada Hayden.)

Cicuta bulbifera L. Bulbous Water-Hemlock

A slender perennial from 1-3 feet high; leaves 2-3 times pinnate; leaflets linear; sparingly toothed, 2 inches long; upper axils bear clustered bulblets; fruit small, ovate.

Distribution. In swamps Nova Scotia to Delaware, Indiana, Iowa, Nebraska and Manitoba.

Poisonous properties. Very poisonous, like the other species described above.

3. Sium (Tourn.) L. Water Parsnip

Smooth perennial herbs with stem leaves pinnate, basal leaves with pinnatifid leaflets; flowers white in large umbels; calyx teeth minute; short styles; fruit ovate to oblong with prominent ribs; 1-3 oil tubes. A small genus of 8 species in temperate regions. The S. latifolium is poisonous especially to cattle. It produces stupor, excitement and gastro-enteritis.

Sium cicutaefolium Schrank. Water Parsnip.

An erect stout marsh herb from 2-6 feet high; lower leaves with long petioles; leaflets 3-8 pairs; segments linear or lanceolate, sharply serrate; flowers in umbels, white; fruit ovate, prominently ribbed.

Distribution. From Nova Scotia across the continent to Florida and California.

Poisonous properties. The water parsnip is reported as poisonous from several different sources. Hyams reports it poisonous in North Carolina.

4. Aethusa L. Fool's Parsley.

Annual glabrous herbs. Leaves 2-3 ternately compound, dissected; umbels compound without involucre; involucels long and narrow; flowers white; calyx teeth obsolete; fruit globose, ovoid, glabrous, flattened dorsally. One species native to Europe and Asia.

Aethusa Cynapium L. Fool's Parsley

An erect leafy branched annual. Leaves 2-3 times pinnate; petiole dilated at the base; umbels long peduncled.

Distribution. In cultivated grounds and waste places from Nova Scotia. New England to Pennsylvania and New York.

Poisonous properties. Contains the alkaloid cynapin, and a coniin-like alkaloid. One physician in England regards the plant as non-poisonous, even recommending it as a pot herb or for salad uses. On the other hand numerous cases of poisoning have been recorded. The following may serve as an illustration which is recorded by Dr. Millspaugh from a statement made many years ago. He records the experiments made on animals. Seven ounces of the juicy leaves were given to a strong dog and the oesophagus tied; twenty minutes thereafter the dog became sick. He stretched out his limbs, and lay on his stomach and it was impossible to arouse the animal. The pupils were scarcely dilated, the pulsations of the heart were slow and strong. The extremities were agitated by convulsive movements; the animal threw himself from one side to the other, and died an hour after taking the poison. The postmortem examination showed that the heart was contracted and the stomach was found to be full of the poison. In domestic animals it causes stupor, paralysis and convulsions. The common name indicates that it is sometimes mistaken for parsley with injurious results.



Fig. 376. Fool's Parsley (Aethusa Cynapium). Flowering branch. Supposed to be poisonous to animals. (After Faguet.)

5. Angelica L. Angelica

Stout perennial herbs with ternately or pinnately compound leaves in large ample umbels with white or greenish flowers; involucre none or of few small bracts; calyx teeth wanting; fruit strongly flattened with prominent lateral wings; oil tubes solitary or several; the 30 or more species found chiefly in the northern hemisphere and New Zealand.

Angelica atropurpurea L. Great Angelica

A stout smooth perennial from 3-5 feet high; large ample, ternately divided

leaves, pinnate segments, ovate-lanceolate and broad dilated petioles; flowers in large umbels 9-25 rayed; fruit broadly oval, oil tubes 25-30.

Distribution. Common in swamps from Labrador, Wisconsin, Minnesota, Illinois to Delaware.

Poisonous principles. Supposed to be poisonous. Although no cases have been reported to the writer, it is known that the Indians of Canada used the fresh roots for suicidal purposes. On drying, however, the roots lose their poisonous properties. Dr. Millspaugh says that they are considered carminative, diuretic, emmenagogue and stimulant. The dried root was often used, especially in combination with other and better known diuretics, in anasarca and various diseases of the urinary organs; and alone in flatulent colic and suppressed menstruation. Dr. Schell claims that doses of 15 to 20 grains of the dried root will cause a disgust for all spirituous liquors. The stems were often made into a candied preserve in some sections of the country—a practice now nearly extinct.

6. Oxypolis Raf. Cobwane

Perennial, glabrous marsh herbs, with clustered tuberous roots; leaves reduced, pinnate or ternate; flowers white, in compound umbels; calyx teeth acute; fruit ovate, dorsally flattened, compressed; dorsal ribs slender, the lateral broadly winged; oil tubes solitary in the intervals, 2-6 on the commissure. Species 4, native to North America. Poisonous.

Oxypolis rigidior (L.) Coult and Rose. Cowbane.

A slender marsh perennial from 2-5 feet high. Roots tuberous, clustered; leaves simply pinnate, petioled; leaflets thick, ovate-lanceolate or oblong-entire or denticulate; involucre of 1-4 bracts or none; flowers white; oil tubes small.

Distribution. In swamps from New York to Florida, to Missouri and Minnesota.

Poisonous property. The roots and leaves are known to be poisonous. Said to poison cattle.

7. Pastinaca L. Parsnip

Tall, branching biennial herbs, pinnate leaves, thick conical roots, compound umbels with yellow flowers; involucres and involucels commonly absent; obsolete calyx teeth; fruit smooth, oval, flattened, the lateral ribs extending into broad wings; oil tubes solitary, 2-4 on the commissure. Six or seven species native to Asia and Europe.

Pastinaca sativa L. Wild Parsnip

Tall branched biennial or annual herb with thick conical roots, pinnately compound leaves smooth or somewhat pubescent; calyx teeth obsolete; petals yellow, fruit oval, glabrous, flattened dorsally, seeds flat.

Distribution. Common in the northern states and on the Pacific coast, where it has escaped from cultivation.

Poisonous properties. Frequently confused with cowbane and said to produce poisoning. The writer has not received a single specimen where Wild Parsnip was said to have produced the poisoning, that the plant did not prove to be cowbane. The wide spread belief of the poisonous nature of the cultivated parsnip running wild is entertained by a large number of people and also to some extent by the medical fraternity. A few years ago Professor Fred-

erick B. Power and one of his pupils (Mr. J. T. Bennett) undertook some experiments to determine whether the cultivated parsnip running wild had any toxic properties. Mr. Bennett failed to detect the presence of any poisonous principle in the root of the true wild parsnip (Pastinaca sativa) and when the boiled roots were fed in considerable amounts to a cat, no symptoms of poisoning were manifest. We may add as a further



Fig. 377. Wild Parsnip (Pastinaca sativa). A plant with yellow flowers and is feral form of ordinary parsnip. Often mistaken for Cowbane. (Charlotte M. King.)

testimony that Professor Power reports that his associate, Dr. Kremers, insisted upon eating one-half of one of the raw roots obtained from Mr. Rynning of West Salem, Wisconsin, which were supposed to cause cases of the poisoning. Dr. Kremers reported no ill effect whatever. One of the largest of the fresh raw roots weighing three and one-half ounces avoirdupois was chopped fine, mixed with some meat and fed to a small dog. The animal ate it greedily and without disturbance. There were no symptoms whatever of poisonous action. I have on different occasions eaten the wild parsnip (Pastinaca sativa), without any ill effect, so that the above results are corroborated. I will admit that I had some hesitancy at first and that Mr. Sexton, the foreman of the horticultural department at the Iowa State College did not expect to see me alive by evening. I must confess also that the roots were somewhat woody and not very palatable.

Dr. J. J. Brown, Sheboygan, Wisconsin, as quoted by Dr. Power, states that he had prepared and dug enough wild parsnips for a good dinner which he ate and can testify that he could discover but little difference cooked or raw, from cultivated parsnip and those which had run wild for about 50 years.

That the parsnip does at times produce dermatitis has been proven. The following verey interesting letter from Professor F. C. Stewart, botanist of the New York Agricultural Experiment Station, is of interest in this connection:

I recollect that some fifteen years ago you were much interested in the reputed poisonous properties of the wild parsnip, and that you reached the conclusion that wild parsnip is not poisonous. I think you may be interested to know of an instance which has recently come to my attention. Henry VanDreser, a prominent lecturer on poultry in this state, last season had a very serious injury to his face and eyes. His face became badly swollen and his eyes were in a terrible condition. It was feared at the time that they would be ruined, but the sight was not lost although it was considerably impaired. The physician in charge diagnosed it as a case of poisoning, due probably to the flowers of wild parsnip. Very shortly before the trouble appeared Mr. VanDreser had been mowing a large patch of wild parsnip which was in bloom. It was a hot day, so that he perspired profusely. He gathered bunches of the wild parsnip plants in his arms and carried them. This brought the plants in contact with his face. Both Mr. VanDreser and the physician feel confident that the wild parsnip was the cause of the trouble. Another gentleman who heard of this case told me that some years ago he lost a little girl with poisoning of a somewhat similar character, and it was attributed to the parsnip blossoms, among which the little girl had been playing immediately before the attack.

The writer has also known of a few cases of dermatitis produced by this plant. Poisoning similar to the above may be produced by other members of this family notably the cowbane. A young high school lad in Boone lost his life in a way similar to that from Pastinaca sativa.

8. Heracleum L. Cow Parsnip

Tall stout perennials with large ternately compound leaves; flowers in large umbels; involucre deciduous and involucels of numerous linear bracts; calyx teeth obsolete or small; petals white, clawed, the outer flowers dilated; fruit broadly obovate, flattened, lateral ribs broadly winged, the intermediate narrow; oil tubes extending to about the middle. About 60 species, of cooler regions.

Heracleum lanatum Michx. Cow Parsnip

A stout, hairy, pubescent perennial from 4-8 feet high; leaflets broad and large, irregularly cut-toothed; flowers white, in broad umbels.

Distribution. From the Atlantic coast, Newfoundland, through the northern states and Allegheny Mountains to California. Common in the Rocky Mountains.

Poisonous properties. Said to be poisonous although the leaves of the fresh plant are eaten by the Indians. It contains the bitter principle heraclin $C_{32}H_{29}O_{10}$. The roots have a disagreeable flavor. Hyams states that it is poisonous and Halsted says that cow parsnip will produce blisters.

Daucus Tourn. Carrot

Bristly annual or biennial herbs with pinnately de-compound leaves; umbels compound, of white or reddish flowers; involucre of numerous entire or toothed bracts; calyx teeth obsolete; petals obovate with apex turned in; the outer flowers often dilated; fruit oblong, flattened with 5 primary ribs, secondary ribs 4 winged, each bearing a row of prickles. About 25 species of wide distribution.

9. Daucus Carota L. Wild Carrot

Biennial or annual bristly herbs and conical tap root; lower leaves 2-3 times pinnate, segments dentate lobed or pinnatifid, bracts of the involucre of linear divisions; flowers white or pinkish.

Distribution. Native to Europe. Common in fields and waste places in the north to the Pacific coast. It is the original of the cultivated carrot.

Injurious properties. The fruit often clings to wool, making it of inferior quality. The leaves of the plant cause vesication. This is especially true when

the plant is wet with dew. In Australia, according to Maiden, the Daucus brachiatus is injurious to sheep, the hooks sometimes entering the skin.

CORNACEAE Link. Dogwood Family.

Shrubs or trees, with opposite or alternate leaves; calyx tube adnate to the ovary, lobes small; petals as many as the stamens, 4-5, inserted on the disk; stamens as many as the petals or more numerous; ovary inferior, 1-2 celled; style 1, ovules 1, in each cavity; fruit a 1-2 seeded drupe.

A small family of 16 genera and about 85 species, containing several plants cultivated for ornamental purposes, such as the red osier (*Cornus stolonifera*), native from Nova Scotia south to Virginia, Kentucky, northeastern Iowa, Nebraska, the Rocky Mountains, California and Alaska.

The Aucuba japonica, with handsome red berries, is cultivated in green-houses; it contains the glucoside aucubin $C_{13}H_{11}OH+H_2O$.

Flowering dogwood (Cornus florida) is a small tree or large shrub, cultivated for its large corolla-like bracts. It is native in dry woods from southern New England to Ontario, Illinois, Missouri, and Texas. The western dogwood (Cornus Nuttallii) of the Pacific, is a beautiful tree from 20-75 feet high and 1-2 feet in diameter. It produces a close grained wood that is capable of receiving a high polish and is used for furniture, cabinet work, and water wheels. Tupelos (Nyssa aquatica) and (N. sylvatica) produce wood of some importance.

The dogwoods, especially the flowering dogwood and the round-leaved dogwood (Cornus circinata) are used in medicine; the bark contains the bitter principle cornin. Some species, like panicled dogwood (C. paniculata) are occasionally troublesome in newly cultivated fields.

Poisonous properties. At various times, members of this family have been suspected of being poisonous. Two species, Cornus Amomum Mill. and C. paniculata L'Her. (= C. candidissima Mill.) are thus listed in Part I of this Manual. Actual poisoning from species of Cornus has not, so far as is known to the present author, been observed. Greshoff, however, lists as poisonous C. Amomum Mill., Alangium Lamarckii Thwait., and Garrya Fremontii Torr.

METACHLAMYDEAE

Petals partly or wholly united, rarely separate or wanting; tubular or funnel-form. This includes the important plants classed as Gamopetalae.

Ericales

Flowers complete, regular with lobed or distinct calyx; corolla cleft; stamens free from corolla; ovary compound. This order includes such families as *Pirolaceae* and *Ericales*.

ERICACEAE. Heath Family.

Shrubs, trees or occasionally herbs, with simple leaves; flowers regular, or nearly so, generally perfect; calyx usually free from the ovary, 4-5 lobed, generally persistent; corolla of 4-5 petals or 4-5 lobed; anthers 2-celled, commonly appendaged or opening by pores; ovary 3-10 celled; style 1; pollen consisting usually of 4 united grains; embryo small.

A large family of wide distribution. About 1400 species. Several species of the family are frequently cultivated for ornamental purposes—notably the rhododendron and the wild laurels (Rhododendron catawbiense, R. maximum), which are hardy out of doors in the south and east. The azalea (Rhododendron indicum) is frequently cultivated in greenhouses, flowering in late winter and early spring; the garden azalea (Rhododendron sinense) is cultivated in gardens.

The California mountain laurel (R. californicum) is a very attractive ornamental shrub or small tree and one of the most handsome of the genus in North America. Among the other cultivated members of this family, mention may be made of the mountain laurel (Kalmia latifolia). The South African species of the genus are frequently cultivated in conservatories. The Scotch heather (Calluna vulgaris) of Europe has been found on Nantucket Island. The May flower or trailing arbutus (Epigaea repens) is well known to people living in the Eastern and Northern States. The rich, spicy fragrance makes it one of the most attractive of our early flowering plants. Several species of the genus Vaccinium are cultivated for their fruits. Among them are cranberries; the larger berry being V. macrocarpon, which is the most common species in North America, and is cultivated extensively in Wisconsin, Michigan, New Jersey and Massachusetts. The smaller cranberry V. Oxycoccus is also sold under the name of Lingon berries and is common in Sweden and Norway and far northward in North America. The berries of this fruit are smaller and more acid than are those of V. macrocarpon.

The blue berries and huckleberries are obtained from several native species of the genus Vaccinium of the East. Dwarf blueberries of our northern dry woods are (V. pennsylvanicum and V. canadense). Another blueberry (V. vacillans) produces a sweet edible berry and occurs from Maine to Missouri. The mountain blueberry of the southern Alleghanies is V. corymbosum, var. pallidum. The tall blueberry (V. corymbosum) has a pleasant acid flavor, and attains a height of 5-10 feet. In the Rocky Mountain, Lake Superior and White Mountain regions, there are several species of blueberries as V. caespitosum with blue sweet berries and V. membranaccum, which produces large black berries, and grows from 1-5 feet high.

The huckleberry (V. stamineum) is not edible, nor is the fruit of the small tree known as farkle berry (V. arboreum). The huckleberry (Gaylussacia baccata) is edible. These come up spontaneously in burnt areas. The fruit is gathered from Maine to Minnesota. The shallon (Gaultheria Shallon) of the Pacific coast, is a shrub or small tree with stout erect branches, and produces edible fruit with a spicy aromatic flavor. There are several species of arbutus, one of which, Madrona (Arbutus Menziesii), forms a tree 1-6 feet in diameter and 20-100 feet high, the wood being used for cabinet work. The strawberry tree (Arbutus Unedo) is a native of Southern Europe to northern Africa. The fruit when ripe resembles the strawberry, but is not edible. Several members of the order are used in medicine. One species only, however, is generally recognized. The Pipsissewa (Chimaphila umbellata) used for cutaneous eruptions, contains chimaphin C24H2,O4. The bear-berry (Arctostaphylos Uva-ursi) is found throughout the mountain regions of North America-Pennsylvania, Illinois, Missouri to Nebraska, the Rocky Mountains and northward. It is an astringent tonic used in diseases of the liver, and contains ericolin; arbutin, ursone C₁₀H₁₈O. The wintergreen (Gaultheria procumbens) is a small creeping shrub with evergreen leaves and edible fruit, and is used for the production of the oil of wintergreen which serves as an aromatic stimulant and also in rheumatism. The fragrance is due to a volatile oil,

Drs. Price and L'Engle quite recently mention a case of poisoning from the oil of wintergreen. In this case a child swallowed the contents of a bottle containing oil of Gaultheria. The symptoms of poisoning were increased pulse, labored and irregular respiration, hearing impaired, some hallucinations of vision, twitching of heart and weak diarrhoea. The child died. Many of the Ericaceae contain the glucoside ericolin $C_{34}H_{56}O_{21}$; this in turn contains ericinol $C_{10}H_{16}O$, a peculiarly fragrant oil; ericolin occurs in the leaves of Ledum, Erica, Calluna, Rhododendron, Gaultheria and Epigaea. Rhododendron $C_{16}H_{22}O_7$, is found in the leaves of Rhododendron chrysanthum; it is converted by hydrolysis into rhododendrol, $C_{10}H_{12}O_2$. Oxycoccin occurs in Vaccinium Oxycoccus.

Andromedotoxin or acetotoxin, found in the leaves of Andromeda Polifolia and A. japonica, and also in many other Ericaceae, as Azalea, Rhododendron Japonicum, A. javanicum, and Kalmia is one of the most toxic substances known.

Asebotin, $C_{24}H_{28}O_{12}$, derived from andromedotoxin, and asebogenin, $C_{18}H_{18}O_7$, are found in Andromeda Japonica. Arbutin $C_{12}H_{18}O_7$, occurring in the leaves of many of the Ericaceae was first detected in Arctostaphylos Uva-ursi; quercetrin is found in Calluna, as is also calluna tannic acid.

In Lehmann's work on the poisonous plants commonly found in Germany only one species of this family is listed as poisonous, namely the Labrador tea (Ledum palustre) of the Arctic region. This pretty shrub, growing in swamps, has alternate entire leaves with rusty wool underneath; revolute margins; white flowers in umbel like clusters; followed by a many seeded capsule. The L. groenlandicum related to it and also occurring in swamps, thickets and on mountain slopes but further southward has wider leaves and is a larger plant.



Fig. 378. Great laurel (Rhododendron maximum): a, flowering branch; b, fruiting capsules—both one-third natural size, Very poisonous. (U. S. Dept. Agr.)





A. Rhododendron, Hydrangea in a Maryland forest. (After Shreve, Chrysler, Blodgett and Besley, Maryland Weather Service).



B. Mountain Laurel, Rhododendron in a hemlock forest in Maryland. (After Shreve, Chrysler, Blodgett and Besley, Maryland Weather Service).

The L. glandulosum, similar to the above but a taller plant, occurs in bogs of the Rockies. All are more or less poisonous; one species is known to contain andromedotoxin. Several species are used as a substitute for tea. The Russians use the first named species. It is also said to be used to protect clothes from moths.

Genera of Ericaceae

Corolla somewhat irregular.

Corolla slightly 2-lipped, leaves deciduous....(Rhododendron-Azalea)

Corolla campanulate; leaves evergreen..................1 Rhododendron

Corolla regular.

Rhododendron L. Rose Bay

Shrubs or trees, with deciduous or persistent leaves; flowers in corymbose or umbellate clusters from scaly cone-like buds; calyx 5-lobed, or 5-parted; corolla bell-shaped, 5-lobed, nearly regular or irregular; stamens 5-10, spreading, declined and slightly or not exserted; anthers opening by terminal pores; style slender, occasionally exserted and somewhat declined; fruit a woody capsule; seeds numerous. About 100 species, native of cooler regions. Several species cultivated for ornamental purposes. The genus includes Azalea.

Rhododendrou californicum, Hook. California Rhododendron.

A shrub or small tree from 3-25 feet high, with stout erect branches; leaves evergreen, broadly oblong, 3-6 inches long; flowers large, in corymbose clusters; calyx 5-lobed; corolla bell-shaped, rose color; capsule 1-11/4 inches long.

Distribution. On the Pacific slope from San Francisco north to British Columbia.

Rhododendron maximum, L. Rose Bay or Great Laurel

A tall evergreen shrub or small tree; leaves 6-10 inches long, thick and leathery, oblong-lanceolate, dark green on both sides, with revolute margins; flowers in corymbose clusters, pedicels viscid pubescent; corolla bell-shaped, 1 inch broad, pale rose color or nearly white or spotted with yellowish orange dots; fruit an oblong capsule.

Distribution. In damp woods from Maine to Ohio and the Alleghenies to Georgia.

Rhododendron catawbiense Michx. Mountain Rose Bay

Shrub from 3-6 feet high, or rarely small tree 20 feet high; leaves oval or oblong, mucronate, dark green above, pale or glaucous beneath; petioles stout; pedicels rather stout; corolla broadly bell-shaped, lilac-purple; petals rusty-brown.

Distribution. On mountain slopes of Virginia, West Virginia, and Georgia. Poisonous properties. According to Chesnut, Rhododendrou californicum

is said, in Oregon, to be poisonous to sheep. The substance andromedotoxin, $\mathbf{C_{81}H_{51}O_{10}}$, a bitter principle, has been found in Rhododendron maximum. It is extremely poisonous, even more so than aconitin, and more emetic than emetin. Rhododendron ponticum contains the glucoside arbutin.

Rhododendron occidentale A. Gray. California Azalea

Shrub 2-6 feet high, shoots glabrous or minutely pubescent when young, not bristly; leaves obovate-oblong, sometimes approaching lanceolate, bright-green and shining above, minutely pubescent, glabrate, the margins minutely hispid-ciliate; scales of the flower-bud somewhat canescent; flowers appearing after the leaves; sepals distinct, oblong or oval, conspicuous; corolla minutely viscid-pubescent outside, white, with the upper lobe yellow inside; the narrow funnel-form tube equalling the deeply 5-cleft slightly irregular limb; the lobes ovate; stamens and style much exserted, moderately curved; capsule oblong.



Fig. 379. California Azalea (Rhododendron occidentale). Flowering stem. Leaves are said to be poisonous to stock. (From American Agriculturist.)

Distribution. In wooded districts along streams in California extending to the mountains east of San Diego and common northward.

Poisonous properties. The leaves of the species have been analyzed chemically and have been found to contain a poisonous substance, said in California to be poisonous to stock. Certain species of Azalea contain the glucoside ericolin $C_{34}H_{50}O_{21}$.

Kalmia, L.

Erect shrubs with entire evergreen coriaceous leaves; naked buds; flowers in umbels or corymbs, or solitary, or 2-3 together in the axils; calyx 5-parted; corolla somewhat wheel-shaped, 5-lobed, with 10 pouches in which 10 anthers are lodged; filaments long, stamens 10, shorter than the corolla; anther-sacs opened by large terminal pores; ovary 5-celled; ovules numerous; capsule obscurely 5-lobed, 5-celled, many-seeded; seeds small. Species 7, 6 native to North America, 1 in Cuba.

Kalmia latifolia L. Calico-bush. Mountain-laurel.

A shrub 4-8 feet high, forming dense thickets, becoming a tree 10-30 feet high in the mountains; branches stiff; leaves alternate, bright green on both sides, ovate-lanceolate or oblong petioled; flowers profuse, large, and showy, tip rose-color to nearly white, in terminal corymbs; capsule depressed, globose, 5-lobed, glandular.

Distribution. Rocky hills, damp soil. Canada to Maine and the Allegheny Mountains, through West Florida, Ohio, Kentucky, and Tennessee.

Poisonous properties. This plant has long been known as poisonous, and is very much dreaded in the Alleghany Mountains. The substance andromedotoxin $C_{31}H_{51}O_{10}$, referred to elsewhere, has been found in all parts of the plant. Prof. Chesnut, in reference to the plants says:

Scores of cattle and sheep are poisoned annually by eating the shrub. Access to it is generally obtained by breaking away from inclosures, or through neglect or accident when cattle or sheep are being driven past laurel thickets to upland pastures in early spring. Laurel leaves (commonly used for decorative purposes in winter), or the flowering branches, are often carelessly thrown into inclosures where animals are kept. The older cattle are not so frequently killed by it, but they are by no means immune. Horses and even goats have died from eating the leaves and in May, 1895, a monkey was killed at the National Zoological Park, at Washington, D. C., by eating a few flowers and leaves offered to it by a visitor. Decr and grouse are said to be immune, although it is claimed that their flesh, especially that of the ruffed grouse, is poisonous when they have fed upon it. It is stated that chickens have been poisoned by eating the vomited matter from poisoned animals. Experiments show, however, that they are able to withstand considerable quantities of the pure poison when it is fed to them. In these experiments the chickens were killed with chloroform after dosing for a few days. The entrails were then cast aside, and the well-boiled meat was fed to cats with nearly fatal results. The honey derived from the nectar of the flower appears to be poisonous under some conditions. Cases of human poisoning occur indirectly in the ways indicated above; directly by overdoses or improper use in domestic medicine, probably by the secret and criminal use of the leaves to increase the intoxicating effects of liquors, and, in children, by their eating the young sboots by mistake for the wintergreen (Gaultheria procumbens).

Dr. Johnson refers to the poisonous properties of the plant as follows:

"The leaves of the plant as also those of Kalmia angustifolia (Sheep-Laurel, Lamb-kill) are said to be poisonous to sheep and calves; and cases are reported of men being poisoned by cating the flesh of partridges which had fed upon the buds and berries. On the other hand, Wilson, the ornithologist, ate freely of the flesh of such birds without any ill effect whatever; and deer feed upon the leaves in winter, not only without ill effect upon themselves, but also without rendering their flesh unwholesome to man. And yet the common opinion is that leaves are very poisonous. The writer himself was cautioned by an old Massachusetts farmer against



Fig. 380. Mountain laurel (Kalmia latifolia). a, flowering spray, one-third natural size; vertical section of flower showing peculiar attachment of stamens, natural size. c, fruiting capsules, natural size. Vcry poisonous. (Chesnut, U. S. Dept. Agr.)

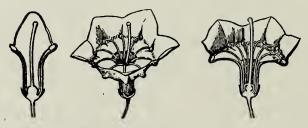


Fig. 381. Mountain Laurel (Kalmia latifolia). 1, section of flower bud showing anthers lodged in the pockets of the corolla; 2, expanded flower showing anthers in pockets; 3, sectional view. (Gray.)

meddling with K. angustifolia because of its poisonous nature; disregarding the advice, he ate several leaves in the presence of the farmer, without, however, convincing the latter, notwith-standing no ill effects were experienced. That persons have been poisoned seriously by eating the flesh of partridges in winter cannot be denied; that these birds may have fed upon kalmia buds and berries is also probable, but that their flesh is thus rendered poisonous does not seem as yet fully established. Some experimenters have reported effects produced in their own persons by strong decoctions of the leaves, similar to those of partridge poisoning; others quite as worthy of credence have failed to observe any sensible effect from them. Taking these contradictory statements in connection with the negative results of the chemical analyses thus far made, one may reasonably conclude either that the poisonous character of these plants has been greatly exaggerated, or that the energy of the poison, whatever it may be, is greatly influenced by the personal idiosyncrasy of the individual experimented with.

Dr. Millspaugh, after digesting the various opinions that have been given regarding the plant, makes this statement:

From the experience of nearly all persons who have experimented upon themselves with a tincture or decoction of the leaves, it is obvious that the effects produced on cattle after grazing on the leaves, and on persons eating of "poisoned" partridges, are due to the plant itself, not to indigestion or putrefaction. Dr. Bigelow's later observations agree in toto with our provings. He gives the following as its action: "The flesh of the bird impairs the functions of the brain and acts directly as a scdative poison, secondarily affecting the digestive and circulatory organs." The symptoms arising in those proving the drug are: Vertigo and headache; almost complete loss of sight; pale, somewhat livid countenance; salivation and difficult deglutition. thirst, nausea and vomiting, with oppression and pressure in the region of the stomach, difficult respiration with great palpitation, and fluttering of the heart, followed by an irregular, feeble, and slow pulse; weakness, weariness and pains in the limbs; coldness of the surface and great prostration."

There would seem to be no question then that this plant is poisonous.

Kalmia angustifolia, L. Sheep-laurel. Lambkill

Shrub from 1-3 feet high; leaves usually opposite, or in 3's; pale or whitish beneath, light green above; acute or narrowly oblong, petioled; flowers in simple or compound corymbs, purple or crimson; pedicels filiform, recurved in fruit; sepals ovate, acute, capsule depressed, globose.

Distribution. In moist soil from Eastern Canada to Newfoundland, from Hudson Bay south to Georgia and Michigan.



Fig. 382. Sheeplaurel (Kalmia angustifolia), showing flowcring branch, onethird natural size. (Chesnut, U. S. Dept. Agr.)

Kalmia polifolia, Wang. Swamp laurel

A low shrub with 2-edged branchlets, opposite nearly sessile leaves; oblong and white-glaucus beneath; margins revolute; flowers in terminal corymbs, few flowered; flowers ½ inch broad, purple; capsule depressed, glabrous, smooth.

Distribution. In bogs, Newfoundland to Alaska, Connecticut to Pennsylvania, Michigan, Rocky Mountains, and California.

Poisonous properties. Both of these species are regarded as poisonous, the K. angustifolia especially so. The symptoms of poisoning are very much the same as those produced by the preceding species.



Fig. 383. Branch ivy (Leucothoe Catesbaei): a, flowering branch; b, fruiting capsules. (Chesnut, U. S. Dept. Agr.)

Leucothoe, D. Don

Shrubs with alternate petioled leaves, and small, usually white, flowers in axillary or terminal spiked, racemes; sepals 5, distinct; corolla cylindrical, 5-toothed; stamens 10, included; anthers naked, or the cells with 1 or 2 erect awns at the apex, opening by a pore; capsule depressed, more or less 5-lobed, 5-celled, 5-valved; seeds mostly pendulous, minute. About 35 species, natives of the Western Continent; a few in Asia.

Leucothoe Catesbaei, (Walt.) Gray

Shrubs 2-4 feet high; leaves ovate-lanceolate; taper-pointed; serrulate, evergreen, with spinulose teeth; racemes dense, many-flowered; bracts borne at the bases of the short petioles; corolla narrowly cylindric; anthers awned; capsules depressed.

Leucothoe racemosa, Gray

Shrubs 4-10 feet high; branches and racemes mostly erect; leaves oblong to ovate, generally acute at each end; sepals ovate-lanceolate; anther cells 2-awned; stigma capitate; capsule depressed-globose; seeds smooth and wingless.

Distribution. Moist thickets and swamps from Massachusetts, to Florida, and Louisiana.

Poisonous properties. Both species are known to be poisonous in the Alleghany Mountain region to all kinds of stock; probably contain the same principles found in other plants of this order.



Fig. 384. Swamp Leucothoe (Leucothoe racemosa). A well known poisonous plant of the Alleghany Mountain region and the Southern states. (Charlotte M. King.)

Andromeda, L

Shrubs or small trees; leaves evergreen, short petioled; flowers in panicles, racemes or umbellate clusters; calyx persistent, without bractlets; corolla globose, urn-shaped, 5-parted; stamens 10, included; anthers fixed near the middle and opening by a pore; ovary 5-celled with columnar style; capsule globular and 5-celled, many seeded; seeds smooth. A small genus of 13 species found in Eastern Asia, the Himalayas, North America and Europe.

Andromeda Polifolia L. Wild Rosemary. Fetter-bush

A glabrous shrub 6-19 inches high, coriaceous; leaves with strongly revolute

margins, glaucus beneath; flowers in few-flowered drooping umbels; bracts persistent; calyx lobes triangular; corolla urn-shaped; stamens 10, anther with a slender terminal awn.

Distribution. From Newfoundland to New Jersey; Michigan to British Columbia and Alaska; Europe and Asia.

Andromeda floribunda, Pursh. Mountain Fetter-bush

A leafy shrub from 2-6 feet high; branches nearly erect, bristly strigose pubescent; leaves oblong to ovate-lanceolate, ciliate and glandular dotted be-



Fig. 385. Andromeda (Andromeda Polifolia). Common in the North, especially in bogs and swamps. (Ada Hayden.)

neath; flowers white in densely flowered panicles; corolla 5-angled; stamens 10; anthers without appendages; capsule somewhat globose, shorter than the style. Distribution. In the mountains of Virginia to Georgia.

Lvonia, Nutt

Shrubs. Leaves coriaceons and evergreen or thin and deciduous; flowers in racemes or panicles, white; calyx of 5 nearly distinct sepals; stamens 5; filaments hairy and often toothed or appendaged; anthers unappendaged; capsule 5-angled, dorsal suture with a thickened ridge; seeds with a thin testa.

Lyonia mariana, B. & H. Stagger-bush

A glabrous shrub with black dots, from 2-4 feet high; oblong or oval leaves, smooth above, slightly pubescent on the veins; flowers in nodding fascicles, racemose on nearly leafless branches; segments of calyx acute; corolla ovoid, white or pinkish; stamens 10; anther filaments 2-toothed near the apex; capsule ovate, as large as the sepals.

Distribution. Low grounds from Rhode Island to Florida, Tennessee and Arkansas.

Poisonous properties. All three species are poisonous. The leaves of the fetter-bush contain a narcotic poison, the andromedotoxin $C_{31}H_{51}O_{10}$, and have been known to kill sheep. Sheep have also been poisoned by the mountain fetter-bush. The stagger-bush received its name because of the intoxicating effect of its leaves on sheep and cattle.

PRIMULALES

Herbs or shrubs; corolla mostly present, gamopetalous; stamens borne on the corolla as many as its lobes or twice as many or more. The family *Plumbaginaceae* contains *Statice*, growing mostly in saline soil along the coast. It is used as an astringent, particularly in diarrhoea. The baycurn (*Statice brasiliensis*) is one of the most powerful astringents, and is used locally as a gargle.



Fig. 386. Stagger-bush (Lyonia mariana), showing flowering branch, one-third natural size. It contains a narcotic poison. (Chesnut, U. S. Dept. Agr.)

PRIMULACEAE. Primrose Family

Herbs with simple leaves and regular flowers; calyx 5-parted; stamens as many as the lobes of the gamopetalous corolla and borne upon it; ovary 1-celled, bearing several or many seeds; calyx free from the ovary or partly adherent A small order with about 28 genera, and 350 species, of wide distribution in the northern hemisphere, many of them arctic.

Few of the members of this order are weedy, or at least they are not troublesome. The honeywort (Lysimachia Nummularia) has escaped from gardens into damp ground and occasionally some species of the loosestrifes, are troublesome as weeds. Several members of this family are cultivated for ornamental purposes, among which are the cowslips or primroses and cyclamens. Native species of Steironema, like S. lanceolatum, and Lysimachia, are pretty ornamental plants. One of the prettiest of the native species in the northern



Fig. 387. Long and short-styled flowers of Primula. a and b styles; c pollen grains, and d papillae of short-styled flower. Modified by C. M. King, after Strasburger.

states is the American cowslip (Dodecatheon Meadia), which is stemless and has a 6-parted reflexed corolla and exserted stamens. The true primroses (Primula) are frequently cultivated, the P. sinensis and P. obconica in greenhouses. The English cowslip, P. vulgaris, is an interesting European plant, and is also found in high altitudes in North America. The beautiful P. Parryi, of the Rocky Mountains, is abundant along brooks and in moist meadows at high altitudes, and is one of the prettiest of the Rocky Mountain plants. The rootstock has a strong odor of musk and is said to be somewhat poisonous. The leaves of P. obconica are poisonous to the touch, being similar in effects to the poison ivy, although not so strong. The European cyclamen is a commonly cultivated plant in greenhouses. The pimpernel or poor man's weather-glass (Anagallis arvensis) is common in sandy soil and occasionally somewhat weedy. It is native to Europe. In Primula and cyclamen is found cyclamin, C₂₅H₄₂O₁₂, with which primulin is supposed to be identified.

Anagallis, L.

Herbs with alternate, sessile, or short-petioled whorled leaves; flowers solitary, on axillary peduncles; calyx 5-parted; corolla wheel-shaped, deeply 5-parted; longer than the calyx; stamens 5; filaments bearded; capsule globose, circumscissile, many-seeded; seeds minute, flat on the back. About 15 species, mostly in the old world.

Anagallis arvensis, L.

Annual; leaves ovate, or oval, membranous, sessile, or somewhat clasping; calyx-lobe keeled, shorter than the obovate corolla-segments, fringed with minute stalked glands; flowers scarlet or white, usually with a darker center; capsule smooth.

Distribution. Newfoundland to Minnesota, Mexico, and on the Pacific Coast. Naturalized from Europe.



Fig. 388. Persian Cyclamen (Cyclamen). Frequently cultivated in greenhouses. (From American Agriculturist.)

Poisonous properties. This plant is especially common on the Pacific Coast, where it is well known as "Poison weed," and according to Prof. Chesnut, is suspected of having caused the death of a horse at Santa Ana. It contains the glucoside $cyclamin\ C_{25}H_{42}O_{12}$, and a saponin-like substance, also a pepsin-like ferment. According to Hyams, the fluid extract in 4 drachm doses is fatal to dogs.

Primula, L.

Perennial scapose herbs, producing a tuft of basal leaves; flowers dimorphous, umbellate, or in bracted racemose whorls; calyx tubular, angled, 5-cleft, persistent; corolla funnelform or salverform; tube longer than the calyx; stamens 5, included; capsule many-seeded, splitting at the top into 5 or 10 valves. About 150 species, mostly in the northern hemisphere. This species is rather rare in eastern North America, but one species, (P. Parryi), is abundant in higher altitudes of the Rocky Mountains, and is suspected of being poisonous.

Primula obconica. Hance. Primrose

Leaves all radical and ovate-cordate; slender scapes 6-12 inches long; flowers blush-lilac or purple, frequently drooping; calyx saucer-shaped and shallow; petals deeply notched, obconical.

Distribution. Native to China, but frequently cultivated in greenhouses.

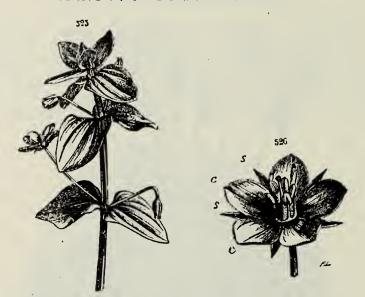


Fig. 389. Poison weed (Anagallis arvensis). Flowering branch. Flower. c, corolla; s, sepals. Poisonous to horses and dogs. (Prom Vesque's Traité de Botanique.)



Fig. 390. Parry's Primrose (Primula Parryi). This primrose is common in high altitudes. The roots have a more or less musky odor and it is regarded as poisonous. (Charlotte M. King.)

Poisonous properties. Primula obconica and P. sincusis both contain embelic acid $C_7H_3O_2(OH)_2C_{11}H_{23}$. It has long been known that some people are sensitive to poisoning by coming in contact with the leaves of P. obconica. The hairs act as an irritant poison. Dr. White calls attention to the injury as follows:

The eutaneous disturbance therein described, dermatitis venenata, subsided in a short time, and the skin of the three affected persons has remained in a healthy condition until recently. Within the last two or three weeks, however, they have all manifested a recurrence of the same symptoms, an eczematous inflammation of the hands and face, and in about the same degree as last year. They now feel assured that the trouble is caused by *Primula obconica*, for the condition did not develop until a few days after this plant was first offered for sale in the shop, where it was freely handled by them. The proprietor informs me that his hands and face became affected immediately after making it up into dinner-table decorations.

Dr. Thurber refers to a case of poisoning quoted by Mr. Meehan:

A person, who after potting a lot of Prinula obconica, had his face so swollen that he remained completely blind for a day. This was the severest case yet reported; but it should be stated that Mr. Cliffe was at that time suffering from poisoning by Euphorbia pulcherrima.

Mr. Thurber says:

Some have likened the effects produced by Primula obconica to those of Rhus Toxicodendron, the Poison Ivy, but it is a well established fact that many persons are poisoned by Rhus without coming in contact with the plant. Merely passing near it or being near a fire where it is being burned is sufficient to cause serious poisoning, indicating that the active principle is volatile.

EBENALES

Shrubs or trees, with alternate simple leaves; flowers generally regular; calyx more or less adnate to the overy; corolla gamopetalous or sometimes polypetalous; stamens as many as the lobes, with corolla opposite them or more numerous.

FAMILIES OF EBENALES

| Stamens as many as the corolla lobes | Sapotaccac |
|---|-------------|
| Stamens twice as many as the corolla lobes. | |
| Flowers dioecious or polygamous | Ebenaceac |
| Flowers perfect | Styracaceae |

SAPOTACEAE. Sapodilla Family.

Shrubs or trees with milky juice; simple and entire, alternate leaves; small and perfect, regular flowers; persistent sepals 4-7; corolla gamopetalous; tube 4-7 lobed, frequently with as many or more lobe-like appendages; stamens as many as the proper lobes of the corolla; fruit a fleshy drupe, generally 1-celled and 1-seeded; seed large, coat hard; endosperm fleshy or none.

A small order of 35 genera and 400 species, of tropical regions. But few of these occur in the United States, the most common in the southern states being the Southern buckthorn (Bumelia lycioides) and B. lanuginosa, both being common in the woods from Illinois southward. The star apple (Chrysophyllum Cainito) with a fruit about the size of an apple, has a pleasant flavor, but is said to be very constipating. The sapodilla plum (Achras sapota) also native to the West Indies, is widely cultivated in the tropics for its fruit, which is about the size of a pear and inferior only to the orange. The marmalade (Lucuma mammosa) of tropical America, from 3 to 5 inches long, is much esteemed for its pulpy fruit. The leaves, however, contain pumic acid, which is often used to adulterate cola nut. The fruits of the cainito (Lucuma obovata) a native of Peru and Chili, are also eaten, as are the fruits of the black bully

tree (Dipholis salicifolia) of Jamaica. Seeds of the buttertree (Bassia butyracea) furnish an oil which is an article of some commercial importance in India. The seeds of the buttertree (Butyrospermum Parkii) of South Africa, furnish an oil which is used as food by the natives. Gutta-percha (Dichopsis Gutta), of Borneo, and the Malayan Peninsula is the source of some of the Gutta-percha of commerce, which is used in surgical dressing, the same sub-



Fig. 391. Gutta-percha Tree (Dichopsis Gutta). The gutta-percha of commerce is obtained from the latex of this tree. (After Faguet.)

stance being also obtained from Minusops Kauki, a large tree, native of British Guiana. The former contains gutta $C_{10}H_{10}N$, fluavil $C_{10}N_{11}O$, and alban $C_{40}H_{64}O_2$. Sapotin $C_{20}H_{22}O_{20}$ occurs in the bark of Achras Sapota and of Symplocos racemosa. In lotus bark occur three basic substances: loturin, colloturin and loturdin. In the seeds of Illipe Maccleyana a toxic glucoside, C_{17}

H₃₂O₁₀, is found.

The Sapotaceae or Sapodilla family contains a number of plants of economic importance in the tropics; among these are Minusops Kauki, Dichopsis Gutta, Palaquium oblongifolium, of the East Indies, and other species which furnish gutta percha. This is obtained from the latex of these plants and because of its plasticity, is particularly valuable in the manufacture of surgical instruments and for temporary filling of teeth. The same substance is produced by Sideroxylon tomentosum and Payena. The Sapodilla tree (Achras Sapota) of Yucatan produces an aromatic gum resin which is largely used in the manufacture of chewing gum. Vegetable butter is obtained from the seeds of Bassia butyracea. The Sapodilla or Sapote (A. Sapota) is commonly cultivated in the tropics, especially the West Indies and Central America, for its wild fruit. The star apple, Chrysophyllum Cainito, also produces a valuable fruit. Hard iron woods are produced by Mimusops Kanki of Australia and Argania Sideroxylon of Morocco. Saponin-like substances are produced by several plants of the family such as Omphalocarpum procerum. The Lucuma salicifolia and Payena latifolia contain hydrocyanic acid.

EBENACEAE. Ebony Family

Trees or shrubs with very hard wood, alternate entire leaves and polygamous regular flowers; stamens from 2-4 times as many as the lobes of the corolla; ovary 3-12 celled; ovules 1-3 in each cavity; fruit a several-celled berry; seeds mostly single in each cell, large and flat, with a smooth coat.

About 6 genera and 275 species. The most widely distributed tree in the United States is the common persimmon (Diospyros virginiana), a tree from 20-70 feet high, found in woods and old fields from Rhode Island to New York and southern Iowa. The fruit is commonly eaten and the unripe fruit is used in medicine for internal hemorrhage. The Japanese persimmon (D. Kaki) with small flowers and a greenish-yellow fruit, of variable size and shape, is cultivated in the Southern States and California for its fruit, which is delicious. In Egypt it is cultivated under the name of Lotus tree.

The Indian persimmon (D. Embryopteris) is a middle-sized evergreen tree native from India to Java, and is used in India in medicine, especially for chronic dysentery. Ebony wood comes from several species like the Ceylon ebony, Indian cbony (D. Ebenum) and (D. melanoxylon), and is highly prized for furniture since it is hard and black. The calamander wood (D. quesita) of Ceylon is a hard wood prized for making boxes. The mangosteen or mabola (D. discolor), a native of the Philippine Islands, is now commonly cultivated in the tropics for its fruit, which is about the size of a quince and has a pleasant flavor. Other hardwooded trees are D. Cargillia and D. pentamera, natives of New South Wales.

Ebony wood is of various colors; the black from D. ebenum; red ebony from D. rubra, native of Mauritius; white ebony from D. chloroxylon. Other species of Diospyros furnish a striped ebony, and others a green.

STYRACACEAE. Styrax Family.

Mostly tropical shrubs or trees with regular perfect, or polygamo-dioecious flowers; calyx free or adherent to the ovary; corolla gamopetalous, or polypetalous; the lobes or petals 4-8; stamens twice as many as the lobes of the corolla, or more, inserted on its tube or base; the filaments monadelphous, or in 4-5 sets; style 1; ovary 2-5-celled; fruit a berry or drupe.

About 75 species and 7 genera, few of them native to the United States. Several species are cultivated for ornamental purposes, especially the snowdrop or silver bell tree (Halesia carolina) which is a small tree native to the southern states. Styrax Benzoin is obtained from Sumatra and Java, benzoin being obtained by cutting incisions into the plant, from which a resin exudes and soon hardens by exposure to the air. Benzoin is used chiefly as incense in the service of the Greek Church.

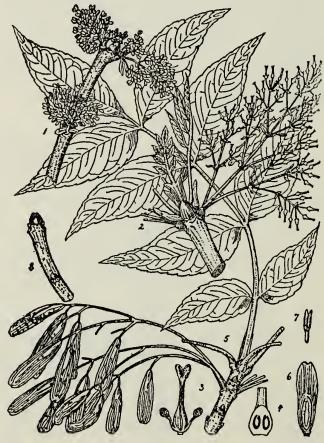


Fig. 392. Black Ash (Fraxinus nigra). 1. Flowering branch of staminate tree. 2. Flowering branch of pistillate tree. 3. Pistillate flower showing rudimentary stamens, enlarged. 4. Longitudinal section of ovary, enlarged. 5. Fruiting branch. 6. Longitudinal section of fruit. 7. Embryo. 8. Winter branchlet. 1, 2, 5, 6, 8, one-half natural size. (M. M. Cheney in Green's Forestry of Minnesota.)

Styracin, $C_0H_{10}O_7$, one of the ketones, occurs in Styrax Benzoin, also resin, benzoic acid $C_7H_6O_2$, and cinnamic acid $C_9H_8O_2$. Siam benzoin contains vanillin.

CONTORTAE

Herbs, shrubs or trees; leaves opposite or rarely alternate; flowers regular; corolla gamopetalous, rarely polypetalous or wanting; stamens generally borne on lower part of corolla when present, as many as the lobes or fewer; ovaries 2, distinct.

FAMILIES OF CONTORTAE

OLEACEAE. Olive Family

Trees, shrubs, or some nearly herbaceous; leaves opposite or rarely alternate, simple or pinnate without stipules; flowers in panicles, cymes or fascicles; calyx small, inferior, sometimes wanting; stamens 2-4; filaments separate; ovary superior, 2-celled; ovules a few in each cavity; style short or none; fruit a capsule, berry or drupe. About 500 species. Contains the lilacs (Syringa vulgaris, S. persica), the jasmine (Jasminum grandiflorum) cultivated in France as the source of a valuable perfume; the fringe tree (Chionanthus virginica), well known in cultivation, and the ashes which make valuable timber trees like green ash (Fraxinns pennsylvanica, var. lanceolata), white ash (F. americana), black ash (F. nigra) and manna of Europe, F. Ornus. The sweet exudate is called manna which contains mannetol and a glucoside resembling aesculin.

The olive (Olea enropaea) of the Mediterranean region, is cultivated extensively in southern Spain, France, Italy to Arabia, California, and other warm countries. The unripe fruit is used for table purposes by steeping in water containing lime and ash, and then pickling in a salt solution. The nearly ripe or ripe olives are used in a similar way. The seed as well as fruit contains an oil consisting of olein $C_3H_5(OC_{18}H_{33}O)_5$, the greenish color of which is due to the chlorophyll. It is nutritive and laxative. The hard wood takes a fine polish.

Edwards and Power have found an alkaloid in the bark of Fraxinus americana, one in the bark of Olea glandulifera, in the leaves and bark of Ligustrum robustum, and in the leaves of Jasminum. The glucoside phillyrin, $C_{20}H_{32}O_{11}$, occurs in Osmanthus fragrans and Forsythia suspensa, and may be converted into phillygenin, $C_{20}H_{22}O_{0}$, which may again be readily converted into ugenol and vanilhin. Ibo $C_{20}H_{23}O_{19}$, a saponin-like glucoside, occurs in Ligustrum and Ibotu; chionanthin is found in the bark of Chionanthus virginica, and is intensely bitter. It may also contain saponin. The plant is probably somewhat poisonous.

Ligustrum (Tourn.) L.

Shrubs or small trees with opposite entire leaves; flowers in a terminal thyrsus or panicle; corolla gamopetalous, funnelform, tube short, the limb 4-



Fig. 393. Large-flowered white Jessamine (Jasminum grandiflorum). A native of Japan but cultivated in France as a source of perfume. (From Vesque's Traité de Botanique.)

lobed; stamens 2; filaments short; ovary 2-celled, with 2 ovules in each cavity; fruit a 1-3 seeded berry. About 35 species, natives of the Old World.

Ligustrum vulgare, L. Privet

A shrub with long and slender branches, firm leaves; smooth, lanceolate or oblong, and short petioled; panicles dense, minutely pubescent; pedicels very short; flowers white.

Distribution. Commonly cultivated for ornamental purposes, but escaped here and there from cultivation.

Poisonous properties. The leaves and fruit of the plant are said to be poisonous. Prof. Chesnut states that accidents have been occasioned to children who have eaten the fruit or the leaves. The plant contains syringin and the bitter glucosidal principle syringopicrin $C_{26}H_{24}O_{17}$. These also occur in the lilac. Edwards and Power have also found an alkaloid in the leaves and bark of an allied species, Ligustrum robustum; and it was discovered also to be in the seeds of L. Ibota.

LOGANIACEAE. Nux Vomica Family.

Herbs, shrubs, vines, or in a few tropical genera, trees; simple, opposite leaves, stipules or small membranes; flowers perfect and regular in axillary or

cymose clusters; calyx of 4 to 5 parted sepals; corolla gamopetalous, 4 to 5 parted; stamens as many as the lobes of the corolla or alternate with them; fruit follicles or drupes.

About 30 genera and 400 species, in warmer regions, several of them very poisonous. Strychnos Nux-vomica, native to India and the islands of southern Asia, is a small straggling tree, in its native country known as the Koochia tree. The fruit resembles an orange, but with a hard rind and numerous flat



Fig. 394. Strychnos (Strychnos Crevauxiana). A curare plant of French Guiana. (After Faguet.)

seeds covered with silky hairs. The seeds contain two deadly alkaloids, strychnin $C_{21}H_{22}N_2O_2$ and brucin $C_{23}H_{26}N_2O_4+_4H_2O$. Strychnin is bitter, used as a tonic and to stimulate the circulation. The bark and root are also bitter. The natives of India use it for snake bites and fevers. This alkaloid is also obtained from other plants of this order, being extracted by water acidulated with hydrochloric acid. It has an intensely bitter taste perceptible in very dilute solution (1 in 700,000).

Delafoy has shown that starving frogs are much more sensitive to strychnin than are normal frogs.¹

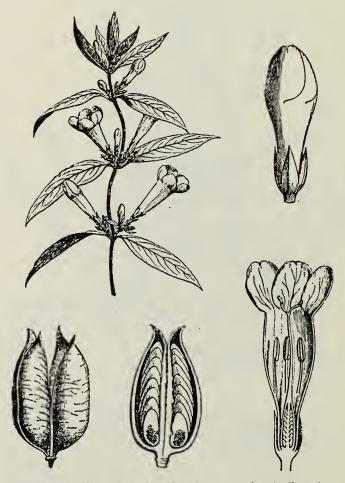


Fig. 395. Yellow Jessamine (Gelsmium sempervirens.) Flowering branch, bud, deliiscent fruit, longitudinal sections of fruit and flowers. Root contains poisonous alkaloid. (After Faguet.)

¹ In a recent paper by Reid Hunt "The Effects of a Restricted Diet and of Various Diets Upon the Resistance of Animals to Certain Poisons," Bulletin Hygienic Laboratory, Public Health and Marine Hospital Service, of the U. S. Treasury Department, he reports that diet has a marked effect upon resistance of animals to certain poisons; the resistance of some

It contains the glucoside loganin C25H34O14 especially in the pulp. The S. toxifera of the Orinoco, used as an arrow poison and called "curarie," or curare, is obtained from the stripped bark, which is steeped in water. It contains the alkaloid curarin C₁₀H₂₆N₂O. Other alkaloids found in curare are protocurin C20H23NO3 which is poisonous, tubocinarin C19H21NO4 and curin C₁₈H₁₇NO₃. The dried seeds of S. potatorum are used for clearing water. The Tieute of Java is a strong poison causing tonic and clonic convulsions and there is no known antidote. It is used as an arrow poison. S. Crevauxiana of French Guiana is used as a curare poison.

Dr. Winslow describes the toxicological effect of strychnin on a dog as follows:

One-twentieth of a grain of strychnin nitrate, injected subcutaneously by the writer into a dog weighing 25 lbs., caused uneasiness and excitement, with protrusion of the eye-balls, and in the space of ten minutes, tetanic convulsions. The breathing was shallow and almost imperceptible, the pulse rapid and irregular, the lips were covered with foam, the tail was stiff and extended, the cars laid back, and there was general muscular rigidity, the animal lying on his side in a state of opisthotonos. This condition lasted about three minutes, and was followed by a period of relaxation. But the slightest noise or irritation of the skin brought on convulsions. The convulsions became less frequent and violent, and ceased altogether within half an hour. The same animal was given gr. 1.40 of the alkaloid on the following day, hut without producing any appreciable result. One-tenth of a grain, given on another day and in the same manner, caused immediate uncasiness and restlessness, and in ten minutes induced a severe convulsion, lasting for three minutes, in which the animal was so rigid that he could be lifted bodily without bending. The cars were drawn back, the limbs were extended and stiff, the tail was straight and rigid, and there was twitching of the muscles of the jaw and limbs. The corners of the mouth were drawn back (risus sardonicus), the mouth was covered with foam, and there was some trismus. The breathing was nearly suppressed, owing to tetanic spasm of the respiratory muscles. Following this convulsion, the jaw dropped, the muscles relaxed and another attack could not be produced by noises or external irritation. Some twitching of the temporal muscles persisted. Evidently the second stage of poisoning had ensued, and the motor nerves and cells of the inferior cornua had become paralyzed. Death occurred in general paralysis within half an hour, and without any recurrence of convulsions or tetanic condition. Death takes place more commonly in strychnin poisoning from asphyxia, during a convulsion, and is caused by spasm of the respiratory muscles, or, more rarely, by spasm of the glottis. Sometimes a fatal result ensues from exhaustion, between the paroxyms, and occasionally death appears to follow the intense action on the nervous system, and depression of the heart.

The lethal dose for dogs has been set at gr. 1-6-gr. 1-3 (Kaufmann). This is much too large, as evidenced by the experiment mentioned above. The fatal amount varies greatly in accordance with the weight of the animal; probably less than gr. 1-20 would kill toy terriers, and eases are reported where they have been destroyed by gr. 1-60 of the alkaloid. The therapeutic dose should, therefore, he proportioned, as advised, to the weight of the animal. Five to eight grains of nux vomica will kill a dog.

animals must be increased forty-fold by changes in dict. It was found that diet causes distinct, but not very marked, differences in resistance to morphin. The experiments further show that foods such as enter largely into the daily diet of man have most pronounced effect upon resistance of animals to several poisons. They produce changes in metabolism but are not readily detectable by means ordinarily used in metabolism studies. Dict causes distinct but not very marked differences in resistance to morphin. It was found by Hunt that the effect of oatmeal diet in increasing the resistance of certain animals to acetonitrile is probably due in part to a specific effect of the diet upon the thyroid gland. The poisonous action of acetonitrile, according to Hunt, is largely due to the formation of hydroeyanic acid from it. This is the view expressed not only hy Dr. Hunt, but also by Heymans and Masoin.

In the paper hy Dr. Hunt referred to above, a statement is taken from the work of Mansfield that starving rabbits are much more susceptible to chlorallydrate, paralelyde, and morphin than well nourished ones. Lewin stated that starving animals are more resistant to quinin, atropin, and nicotin, than well nourished ones. Salant and Rieger found the resistance of rabbits to caffein to be diminished when the animals were starving for four or five days. The fatal dose was about 30 percent less than in well fed rabbits. In this connection it is also of interest to state that Hunt found that season has an important effect upon the resistance of animals to certain poisons. In some cases these effects seem to depend upon seasonable variations in the activity of the thyroid.

Foster, who experimented with dogs and ricin, found that when dogs were kept upon a high protein nutrition all died from 1 mgm. ricin per kilo, and of three kept on medium low plane of protein nutrition, two survived this dose.

The minimum fatal dose of strychnin for man is one-half a grain. Usually four to seven grains constitute a lethal quantity, but recovery has ensued following the ingestion of 22 grains, after a full meal.

Horses—The toxic symptoms in horses resemble those already described in the dog. They do not appear for some time (20 minutes to 6 hours), depending on the rapidity of absorption when the drug is swallowed, and include excitement, muscular spasm and convulsions, increased frequency of the pulse, and difficult respiration. Death occurs in convulsions or in the interim between them. The minimum fatal dose of strychnine, when given under the skin, is about 1½ to 3 grains, and when swallowed, 3 to 5 grains of the alkaloid, or 1 to 2 ounces of nux vomica.

Cattle are similarly affected with horses and dogs. There are exhibited muscular spasms, frequent pulse, difficult respiration, sensitiveness to light, sounds and external stimuli, protrusion of the eye-balls and convulsions. The fatal dose, by the mouth, varies greatly owing to difficulty of absorption in the complicated and capacious digestive apparatus of these ruminants. This is true of all medicines. When given under the skin, the lethal dose is a little larger than that for horses. The fatal dose for swine is said to be from gr. 1-6-gr. 34. Chickens are comparatively insusceptible; also guinea pigs and some monkeys.

Strychnin poisoning differs from tetanus in the fact that muscular rigidity is continuous in the latter, but disappears to a considerable degree, if not completely, in the periods between the convulsions, in the case of strychnin poisoning. Moreover, in tetanus the body and limbs are less, and the jaw more affected; while in strychnin poisoning the condition is reversed.

Treatment—The treatment embraces the use of chemical antidotes, as iodine or its salts, or tannic acid; animal charcoal and emetics or the stomach tube, before absorption has occurred. The best physiological antidote is chloral in large doses per rectum. Chloroform and nitrate of amyl may also he given by inhalation, and quiet and rest enforced. Artificial respiration is of no service on account of the muscular spasms, unless air he forcibly driven into the trachea through a canula. Calabar hean and gelsemium both depress the inferior cornua, but neither is of much value in strychnin poisoning.

A large number of the cases of poisoning are fatal; according to Schauenstein 62 out of the 130 reported by him proved fatal. According to Falk, the minimum lethal dose is as follows: .6 mgrm. strychnin nitrate for rabbit per kilogramm; cats, .75 mgrm; frogs, 2 mgrm. In one case ½ grain strychnin sulphate produced death in 20 minutes, in another case 7/10 grain was required. The so-called vermin killers contain strychnin.

Strychnin is the best remedy to stimulate the action of the heart, and to promote appetite and digestion. It is valuable in chronic tympanites of cattle.

The St. Ignatius or False Angustura Bark (S. Ignatii) of the Philippine Islands, much used in medicine in the Philippines, contains the same substance as S. Nux-vomica. The S. malaccensis of Tonquin is used as a remedy for leprosy. Pink root (Spigelia marilandica) is a perennial herb; with opposite sessile leaves; tubular funnel-form corolla, red outside, yellow within; 6 stamens; slender style; short 2-celled capsule; found from Ohio to Florida and Texas. It contains a volatile alkaloid spigelin resembling coniin and a bitter acrid principle. It is a powerful anthelmintic. According to Hyams, when taken in overdoses it excites the circulation, causes dimness of vision, vertigo, dilated pupils, spasms of facial muscles and general convulsions, followed by death. It is especially fatal to children. Dr. True reported the plant to be poisonous. According to Dr. Stockberger this is frequently adulterated with Ruellia ciliosa (East Tennessee Pink-root). The prepared drug may contain, as impurities, roots of golden seal, serpentaria, soapwort, wild yam, and stonewort.

Gelsemium, Juss. Yellow Jasmine

Smooth vines with opposite or whorled leaves; flowers in axillary cymes, regular, perfect; calyx deeply 5-parted; corolla funnel-form, 5-lobed; stamens 5, inserted on the tube of the corolla; pistil with a 2-celled ovary; style slender,

or filiform, 4-cleft; fruit a capsule, flattened, seeds winged. A small genus of 2 species, one in North America and the second in Asia.

Gelsemium sempervirens (L.) Ait. Yellow Jessamine.

A well known poisonous woody vine climbing over shrubs and trees to the height of thirty feet or more; with opposite or entire ovate or lanceolate leaves, shining and evergreen; flowers showy and fragrant in short axillary clusters, yellow.

Distribution. This plant grows in woods and low grounds from eastern Virginia to Florida, and as far west as Mexico to Guatemala.

Poisonous properties. It contains a poisonous alkaloid, gelsemin $C_{24}H_{28}N_2O_4$. The root is frequently used in medicine and poisoning has been caused by overdoses. It is used in febrile and inflammatory infections, and on the nervous system it is an antispasmodic sedative. It is, however, a powerful drug and should be used with great caution. In overdoses it produces nausea, pain in brows and eye-balls, dilation of the pupils, paralysis and dimness of vision. When overdoses have been administered, stomach pump and coffee should be used at once in conjunction with whiskey. Gelsemin is a colorless, crystalline, bitter principle. This plant also contains the amorphous, bitter alkaloid, gelseminin $C_{22}H_{23}(OH)N_2O_2$.

Dr. Winslow gives the toxicological effects on animals as follows: Muscular weakness, especially in the fore legs, staggering gait and falling. These symptoms are followed by convulsive movements of the head, fore legs, and sometimes of the hind legs. The respiration is slow and feeble, temperature reduced, and there is sweating. Death occurs because of respiratory failure. Morphine subcutaneously has proved a good antidote.

Blyth records that 10 mgrs. killed a frog in 4 hours, 8 mgrs. killed a cat in 15 minutes; 1/6 grain killed a woman in 7½ hours.

GENTIANACEAE. Gentian Family.

Smooth, bitter herbs with opposite and sessile entire mostly simple leaves without stipules; flowers regular and perfect in clusters or solitary; calyx persistent, 4-12 lobed; corolla gamopetalous, 4-12 lobed or parted, mostly persistent but withered; stamens as many as the lobes of the corolla, alternate, inserted in the throat of the corolla; pistil with a 1-celled or partly 2-celled ovary; ovules numerous, anatropous or amphitropous; fruit a capsule, mostly dehiscent by 2 valves; seeds small, with a minute embryo and fleshy endosperm.

About 600 species of wide distribution. Several plants of the family are used in medicine. The American centaury or rose pink (Sabatia angularis) furnishes a simple bitter tonic formerly much used in intermittent fevers. An allied species contains the glucoside erythrocentaurin, $C_{27}H_{24}O_8$. The Chirata (Swertia Chirata) a bitter tonic, contains the glucoside chiratin $C_{26}H_{48}O_{15}$, a very bitter substance, and ophelic acid $C_{13}H_{20}O_{10}$. The root of the American columbo (Frasca carolinensis) is used as an emetic and cathartic or as a tonic The F. speciosa is common in the Rocky Mountains. Several species of the gentian, as Gentiana Andrewsii, G. puberula, are used as bitter tonics.

The gentians are largely used in the preparation of the so-called stock foods, the chief constituents of the root being a bitter crystalline glucoside, gentiopicrin $C_{28}H_{30}O_{12}$ and gentisic acid $C_{14}H_{10}O_5$. The plants act as stomachies and bitter tonics.

Gentians are pretty flowers, but few of them are cultivated. The prettiest of these are the closed gentian (G. Andrewsii), with large blue flowers, the white closed gentian (G. flavida), the fringed gentian (G. crinita), and the Texas lily (Eustoma Russellianum), native from Texas to Nebraska and introduced into Colorado, with handsome large blue and white flowers.

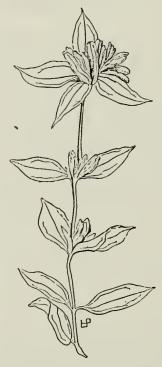


Fig. 396. Yellowish Gentian (Gentiana flavida). A plant with bitter properties frequently used in connection with stock foods. (Lois Pammel.)

Menyanthes (Tourn.) L.

Smooth marsh herb with creeping rootstocks; leaves 3-foliolate; flowers conspicuous white or purplish, in racemes or panicle; calyx 5-parted; corolla short, funnel-shaped, 5-cleft; stamens 5, inserted on the tube of the corolla; ovary 1-celled; capsule oval, indehiscent.

Menyanthes trifoliata L. Buckbean. Bog-bean

Perennial from scaly rootstocks; leaves petioled, sheathing at the base; leaflets oblong or obovate; raceme 10-20 flowered; calyx shorter than the white or purplish corolla.

Distribution. Europe and Asia, and North America, especially northward from New England to Greenland and Alaska, Iowa and Minnesota.

Poisonous properties. Buckbean is bitter and suspected of being poisonous. It causes nausea, and contains the substance menyanthin $C_{33}H_{50}O_{14}$, in the green parts of the plant.



Fig. 397. Buckbean (Menyanthes trifoliata). This plant is said to be more or less poisonous. It is very common in low lands. (Charlotte M. King.)

APOCYNACEAE. Dogbane Family

Perennial herbs; shrubs or vines with acrid or milky juice; entire, mainly opposite leaves; without stipules; and perfect, regular 5-parted flowers; the 5 lobes of the gamopetalous corolla convolute and twisted in the bud; stamens as many as the lobes of the corolla, alternate with them and inserted on the tube, or throat; pollen granular; ovary superior, with 2 distinct ovaries; fruit follicles or drupes; seeds with a large straight embryo, often bearing a tuft of hairs. About 125 genera and 1000 species. Very widely distributed, chiefly in the tropics.

The best known representative, perhaps, under cultivation is the oleander (Nerium Oleander) from the Levant, naturalized in Southern Europe, the Southwest and Mexico, and the Nerium odorum, of India. The periwinkles, Vinca minor and Vinca major, are common in cultivation, the most widely distributed being the common periwinkle (Vinca minor) erroneously called myrtle, found especially in country gardens and running wild in cemeteries and shady places. Some species of Gynopogon have a cumarin-like fragrance. Rubber is obtained from Ochrosia, Laudolphia, Kickxia, Haucornia, Urceola and others. Of the medicinal species, the Alstonia, or Dita Bark, is a handsome forest tree from 50 to 90 feet high; common throughout the Indian Peninsula

to Ceylon. The Dita Bark contains the alkaloid ditamin $C_{16}H_{10}NO_2$, recommended as a tonic and an antiperiodic; also the alkaloids echitamin $C_2H_{28}N_2O_4$ and echitemin $C_{20}H_{27}NO_4$. In the bark of Alstonia spectabilis, besides ditamin and echitemin there is found a third alkaloid alstonamin. An allied species native of eastern Australia, is used for ague. In North America the dogbanes are also used in medicine. The ordeal tree of Madagascar (Cerbera Tanghin) produces a very poisonous narcotic seed. It is used by the natives to poison persons, and formerly criminals were put to death by being pricked with a lance dipped in the juice of the kernel. A single seed is said to contain enough poison to kill 20 people.

Both species of Apocynum are used in medicine. The Indian hemp used as an emetic, cathartic and diuretic contains the glucoside apocynein and a bitter resin-like extractive, apocynin, both of which Schmiedeberg isolated. The Thevetia neriifolia of tropical America contains a powerful heart poison, thevetin $C_{54}H_{48}O_2$, and theveresin. Strophanthus Kombe of East Africa is a woody climber, and contains strophanthin $C_{10}H_{10}O_8$ which acts like digitalin, being valuable in cardiac disease; it is also used as an arrow poison.* The S. hispidus contains the same glucoside and an alkaloid inaein. The action of this drug is similar to Digitalis although not as efficient. The Querbracho (Aspidosperma Querbracho blanco) of Argentina furnishes a valuable tanning bark containing 3-4 per cent of tannin; it also contains alkaloids, six of which have been discovered, among them being aspidiospermin $C_{22}H_{30}N_2O_2$, aspidospermatin and querbrachin $C_{21}H_{20}N_2O_2$, aspidosamin and quebrachinamin. The yellow flowered Nightshade (Urechitea) of the West Indies is a cardiac poison not unlike Digitalis,

Apocymum. Dogbane

Perennial herbs with opposite entire leaves; small white or pink flowers; calyx 5-parted, with acute lobes and corolla bell-shaped, 5-cleft with 5 small triangular appendages below the throat opposite the lobes; stamens 5, inserted on the base of the corolla, with arrow-shaped anthers; style none; stigma large, ovoid, slightly 2-lobed; fruit of 2 long slender follicles; seeds numerous, small, each with a long silky coma. A small genus of about 15 species, found chiefly in the north temperate zone.

Apocynum androsaemifolium L. Spreading dogbane.

Rootstock horizontal, smooth, or rarely soft-tomentose, branched above,

The common names of a great many plants indicate their supposed virtues as antidotes for snake bites. In the last edition of Gray's Manual, a few of the common names of this class are among the Compositae; as Rattlesnake weed (Hieracium venosum), Rattlesnake root (Prenanthes alba, P. serpentaria), Rattlesnake master or Button snakeroot (Eryngium yuccifolium), Black snakeroot (Sanicula marilandica); among the Orchidaceae Rattlesnake plantain (Epipactis pubescens); of the Gramineae, Rattlesnake grass (Glyceria canadensis), perhaps so named because of the rattling of the spikelets; Snakegrass (Eragrostis megastachya), so called because of the odor of the grass; of the Ophioglossaceae, Rattlesnake fern (Botrychium Virginianum).

^{*}There are also many snake poisons belonging to different families.

In the family Acanthaceae are Andrographis paniculata, Rhinacanthus communis, Barleria cristata and B. lupulina, the latter of which contains not only a large amount of potassium but some alkaloids also.

spreading; leaves ovate, petioled; cymes loose, spreading, both terminal and axillary; calyx segments shorter than the tubes of the corolla; the latter pale rose color, open, bell-shaped.

Distribution. Common borders of thickets from Canada to British Columbia to Arizona and Georgia.

Apocynum cannabinum L. Indian Hemp

Glabrous or more or less soft-pubescent, 2-3 feet high, smooth, terminated by an erect, close, many-flowered cyme; corolla lobes nearly erect; the tube not longer than the lanceolate segments of the calyx; greenish white; appear in July and August.

Distribution. Common species eastward and troublesome as a weed in northern Mississippi Valley.

Poisonous properties. Said to be poisonous to live stock.

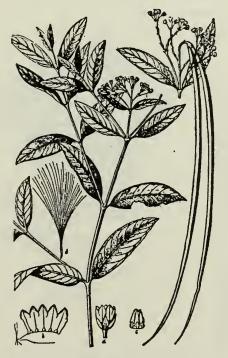


Fig. 398. Indian Hemp (Apocynum cannabinum). Flowering and fruiting branches. Bundle of fibers from stem. Section of flower. Furnishes a good bast fiber. (Dodge, U. S. Dept. Agr.)

Nerium. Oleander.

Shrubs; leaves coriaceous, rigid, closely and transversely veiny; flowers showy, in terminal cymes; corolla salverform or tube narrow, funnelform; stamens attached to the middle of the tube; style 1; ovaries 2, forming pods; seeds tufted. Native to the Levant and India.

Nerium Oleander L.

Leaves lanceolate; coriaceous, rigid, closely and transversely veiny; flowers in terminal cymes, rose-color or white; anthers scarcely protruding.

Distribution. Native to the Levant, naturalized in Southern Europe, and Southwest United States. Frequently cultivated in greenhouses.

Poisonous properties. John Smith, in his Domestic botany, says, with reference to its poisonous properties:

It grows abundantly in the valley of the Jordan, and when in flower is very beautiful. The whole of the plant is poisonous, and it is recorded that soldiers in Spain were poisoned through their meat being roasted on spits made of the peeled stem.

Prof. Chesnut states that stock are occasionally poisoned by cating the leaves, as the plant grows wild in northern Mexico and is abundant in the Southwest. The oleander is a heart stimulant acting like digitalis. Dr. S. Wateff recently reported a case in which gastro-enteritis occurred; nausea, vomiting and irritation were prominent symptoms. He also reports the odor of the flower as poisonous.



Fig. 399. Oleander (Nerium Oleander). At the left, a stamen; fl. thread enlarged; an, cells of anther; elongated end of connective. At the right, flowering branch. Whole plant poisonous. (From Vesque's Traité de Botanique.)

Prof. F. W. Wilson of the Arizona Station 1 has recently brought together the literature on the subject of oleander poisoning, also giving some of his own experiments.

Prof. Wilson finds that both the pink and white varieties are poisonous. He conducted some experiments with two horses, a cow, a mule, and three sheep:

¹ Bull. Ariz. Exp. Sta. 59:381.

The amount of oleander necessary to cause death in horses ranges from 15 to 20 gm, of green leaves, and from 15 to 30 gm, of dry leaves. This depends on the condition of the animal at the time the poison is obtained. A full stomach will necessitate more poison. In the case of cows it is safe to say that from 10 to 20 gm, of green leaves and 15 to 25 gm, of dry leaves are sufficient to cause death. For sheep the fatal dose of either green or dry leaves is from 1 to 5 gm. There is little danger in the bark, roots, or flowers since live stock would hardly obtain sufficient poison in that way. . .

The general symptoms are increased temperature and pulse, coldness of the extremities, warm body temperature, dilation of the pupils of the eyes, and discoloration of the mouth and nostrils followed by sore mouth. The body becomes wet with sweat, due to the exertion caused by the powerful heart stimulation. The animal generally refuses to eat or drink during the 24 hours preceding dealls. This is usually due to soreness of the mouth and throat, making it painful to masticate and swallow food. The bowels act often and feces are usually greenish in color. The action of the kidneys is increased slightly and color of urine is normal. There is little doubt that numerous cases of oleander poisoning have never been brought to light because of death being attributed to other sources. It is safe to say, however, that many hundreds of animals have been lost in southern Arizona from this shrub.

ASCLEPIADACEAE. Milkweed Family.

Perennial herbs, vines, or shrubs, with milky juice and opposite or whorled leaves, entire; flowers in umbels, regular; calyx inferior; corolla bell or urnshaped, rotate or funnel-form, 5-lobed or 5-cleft, the segments generally reflexed; a crown between the corolla and stamens; stamens 5, inserted on the corolla; generally monadelphous; anthers connivent around the stigma or more or less united with each other; commonly bearing an erect or inflexed membrane; pollen collected in masses, generally 10, known as pollinia; ovary consisting of 2 carpels, in fruit of 2 follicles; seeds flattened, usually appendaged by a long tuft of hairs called a coma.

About 200 genera, and 1800 species, of wide distribution, many members of the family being weedy; some are medicinal, but few are economic. Several plants of the family are cultivated. The Periploca gracea of the Old World is an ornamental climber and produces granular pollen in place of pollinia. The Stapelia, several species of which are cultivated, are natives of the Cape of Good Hope. They produce flowers of dull purple color with transverse stripes, exhaling a very disagreeable odor not unlike that of putrid meat.

The wax plant (Hoya carnosa), is a well known house plant of India, with rooting stems; thick, fleshy, oval leaves; and flesh colored flowers. The Vincetoxicum is a European climber sometimes cultivated in the Eastern States. Several species native from South America are sometimes cultivated for ornamental purposes. Some species of the order are used in medicine. The Indian sarsaparilla (Hemidesmus indicus) growing in the Indian peninsula and Ceylon, has a medicinal root which is used as a tonic and diuretic. The root has the odor of the tonka bean or of sweet clover. The mudar (Calotropis procera) also a native of India, from Ceylon and the Moluccas, is common in waste grounds. The bark contains one bitter principle, mudarin, used as a tonic and diaphoretic and in large doses as an emetic. It produces a strong fiber, the silk being exported as "kapok," or tree cotton. The Indian ipecacauanha (Tylophora asthmatica) is a climbing perennial of India and Mauritius; anciently much used by the Hindoos in dysentery. The pleurisy-root or butterfly-weed (Asclepias tuberosa) is used as a diuretic and an emetic. A. Curassavica of the West Indes is a vermifuge. The flowers are said to produce excellent honey. Generally, however, honey bees are killed by becoming entangled in the pollen masses of some of the species of Asclepias. Many other plants of the order have an acrid poisonous juice. Two of our common species of milkweed (A. tuberosa) and (A. incarnata) and the (Vincetoxicum officinale) contain the bitter glucoside asclepidin which is an amorphous, bitter, yellow emetic substance. The root especially, which acts as an emetic, is recorded as poisonous in Europe and cattle and other domestic animals will not cat it. Friedberger and Fröhner state that it causes diabetes and general weakness. The caustic bush Sarcostemma australe of Australia is regarded in that country as poisonous. The Condurango (Marsdenia Cundurango), a South American vine, is used as an alterative. It is bitter and aerid. The root of Asclepias stellifera of South Africa, according to J. Burtt Davy, yields an excellent rubber.

Asclepias (Tourn.) L. Milkweed

Perennial herbs with milky juice, entire leaves; flowers in umbels; calyx 5-parted, persistent, the lobes spreading; corolla deeply 5-parted, reflexed during flowering, deciduous; the crown consists of 5 hooded processes each containing an incurved horn, enclosing the stamen tube; stamens 5, inserted at the base of the corolla; anthers adherent to the stigma, each with 2 cells and containing a pair of pear shaped masses of pollen (pollinia); ovaries 2; follicles 2; the stigma 5-angled or 5-lobed; numerous seeds with a coma. About 85 species, mostly of the new world. Several members of the order are quite weedy, especially in grain fields and pastures.

Asclepias vestita Hook and Arn.

Densely floccose-woolly, the white wool deciduous in age; leaves from ovate to oblong-lanceolate very acute or acuminate, often subcordate, short petioled or the upper sessile, 4-6 inches long; umbels 1-4, the terminal usually peduncled, the lateral all sessile; corolla greenish-white or purplish, the lobes ovate, 3 inches long, column very short; hoods nearly erect, ventricose, slightly surpassing the anthers, entirely at the back of the somewhat truncate summit, auriculate extended at the inner angle, the auricles or angles involute; the crest not horn-shaped attached up to the summit of the hood, blunt not exserted; an interior crown of 10 tooth-like processes in pairs between the hoods; ovaries glabrous; follicles at first canescent.

Distribution. California and adjacent regions.

Poisonous properties. Said to be poisonous. The juice of this species and of A, eriocarpa are irritating.

Asclepias mexicana Cav.

Stem 3-5 feet high; leaves in whorls of 3-6 or uppermost and lower opposite, sometimes also in axillary fascicles, linear or narrowly lanceolate (3-6 inches long, 2-6 lines broad); umbels corymbose, densely many flowered, on peduneles longer than the pedicels; flowers greenish white, sometimes tinged with purple; corrolla lobes oblong; hoods broadly ovate, entire, shorter than the anthers, exceeded by the stout-subulate incurved horn.

Distribution. In the southwest.

Poisonous properties. Said to be poisonous.

Asclepias speciosa Torr. Showy Milkweed

A perennial from 1-4 feet high, white tomeritose or canescent; leaves thick, broadly ovate or oval, petioled; pedicel glabrate above; flowers greenish purple,





Common Milkweed (Asclepias syriaca). Listed by Lehmann as poisonous. (Bull. 1a. Agr. Exp. Sta. 70).

borne in dense umbels or rarely solitary follicles, erect or spreading on the recurved pedicels.

Distribution. This weed is found in moist soil from Minnesota and northern Iowa to Kansas, in the Rocky Mountain region, common in Colorado, Wyoming and Utah. Troublesome not only in our meadows but occasionally also in our grain fields and gardens.



Fig. 400. Showy Milkweed (Asclepias speciosa) A well known troublesome weed with milky juice. This plant and other species are known to be poisonous; it is common from Western Missouri and Iowa westward. (Ada Hayden.)

Asclepias syriaca L. Milkweed

A perennial herb with a stout stalk from 2-5 feet high, finely soft, pubescent leaves oval-oblong, or ovate, obtuse or roundish at the base, the young leaf somewhat pubescent above, soon becoming glabrate; petioles stout; flowers borne in umbels, from a few to many, peduncles pubescent or tomentose; corolla dull purple or whitish in color; follicle borne on erect or recurved pedicels.

Distribution. This species is widely distributed in the north, occurring in waste places from New Brunswick to the Saskatchewan, along the Atlantic coast to North Carolina and south and west to Missouri and Kansas.

Poisonous properties. Said to be poisonous to live stock. It contains asclepion. Bees often become entangled in the pollen manes of this and preceding species and are unable to extricate themselves. Lehmann and other European writers list this species as poisonous.

TUBIFLORAE

Mostly herbs, rarely shrubs or trees; corolla generally gamopetalous, irregular or regular; stamens adnate to the corolla tube and generally alternate with them; overy 1-celled, occasionally deeply 4-lobed.

Among the important families of this order are the Polemoniaceae, containing the ornamental Phlox Drummondii, the perennial P. maculata, P. divaricata, and P. pilosa, many cultivated varieties of Phlox and of Gilia, of which G. aggregata and Collomia gracilis are examples. According to Greshoff Gilia aggregata contains saponin. It is regarded as a poisonous plant. The order also includes Pedaliaceae, including the sesame (Sesamum indicum) which furnishes the valuable sesame oil, the plant being indigenous to the East Indies; the Orobanchaceae which contains the troublesome broom rapes, (O. ramosa and O. minor); the Gesueriaceae containing the Gloxinias, native to Mexico but cultivated in this country as greenhouse plants; the Leutibulariaceae which includes the bladderwort (Utricularia vulgaris) and other insectivorous plants which are sometimes destructive to fish, and the butterwort (Pinguicula vulgaris) found in northern regions, the leaves of which are used by the Lapps to curdle reindeer's milk and also to thicken fresh warm milk so that it will neither curdle nor form cream afterward but makes a palatable tenacious mass, a small portion of which will act similarly upon another quantity of fresh milk; and the Acanthaceae containing several plants cultivated as ornamentals like the Thunbergia alata and the Ruellias. The Ruellia ciliosa is used as a substitute for Spigelia and is common in the central states to Iowa and Southward. The Barleria Prionitis of Siam is used for snake bites. The Strobilanthes callosus, a shrub found in India, is an irritant poison, according to Major Kirtikar, these irritant properties being due to hairs on the leaves.

FAMILIES OF TUBIFLORAE

| Corolla generally regular. |
|---|
| Ovary not 4 lobed, ovules 2 or more. |
| Style 1, ovary 2-celled; fruit a berry or capsuleSolanaceae. |
| Style 1, entire 2-cleft or 2-parted; frequently twining plants |
| |
| Style 1, 2-lobed or 2-parted; herbs not twiningHydrophyllaceae. |
| Ovary generally 4-lobedBoraginaceae |
| Corolla generally irregular. |
| Ovary generally 4-lobedLabiatae. |
| Ovary not 4-lobed. |
| Placentae axillaryScrophulariaceae. |
| Placentae parietalBignoniaceae. |
| |

CONVOLVULACEAE

Chiefly twining or trailing herbs, shrubs or trees, some with milky juice; alternate leaves without stipules; flowers regular and perfect; calyx inferior, 5-parted or 5-divided; a 5-lobed or plaited corolla, convolute or twisted in the bud; stamens 5, inserted on the tube of the corolla and alternate with its lobes; a 2-celled, rarely 3-celled ovary with a pair of erect ovules in each cell. About 900 species of wide distribution, but chiefly in warm regions. Contains a number of important economic plants, among them the sweet potato (*Ipomoea*

Batatas), which has been widely cultivated in all tropical and sub-tropical countries and is a well known and cultivated plant in the United States. There are many varieties. In the South the large varieties are called yams, but these should not be confused with the Chinese yam (Dioscorea), which forms an important article of food in tropical countries, in the islands of the Pacific and in New Zealand. The man of the earth (Ipomoea fastigiata) was used by the Indians as food. It has properties similar to Jalap, for which it is sometimes substituted. The Ipomoea Jalapa, native to Mexico, produces a large root which is also a purgative and contains a glucoside convolvulin C₈₁H₅₀O₁₈. Several plants of the order, like the moon-flower (Ipomoca Bona-nox), the morning-glory (Ipomoea purpurea) and cypress vine (Ipomoea Quamoclit) are cultivated for ornamental purposes. Several, like Jalap (Ipomoca Purga) and (I. congesta) are used in medicine and are strong purgatives. The scanmony (Convolvulus Scammonia) of Western Asia is also used as a purgative, the milky juice being collected when hard, it contains scammonin C34 H56 O18. Convolvulus scoparius and C. floridus, furnishing the oil of rhodium, are small shrubby species of the Canary Islands. The wood is strongly scented and the oil is used to adulterate attar of roses.

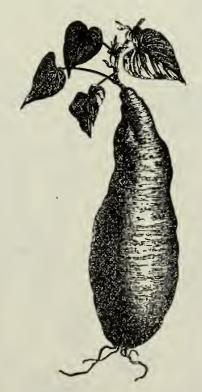


Fig. 401. Sweet Potato (Ipomoea Bata(as). A well known cultivated plant in tropical and sub-tropical countries. (From Vesque's Traité de Botanique.)

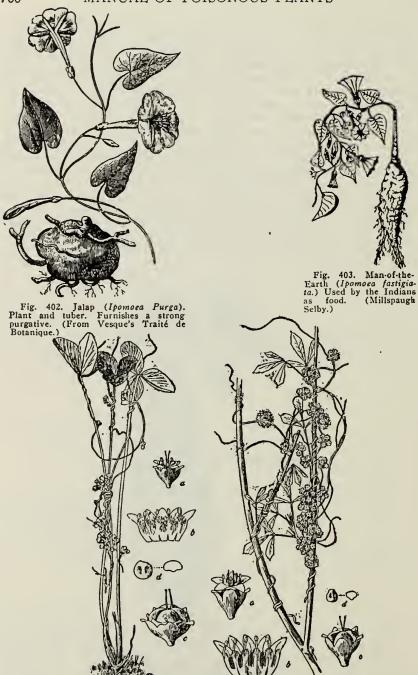


Fig. 404. Dodder. To the left—Field dodder (Cuscuta arvensis), a, flower; b, flower spread apart; c, capsule with stamens and styles; d, seed. To the right—Alfalfa dodder (C. epithymum), a, flower; b, flower spread apart to show stamens and corolla; c, capsule showing styles; d, seed. (Dewey, U. S. Dept. Agrl.).

The dodders (Cuscuta) belong to the family Cuscutaceae, and are of interest in this connection mainly because they are parasitic on clover, alfalfa and other plants. Those growing on the above named plants are Cuscuta arvensis and C. Epithymum; the flax dodder (C. Epithymum) occurs on flax. The plants contain cuscutin. From two independent sources in this country there have come reports that clover dodder is injurious. Dr. J. L. Taylor of Lisbon, Ohio, wrote Prof. A. D. Selby that when dodder was fed to horses it caused bowel trouble. Dr. E. H. Jenkins of Connecticut states that cattle were seriously troubled with scours when clover hay in which this plant occurred was a part of the ration. He could not determine whether it was the dodder or the mould which had developed because the dodder had so matted with the clover the hay was not cured well. When this hay was cut out from the ration the trouble ceased.

Ipomoea L. Morning Glory

Twining or erect herbs; flowers large, showy, axillary; calyx 5-parted; corolla broadly bell-shaped, 5-cleft; stamens included; pistil with 2-4 celled ovary, 4-6 ovules; style undivided; stigmas capitate, 1, 2 or 3; fruit a globular capsule, 4-6 seeded. About 350 species, tropical, or of warm regions.

The *Ipomoea leptophylla* of the plains of Nebraska to Kansas and the Rocky Mountains produces a large root weighing from 10-100 pounds; it has an crect non-twining stem from 2-4 feet high; narrow and long leaves; large pink purple flowers.

Ipomoea fastigiata Sweet. Man of the Earth. Wild Potato

A deep rooted, smooth, trailing or twining perennial with a stout, large root occasionally weighing 30 pounds; leaves heart-shaped, acuminate or occasionally fiddle-shaped; peduncles 1-5-flowered; sepals smooth, ovate, oblong, very obtuse; corolla funnel-shaped, 3 inches long, tube purplish; pistil with a 2-celled ovary; stigma 2-lobed; each cell 2-seeded.

Distribution. Common in fields, dry or alluvial grounds from New England to Florida to Ohio, Michigan, Illinois, to Texas.

Poisonous properties. The large root of this plant is said to be poisonous; it contains the glucoside ipomoein and is purgative. The same is true of I. leptophylla. The I. Purga is regarded as somewhat poisonous.

Convolvulus (Tourn.) L. Morning Glory or Bindweed

Herbs or somewhat shrubby plants; twining, erect or prostrate; leaves generally cordate or sagittate and petioled; flowers large and axillary; calyx bractless or with a pair of bracts; sepals nearly equal; corolla funnel-form or campanulate; stamens included, inserted on the tube; style undivided or 2-cleft at the apex; stigmas 2, filiform, oblong or ovoid; ovary 1 or 2 celled with 4 ovules; fruit a capsule, 1-4 celled, 2-4 valved. About 175 species of wide distribution.

Convolvulus sepium L. Hedge Bindweed

Smooth, occasionally pubescent, twining around supports or trailing; leaves triangular, halberd or arrow-shaped, the tip acute or pointed, the basal lobes obliquely truncate or sinuate lobed; the flowering peduncles 4-angled, with 2 leaf-like bracts which are commonly acute; corolla is white or tinged with rose purple.

Distribution. The weed is common in the Mississippi Valley in the moist alluvial bottoms along streams and in fields. It is also found extensively creeping over weeds and other herbaceous vegetation and roadsides throughout the state. The species is native to North America and is common from Nova Scotia to Maine, south to North Carolina and Texas to Kansas, Utah, Montana, Minnesota and eastward. It also occurs in Europe and Asia,



Fig. 405. Bindweed (Convolvulus sepium.) Supposed to be poisonous to swine. (After Vasey.)

. Poisonous properties. The plant has a somewhat disagreeable odor. Dr. Schaffner states that it is supposedly poisonous to swine. It is more than likely that some of the same substances are found in hedge bindweed that occur in Jalap. Jalap contains the glucosides convolvulin $C_{31}H_{50}O_{16}$, jalapin $C_{34}H_{50}O_{16}$, turpethin $C_{34}H_{50}O_{16}$, tampicin $C_{34}H_{54}O_{14}$. The rootstock is rich in starch.

Convolvulus arvensis L. European Bindweed

A deep-rooting perennial; procumbent stem, twining or creeping; propagates freely by underground rootstocks; the leaves from 1 to 2 inches long, ovate, oblong, arrow-shaped, the lobes at the base running to a point; the flowers are borne in 1-flowered peduncles with very small leaf-like bracts some distance from the flowers; flowers an inch or less long, short; broadly funnel-shaped, white, or commonly of a rose tinge.

Distribution. This weed has been known for a considerable length of time in eastern North America, where it has been sparingly naturalized for some time. Its distribution may be given as Nova Scotia to Ontario, New Jersey, Nebraska and Kansas.

Poisonous properties. Probably the same as the preceding. European authorities list this and the common morning glory, especially the latter, as somewhat poisonous because of their purgative properties.

HYDROPHYLLACEAE. Water-leaf Family

Herbs, generally hairy; with alternate leaves; perfect, regular 5-parted flowers; calyx inferior, deeply cleft or divided; corolla gamopetalous; stamens 5, inserted on the corolla; ovary superior, 2-celled, with 2 parietal placenta; styles 2-cleft or partially united; fruit a capsule, seeds generally reticulated or pitted.

A small family of 17 genera and about 160 species, chiefly in western North America. Very few plants of the family are ornamental. Some of the western species are occasionally cultivated. Of these the *Phacelia* is the most important. A few of the plants are weedy. The Yerba Santa (*Eriodictyon crassifolium*) is an evergreen shrub with funnel-shaped, white or purple flowers in cymose clusters; it contains a yellow aerid resin, a crystallin principle, *eriodictyonic acid* and *eriocolin*.

Phacelia Juss.

Mostly hirsute, hispid, or scabrous herbs; leaves alternate or the lower opposite; flowers blue or purple, violet, or white; inflorescence hispid; calyx naked at the cymes; deeply 5-parted; stamens attached near the base of the corolla; ovary 1-celled; capsule 1-celled or falsely 2-celled; seeds reticulated. About 80 species, natives of the New World.



Fig. 406. Hairy Phacelia (Phacelia sericea). The Phacelia is common in the Rocky Mountains. The bristly hairs produce mechanical injuries. (Charlotte M. King.)

Phacelia sericea A Gray

Herbs 6 inches to a foot high from a branching caudex, silky-pubescent or canescent, or the simple virgate stems and inflorescence villous-hirsute, rather leafy to the top; leaves pinnately parted into linear or narrow-oblong numerous and often again few-cleft or pinnatifid divisions, silky canescent or sometimes greenish; the lower petioled; the uppermost simpler and nearly sessile; short spikes crowded in a naked spike-like thyrsus; corolla violet-blue or whitish; stamens long exserted; capsule a little longer than the calyx.

Distribution. Common in the mountains from Colorado and northward in rather dry soil.

Phacelia Menziesii Torr.

Herbs 6 inches to a foot high, at length paniculate-branched, hispid or rough-ish-hirsute; leaves mostly sessile, linear or lanceolate and entire, or some of them deeply cleft; the lobes few or single, linear or lanceolate, entire; spikes or spike-like racemes thyrsoid-paniculate, at length elongated and erect; corolla bright violet or sometimes white; stamens about the length of the corolla; capsule shorter than the calyx.

Distribution. Common in the Rocky Mountains from Montana to Utah and westward.

Poisonous properties. The stiff bristles upon these plants certainly produce mechanical injuries. A form of dermatitis venenata occurs after handling the plants. The writer has had abundant experience in contact with these plants in the Rocky Mountains.

BORAGINACEAE. Borage Family

Herbs, shrubs or trees, with alternate entire, rough or frequently scabrous or setose leaves; flowers perfect, usually regular, generally blue, borne in one-sided spikes; racemes, cymes or scattered; calyx inferior, mostly 5-lobed or 5-cleft; corolla short, bell or wheel-shaped 5-cleft or 5-parted; stamens as many as the lobes of the corolla and alternate with them; ovary superior, deeply 2-lobed; style entire; capsule globular; fruit forming 4 seed-like, 1-seeded nutlets, or into two 2-seeded or four 1-seeded nutlets.

A large family of wide distribution, consisting of about 80 genera and 1500 species. Some of the members are ornamental and are frequently cultivated, like the common heliotrope (Heliotropium peruvianum), native to Peru, used for bedding and in greenhouses. Borago or borage (Borago officinalis), used in old gardens for ornamental purposes is an excellent honey plant. Lungwort (Mertensia virginica), an early spring blooming plant of the North, with handsome blue flowers, is occasionally cultivated. The Rocky Mountain M. sibirica is an equally handsome species. The forget-me-not (Myosotis scorpioides), with small but pretty blue flowers, native of Europe, is occasionally cultivated. Alkanet (Alkana tinctoria-Anchusa tinctoria), a native of southern Europe, yields a red dye used for coloring oils and wax. Other plants of this order yield a similar product and one species is known to color the wool of sheep. The roots of the common puccoon (Lithospermum), also yield a dye. The comfrey (Symphytum officinale) is used as a forage plant in Europe. but in the United States is seldom cultivated. The wood of Cordia alba is used by military authorities in San Juan in the manufacture of gun carriages, carpenters' benches, vises, etc.

Genera of Boraginaceae

| Ovary not lobed | Heliotropium |
|-----------------------|--------------|
| Ovary deeply 4-lobed. | • |
| Flowers regular | |

Nutlets horizontally radiate, covered with prickles......Cynoglossum Nutlets erect, armed with prickles......Lappula Corolla irregular.

Stamens exserted Echium

Heliotropium (Tourn.) L. Heliotrope

Herbs or shrubs with entire alternate leaves; the small blue or white flowers, scattered or borne in scorpoid spikes; corolla salver or funnelform without appendages, more or less plaited in the bud; stamens not exserted, with nearly sessile anthers; stigmas conical or capitate; fruit 2-4-lobed, separating into 4, 1-seeded nutlets or into 2, 2-seeded carpels.

About 115 species, mostly tropical. The best known representative in the North is the cultivated H. peruvianum. The seaside heliotrope (H. curassavicum) occurs in saline soil along the Atlantic coast, to Maine, and from Mexico to Illinois and westward.

Heliotropium indicum L. Indian Heliotrope

An erect, hairy annual with hirsute or hispid stem; leaves petioled, ovate or oval and somewhat heart-shaped; flowers blue, borne in spikes; fruit deeply 2-lobed, smooth, 2 seeds in each cell.

Distribution. Common in waste places from North Carolina to Southern Indiana and Missouri. Naturalized from India.

Heliotropium europaeum L. European Heliotrope

An erect annual, branched, roughish pubescent herbs; leaves oval, long petioled, narrow at the base; flowers borne in 1-sided bractless spikes, the terminal spikes in pairs; calyx spreading, the segments shorter than the corolla tube; anthers distinct, obtuse.

Distribution. Common in waste places southward. New York to Pennsylvania and Florida.

Poisonous properties. The European heliotrope contains heliotropin, a bitter, volatile, alkaline, poisonous alkaloid.

Cynoglossum (Tourn.) L. Hound's Tongue

Hirsute or hispid, rarely smooth herbs; alternate or entire leaves; purple, blue or white flowers in panicled racemes; calyx 5-cleft or 5-parted; corolla funnelform or salverform, the throat clothed with 5 obtuse scales; ovary deeply 4-lobed fixed near the apex to the base of the style, roughened with short barbed or hooked prickles. About 75 species, of wide distribution.

Cynoglossum officinale L. Hound's Tongue

A coarse biennial herb, clothed with short, soft hairs; lower leaves oblong or oblong lanceolate, the upper closely sessile with a slightly heart-shaped base; racemes nearly bractless, elongated in fruit; divisions of the calvx oyate. lanceolate, acute; corolla reddish-purple, rarely white; nutlets flat on the broad upper face, splitting away at maturity.

Distribution. In fields and waste places, especially eastward from New England to Quebec, Ontario, Minnesota, Manitoba, and Kansas.

Cynoglossum virginianum L. Wild Comfrey

Perennial hirsute herb with simple stem, 2-3 feet high, leafless above; stem leaves lanceolate oblong, clasping by a heart-shaped base; flowers long-peduncled, pale blue, small; fruit broad, nutlets not margined, convex on the upper surface.

Distribution. Common in woods of the central Mississippi Valley states from New Brunswick to Ontario, Florida, Louisiana to Texas.

Poisonous properties. The common hound's tongue is suspected of being poisonous. The European species, C. officinale, contains a powerful alkaloid, cynoglossin, which resembles curare in its action. It also has the principle consolidin.



Fig. 407. Hounds-tongue (Cynoglossum officinale). Suspected of being poisonous. (From Darlington's Weeds and Useful Plants.)

Lappula (Rivinus) Moench. Stickseed

Roughish pubescent or hairy herbs with alternate narrow or entire leaves, small blue or white flowers in racemes or spikes; calyx deeply 5-cleft or 5-parted with narrow segments; corolla salverform or funnelform.

About 40 species in north temperate regions, several of which are weedy in North America.

Lappula virginiana (L.) Greene. Beggar's Lice

A coarse pubescent biennial from 2-4 feet high; lower leaves ovate, orbicular cordate, long, petioled; stem leaves ovate-oblong or oval; flowers nearly white; globose nutlets, flattened and barbed.

Distribution. Common especially in woods northward from New Brunswick to Wisconsin, Minnesota, Kansas to Louisiana.

Lappula echinata Gilbert. Stickseed

An erect annual from 1-2 feet high; pale, leafy, hispid with erect branches; leaves linear or linear-oblong; racemes 1-sided, bracteolate; calyx segments lanceolate; corolla blue; nutlets rough-granulate or tuberculate on the back, the margins with a double row of slender prickles.

Distribution. Abundant in waste places along roadsides from eastern Canada and New England to Minnesota, Kansas and British Columbia. Weedy also in Europe, where it is native.

Lappula floribunda (Lehm.) Greene. Large-flowered Stickweed

An erect perennial or biennial, rough-pubescent; 2 feet or more high; leaves oblong to linear-lanceolate, the lower tapering on margined petioles; racemes erect or nearly so; pedicels deflexed in fruit; flowers pretty; blue, occasionally white; nutlets scabrous on the margin with a row of flat prickles.

Distribution. Common in Saskatchewan and Minnesota and abundant in the Rocky Mountains.

Poisonous properties. perhaps the fruits of all the species are somewhat injurious to animals, causing inflammation. They are frequently found in wool.

Echium L. Blueweed

Bristly annual, biennial, or perennial; erect stems; corolla with a funnelform tube and a 5-lobed spreading border; stamens many, exserted. The 30 species are natives of the old world.

Echium vulgare (Tourn.) L. Blueweed

An erect, rough, bristly biennial, 18 inches to 2 feet high, with an erect, mostly simple stem; leaves of the stem linear-lanceolate, sessile; flowers in cymose clusters; corolla reddish-purple, changing to blue; tube funnelform, border unequal, spreading, 5-lobed; stamens 5, inserted on the tube; style threadlike; nutlets roughened or wrinkled.

Distribution. A common weed along roadsides, fields, and meadows from New England to Indiana.

Poisonous properties. Probably poisonous. According to Friedberger and Fröhner it causes slavering.

VERBENACEAE. Verbena Family

Herbs, shrubs or trees, with opposite or alternate leaves; perfect, regular, or somewhat irregular flowers borne in spikes, racemes, eymes or panieles; calyx generally persistent, inferior, 4-5-lobed or cleft; corolla 4-5-cleft, tube cylindrical; stamens 4, didynamous or 2; ovary superior, 2-4-celled or more; fruit dry or drupaceous, splitting into 1-4 nutlets.



Fig. 408. Blueweed (Echium vulgare). Probably poisonous. (Vasey-Selby).

A large family which is found chiefly in tropical regions, there being about 1200 species. The wood of some of the tropical members of the family is valuable. Among these is the Indian teak tree, (Tectona grandis), native to India. The wood is hard, heavy and very durable and is used in ship-building and for railway cars. The Vitex littoralis, a large tree, native of New Zealand, produces a hard and heavy wood. The fragrant leaves of V. Negundo are used to stuff pillows.

This family includes many ornamental plants, the most commonly cultivated in green houses being the Clerodendron Thompsonae, which has a showy crimson corolla and white calyx. The lemon verbena, (Lippia citriodora), a stiff, branching shrub from Chili, is commonly cultivated in the south on account of its very fragrant leaves; an oil, from it, contains citral. The fog-fruit, (L. nodiflora), is valued in Egypt as a lawn plant, proving a success where all grasses or other plants tried for that purpose have failed, the lawns lasting five or six years without renewal. The Lippia mexicana, used in medicine, contains lippiol. The extensively cultivated verbena of our flower gardens is the V. Aubletia, which is produced in many colors, the species being native from southern Indiana to Texas. Lantanas are also cultivated. The French mulberry, (Callicarpa americana), is said to be poisonous. The vervains (Verbena stricta and V. hastata) with blue flowers, are common pasture weeds. The white vervain (V. urticaefolia) contains a bitter glucoside.

LABIATAE Mint Family

Chiefly aromatic herbs, some shrubs and trees; with square stems; opposite leaves without stipules; flowers with cymose inflorescence, perfect, irregular, more or less 2-lipped; calyx 5-toothed or 5-lobed; corolla 4-5-lobed, commonly 2-lipped, upper 2-lobed or entire, the lower usually 3-lobed, stamens borne on the corolla tube; ovary superior, deeply 4-lobed or 4-parted, in fruit forming 4 small seed-like nutlets or achenes surrounded by the persistent calyx; usually exalbuminous or with some albumin. A large family of about 3000 species

of very wide distribution. Many of these plants are used in medicine. Lavender (Lavandula vera) of the mountain regions of northern and eastern Europe, is cultivated for the oil of lavender, much used in perfume and contains linalool and geraniol. Spearmint (Mentha spicata) a fragrant perennial plant of Europe is used by confectioners and in the manufacture of perfumed soap. The volatile oil contains carvol C10H14O. Peppermint (Mentha piperita) a native to Europe and naturalized in North America, is cultivated in New York and Michigan for the manufacture of peppermint oil. It contains a volatile oil and menthol C,0H20O, and is used for flavoring mutton and sweetmeats and as a cordial. It is a stimulant. The Japanese peppermint is obtained from M. arvensis var. piperascens. Pennyroyal (Mentha Pulegium), native to Europe, is used for the same purposes for which peppermint is used. Garden thyme (Thymus vulgaris) is a small erect woody shrub of southern Europe; it is fragrant, has a pungent taste and contains thymol C,0H,8ON, which is used as an antiseptic. It also contains cymene, borneol, and linalool. Rosemary (Rosmarinus officinalis), an evergreen shrub of Europe, is chiefly used as a perfume. This and Lavandula Spica both contain borneol C,0H,8O, one of the pinene group of terpenes and camphene. Germander (Teucrium canadense) is a stimulant and has aromatic properties. Horse mint (Monarda fistulosa) is a stimulant and is used to remove colic pain. Oswego tea (Monarda didyma) is used as a substitute for tea, and catnip is used for the same purpose. M. punctata contains thymol, carvacrol, etc. Horehound (Marrubium vulgare), a perennial weed native to Europe, is a stimulant and tonic and is also slightly laxative. It contains the bitter principle marrubin. Sweet basil (Ocimum Basilicum) of India is a strong aromatic herb used for culinary purposes and in the manufacture of Chartreuse liquors while the mucilaginous



Fig. 409. Horse Mint (Monarda fistulosa). A common roadside weed with pungent properties. (Charlotte M. King).

"seeds" are used for genito-urinary troubles. Savory (Satureia hortensis) an annual herb of Europe, sparingly run wild in western United States, is used as a pot herb. The patchouli (Pogostemon Heyneanus) is used by the Arabs to scent mattresses and shawls. Sage (Salvia officinalis) is cultivated as a pot herb and contains pinene, cineol, thujone and borneol. Marjoram (Origanum majorana), a perennial plant native to South of Europe, and cultivated as an aromatic herb, is also an excellent honey plant containing citral. Many species of mints, like the brilliantly colored South American sage (Salvia coccinea, S. splendens, etc.) and others native to the southern states and the west, are cultivated for ornamental purposes. The lance leaved sage (S. lanceolata) is a troublesome weed in the West. S. officinalis of Europe, a stimulant and tonic, contains cineol and salviol C₁₀H₁₈O, and is commonly used with meat and sausage in German communities. The Japanese potato (Stachys Sieboldii), is used as the Jerusalem artichoke is. Sweet balm (Melissa officinalis), a pot herb, contains a bitter principle.

Cymol, C₁₀H₁₄O, one of the benzol derivatives, occurs in many of the Labiates, especially in Thymus officinalis, Monarda punctata, and Micromeria punctata. Thymol, a benzol derivative, is also found in Origanum floribundum and Monarda citriodora.

Genera of Labiatae

| Calyx rigid, spiny, pointed |
|--|
| Calyx not spiny toothed. |
| Anthers approximate. |
| Fertile stamens 2 Hedeoma |
| Fertile stamens 4 |
| Calyx tubular curved |
| Calyx bell-shaped, |
| Aromatic herbs, stamens exserted Mentha |
| Not aromatic herbs ascending under the upper lip4 Lamium |
| 1. Nepeta I. Catnip |

Herbs with dentate or incised leaves; flowers white or blue, in clusters; calyx tubular; corolla 2-lipped, lower lip spreading, 3-lobed. About 150 species native to Europe and Asia.

Nepeta Cataria L. Catnip

A perennial erect herb, 1-3 feet high; leaves ovate, cordate, coarsely serrate, petiolate, whitish, downy underneath; flowers in cymose clusters; corolla whitish, dotted with purple.

Distribution. Native to Europe; widely naturalized in northern states.

Nepeta hederacea (L.). Trevisan. Ground Ivy

A creeping, trailing perennial, with leaves all alike; petioled, round, kidney-shaped, crenate, smooth green on both sides; flowers light blue in axillary whorls of about 6, appearing in early spring and summer.

Distribution. Native to Europe, widely naturalized in the northern states, especially in shady places.

Poisonous properties. According to Dr. Schaffner, Ground Ivy is said to be poisonous to horses. Contains a volatile oil and a bitter principle. The common catnip also contains a volatile oil and bitter principle.

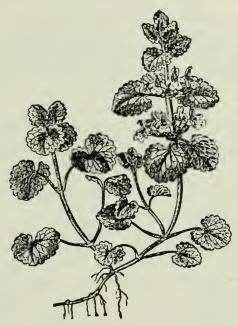


Fig. 410. Ground Ivy (Nepcta hederacea). A somewhat weedy plant, naturalized from Europe. Said to be poisonous to horses. (From Johnson's Med. Bot. of N. A.).

2. Hedeoma Pers.

Aromatic, pungent herbs; leaves small; flowers in axillary clusters, crowded into terminal spikes or racemes; calyx ovoid or tubular, bearded in the throat, 2-lipped; the upper lip toothed; corolla 2-lipped, the upper 2-lobed, the lower spreading, 3-cleft; fertile stamens, 2; the upper pair reduced to sterile filaments or wanting; nutlets ovoid, smooth.

Hedeoma pulegioides Pers. American Pennyroyal

An erect, branching, hairy annual; leaves ovate to ovate-oblong, petioled, sparingly serrate; whorls few flowered; upper calyx teeth triangular, gibbous; corolla bluish; rudimentary stamens, evident but not usually anther-bearing.

Poisonous properties. It has been regarded with suspicion. It has the odor and taste of true Pennyroyal. Hedeoma pulegioides contains a volatile oil, hedeomol $C_{10}H_{18}O$.

Leonurus L. Motherwort

Tall herbs with palmately cleft or dentate leaves; flowers small, white or blue, in axillary clusters; calyx tubular, 5-nerved and 5 rigid teeth; corolla 2-lipped. About 10 species in the old world.

Leonurus Cardiaca L. Motherwort

Tall perennial herb with erect stem, 2-6 feet high; leaves long petioled, the

lower round and palmately lobed, the upper crenate at the base, 3-cleft; flowers pale purple in close axillary whorls; corolla bearded.

Distribution. Native to Europe, widely naturalized in the northern states. Injurious properties. The stiff bristles of the calyx are often injurious, producing mechanical injuries. It contains a volatile oil with an unpleasant odor, a bitter principle, etc.

Lamium L. Dead Nettle

Decumbent herbs; leaves usually cordate, doubly toothed; flowers small, axillary and terminal clusters; calyx tubular; 5-toothed, the teeth nearly equal, the upper ones larger; corolla dilated at the throat, upper lip ovate or oblong arched, narrowed at the base, the middle lower lip spreading, the lobe notched at the apex; stamens 4, small; the anthers nearly in pairs, nutlets truncate. About 40 species in the old world; some troublesome weeds like L amplexicaule.

Lamium amplexicaule L. Dead Nettle

An annual or biennial herb with rounded, deeply toothed, crenate leaves; the upper leaves small, clasping; calyx tubular, 5-toothed; flowers small; corolla purple, upper lip bearded, the lower spotted. The *L. album* is perennial, has larger flowers, and has slender calyx teeth.

Distribution. Common in the eastern states to Missouri. The L. album escaped and not infrequent westward.

Injurious properties. The dead nettle is regarded as injurious.



Fig. 411. Dead Nettle (Lamium, album), frequently escaped from cultivation. (Ada Hayden).

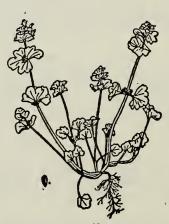


Fig. 412. Dead Nettle (Lamium amplexicaule). Regarded as poisonous. (Selby).

Mentha (Tourn.) L. Mint

Herbs with the odor of mint. Leaves usually with punctate spots; flowers small in whorled clusters, pink or white; calyx bell-shaped or tubular, 5-toothed; corolla tube shorter than the calyx; limb 4-cleft; stamens equal, erect, included or exserted; filaments smooth; nutlets ovoid, smooth. About 30 species of temperate regions. Our native species (M. arvensis, var. canadensis) is common in low marshy ground.

Mentha crispa contains linacol, $C_{10}H_{18}O$, one of the terpenes. It may be mentioned in passing that this same substance recurs in Ocimum Thymus and Darwinia. A ketonc-carbon, $C_{10}H_{14}O$, is found in several species of the genus.

Mentha piperita L. Peppermint

Smooth, erect, perennial herb with creeping rootstocks from 1-2 feet high; leaves petioled, ovate, oblong to oblong-lanceolate, acute and sharply serrate; flowers whorled in interrupted loose, leafless spikes; purplish or whitish.

Distribution. Commonly escaped from cultivation and troublesome in the East.



Fig. 413. Peppermint (Mentha piperita). One of the sources of the peppermint of commerce. (From Vesque's Traité de Botanique).



Fig. 414. Tomato (Lycopersicum esculentum). An important food plant. (W. S. Dudgeon).

SOLANACEAE. Nightshade Family

Herbs, rarely shrubs, vines; a few of the tropical species, trees with alternate leaves without stipules; flowers regular or nearly so, borne in cymes; calyx inferior, 5-lobed; corolla gamopetalous, generally 5-lobed; stamens as many as the lobes of the corolla and alternate with them, inserted on the tube, generally equal; style and stigma 1; ovules numerous; fruit a berry or capsule. A large family, chiefly tropical, consisting of 70 genera and 1600 species. Several of these are important medicinal plants and several important food plants. Many plants of the order have poisonous properties.

The red or Cayenne pepper (Capsicum annuum), native to North America, is much used in tropical countries to stimulate the appetite. It seems to run to many forms, but has been shown by Prof. Irish that many of the so-called species belong to C. annuum. A shrubby species, C. frutescens, is native to western Texas and Mexico. The tobacco (Nicotiona Tabacum), was undoubtedly native to America. It was used by the Indians in North America at the time of the discovery by Columbus, and was introduced into England in 1585 by Lane who was a deputy of Sir Walter Raleigh. Tobacco is now cultivated in many civilized countries, as Cuba, Philippine Islands, Sumatra, also in Florida, the Carolinas, Connecticut, Kentucky, and Wisconsin in the United States. It is used for making cigars, snuff and for chewing. It is an important article in commerce. The potato (Solanum tuberosum) is indigenous to Peru and Chili, but was introduced into Spain about the beginning of the 16th century, and into England from Virginia in 1586 by Sir Walter Raleigh. The greatest yields occur in irrigated districts. It is one of the most important food

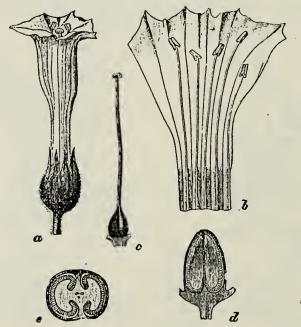


Fig. 414a. Tobacco Plant (Nicotiana Tabacum). a, Flower; b, corolla; cut open; c, ovary; d, e, young fruit. (a, b, c, natural size; d. e. x 2.). (After Strasburger, Noll, Schenck, and Schimper).

plants. Other species of tuberous Solanum occur, as the S. Jamesii, in south-western United States, and several others in Mexico and South America. The egg-plant (Solanum Melongena) is native to India, but is now widely cultivated in tropical countries and temperate regions, the fruit being used for culinary purposes. The tomato (Lycopersicum esculentum), a native of tropical America, of which there are many varieties, is now widely distributed in tropical and temperate regions. The fruit is eaten fresh or canned, or made into

various products. The currant tomato (L. pimpinellifolium), a fruit about the size of a currant, is cultivated as a curiosity. Ground cherries are much cultivated. The strawberry tomato (Physalis Alkekengi), a perennial native to southern Europe, has a large fruiting calyx which turns red. The fruit is caten, but it is not so palatable as the fruit of the Cape gooseberry (P. peruviana), which is native to Peru. The fruit of a native species (P. pubescens) is also caten, but this also is inferior to the Cape gooseberry.

Belladonna is derived from Atropa Belladonna, a tall glabrous or slightly downy herb, with a perennial rootstock, native to southern Europe and eastward to Asia Minor. This furnishes the atropin of commerce used for dilating the pupil of the eye. The earliest investigations of the alkaloids of belladonna were made by Bauberlein in 1809, who first determined their presence. Esse was the first to find atropamin $C_{17}H_{21}NO_2$, in the roots of the plant, although Schmidt denies the presence of atropin, affirming the presence of hyoscyamin only. Belladonnia, an isomer of the above, is probably also present. The root of Atropa Belladonna sometimes contains from 4/10-1% of the alkaloid and the leaves about half as much. The greatest amount of the alkaloid occurs during the flowering period. Pseudo-hyoscyamin occurs in the roots of Mandragora. The alkaloids mandragorin, $C_{11}H_{19}NO_2$, and manacin $C_{22}H_{33}N_2O_{10}$, occur in Brunfelsia Hopeana. Several other undetermined alkaloids, such as jurubebin, have also been found in the family.

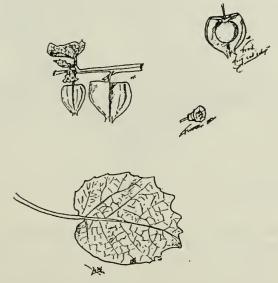


Fig. 415. Ground Cherry (Physalis viscosa). Cultivated for its fruit. (W. S. Dudgeon).

The mandrake (Mandragora officinarum) of the ancients was, at one time, supposed to have medicinal virtues. The flowering tobacco (Nicotiana alata) is a well known cultivated plant of Brazil. The thorn apple (Datura Metel) is much cultivated, as is the D. meteloides, which has large, sweet-scented flowers. The bittersweet (Solanum Dulcamara), the hardy annual (Schizanthus pinnatus), the Brunfelsia latifolia, which has fragrant flowers, and the Cestrum



Fig. 416. Sleeping or Deadly Night-shade (Atropa Belladonna). Tip of flowering and fruiting branch; entire fruit; cross section of fruit; corolla cut open and spread out. Source of the belladonna of commerce. (From Vesque's Traité de Botanique).

elegans are frequently cultivated. The odor from the flowers of the latter is very overpowering. The berries of C. pallidum are said to be poisonous, but birds have scattered the plant very widely in the tropics. Petunia violacea, Lycium halimifolium and L. clinense are cultivated. The Duboisia myoporoides of Australia is a tall shrub, its leaves having narcotic properties and containing the substance duboisin, a mixture of hyoscyamin and atropin producing an action like that of hyoscyamus but more hypnotic. According to Maiden this plant is poisonous to stock. Other species like D. Leichardtii contain the same substance. The piturie (D. Hopwoodii) contains a liquid volatile alkaloid piturin C_0H_8N resembling nicotin. The natives mix the piturie leaves with the ashes of some other plant and chew them like tobacco. In its action it resembles nicotin.

The scopola (Scopolia carniolica) of Austria and Hungary, is a perennial herb used like Belladonna in medicine. The leaves and rhizomes of this species and S. japonica are poisonous. The S. carniolica plant contains atropin $C_{17}H_{21}NO_4$, hyoscyamin and scopalamin. The latter substance is broken up into scopolin $C_8H_{18}NO_2$ and tropic acid $C_9H_{10}O_3$. The hyoscin $C_{17}H_{23}NO_3$ is impure scopolamin. Scopalin causes dilation of the pupils; the heart action is at first diminished, then increased, due to the stimulation of the imhibitory nervous apparatus. The pichi used in kidney troubles is the dried leafy twigs of the Chilian shrub (Fabiana imbricata). The tree tomato (Cyphomandra betacea) produces a fruit similar, in taste, to that of the common tomato, if eaten when raw; but after it is stewed, provided the skin and seeds have first been removed, an apricot-like flavor is produced. It is much used in tarts and pastry in the mountainous districts of the tropics.



Fig 417. Corkwood (Duboisia myoporoides). A shrub, bearing leaves that possess narcotic qualities. (After Faguet).

Genera of Solanaceae

| Fruit a berry. |
|---|
| Corolla wheel shaped. |
| Anthers opening by uplifted valves Solanum |
| Anthers opening longitudinally, widely spreading 2 Capsicum |
| Corolla not wheel shaped |
| Corolla funnel form |
| Fruit a capsule. |
| Calyx urn shaped somewhat irregular4 Hyoscyamus |
| Calyx prismatic, corolla funnel form |
| Calyx tubular Nicotiana |

1. Solanum (Tourn.) L. Nightshade

Herbs or shrubs, often with stellate pubescence; flowers in cymose, umbellate, or racemose clusters; calyx bell-shaped or rotate generally 5-parted or 5-cleft, corolla rotate 5-lobed or cleft, plaited in the bud; stamens exserted, filaments short inserted on the throat, anthers converging around the style opening at the top; ovary usually 2-celled, stigma small; berry with persistent calyx at the base or enclosing it; seeds numerous. About 900 species of wide distribution. Several are troublesome weeds as horse nettle (Solanum carolinense) and buffalo bur (S. rostratum). The potato (S. tuberosum) and eggplant (S. Melongena), are cultivated.



Fig. 418. Common potato (Solanum tuberosum). The potato under some conditions is very poisonous, especially when the tops are green. (Lois Pammel).

Solanum Dulcamara L. Bittersweet

A more or less pubescent perennial, stem climbing or straggling, somewhat woody below; leaves petioled ovate or hastate, the upper usually halberd shaped: flowers purple or blue in cymes; berry globose, red.

Distribution. Naturalized from Europe, New Brunswick to New Jersey, Pennsylvania, Kansas to Minnesota.

Poisonous properties. The plant contains the alkaloid solanin $C_{28}H_{48}NO_{11}+H_2O$, with a hot, bitter taste, dulcamarin a bitter principle $C_{22}H_{34}O_{10}+20H_2O$ and the alkaloids solanidin $C_{40}H_{61}NO$ and solanein $C_{52}H_{33}NO_{13}$.

Chesnut says with reference to this plant:

Besides solanin, (0.3 percent), this plant contains another less poisonous compound, dulcamarin, which gives it its peculiar bitter-sweet taste. Neither of the compounds is abundant. The berry, though its taste is not remarkably disagreeable, is somewhat poisonous, and it has been shown that an extract of the leaves is moderately so. The plant has nevertheless caused some ill effect. The treatment is the same used in case of the above species.

According to Schimpfky the berries of this plant have been used to poison dogs and the juice of the fruit acts as a poison to rabbits. Flückiger and Hanbury in their Pharmacographia, make this statement with reference to Dulcamara:

Dulcamara is occasionally given in the form of decoction, in rheumatic or cutaneous affections; but its real action, according to Garrod, is unknown. This physician remarks that it does not dilate the pupil or produce dryness of the throat like belladonna, henbane or stramonium. He has given to a patient 3 pints of the decoction per diem without any marked action, and has also administered as much as half a pound of the fresh berries with no ill effect.

Johnson in his Medical Botany of North America refers to the use of the plant as follows:

Bittersweet, in full doses, produces a certain mount of cerebral disturbance of a narcotic character, together with dryness of the throat, and sometimes an erythematous eruption of the skin, with a tendency to diaphoresis. It has been employed with benefit in a variety of cutaneous eruptions, in muscular rheumatism, and in chronic bronchial and pulmonary affections.

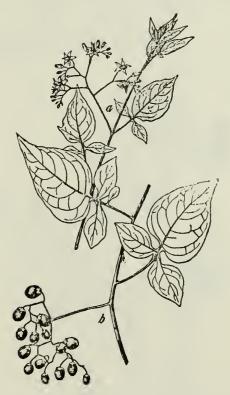


Fig. 419. Bittersweet (Solanum Dulcamara), a, flowering spray; b, fruit—both onethird natural size. Berries somewhat poisonous. (U. S. Dept. Agr.).

Lehmann states that it is a narcotic poison when given in large doses, even causing death in rabbits.

It will be seen from the above quotations that the plant is not a violent poison and yet ill effects are probably produced by it under some conditions.

Solanum nigrum L. Common Nightshade or Stubbleberry.

Annual, low branched and often spreading; stem glabrous or hairy, hairs simple, roughened on the angles; leaves ovate, petioled, flowers white in small, umbel-like drooping lateral clusters; calyx spreading, the lobes obtuse, much shorter than the white corolla; berries glabrous, globose, black; occasionally large,

Distribution. Found in northern United States. Also occurs in Europe. Shady grounds and fields. A cosmopolitan weed.

Poisonous properties. Stubbleberries are occasionally cultivated for their fruit. They are sometimes sold as huckleberries and used for pies and pre-



Fig. 420. Black Night-shade (Solanum nigrum). Unripe fruit thought to be poisonous. (Charlotte M. King).

serves. The fruit should, however, be used with caution, especially before it is ripe. A decoction of the ripe stubbleberry when fed to cats does not appear to be poisonous, according to Dr. Buchanan, who fed considerable quantities to cats in the writer's laboratory without injurious effects. Mr. Chesnut says:

The amount of poison present in any part of this plant varies with the conditions of growth. The more musky-odored plants are the most poisonous. In some, the amount of alkaloid in the ripe fruit and leaves is so small that these parts may be, and are, consumed in considerable quantity without any ill consequences. Poisoning does sometimes follow, but it is not clear whether this is due to improper preparation or to careless selection of the parts used. The use of black nightshade for food is certainly not to be recommended. Cases of poisoning are recorded for calves, sheep, goats and swine.

The characteristic symptoms are about the same in man and animals. They are stupefaction, staggering, loss of speech, feeling and consciousness; cramps, and sometimes convulsions. The pupil of the eye is generally dilated. Death is directly due to a paralysis of the lungs, but fortunately few cases are fatal.

In June, Dr. Flickinger reported from Greenfield, Iowa, several cases of sheep poisoning probably due to eating plants of *Solanum nigrum*. A subscriber of the Iowa Homestead also says that for a number of years poisoning has affected his

sheep in a pasture where this weed has been abundant. Dr. Koto, Iowa State Veterinarian, also reports cases due to this plant.

This Black Nightshade contains the alkaloid solanin C52H97NO18which is probably present in larger quantities in the fruit before it is entirely ripe, also solanidin CanHanNO2 with stronger basic properties. From all the evidence I can get, I may say that the fruit should be eaten with caution. In Europe it has been looked upon with suspicion for a long time. The ancients held it in suspicion and many superstitious beliefs were connected with it. Schimpfky in discussing the poisonous and non-poisonous action of the plant states that the amount of poison produced depends upon climatic conditions and the character of the soil. In some places it may be entirely harmless and in other places poisonous. In Europe the plant is sometimes used as a salad plant, but the author above referred to, remarks that when the odor is unpleasant and disagrecable it should not be eaten.

According to Lehmann, Schreber and Haller the berries are poisonous to ducks and chickens. Cases of poisoning from the berries of this plant have been recorded in Europe by Hirtz,1 Manners,2 etc.



Fig. 421. Spreading nightshade (Striflorum), one-third natural size. Stof being poisonous. (U. S. Dept. Agr.). (Solanum

¹ Gaz. Med. d. Strasbourg. 1842.
2 Edin. Med. Jour. 1867.
For these and other references see Blyth "Poisons" 4th ed., 398. Poisoning by bitter sweet berries is recorded in Lancet, 1856. Berries of S. tuberosum, Brit. Med. Jour. 1895.

Solanum triflorum Nutt. Three-flowered Nightshade

Annual, low spreading, slightly hairy or nearly glabrous; leaves acute; pinnatifid, 7-9 lobed; peduncles 1-3 flowered; corolla white; berries greenish or inclined to blackish, about the size of a small cherry; pedicels reflexed in fruit.

Distribution. On the plains, and waste places from Nebraska and Kansas to Northwest Territory and Arizona; introduced castward.

Poisonous properties. Prof. Chesnut says experiments on guinea pigs show that the berries are poisonous. No cases of human poisoning have been reported. The berry is not attractive to the eye, but has an agreeable odor and taste. It is therefore to be suspected in cases of poisoning which occur in localities where the weed is abundant. The writer has also received complaints of the poisoning by this plant from Nebraska and other western states. The active constituent is, no doubt, solanin.

Solanum rostratum Dunal. Buffalo Bur. Sand Bur

Herbaceous, woody when old; somewhat hoary or yellowish; 8 inches to 2 feet high; covered with copious stellate pubescence; the branches and stems covered with sharp yellow prickles; leaves somewhat melon like, 1-3 times

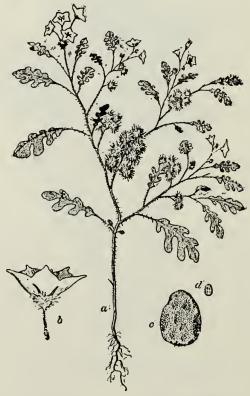


Fig. 422. Buffalo Bur (Solanum rostratum). a, branches of the plant with burs; b, yellow flowers; c and d, seeds; c, very much enlarged; d, natural size. The prickles cause mechanical injuries to stock.

pinnatifid; lobes roundish or obtuse and repand, covered with soft pubescence, hairs stellate; flowers yellow; corolla gamopetalous, 1 inch in diameter, nearly regular, the sharp lobes of the corolla broadly ovate; stamens, 5, declined, anthers tapering upward, linear lanceolate, dissimilar, the lowest much larger and longer with incurved beak, hence the technical name rostratum; style much declined; fruit a berry but enclosed by the close fitting and prickly calyx, which has suggested the common name buffalo bur or sand bur; pedicels in fruit erect; seeds thick, irregular, round or somewhat longer than broad, wrinkled showing numerous small pits; seeds surrounded by a gelatinous substance. The related species S. citrullifolium of the southwest is glandular pubescent with slender yellow subulate prickles, lowest anther violet.

Distribution. The buffalo bur was undoubtedly a native to the region of the

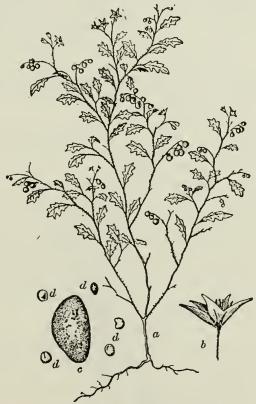


Fig. 423. Horse nettle (Solanum carolinense), a, plant with flowers and fruit; b, flower; c, seed enlarged. Considered poisonous by Bessey and others. (U. S. Dept. Agr.).

plains occurring in the bare places where grass is scant and in former times was most abundant around the "buffalo wallows." Its range is from New Mexico to Wyoming and across the plains. The general traffic from the west to the east has caused the weed to be distributed in various eastern and middle states, Iowa, Minnesota, Wisconsin, Missouri, Illinois, Indiana, Ohio, New York, Massachusetts and Tennessee.

Injurious properties. This plant is carefully avoided by stock, but so far as known is not poisonous although the prickles on the plant produce mechanical injuries. When the prickles enter the tissues of animals inflammation occurs and pus is formed.

Solanum carolinense L. Horse Nettle

Horse nettle is a deep rooting perennial, propagating freely by its underground roots; these running roots are often 3 feet long; stem from 1-2 feet high, somewhat straggling, half shrubby at the base; stems hairy or merely roughish with minute hairs which are usually numerous; leaves oblong or sometimes ovate, obtusely sinuate, toothed or lobed or deeply cut, 2-4 inches long; flowers borne in racemes which later become 1-sided; the outer part of the flower, the calyx, consists of slender lobes; the corolla is light blue or white, an inch or less in diameter and resembles that of the common potato; the flowers are followed by yellow globose berries, 1/2-3/4 inch in diameter; the small seeds are yellowish, a little less than 1/12 of an inch long, minutely roughened.

Distribution. Its distribution in North America is from Connecticut through New York, Pennsylvania and New Jersey, West Virginia along the Atlantic seacoast to Florida, west along the Gulf Coast to Texas, through Kansas, Nebraska, Iowa, Illinois and Michigan.

Poisonous properties. The root as well as the fruit of the plant has a very disagreeable narcotic odor; according to several authors, the plant is poisonous. Dr. Bessey reports it as possibly poisonous. It contains solanin, according to Kraemer, 0.8 per cent in the berries.

Solanum elacagnifolium Cav. Horse-weed. Bull Nettle

A deep-rooted spreading perennial from 1-3 feet high; stem silvery canescent, finely pubescent; leaves lanceolate, oblong or linear, petioled entire or repand-dentate; flowers in cymose clusters; peduncle stout and short; corolla gamopetalous, blue; calyx lobes lanceolate; berry yellow; smooth globose.

Distribution. Common on the prairies of Kansas to Texas and New Mexico. Poisonous properties. The berries of this fruit are used to curdle milk in northern Mexico and southern Texas. They are crushed into a powder, put into a muslin bag, suspended in the milk until coagulation occurs. It is also used as a medicine by the Mexicans.

Solanum tuberosum L. Potato

An erect herb, cultivated as an annual for the esculent tubers; leaves pinnate of several ovate leaslets and smaller ones between; flowers blue or white, berries round, green.

Distribution. Native to Chile north to Mexico and Arizona. Introduced into Europe between 1580 and 1585.

Poisonous properties. The wilted green stem and leaves are poisonous, containing the alkaloid solanin. The water from boiled potatoes contains a poisonous substance. Some persons cannot eat potatoes because poisonous to them. According to Kassner, healthy potatoes do not contain solanin but diseased potatoes contain this substance; from 150 gms. he separated 30-50 mgrs. of solanin. It is probable that this substance occurs in other species of Solanin. Thos. Maiden states that the S. eremophilum poisons sheep and cattle when they cat the tops. Friedberger and Fröhner state that potato tops are injurious; that the diseased animals show symptoms resembling foot and mouth disease.

Dr. Doerr has recently reported a case of poisoning in a cow that had been fed exclusively upon refuse from the Club-house kitchens near the Iowa State College Campus. These contained potato parings among other things and to these the poisoning was charged. The trouble was diagnosed as gastro-enteritis. The post-mortem revealed diffuse intestinal hemorrhages with enlarged liver and spleen.

2. Capsicum. Pepper

Herbs or shrubs with sharp taste; leaves fleshy; flowers white; corolla wheel shaped; 5-lobed; tube short; stamens separate with filaments longer than the heart shaped anthers which open longitudinally; fruit a berry. The Guinea pepper and the Indian goat pepper (C. frutescens) are much more powerful stimulants than cayenne and often produce violent pain and purging. This shrub is native to the Southwest. The genus Capsicum has two species. A monograph by Prof. Irish published in the Missouri Botanical Garden Reports describes many of the varieties.

Capsicum annuum L. Cayenne Pepper

Annual. Leaves ovate entire; flowers with truncate calyx and white corolla; fruit a berry, oblong or globular, red or green.

Distribution. Widely cultivated. Native to the Southwest.

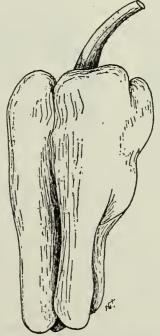


Fig. 424. Red Pepper (Capsicum annuum). The fruit of this as well as the leaves are powerfully pungent. (W. S. Dudgeon).

Poisonous properties. The peppers are often used in domestic practice in making a stimulating plaster; if its action is continued long enough, however, a vesicular formation makes its appearance. In domestic animals it causes

gastro-enteritis. Death has even occurred where too much of the Cayenne pepper has been used. Thresh isolated a principle to which he gave the name of capsaicin $C_9H_{14}NO_2$, however, the bitter pungent substance has been called by Morbitz, capsacutin, a crystalline nitrogenous compound containing the substance $C_{35}H_{54}N_3O_4$. This is so powerful that 1 part in 11,000,000 will impart the pungent taste. A volatile alkaloid resembling contin has been found in small quantities.

3. Nicandra Adans. Apple of Peru

Tall smooth annual with alternate leaves, calyx 5-parted, angled; corolla wheel shaped or somewhat funnel shaped with widely spreading border; tube short; anthers 5 connivent; fruit a somewhat 3-5 celled berry.

Nicandra Physalodes (L.) Pers. Apple of Peru

A tall smooth annual, 2-5 feet high; leaves ovate angled or sinuate toothed; flowers solitary; corolla pale blue rather large; fruit a globular dry berry; calyx, 5-parted, 5-angled, enlarged and bladder like in fruit.

Distribution. Native to Peru but sparingly naturalized in the United States. *Poisonous propertics*. Said to be poisonous; used as a fly poison in parts of the United States.

4. Hyoscyamus (Tourn.) L. Henbane

Clammy-pubescent, fetid, narcotic herbs; leaves alternate, mostly lobed or pinnatifid; flowers large, calyx bell-shaped or urn-shaped, 5-lobed; corolla funnelform, oblique; 5-cleft, the lobes unequal; capsule enclosed in the persistent calyx, 2-celled.

About 15 species, native to the Mediterranean region. Medicinal and poisonous plants.

Hyoscyamus niger L. Black Henbane

Biennial or annual. Stem 1-3 feet high; leaves ovate, sinuate toothed and angled, the upper clasping; flowers short pedicelled in one sided leafy spikes; corolla dull yellowish, reticulated, with purple veins; capsule globose oblong.

Distribution. Common only eastward in waste places from Nova Scotia to Michigan, also in Montana, Utah, Idaho and the Pacific Coast.

Poisonous properties. A well known medicinal plant from which hyoscyamin is obtained. Hyoscyamin is an anodyne and hypnotic and is poisonous. Dr. Chesnut says:

One or two cases are recorded in foreign literature in which stock have been poisoned by eating the plant of their own accord, but there is very little danger from it, on account of its ill odor and harsh texture.

It contains hyoscyamin, $C_{17}H_{23}NO_3$, causing a dilation of the pupils and having a sharp and disagreeable taste. It also contains pseudo-hyoscyamin $C_{17}H_{23}NO_3$, another alkaloid, and hyoscin $C_{17}H_{23}NO_3$; the latter of which also dilates the pupils. The hyoscyamin resembles atropin in its composition and action and is obtained from the Hyoscyamus seed. When damp the alkaloid has a tobacco-like odor and a bitter taste. According to Dr. Winslow, the hyocsyamin is practically atropin except that its mydriatic action is shorter. Hyoscin is a powerful depressant to the cerebrum, respiratory center, spinal reflex centers, and motor tract. It is a cerebral sedative. According to Winslow:

The tetanic stage succeeding spinal paralysis, observed in atropin C $_{17}^{}H_{23}^{}$ NO poisoning, does not ensue with hyoscin. The latter alkaloid slightly depresses and slows the

heart, and does not paralyze the vagus terminations, nor depress the motor and sensory nerves or muscles. The circulation is but slightly influenced, and vasomotor depression only occurs in the latter stage of lethal poisoning. Death occurs from paralysis of the respiratory centers. Poisoning in animals is exhibited by loss of muscular power, slowing and failure of respiration, dryness of the mouth, stupor and asplyxia. The pulse may be infrequent, the pupils are dilated and the skin is moist rather than dry. Delirium and convulsions sometimes occur in man. The effect of the combined action of hyoscyamia and hyoscin in Hyoscyamus is shown when we compare the drug with belladonna. Hyoscyamus is more of a cerebral sedative and hypnotic, and less of a heart and respiratory stimulant. It is said to possess more power in overcoming spasm, and griping of cathartics, and in aiding intestinal movement. Hyoscyamus is also thought to exert a more pronounced antispasmodic action than belladonna upon the smooth muscles of the bladder and urethra.

Blyth gives the action of hyoscyamin as follows:

Thirty-two mgrs. (½ gr.) hegins to act within a quarter of an hour, the face flushes, the pupils dilate, there is no excitement, all muscular motion is enfectled, and the patient remains quiet for many hours; 64.8 mgrs. would possibly be a fatal dose. The root is

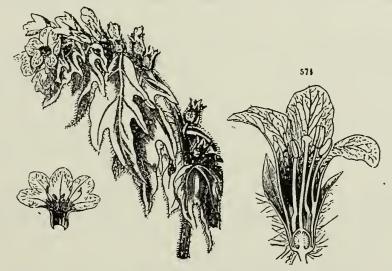


Fig. 425. Black Henbane, (Hyoscyamus niger). At the left, open corolla, and flowering branch. At the right, longitudinal section of flower. A well known medicinal plant. (From Vesque's Traité de Botanique).

more poisonous than the leaves, and the seeds of Datura contain a considerable quantity of hyoscyamin; they are often mistaken for other seeds such as poppy.

Many cases of children being poisoned by this seed are recorded. One instance is given by Schimfky where of two children who had eaten the seeds' of the plant, one died before purgative action could be produced. The second child slowly recovered but growth was checked.

5. Nicotiana (Tourn.) L. Tobacco

Rank, viscid-pubescent narcotic herbs or shrubs; leaves alternate, entire; flowers borne in panieled racemes; calyx tubular, bell-shaped, 5-cleft; corolla funnel-form or salver-form, the limb with five separating lobes; stamens 5, inserted on the tube of the corolla; dehiscence of the anthers longitudinal; ovary. 2-celled; stigma capitate; capsule 2-valved; seeds numerous, small. About 50 species nearly all native of North America.

Nicotiana Tabacum L. Common Tobacco

A coarse annual from 4-6 feet high; leaves lanceolate, ovate, decurrent,

1-2 feet long; flowers panicled, rose-purple; corolla funnel-form, 2 inches long; lobes short, somewhat inflated.

Distribution. Native to South America but widely cultivated; introduced into Europe by the Spaniards shortly after the discovery of America. The best types of tobacco are cultivated in Cuba and Porto Rico; and this forms an extensive industry in North America, especially in Connecticut, Wisconsin, Virginia, etc.

Poisonous properties. Various opinions are expressed in regard to its poisonous properties. It is known, however, that an alkaloid occurs in N. Tabacum, N. macrophylla, N. rustica and N. glutinosa, which apparently does not occur in any other plant.

The active principle of the tobacco leaf is the alkaloid nicotin $C_{10}H_{14}N_2$, which is easily extracted from tobacco by means of alcohol or water; it occurs to the extent of 6 per cent in the dry leaves; it has a sharp, burning taste, is very poisonous and is said to have sixteen times the toxic power of coniin. On application of heat, nicotin is changed into pyridin, C_5H_5N , and other similar alkaloids like picolin, CHN. Pyridin depresses the spinal motor tract and causes paralysis of respiration. Moderate doses cause contraction of the pupil. Nicotein $C_{10}H_{12}N_2$ was found by Pictet and Rotschy in leaves of tobacco; also nicotemin $C_{10}H_{14}N_2$, and nicotellin $C_{10}H_8N_2$. According to more recent investigations the seeds of Nicotiana are free of nicotin. The following statement is made with reference to the toxicology of nicotin by Dr. Winslow:

Nicotin is one of the most powerful and rapidly acting poisons. When swallowed, it causes, in animals, local irritation and pain in the throat and stomach; muscular tremors and weakness, on account of which the animal falls. These symptoms are followed, first, by severe tonic and clonic convulsions, and then by abolition of voluntary motion and quietude. The pupils are contracted, and there is vomiting (in the case of some animals), purging and micturition. The respiration is at first shallow and rapid, but becomes weaker and slower, and death occurs from respiratory failure and general collapse. The pulse is primarily slow and intermittent, but later becomes rapid. The treatment of poisoning consists in evacuation of the stomach; the use of tannic acid; respiratory and heart stimulants, as strychnin, atropin, and alcohol; together with external heat and artificial respiration. The minimum lethal dose is about one drachm of tobacco, or one minim of nicotine, for small dogs. For horses, five to ten drops of nicotin or one-half pound of tobacco.

Friedberger and Fröhner state that animals have convulsive spasms, great muscular weakness, with acute paralysis.

The chronic effects of the use of tobacco, according to Millspaugh, are as follows:

The effects produced upon smokers are almost useless in the study of the drug itself, and it is only in that class of chewers who swallow the juice, that positive data could be looked for; still here, as well, we are at a loss to determine facts, for in manufacturing the narcotic, processes are used which alter the product greatly; nevertheless some few symptoms seem to be more or less common to all who have been, for protracted periods, subjected to the drug. Mental anxiety and irritability, with at times confusion of ideas; dilation of the pupils; ringing in the ears; increased secretion of saliva; uncertainty of speech, dryness of the throat; at times weakness of the stomach and nausea; increased secretion of urine; dry cough especially at night; præcordial oppression with palpitation of the heart and at times an irregular pulse; trembling of the extremities when held long in one position; general an, minic condition of the blood spasmodic contraction or jactation of single muscles; sensations of exhaustion and especially lassitude; sleepiness; profuse perspiration and sensitiveness to cold.

A writer in the London Lancet quoting from the Therapeutic Gazette* states that the injury from tobacco smoke comes largely from the inhalation of carbon monoxide. Cigarette smoking is more harmful than smoking a pipe because more of the gas is inhaled.

^{*} Vol. 32:78C.

According to Zalackas, eserin and strychnin are not antagonistic to nicotin but the juice of Nasturtium officinale counteracts it.

Nicotiana rustica L. Wild Tobacco

Annual, with obovate, petioled leaves; flowers greenish-yellow, panicled, longer than the calyx; capsule globose.

Distribution. In fields and waste places from Canada to Florida, and Minnesota. Cultivated by the Indians.

Poisonous properties. Probably the same as those of the preceding species.

Nicotiana alata Link & Otto

An annual, pubescent plant from 3-4 feet high; leaves lanceolate, flowers large tubular; tube 5-6 inches long; the limb deeply 5-cleft.

Distribution. Native to Brazil but a frequent escape in gardens in the east.

Nicotiana quadrivalvis Pursh. Wild Tobacco

An annual 1 or more feet high, leaves oblong or the upper lanceolate and the lower obovate lanceolate, acute at both ends; flowers few; corolla white, tubular funnel-form; tube 1 inch long.

Distribution. Oregon to the plains. Often cultivated by the Indians for tobacco. The allied species, N. attenuata, Torr., is found from Colorado to Nevada and California.

Poisonous properties. The poisonous properties are probably the same as those of common tobacco. According to Maiden the N. suaveolens is poisonous to stock in New South Wales.

6. Datura L. Thorn Apple. Jimson Weed

Rank-scented, tall, narcotic herbs; or a few tropical shrubs or trees with alternate petioled leaves; large flowers; calyx 5-cleft; corolla funnelform, 5-lobed, the limb plaited; stamens generally included, inserted at or below the middle of the corolla tube; ovary 2-celled, forming a capsule which is globular and prickly.

A small genus of about 12 species of wide distribution, 2 being cosmopolitan weeds. Several of the species are used for ornamental purposes. Among these are the common white-flowered thorn apple (Datura Metel) which is native to tropical America, also the Datura meteloides, native to New Mexico and cultivated for its large sweet-scented flowers. Several tropical American tree-like shrubs like Datura suaveolens are often cultivated in conservatories. The seeds of D. fastuosa are used in India as a poison according to Gimlette, and are commonly used on the Malay Peninsula. The D. alba is common in India, about Madras, and D. atrox occurs on the coast of Malabar. The Daturas are all important in India from the point of view of poisoning. The seeds of D. alba are often mistaken and eaten for the seeds of Capsicum.

Datura Tatula L. Purple Thorn-apple or Purple Stramonium

A glabrous annual from a few inches to 5 feet high; stem purplish; leaves thin, ovate, acute or acuminate; flowers consisting of a 5-toothed calyx and a 5-lobed funnel-form corolla, with stamens included; filiform filaments inserted below the middle of the corolla tube; capsule globular, prickly, 4-valved and 2-celled.

Distribution. Abundant in fields and waste places from New England to

Ontario to North Dakota, Nebraska, Texas, and Florida, naturalized from tropical America.

Poisonous properties. The poisonous alkaloids found in this plant are: atropiu $C_{17}H_{23}NO_3$, hyoscyamin, and hyoscin. The daturin is a mixture of hyoscyamin and atropin.

Professor Chesnut, in his work on the Poisonous Plants of the United States, referring to the jimson weed, says:

The poisonous alkaloids, atropin and hyascyamin, the active constituents of belladonna are found also in both of the jimson weeds. Hyascyamin is the poison of the henbane and as it is identical in its physiological action with atrapin, the abovenamed plants present the same symptoms of poisoning, which must be met in the same manner. The alkaloids exist in all parts of the two daturas. The seeds are especially poisonous.



Fig. 426. Jinson Weed (Datura Stamonium). a, leaf and flowers; b, fruiting capsule.

Cases of poisoning arise in adults from excessive use of a stImulant or a medicine. Children are sometimes tempted to eat the fruit, if they are permitted to play where the weed is to be found. Several cases of this kind were reported to the Department during the fall of 1897. At Alpena, Michigan, five children were badly poisoned in August by eating the seeds of the purple-flowered species, which was cultivated in a garden as a curiosity under the fanciful, trade name of "Night-blooming Cactus." In Sept. a boy was killed in New York by eating the seeds of a jimson weed, which was permitted to grow in a vacant lot; his brother poisoned at the same time was saved only with difficulty. In October two other cases occurred in New York. Four children were playing in one of the public parks of the city where jimson weed were growing luxuriantly. The boys imagined themselves Indians and roamed about and ate parts of various plants. Three of them ate the seeds of the jimson weed. One died in a state of wild delirium; another was saved after heroic treatment with chloral hydrate and morphine;

the third, who ate but few of the seeds, was but little affected. Children are also poisoned by sucking the flower, or playing with it in the mouth. The fresh green leaves and also the root have occasionally been cooked by mistake for other wild edible plants. One or two instances are recorded in which cattle have been poisoned by eating the leaves of young plants which were present in grass hay, but these animals generally either avoid the plants or are very resistant to its poison.

The symptoms of the poisoning are about the same in all cases, those characteristic of large doses being headache, vertigo, nausea, extreme thirst, dry, burning skin, and general nervous confusion, with dilated pupils, loss of sight and of voluntary motion, and sometimes mania, convulsions, and death. In smaller amounts the effects are like those of the ordinary narcovies. As vomiting is not a common symptom, the contents of the stomach must be quickly removed by the use of the stomach tube or emetics. It is well then to wash out that organ thoroughly with strong tea, tannic acid, or an infusion of oak bark, and to administer stimulants, such as brandy and hot, strong coffee. Pilocarpin is recommended by physicians to counteract the drying effect upon the secretions (licorice is very useful), and prolonged artificial respiration must often be resorted to to maintain the artation of the blood.

As nothing has been said in regard to the atropin which is found in the jimson weed, it might be said that the commercial atropin is derived from the root of belladomia and when used externally it is a local anodyne. Dilute solutions of acropin paralyze and stop the corpuscular movement in the blood and large doses give rise to slowing of the pulse. In poisoning it causes a paralysis of the vascular motor centers and stimulates the brain; large doses produce restlessness and excitement and delirium in man and occasionally delirium in lower animals. With reference to the spinal cord, large doses cause complete loss of motion. Its action upon the nerves is very important and on this depends much of the value of the drug. Dr. Winslow says:

The peripheral motor nerve terminations, and to a less extent, their trunks, are depressed and paralyzed. This is never so complete, however, but that there is some voluntary power left in an animal fatally poisoned.

Dryness of the mouth is one of the first symptoms following the use of belladonna because of the paralysis of the peripheral terminations of the secretory nerve. The involuntary nerves are not affected by moderate doses of belladonna. The motor nerves ending in the voluntary muscles are paralyzed by poisonous doses of belladonna. Small doses do not affect the respiration, large doses make it quicker and deeper. Fatal doses cause asphyxia. Moderate doses cause a rise of temperature, but fatal doses lessen the bodily heat. Dr. Winslow gives the following summary of the action of the drug:

It will be observed that belladonna, generally speaking, first stimulates and then depresses the nerve centres, while it chiefly paralyzes the motor nerve terminations, including the inhibitory (vagus and splanchnic), the secretory (Chorda tympani, etc.), and, to a less extent, the sensory nerves. Secondary depression of the cerebrum is not so profound as that of the great medullary centres, especially the respiratory centre, and there is sometimes a slight and brief stimulation of the motor nerves of the smooth muscles, viz., vagus, splanchnic, and possibly vaso-motor nerves.

Full medicinal doses depress the peripheral filaments of the inhibitory and secretory nerves, and those of the unstriped muscles, lessen the functional activity of the voluntary motor system, and, to a less degree, that of the afferent nerves. The pulse becomes quickened because of paralysis of the peripheral vagus endings and stimulation of the heart or its ganglia; the blood tension is augmented because of the increased cardiac action and stimulation of the vasomotor centers; and the respiration is accelerated because of excitation of the respiratory centers. The temperature is elevated owing to the circulatory exaltation and stimulation of the heat-producing centers. Slight delirium may be present from the exciting action of the drug upon the cerebral motor centers.

The spinal cord is unaffected by therapeutic doses. Locally applied, belladonna is a direct paralyzant to nerves, muscles, vessels and cells.

Toxic doses of belladonna cause in animals dryness of the mouth, increased frequency of the pulse and respiration, elevation of temperature, dilation of the pupil and partial blindness, restlessness, nervousness, delirium, twitching of the muscles (occasionally erythema), and frequent micturition. These symptoms are succeeded, in fatal poisoning, by fall of temperature, retention of urine, muscular weakness, staggering gait, partial anæsthesia, convulsions and paralysis (one preponderating over the other), weak, slow, irregular respiration, feeble, rapid pulse, paralysis of the sphincters, stupor and death. Death occurs mainly from asphyxia, but is due in part to cardiac failure. The physiological test consists in placing a drop of urine (secreted by the poisoned animal) into the eye of a healthy animal, when mydriasis should follow if the case be one of belladonna poisoning. Three-quarters of a grain of atropin under the skin has proved fatal to dogs. Two grains of atropin produce mild toxic symptoms in the horse. Small dogs are slightly poisoned by gr. 1-80 of atropin; medium sized dogs by gr. 1-60, given hypodermatically. Cattle are as susceptible as horses, although herbivora are not so easily influenced as carnivora. The pulse in dogs is greatly accelerated, sometimes as high as 400, while the pulse rate of the horse is not generally more than doubled. Rodents, as guinea pigs and rabbits, and pigeons, are particularly insusceptible to belladonna, in regard to its effect upon the pupil, circulation, etc.

The treatment for poisoning includes the use of the stomach pump, emetics, cardiac stimulants, and pilocarpin under the skin. Also external heat, general faradism and artificial respiration.

Datura Stramonium L. Jamestown or Jimson Weed

Much like the last; an annual; glabrous or the young stem somewhat pubescent, stout and green; branches and leaves sparingly pubescent; leaves thin ovate, sinuate toothed or angled; calyx less than ½ the length of the corolla; corolla white, 3 inches long, the border 5-toothed; capsule ovoid, prickly, the lower prickles mostly shorter.

Distribution. Nova Scotia, New England to Minnesota, Iowa, Nebraska, Texas and Florida. Naturalized, native to tropical regions of the Old World, probably Asia. De Candolle says that it is probably native to the borders of the Caspian Sea.

Poisonous properties. The Datura Stramonium has been used in medicine since the close of the sixteenth century. Earlier than this it was used by the people of western Asia and eastern Europe. Miss Henkel describes the method of collecting as follows:

The leaves are collected at the time of flowering, the entire plant being cut or pulled up and the leaves stripped and dried in the shade. The unpleasant narcotic odor diminishes upon drying. The leaves are poisonous, causing dilation of the pupil of the eye, and are used principally in asthma.

All three species of *Datura* are poisonous, the seeds being especially poisonous. Dr. Halsted records a case of poisoning of a boy five years old in Newark, New Jersey, who ate freely of a half grown capsule of this species and died the next morning. The seeds of *D. Stramonium* are known to have poisoned a child in eastern Iowa. They contain hyoscyamin, atropin and scopolamin. In some analyses, as much as 0.33 per cent of the alkaloid atropin has been found in the seeds, and about 0.2 per cent in the leaves.

Datura Metel L. Thorn Apple

A clammy pubescent annual 3-4 feet high, leaves ovate, entire or obscurely angular toothed, rounded at the base; flowers large, white, calyx about ½ as long as the corolla; capsule globose prickly.

Distribution. Native to tropical America. Naturalized from New England to Florida and westward,

Datura meteloides DC. Wright's Datura

A spiny pubescent annual, pale in color, leaves obovate entire; flowers large showy, white or pale violet, sweet scented; corolla with a 5-toothed border; capsule nodding spiny.

Poisonous properties. The writer a few years ago saw a notice in a local

paper of a child being poisoned by sucking the nectar of a flower of Wright's Datura. Professor Chesnut, in speaking of the poisonous properties of the same species, says:

Datura meteloides is a very large-flowered species, which is native from southern California to Texas, and in some localities is common in cultivaton. No cases of poisoning have yet been recorded against it, but it is largely used as an intoxicant by Indians, and is used in general for the same purposes as jimson weed. It undoubtedly contains the same poisons.

7. Lycium L. Matrimony Vine

Shrubs or woody vines, often spiny; leaves small, entire, alternate, with smaller ones between; calyx 3-5 toothed or cleft, persistent, corolla funnel-form or salver-shaped, usually 5-lobed, the lobes imbricated; stamens 5 rarely 4; anthers opening lengthwise; style slender; stigma capitate; ovary 2-celled; berry globose, ovoid or oblong.

About 75 species of wide distribution.

Lycium halimifolium Mill. Common Matrimony Vine

A glabrous, spiny, or unarmed shrub; slender climbing or trailing stem; leaves lanceolate, oblong or spatulate; petioles short; peduncles filiform; calyx lobes ovate; corolla short, funnel-form; greenish-purple stamens slightly exserted; berry oval; orange red.

Distribution. In thickets and waste places, escaped from gardens from Canada to Texas. Introduced from Europe.

Poisonous properties. Supposed to be poisonous.

SCROPHULARIACEAE. Figwort Family

Mostly herbs, shrubs or rarely trees; leaves without stipules; flowers perfect, regular or irregular; calyx 4-5 toothed, cleft or divided; corolla irregular 2-lipped or nearly regular; stamens 2-5, didynamous or nearly equal, inserted on the corolla; pistil 1, 2-celled, many ovuled; fruit a capsule; seeds numerous, with a small embryo in copious albumen.

About 2500 species of wide distribution. Few plants of the family are of economic importance. Several species are medicinal. The most important are foxglove (Digitalis purpurea); mullein (Verbascum Thapsus), used as a stimulant because of its mucilaginous properties, and speedwell or Culver's root or Culver's physic (Veronica virginica), used for digestive disorders, when fresh being a violent emetic-cathartic and containing a glucoside leptandrin. Several species of the order are cultivated for ornamental purposes. The Paulownia tomentosa, native to Japan, is hardy in the south. The foxglove is also much cultivated. Several species of monkey flowers (Minulus luteus) and musk flower (M. moschatus) are cultivated. The best known of all is the snap dragon (Autirrhinum majus), native to Europe. The genus Calceolaria, of which there are numerous species, is native to Chili and other parts of South America, and is cultivated. The C. crenatiflora is a showy herbaceous plant cultivated for its pretty slipper-shaped, sac-like flowers. The Maurandias are Mexican climbers with heart-shaped or halberd-shaped leaves and openmouthed, somewhat bell-shaped, purple, rose-colored or violet corollas. The Torenia asiatica, native to Asia, is cultivated for its handsome pale violet or purple flowers. The turtle-head (Cheloue glabra) is occasionally cultivated and has large, white or rose-tinted corollas that are very pretty. The beard-tongue (Pensteinoii) contains many species, found mostly in western America and Mexico, the most beautiful of our western species being P. grandiflorus, with

large, lilac-purple flowers collected in ample racemes. Many of the Rocky Mountain forms are handsome perennials. The Mexican Russellia juncea is a showy bedding and greenhouse plant with carmine flowers and leaves reduced to scales. The painted cup (Castilleia coccinea) is a pretty species, native of the northern states. The roots of most species are parasitic. The common lousewort (Pedicularis canadensis) is an early spring blooming plant of northern prairies. P. groeulandica of Europe, is found in the colder regions of North America also and has handsome purple flowers borne in spikes. The flowers of Lypevia atropurpurea or Cape Saffron resemble true saffron very greatly in odor, taste and drying qualities.

Vanquelin isolated the glucoside gratiolin C₄₀H₇₀O₁₅ from Gratiola of-ficinalis. This species is poisonous to stock; strong medicinal doses are poisonous to man as well. Some species, according to Maiden, are often poisonous to stock in Australia. The cow wheat of Europe (Melampyrum arvense) cause-colic and sleepiness.

Genera of Scrophulariaceae

| Flowers | regular | or | nearly | so. |
|---------|---------|----|--------|-----|
|---------|---------|----|--------|-----|

Stamens 4 not in pairs.

1. Verbascum (Tourn.) L. Mullein

Biennial or perennial, generally tall herbs with alternate leaves; flowers in spikes, racemes or panicles; calyx 5-parted; corolla flat with 5 broad rounded or slightly unequal divisions; stamens 5, inserted on the base of the corolla unequal; filaments of all of the stamens woolly or only the 3 upper; style flattened at the apex; fruit a capsule, 2-valved; seeds rough.

About 125 Old World species. Several naturalized in North America.

Verbascum Thapsus L. Common Mullein

A tall, densely woolly annual from 2-6 feet high; leaves oblong, thick, covered with branched hairs, the basal leaves margined petioled; flowers in long dense spikes; corolla rotate, yellow or rarely white; stamens unequal, the 3 upper shorter, woolly with short anthers; the 2 lower smooth with large anthers.

Distribution. From Nova Scotia north across the continent; south to Missouri and Kansas and west to Utah.

Verbascum Blattaria L. Moth Mullein

Stem round, sparingly branched, biennial with smooth leaves, the lower petioled, oblong, ovate, lanceolate, laciniate, serrate, upper clasping; flowers in loose racemes, yellow or white with a tinge of purple; all the stamens bearded with violet hairs; capsule nearly globose; numerous seeds.

Distribution. Common eastward, rarely in the Mississippi Valley. Abundant in the west in Salt Lake basin.

Poisonous properties. The common mullein produces an irritation but is probably not very poisonous to stock.

2. Linaria (Tourn.) Hill. Toad Flax

Herbs or shrubs, with alternate leaves, or those of the sterile shoots opposite or whorled; flowers in raceines or spikes; ealyx 5-parted; corolla per-

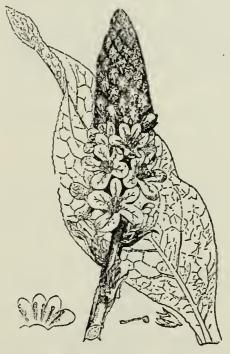


Fig. 427. Verbaseum Thapsus. (From Johnson's Medical Botany of N. A.).



Fig. 428. Toad flax (Linaria tulgaris). a, seed. Regarded with suspicion. (Selby).

sonate and with a spur at the base; the upper lip erect, 2-lobed, the lower 3-lobed; stamens 4, didynamous, not exserted; fruit a capsule, opening by 1 or more holes in the top; seeds small, numerous.

About 150 species of wide distribution. One native species in the northern states.

Linaria vulgaris Hill. Ramsted. Butter and Eggs.

A pale green perennial with erect, leafy globose or sparingly pubescent stem; leaves sessile, entire, upper, at least, alternate; flowers in dense racemes; calyx segment oblong, spur subulate; corolla orange color, nearly erect, 1 inch long, spur subulate, nearly as long as the body of the corolla.

Distribution. Native to Europe. In fields and waste places from Nova Seotia to Kansas, North to Manitoba.

Poisonous properties. It is regarded with suspicion. The plant has a very disagreeable odor. A glucoside linariin $C_{64}H_{50}O_{40}$ has been isolated.

Veronica peregrina L. Purslane Speedwell, Neckweed

Glabrous, glandular, or nearly smooth, branehing annual, 4-9 inches high;

leaves petioled, upper oblong, linear and entire; floral leaves like those of the stem but reduced; flowers axillary and solitary, white; capsule orbicular.

Distribution. A common weed in fields in eastern North America from Nova Scotia to Florida, Missouri, Kansas, Texas, British Columbia and California. Also found in Mexico, South America and in Europe, almost cosmopolitan.

Veronica virginica L. Culver's Root. Culver's Physic

A tall, smooth, or occasionally somewhat hairy perennial, simple stem; leaves lanceolate pointed in whorls finely serrate; flowers in panicled spikes, small nearly white, salverform, tube longer than the calyx; stamens 2, exserted; pistil 1, style 1; fruit a capsule, oblong-ovate.

Distribution. From western New England to Minnesota, Manitoba to Nebraska and Kansas.

Poisonous properties. The V, peregrina has been reported as poisonous. The root of V, virginica contains leptandrin. It is a violent emetic, cathartic, and, according to Johnson, cannot be used with safety in medicine.

3. Digitalis L. Foxglove

Tall herbs, leaves alternate; large purple yellowish or white flowers borne on 1-sided racemes; calyx 5-parted; corolla irregular; tube contracted, upper lip 2-cleft, lower lip 3-lobed, middle largest; stamens 4 didynamous; style slender; fruit a capsule; seeds numerous, roughened. About 20 species in Asia and Europe.

Digitalis purpurea L. Purple Foxglove

Biennial or annual pubescent herb with stout stem; lower leaves ovate or ovate-lanceolate, slender petioled, upper leaves smaller, sessile; flowers borne in long drooping racemes; corolla spotted.

Distribution. Native to Europe but widely naturalized in the Pacific Northwest.

Poisonous properties. The plant has long been used in medicine. For this purpose the leaves of the second year's growth are collected. The active constituents are such glucosides as digitoxin, $C_{34}H_{50}O_{10}$. The most poisonous are active, digitalin, $C_{35}H_{56}O_{14}$, digitalein, an amorphous bitter substance soluble in water, digitonin, $C_{27}H_{44}O_{14}+H_2O$ and digitophyllin $C_{32}H_{52}O_{10}$. The leaves also contain luteolin which occurs in mignonette.

In large doses digitalis is a gastro-intestinal irritant and in poisoning causes nausea. It causes the pulse to become slower, fuller and stronger and more regular. It causes stimulation of the heart muscles. In poisonous doses it is rapid, weak and irregular. The respiratory centers are unaffected except by toxic doses. The temperature is reduced by toxic doses. Dr. Winslow, in speaking of the cumulative action of digitalis and the toxic action, says:

Digitalis and strychnin are said to be cumulative in their action. Evidence is stronger in the case of the former drug than in that of the latter. By cumulative action is meant sudden transition from a therapeutic to a toxic effect. This may be due to three causes. 1. Tardy absorption. 2. Increasing susceptibility. 3. Delayed climination and accumulation of the drug in the system. The cumulative action of digitalis is chiefly due to the latter cause. It should never be administered in full medicinal doses uninterruptedly for any considerable length of time.

Toxicology.—Poisoning may occur from large single doses within 3 to 10 hours of their ingestion, and last for 16 or more hours with a fatal result; or may appear suddenly after the administration for several days of large medicinal doses (cumulative action). A minimum fatal dose for the horse is about 3 vi. of digitalis, or gr. i. ss. of Honolle's digitalin. For dogs,

3 i. of digitalis, or gr. ¼ of digitalin. The symptoms exhibited are chiefly concerned with the digestion and circulation. They consist in dulness, lassitude, loss of appetite, nausea, flatulence, diarrhoea, infrequent, full pulse (reduced 6-10 heats in the horse), and contracted pupils. There is vomiting in dogs. In fatal cases these symptoms are followed by severe colic and tympanites; rapid, feeble, dicrotic, irregular or intermittent pulse (120-140 in horses), while the heart may be heard and felt heating wildly and strongly, and a systolic blowing murmur can frequently be detected. This is due to mitral or tricuspid regurgitation caused by irregular contraction of the columnæ carnæ. The pulse is imperceptible hecause of the failure of the heart to fill the vessels. The extremities are cold, the eye is protruding, and salivation occurs. Bloody diarrhoea is very often present and the urine may be suppressed. The breathing finally becomes difficult and death ensues within a few hours, or as late as several days.

Treatment.—Evacuation of the stomach and howels. Tannic acid, as a chemical antidote, alcohol, opium, and aconite, which is the physiological antagonist in depressing the action of the heart and lowering blood tension. In addition, external heat should be applied and complete quiet and rest secured.

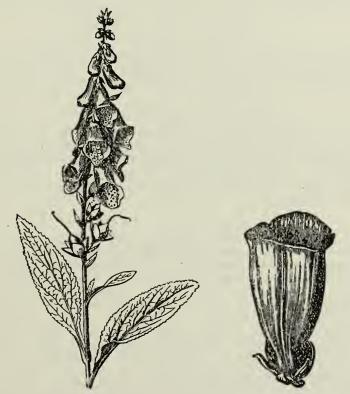


Fig. 429. Digitalis purpurea. Flowering hranch-diminished. Flower natural size. A well known medicinal and poisonous plant. (From Vesque's Traité de Botanique).

4. Gerardia (Plumier) L. Gerardia

Erect herbs or a few shrubs; leaves generally opposite and sessile or the upper alternate; flowers showy purple or yellow; calyx bell-shaped, 4-toothed or 5-lobed; corolla somewhat irregular, bell-shaped, 5-lobed, slightly 2-lipped; stamens 4, somewhat unequal; filaments pubescent; capsule many seeded. About 40 species native to America.

Gerardia tenuifolia Vahl. Slender Gerardia

A glabrous annual; leaves narrowly linear acute; calyx teeth very short acute; corolla light purple spotted ½ inch long.

Distribution. In low grounds Quebec to Minnesota, Iowa, Kansas and Louisiana.

Gerardia grandiflora Benth

A common pubescent annual or biennial 2-4 feet high; leaves ovate lanceolate coarsely toothed or cut, the lower pinnatifid; pedicels shorter than the calyx; calyx-lobes oblong or ovate; corolla longer than calyx-lobes.

Distribution. In dry woods Wisconsin, Minnesota to Texas and Tennessee. *Poisonous properties*. The first species said to be poisonous to sheep and calves. Other species probably poisonous.

5. Pedicularis (Tourn.) L. Lousewort

Perennial herbs with pinnately lobed or cleft or pinnatifid leaves; calyx tubular; corolla 2-lipped, the upper lip arched, frequently beaked at the apex; lower lip erect; stamens 4, under the upper lip, anthers transverse; capsule generally oblique; several-seeded. About 125 species mostly in the Northern Hemisphere.

Pedicularis canadensis L. Lousewort

Hairy simple stemmed plant from 6 inches-1 foot high; leaves scattered, the lowest pinnately parted; flowers in short spikes; calyx split in front, oblique; corolla greenish yellow and purple, upper lip of the corolla hooded, 2-toothed under the apex.

Distribution. In woods and prairies from eastern Canada to Florida, to Missouri and New Mexico, to Manitoba.

Pedicularis lanceolata Michx. Swamp Lousewort

An upright glabrous perennial from 1-3 feet high; leaves opposite and alternate, lanceolate or linear-lanceolate, pinnately lobed, the lower petioled; flowers in spikes; calyx 2-lobed, leafy-crested; corolla pale yellow, bearing a short truncate beak, the lower lip nearly erect; capsulc ovate, about as long as the calyx.

Distribution. In swamps from Ontario to Connecticut, Virginia, Iowa, Nebraska, Minnesota to Manitoba.

Poisonous properties. P. canadensis is said to be poisonous to sheep and P. lanceolata is also suspected. Rocky Mountain species such as P. groenlandica, P. racemosa and P. bracteosa, are frequently eaten by sheep without any ill effects. Lehmann lists three European species as poisonous, the P. palustris, P. sylvatica and P. sudetica. A decoction made from these European plants is used to destroy animal parasites. In cattle these plants cause anemia,

Dr. Lindley, in speaking of the European species, says, "they are acrid but are eaten by goats." The European P. palustris was formerly officinal and is much used in Europe as a domestic remedy. The glucoside, rhinanthin, is found in the different species of the genus. It is identical with the material found in the common Butter and Eggs (Linaria vulyaris) ($C_{64}H_{56}O_{40}$).

BIGNONIACEAE. Bignonia Family

Woody plants, trees, shrubs or woody climbers or some exotic herbs; leaves opposite or rarely alternate; flowers mostly large and showy; calyx 2-

lipped, 5-cleft or entire; corolla tubular bell-shaped, 5-lobed, somewhat irregular; stamens inserted on the corolla; some of the stamens sterile or rudimentary, inserted on the tube of the corolla, anther bearing 2 or 4; ovary usually 2-celled; fruit a 2-valved capsule; seeds flat, winged; cotyledons broad and flat.

About 500 species mostly tropical. The trumpet creeper (Tecoma radicans), native from Pennsylvania to Minnesota and southward, produces large scarlet or orange flowers, and is much cultivated as an ornamental plant. It contains narcotic principles. T. jasminoides, much cultivated in greenhouses for its pretty white, pinkish or purple flowers, is a native of Brazil. Bignonia capreolata, from Virginia to southern Illinois and southward, produces pretty orange red flowers and is cultivated southward. B. venusta is a greenhouse plant native to Brazil. The leaves of Caroba (Jacaranda procera) furnish a valuable alterative. The Newbouldia lacvis is used in dysentery.

Catalpa Scop. Catalpa

Trees or shrubs; leaves opposite or verticillate, simple, petioled; flower large in terminal panicles; calyx deeply 2-lipped; corolla bell-shaped with spreading margin; some stamens with fertile anthers 2, the others sterile or rudimentary; capsule large and slender, 2-celled; seeds numerous, winged. Seven species in Asia, North America and the West Indies.

Catalpa speciosa. Warder. Common Catalpa.

A large tree with thick bark; leaves large, heart-shaped, long, acuminate; corolla 2 inches long, white and mottled; capsule thick with numerous seeds.

Distribution. The common hardy catalpa (C. speciosa), a native to the United States from Illinois to Arkansas, is a tall tree largely planted for its wood, which is used for posts, and railroad ties. It is hardy as far north as northern Iowa.

Catalpa bignonioides Walt.

Tree with thin bark, spreading branches; leaves strongly scented and broadly ovate entire or 3-lobed acute or acuminate, deeply pubescent beneath; flowers in panicles, white mottled with yellow and purple; corolla tube, bell-shaped, the lower lobe entire; capsule rather thin walled, drooping.

Distribution. Commonly cultivated but less hardy than the preceding, native to the Gulf States.

Poisonous properties. The odor coming from the fragrant flowers is poisonous and Dr. White in his Dermatitis Venenata states that the flowers are irritating to many persons. Dr. Millspaugh, also, says that it is considered to be dangerous to inhale the odor of the flowers for a long time, which, however, is probably not generally the case. The allied caroba (Jacaranda procera) contains the bitter principle carobin. The glucoside, catalpin, comes from the bark and pod of C. bignonioides. The Oroxylon indicum contains oroxylin $C_{10}H_{14}O_{6}(OH)_{3}$.

PLANTAGINALES

This order contains but one family, the Plantaginaceae.

PLANTAGINACEAE Lindl. Plantain Family.

Mostly stemless herbs; leaves, in species with stems, opposite or alternate; flowers small, perfect, polygamous or monoecious; calyx 4-parted, persistent;

corolla 4-lobed, hypogynous; stamens 4 or 2 or only 1, inserted on the throat or tube of the corolla; ovary 1-2 celled, or falsely 3-4-celled, sessile; ovule 1, several ovules in each cavity; fruit a pyxis, circumscissile, at or below the middle or a nutlet.

3 genera, 2 native to North America, and more than 200 species, of wide distribution. None of the plants are of any economic importance. The seeds of several species (*Plantago major* and *Plantago Rugelii*) are used to feed birds. The *P. ovata* is used in France as a salad. The seeds of *P. arenaria* of Europe and *P. indica* are used for sizing in the manufacture of muslin. All of the seeds of the genus have a mucilaginous testa.



Fig. 430. Hardy Catalpa (Catalpa speciosa). 1, Panicle of flowers. 2, Longitudinal section of flower. 3, Single fruit. 4, Seed. 5, Longituinal section of seed. All one-half natural size. (M. M. Cheney in Green's Forestry in Minnesota).

RUBIALES

Leaves opposite or whorled; flowers with gamopetalous corolla, separate anthers; stamens as many as the lobes of the corolla and alternate with them

or occasionally fewer or twice as many; ovary compound, adnate to the calyx tube; ovules one or two in each cavity of the ovary. The important families are Rubiaceae, containing cinchona coffee, asperula, and galium; Adoxaceae containing a single genus Adoxa the Musk-root, A. Moschatellina; Valerianaceae containing the corn salad (Valerianella olitoria, and Valeriana officinalis), native to Europe and North America, the roots of which are used in medicine; Dipsaceae, containing fuller's teasel (Dipsacus fullonum), whose rigid chaff hooked at the end is used for carding woolen cloth, and scabious (Scabiosa maritima) frequently cultivated for ornamental purposes; and Caprifoliaceae.

Families of Rubiales

Herbs or shrubs; flowers regular; leaves with stipules. Rubiaceae. Herbs or shrubs; flowers regular or irregular; leaves without stipules. Caprifoliaceae.



Fig. 431. Teasel (Dipsacus sylvestris). A common American weed allied to Fuller's Teasel. (Millspaugh-Selby).

RUBIACEAE B. Juss. Madder Family.

Herbs, shrubs or trees, with simple opposite leaves connected by stipules, or the leaves sometimes in whorls without stipules; calyx tube adnate to the ovary; flowers regular and perfect, often dimorphic; corolla funnel-shaped, club-shaped, bell-shaped or rotate, 4-5 lobed; stamens as many as the lobes of the corolla and alternate with them; pistil with a simple or lobed style; ovary 1-10 celled; ovules one to many in a large cell; fruit various, capsule, berry or drupe; seeds small or large, the coat thin or hard; endosperm fleshy or horny.

A large order, chiefly tropical, consisting of about 350 genera and 5000 species. Only a few of them found in northern United States. Some species are abundant in southern United States, a few being weedy. Cinchona or Peruvian Bark, from which quinine is derived, is found in several species of the genus Cinchona, a tree with evergreen leaves. Quinine is derived chiefly from Chincona officinalis, which is a native of South America; C. lancifolia is native to Peru. The Cinchona is now, however, extensively cultivated in India. Its use in fevers has been recognized since its earliest introduction from the wild plants gathered in the Andes Mountains by the Indians. The Red cinchona is obtained from C. succirubra and Calisaya bark from C. Ledgeriana.

Cinchona contains a large number of alkaloids of which the following are more important: Cinchonin, $C_{19}H_{22}N_2O$, quinamin, $C_{19}H_{24}N_2O_2$, quinin, $C_{20}H_{24}N_2O_2$, hydroquinin, $C_{20}H_{26}N_2O_2$, aricin, $C_{23}H_{26}N_2O_4$. Another alkaloid belonging to this group is disinchonin, $C_{23}H_{44}N_4O_2$. Javanin, $C_{23}H_{26}N_2O_4$, occurs in Calisaya bark. The Cuprea bark (Remijia pedunculata) from the U.S. of Colombia is also used in the manufacture of quinine and contains cinchonanin, $C_{19}H_{24}N_2O$. The partridge berry (Mitchella repens) is a tonic. Gambier (Uncaria) of the East Indies is used for tanning. The root of ipecac (Psychotria Ipecacuanha) of Brazil is a systemic emetic used as a remedy in dysentery and contains emetin, $C_{15}H_{22}N_2O_5$ and cephaëlin, $C_{14}H_{20}NO_2$. Madder (Rubia tinctorum) of the Levant and Southern Europe is used for dyeing and contains a red coloring matter, alizarin. The Morinda citrifolia contains a yellow coloring principle morindin. The cape jasmine (Gardenia jasminoides) also contains a yellow coloring resembling crocin.

Coffee obtained from the Coffee arabica and other species, contains the chemical principle caffein $C_bH_{10}N_4O_2$. This is the same as thein. This substance occurs in a large number of plants including cocoa (Theobroma Cacao), cola (Cola acuminata), yopan (Ilex Cassine), maté (I. paraguensis), Sterculia platanifolia, Paullinia Cupana.

Green seeds of Coffee arabica contain 1.22 per cent of caffein or thein $C_8H_{10}N_4O_2$, the young leaves of Chinese tea 2.12 per cent. Caffeidin $C_7H_{-2}N_4O$ is obtained from caffein. Caffein causes the heart to beat more



Fig. 432. Cinchona (Chinchona lancifolia). A native of Peru and one of the species furnishing the Peruvian bark of commerce. (From Strasburger, Noll, Schenck and Schinper).

forcibly; it is a cerebral stimulant, producing wakefulness and restlessness; in lower animals it produces excitement and mania. From a toxicological point it is a spinal and muscle poison to the frog. In dogs and mammals it causes restlessness, and in dogs it produces vomiting. The minimum fatal dose according to Winslow is 1 gr. to 1 lb. of live weight.



Fig. 433. Coffee-plant (Coffee arabica). Fruiting branch. Furnishing the coffee berry of commerce. (After Faguet).



Fig. 434. Coffee-plant (Coffee arabica). 1, Flowering branch. 2, Fruit. 3, Transverse section of fruit. 4. Seeds. (After Wossidlo).

The coffee plant is a small evergreen tree native to the tropical mountain districts of Africa but now cultivated in all warm countries. It was introduced into Arabia early in the 15th century or perhaps earlier. Brazil supplies a large amount of coffee. Large amounts of coffee also come from Ceylon, Java and the Celebes. It is also grown in Puerto Rico and Cuba. Its first introduction is said to have occurred in the middle of the 16th century. The Mocha coffee comes from southwestern Arabia.

The sweet-scented bedstraw (Galium triflorum) is used in making an aromatic drink, especially in German communities in this country. It contains coumarin. In Europe the sweet woodruff (Asperula odorata) is used like G. triflorum and when added to wine, the drink is known as "Mai-trunk."

Cephalanthus L. Button Bush

Shrubs or small trees; leaves opposite or verticillate; flowers in spherical peduncled heads, white or yellow; calyx tube obpyramidal with 4 obtuse lobes; corolla tubular 4-toothed; stamens 4, inserted on the throat of the corolla; ovary 2-celled ,ovules solitary in each cavity; style thread-like, stigma capitate; fruit dry obpyramidal 1-2 seeded.

About 7 species native to America and Asia.

Cephalanthus occidentalis L. Button Bush

A shrub or small tree; leaves petioled ovate or lanceolate-oblong pointed, opposite or whorled with small petioles; flowers borne in globular head; sessile white; style longer than the corolla.

Distribution. In swamps and low grounds from Canada to Minnesota, Texas and Florida.

Poisonous, properties. The leaves contain a poisonous, bitter glucoside cephalanthin $C_{22}H_{84}O_{e}$. It has been used in medicine on account of its bitter properties.

CAPRIFOLIACEAE Vent. Honeysuckle Family.

Shrubs, trees or vines, or rarely herbs, with opposite leaves; stipules absent or present; flowers perfect, mostly cymose; calyx adnate to the ovary, 3-5 toothed or 3-5 lobed; the gamopetalous corolla with a 5-lobed limb or 2-lipped; stamens 4-5, inserted on the tube of the corolla and alternate with its lobes; ovary 2-5 celled; style slender; stigma capitate; fruit a berry, drupe or pod; seeds with a membranous or hard coat.

About 275 species. Generally found in the northern hemisphere. The plants of this order are of small economic importance. Several are used in medicine, as the feverwort (*Triosteum perfoliatum*).

Many members of the order are used for ornamental purposes. The most important are members of the genus Lonicera. Of the native species, the trumpet honeysuckle (Lonicera sempervirens) is widely cultivated, also Sullivanti's honeysuckle (L. Sullivantii), Fraser's honeysuckle, (L. flava), and the western honeysuckle (L. involucrata). Some of the Loniceras are possibly poisonous. Of the European and Asiatic species, the L. tatarica, L. japonica, L. fragrantissima and L. Periclymenum are cultivated. The elders (Sambucus canadensis and S. racemosa) are likewise cultivated in the North. The former is often weedy. Several species of the snowberries, like the wolfberry (Symphoricarpos occidentalis) and the snowberry proper (S. racemosus) are native to the northern states. The Indian currant (S. orbiculatus) is sometimes weedy in Iowa and Missouri. Suckers used in Arkansas for making

baskets. The Linnaea borealis or twin-flower is native in the cool damp woods of the North.

Several of the viburnums are cultivated. The hobblebush (V. alnifolium), a native from Ontario and southward, is frequently cultivated in the East. The cranberry-tree or guelder rose (V. Opulus) found along streams from New Brunswick to northeastern Iowa, is cultivated both in its native and cultivated forms. The well-known snowball is a cultivated form of the cranberry tree. The fruit of this plant is used in the North. The root of horse gentian (Triosteum perfoliatum) has a bitter taste and is used as a cathartic. The flowers of the elder berry (Sambucus canadensis) are sudorific. The bark of Viburnum prunifolium, the black haw, is officinal, an antispasmodic, nervine and an astringent. It is also used as an uterine sedative, contains viburnin, oxalic acid, tannic acid, etc. The bark of V. Opulus is said to be antispasmodic. Xylostein occurs in Lonicera Xylosteum. According to Greshoff the leaf of Viburnum macrophyllum and of Symphoricarpos mollis contain saponin.

Genera of Caprifoliaceae

| Flowers in compound cymes; corolla rotate | . Sambucus |
|---|-------------|
| Flowers not in cymes, tubular to campanulate. | |
| Erect perennial herbs | Triosteum. |
| ShrubsSymp | horicarpos. |

Triosteum L. Horse Gentian. Feverwort

Coarse hairy perennial herbs with simple stems; leaves connate perfoliate or sessile; flowers axillary perfect sessile; calyx tube ovoid with a 5-lobed limb, persistent; corolla tubular, gibbous at the base, 5-lobed; stamens 5, anthers linear; ovary 3-5 celled; ovule 1 in each cavity; style filiform; fruit a dry drupe orange or red, enclosing 2-3 or rarely more 1 seeded nutlets, embryo minute.

Triosteum perfoliatum L. Feverwort. Wild Coffee

A soft hairy perennial 2-4 feet high; leaves oval, abruptly narrowed below, downy beneath; flowers brownish purple, clustered; corolla purplish; fruit orange in color.

Distribution. In rich woods New York to Minnesota, Kansas and Alabama. Poisonous properties. Some species of the genus were used by the Indians as a cure of fevers and early practitioners in this country used the root as an emetic. In early days, the berries of this plant were used as a substitute for coffee. The physiological action of the plant is to produce vomiting. It has a bitter nauseous taste.

Sambucus (Tourn.) L. Elder

Shrubs, trees or occasionally herbs; leaves opposite, pinnate; flowers small in compound cymes; calyx-lobes minute or obsolete; corolla rotate or somewhat campanulate, regular with a 5-cleft wing; stamens 5, inserted at the base of the corolla; stigmas 3; ovary 3-5-celled; ovules 1 in each cavity; fruit a berry-like drupe with 3-5 1-seeded nutlets; endosperm fleshy.

About 20 species of wide distribution.

Sambucus canadensis L. Common Elder

A shrub from 5-10 feet high, wood with large lenticels and large pith; leaflets 5-11 ovate or oval acuminate or acute, short stalked, smooth above, sharply serrate; flowers white.

Distribution. From Canada to Manitoba, Kansas, Texas to Florida.

Poisonous properties. In regard to the poisonous properties Dr. Rusby says: The common black elder or Sambucus canadensis L., a plant very common throughout the entire eastern and central United States, and represented by other species, apparently with similar properties, upon the Pacific Coast and in the old world, has dangerous properties which have remained unrecognized, or, to say the least, very obscure, to the present time.

Of the last mentioned, Dr. Robert Christian reports in the Edinburgh Med-

ical and Surgical Journal, 1830, page 73, as follows:

Two boys in the vicinity of Edinburg encountered a clump of the S. Ebulus, and one of them ate freely of the flowers, the other of the leaves. The boy who had caten the leaves was attacked with enteritis, the abdomen at length becoming so sore that it could scarcely be touched. There was continuous vomiting, the matter containing blood. Obstinate constipation existed throughout. The boy was saved by vigorous treatment. The one who had eaten the flowers suffered considerably, and for a considerable time, from vertigo with some headache, but the symptoms were not very serious.

Dr. Christian observed that both the berries and the flowers were known to kill fowls which fed upon them and that when berries were freely eaten they often caused giddiness. He also quotes a report of a case of a woman who dressed the shoots with vinegar and ate them as a salad, and who was promptly seized with violent purging, forty times in two days, coma resulting on the third day. Of our own species, S. canadensis, Dr. Johnson states that the bark and the root are actively cathartic and hydragogue when freely used. There is little doubt that he refers in this instance to the bark and the root in the green condition, since it is well-known that the properties become much less active upon drying and keeping.

Our most direct evidence bearing upon the poisonous character of the elder-berry root rests upon a case which occurred in the spring of 1894, at the Institution of Mercy, a Roman Catholic institution for children at Tarrytown, on the Hudson, and which attracted a great deal of attention at the time in the public press. The grounds of this institution were comparatively new, and ditching and fencing were still in progress at the time stated. A workman in digging a drain, uncovered a large number of roots to which the children took a fancy and which they began eating. Within a very few minutes, and while still engaged in eating, a large number of the boys were scized with convulsions and several of them died. One of them had the remainder of the root, the marks of his teeth upon it, still clutched in his hand after death. The symptoms correspond in most features to those of the Cicuta poisoning above described and to that agent the accident was ascribed in the public press. Several months later I visited the institution in company with Mr. Frederick V. Coville, the botanist of the United States Department of Agriculture and Prof. Edward L. Greene, Professor of Botany, in the Catholic University at Washington. At this time, and subsequently through correspondence, a pretty thorough investigation of the case was made. We found that it was not a locality where Cicuta would be apt to grow and no evidence existed that any had grown there. Three poisonous plants grew upon the spot, viz., the locust, poke-berry and elder. The workman who had dug the drain, the surviving boys and the Sisters in attendance were positive that it was the elder root which had occasioned the poisoning. They did not know the name of the plant, and had accepted the statements of the papers that it was Cicuta; but they positively identified it by its appearance and by the young purple shoots and compound leaves which they had observed carefully while still attached to the pieces of root which had been taken from the hands of the boys poisoned. Their story was so clear, connected and positive that it was difficult to doubt that the elder root was the poisoning agent. Furthermore the locust would not have produced the symptoms that were observed, and the poke should have at once been distinguished by even a casual observer. Nevertheless, since the root was described as "like a carrot or parsnip," and since the symptoms in some respects resembled those of Pokeroot poisoning, the question cannot be regarded as settled beyond a doubt. In the case of so large a number of victims it is even possible that both of the roots were concerned. The attending physician, Dr. Luke Fleming, does not believe that the poisoning was caused by Elder. The active constituent of the elder is not known farther than that a report has isolated coniin from the twigs and leaves of the related European species, S. nigra. This would, of course, explain the very similar symptoms to those of Cicuta poisoning. The chemistry of the plant is now receiving thorough investigation in the division of pharmacology in the United States Department of Agriculture.

In Part I attention was called to Treub's theory in regard to the role of hydrocyanic acid in plants. Prof. Treub in a recent paper1 reiterates his former conclusions that it performs some part in the products of assimilation. It was found that the amount of hydrocyanic acid in plants of Sorghum increases during the day because of its relation to the products of assimilation of carbon. It had previously been shown in the case of Pangium edule and Phaseolus lunatus that light plays no part in the formation of this substance, except as it favors photosynthesis. The same results have now been obtained with Prunus javanica, Passiflora foetida, and some other plants. The results of the investigation with these plants show a direct proportion between the formulation of hydrocyanic acid and the function of the chlorophyll. The amount of acid is usually greatest in the young leaves and gradually diminishes as the leaves grow older. Leaves about to fall contain very little hydrocyanic acid. Sambucus nigra, according to Guignard is one of the exceptions to the rule, and Treub has also found this to be true for Indigofera galegoides. Hydrocyanic acid is probably the first recognizable product of the assimilation of nitrogen "and perhaps the first organic nitrogen compound formed." The amounts of the acid in the plants could be increased and decreased in proportion to the amount of nitrate used. Ravenna and Peli think that the nitrates are necessary for the formation of the acid. Treub agrees with this and adds that dextrose is especially essential. The acid probably occurs in the form of a glucoside and is liberated by an enzyme or by boiling water.

The investigations of Em. Bourquelot and Em. Danjon² with the glucosides found in various plants show that they are not identical. In Sambucus nigra they find sambunigrin. They also studied the character of glucosides from which hydrocyanic acid is derived in S. racemosa and S. Ebulus.

The flowers have long been used in domestic practice. The physiological action recorded for the drug by Dr. Millspaugh are as follows:

Dr. Ubelacker's experiments with from 20 to 50 drops of the tincture gave the following symptoms of physical disturbance: Drawing in the head, with anxious dread; flushed and blotched face; dryness and sensation of swelling of the mucous membranes of the mouth, pharynx, and trachia; frequent and profuse flow of clear urine; heaviness and constriction of the chest; palpitation of the heart; pulse rose to 100, and remained until perspiration ensues; sharp, darting rheumatic pains in the hands and feet; exhaustion and profuse perspiration, which relieved all the symptoms.

Prof. Hyams states that the young buds of the American elder are especially poisonous.

The European Sambucus nigra contains the alkaloid sambucin; according to Sanctis the same plant contains contin. Sambunigrin has also been isolated and hydrocyanic acid is known to be present.

Symphoricarpos (Dill.) Ludwig. Snowberry

Low, branching shrubs; leaves oval, short petioled entire or wavy-toothed, downy underneath; flowers white tinged with purple; calyx-tube nearly globular, teeth short; corolla bell-shaped 4-5 lobed; stamens 4-5 inserted in corolla; ovary 4-celled, 2 with a fertile ovule; fruit a berry 4-celled and 2-seeded; embryo minute.

About 10 species of North America. One species frequently cultivated for ornamental purposes. The wolf-berry (S. occidentalis) northward is com
1 Nouvelles recherches. Ann. Jard. Bot. Buitenzorg. II. 8:84-118. 1910. See R. Catlin

Rose, Bot. Gazette 50:156.

2 Soc. Biol. Paris. 7. July, 1905. 1. c. Oct. 9, 1905. Jour. de Pharm. et de Chimie, Aug. 16, and Sept. 1, 1905. Compt. rend. Acad. Sci. Paris, July 3, 1905.

mon in the Rocky Mountains and frequently cultivated. The snowberry (S. racemosus) is common in rocky woods, and abundant along river course from Minnesota to Arkansas and westward.

Symphoricarpos orbiculatus Moench. Indian Currant. Coralberry

A shrub 2-4 feet high, purplish, usually pubescent, branches; leaves oval or ovate entire or undulate, nearly glabrous above, pubescent underneath; flowers in short axillary clusters; corolla bell-shaped sparingly bearded, pinkish, stamens included; fruit a purplish berry.

Distribution. Rocky woods and along streams; from New Jersey, Illinois, Southern Iowa, South Dakota, Nebraska, Texas to Georgia.

Poisonous properties. It is suspected of being poisonous, but there is no direct evidence to support this view.

CAMPANULATAE

Herbs or rarely shrubs; corolla gamopetalous; petals occasionally separate; stamens as many as the lobes of the corolla; anthers united; ovary inferior. It contains the families Cucurbitaceae, Candolleaceae (mostly Australian), and Compositae, this last order including Cichoriaceae, Compositae and Ambrosiaceae of some authors.

Families of Campanulatae

Herbs or rarely shrubs; flowers in an involucrate bead..........Compositae. Flowers not in involucrate heads.



Fig. 435. Water melon (Citrullus vulgaris). (W. S. Dudgeon).

CUCURBITACEAE B. Juss. Gourd Family

Herbaceous vines, usually with tendrils; leaves alternate, petioled, palmately lobed or dissected; flowers dioecious, monoecious or rarely perfect; calyx tube adnate to the ovary, 5-lobed; petals usually 5, inserted on the limb of the calyx; stamens 1-3, 2 of them with 2-celled anthers, the other with a 1-celled anther; filaments short, frequently monadelphous; ovary 1-3-celled; stigmas 2 or 3; fruit indehiscent or rarely dehiscent; seeds flat in the large embryo, exalbuminous.

About 650 species, mainly in tropical regions. A few of the species are medicinal. The squirting cucumber (*Ecballium Elaterium*) is a fleshy decumbent herb used for making elaterium, a powerful hydragogue cathartic and an irritant poison. Its poisonous nature was known to Pliny. It contains elaterin, $C_{20}H_{28}O_{5}$. E. officinale contains prophetin, another glucoside. The colocynth (*Citrullus Colocynthis*) a slender scabrous plant with perennial roots, is native to the dry regions of the Old World, Palestine and North Africa. Its gourd is about the size of an orange and used as a purgative, while the seeds are roasted and boiled and used as food by some of the tribes of the Saharah. Tea made from this fruit is used by the people of the Nile to smear their water bags to prevent camels from cutting them. It contains colocynthin,



Fig. 436. Bryony (Bryonia dioica). A a branch with flowers; b, female flowers; c, male flowers; d, stamens; e, fruits; f, section of fruit. (After Strasburger, Noll, Schenck and Schimper).

C₉₈H₁₄₀O₄₃. The plant is intensely bitter. The towel or sponge gourd (Luffa aegyptiaca), a native of Egypt, produces a fruit one foot or more long, filled with a spongy fiber, which when the outer part is removed is used to rub the skin, and for many other domestic purposes. The fruits of many plants of the family are economic. The nara (Acanthosicyos horrida) in Southern Angola is used as food and has medicinal virtues. The chayote (Sechium edule) is cultivated in the West Indies for its fruit. The green and ripe fruit



Fig. 437. Colocynth (Citrullus colocynithis). An intensely bitter plant of economic importance. (After Faguet).

of the cucumber (Cucumis sativus) native to India, has long been used for food, especially for pickles, and the West Indian gherkin (C. Anguria) is also cultivated for the same purpose. According to Greshoff the foliage of Cucumis metuliferus contains saponin. He also states that he found saponin in the seeds of Lagenaria vulgaris, and Cucurbita maxima. The C, myriocarpus contains the toxic alkaloid myriocarpin. The musk melon (Cucumis Melo), a native to British India, is now widely cultivated. Sugar and nutmeg melons are well known everywhere in North America. The water melon (Citrullus vulgaris), native to tropical Africa, where large areas of wild plants occur, has long been cultivated in Mediterranean countries, and is well known everywhere in North and South America. The citron is a form of the water melon. The common pumpkin (Cucurbita Pepo) is native to southwestern North America, Arizona and Mexico. It is used for stock food and for culinary purposes. The nest-egg gourd, bush scalloped squash, crookneck squash and common pumpkin are all from the same species. The seeds of pumpkin and squash are used in North Africa and Egypt much as peanuts are in this country and have taenifuge properties. The winter squash (C. maxima) is probably also of

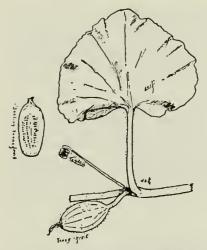


Fig. 438. Muskmelon (Cucumis Melo). A part of a plant of the common melon. (W. S. Dudgeon).

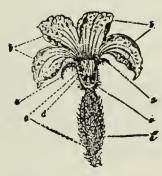


Fig. 439. Pistillate flower of cucumber (Cncumis). a, sepal; b, petal; c, pistil; d, stigma; e, style; f, ovary. (Pieters, U. S. Dept. Agr.).

American origin. The hubbard squash is a well known representative of the species and is used in various food preparations. Other species like cushaw (C. moschata) are also cultivated. The Missouri gourd (C. foetidissima), with a large root, sometimes a foot in diameter, occurs from Nebraska to Texas. The wild cucumber or wild balsam apple (Echinocystis lobata) is frequently used as a climber and is sometimes weedy. The star cucumber (Sicyos augulatus) is used in a similar way. The cocoon antidote, (Fevillea cordifolia), a native of Jamaica, has seeds which are used as a cure for snake bites and as an antidote against Entada scandens. They also contain an oil. The seeds of Tefaria pedato, a native of the East coast of Africa and Zanzibar, contain an excellent oil. The bryony of Europe (Bryonia dioica) is a climbing plant producing a pretty colored fruit. It is a drastic purgative and is poisonous. It contains the alkaloid bryonicin C10H17NO2, and the root of Bryonia alba contains the glucoside bryonin C₈₂H₉₃O₃₁. Dr. Halsted states that a friend of his has been repeatedly poisoned by handling the star cucumber (Sicyos angulata). Friedberger and Fröhner state that poisoning has occurred from Cucurbita Pepo, causing symptoms of dullness and in some cases the animals showed excitement. Major Kirtikar says that the pulp of a cucurbitaceous plant of India, Trichosanthes palmata, a perennial herb, is used in India to poison cattle, and that the T. cucumerina also of that country is an emetic and a drastic purgative. Echinocystis macrocarpa, according to Trimble and Sayre contains a glucoside. The marimbo or dipper gourd (Lagenaria vulgaris) yields gourds which are edible when small.

LOBELIACEAE. Lobelia Family

Herbs with acrid milky juice; leaves alternate; flowers perfect, irregular, 5-lobed; gamopetalous corolla; stamens 5, free from the corolla, united into a tube; stamens monadelphous and syngenesious; flowers proterandrous, the stigma of the single style often fringed with hairs; fruit a capsule with numerous small seeds; embryo minute and straight.

About 600 species of wide distribution, comparatively few in northern United States. Some of the tropical species shrubby. Several species are medicinal, among them Indian tobacco (Lobelia inflata). The Lobelia erinus of the Cape of Good Hope is frequently cultivated in the conservatories and in gardens. It has small azure blue flowers. It has escaped on the Pacific Coast.

Lobelia (Plummer) L. Lobelia

Herbs or occasionally shrubs with alternate or radical leaves; flowers racemose or spicate; calyx 5-cleft with a short tube; corolla irregular, with a straight tube split down on one side, the upper lip of 2 erect lobes, the lower lip spreading and 3 cleft; stamens 5, free from the corolla tube, monadelphous; two of the anthers or all of them bearded at the top; ovary 2-celled; fruit a 2-celled pod, many-seeded.

About 200 species, of wide distribution. Some 25 species native to the United States.

Lobelia inflata L. Indian tobacco

A pubescent or hirsute, much branched annual from 1-2 feet high; leaves dentate or denticulate, the lower larger, the upper small, bract-like, but longer than the pedicels of the flower; flowers pale blue; calyx tube ovoid; capsule ovoid, inflated.

Distribution. In fields, especially clay soils, from Labrador to Georgia, Tennessee, Arkansas, Kansas, Iowa to Northwest Territory.

Poisonous properties. It is used medicinally for largngitis and spasmodic asthma. In full doses it produces nausea, vomiting and great prostration; in overdoses it produces prostration, stupor, coma, convulsions and death.

We quote from Dr. Millspaugh in regard to poisoning:

Tbanks to much reckless prescribing by many so-called Botanic physicians, and to murderous intent; as well as to experimentation and careful provings, the action of this drug is pretty thoroughly known. Lobelia in large doses is a decided narcotic poison, producing effects on animals generally, bearing great similitude to somewhat smaller doses of tobacco. and lobelina in like manner to nicotia. Its principal sphere of action seems to be upon the pneumogastric nerve, and it is to the organs supplied by this nerve that its toxic symptoms are malnly due, and its "physiological" cures of pertussis, spasmodic asthma, croup and gastralgia gained. Its second action in importance is that of causing general muscular relaxation, and under this it records its cures of strangulated hernia (hy enemata), tetanic spasms, convulsions, bysteria, and mayhap, hydropbobia. Its third action is upon mucous surfaces and secretory glands, increasing their secretions.

The prominent symptoms of its action are: great dejection, exhaustion, and mental depression, even to insensibility and loss of consciousness; nausea and vertigo; contraction of the pupil; profuse clammy salivation; dryness and prickling in the throat; pressure in the œsophagus with a sensation of vermicular motion, most strongly, however, in the larynx and epigastrium; sensation as of a lump in the throat; incessant and violent nausea, with pain, heat, and oppression of the respiratory tract; vomiting, followed by great prostration; violent and painful cardiac constriction; griping and drawing abdominal pains; increased urine, easily decomposing and depositing much uric acid; violent racking paroxysmal cough with ropy expectoration; small irregular slow pulse; general weakness and oppression, more marked in the thorax; violent spasmodic pains, with paralytic feeling, especially in the left arm; weariness of the limhs, with cramps in the gastroenem; and sensation of chill and fever. Death is usually preceded by insensibility and convulsions.

It contains lobelic acid, lobelacrin, inflatin and the alkaloid lobelin $C_{12}H_{23}NO_2$ has been isolated which according to Lloyd is a powerful emetic. Lobelia nicotianaefolia of India and L. purpurascens contain the same alkaloids. According to the late Baron Ferdinand von Müller the L. Breynii of Australia and other species are poisonous.

Lobelia siphilitica L. Great Lobelia

A somewhat hairy stout, perennial herb from 1-3 feet high; leaves thin, acute or acuminate at the apex, dentate or crenate-dentate, sessile or the lower petioled; flowers large, spicate, racemose, leafy bracts; calyx hirsute; corolla bright blue or occasionally white.

Distribution. In moist soil near springs and in marshes from New England to South Dakota, Kansas, Louisiana and Georgia.

Poisonous properties. It is suspected of being poisonous. Johnson in his manual says of the action of the species of Lobelia:

In full doses lobelia produces severe nausea, obstinate vomiting, and great prostration. In overdoses the prostration becomes extreme, there is failure of voluntary motion, followed by stupor, coma, and not infrequently convulsions and death. Though formerly much used for emetic effect by empirics, dangerous effects were so often produced that it is now seldom employed in this manner. It is chiefly employed in spasmodic affections of the air-passages, as spasmodic laryngitis and spasmodic asthma. In the latter disease it often produces the happlest effects.

The great lobelia is probably not as poisonous as L. inflata to which the above remarks chiefly apply.

Lobelia cardinalis L. Cardinal-flower

A tall smooth or slightly pubescent perennial 2-4 feet high; leaves thin, oblong, lanceolate, smooth or slightly pubescent, crenulate; flowers racemose, bright scarlet or red.

Distribution. In moist soil, usually alluvial bottoms, from New Brunswick to Manitoba, Kansas, Texas, to Florida.

Poisonous properties. Reported as poisonous.

Lobelia spicata Lam. Spiked Lobelia

A perennial or biennial, smooth or pubescent herb; leaves smooth or minutely pubescent; leaves thickish, the lower obovate or spatulate, the upper linear or club-shaped bracts, entire or dentate or crenulate; flowers in a racemose spike, pale blue; calyx tube short, obconical or nearly hemispherical.

Distribution. Prairies or dry sandy soil. From Nova Scotia to Manitoba, Louisiana to North Carolina.

Poisonous properties. Reported as poisonous.

Compositat Adans. Thistle Family

Herbs or rarely shrubs; flowers borne in a close head on the receptacle, surrounded by an involucre of a few or many bracts; anthers usually united into a tube, syngenesious, sometimes caudate; calyx adnate to the ovary; limb crowning the summit in the form of capillary or plumose bristles or chaff called the pappus; corolla tubular or strap-shaped, when tubular, usually 5-lobed; ligulate or bilabiate in one small division of the family; the flowers of a head may be all alike when they are called homogamous; or of two kinds, heterogamous; bracts or scales on the receptacle are often present; flowers inside of the rays are disk flowers and a flower without rays is said to be discoid; stamens 5 or rarely 4; style 2-cleft at the apex or in sterile flowers usually entire; fruit a dry, indehiscent achenium containing a single seed without endosperm.

A large family consisting of 840 genera and 13,000 species, found in all parts of the world. This is the largest order of flowering plants. Sometimes it is divided into the families Cichoriaceae, Ambrosiaceae and Compositae.

A few of the plants of the family are medicinal. Inulin is obtained from

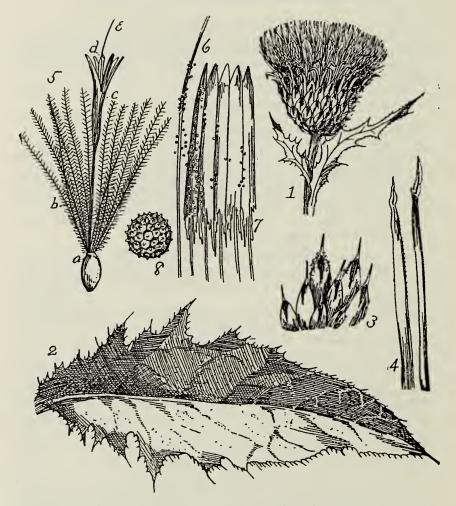


Fig. 439a. Illustrating structure of Compositae. Thistle (Cirsium altissimum). 1, head; 2, leaf; 3, outer bracts of head; 4, inner bracts; 5, single flower; a, achene; b, pappus; c, tubular corolla with 5 lobes; d, anthers; e, style. 6, style enlarged with two stigmas and pollen grains on style; 7, syngenesious anthers cut lengthwise to show pollen grains and tailed appendages; 8, single pollen grain. (Charlotte M. King).

the Inula Helenium, native to Europe and occasionally naturalized in the northern states. It is a mild tonic and contains inulin $C_6H_{10}O_5$, helenin and a volatile oil. The pellitory root (Anacyclus Pyrethrum) contains pyrethrin with a pungent taste, which, according to Dunstan, is apparently identical with piperovatin C_{16} II₂₁NO₂, used for toothache; the flowers of Roman chamomile (Anthemis nobilis) are used as a tonic and stomachic. However, German chamomile (Matricaria Chamomilla) is sometimes substituted for the preceding and contains anthemidin and a deep-blue volatile oil. Santonica, a species of wormwood, Artemisia Cina, contains santonin $C_{16}H_{18}O_3$, and cineral $C_{10}II_{18}O_6$, is

found in the volatile oil, an anthelmintic. Artemisia maritima, A. pontica, A. Absinthium, A. biennis and A. Abrotanum are also used for the dislodgement of worms. The latter contains the alkaloid abrotanin $C_{21}H_{22}H_2O$. Sage brush (Artemisia tridentata) and other species produce sneezing.

Wormwood (Artemisia Absinthium) is a stimulant and tonic; the volatile oil produces cerebral disturbances and enters into the familiar composition of absinthc, made by the French, and contains absinthin C₁₅H₂₀O₄. Arnica root (Arnica montana) native to arctic Asia and America, is used as a popular remedy for chilblains and bruises and contains the bitter principle arnicin C20 H₃₀O₄. The tincture is liable to produce a form of dermatitis. lion root (Taraxacum officinale) is used as a mild laxative and tonic and contains a bitter principle taraxacin, and taraxaccrin C,H,3O. Lactucarium, the milky juice from several species of the genus Lactuca occurs in lettuce, in which is also found lactucopicrin, a bitter acrid substance, and lactucol C21 H34O. Colt's foot (Tussilago Farfara), a bitter astringent containing much mucilage, is used for asthma. The costus (Saussurea Lappa) produces flowers with thistle-like heads and large roots, the latter of which are used as a perfume and an incense; according to Kraemer, it contains a ketone. The musk tree (Olearia argophylla) of Tasmania, whose leaves emit a musk-like odor, grows to a height of 20 feet and is often 1 foot in diameter. The wood takes a nice polish. Other trees of the order are found in the genus Senecio (Senecio Forsteri). The genus Baccharis, found along the sea coast, is shrubby. The B. cordifolia contains baccharin, an alkaloid poisonous to sheep. It is the Mio Mio of South America.

The button snake root (Liatris spicata) has been used as a remedy for snake bites, but it probably contains no antidotal properties whatever. Boneset (Eupatorium perfoliatum) and other species much used in domestic medicines for colds, and in large doses are emetic. They are tonics, emetics, cathartics, and diaphoretics containing the bitter glucoside eupatorin. Horseweed (Erigeron canadensis) and other species are used as tonics and astringents.

Golden-rod (Solidago odora) is used to relieve colic, and gumwced (Grindelia squarrosa) is beneficial in catarrhal affections. It is said to contain an alkaloid known as grindelin. Madia oil is obtained from tarweed (Madia sativa). The niger seed, the fruit of Guizotia abyssinica, is an important source of oil in Abyssinia and India.

The root of the burdock (Arctinm Lappa), used by the laity as a remedy in skin diseases, contains a bitter glucoside, lappin. Chicory (Cichorium Intybus) is used to increase the appetite and to aid digestion. Rattlesnake weed (Hieracium venosum) is a popular antidote to the bites of poisonous snakes. Rattlesnake root (Prenanthes alba) is used as a remedy for toothache. The ragweeds (Ambrosia artemisiifolia and A. trifida) are stimulants and astringents, the larger weed being also supposed to cause hay fever.

Dunbar has demonstrated that the producing cause of hay fever may be pollen, and that pollen of all grasses, lillies of the valley, asters, and certain other plants may produce an irritation similar to that accompanying hay fever. He isolated an active principle which is believed to be a tox-albumin. A very interesting account of his experiments is given by Rochussen in the twenty-sixth volume of the Pharmaceutical Review.

According to Maiden, the Helichrysum apiculatum of Australia causes death from irritation and from the formation of hair balls. The African mari-

gold is poisonous to stock and the may weed (Anthemis Cotula), when applied to the surface, causes vesication. The European prickly lettuce (Lactuca virosa) contains a bitter principle, hyoscyamin; the prepared milky juice is called Lactucarium. It is listed as poisonous by Lehmann; a small amount of hyoscyamin also occurs in the cultivated lettuce.

The seeds of the sunflower (Helianthus annuus) are said to be diuretic. The oil cake from these seeds is used as stock food. Sneeze weed (Helenium autumnale) is used by the Indians to produce sneezing; a decoction made from it is used as a tonic. The mayweed (Anthemis Cotula) acts like chamomile, and is used as a tonic and stimulant in colic; when applied to the skin, it causes vesication. Yarrow (Achillea Millefolium), a stimulant and tonic, contains achillein, $C_{20}H_{38}N_2O_{15}$. The oil of tansy (Tanacetum vulgare) is used as an abortifacient, in many cases with fatal results. It contains the substance thujone, found in Thuja occidentalis. This has commonly been called tanacetin $C_{11}H_{16}O_4$, and is identical with absinthol obtained from absinthium, and with salviol from salvia. The Cnicus benedictus contains cnicin $C_{20}H_{37}O_{10}$.

Of the many cultivated plants of this order used for ornamental purposes, the best known in the northern states are probably the bachelor's button (Centaurea Cyanus), a native of Europe, ageratum (Ageratum conyzoides), Chinese aster (Callistephus hortensis), garden daisy (Bellis perennis), and dahlia (Dahlia variabilis), native to Mexico, running into many varieties. The last named produces thickened roots, that contain a great deal of inulin, CoH100s. Among the species of the order which are common in the gardens are the zinnia (Zinnia elegans), golden glow (Rudbeckia laciniata), sunflower (Helianthus annuus), coreopsis (Coreopsis tinctoria), gaillardia (Gaillardia pulchella), dusty miller or cineraria (Senecio Cineraria), common cineraria (S. cruentus) from the Teneriffe, and the purple ragwort (S. elegans) from the Cape of Good Hope, yarrow (Achillea Ptarmica), whiteweed (Chrysauthemum Parthenium), several species of the genus chrysanthemum from Japan (C. sinense and C. indicum), Marguerite (C. frutescens) and summer chrysanthemum (C. coronarium), marigold (Tagetes erecta), the pot marigold (Calendula officinalis), cosmos (Cosmos bipinnatus) of Mexico. The marigold (Calendula officinalis) cultivated in country gardens contains calendulin CoH10O5.

Of the economic plants, the following are the more important: the cultivated lettuce (Lactuca sativa), probably native to Asia, slightly narcotic, comprising many varieties; the safflower or saffron (Carthamus tinctorius), native to Egypt, used for dyeing; marigold flowers (Calendula officinalis) sometimes used as an adulterant for saffron; wormwood (Artemisia Absinthium), used for making absinthe; tarragon (Artemisia Dracunculus) used as a pot-herb and in making vinegar, a native of Asia, but cultivated now in Holland and England; the Jerusalem artichoke (Helianthus tuberosus), with thickened roots, cultivated as food for hogs and stock, native to North America; the great sunflower (Helianthus annuus), the seeds of which are eaten in Russia, and from which an oil is obtained; the chicory (Cichorum Intybus); the burdock (Arctium minus), cultivated for its thick root which is sometimes dried and mixed with coffee and also used as a forage plant; the endive (Cichorum endivia) cultivated as a winter salad plant; the cardoon (Cynara Cardunculus), also known as the European artichoke, used as a vegetable, the thickened scales being used as food; salsify (Tragopogon porrifolius) a vegetable, and the dandelion (Taraxacum officinale) used like lettuce.

The Compositae are divided into two sub-families: the Tubuliflorae with corolla tubular in all the perfect flowers, 5-lobed, rarely 3-4-lobed, ligulate only in the marginal flowers, called the ray flowers, which are absent in some species; the Liguliflorae with corolla ligulate in all the flowers of the head and all the flowers perfect; herbs with milky juices and alternate leaves: The former contains the tribes Vernonieae, with the large genus Vernonia of 450 species mostly of the tropics; the Eupatorieae, containing Eupatorium; Astereae, containing Solidago, Aster, Erigeron, Grindelia, Bigelovia; Inuleae, containing Inula, Antennaria; Heliantheae, containing Silphium, Parthenium, Helianthus, Bidens, Coreopsis; the Helenieae with Helenium, Actinella; the Anthemideae with Anthemis, Achillea, Tanacetum; the Senecioneae with Senecio, Tussilago; the Cynareae with Arctium, Cnicus, Cirsium and Centaurea. The Liguliflorae contain but one tribe, the Cichorieae, which contains the genera Taraxacum, Cichorium, Lactuca, Hieracium, Sonchus and Scorsonera.

Genera of Compositae

Corolla ligulate in all of the flowers of the head; flowers perfect.

| LIGULIFLORAE CICHORIEAE |
|---|
| Pappus none |
| Pappus composed of capillary bristles. |
| Flowers yellow, achenes not beaked |
| Flowers yellow, purplish or cream colored. |
| Achenes beaked |
| Achenes not beaked4. Lygodesmia |
| Corolla tubular in perfect flowers, 5 or rarely 3 or 4-lobed; lingulate only in |
| the marginal flowers. TUBULIFLORAE |
| Stamens distinct or nearly so. Ambrosieae |
| Staminate and pistillate flowers in the same head |
| Staminate and pistillate flowers in a separate head. |
| Involucre of pistillate heads with several tubercles6. Ambrosia |
| Involucral bracts of pistillate flowers forming a bur7. Xanthium |
| Stamens generally united by their anthers into a tube around the style. |
| Anthers not tailed. |
| Style branches thickened upward, papillose. EUPATORIEAE. |
| Achenes 3-5 angled, flowers discoid8. Eupatorium |
| Achenes 8-10, ribbed or striate. |
| Bracts of involucre in several series9. Liatris |
| Bracts of involucre in 2 or 3 series |
| Style branches of perfect flowers flattened with terminal appendages. ASTREAR |
| pendages. ASTEREAE. Ray flowers yellow. |
| Pappus of scales or awns |
| Pappus of numerous capillary bristles12. Solidago |
| Ray flowers not yellow, bracts nearly equal14. Erigeron |
| Bracts in several unequal series |
| Style branches truncate or hairy appendaged. HELIANTHEAE |
| Pappus wanting or minute crown |
| Pappus of 2 persistent downwardly barbed awns or tubes |
| 16. Bidens |
| |

| Style branches truncate or with hairy tips. | Helenieae. |
|--|------------------------|
| Bracts of the involucre spreading. Receptacle naked; bracts of involucre sp | reading or reflexed at |
| maturity | |
| Bracts of the involucre united | 18. Dysodia |
| Style branches mostly truncate with brush hairs | on the tip. |
| | Anthemideae. |
| Pappus of short scales or a crown, | |
| Scales of involucre scarious and imbricated. | |
| Receptacle chaffy. | |
| Heads small; involucre obovoid or | campanulate; achenes |
| flattened | |
| Heads large; achenes terete | 20. Anthemis |
| Heads solitary or corymbose. | • |
| Receptacles not chaffy. | |
| Ray flowers usually present, conspic | |
| | |
| Ray flowers inconspicuous | |
| Heads small spicate or racemose par | |
| Style branches truncate or triangular with brush | hairs. |
| Heads radiate or discoid. | |
| Pappus of capillary bristles. | Senecionideae. |
| Heads showy, leaves oppositc | |
| Heads usually showy, leaves alterna | |
| Style branches short or united, anthers caudate. | |
| Achenes basi-fixed | |
| Involueral bracts pointed or prickly. | 07 01 1 |
| Receptacle densely bristly | |
| Receptacle honey-combed | 29. Silybum |
| Achenes attached laterally. | 20 0 |
| Pappus of short scales | |
| | |

LIGULIFLORAE

Herbs with milky juice. Corolla ligulate in all of the flowers of the head, and all of the flowers perfect.

1. Cichorium (Tourn.) I. Chicory

Erect branching perennial or biennial herbs with alternate leaves; involuce of two series of herbaceous bracts, the inner of 9-10 scales, the outer of 5 short spreading scales; receptacle flat, naked or slightly fimbriate; flowers bright blue, purple or pink; rays 5-toothed; Achenes striate; pappus of numerous small chaffy scales forming a crown.

About 8 species of the old world.

Cichorium Intybus L. Chicory or Succory

A branching perennial with deep roots and alternate leaves; basal leaves spreading on the ground; stem leaves oblong or lanceolate, partly clasping.

Distribution. Common along roadsides and in fields and waste places from New England to Canada and Nebraska, especially where chicory has been cultivated. It has become a troublesome weed in Wisconsin and Minnesota. It is allied to endive (Chicorium Endivia), cultivated as a salad plant.

Poisonous properties. When fed in large quantities it imparts a bitter flavor to milk and butter. It contains the bitter glucoside chicorin $C_{32}H_{24}O_{10}$. Chicory root is used as an adulterant of coffee.



Fig. 439b. Dandelion (Taraxacum officinale). 1, Single head during flowering, single head after flowering. 2, Single flower with corolla stamens and style. 3, Achenium. 4, Receptacle and single achenium. (After Strasburger, Noll Schenck and Schimper).

2. Sonchus (Tourn.) L. Sow Thistle

Annual or perennial herbs with alternate, mostly auriculate-clasping, entiredentate, lobed, or pinnatifid leaves with soft prickly margins; flower heads in corymbose or paniculate clusters; involucre bell-shaped; scales imbricated in several rows; receptacle flat and naked; achenes oblong, more or less flattened; 10-20-ribbed; pappus of soft white capillary bristles.

About 45 species of the old world.

Sonchus olcraceus L. Annual Sow-thistle

Annual or perennial succulent herbs with leafy stems, smooth and glaucous with corymbed or umbellate heads of yellow flowers. Stem leaves dentate, runcinate-pinnatifid, terminal segments large and triangular; heads numerous; flowers pale yellow, occurring in summer and fall.

Distribution. Common in fields and waste places throughout North America, except far northward. Also from Mexico to South America.

Sonchus arvensis L. Field Sow-thistle

A glabrous perennial, producing deep creeping root-stock, stem leafy, branched, basal leaves runcinate-pinnatifid, spiny-toothed, clasping by a heart-shaped base; flowers yellow; achenes transversely wrinkled.

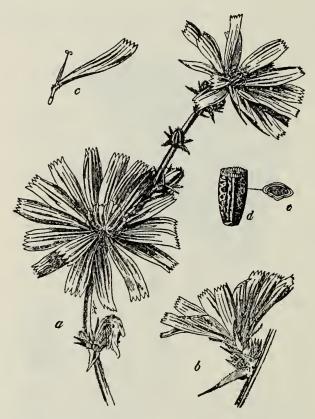


Fig. 439c. Chicory (Chicorium Intybus). a, part of plant with several heads; b, single head side view; c, single flower with strap shaped corolla; d, achenium with small chaffy scales. (U. S. Dept. Agrl.)

Distribution. Common in eastern states, Canada (Manitoba) and oecasionally in Wiseonsin, Illinois, Iowa and Utah.

Poisonous properties. The plants are more or less bitter and not liked by

Poisonous properties. The plants are more or less bitter and not liked by cattle. Their milky juice probably contains some active principle.

3. Lactuca (Tourn.) L. Lettuce

Tall, leafy-stemmed herbs with milky juice and alternate leaves; flowers white, yellow or blue, in panieled heads; involuere cylindrical, bracts imbricated in two or more series; receptacle flat, naked; anthers sagittate at the base; achenes oval, oblong or linear, abruptly contracted into a beak, dilated at the apex, bearing a soft white capillary or brown pappus.

About 90 species, natives of the northern hemisphere. Garden lettuce (L. sativa), native to Europe, is cultivated.

Lactuca Scariola L. Prickly Lettuce

Tall, erect herbs, annual or winter annual, 2-6 feet high, simple or branched except the lower part of the stem which has stiff bristles; leaves glaueous, green, smooth except the midrib which is beset with weak prickles; lanceolate

to oblong in outline with spinulose, denticular margins occasionally sinuate toothed, sometimes pinnatifid; base sagittate clasping; leaves becoming vertical by a twist; the leaves are not twisted in shady situations; flowers in small open panicled heads; each head has from 4-18 yellow flowers; achenes flat, striate nerved, obovate, oblong, produced in long filiform beak which is paler in color than the achene; pappus consists of delicate white bristles arising at the end of the beak.

Distribution. Prickly lettuce is a native of temperate and southern Europe, Canary Islands, Maderia, Algeria, Abyssinia, and the temperate regions of eastern Asia. It was introduced into North America about 1863.



Fig. 440. Prickly Lettuce (Lactuca Scariola). After Fitch.

Lactuca canadensis L. Wild Lettuce

A tall, leafy, smooth or occasionally somewhat hairy biennial 4-9 feet high; leaves 6-12 inches long; stem leaves sessile or auriculate clasping, the upper leaves smaller, lanceolate acuminate and entire; heads with about 20 flowers in spreading panicles; involucre cylindrical; rays yellow; achene somewhat longer than the beak.

Distribution. In moist places, borders of thickets and in fields from Nova Scotia to Manitoba, south to Arkansas, Louisiana and Georgia.

Lactuca pulchella (Pursh.) D. C. Blue Lettuce

A glabrous perennial, with milky juice, simple stem from 1-2 feet high, sessile, oblong or linear lanceolate entire leaves the lower runcinate-pinnatifid; heads corymbose paniculate, peduncles with scaly bracts; scales of the involucre imbricated in 3 or 4 ranks; flowers blue; achenes oblong lanceolate, somewhat flattened.

Distribution. Native to the plains from western Iowa north to Manitoba and west to the Great Basin and California, and rare as far east as Michigan. A troublesome weed in grain fields of Montana and Utah.

Poisonous properties. The wild blue lettuce is common in the west but is not liked by cattle although sometimes eaten by sheep. It has been regarded

with suspicion. The cultivated lettuce contains several active principles as follows: Lactucorol C₁₈H₃₀O, lactucol C₂₁H₃₄O, a small amount of hyoscyamin. The bitter taste of L. canadensis is due to lactucrin C₂₂H₃₄O₈, and lactucopicrin C₄₄H₆₄O₂₁. I quote Dr. Millspaugh in regard to the physiological action of L. canadensis. "Lactucarium, in large doses, causes: delirium, confusion of the brain, vertigo and headache, dimness of vision, salivation, difficult deglutition, nausea and vomiting, and retraction of the epigastric region, with a sensation of tightness; distension of the abdomen, with flatulence; urging to stool followed by diarrhoea; increased secretion of urine; spasmodic cough, oppressed respiration, and tightness of the chest; reduction of the pulse ten to twelve or more beats; unsteady gait; great sleepiness; and chills and heat, followed by profuse perspiration." The L. virosa, a wild lettuce of Europe, and occasionally in the Mississippi Valley, but never abundant westward, is regarded in Europe as poisonous.

4. Lygodesmia D. Don.

Low smooth perennial herbs with linear leaves or the lower somewhat pinnatifid, the upper of scales; heads 3-12 flowered, a single one terminating the branch; flowers purple or pink; achenes smooth or striate; pappus of copious, somewhat unequal simple bristles.

About 6 species of western North America.

Lygodesmia juncca (Pursh.) D. Don.

A tufted smooth, frequently glaucous perennial, a foot or more high, coming from a thick woody root, copious milky juice and stems; lower leaves rigid, linear lanceolate, entire, the upper scale-like; heads erect with purple flowers; achenes narrow-ribbed, pappus light brown.

Distribution. Common on the plains from the Missouri river to western Montana, Northwest Territory and east to the St. Croix river in Wisconsin. This has been reported as a troublesome weed in corn fields in northwest Iowa, troublesome also in Colorado and Montana.

Poisonous properties. The plant is bitter like many others belonging to sub-family Cichoriaceae. The milky juice no doubt contains some deleterious properties. The plant is not liked by stock.

Professors Chesnut and Wilcox say with reference to the species in Colorado:

"This species, sometimes known as prairie pink, grows abundantly on dry prairies and plains in Park, Sweet Crass, Gallatin, Meagher, Lewis and Clarke, Choteau, and Teton counties. The general distribution of the plant is from Minnesota to New Mexico and Nevada. It has been suspected by stockmen both in Montana and Utah of being poisonous to stock. The plant was not investigated, but it was ascertained that the milky orange-colored juice of nonflowering plants gathered at Toston was extremely bitter and disagreeable to the taste."

TUBULIFLORAE

Corolla tubular in all the perfect flowers, 5-lobed, rarely 3-4-lobed, or ligulate only in marginal flowers, called the ray flowers which are absent in some species.



Fig. 440a. Boneset (Eupatorium altissimum L.) Common in the Mississippi valley. At the right Maxmillian's Sunflower (Helianthus Maximiliani) common in Mississippi valley. (Charlotte M. King).

Iva L. Marsh Elder

Herbaceous or some shrubby plants, pistillate and staminate flowers in the same head. The lower leaves opposite, the upper alternate; flowers greenish, rays absent; subtended by an involucre of hemispherical or cup-shaped bracts; achenes obovoid or lenticular, without pappus.

About 12 species of western and southern America.

Iva axillaris Pursh. Small-flowered Marsh-elder

A smooth or sparingly pubescent perennial with herbaceous stems, from 1-2 feet high, with woody roots; leaves sessile, entire or nearly so; obovate, oblong or linear oblong, the lower opposite, the upper smaller and alternate; heads generally solitary in the axils of the leaves, short petioled, involucre hemispherical, pistillate, flowers with tubular corolla.

Distribution. Common especially in the saline soils from Nebraska to the Dakotas, British Columbia, California and New Mexico.

Iva xanthifolia Nutt. Marsh Elder

An annual from 1-8 feet high; stem frequently pubescent when young; all the leaves opposite, rhombic, ovate or the lowest heart-shaped, doubly serrate or cut-toothed, obscurely lobed; the upper surfaces minutely scabrous, canescent beneath, especially when young; petiole frequently ciliate at its upper end; flowers born in spike-like clusters forming a compound panicle; heads small, crowded; outer bracts of the involucre broadly ovate, greenish; inner membranaceous; achenes glabrate.

Distribution. In alluvial ground or along streams, Saskatchewan and Nebraska to New Mexico, Utah and Idaho. In the western part of the state of Iowa this weed is extremely common as in Woodbury, Harrison, Monona and Fremont counties. It occupies not only the vacant lots but is found in the streets and cornfields. To the west in Nebraska it becomes increasingly abundant and in the irrigated fields of portions of Colorado it is frequently 8 feet high. It is common in the Red River Valley of the North and other parts of Minnesota and Dakota, and Manitoba. It is a most aggressive weed.

Poisonous properties. This plant and the preceding produce an unusually large amount of pollen and have been looked upon as in part responsible for hay fever, the pollen being simply an irritant of the nasal mucous membrane.

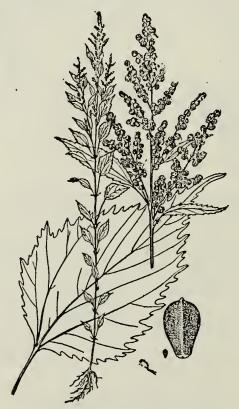


Fig. 441. False ragweed (*Iva xanthifolia*). The pollen is possibly a cause of hay fever. (Dewey, U. S. Dept. Agr.).

6. Ambrosia (Tourn.) L. Ragweed

Herbs; leaves alternate or opposite, lobed or dissected; flowers in heads, fertile, 1-3 together, sessile in the axils of leaves or bracts; involucre of the pistillate flowers top-shaped, ovoid or globose, closed, 1-flowered, armed with 4-8 tubercles or spines; corolla none; pappus none; involucre of the staminate flowers saucer-shaped, 5-12 lobed, many flowered; receptacle nearly flat or with

filiform chaff; corolla funnelform, 5-toothed, anthers but slightly united; achenes ovoid.

The twelve species are mostly native of North America.

Ambrosia trifida L. Great Ragweed, Ironweed

A stout, scabrous, hispid or nearly glabrous annual, 3-12 feet high; leaves opposite and petioled, 3-nerved, deeply 3-5-lobed, the lobes ovate, lanceolate and serrate, the upper leaf sometimes ovate and undivided; flowers monoecious,



Fig. 442. Ambrosia showing flowering stalk, pistils, stamens, and longitudinal sections of stamens in perianth and ovary. (After Faguet).

staminate borne in spikes surrounded by the larger bract-like leaves; the involucre is turbinate to ovoid, 5-7-ribbed, beaked, each rib bearing a tubercle near the summit; the involucre enclosing a single achene.

Distribution. The greater ragweed is distributed from Quebec to Florida throughout the Atlantic region to Texas, common throughout the Mississippi Valley west to Colorado and Northwest Territory, in Manitoba and Saskatchewan.



Fig. 443. Tall ragweed (Ambrosia trifida). (U. S. Dept. Agr.)

Fig. 444. Small Ragweed (Ambrosia artemisisfolia). In the lower left hand corner is a bur that contains the seed; upper right hand corner, a flower; in middle, branch and leaf. Probably poisonous. (Holm-Britton).

Ambrosia artemisiifolia L. Hogweed. Bitterweed

A puberulent or hirsute branched annual 1-3 feet high; leaves thin, once to twice pinnatifid; the upper alternate, the lower usually opposite, pale or cancscent beneath; flowers monoecious, the staminate above and the pistillate in the lower axils of the leaves; the fertile heads are obovoid or globose, short beaked and 4-6 spined.

Distribution. A troublesome weed in northern states. Its distribution in North America is from Nova Scotia to Florida throughout the Atlantic states and Mississippi Valley; in the Rocky Mountain region and west to British Columbia; also in Mexico and the West Indies and South America.

Poisonous properties. The greater ragweed is regarded as especially

767

troublesome as an exciting cause with reference to hay fever. The following from Dr. Johnson on this point is of interest:

These plants are said to be stimulant, tonic, and astringent. A decoction has been used, chiefly in domestic practice, as a topical astringent in chronic catarrhal affections.

Of late years A. artemisiifolia has attracted considerable attention on account of its real or assumed agency in the production of hay fever. The plant produces pollen in great abundance, which is extremely irritating to the air-passages of many people, and is capable of exciting asthmatic attacks in susceptible persons. Now as the weed is so very abundant, and its time of flowering coincident with the greatest development of hay-fever, the relation of cause and effect has been asserted by many writers. That it may be so in a certain proportion of cases is quite probable, but that its influence in this direction has been overrated is still more probable. The pollen of all plants is irritating to the air-passages of sensitive people, but probably little more so than any other dust of an organic character; and the proportion which rag-weed pollen in the air of any specified locality bears to that of all other plants combined must be very small indeed. Much less still must its proportion be to other pollen and organic dust in the air of cities, where this affection has become endemic-and fashionable.,

The ragweeds, marsh elder, goldenrod, and chrysanthemum, as well as the pollen of some grasses, are said to produce hay fever. In recent years a toxic substance has been isolated which belongs to a class of poisonous substances known as toxalbumin,

The fact that this troublesome disease is caused by a poisonous toxin has led to a study of serum treatment by Dunbar* who has produced an antitoxin which he calls "pollantin," and Weichardt** another called "graminol." Pollantin is obtained from the blood serum of horses which have been immunized with the pollen toxin. I ambar's hav fever serum is sold both as a powder and as a fluid. There are those, however, who believe that hay fever is not due to poison by pollen toxin so that this treatment can be of no use. A. Wolffeisner*** attributes the action of the serum not to antitoxins but to colloidal Weichardt prepared the serum from the blood of herbiverous animals. Sattistics of The Hay Fever Union of Germany for 1906 indicate that the results with the treatment of "graminol" were favorable.

A correspondent from Nebraska sent to the writer a specimen of the small ragweed stating that it was abundant in his pasture and that where cattle used it as forage, the mouths of these animals became very sore. The plant is bitter and possibly may be irritating at times. There were no parasitic fungi on the specimens sent us.

7. Xanthium (Tourn.) L. Cocklebur. Clothur

Coarse low branching annual herbs with alternate toothed or lobed petioled leaves; monoecious flowers; staminate flowers with a short involucre of several distinct bracts, receptacle cylindrical; pistillate flowers with a closed involucre, covered with hooked prickles; 2-celled, 2-flowered, in fruit forming a bur; achenes oblong, flat, without pappus.

The 12 species are widely distributed.

Xanthium spinosum L. Clothur

A pubescent branched annual herb with slender yellow 3-parted spines in the axils; leaves lanccolate or ovate lanceolate, white downy underneath; bur oblong cylindrical, armed with single short beak and numerous glabrous prickles.

Berlin Klin. Woch. 1903:140.

Berlin Klin. Woch. 1903:24, 25, 26, 28.

** Klinisch-ther. Woch. 1903:1457.

*** Das heufieber, sein wesen und seine Behandlung. München 1906. In this connection the very excellent review on Serum Therapy in E. Merk's Annual Report, 1909, Vol. 23 should be consulted.

Distribution. Common in waste grounds from Ontario to Missouri and Texas to Florida. Native to the old world.



Fig. 445. Thorny Clot-bur (Xanthium spinosum). After Hochstein.

Xanthium Strumarium L.

A low, rough, branching annual from 1-2 feet high; leaves slender, petioled, broadly ovate or orbicular, 3-5-lobed, both surfaces rough; bur oblong, smooth or nearly so, with two straight or nearly straight beaks.

Distribution. In waste places along the Atlantic seacoast. Native to Europe,

Xanthium canadense Mill. Cockle Bur

A coarse rough annual from 1-3 feet high, stem marked with brown punctate spots; leaves alternate, cordate or ovate, 3-nerved, long petioled; flowers monoecious, staminate and pistillate flowers in different heads, the pistillate flowers clustered below; the involucre of the staminate flowers somewhat flattish of separate scales, receptacles cylindrical; scales of the fertile involucre closed in fruit, 2-beaked, containing 2 achenes; the bur is densely prickly and hispid, achenes oblong without pappus.

Distribution. In Iowa this species is very common along the sandy bottoms of our streams and river courses. It is less troublesome in uplands but here and there it does occasion some trouble even in the central and southern part of the state. However, in southern and southeastern Iowa the weed is often quite troublesome in cornfields, coming up in enormous quantities. In Texas, too, it appears in the very richest soil. Its distribution in North America is

from Nova Scotia to North Carolina, southwest to Texas and west through Colorado, Utah and Nevada, and north to Saskatchewan and Manitoba. Most abundantly developed in the Mississippi Valley region from Minnesota to Texas.

Poisonous properties. The injury from this plant probably comes largely from its mechanical action. As the involucre is indigestible, its barbs sometimes injure an animal feeding upon it. Stock will probably not eat very much of it, but on account of the hooked awns of the involucre the animal may have considerable difficulty in removing them. The hairs of the plant cause itching. Several cases of poisoning of hogs, probably due to this plant, have been reported to me. The plant contains the poisonous glucoside xanthostrumarin which resembles datiscin. On heating, the odor of succinic acid is given off. According to Chesnut, the young seedlings of three species of cockle bur, among them our Canadian cockle bur (Xanthium canadense) are poisonous to hogs. Dr. Bitting was unable to find a poisonous substance in the growing plants. He thinks the injurious properties are largely mechanical.



Fig. 440. Cockle-bur (Xanthium canadense). Cause of mechanical injuries to animals. (Dewey, U. S. Dept. Agrl.)

A writer in the American Agriculturist says:

When the seed is ripe the bristles are very hard and sharp, and in the stomach and intestines of the animal, mat or felt together by the aid of their barbs, forming large balls, which obstruct the intestines and prove fatal. In these cases, death is evidently not from any poisonous quality of the hay, but is due to mechanical causes. While Cocklebur may not be poisonous, it may in a similar manner kill swine by mechanical obstructions. When the burs are ripe, they readily leave the plant, and attach themselves to the coat of any passing animal. The plant is especially annoying to sheep owners, as these burs become entagled in the wool. Those who have traveled in Texas and Mexico, soon make acquaintance with the cockle bur. Horses and mules, while grazing for a single night, will have their tails clotted with these burs, and converted into a useless club. It can be understood, that if swine eat the burs in considerable quantity, the pieces of them can, by means of the prickles, form masses which may prove fatal. Both on account of its probable danger to swine and its injury to sheep, there should he an united effort to destroy the plant. Being an annual, its extermination would not be difficult. If the plants are cut down before the seed is ripe, new ones can not appear unless the ground is re-seeded.



Fig. 447. Boneset (Eupatorsium urticaefolium). A common plant in woods. It is supposed by some to cause milk fever or trembles. Branches with numerous small heads. (Lois Pamnel.)

8. Eupatorium (Tourn.) L. Boneset

Erect perennial herbs with opposite, whorled or alternate leaves, often sprinkled with resinous dots; flowers in corymbose heads, white, bluish, or purple; bracts of the involucre in two series; receptacle naked; corolla regular, tube short 5-lobed; branches of the style slender, thickened upward or clubshaped, very minutely and uniformly pubescent, with stigmatic lines indistinct; achenes 5-angled, truncate; pappus of numerous fine capillary bristles, arranged in one row.

About 450 species in warm temperate and tropical regions. An oil is made from the southern E. capillifolium, which has an aromatic, pepper-like odor.

The boneset (E. perfoliatum), is commonly used as a domestic remedy, employed in catarrhal affections and fevers. The Joe-Pye weed (E. purpureum) is used in urinary affections.

Eupatorium urticaefolium Reichard. White Snake-root

A smooth or nearly smooth branching herb from 2-3 feet high; leaves opposite, thin rounded, cordate at the base or abruptly narrowed into a slender petiole, coarsely and sharply toothed; heads in ample and loose corymbose clusters; flowers white; involucre narrowly campanulate; bracts linear, acute or acuminate.

Distribution. Common in rich woods from New Brunswick to South Dakota and Nebraska to Indian Territory and Louisiana.

Poisonous properties. This plant is said to produce the disease known as trembles in cattle, horses and sheep, and milk sickness in people. Mr. E. L. Mosely states that:

Milk-sickness is known to be due to the use of milk, butter, cheese or meat of animals afflicted with the trembles, but what causes the trembles has not been well understood. It has long been known that only the animals allowed to run in the woods were affected, and experience showed that certain woods were very dangerous, while others were safe. People who came from Pennsylvania with a view to settling here returned to their own State on learning of the peril of pasturing animals in Ohio. To this day many woods in this district are not pastured, because animals would soon die if turned into them.

The Eupatoriums are not palatable. Anyone who has tasted boneset will admit that this is true of Eupatorium perfoliatum. In the South I have observed that animals leave Eupatorium serotinum untouched even where they have been confined so as to eat almost every other green thing in reach. In northern Ohio I have found Eupatorium urticaefolium, the white snake-root, growing abundantly in a number of woods where animals were pastured but no sign of their having eaten it. But if the pasture becomes poor, some are likely to eat it.

On the 8th of last October I visited a piece of woods in Sandusky county where there was nothing fit for an animal to eat, the principal herbs being nettle, white snake-root, poke and black nightshade, with some clearweed, basil, and bedstraw. Every plant of snake-root had been nipped off so that I did not see one more than about half the normal height. This had probably been done by cattle from the adjoining pasture which were doubtless accustomed to spending a portion of hot sunny days in the shade of the woods. A few weeks before my visit a man and his wife who had been using butter made from the milk of cows in this pasture had milk-sickness and the wife died.

Elisha IIaff, Townsend township, Sandusky county, did not think trembles were due to any weed, until he found that western sheep which he turned into his woods ate the white snake-root and died of trembles. Sheep whose ancestors had long been in the region did not eat it, and did not have trembles. Since that he has been destroying the weed.

Mr. William Ramsdell of Bloomingville informs me that about 1842 when there was so much discussion of the subject the boys of the neighborhood used to assemble evenings at the old lime-kiln southeast of Castalia and experiment on dogs. They would boil or steep the white snake-root and putting the extract in milk give it to the dogs, in which it would induce the trembles; a large number were killed in this way. Some one experimented on sheep with the same result. He informs me also that a Mr. Redmond (who did not believe that-the weed was the cause of trembles) chewed some of the weed and died after suffering for about four weeks.

On November 26th my pupil, Oscar Kubach, using snake-root I had recently gathered, broke up the stems and leaves of two plants and soaked them over night in about a pint of milk, of which he gave about a gill at about 9 A. M. to his tom-cat. The cat took about one-half of it.

About 9:30 it seemed to take effect and he tried very hard to vomit but could not. He took long, deep breaths. He was quiet and wanted to sleep very hard. All of a sudden be would tremble very hard, then again very little. A watery fluid passed from his eyes and mouth. He chose a spot in the sun and when driven away walked back in a staggering manner. He had no appetite. His senses seemed to be duller, as he did not care for anything. He went to sleep about 10:30 but did not sleep sound. He seemed to be in an unconscious state for the rest of the day. The next morning about 10:30 he walked about three rods and there died about noon.

Mr. Mosely estimates that 5000 animals have succumbed from the disease in a small area in northern Ohio. There are many cases commonly in this same region from milk sickness.

Dr. Albert C. Crawford of the Bureau of Plant Industry, investigated the so-called milk sickness occurring in Minooka, Illinois, which resulted in the death of about 50 head of cattle, and gives the following conclusions:

To sum up, it certainly can not be said that it has been proved that milksickness is due to any constituent of E. urticaefolium. The transmission of the disease by eating small quantities of meat or milk of animals sick with the "trembles" and the fact that cooked meat or boiled milk does not produce this disorder point primarily rather to a parasitic origin, while the fact that Eupatorium urticae folium is abundant in areas where the disease is not known and absent in some milksick districts also indicates that the plant has no relation to the disease. If it does, it would be only an accidental carrier of some pathogenic organism. According to reports, the same flora may be in areas in which "trembles" occur as in those free from it, and milksickness is also said to occur where no vegetation grows (inclosed pens). The disease also has disappeared from an area after simply clearing the woodland where it occurred and turning it into pasture. Again, severe epidemics have occurred in winter when the foliage has disappeared, which would tend to exclude the higher non-evergreen plants as the cause of this disorder. In fact, all the evidence in hand is against the causation of this disease by such plants, and certain analogies with cases of botulismus suggest a somewbat similar cause. If there is any truth in the statement that cattle exposed in pasture to night air especially contract the disease, this fact might suggest the more or less direct connection of some night organism as a carrier of the parasite, and certain parasites are supposed to be associated with certain localities. Very little is known chemically of Eupatorium urticaefolium.

And this seems to confirm the investigation by Dr. Bitting of the Indiana Agricultural Experiment Station. It seems very doubtful that this plant causes milk sickness, since it is very common in many pastures in the west where trembles does not occur.

In regard to boneset (E. purpureum and E. perfoliatum) Dr. Johnson states as follows:

Of domestic remedies few are better known or more largely used than boneset. It is tonic, diaphoretic, emetic, and cathartic, the different effects depending largely upon the size of the dose and mode of administration. The infusion, taken cold in moderate doses, is tonic, and is employed in debility of the digestive organs and in convalescence. Taken warm in large doses, the infusion or decoction produces copious diaphoresis, and is employed in the acute stages of catarrhal affections and in fevers, especially those of an intermittent or remittent type. In still larger doses the warm infusion or decoction produces emesis or catharsis; these effects are, bowever, seldom sought.

E. purpureum, or gravel-root, is said to be diuretic and to have been employed in urinary affections, but it has not attained an established reputation and is seldom used.

Boneset (E. perfoliatum) contains the glucoside eupatorin; the E. purpureum contains the glucoside euparin $C_{12}H_{11}O_{3}$.

Dr. T. Holm gives an extended account of the medicinal qualities accompanied by the anatomical structure of this plant. *Eupatorium perfoliatum** according to Dr. Holm is now prescribed as a tonic and in large doses is an emetic.

9. Trilisa Cass. Trilisa

Erect perennial herbs, fibrous roots, leaves alternate and simple; heads in terminal corymbose panicles, discoid, 5-10-flowered, flowers white, receptacle flat; corolla regular, 5-lobed; achenes nearly terete, 10-ribbed. A small genus closely allied to Liatris. Two species native to eastern North America.

Trilisa odoratissima Cass. Vanilla Plant

A rather stout, somewhat glabrous perennial, leaves pale obovate-spatulate,

[•] Merck's Rep. XVII:326-328. f. 1-11.

COMPOSITAE—THISTLE FAMILY—VANILLA PLANT 773.

or oval, thickish and clasping; head in corymbose clusters; achenes glandular pubescent.

Distribution. From Virginia to Louisiana.

Trilisa paniculata Cass. Hairy Trilisa

Viscid-hairy, perennial, leaves entire, base lanceolate, narrowly oblong, acute, or obtusish, those of the stem small; heads paniculate; achenes finely pubescent.

Distribution. From Virginia to Georgia and Florida.

Poisonous properties. The former plant has the odor of vanilla and contains the substance cumarin CHO. Dr. Johnson says:

Odoratissima deserves much more attention from the fact that it is largely used as an adulterant of smoking tobacco, than from any demonstrated medicinal virtues. There is abundant evidence to show that the leaves of this plant enter largely into the manufacture of many grades of smoking tobacco, especially those employed in our domestic cigarettes. And the author is convinced, from personal experience and observation, that the deleterious effects produced by smoking tobacco thus adulterated are much greater than those produced by the consumption of pure tobacco in even great excess. The inhalation of a few whiffs of the smoke from a cigarette made from this adulterated material, provided the inhalations are made in quick succession, produces a train of cerebral sensations of an intoxicating character as much different from any effect of tobacco alone as could be imagined; and prolonged use of such cigarettes invariably produces great derangement of the digestive organs, very littleresembling the dyspepsia induced by excessive use of tobacco, together with cardiac symptoms often of a distressing character. And again, the habit of smoking coumarin in this form appears to become more inveterate, more exacting, than that of the use of tobacco alone, so that the unhappy victim-for such he should be called-is never comfortable except when indulging. Hence it happens that cigarette-smoking in this country in its effects upon adolescents especially, is assuming the proportions of a great national evil, and is producing far more deleterious effects than in other countries where it is practised to a greater extent but with different material.

10. Liatris Schreb. Button Snakeroot. Blazing Star.

Perennial herb, usually from a corm-like tuber; leaves alternate entire, narrow; flowers spicate or racemose, discoid, scales of the involucre few or many imbricated in several series, the outer shorter, corolla regular, 5-lobed or 5-cleft; branches of the style exserted; achenes 10-ribbed, slender tapering to thebase; flowers rose-purple, rarely paler in color.

Distribution. About 20 species in North America.

Liatris spicata (L.) Willd. Snakeroot

Smooth or somewhat hairy perennial; leafy stem; leaves linear the lower 3-5 nerved; heads crowded in a long spike, 8-12 flowered; involucre cylindrical, bell-shaped, flowers purple; pappus not very plumose.

Distribution. In moist soil from New England to Florida, Kentucky, Arkansas, and South Dakota.

Liatris pycnostachya Michx. Snakeroot

Hairy or smooth perennial with a stout stem 3-5 feet high; leaves linear-lanceolate, the upper very narrowly linear; spikes 6-20 inches long, densely-flowered, flowers purple; pappus not very plumose.

Distribution. Prairies, Indiana to Minnesota and southward.

Liatris punctata Hook. Western Snakeroot

A glabrous or sparingly pubescent perennial; stout rootstock; leaves rigid punctate; spike many flowered; heads 3-6 flowered, purple; bracts of involucre-oblong, often ciliate on the margins; pappus very plumose.

Distribution. From Minnesota, Western Iowa, to Montana, Texas, New Mexico and Sonora.

Poisonous properties. Several species of Liatris are powerful diaphoretics, formerly these species were used as antidotes against snake bite. Dr. Johnson says that they probably possess no antidotal properties whatever and beneficial effects attributed to them are doubtless due to the diaphoresis induced by the administration of large quantities of the hot decoction.

11. Grindelia Willd. Gum Weed.

Coarse perennial or biennial herbs, occasionally woody at the base; leaves alternate, sessile or clasping, spinulose serrate; involucre hemispherical; scales imbricated in several series; heads large, terminating leafy branches; radiate or rayless; ray flowers yellow, pistillate, disk flowers perfect or staminate; achenes short, thick, compressed or turgid; pappus of 8 awns, soon falling.

About 25 species, from western Minnesota and Iowa westward and south-ward to Peru and Chili.

Grindelia squarrosa (Pursh.). Gum Weed

A resinous, viscid, glabrous perennial from 1-3 feet high; leaves alternate, spatulate to linear oblong, sessile or clasping, spinulose serrate; heads many flowered; ray flowers yellow, pistillate; scales of the involucre hemispherical, imbricated in several rows with green tips; achenes short and thick; pappus consisting of 2 or 3 awns.

Distribution. Common west of the Missouri river from Mexico, Nevada and Texas to British America and east to Minnesota, Illinois and Missouri, occasionally naturalized eastward.

Poisonous properties. Very abundant in the west; is not liked by stock. The G. robusta and G. squarrosa are used in medicine in moderate doses to stimulate the mucous membrane and are beneficial in catarrhal affections. They are also antispasmodics. An alkaloid has been isolated from G. robusta. It contains grindelin, a bitter alkaloid, and two glucosides which resemble the saponins of Polygala.

12. Solidago L. Golden-rod

Perennial erect herbs, simple or branched; leaves alternate, simple, toothed or entire; heads small, in terminal or axillary panicles, cymose corymbose; ray flowers yellow, rays few or many pistillate; disk flowers yellow, perfect; involucre hemispherical or bell-shaped, bracts appressed, destitute of green tips, achenes many-ribbed, terete or nearly so; pappus of simple capillary bristles.

A genus of about 100 species mostly in North America. About one-half of the number found in northern states east of the Missouri river. A few of the species are handsome and ornamental, like Solidago speciosa, S. Drummondii, S. Missouriensis and S. odora. The latter species is used as a stimulant and carminative. From this species there is derived an aromatic oil. An oil is also derived from Solidago canadensis, which strongly resembles the oil obtained from pine needles. The leaves of the fragrant golden-rod (S. odora) are often used as a substitute for tea; it contains an aromatic volatile oil.

Solidago canadensis L. Golden-rod

A perennial with rough stem, from 3-6 feet high; leaves hairy beneath, rough above, lanceolate and pointed, sharply serrate; heads small, few flowered;

rays yellow, short pistillate; scales of the involucre appressed, not herbaceous; receptacle small, not chaffy; achenes ribbed; pappus simple, of capillary bristles.

Distribution. Widely distributed from New Brunswick to Florida. Common in the Mississippi Valley, especially along fences and in pastures and borders of fields in the Rocky Mountains to the Northwest Territory, British, Columbia to Arizona.

Solidago rigida L. Rigid Golden-rod

An erect perennial from 2-5 feet high, rough and somewhat hairy, with minute pubescence; leaves oval or oblong, thick and rigid, not 3-nerved, the upper sessile, slightly serrate; heads in a compound corymb, large, 3 or more flowered; rays large, 7 to 10, yellow.

Distribution. Common in the upper Mississippi Valley, especially on theprairies; occurs east to New England.

Poisonous properties. It is thought by some that hay fever is caused by this and other species. A disease of horses in Wisconsin a few years ago was attributed to the eating of golden-rod, but Chesnut thought this might be caused by a rust fungus, Coleosporium solidaginis.

Mr. J. L. Scott, who made an investigation of this disease, reported in Garden and Forest as follows:

During the past four years a large number of horses have died in the northern part of this state from the ravages of a disease which has baffled the skill of veterinarians, and I have been called upon to make investigations as to the cause and nature of the malady. At first it was thought to be anthrax, and samples of the blood and sections from the spleen and other internal organs were sent to the Bureau of Animal Industry and to Dr. Russell, of the State University, for bacteriological examination. Numerous bacteria were found, but the bacillus anthracis was not present.

The horses affected were in the majority of cases heavy draught horses from the lumber camps. These animals were brought from the woods in the spring, usually in good condition, and turned out to pasture. Most of them were fed grain while on pasture. On the farm of Mr. C. F. Reynolds, Hayward, Wisconsin, over seventy horses died during the past four years from this peculiar malady. The pasture contained about four hundred acres, three hundred acres of which had been broken and seeded to timothy. Adjoining this was one hundred acres of "slashings" or land from which the timber had been cut, but which had never been broken. This was thickly covered with Golden-rod. On one side of the farm is a lake with a clean gravel bottom and shore. The lake is fed by springs. There is no marsh or low land on the farm. Upon investigation I became convinced that the cause of the trouble was to be found either in the food or water, and watched the horses closely for several days, and saw them eating the Golden-rod greedily—some of them, especially those affected, seeming to prefer the plant to anything else.

I also visited the farm of Peter Traux, near Eau Claire. There is no Goldenrod to be found on this farm and the disease has not made its appearance. During the past summer,. Mr. Traux placed ten horses in pasture near by, where the plant was plentiful, and eight of them died during the summer and the remaining two were affected. When the healthy horses are taken from the pasture in the fall the disease disappears. None of the animals attacked by the malady bave recovered, and medicinal treatment does not seem to produce any beneficial effect.

Symptoms: The animal appears dull, ears drooped, temperature elevated, ranging from 103° to 107°, Fahrenheit, during the entire course of the disease. The visible mucous membranes are pallid. On the mucous membranes of the vulva small petechial spots are seen. Occasionally the legs swell and oedematous enlargements appear under the abomen. The appetite remains fairly good during the entire course of the disease. Emaciation takes place rapidly as the disease advances. Loss of coordination with staggering gait. Death takes place in from two weeks to two months from the onset.

Post mortem: On cutting open the body the blood appears to be completely disintegrated, resembling ordinary blood serum. Intestines bloodless, with numerous petechial spots on themucous membrane. Spleen enlarged, weighing from six to ten pounds. No structural changes.

apparent to the naked eye. The lungs and kidneys apparently normal. The brain and spinal cord were not examined.

I am fully convinced that this disease is due either to some poisonous principle in the plant or some parasitic fungus upon the surface of the same. It is now too late in the season for any investigation to be carried on in this direction this year, but I intend to have the matter thoroughly investigated next summer.

13. Aster (Tourn.) L. Aster

Herbs generally perennial with corymbose panicled or racemose heads; heads many flowered, radiate; the ray flowers in a single series, fertile. Bracts of the involucre more or less imbricated, generally with herbaceous tips; receptacles flat; achenes somewhat flattened; pappus simple, consisting of capillary bristles. A large genus of 275 species, chiefly in eastern North America. Flowering in the autumn. A few of the species are cultivated for ornamental purposes.

The New England aster (A. novae-angliae) occurs in moist ground. The A. laevis with somewhat clasping leaves is common in dry soil or prairie regions of the West. The small white aster (A. multiflorus) with small leaves is common along roadsides in dry soil. Very few of the species have deleterious properties.

Aster Parryi Gray. Parry's Aster or Woody Aster.

A somewhat hoary perennial with deep woody roots and a short more or less branched stem; hoary leaves, spatulate-linear, cuspidate; heads solitary, bracts of the involucre oblong-lanceolate, long acuminate, pubescent, rays white over half inch long, achenes white villous. This is the *Xylorhiza Parryi* Gray.

Distribution. Common in saline soils. In Colorado, West Wyoming and Utah.

Poisonous properties. This plant has recently come into prominence in western Wyoming where the disease "grub in the head" has been attributed to the plant but Dr. Aven Nelson 1 has attributed this disease to a fungus Puccinia xylorrhizae which according to Dr. Nelson abundantly occurs on the plant. He says:

If it should prove to be true that the malady is due to the cating of the aster, then it may be the aster itself that is the source of the trouble, but the chances are rather hetter that the specific poisonous qualities are due to the fungus. Some other parasitic fungi have been proved poisonous and we may well, in the absence of evidence to the contrary, also suspect this one.

Dr. Nelson states that the trouble is not a new one and that similar outbreaks have occurred in previous years, the disease recurring in the same localities. The disease was prevalent near Medicine Bow and northward to the Shirley Basin as well as Natrona county. The farmers in the vicinity where this disease occurs speak of the area in which the plant occurs as the "poison patch." Dr. O. L. Prien, Wyoming Agricultural Station, and Dr. Frederick of the Utah Agricultural Station, are making a careful study of this disease. To prevent the trouble, sheep should be kept, so far as possible, away from areas in which this plant occurs.

14. Erigeron L. Fleabane, Daisy

Branching or scapose herbs with entire or toothed leaves; heads in corymbose, paniculate or solitary peduncled heads; scales of the involucre narrow, nearly equal, not foliaceous or green-tipped; flowers radiate, white violet or purple, numerous, fertile; disk flowers yellow, tubular and perfect; branches of

¹ Press Bull. Wyoming Agr. Exp. Sta. No. 10.

the style more or less flattened with short appendages; achenes pubescent and 2-nerved; pappus a single row of fine capillary bristles.

About 120 species of wide distribution, most numerous in North America. Several species of fleabane (E. philadelphicus, E. annuus and E. canadensis) are used in medicine. They are reputed diuretics, tonics and astringents.



Fig. A. Small Horseweed (Erigeron divaricatus). From Indiana to Minnesota to Nebraska and southward. Common in sterile grounds. (Charlotte M. King).



Fig. B. Horseweed, Mares' Tail (Erigeron canadensis). A common weed throughout the United States, especially northward, also in Europe. Acrid said to be irritating. (Charlotte M. King).

Erigeron canadensis L. Horseweed

Bristly herb, stem hairy or somewhat glabrate, 1-6 feet high, simple or paniculately branched; leaves usually pubescent or ciliate, the lower spatulate, incised or entire, obtuse or acutish, the upper generally linear and entire; heads numerous, with inconspicuous white ray flowers shorter than the pappus; achenes small, flattened; pappus of numerous small fragile bristles.

Distribution. Common throughout the eastern part of North America except far northward. Common also in the Rocky Mountain region, and in

waste and cultivated grounds along the Pacific coast appearing as an introduced weed. Also an introduced weed in Europe and South America.

Erigeron ramosus (Walt.) BSP. White Weed. Fleabane.

Stem and leaves somewhat hirsute and hairy, roughish; leaves entire or nearly so; the upper lanceolate, the lower oblong or spatulate; heads borne in corymbose panicles; ray flowers white and twice as long as the scales of the involucre; achenes small, pappus double, the inner of fragile bristles.

Distribution. From Nova Scotia to Florida, west to Louisiana and Texas to Northwest Territory.

Erigeron annuus (L.) Pers. Fleabane. White Weed.

A sparingly pubescent annual from 3-5 feet high; leaves thin, coarsely and sharply toothed, the lower one ovate, or ovate-lanceolate, acute and entire on both ends; heads corymbed and rays white, tinged with purple.

Distribution. A common weed in meadows, pastures and woods in northern United States.



Fig. C. White weed (Erigeron annus). Common in clover and timothy meadows. A troublesome weed. Leaves and stem with a somewhat bitter flavor. (Charlotte M. King).

Poisonous properties. Common horseweed (E. canadensis) abundant in many places has been looked upon as being suspicious. It contains an oil, mainly a terpene $C_{10}H_{10}$. It has an acrid taste and is neutral in its reaction. It causes smarting of the eyes, soreness of the throat, aching of extremities, and colic. It causes irritation to people handling the plant. When in hay it

779

may be injurious because the leaves are very bitter and cause much dust. The other species are very abundant in meadows in central Mississippi Valley states, and are regarded with much disfavor.

15. Rudbeckia L. Cone-flowers

Annual, biennial or perennial herbs, usually rough or rarely smooth, with alternate leaves; flowers in terminal clusters; involucre hemispherical; bracts imbricated in 2-4 series; scales leaf-like, in 2 rows, spreading; receptacle conical or columnar with chaffy scales; ray flowers neutral; disk flowers perfect, fertile; achenes 4-angled, obtuse or truncate at the apex; pappus wanting or a minute crown.

A small genus of some 30 species in North America and Mexico. Several species are common in cultivation.

Rudbeckia hirta L. Cone-flower. Nigger-head

A rough, bristly, hairy biennial from 1-21/2 feet high with stems simple or branched near the base, bearing a long pedunculate head; leaves nearly entire, the upper sessile, oblong or lanceolate, the lower petioled and spatulate; heads many-flowered, radiate, the rays about 14, neutral; receptacle columnar or conical; chaff hairy at tip, acutish; ray flowers yellow, disk dull brown; achenes four-angled; pappus none.

Distribution. Common in dry soils and on the prairies and meadows in the northern Mississippi Valley, also a common weed in meadows in the eastern states. Introduced largely with clover seed.

Rudbeckia laciniata L. Golden Glow or Tall Cone-flower.

A tall branching smooth perennial; leaves thin, minutely pubescent; the upper leaves irregular; ray flowers yellow; disk flowers dull greenish; bracts of the involucre unequal; pappus a short crown. Improved forms of this species are cultivated under the name of golden glow.

Distribution. From Canada to Florida, Kentucky, Kansas and New Mexico to Manitoba and Montana.

Rudbeckia occidentalis Nutt. Western Cone-flower

Nearly glabrous and smooth, or somewhat scabrous-puberulent; leaves undivided, ovate or ovate-lanceolate, acuminate, entire or irregularly and sparingly dentate, 4-8 inches long; upper sessile by a rounded or subcordate base; lower abruptly contracted into a short winged petiole, rarely a pair of obscure lateral lobes; disk in age becoming 1½ inches long, and achenes 2 lines long.

Distribution. Common in the Rocky Mountains from Utah, Wyoming and Idaho.

Poisonous properties. Dr. Schaffner states that the R. laciniata is poisonous to sheep. It is eaten by cattle. The R. occidentalis is eaten by sheep but not relished. May be suspicious. A correspondent in Kansas and one in Iowa state that the plant is poisonous to cattle.

16. Bidens L. Bur Marigold or Beggar-ticks

Annual or perennial herbs with opposite leaves or the upper alternate; involucre double, campanulate, the outer scales commonly foliaceous; heads manyflowered; receptacle chaffy; rays present or absent; when present, neutral, in our species yellow; disk flowers perfect, fertile; achenes flattened or 4-sided; pappus of 2-6 teeth, upwardly or downwardly barbed.

About 70 species of wide distribution,

Bidens discoidea (T. & G.) Britton. Small Tickseed

A diffusely branched annual from 1-2 feet high; leaves petioled, ternately divided, leaflets ovate-lanceolate, pointed and serrate; rays wanting; disk flowers yellow; achenes narrow wedge-shaped, bearing a pair of short and stout upwardly or downwardly barbed awns.

Distribution. Common on wet banks throughout the northern Mississippi Valley.

Bidens frondosa L. Beggar-ticks. Stick-tights.

A branching hairy or smooth annual from 2-6 feet high; leaves petioled, 3-5 divided, the stalked leaflets lanceolate, pointed, coarsely toothed; rays small and yellow; involucre double, the outer foliaceous, longer than the head; receptacle flattish with deciduous chaff; achenes wedge-obovate, 2-awned, the awns downwardly barbed.



Fig. 447a. Spanish Needle, Boot Jacks (Bidens frondosa). The flattened achenes with downwardly barbed arms more or less irritating. (Charlotte M. King).

Distribution. Widely distributed in moist places throughout the northern states; often a very troublesome weed in gardens. Widely scattered because of the "seeds" adhering to clothing, pelts of animals, etc.

Bidens bipinnata L. Spanish Needle

A smooth branched annual from 1-5 feet high; leaves petioled and 1-3

pinnately dissected; heads many flowered, small, long peduncled; involucre double, outer of linear scales; ray flowers yellow or none; achenes linear, 4-angled, narrowed upward into a beak, nearly smooth, the four downwardly barbed awns usually spreading, shorter than the achenes.

Distribution. Native to Mexico and tropical America and the southern states, now occurring as far north as Rhode Island to Ohio, southern Iowa and Nebraska. A weed also in southern Europe and Asia.

Poisonous properties. All of the species are local irritants but especially B. frondosa,

17. Helenium L. Sneeze-weed.

Erect branching annual or perennial herbs; plants often sprinkled with resinous matter; leaves alternate decurrent on the stem; branches terminated by a single or corymbose heads, yellow or purple in color; involucre short and broad but small; bracts in 1 or 2 series, linear or subulate reflexed or spreading; ray flowers pistillate and fertile or neutral, rays 3-5 cleft; disk flowers perfect, fertile, corolla 4-5 toothed; style branches dilated and truncate at the apex; achenes top-shaped and ribbed; pappus of 5-8 thin 1-nerved chaffy scales extended into a point.

About 25 species in North America. All of the species of the genus more or less poisonous. Among these are the *H. Bigelovii* of Oregon, California, and *H. nudiflorum* from Texas to Kansas, Illinois and Florida.

Helenium Hoopesii Gray

A stout perennial from 1-2 feet high, tomentose or pubescent, when young becoming smooth with age; leaves rather thick, entire oblong lanceolate, the lower spatulate and tapering base, somewhat nerved; heads large, single or several; ray flowers fertile, yellow, an inch long; soon reflexed; bracts of the involucre in 2 series.

Distribution. Common in the Rocky Mountains to eastern Oregon to California. It covers large stretches between 6,000 and 8,000 feet in the Uintah mountains.

Poisonous nature. It is said to be poisonous like other species of the genus. Sheep carefully avoid it, feeding on the grass and other herbaceous plants, leaving the plant standing.

Helenium autumnale L. Sneeze-weed

A smooth or pubescent perennial from 1-6 feet high; leaves toothed, lanceolate to obovate, oblong decurrent on the stem, acuminate at the apex; heads numerous, many flowered; radiate; rays yellow, wedge shaped, 3-5 cleft, fertile; disk flowers yellow, involucre small, reflexed; scales linear or awl-shaped; receptacle glabrous or oblong; achenes pubescent on the angles; scales of the pappus ovate or acuminate; the variety grandiflorum with larger heads; scales of the pappus narrowed and longer awned.

Distribution. Common in low meadows and alluvial banks from eastern. Canada to South Dakota, Florida to Alabama, and especially common in the northern Mississippi Valley to Kansas. Occurs also in the Rocky Mountains, Wyoming to Montana. The variety in Washington to California.

Poisonous properties. This plant is considered poisonous to cattle and sheep. Numerous complaints have been made in various parts of the state of Iowa in regard to the poisonous nature of this weed. This plant is more or less bitter, acrid and pungent, especially the flowers. Parts of the plant

rubbed were used by the Winnebago Indians for colds and to produce sneezing. Dr. Chesnut says:



Fig. 448. Sneeze-weed (Helenium autumnale).

Sheep, cattle and horses that are unfamiliar with the plant are often poisoned by it when driven to localities where it is abundant. As a rule these animals avoid it, but it is said they sometimes develop a taste for it and are quickly killed by eating it in large quantity. The poisonous constituent has not been closely investigated, but it is known that it exists principally in the flowers. The young plants appear to be only very slightly dangerous. In the mature ones the amount of poison present seems to vary greatly even in the same field. The symptoms as determined by experiments made in Mississippi upon calves, are an accelerated pulse, difficult breathing, staggering, and extreme sensitiveness to the touch. In fatal cases, death is preceded by spasms and convulsions.

Melted lard, when given before the spasms begin, has been used with good effect in offsetting the action of the poison. The sneeze weeds are said sometimes to taint milk, imparting to it a bitter flavor.

Dr. Phares, in an early account of the poisonous nature of the plant states: Experiment 1.—Calf about four months old and not fed in the morning. Upon three ounces of the dried plant was poured a pint of boiling water. After an hour the water was decanted with pressure. Mr. J. W. McWilliams, of the senior class, gave all the doses in all the experiments and noted the symptoms. October 6th the above infusion was ordered to be divided into four parts and one part given every half hour, beginning at 11:10 a. m. The first dose produced no very marked effect—only a flow of a few tears. After second dose became restless, later laid down. After third dose breathing impaired, continued lying down and appears sicker for a short while. Fourth dose, breathing harder,

COMPOSITAE — THISTLE FAMILY — SNEEZEWEED 783

but at 3 p. m. the calf was resting easy. Next day he seemed to be nearly as well as usual and took his feed.

Experiment 2.—October 13th; same calf. A hot infusion of six ounces of the drug was prepared in one quart of water and given between 8 a. m. and noon. Soon after the first dose some weeping began, but no other marked effect was observed. After second dose the calf laid down. Third dose, he continues down, but somewhat restless and sick. Fourth dose, becomes very restless, bowels loose, staggering, and very sick, holding head down, sucking wind, left flank tympanitic but not very tense, right flank contracted, trying to put head between forelegs, and very sick greater part of the day; movements weak and uncertain, purposeless, hyperesthesia very marked, especially along the back, shrinking from least touch, but by persistent handling submits to touch, respiration and pulse quickened, appearance of slight spasm along the back, neither eats nor drinks of food and water and milk in his stall, a little straw from floor in his mouth, and although the jaw is in constant motion the straw is undisturbed and he seems unconscious of its presence. Next day apparently well.

Experiment 3.—October 20th; another calf about four months old. One pound of the dry plant was taken, one gallon water poured on it and boiled down to a quart; all given by 10:35 a. m. After third dose the left flank tympanitic, feces and urine discharged, slight jerking as though chilled, tremor of muscles of hindquarters, lying on right side over an hour, breathes with some noise and distress, very sensitive from withers to sacrum; at 11 o'clock fullness of left flank subsided, limbs showed weakness and later stiffness, very sick, frothing at mouth and flow from nose. Next day the calf was well.

The large quantity of the drug given this calf producing little more effect than the smaller dose on the former one leads me to suppose that the prolonged boiling may have dissipated a portion of the poisonous principle. Knowing that none of the symptoms observed would lead on to death no antidote was administered in any of the experiments. The weeping and movtment of the jaw were probably due to the pungent effect of the drug on the throat and stomach; and I incline to the opinion that the nervous symptoms were not caused by direct action of the drug on nerve centers, but were reflexes for the irritated throat and stomach.

In the sheep the spasins in severe cases are epileptiforin and a sheep may have many such convulsions and yet recover without treatment and after many hours rise up and walk.

The horse and mule succumbed to the injurious effects of the poison quicker and more completely than other animals. The effect is manifested very soon after ingestion and with great violence. The animal cannot control his motions, plunges about blindly, falls dead or perhaps breaks his neck in falling forward with the head under the body.

An antidote which has proved effective is a pint or two of melted lard poured down the animal's throat. The relief is very prompt. But it must be given before the horse loses control of his limbs, or it cannot be administered at all. The relief is so prompt and complete that it is difficult to believe that it is the direct effect of the lard on the nerve centers, but as a local emollient applied directly to the burning throat and stomach. The local trouble being quickly relieved, the violent reflex effect on the nervous system suddenly ceases.

When we consider the wide distribution of this plant and that in some places it is so abundant as to hide the earth from view over whole acres, it is remarkable that so few poisonings occur. But animals have no special fondness for it. When they have been long on the road and deprived of green food, stopping where this weed is found and other green plants scarce, they are disposed to eat a little of it. It is under such conditions that the plant is most frequently eaten and the poisoning occurs.

Animals, notably sheep, once intoxicated by the plant and recovering, seem to acquire a mania for it, and when finding themselves in localities where it grows hurry to and devour it.

Within 200 miles of the Gulf of Mexico the H. quadridentatum takes the place of the preceding, is very abundant, harmless, blooms early and dies by August.

Helenium tenuifolium Nutt. Narrow-leafed Sneeze-weed.

A perennial, erect branching herb from 1½-2½ feet high; leaves alternate decurrent on the stem, linear filiform; flowers borne in corymbed clusters; heads many flowered, ray flowers yellow, fertile, disk-shaped, ribbed; pappus of 1-nerved chaffy scales, the nerve extended into a point.

Distribution. Common from Texas to Missouri, Florida and Virginia, especially in pastures and meadows and along roadsides. It is said to have been widely scattered in the south after the war of the rebellion.

Poisonous properties. It is often the cause of bitter milk in the south. Dr. Chesnut says:

The fine-leaved Sneezeweed has been reported from several of the Gulf States, where it is a troublesome weed, fatal to horses and mules. It is not known to what extent cattle may feed on the plant with impunity, but the bitter principle in milk and meat sometimes met with in the Southern States is quite generally supposed to be due to these plants.

It contains a narcotic poison.



Fig. 449. Fine-leaved sneezeweed (Helenium tenuifolium). Often the cause of "bitter-milk" (Dcwey, U. S. Dept. Agr.)

18. Dyssodia Lag. Fetid Marigold

Mostly annual or biennial herbs with strong scent, dotted with large pellucid glands; leaves mostly finely dissected; heads many-flowered, small, of both tubular and ray flowers; involucre cylindrical or hemispherical, bracts in one series united into a cup; receptacle flat, not chaffy, but with short bristles; ray flowers pistillate, short; disk flowers perfect; achenes slender, 4-angled; pappus a row of chaffy scales dissected into rough bristles. A single species.

Dyssodia papposa (Vent.) Hitchc. Fetid Marigold.

A nearly smooth branched annual from 6 inches-2 feet high; leaves opposite, sessile, pinnately parted, bristly-toothed with large pellucid glands, which give to the plant a strong odor. Heads many-flowered; rays small, yellow, pistillate; disk yellow; involucre with a few scales at the base, one row of scales united to form a cup; achenes slender, 4-angled, pappus a row of chaffy scales finely divided into numerous rough bristles.

Distribution. Common from western Iowa and Minnesota to Illinois and the southwest. In this section of the country it occupies the same ground occupied by the Mayweed.

Poisonous properties. It is a very disagreeable smelling herb. Not relished by stock. It is probably injurious.

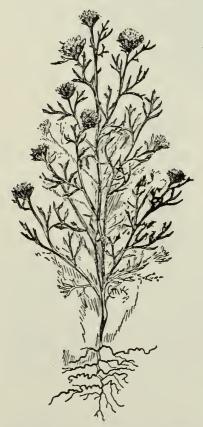


Fig. 450. Fetid Marigold (Dyssodia parposa). Pungent odor said to be injurious. (Charlotte M. King.)



Fig. 451. Dog Fennel (Anthemis Cotula). Contains a pungent principle.

19. Achillea (Vaill.) L. Yarrow

Perennial herbs with erect leafy stems and finely dissected alternate leaves; heads many flowered, radiate, in corymbose clusters, ray flowers few, white-

or pink; disk flowers perfect, fertile; corollas yellow and 5-lobed; receptacle nearly flat; achenes oblong or obovate, slightly compressed; pappus wanting.

About 75 species mostly of the Old World. The yarrow (Achillea Ptarmica) occasionally cultivated.

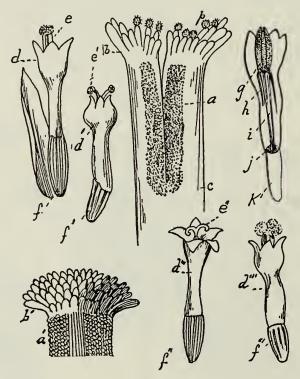


Fig. 451. Yarrow (Achillea Millefolium). Structure of flower. a, stigmas; b, papillae; c, style, pollen; d, single flower; e, f, g and f, flowers in different stages. (Müller).

Achillea Millefolium L. Yarrow.

Perennial herb from 1-2 feet high; simple stems; leaves twice pinnately parted, consisting of crowded linear divisions; flowers in compound flat-topped corymbs; heads many-flowered, radiate; rays 4-5, white; scales of the involucre with scarious margins; receptacles chaffy; disk flowers small, tubular; achenes oblong, flattened; pappus none.

Distribution. A common weed from the Atlantic to the Pacific. In the East it is found in old fields and dry hills and is abundant in open parks and rocky dry soils in the Rocky Mountains up to the timber line. It is native also to Europe, where it is used as an ornamental plant. Sheep will eat the weed. The form with rose-colored ray flowers is frequently cultivated.

Poisonous properties. The alkaloid achillein C₃₀H₃₈N₂O₁₅ has been isolated from the plant. Millspaugh describes the action of the drug as follows: "Yarrow seems to have a decided action upon the blood vessels, especially in the pelvis. It has been proven to be of great utility in controlling hemorrhages, especially of the pelvis, viscera, where hemorrhage is caused by it. Its common

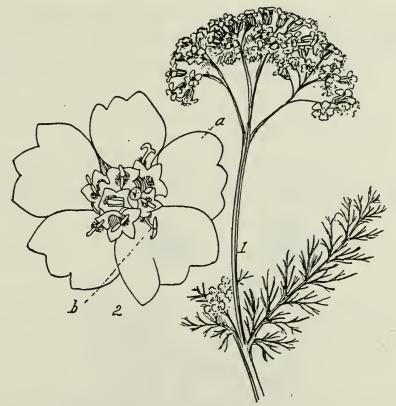


Fig. 451a. Yarrow (Achillea Millefolium). 1, plant with leaf and flowers. 2, A single head; a, ray flowers; b, a single flower. (Charlotte M. King).

European name, Nosebleed, was given from the fact that the early writers claimed hemorrhage of the nose followed placing its leaves in the nostrils; this may have been either due to its direct irritation, or the use of Achillea Ptarmica, its leaves being very sharply serrate and appressed toothed. Millefolium causes burning and raw sensations of the membranes with which it comes in contact, considerable pain in the gastric and abdominal regions, with diarrhoea and enuresis. An alkaloid having the same formula as achillein has been isolated in A. moschata; a second alkaloid, moschatin, $C_{21}H_{27}NO_7$ is said to occur in the same plant. In Europe sometimes regarded as a forage plant.

20. Anthemis (Mich.) L. Mayweed

Annual or perennial herbs with finely dissected leaves and a strong scent; heads peduncled; involucre hemispherical; bracts imbricated in several series; ray flowers white or yellow, 2-3 toothed; pistillate and fertile; style branches of the disk flowers truncate; achenes oblong angled, ribbed; pappus none or short crown. There are about 60 species in the Old World. They are strong scented or aromatic herbs.

Anthemis Cotula. L. Mayweed. Dog Fennel

An acrid branching scented annual from 1-2 feet high; leaves thrice pin-

nately dissected; heads solitary, many flowered; ray flowers white, pistillate, fertile or neutral; involucre of numerous small imbricated dry and scarious scales; disk flowers yellow, small, tubular; achenes terete or ribbed, smooth, truncate; pappus none or a minute crown.

Distribution. A common weed in Europe where it is a native. Widely scattered in the Northern States from the Atlantic to the Dakotas and Nebraska. Occurs also in Washington and Oregon. In the north Mayweed is common in yards and along roadsides.

Poisonous properties. The strong odor and acrid taste makes it extremely undesirable for stock. The Mayweed blisters the skin.

Anthemis arvensis L. Corn Chamomile

A much branched annual or biennial pubescent herb, not ill-smelling; leaves sessile, finely 1-2 pinnately parted, less divided than the preceding; heads numerous; bracts of the involucre oblong, obtuse, usually somewhat pubescent with

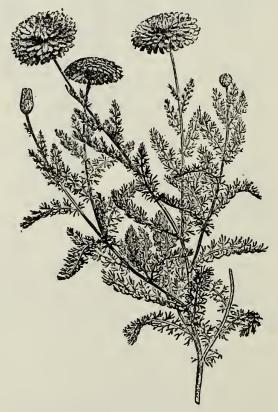


Fig. 452. Anthemis nobilis. Cultivated as an ornamental plant. Faguet.

broad scarious margins; ray flowers white, pistillate, mostly 2-toothed; chaff of the receptacle obtuse; achenes oblong, obtusely 4-angled; pappus a small border.

Distribution. In fields and waste places in the Eastern States. Nova Scotia to Virginia, Missouri, and Wisconsin to the Pacific coast.

Poisonous properties. The oil of Anthemis nobilis contains the terpene anthemin $C_{18}H_{80}$, angelic acid $C_5H_8O_2$, tiglinic acid $C_5H_8O_2$. The oil produces abdominal pain, increase of heart action and freeness of the bowels. It is not known that our species is poisonous but stock carefully avoid the plant.

21. Chrysanthemum (Tourn.) L. Ox-eye Daisy

Perennial or annual herbs; leaves alternate, dentate, pinnatifid or dissected; heads single or corymbose, or both ray and tubular flowers rarely wanting; involucre with broad imbricated scales; receptacle flat or convex; ray flowers pistillate and fertile, the rays white; style, branches of the pistillate flowers, truncate and penicillate; achenes angled ar terete, striate with pappus.

About 100 species of wide distribution. The common fever-few (Chrysanthenium Parthenium) is frequently cultivated in Germany and is rarely an escape in eastern North America. It contains the oil of feverfew.

Insect powders are derived from several species of Chrysanthemum, the Dalmatian from C. cinerariifolium cultivated in Europe, North Africa, New York and California; the C. coccineum Willd, and C. Marshallii, found in the region of the Caucasus, furnish the Persian insect powders.

Chrysanthemum Leucanthemum L. Ox-eye Daisy

A perennial herb with erect stem and spatulate, petioled root leaves, those of the stem partly clasping; all the leaves cut or pinnatifid toothed; the nearly simple stem bears a large many-flowered head with numerous white rays, scales of the involucre with scarious margins; disk and ray flowers pistillate and fertile; achenes marked with longitudinal lines, pappus absent.

Distribution. Common in fields and meadows of the eastern states and eastern Canada, less common in the middle and western states.

Poisonous properties. This abundant weed probably contains an active principle. The C. coccineum contains the alkaloid chrysanthemin C₁₄H₂₈N₂O₈. The flowers are more or less irritating.

22. Tanacetum L. Tansy

Bitter acrid strong-scented aromatic annual or perennial herbs; leaves alternate, 1-3 pinnately dissected and corymbed heads; involucre hemispherical, depressed, scales imbricated into several series; rays wanting or present, yellow marginal flowers pistillate and fertile; corollas 5-toothed; style branches truncate and penicillate; achenes angled or ribbed; pappus a short crown. A small genus of 35 species, natives of the northern hemisphere. The *Tanacetum balsamita* contains a balsamic oil.

Tanacetum vulgare L. Tansy

A bitter acrid strong scented simple or branched perennial herb from 2-4 feet high; leaves pinnately divided into linear pinnatifid divisions, lobes serrate; heads corymbose, many flowered; ray flowers few, disk flowers yellow; marginal flowers fertile, scales of the involucre imbricated in several series; receptacle flat or convex, naked; branches of the style brush-like at the summit; achenes 5-angled or 5-ribbed, truncate or obtuse; pappus none or a short crown.

Distribution. Native to Europe, frequently escaped from gardens to roadsides along the Atlantic Coast as far west as Iowa and Kansas.

Poisonous properties. The oil of tansy is obtained from this plant. The

bulk of the oil consists of tanaceton or thujon $C_{10}H_{15}(OH)$. This oil has been used as an anthelmintic since the middle ages. Tansy is a very poisonous, producing a condition similar to rabies. From this plant many serious cases of poisoning in the human race occur, but poisoning of animals is very infrequent. It is an irritant narcotic poison. Dr. Millspaugh cites a few cases as follows:

A young woman had been in the habit of using tansy tea, made from the herb, at nearly every menstrual period, for difficult menstruation. On this occasion about two and a half drachms of the oil was poured into half an ordinary tin cupful of water; this, with the exception of a small portion of the water containing about one-half drachm of the oil, was taken at one dose. Convulsions were almost at once produced, and when Dr. Bailey was sent for the patient was unconscious, foaming at the mouth, and in violent tonic spasms, with dilated pupils, frequent and feeble pulse. Constant kneading on the stomach had produced partial emesis, and then ipecac, mustard, and large draughts of hot water, emptied the stomach. Two drachms of magnesia were then given, and a full dose of acetate of morphine; consciousness then returned, no unfavorable symptoms followed, and, after thirty-six hours, without additional medication she was entirely restored.

A woman took half an ounce of the oil; the most violent rigid kind of clonic spasms occurred once in about twelve minutes, coming on generally and instantly, and continuing about one minute. They were attended with slight, if any motion of the arms; it might be called a trembling. The arms were peculiarly affected, and invariably in the same way; they were thrown out forward of, and at right angles with, the body; the hands at the wrists bent at right angles with the fore-arm supinated, the points of the fingers nearly in contact, the fingers straight and slightly bent at the metatarso-phalangeal joints. The muscles of respiration were strongly affected during each paroxysm; the air was forced from the chest slowly but steadily, and made a slight hissing noise as it escaped from between the patient's lips. During the intermission of spasm, the muscles were perfectly flexible, and the transition seemed very sudden. The jaws were the only exception to this rule; they were, for the first hour and a quarter, rigidly closed, and were with difficulty opened, but after that they were subjected to the same action as the rest of the body—when the spasms were on they were rigid; when off, they were relaxed. After the patient grew weaker, the spasms were more frequent, but had about the same severity and length. Death ensued in two hours.

On animals the symptoms are as follows according to a statement condensed by Dr. Millspaugh:

Dr. Ely Van DeWarker records cases of the action of the oil upon dogs. In one case two drachms were given, causing salivation, vomiting, dilation of the pupils, muscular twitchings, followed by clonic spasms, and a cataleptic condition from which the animal recovered. Recovery also followed a half ounce after the same class of symptoms, but, however, on repeating the dose, the already poisoned animal was plunged into a long and fatal convulsion. Postmortem examination disclosed the cerebral veins and spinal cord itself highly congested, and serous effusions had taken place in the pia mater. The lungs were found to be engorged, the left heart empty, and the right distended with dark, liquid blood. Congestion of the kidneys had also taken place, and the bladder was found contracted.

The safe maximum dose of the oil is indeterminable, a few drops only sometimes proving serious.

The symptoms occurring in a number of cases of poisoning and experiments were substantially as follows: Mental confusion, loss of consciousness; vertigo, with cephalalgia, at first contraction, then wide dilation, of the pupils, staring, immovable eye-balls; ringing in the ears; face congested; roughness of the mouth and throat; difficult deglutition; eructations, nausea, free vomiting, and burning of the stomach; sharp colic pains in the abdomen; diarrhoea; constant desire to urinate—urine at first suppressed, then profuse; respiration hurried and laborious; pulse at first high, then very low and irregular; numbness of the extremities; tonic and clonic spasms, and nervous tremblings; drowsiness and cold sweat. Death appears to ensue from a paralysis of the heart and lungs,

23. Artemisia L. Wormwood, Sage Brush

Bitter and aromatic herbs or shrubs with alternate leaves; heads discoid, few or many flowered, collected in panicles, racemes or spikes, with greenish or yellow flowers; involucre ovoid or oblong, scales imbricated, the outer shorter; receptacle small, naked; marginal flowers pistillate, disk perfect, or

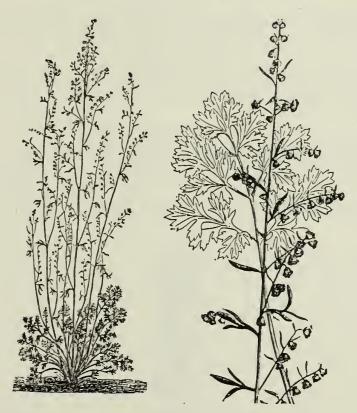


Fig. 453. Artemisia. Sage Brush. Absinthe and Wormwood belong to this geniue. (After Faguet).

perfect and fertile with branches of the style truncate or sometimes sterile with ovaries abortive; styles undivided; marginal flowers usually pistillate and fertile, or flowers all perfect and fertile in some species; anthers often tipped with subulate appendages; achenes obovoid and no pappus.

About 200 species, mostly native of the northern hemisphere. A few in southern South America. The oil of Levant wormseed (A. maritima var. Stechmanniana) contains the substance cineol. Santonin $C_{15}H_{78}O_3$ is the active principle found in the flowers of A. cana. The A. maritima contains artemisin $C_{15}H_{18}O_4$. Absinthe is furnished by Artemisia absinthium of Europe. The A. Barrelieri furnishes the Algerian absinthe. Several species of sagebrush (Artemisia tridentata and A. cana) are common in the west and much used as forage for sheep. The alkaloid abrotanin $C_{21}H_{22}N_2O$ is obtained from A. Abrotanum.

Artemisia biennis Willd, Wormwood

An aromatic, somewhat bitter, smooth annual or biennial, from 1-3 feet high, with leafy stems and erect branches; lower leaves twice pinnately parted, the upper pinnatifid; the lobes linear or linear-oblong, serrate or cut-toothed; ray flowers absent; heads numerous in short axillary spikes; the bracts of the involucre green, scarious, margined.

Distribution. Common in the northern Mississippi Valley, now widely scattered east to Nova Scotia and south to Kentucky.

Artemisia tridentata Nutt.

From 2-12 feet high, much branched, silvery canescent; leaves cuneate, obtusely 3-toothed or 3-lobed, or 4-7 toothed at the summit, upper leaves cuneate-linear; heads homogamous; flowers all perfect and fertile; heads densely paniculate; 5-8 flowers.

Distribution. From Montana and Colorado westward covering immense areas. The characteristic sage brush; similar species are the A. trifida, Nutt, a smaller plant common westward, and the A. arbuscula, Nutt., also a dwarf plant found in the high mountains and elevated plains; the A. cana, Pursh, from 1-2 feet high; silvery canescent leaves; found from Saskatchewan to Dakota, Montana and Colorado.

Poisonous properties. The sage brush is consumed by sheep and cattle,



Fig. 454. Wormwood. (Artemisia biennis). A common weed in the Mississippi Valley. Ada Hayden.

although the latter do not relish the plant. The inhalation of the dried powder of the plant causes violent sneezing. Chesnut and Wilcox, in speaking of the sage brush of Montana, say that some species are recognized by stockmen as valuable forage plants for the late fall and winter grazing. Various species are suspected in Montana of being poisonous to stock, but no specific cases have been reported to the department. At Toston in May, 1900, it was noticed that the terminal branchlets of A. tridentata had all been caten off from a considerable number of plants.

A. Absinthium L. Common Wormwood. Absinth.

A shrubby, silky, hairy plant from 2-3 feet high; leaves 2-3 pinnately parted, the lobes lanceolate, obtuse; heads in racemose panicles, nodding yellow, marginal flowers pistillate, fertile or sterile, central flowers fertile; involucre hemispherical outer bracts linear.

Distribution. Native to the old world but escaped from gardens and found along roadsides. From Newfoundland to Massachusetts and North Carolina; occasionally westward to Wisconsin and Montana.

Poisonous properties. Wormwood is a stimulant and tonic and has been employed chiefly for digestive disorders, although seldom used medicinally. The volatile oil of wormwood in large doses produces cerebral disorders, convulsions and even death. The well-known absintle is made from this plant and used with alcoholic drinks. For this purpose the plant is extensively cultivated in Europe and to some extent, in Nebraska, Michigan and Wisconsin. Absinthe contains the principle absinthin C₁₅H₂₀O₄. Dr. Rusby, in speaking of absinthe says that absinthium is in a general way similar in composition and properties as a poison to Tanacetum vulgare though the symptoms are more largely cerebral. Neither is it liable to be taken except for medicinal purposes or as an addition to spirits. As a result of the latter, poisoning is usually chronic and extremely difficult to cure. Dr. Millspaugh reports the experience of a clerk who took about a half ounce of the oil; he was found on the floor perfectly insensible, convulsed, and foaming at the mouth; shortly afterward the convulsions ceased, the patient remained insensible with the jaws locked, pupils dilated, pulse weak, and stomach retching. After causing free emesis and applying stimulants, the man recovered, but could not remember how or when he had taken the drug. According to Dr. Legrand, the effects prominent in absinthe drinkers are: Derangement of the digestive organs, intense thirst, restlessness, vertigo, tingling in the ears, and illusions of sight and hearing. These are followed by tremblings in the arms, hands, and legs, numbness of the extremities, loss of muscular power, delirium, loss of intellect, general paralysis, and death. Dr. Magnan, who had a great number of absinthe drinkers under his care, and who performed many experiments with the liquor upon animals, states that peculiar epileptic attacks result, which he has called "absinthe epilepsy."

Post-Mortem.—Great congestion of the cerbro-spinal vessels, of the meninges of the brain, extreme hyperaemia of the medulla oblongata, injection of the vessels of the cord, with suffusion of the cord itself. The stomach, endocardium, and pericardium show small ecchymoses.

Absinthe is sometimes added to hops to make beer more exhilirating.

A. vulgaris L. Common Mugwort

Tall, branching perennial with fine and closely appressed hairs; leaves mostly

smooth and green above, but quite woolly underneath; pinnatifid, the divisions cut-toothed, linear-lanceolate; heads small in open leafy panicles.

Distribution. In waste places from Canada to North Carolina to Pennsylvania and occasionally westward.

Poisonous properties. The common mugwort was used medicinally by the physicians of the 16-18 century, but in modern times the plant is not used to any extent. Mugwort causes epileptic spasms, profuse sweat with the odor of garlic; violent contractions of the uterus; labor-like pains; prolapsus and rupture of the uterus; miscarriage; metrorrhagia and increase of lochial discharges.

24. Arnica L.

Perennial herbs, erect, simple or branching; leaves opposite, or rarely with the upper alternate; heads many-flowered, radiate on long peduncled heads; scales of the involucre bell-shaped; bracts in 1-2 series; the ray flowers pistillate and fertile; disk flowers perfect and fertile; achenes linear, 5-10 ribbed; pappus of roughened-denticulate bristles.

About 30 species native to the Northern Hemisphere. The tincture of arnica is a well-known house remedy used for bruises and chilblains. A. montana contains a bitter principle arnicin C20H38O4. The value of arnica as a germicide has been largely overestimated. The germicidal action is probably due to the presence of alcohol. Dr. White, in his Dermatitis Venenata, records several instances of injurious effects from the use of arnica. In one case a gentleman descending the stairs to mount his horse for a ride, slipped and scraped the lower part of his back. A handkerchief dipped in the tincture was applied to the bruised skin and worn in contact with the part during the ride. During the ride he felt considerable itching and upon his return home found that the skin was greatly congested. On the next day I was called to see him. The skin of the back, nearly to the shoulders, was in a state of active hyperaemia, and already covered with innumerable papules. The inflammatory process extended rapidly downwards nearly to the knees, and forward upon the abdomen and genitals. In a few days these parts presented all the characteristic appearances of acute eczema in its various stages of progression: general hyperaemia, papules, vesicles, excoriated and exuding surfaces, and crusts. The subjective symptoms were intense itching, stinging, and burning in the parts. Scarcely any clothing could be borne in contact with the skin by day, and sleep was for a few nights almost impossible, but the system generally was only slightly disturbed. Dr. White records two other cases. All three seem to have been old gentlemen and he states: "In all of them we have an acute inflammatory process, confined to the upper dermal layers, and manifesting itself, according to the stage reached, by the following appearances: hyperaemia, papules, vesicles, excoriations, crusts, and scales, in regular sequence. The local sensations were intense itching and some degree of burning in the parts affected."

Arnica cordifolia Hook. Arnica

A low perennial, 1-2 ft. high or less; pubescent, or the stem hirsute and peduncles villous; the root and leaves deeply cordate or ovate at the base, on slender petioles; stem leaves opposite, in 1-3 pairs, dentate or denticulate; upper small, sessile; heads few, solitary; involucre pubescent or villous; rays an inch long; achenes more or less hirsute.

Distribution. Higher altitudes of the Rocky Mountains, from Colorado to California and British Columbia.

Arnica mollis Less. Hook.

A leafy branching perennial, tomentose or villous-pubescent or nearly smooth; leaves denticulate or dentate, acute or obtuse; lower leaves tapering into a margined petiole, upper broader at the base and clasping; flowers much smaller than the preceding; ray flowers yellow; achenes hirsute-pubescent.

Arnica monocephala Olin.

An erect, simple, branching perennial, from 1-2 feet high; leaves opposite; long peduncled heads; leaves narrow, oblong and lanceolate, the upper linear; ray flowers, yellow.

Distribution. Grows in woods and moist localities, Montana to New Mexico and westward.

Poisonous properties. This well-known arnica has, for some time, been suspected of being poisonous. When bruised all the plants exhale the odor of arnica. Chesnut and Wilcox state:

It has an odor and taste much like that of the official drug, and, in fact, an extract made from the heads is considerably used locally as a liniment. No specific cases of poisoning caused by the plant have been reported from Montana, and we are not aware that it is eaten, but it has, nevertheless, been suspected of being poisonous to stock.

Dr. S. B. Nelson states that a sheep ate 2 pounds (of the leaves?) of A. alpina within a day without experiencing any bad effect. The plants had been gathered eighteen hours.

25. Senecio (Tourn.) L. Groundsel

Perennial or annual herbs; shrubs or trees; leaves alternate; heads many flowered, solitary, corymbose or paniculate; involucre cylindrical or bell shaped; principal bracts in one series, distinct or united at the base, usually with some shorter rays, outer ones pistillate or none; disk flowers perfect; corolla tubular; achenes terete or those of the marginal flowers compressed; 5-10 ribbed; pappus of numerous capillary bristles. A large genus of 1000 species of wide distribution. Some ornamental. Some species have been looked upon as poisonous.

Dr. Day, in the Twenty-third Annual Report of the Bureau of Animal Industry, 1906, gives the following in regard to a disease supposed to be associated with a species of this genus:

At the present stage of the work there appears reason for associating this malady with an affection of cattle known in Nova Scotia as Pictou disease, with Winton disease of horses of New Zealand, and with Molteno cattle disease of South Africa. The tissues examined present practically the same lesions as those described for the above-mentioned affections—namely, interlobular connective tissue hyperplasia, fatty infiltration of the liver cells, and hemorrhages into the spleen.

Pictou disease, Winton disease, and Molteno cattle disease have been investigated time and again in an endeavor to find a causative agent. All inoculation experiments have failed, and it has only been found of late that feeding experiments with certain weeds have given seemingly reliable results.

Pictou disease, occurring in Pictou and Antigonish counties, Nova Scotia, is now ascribed to the ingestion of a weed called ragwort. Winton disease is thought to be caused by a member of the ragwort family, Senecio Jacobaea. The Molteno disease of South Africa is probably induced, according to recent investigations, by still another ragwort, Senecio Burchelli. At least these weeds when fed to animals in their respective localities have produced diseases followed by a train of symptoms corresponding to those seen in the naturally induced disease. The post-mortem findings were also quite similar. It was also proved that these plants were most dangerous in the young state. Yet another variety known as Senecio latifolius was found to he deadly to stock.

In some parts of the states in the west Senecios are common but they are not

commonly found when this forage is cut for hay. It is extremely doubtful whether the Senecio occurring in the western part of Iowa is responsible for this disease. The groundsel or mug-wort (Senecio Balsamitae) blooms in May



Fig. 454a. Burdock (Arctium Lappa). 1, general view of plant; 2, single flower; 3, scales of involucre hooked. These heads annoying to stock. (U. S. Dept. Agrl.)

797

and after blooming disappears except a few of the root leaves. The stinking Willie (S. Jacobaea) is naturalized in the east and of course may be responsible for the Pictou disease,

26. Arctium L. Burdock

Large coarse biennial, rough or pubescent herbs; leaves broad, alternate and petioled; heads large, recemose, corymbose or paniculate; flowers purple or white; all tubular perfect and similar; involucre globular; the scales imbricated; lanceolate, tipped with hooked points; receptacle flat, bristly; achenes oblong, somewhat compressed and 3-angled and wrinkled; pappus short, of numerous short bristles.

A small genus of 5 or 6 species in the old world. The root of the common burdock is used in medicine, while in Japan it is also used for food.

Arctium Lappa L. Burdock

A coarsely branched biennial from 1-3 feet high, hairy; leaves large, roundish or heart-shaped, thin obtuse, entire or dentate, floccose tomentose beneath; petioles deeply furrowed; heads of purplish or whitish flowers, clustered or somewhat corymbose; the involucre lengthened into hooked tips, glabrous or slightly cottony.

Distribution. Common from New Brunswick westward in the Canadian provinces in Canada, southward to Alabama, general in the Mississippi Valley, Rocky Mountain region, and Utalı.

Poisonous and injurious properties. Where the burdocks are allowed to grow freely the burs cling to the fleece of animals, often forming large balls which may be found on the tails of cattle and horses. When they are common where sheep pasture, the burdocks work into the wool which soon becomes of little value. Formerly burdock was used medicinally more than it is at the present time. The root has diuretic properties. Dr. Johnson gives the chief medical properties as follows:

Burdock, though chiefly used by the laity, is highly esteemed by some of the profession as a diuretic and alterative. It is, perhaps, more frequently used in rheumatism than in any other disease, but is also recommended in chronic cutaneous diseases, catarrhal affections, syphilis, and scrofula. In the form of an ointment or liniment it is used as an application to burns, ulcers, etc.

Some forms of the burdock are used for food; the young stems when stripped of their rind may be eaten raw or boiled, or in a pottage with vinegar.

27. Cirsium (Tourn.) Hill. Thistle

Perennial, annual or biennial, erect, branched or simple, herbs; leaves sessile, alternate, often pinnatifid, prickly; heads usually large, many flowered, solitary or clustered; heads discoid, flowers purple, yellow or white, tubular, perfect and fertile or dioecious; involucre ovoid or globose, scales imbricated in many rows, often glutinous, tipped with a prickle or unarmed; receptacle flat or convex, bristly; corolla tube slender, the limb deeply cleft; anthers sagittate at the base; style elongated, branches short or obtuse, unappendaged, often with a pubescent ring below; achenes obovate or oblong compressed or obtusely four-angled, glabrous, often ribbed; pappus plumose or minutely serrulate.

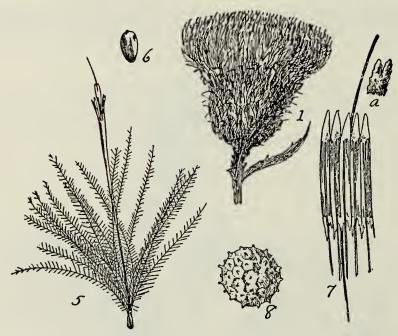


Fig. 455. Woolly Thistle (Cirsium canescens). 1, Head; 5, Flower and pappus; 6, Achene; 7, Anthers and style cut open; 8, Pollen grain; a, end of style. (Charlotte M. King).

About 200 species of the north temperate regions. Some of them are cultivated for ornamental purposes; many of the species are pleasantly scented; many of them are troublesome weeds. $Cnicin\ C_{20}H_{37}O_{10}$ has been found in a related genus ($Cnicus\ benedictus$). Horses are fond of the heads of many species like $C.\ eriocephalum,\ C.\ Drummondii,\ C.\ undulatus$ and others that occur on the Rocky Mountains. None of the species are probably poisonous but many of them have irritating properties. The Canada thistle is used in domestic practice.

Cirsium arvense (L.) Hoffm. Scop. Canada Thistle

Smooth perennial herbs, spreading by creeping rootstocks, 1-3 feet high, corymbosely branched at the top; stem smooth; leaves lanceolate, sessile and deeply pinnatifid, lobes and margins of leaf with spiny teeth; heads small, 34-1 in. high; bracts appressed, the outer with a broad base, inner narrow, all with an acute tip, never spiny; flowers purple, dioecious; in staminate plant, flowers exserted with abortive pistil; in pistillate, less so, scarcely exceeding the bracts; long stamens with abortive anthers, tube of the corolla 6 lines long, anther tips acute, filaments minutely pubescent, all of the bristles of the pappus plumose.

Distribution. The Canada thistle is found in waste places from Newfoundland, Nova Scotia, various provinces of Ontario, to New York, Virginia, southwest to Missouri and Kansas, Colorado to Idaho, Montana and Oregon.

Cirsium lanceolatum (L.) Hill. Bull Thistle

Branching biennial, 3-4 feet high, tomentose, becoming dark green and villous or hirsute with age, branchlets bearing large heads; leaves lanceolate,



Fig. 456. Canada Thistle (Cirsium arvense). Common in the North, causes inflammation. (Charlotte M. King). decurrent on the stem with prickly wings, deeply pinnatifid, the lobes with rigid

prickly points, upper face roughened with short hairs, lower face with cottony tomentum; heads 1¾ to 2 inches high, bracts of the involucre lanceolate, rigid when young, more flexible with age, long attenuated prickly pointed spreading tips, arachnoid woolly; flowers perfect; anther tips acute; filaments pubescent; achenes smooth; pappus of numerous plumose bristles.

Distribution. Distribution in North America in fields and waste places from Newfoundland to Georgia, Missouri, Kansas to Nebraska, the Dakotas, Montana, Washington, Oregon and Idaho. Naturalized from Europe, native also to Asia.

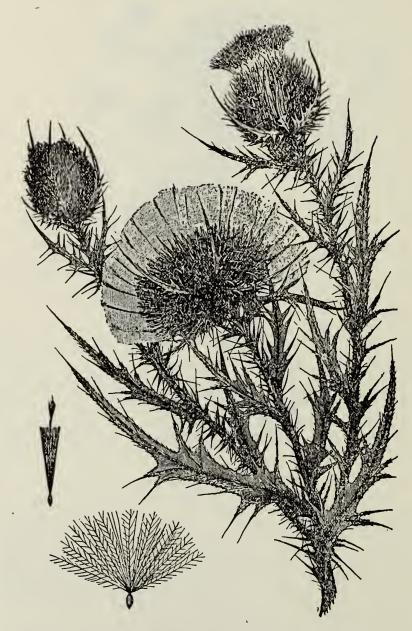


Fig. 457. Bull thistle (Cirsium lanceolatum). A common pasture weed. Causes mechanical injuries. (U. S. Dept. Agrl.)

COMPOSITAE — THISTLE FAMILY — BULL THISTLE 801

Cirsium canescens Nutt. Woolly Thistle

Branching perennial 2-4 feet high, woolly throughout bearing single medium sized heads, stem angled, white woolly; leaves, radical 8 inches to 1 foot long, the divisions usually 2-lobed, prominently ribbed, ending in stout spines; stem leaves except the lower, 1-4 inches long, pinnatifid, the upper sessile, slightly roughened, with a slight cottony down, the lower white, woolly; heads 1½ to 2 inches high, bracts of the involucre somewhat arachnoid, lower scales with a broad base, glutinous ridge, and ending in minutely serrated spine, inner scales long, attenuated, tips straw colored; flowers purple. This is Carduus Flodmannii Rydb.

Distribution. This species is distributed from Mason City, Iowa, to south-western Minnesota, west to the Rocky Mountains. Collected by Charles A. Geyer in 1839, and described by Nuttall. The writer has seen it very abundant in Wyoming, Montana and Colorado.

Cirsium discolor (Muhl.) Spreng. Prairie Thistle

Tall, branching, leafy biennial, 5-7 feet high, with heads larger than the Canada thistle; stem striate, slightly hirsute; leaves radical 12-14 inches long, deeply pinnatifid, the divisions frequently divided, prickly toothed, the upper surface smoothish, and the lower white, woolly single heads terminating the branches, with purple flowers, heads 1½ inches high; bracts of the globose involucre slightly arachnoid, lower bracts ovate, with a broad base and a weak prickly recurved bristle, slight dorsal gland, inner linear lanceolate with a nearly colorless entire appendage; flower purple; lobes of the corolla terminating in clavate tips, anther tips acute, filaments pubescent; bristle of pappus plumose; achenium smooth, upper part yellow.

Distribution. In fields and along roadsides from Quebec, Ontario, south through New England, New York and Georgia, west to Missouri, Nebraska and South Dakota.

Cirsium ochrocentrum Gray

Biennial, 2-8 feet high; white tomentum; leaves commonly deeply pinnatifid and armed with long yellowish prickles; heads 1-2 inches high; involucral scales with a viscid line on the back, with a prominent spreading yellowish prickle; corolla purple, rarely white.

Distribution. From Arizona to Colorado and Utah.

Cirsium undulatum (Nutt.) Spreng.

Biennial 1-2 feet high, white tomentose; leaves pinnately parted, somewhat prickly; heads about 2 inches high; the outer bracts thickened by glandular-viscid ridge; corolla rose-color, purple or pale purple; the variety megacephaleum with larger heads.

Distribution. West of the Missouri River to Oregon and New Mexico.

Injurious properties. None of the species is poisonous so far as the writer knows. The spiny involucre and spiny leaves inflict inflammation and cause the formation of pus.

28. Centaurea L.

Perennial or annual herbs, with leaves alternate, entire, dentate, incised or pinnatifid; involucre ovoid or globose; bracts in many series; marginal flowers usually neutral and larger than the central ones in some species; all the flowers perfect and fertile; heads middle-sized, tubular, purple-violet, or white or rarely

yellow; receptacle flat and bristly; corolla-tube slender and 5-cleft or 5-lobed; anthers sagittate at the base; achenes oblong or obovoid; pappus of several series of bristles and scales.

About 350 species mostly of the old world where they are troublesome weeds. The common bluebottle or corn flower is frequently cultivated in gardens and is a frequent escape from cultivation.

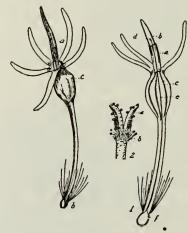


Fig. 458. Flowers of centaurea. 1, Flower; a, anther; 2, style. At the left, Achene and Pappus.

Centaurea solstitialis Linn. Knapweed

Erect, branched, cottony, stems winged, lower leaves lyrate, upper linear entire, decurrent, spines of upper bracts long spreading with a few smaller ones at the base, pappus soft.

Distribution. Fields in California, also in Europe and England, rare; introduced with alfalfa. Introduced into Iowa with alfalfa seed.

Injurious properties. Because of the spiny character of the plant it is often troublesome to man and animals.

29. Silybum (Vaill.) Adans. Milk Thistle

Annual or biennial, much branched herbs; leaves large, alternate, clasping, white-blotched; heads discoid, solitary at the end of the branches; involucre large; bracts rigid, some armed with large spreading or recurved spines; flowers all tubular; corolla-tube slender, top 5-cleft; anthers sagittate; achencs oboyate-oblong; pappus-bristles in several rows.

A single species, native to the Mediterranean region.

Silybum Marianum (L.) Gaertn. Lady's Thistle

A tall branched glabrous annual with striate stem; leaves oblong lanceolate, prickly clasping.

Distribution. Occasionally found eastward especially in ballast and common on the Pacific coast, from British Columbia to Southern California.

Injurious properties. The spiny leaves and involucre have been trouble-some; produces mechanical injuries.

A CATALOGUE OF THE POISONOUS PLANTS OF THE WORLD

In the following catalogue an attempt has been made to record all of the species that have been in some way or another regarded as poisonous or injurious to man. This catalogue, however, is based mainly on the plants listed by Greshoff, Cornevin, Chesnut, Ernst, Lyons, Maiden, Miquel, Radlkofer, Rusby, Smith, White in "Dermatitis venenata," but includes also a few from other scattered sources. The poisonous fungi were listed from Atkinson, Hard, Clements, and some from Fries, Peck, Farlow, and Bulliard. It was not thought best to list the fungi from the works of Ratzeburg and Phoebus entirely because of the uncertainty as to the poisonous qualities. It should be stated that many of the plants in this list are not poisonous nor are they always given as poisonous by the authors credited to them after the species, in fact, many of them on the whole must be regarded as plants simply having medicinal virtues. It was thought best, however, to list them, to permit future investigators to take up the problem of the chemistry and poisonous qualities of these plants from an experimental standpoint. It is simply an indication of the lines along which work should be done. We have also indicated the action of the plant, or in some few cases the important substance found in the plant. The alphabetical arrangement has been adopted except as to the lower forms. In the last column the distribution is indicated usually only where the plant is indigenous, but in some cases North America is added where the plant has been introduced.

I have been greatly aided in this work by the excellent treatise of A. B. Lyons, "Plant Names, Scientific and Popular," published by Nelson Baker & Co., and also by the excellent work of Sayre, "Organic Materia Medica and Pharmacognosy."

In the preparation of this catalogue I have received very substantial help from Miss Harriette S. Kellogg, who has looked after the details in catalogueing the species and the synonyms. In some cases the synonyms could not be found in the Kew Index, and these have been allowed to stand as they were given by the authors. Some duplication of names may occur, and possibly through an oversight some have been placed in the wrong orders, but I ask the reader's indulgence in errors of this kind.

Finally, I am indebted to Miss Bertha D. Herr for the laborious task of getting the copy ready for the printer.

L. H. Pammel.

The following abbreviations refer to the properties mentioned in the third column of the catalogue.

Abort.—abortifacient
Acr.—acrid poison
Alk.—alkaloidal poison
Amyg.—contains amygdalin
Androm.—contains andromedotoxin
Anes.—anesthetic
Ant.—anthelmintic
Antisc.—antiscorbutic
Antisp.—antispasmodic

Astr.—astringent
Berb.—contains berberin
Card.—cardiac poison
Cath.—cathartic
Con.—convulsive
Cou.—contains coumarin
Cur.—curare poison
Cyt.—contains cytisin
Del.—deleriant

A. P.—arrow poison
Ast.—asthenic
Dr. Em.—drastic-emetic
Em.—emetic
Ent.—causes enteritis
Eser.—contains eserin
Exp.—expectorant
F. P.—fish poison
Gastr.—causes gastro-enteritis
Hyd.—contains HCN.
Hyp.—hypnotic
Ineb.—inebriant
Ins.—insecticide
Intox.—intoxicant
Ir.—irritant
Lax.—laxative
Loco.—causes loco poisoning
Mech.—causes mechanical injuries

Nar.—narcotic

Diaph.—diaphoretic
Diur.—diuretic
Naus.—nauseant
Ord.—ordeal poison
Par.—paralyzing
Pur.—purgative
Sap.—contains saponin
Sed.—sedative
Sial.—sialagogue
Sol.—contains solanin
Sop.—soporific
Stim.—stimulant
Taen.—taenifuge
Toxal.—Contains a toxalbumin.
Tymp.—causes tympanites
Ur.—causes uraemic poisoning
Urt.—urticarial
Ver.—vermifuge

Ves.-vesicant

The abbreviations for the literature referred to are as follows:

C.—Cornevin's Les plantes vénéneuses et ses empoisonnements qui elles determinent,

Ches.—Chesnut's Preliminary Catalogue of Plants Poisonous to Stock.

E.—Ernst's Ueber fischvergiftenden Pflanzen.

G.—Greshoff's The Distribution of Prussic Acid in the Vegetable Kingdom. Phytochemical Investigations at Kew.

K.-Kirtikar's The Poisonous Plants of Bombay.

L.-Lyon's Plant Names, Scientific and Popular.

M.-Maiden's Plants reputed to be Poisonous to Stock in Australia.

M.i-Miquel's Poisonous Plants of North Netherlands.

R.-Radlkofer's Plants said to be Poisonous to Fish,

Rusby-Rusby's The Poisonous Plants of the Vicinity of New York City.

Sm.—Smith's Poisonous Plants of all Countries.

Wh.-White's Dermatitis venenata.

ACANTHACEAE

| NAME OF PLANT | Authority | Properties | Locality |
|----------------------------------|----------------|------------|----------------|
| Hygrophila hispida Nees. | G. | Em. | Cuba |
| Justicia Gendarussa Burm. | G. G. G. | | Tr. As. |
| Lepidagathis Wightiana Benth. | G. | Abort. | |
| Paulowilhelmia polysperma Benth. | G. G. | F. P. | Tr. Afr. |
| Paulowilhelmia speciosa Hochst. | G. | Em. | Abyss. |
| Rhinacanthus communis Nees. | G. | | Ind. Malay. |
| | | | Burma. Abyss. |
| Ruellia patula Nees. | G. G. | Em. | Arab. Ind. |
| Ruellia strepens L. | | Em. | N. Am. W. Ind. |
| Ruellia tuberosa L. | G. L. | Em. | N, Am. Tr. Am. |

ADOXACEAE ALISMACEAE

| Alisma Plantago L. | G. M. L. | Ir. | Tem, N. Am. |
|--------------------|----------|-----|-------------|
| | | | Austr. |

AMARANTHACEAE

| NAME OF PLANT | Authority | Properties | Locality |
|--------------------------------|-----------|------------|----------------|
| Achyranthes bidentata Bl. | G. | Sap. | Tr. As. |
| Amaranthus gangeticus L. | G. G. | Sap. | Trop. Reg. |
| Amaranthus hybridus L. | L. | Astr. | Eur. N. A. |
| Amaranthus hypochondriacus L. | G. | Sap. | N. Am. Eur. |
| Amaranthus viridis L. | G. | Sap. Ant. | Tr Reg |
| Celosia anthelmintica Aschers. | G. | Ant. | Tr. Afr. |
| Celosia trigyna L. | G. | Taen. | Tr. Afr. |
| Deeringia celosioides R. Br. | G. | | As. Tr. Austr. |

AMARYLLIDACEAE

| Amaryllis Belladonna L. | G. | | S. Afr. |
|----------------------------------|--|-------|--------------|
| Buphane disticha Herb. | G.C. Sm.L. | A. P. | S. Afr. |
| Crinum asiaticum L. | G. | | Tr. As. |
| Crinum zeylanicum L. | G. | | Ind. |
| Curculigo scorzoneraefolia Baker | G. | | Tr. Am. |
| Furcraea gigantea Vent. | G. | Sap. | Tr. Am. |
| Galanthus nivalis L. | G. | | Eur. |
| Haemanthus | G. | | Afr. |
| Haemanthus coccineus L. | G. G | Diur. | S. Afr. |
| Hippeastrum equestre Herb. | G. | | Mex. |
| Hippeastrum rutilum Herb. | G. | | Brazil |
| Hymenocallis sp. | G. | | Tr. Am. |
| Leucojum aestivum L. | C. | | Ori. |
| Leucojum vernum L. | G. | | Eur. |
| Lycoris radiata Herb. | G. | Em. | China. |
| Narcissus poeticus L. | Mi. | Ir. | Eur. |
| Narcissus Pseudo-Narcissus L. | M.G.C.L. | Ir. | Eur. |
| Narcissus Tazetta L. | Mi. | | Med. Reg. |
| Pancratium Illyricum L. | G. | | Eur. Austr. |
| Pancratium maritimum L. | G. C. L. | Diur. | Med. Reg. N. |
| | | | Am. |
| Pancratium zeylanicum L. | G. | | Ind. |
| Sprekelia formosissima Herb. | G. | | Mex. |
| Zephyranthes Atamasco Herb. | Clies. | | N. A. |

ANACARDIACEAE

| Buchanania sp. | G. | | Tr. As. Pac. Is. |
|---|--|-------|-------------------------|
| Comocladia glabra Spreng. | G | | Austr. Cuba. |
| Corynocarpus laevigata Forst. | G. G | Con. | N. Zeal. |
| Gluta Benghas L. | G. | | Java. |
| Holigarna caustica Roxb. | G. | ł | Ind. |
| Holigarna longifolia Roxb. | G. | i | Ind. |
| Lithraea venenosa Miers. | ا د | | S. Am. |
| Mangifera indica L. Oncocarpus vitiensis A. | Ğ. | | Malay, Ind. Fiji Is. |
| Pistacia Khinjuk Stocks | Ğ. | Ir. | Egyp. Pers. |
| I istacia zimijan stovils | | | Him. |
| Pseudosmodingium perniciosum Engl. | G. C. | Ir. | Mex. |
| Rhus Cotinus L. | J C. | | Eur. China. |
| | | | Med. Reg. |
| *Rhus glabra L. | L. | Astr. | N. A. |

^{*} Just as this part of the work is going to press I have received two important papers on Rhus by L. F. Warren in which the statement is made that the poisonous substance of Rhus is a powerful escharotic, one milligram producing very severe blistering when

| NAME OF PLANT | Authority | Properties | Locality |
|---|-------------|---------------|----------------------------------|
| Rhus Metopium L. | L. | Emeto-Cath | N. A. W. I. |
| Rhus Toxicodendron L. | G. L. | Ir. | N. Am. |
| Rhus venenata DC. | G. | Įr. | N. Am. |
| Rhus vernicifera DC. | G. L. | Ir. | Thib. Jap. |
| Sclerocarya Caffra Sond. | G. G. L. | Ir. Ir. | S. Afr. Austr. As. Tr. Austr. |
| Semecarpus Anacardium L. Semecarpus heterophylla Bl. | G. L. | Ir. | I I ava. |
| Spondias mangifera Willd. | G. | Ir. | Trop. As. |
| | 1 0. | 1111 | 110p. 115, |
| ANONA | | | |
| Anona amara Raeusch. | G. | | Guiana |
| Anona Cherimolia Mill. | G. | F. P. | Tr. Am. |
| Anona muricata L. | G. L. | Astr. | Tr. Am. |
| Anona palustris L. | G. L. | F. P. Acr. | Tr. Am. & Afı |
| Anona reticulata L. Anona spinescens Mart. | G. L. G. | Acr. | Tr. Am. Brazil |
| Anona squamosa L. | G. L. | г. Р. | Ind. |
| Asimina triloba Dun. | G. L. | Em. | N. Am. |
| Goniothalamus macrophyllus Hook. | G. | | Malacca. |
| Guatteria veneficiorum Mart. | G. |) I | Brazil. |
| Popowia pisocarpa Endl. | G. | | Java. |
| Xylopia odoratissima Welw. | G. | 1 | Tr. Afr. |
| Xylopia polycarpa Oliv. • | L. | Berb. | Tr. Afr. |
| Xylopia salicifolia HBK. | 1 | H | Tr. S. Am. |
| APOCYN | NACEAE | | |
| Acokanthera Lamarkii G. Don. | I G. | | S. Afr. |
| Acokanthera venenata G. Don. | l G. | | S. Afr. |
| Adenium Boehmianum Schinz. | I G. | | Afr. |
| Adenium obesum Roem & Schult. | G. | | Afr. Arab. |
| Adenium somalense Oliv. | G. | A. P. | Afr. |
| Aganosma calycina A.DC. | G. | F. P. | Ind. |
| Allamanda cathartica L. | G. | Em. | Tr. Am. |
| Alstonia constricta F. Muell. | L. | Bitter | M 014 |
| Alstonia Scholaris R. Br. | G. L. | | Trop. Old World |
| Alstonia venenata R. Br. | G. 17. | | Ind. |
| Anstoina Venenata It, Di. | J . | | LIICI. |

placed on the arm for fifteen minutes. When in contact with the air it produces the characteristic non toxic varnish. The resinous products from the latex of the Rhus give black compounds with the alkali hydroxides. The irritating properties he thinks will be found to be connected with the presence or the relations of these hydroxyl groups.

G.

Austr.

This writer found that the milk sap of Rhus vernix is analogous in almost every particular to the Japanese lac. Warren agrees with nearly everyone else that the bacterial infection theory has very little to support it. Nor is there much to support the recently elaborated theory of immunity. He shows how a popular impression in regard to supposed immunity has become widespread even among scientists. It has not, however, been demonstrated. In the same way, contrary to the early published records, Rhus Michauxii has been shown by Warren and Trelease not to be poisonous. As to treatments, Warren, after an extensive investigation of the subject, states that the great majority are empirical. "There is no specific for this troublesome complaint. Remedies which have alleviated the symptoms in one case have proved utterly valucless or worse in others." The following species are listed by Warren as poisonous:

Rhus venenata D. C. (R. vernix L.); Rhus radicans L.; Rhus Toxicodendron L.; Rhus diversiloba Torr. & Gray; Rhus Rydbergii Small; Rhus Metopium L.; Rhus floridana Mearns; Rhus littoralis Mearns; Rhus vernicifera DC; Rhus succedanea L.; Rhus sylvestris Sieb. & Zucc.; Rhus Wallichii Hook. fils; Rhus Griffithii Hook. fils; Rhus striata R. & Pav.; Rhus perniciosa H. B. & K.; Rhus chinensis Mill.; Rhus javanica L.; Rhus caustica Hook. & Arn.; Rhus lucida L.

Alyxia buxifolia R. Br.

| NAME OF PLANT | Authority | Properties | Locality |
|---|----------------------|-------------|------------------|
| Alyxia daphnoides A. Cunn. | G. | 1 | Norfolk Is. |
| Amsonia Tabernaemontana Walt. | } | | N. Am. |
| Apocynum androsaemifolium L. | Ches. G.C. | Card. | N. Am. |
| | Rusby L. | | 1 |
| Apocynum cannabinum L. | Ches. G.C. | F. P. Card. | N. Am. |
| | Rusby L. | Ì | |
| Apocynum venetum L. | G. | | Med. Reg. Ori- |
| Aspidosperma quebracho-blanco Schlecht. |] G. L. | | ent N. As, Ind |
| | [| | S. Am. |
| Aspidosperma sessilifolium Fries. | G. R. | F. P. | S. Am. |
| Cameraria latifolia L. | G. |] | Ind. |
| Carissa ovata R. Br | G. | | Austr. |
| Carissa Xylopicron Thou. | G. | | Mascar Is. |
| Cerbera Odollam Gaertn. | G. C. R. | F. P. | Ind. Malay. |
| Cerbera Tanghin Hook. | C. L. | | _ |
| Echites alexicaca Mart. & Stadelm. | L. | Cath. | Braz. |
| Echites maculata A. DC. | 1 G. C. | | St. Doming. Is |
| Echites venenosa Mart. | G. L. | A. P. | Brazil. |
| Ellertonia Rheedi Wight. | G | | Ind. |
| Geissospermum vellosii Allem | G. L. | Bitter | Braz. |
| Haplophyton cimicidum A. DC. | G. | Ins. | Mex. |
| Malouetia nitida Spruce | | A. P. | Braz. |
| Melodinus monogynus Roxb. | G. R. | F. P. | Him. Malaya. |
| N : 1 C 1 1 | | l | China. |
| Nerium odorum Soland. | G. C. Mi. | Car. | Per Jap. Ind. |
| Nerium Oleander L. | | Car. | Med. Reg. Ori |
| Ochrosia Moorei F. von Muell | Ches. L. | } | Austr. |
| Plumiera rubra L. | C. L. | | Trop. Am. |
| Pottsia cantonensis Hook & Arn. | G. | | Ind. Java China |
| Prestonia toxifera | G. | | Ind. Java Cillin |
| Rauwolfia serpentina Benth. | G. | F. P. | Ind. Java. |
| Rauwolfia trifoliata Gaertu. | G. | | Tava. |
| Rauwolfia verticillata Baill. | | F. P. | java. |
| Strobilanthes callosus Nees. | K. | 1 | Ind. |
| Strophanthus Eminii Asch. | L. | A. P. | Cent. Afr. |
| Strophanthus hispidus DC. | G. L. | F. P. A. P. | S Afr |
| Strophanthus Kombe Oliver | L. | F. P. | Trop. Afr. |
| Strophanthus Pierrei Heim. | Ğ. | | W. Afr. |
| Tabernaemontana Borbonica Lam. | Ğ. | | |
| Tabernaemontana citrifolia L. | Ğ. | | Ind. Mex. |
| Taberaemontana coronaria Willd. | Ğ. | | Ind. |
| Taberaemontana malaccensis Hook. | Ğ. | | Malacca. |
| Taberaemontana Mauritiana Poir. | | F. P. | Mascar. Is. |
| Tabernanthe Iboga Baill. | G. | | Trop. Afr. |
| Thevetia Ahouai A.DC. | G. R. G. C. R. L. | F. P. | Brazil. |
| Thevetia nereifolia Juss. | G. C. R. L. | F. P. | Tr. Am. |
| Urechites suberecta Muell. | G. L. | Sed. | St. Doming, Is |
| Vinca major L. | G. L. | | Med. Reg. |
| Vinca minor L. | L. | | Eur. |
| Vinca pusilla Murr. | G. | | Ind. |

AQUIFOLIACEAE

| T1 C ' 117.14 | CT | IT C (1 | 137 4 |
|---------------------------|--------|-----------|---------|
| Ilex Cassine Walt. | 5m. L. | Em. Cath. | IN. Am. |
| Ilex glabra A. Grav | L. | Astr. | N. A. |
| Ilex verticillata A. Grav | T. | Astr. | N. A. |
| HEX VEHICINALA A. Gray | L. | Insu. | 114. A. |

ARACEAE

| NAME OF PLANT | Authority | Properties | Locality |
|-------------------------------------|----------------|------------|----------------|
| Alocasia indica Schott. | G. | 1 | Ind. Malay. |
| Alocasia montana Schott. | G. | | Ind. Malay. |
| Amorphophallus viridis Bl. | G. | | |
| Anthurium Ferrierense | | Ir. | |
| Arisaema curvatum Kunth. | G. | Ir. | Him. Reg. |
| Arisaema tortuosum Schott. | G. | Ir. | Him. Reg. |
| Arisaema triphyllum Schott. | Rusby | Em. | N. A. |
| Arisaema vulgare Targ. | G. | Ir. | Med. Reg. |
| Arum Dioscoridis Sibth. | G. | | Greece As. Min |
| Arum Dracontium Schott. | L. | Acr. | N. A. |
| Arum italicum Mill. | G. C. Mi. | Sap. | Eur. |
| Arum maculatum L. | G. Mi. | Ir. | Eur. |
| Arum triphyllum Schott. | L. | Acr. | N. A. |
| Caladium bicolor Vent. | G. | Ins. | Am. Austr. |
| Calla palustris L. | C. L. | Acr. | Eur. As. N.Am |
| Colocasia antiquorum Schott. | G. L. | Acr. | Tr. As. |
| Colocasia gigantea Hook f. | G. G. | Acr. | |
| Colocasia virosa Kunth | G. | Acr. | Ind. |
| Cryptocoryne spiralis Fisch. | G. | Em. | Ind. |
| Cyrtosperma Merkusii Schott. | G. G. C. G. L. | | Sumatra. |
| Dracontium asperum C. Koch. | G. | | Brazil. |
| Dracunculus vulgaris Schott. | C. | | S. Eur. |
| Epipremnum mirabile Schott. | G. L. | Ant. | Java. |
| Homalomena aromatica Schott. | I G. | | Malay. Ind. |
| Lasia aculeata sp. Lour. | G. G. | Hyd. | Tr. As. |
| Monstera pertusa Schott. | G. | Ir. | Tr. Am. |
| Philodendron bipinnatifidum Schott. | G. | }_ | Brazil. |
| Philodendron hederaceum Schott. | G. | P. | S. Am. |
| Philodendron Imbe Schott. | G. | Em | Argentine. |
| Philodendron Simsii Sweet. | Sm. | Ir. | Guiana. |
| Pinellia tuberifera Ten. | G. L. | Ir. | Japan. |
| Scindapsus officinalis Schott. | G. | Ant. | Burma. |
| Spathiphyllum candicans Poepp. | ! G. | 1. | Trop. Am. |
| Symplocarpus foetidus Nutt. | Sm. | Tr. | E. N. Am. |

ARALIACEAE

| Aralia sp. | G. | Sap. | N. Am. Asia |
|--------------------------------|----------------------|---------|-----------------|
| Aralia hispida Vent. | [L, | Dur. | N. E. U. S. |
| Aralia nudicaulis L. | L. L. | Dur. | N. E. U. S. |
| Aralia racemosa L. | G. L. | Em. | N. A. |
| Aralia spinosa L. | | Diur. | N. Am. |
| Hedera Helix L. | G. C. L. | Em. Nar | Eur. Afr. As. |
| Heptapleurum emarginatum Seem. | | | Zeylan. |
| Heptapleurum scandens Seem. | G. G. G. G. | | Malay. |
| Heptapleurum venulosum Seem. | G. | | Austr. |
| Panax sp. | Ğ. | Sap. | N. Am. As. |
| Polyscias sp. | Ğ. | Sap. | Austr. N. Zeal. |
| z oty serias sp. | J. | Dap. | Afr. Madagas. |
| | | | Malay. Pac. Is. |
| Polyscias nodosa Seem. | G | F. P. | Malay. |
| Trevesia sp. | G. G. | Sap. | Java. Sumatra. |
| ricvesia sp. | <u> </u> | 11.01/. | ,3474. |

ARISTOLOCHIACEAE

| Aristolochia anguicida Jacq. | G. | Diaph. | S. Am. |
|----------------------------------|------|--------|--------------|
| Aristolochia antihysterica Mart. | l G. | Em. | Brazil. |
| Aristolochia argentina Griseb. | G. | Em. | Argent, Reg. |

| NAME OF PLANT | Authority | Properties | Locality |
|---|-----------|-------------|-----------------|
| Aristolochia brasiliensis Mart. & Zucc. | G. | Em. | Brazil. |
| Aristolochia Clematitis L. | G. C. Mi. | Em. | Eur. Caucas. |
| | | | Reg. As. Minor. |
| Aristolochia elegans Mast. | G. | Em. | Brazil. |
| Aristolochia grandiflora Arruda | G. C. | | Pernamb. |
| Aristolochia indica L. | G. G. | | Ind. |
| Aristolochia Kaempferi Willd. | G. | | Japan. |
| Aristolochia longa L. | C. L. | | Med. Reg. |
| Aristolochia pallida Willd. | G. | F. P. Nar. | Eur. As. Minor. |
| Aristolochia Pistolochia L. | l C. L. | | Med. Reg. |
| Aristolochia rotunda L. | G. L. | Nar. | Med. Reg. |
| Aristolochia Serpentaria L. | L. E. G. | Diaph. | E. U. S. |
| Aristolochia sp. | E. | F. P. | Italy. |
| Asarum albivenium Regel. | G. | Em. | Japan, |
| Asarum arifolium Mx. | L. | Ir. | N. A. |
| Asarum canadense L. | G. L. | Ir. | N. Am. |
| Asarum caudatum L. | L. | lr. | Calif. |
| Asarum europaeum L. | | [Ir. EmCat. | |
| Asarum virginicum L. | L. | Ir. | N. A. |
| Bragantia tomentosa Bl. | G. | F. P. | Ind. Malay. |
| Thottea dependens Klotzsch. | G | lr. | Malay. |

ASCLEPIADACEAE

| Araujia sericifera Brot. | G. | Em. | Peru. | | |
|------------------------------------|----------------|------------|---------------|--|--|
| Asclepias curassavica L. | G. L. | Ent. | S. Am. | | |
| Asclepias eriocarpa Benth. | 1 | Em. | W. N. Am. | | |
| Asclepias incarnata L. | Ches. C. L. | | N. Am. | | |
| Asclepias mexicana Cav. | Ches. | Em. | W. N. Am. | | |
| Asclepias syriaca L. | Ches. C. L. | | N. Am | | |
| Asclepias tuberosa L. | C. Ches. | | N. Am. | | |
| Calotropis gigantea R. Br. | | Em. | Ind. | | |
| | 1 - | | Peru N. Afr. | | |
| Calotropis procera R. Br. | G. | Ineb | | | |
| Consider to the second | 0 | | Ind. | | |
| Ceropegia bulbosa Roxb. | G. | | Ind. | | |
| Chlorocodon Whiteii Hook. | G. G. G. | Cou. | S. Afr. | | |
| Choristigma Stieckertianum. | G. | | Arg. | | |
| Cosmostigma racemosa Wight. | G. | | Ind. | | |
| Cryptostegia grandiflora R. Br. | G. | | Tr. Afr. Ind. | | |
| | 1 | l | [(Cult.). | | |
| Cynanchum acutum L. | | Cath. | Eur. Ori. As. | | |
| Cynanchum sarcostemmoides K. Schum | .] <u>G</u> . | _ | | | |
| Gonolobus laevis Mx. | G. G. G. | A. P. | N. Am. | | |
| Gonolobus obliquus R. Br. | G. | | N. Am. | | |
| Gymnema latifolium Wall. | | | Ind. Burma. | | |
| Gymnema sylvestre R. Br. | G. L. | Destroys | Ind. Tr. Afr. | | |
| | | sense of | Austr. | | |
| | | taste | ì | | |
| Hemidesmus indicus R. Br. | G. L. | Cou. | Zeylan. | | |
| Marsdenia Cundurango Nich. | L. | Dog Poison | S. A. | | |
| Marsdenia erecta R. Br. | G. L. | Ì | Syr. | | |
| Menabea venenata Baill. | G. L. | Ord. | Madagas. | | |
| Metaplexis Stauntoni Schult. | G. | | China. | | |
| Morrenia brachystephana Griseb, | G. | | Argentine. | | |
| Periploca graeca L. | G. L. | | S. Eur. | | |
| 9 | | Poi. | | | |
| Periploca vomitoria Leschen. | G. | Em. | Java. | | |
| Sarcolobus carinatus Wall. | G. G. | | Ind. Burma. | | |
| Sarcolobus narcoticus Mig. | Ğ. | Nar. | Java. | | |

| NAME OF PLANT | Authority | Properties | Locality |
|--|---|--|---|
| Sarcostemma australe R. Br. Sarcostemma glaucum HBK. Solenostemma Argel Del. Stapelia sp. Tylophora asthmatica W. & A. Tylophora fasciculata Ham. Tylophora laevigata Decne. Vincetoxicum nigrum Moench. Vincetoxicum officinale Moench. | G. M. G. L. C. G. L. G. G. C. C. C. C. Mi. L. | Em. Em. Em. Em. Dr. Em. Dr. Em. | Aust. S. Am. Venez. Nubia. S. Afr. Ind. Malay. Ind. Maurit Is. Eur. As. Minor. Eur. Cau. C. |
| BALSAMI | NACEAE | | |
| Impatiens amphorata Edgew. Impatiens aurea Muhl. Impatiens biflora Walt. Impatiens Noli-tangere L. | G. L. L. G. L. | Diur, Diur, Diur, Ir, Diar, | Him. N. A. N. A. Eur. Siberia. |
| BEGON | ACEAE | | |
| Begonia gracilis HBK. | G. | Em. | Mex. |
| BERBERI | DACEAE | | |
| Berberis Aquifolium Pursh, Berberis aristata DC, Berberis vulgaris L, Caulophyllum thalictroides Michx. Leontice Leontopetalum L, Nandina domestica Thunb, Podophyllum Emodi Wall, Podophyllum peltatum L, | G. C. L. G. L. G. L. G. L. G. L. G. L. G. L. Sm. Rusby C. Ches. | F. P. F. P. Ir. Sap. Ant. for Opium Cath. Pur. | N. Am. Ind. Eur. Tem. As. N. Am. Italy Orient, Japan China. Him. Reg. N. Am. |
| BETUL | ACEAE | | |
| Betula lenta L. Betula lutea Michx. | Witthaus Witthaus | Astr. Astr. | E. N. Am. E. N. Am. |
| BIGNON | IACEAE | | |
| Bignonia capreolata L. Crescentia cucurbitina L. Crescentia Cujete L. Dolichandrona falcata Seem. Jacaranda procera Spr. Osmohydrophora nocturna Tanaecium crucigerum Seem. Tecoma ceramensis Teijsm & Binn. Tecoma leucoxylon Mart. Tecoma obtusata DC. Tecoma radicans Juss. Tecoma speciosa DC. Tecoma stans Juss. Tecoma toxophora Mart. | G. G. R. G. | F. P. Abort. F. P. Hyd. F. P. F. P. | N. Am. Tr. Am. Trop. Am. Ind. Guiana. Braz. W. Ind. Ceram Is. Guiana W. Ind. Brazil. N. Am. Brazil. N. Am. Brazil. N. Am. Brazil. |
| BIXAC | | F D | |
| Gynocardia odorata R. Br. Hydnocarpus anthelminticum Pierce. Hydnocarpus castanea H. F. & Thoms. | G. L. L. G. | F. P. Ant. F. P. | Ind. Andoman Is. Malacca. |

| NAME OF PLANT | Authority | Properties | Locality |
|--|---|----------------------|--|
| Hydnocarpus heterophylla Bl. Hydnocarpus Kurzii Hydnocarpus piscidia Hyndocarpus venenata Gaertn. Hydnocarpus Wrightiana Bl. Pangium edule Reinw. Taraktogenos Blumei Hassk. Trichadenia zeylanica Thw. | R, L. Sm. E. G. L. R. L. R. G. R. E. G. G. | Ant. | Java. Trop. As, Zeylan. Ind. Java. Sumatra. Zeylan. |
| BORAGI | NACEAE | | |
| Asperugo procumbens L. Borago officinalis L. Pourreria Havanensis Miers. Cynoglossum officinale L. Echium vulgare L. Heliotropium europaeum L. | Sm. Whi. C. G. G. L. G. G. | Ir. Ir. Ir. | N. As. Eur. Eur. Afr. As. Cuba. Eur. Orient. Eur. N. Am. Eur. Afr. Ori. Austr. N. An. Int. |
| BROMEL | IACEAE | | |
| Ananas sativus Schult. Bromelia Pinguin L. Karatas Plumieri Emorr. | G. L. G. L. G. | Ant. Sap. Ant. | Tr. Am. Tr. Am. Panama. |
| BURMAN | NIACEAE | | |
| Dipodium punctatum R. Br. | G. M. | | Aust. |
| BURSER | ACEAE | | |
| Canarium sp. Canarium commune Linn. | G. L. | F. P. Cath. | Aust. Pac. Is. Philipp. Molucca Is. |
| BUXA | CEAE | | |
| Buxus balearica Lam. Buxus sempervirens L. | G. Ches. G.C. L. | Cath. Ir. | Balearic Is. Eur. Ori. Tem. As. |
| CACTA | CEAE | | |
| Anhalonium Lewinii Henn. Cereus Bonplandii Parm. Cereus flagelliformis Mill. Cereus grandiflorus Mill. Cereus McDonaldii Hook. Echinocactus sp. Echinocactus Williamsii Lem. Mammillaria sp. Rhipsalis sp. | G. L. G. L. G. L. G. C. G. C. G. C. G. C. | Ir. Nar. | Mex. Brazil. S. Am. W. Ind. Honduras. N. Am. Mex. Tex. W. N. Am. S. Am. Trop. |
| CALYCAN | ГНАСЕАЕ | | |
| Calycanthus glaucus Willd. | G. L. Ches. | Sheep Poi. | N. Am. |

CAMPANULACEAE (Including LOBELIACEAE)

| CAMPANULACEAE (Including LOBELIACEAE) | | | | | |
|---|---|--|--|--|--|
| NAME OF PLANT | Authority | Properties | Locality | | |
| Isotoma axillaris Lindl. Isotoma Brownii G. Don. Isotoma longiflora Presl. Lobelia cardinalis L. | G. M. M. G. M. C. Mi. L. | Ant. | Austr. Austr. W. Ind. E. N. Am. | | |
| Lobelia fulgens Willd. Lobelia inflata L. | Rusby C. Mi. Sm. Ches. L. Rusby. | Ac. Dep. Em. | Eur. N. Am. | | |
| Lobelia Kalmii L. Lobelia nicotianaefolia Heyne. Lobelia pratioides Benth. Lobelia rhynchopetalum Hemsl. Lobelia siphilitica L. | Ches. G. M. G. Rusby. L. Ches. | Ac. Acr. Acr. | N. A. Ind. Austr. Tr. Afr. E. N. A. | | |
| Lobelia spicata Ruiz. & Pav. Lobelia Tupa L. Lobelia urens Willd. Pratia erecta Gaud. Siphocampylus giganteus Don. | Ches. Ches. G. R. G. C. G. M. | F. P. Acr. | Peru. Chili. Peru. Eur. Austr. Equad. | | |
| CANELI | LACEAE | | | | |
| Canella alba Murr. | G. L. | Sap. F. P. | W. Ind. | | |
| CAPPARIDACEAE | | | | | |
| Boscia urens Welw. Cadaba indica Lam. Capparis Cynophallophora L. Capparis ferruginea L. Capparis globifera Del. Capparis odoratissima Jacq. Cleome Chelidonii Lf. Cleome frutescens Aubl. Cleome gigantea L. Cleome graveolens Raf. Cleome psoraleaefolia DC. Cleome rosea Cleome spinosa Jacq. Maerua angolensis DC. Morisonia americana L. | G. G. Sm. L. G. | Ant. Diur. Ant. Diur. Ir. F. P. | Tr. Afr. Ind. S. Am. W. Ind, Tr. Afr. S. Am. Venez. Ind. Java. Guiana, Braz. S. Am. N. Am. Brazil. Texas. S. Am. Tr. Afr. S. Am. | | |
| CAPRIFO | | | | | |
| Abelia uniflora R. Br. Diervilla japonica DC, Diervilla trifida Moench. Lonicera chrysantha Turcz. Lonicera involucrata Banks Lonicera japonica Thunb. Lonicera Standishii Hook. Lonicera tatarica L. Lonicera tomentella Hook f. & Thoms. Lonicera Xylosteum L. Sambucus canadensis L. Sambucus Ebulus L. | G. C.Rusby L. G. C. | Sap. | China. Japan. N. Am. Siberia. N. Am. Ry.Mts. Him. Reg. China. Siberia. Him. Reg. Eur. N. As. E. N. Am. Reg. | | |
| Sambucus mexicana Presl. | L. | Em. | W. N. A. | | |

| NAME OF PLANT | Authority | Properties | Locality |
|---------------------------------|------------|------------|-----------------|
| Sambucus nigra L. | G.C. Rusby | Pur. | Eur. N. As. |
| Sambucus racemosa L. | G. C. | | N. Tem. Reg. |
| Symphoricarpos mollis Nutt. | G. | Sap. | W. N. Am. |
| Symphoricarpos racemosus Michx. | l G. | Jr. | N. Am. |
| Triosteum majus Michx. | | | E. N. Am. |
| Triosteum perfoliatum L. | G.Rusby L. | Cath. | N. Am. |
| Viburnum Lantana L. | G. L. | Astr. | Eur. Cauc. Reg. |
| Viburnum macrophyllum Thunb. | G. | Sap. | Japan. |
| Viburnum Opulus L. | G. L. | Sed. | Eur. As. N.Am. |

CARYOPHYLLACEAE

| Acanthophyllum C. A. Mey. sp. | G. | | |
|-------------------------------|----------------|-----------|-----------------|
| Agrostemma Githago L. | G. C. Sm. | Sap. Ir. | Eur. As Int. N. |
| | Ches. | 1 | & S. Am. Austr. |
| Arenaria serpyllifolia L. | | Abort. | Eur. |
| Dianthus chinensis L. | j G. | Sap. | Eur. Tem. As. |
| Gypsophila Struthium L. | G. L. | Sap. | Spain. |
| Herniaria glabra L. | G. | Sap. | Eur. N. As. |
| Lychnis chalcedonica L. | G. G. G. | Sap. | Japan. |
| Lychnis dioica L. | G. | Sap. | Eur. N. As. |
| Lychnis Flos-cuculi L. | G. | Sap. | Eur. N. As. |
| Lychnis indica Benth. | G. | Sap. | Him. Reg. |
| Polycarpaea sp. | G. | Sap. | Austr. Pac. Is. |
| a | 0.01 | | Trop. |
| Saponaria officinalis L. | G. C. L. | IIr. Sap. | Eur. W. As. |
| Saponaria Vaccaria L. | G. | Sap. | Eur. As. Min. |
| 0.1 | | | Siber. |
| Silene antirrhina L. | Ches. | ļ | E. N. Am. |
| Silene Cucubalis Wibel. | G. | 1 | Eur. N. Afr. |
| C'1 | 1 | | Him. Reg. |
| Silene Griffithii Boiss. | G. | | Him. Reg. |
| Silene macrosolen Steud. | G. | | Abyss. |
| Silenc virginica L. | G. G. G. | Ant. | N. Am. |
| Silene viscosa Pers. | G. | | Eur. N. Am. |
| Stellaria crassifolia Ehrh. | G. | 1 | N. & Arct. Reg. |

CELASTRACEAE

| Catha edulis Forsk. | G. | Stim. | Trop. Afr. |
|------------------------------|----------|-------|-----------------|
| Celastrus scandens L. | Ches. L. | Em. | E. N. Am. |
| Elaeodendron orientale Jacq. | G. | İ | Madag, Is. |
| • | l | 1 | Maurit. |
| Euonymus atropurpureus Jacq. | G. L. | Em. | N. Am. |
| Euonymus Europaeus L. | | | Eur. As. Min. |
| Euonymus latifolius Mill. | | Em. | Eur. As. Minor. |
| Kurrimia zeylanica Arn. | G. | Hyd. | Zeyl. |
| Lophopetalum pallidum Laws. | G. | | Malacca. |

CHENOPODIACEAE

| Atriplex Halimus L. | G. | Sap. | N. & S. Afr. |
|-----------------------------|------------|----------|-----------------|
| Atriplex hortensis L. |] G. L. | Sap. Em. | Cosmon. Cult. |
| Atriplex laciniata L. | G. | Sap. | Eur. N. Am. |
| Atriplex Nuttallii S. Wat. | G. | Sap. | W. N. Am. |
| Atriplex rosea L. | G. | Sap. | Eur. N. Am. N. |
| • | 1 | | Afr. As. Minor. |
| Atriplex tatarica L. | G. | Sap. | Cent. As. |
| Atriplex vesicaria Hew. | G. | Sap. | Austr. |
| Chenopodium ambrosioides L. | G.Rusby L. | Ant. | Temp. & Trop. |
| | | | |

| NAME OF PLANT | Authority | Properties | Locality |
|--|----------------|----------------------|---|
| Chenopodium anthelminticum L. | Sm. L. | Ant, | Reg. N. Am. Temp. & Trop. Reg. N. Am. |
| Chenopodium Bonus-Henricus L. Chenopodium Botrys L. | L. G. L. | Ant. | Eu. N. Am. N. Reg. N.Am. |
| Chenopodium californicum S. Wat. Chenopodium hybridum L. | G. G. | Sap. | Eur. Cal. N. Temp. Reg. |
| Chenopodium mexicanum Moq. Chenopodium polyspermum L. | G. R. G. | Sap. F. P. | N. Am. Eur. Mex. Eur. N. As. |
| Chenopodium rubrum L. Chenopodium Vulvaria L. Eurotia ceratoides C. A. Mey. | Sm. L. G. | Ant. Sap. | Eur. N. As. Eur. N. Am. Cal. Eu. N. As. |
| Halocnemum fruticosum D. Dietr. Kochia arenaria Roth, Kochia scoparia Schrad. | G. G. G. | Ant. Sap. Sap. | W. N. Am. S. Eur. N. Afr. S. Eur. W. As. Eur. W. As. |
| Kochia trichophylla Hort. Salsola Kali L. | G. G. | Sap. Ant. | N. Am. N. Eur. As. N. |
| Salsola Kali var tenuifolia G. F.W. Mey. Salsola tamariscifolia Cav. Sarcobatus Maximilianii Nees. | G. Ches. | Ir. Ant. | Am, Eu. As. N. Am. Eur. Ry. Mts. W. N. Am. |

CISTACEAE COMBRETACEAE

| Combretum bracteosum, Brandis | G. | Sap. | |
|--------------------------------|-------|-------|------------|
| Combretum erythrophyllum Sond. | G. | | S. Afr. |
| Combretum phaneropetalum Bak. | G. | | S. Afr. |
| Combretum racemosum Beauv. | G. | ĺ | Trop. Afr. |
| Combretum trifoliatum Vent. | G. | Ant. | Hab. |
| Gustavia augusta L. | G. R. | F. P. | N. Granat. |
| Gustavia brasiliana DC. | G. R. | F. P. | Brazil |
| Quisqualis indica L. | | | Tr. As. |
| Terminalia Bellerica Roxb. | G. L. | F. P. | Ind. Malay |
| Terminalia Chebula Retz. | L. | Astr. | Tr. As. |
| Terminalia tomentosa Wight. | G. | | Ind. |

COMMELINACEAE

| Athyrocarpus persicariaefolium Hems | |]_ | Panama |
|-------------------------------------|-------|------|----------|
| Commelina deficiens Herb. | G. | Sap. | Braz. |
| Commelina nudiflora L. | G. | 1 | Tr. Reg. |
| Commelina tuberosa L. | G. L. | | Mex. |
| Tradescantia crassifolia Cav. | P. | | S. Am. |
| Tradescantia elongata G. F. W. Mey. | G. | ĺ. | IS. Am. |

COMPOSITAE

| Achillea Ptarmica L. | C. L. | lAcr. | N. Temp, Reg. |
|------------------------------|-------|-------|-----------------|
| Anacyclus officinarum Hayne. | G. L. | Hyd. | S. Eur. |
| Anacyclus pedunculatus Pers. | G. | lr. | Barbar. |
| Anacyclus Pyrethrum DC. | | | N. Afr. Orient. |
| Anthemis aetnensis Schouw. | G. | Hyd. | Mt. Aetna |
| Anthemis altissima L. | G. | Hyd. | Eur. Orient. |

| Anthemis arvensis L. Anthemis austriaca Jacq. Anthemis Blancheana Boiss. Anthemis Cota L. Anthemis Cotula L. Anthemis cotula L. Anthemis montana L. Anthemis montana L. Anthemis montana L. Arrica alpina Olin and Ladau. Arnica andicaulis Ell. Artemisia maritima L. Artemisia cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centaurea Calcitrapa L. Centaurea Calcitrapa L. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum parthenium Bernh. Chrysanthemum parthenium Bernh. Chrysanthemum parthenium Bernh. Chrysanthemum pardalianches L. Dysodia papposa Hitch, Comono sulphureus Cav. Crepis lacera Ten. Cyunara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Edinion Schata G. L. G. | | | | |
|--|-------------------------------|-----------|-------------|----------------|
| Anthemis austriaca Jacq. Anthemis Blancheana Boiss. Anthemis Cota L. Anthemis Cotula L. Anthemis Cotula L. Anthemis Cotula L. Anthemis elbuensis Anthemis montana L. Anthemis montana L. Arnica alpina Olin and Ladau. Arnica alpina Olin and Ladau. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia arenaria DC. Artemisia mexicana Wild. Artemisia pontica L. Artemisia pontica L. Artemisia pontica L. Artemisia yulgaris. Athanasia amara L. Artensisia yulgaris. Athanasia amara L. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centaurea Calcitrapa L. Chrysanthemum Balsamita L. Chrysanthemum Barthenium Bernh. Chrysanthemum pinnatifidum L. Chryscocma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elystopappus glandolusus Less. Eupatorium amarissimum Eupa | NAME OF PLANT | Authority | Properties | Locality |
| Anthemis austriaca Jacq. Anthemis Blancheana Boiss. Anthemis Cota L. Anthemis Cotula L. Anthemis Cotula L. Anthemis Cotula L. Anthemis elbuensis Anthemis montana L. Anthemis montana L. Arnica alpina Olin and Ladau. Arnica alpina Olin and Ladau. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia arenaria DC. Artemisia mexicana Wild. Artemisia pontica L. Artemisia pontica L. Artemisia pontica L. Artemisia yulgaris. Athanasia amara L. Artensisia yulgaris. Athanasia amara L. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centaurea Calcitrapa L. Chrysanthemum Balsamita L. Chrysanthemum Barthenium Bernh. Chrysanthemum pinnatifidum L. Chryscocma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elystopappus glandolusus Less. Eupatorium amarissimum Eupa | Anthemis arvensis L. | G. | Hvd | Eur Orient |
| Anthemis Blancheana Boiss. Anthemis Cota L. Anthemis Cotula L. Anthemis Cotula L. Anthemis elbuensis Anthemis montana L. Anthemis montana L. Anthemis montana L. Anthemis nobilis L. Antrica alpina Olin and Ladau. Arnica Chamissonis Less. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia maritima L. Artemisia maritima L. Artemisia pontica L. Artemisia maritima L. Artemisia pontica L. Artemisia pontota L. Cardopatium corymbosum Pers. Carlina acaulis L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum pinnatifidum L. Chrysonthemum Pinn | | | | |
| Anthemis cota L. Anthemis Cota L. Anthemis Cota L. Anthemis Cota L. Anthemis elbuensis Anthemis montana L. Anthemis montana L. Arnica alpina Olin and Ladau. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia maritima L. Artemisia mexicana Willd. Artemisia pontica L. Artemisia pontica L. Artemisia vulgaris. Athanasia amara L. Artensisia vulgaris. Athanasia amara L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum pinnatifidum L. Chrysanthemum Parthenium Bernh | Anthomia Plancheana Poice | | | |
| Anthemis Cota L. Anthemis Cotula L. Anthemis elbuensis Anthemis montana L. Anthemis nobilis L. Anthemis nobilis L. Antrica alpina Olin and Ladau. Arnica alpina Olin and Ladau. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia abrotinlium L. Artemisia arenaria DC. Artemisia maritima L. Artemisia maritima L. Artemisia maritima L. Artemisia pontica L. Cartina acaulis L. Cardopatium corymbosum Pers. Carlina acaulis L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum pinnatifidum L. Chrysonthemum pinnatifidum L. Chrysonthemu | | | | |
| Anthemis Cotula L. Anthemis elbuensis Anthemis montana L. Anthemis nobilis L. Arnica alpina Olin and Ladau. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia maritima L. Artemisia pontica L. Artemisia pontica L. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Chrysanthemum Balsamita L. Chrysanthemum Banthan L. Chrysanthemum Parthenium Bernh. Chrysanthemum Balsamita L. Chrysanthemum Balsamita | | | | |
| Anthemis elbuensis Anthemis montana L. Anthemis montana L. Anthemis nobilis L. Antrica alpina Olin and Ladau. Arnica alpina Olin and Ladau. Arnica chamissonis Less. Arnica montana L. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia mexicana Willd. Artemisia in exicana Willd. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centaurea Calcitrapa L. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum marifidum L. Chrysanthemum marifidum L. Chrysanthemum parthenium Bernh. Chrysanthemum parthenium Bernh. Chrysanthemum parthenium Bernh. Chrysanthemum pardalianches L. Dynorphotheca Ecklonis DC. Dimorphotheca Pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium annarissimum Euratorium annarismum Euratorium annarismum Euratorium annarismum Euratorium annarismum Euratorium ann | | | | |
| Anthemis elbuensis Anthemis montana L. Anthemis montana L. Anthemis nobilis L. Arnica alpina Olin and Ladau. Arnica chamissonis Less. Arnica montana L. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia mexicana Willd. Artemisia pontica L. Artemisia pontica L. Artemisia wilgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Chrysanthemum Balsamita L. Chrysanthemum cinerariaefolium Vis. Chrysanthemum Parthenium Bernh. Chrysanthemum pantaliidum L. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysanthemum corea cardunculus L. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Euratorium amarisma Pers. G. L. G. L | Anthemis Cotula L. | G. L. | Ir. | |
| Anthemis nobilis L. Arnica alpina Olin and Ladau. Arnica Chamissonis Less. Arnica montana L. Arnica mudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia mexicana Willd. Artemisia pontica L. Artemisia pontica L. Artemisia vulgaris. Athanasia amara L. Artactylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Barbanco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca Pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. G. L. Ant. Trop. Am. Austr. Cal. Cal. Cal. Cal. Cal. Cal. Cal. Cal | Anthemis elbuensis | G. | Hyd. | |
| Anthemis nobilis L. Arnica alpina Olin and Ladau. Arnica Chamissonis Less. Arnica montana L. Arnica mudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia mexicana Willd. Artemisia pontica L. Artemisia pontica L. Artemisia vulgaris. Athanasia amara L. Artactylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Barbanco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca Pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. G. L. Ant. Trop. Am. Austr. Cal. Cal. Cal. Cal. Cal. Cal. Cal. Cal | Anthemis montana L. | l G. | Hvd. | Eur. Orient. |
| Arnica Alpina Olin and Ladau. Arnica Chamissonis Less. Arnica montana L. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia maritima L. Artemisia maritima L. Artemisia trifida Nutt. Artemisia trifida Nutt. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysooma Coma—aurea L. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Eur. N. As. L. L. Nar. Nar. L. Nar. Nar. L. Nar. Nar. L. Nar. L. Nar. L. C. L. Rusby Sm. L. C. L. Ant. G. L. G. L. G. L. G. L. G. L. J. Ant. G. L. G. L. J. Ant. G. L. J. Ant. G. L. C. Diur. N. Af. As. As. As. As. As. As. As. As. C | Anthemis nobilis L. | L. | Naus, Stim. | Eu. As. |
| Arnica Chamissonis Less. Arnica montana L. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia mexiciana Willd. Artemisia pontica L. Artemisia mexicana Willd. Artemisia pontica L. Antisp. G. L. G. L | | | | |
| Arnica montana L. Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Abrotanum L. Artemisia arenaria DC. Artemisia maritima L. Artemisia pontica L. Artemisia pontica L. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centaurea Calcitrapa L. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Clibadium Barbasco DC. Clibadium Barbasco DC. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Comun. G. G. C. L. G. L. Ant. Ant. Eur. Cau. Reg. L. Ant. Eur. Cau. Reg. L. Ant. Eur. Ant. Eur. Cau. Reg. L. Ant. G. C. L. G. L. Ins. G. L. G. L. G. L. Ins. Diur. Eur. N Af. Eur. N Af. Eur. Na. Feur. Na. Sca. Ing. Na. Med. Reg. Beru. F | | | | |
| Arnica nudicaulis Ell. Artemisia Abrotanum L. Artemisia Absinthium L. Artemisia arenaria DC. Artemisia maritima L. Artemisia mexicana Willd. Artemisia pontica L. Artemisia pontica L. Artemisia pontica L. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum Parthenium Bernh. Chrysanthemum Parthenium Bernh. Chrysanthemum Parthenium Bernh. Chrysanthemum Barbasco DC. Clibadium Barbasco DC. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca pluvalis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. G. L. Anti. Heh. Ant. Ant. Eur. Cau. Reg. L. Anti. B. Ant. G. L. Anti. G. L. Ins. G. L. Ins. Ant. G. L. Ins. Ant. Eur. Cau. Reg. L. Ant. Diaph. W. N. A. Ineb. Ant. L. Antisp. Ca. L. Ant. G. L. G. L. G. L. G. L. Ins. Ant. Eur. Cau. Reg. L. Ant. Ca. L. Ins. Ant. G. L. F. P. G. R. E. G. L. Ins. Ant. G. L. Ins. Ant. Eur. Cau. Reg. L. Ant. Ca. L. Ins. G. L. Ins. G. L. Ins. G. L. Ins. G. L. Ins. O. Ins. O. Ins. O. | | | | |
| Artemisia Absinthium L. Artemisia arenaria DC. Artemisia maritima L. Artemisia mexicana Willd. Artemisia prontica L. Artemisia trifida Nutt. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centipeda orbicularis Lour. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum cinerariaefolium Vis. Chrysanthemum pinnatifidum L. Chrysanthemum prinnatifidum L. Chrysocoma Coma—aurea L. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia pappos Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elephantorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. General G. L. Incb. Nar. Ant. Eur. Cau. Reg. L. G. Ant. G. L. Diiaph. Ant. Cau. Reg. Lu. Cau. Reg. Lu. Ant. Cau. Reg. Lu. Ant. Cau. Reg. Lu. Cau. Reg. Lu. Ant. Cau. Reg. Lu. Ant. Eur. Cau. Reg. Ant. Cau. Reg. Lu. Ant. Cau. Reg. Ant. Cau. Reg. Ant. Cau. Reg. Ant. Cau. Reg. Lu. Ant. Cau. Reg. Ant. Ca. L. G. L. Ins. Ant. G. L. Ant. W. As. Austr. Pac. Is. Ant. G. L. Ant. Cum. Ant. Cau. Reg. Ant. Cau. Reg. Ant. Cau. Reg. Ant. Cau. Reg. Ant. G. L. G. L. G. L. G. L. G. L. G. L. G. | Timea montana 2, | | | 12ai. 14. 11. |
| Artemisia Absinthium L. Artemisia arenaria DC. Artemisia maritima L. Artemisia mexicana Willd. Artemisia prontica L. Artemisia trifida Nutt. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centipeda orbicularis Lour. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum cinerariaefolium Vis. Chrysanthemum pinnatifidum L. Chrysanthemum prinnatifidum L. Chrysocoma Coma—aurea L. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia pappos Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elephantorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. General G. L. Incb. Nar. Ant. Eur. Cau. Reg. L. G. Ant. G. L. Diiaph. Ant. Cau. Reg. Lu. Cau. Reg. Lu. Ant. Cau. Reg. Lu. Ant. Cau. Reg. Lu. Cau. Reg. Lu. Ant. Cau. Reg. Lu. Ant. Eur. Cau. Reg. Ant. Cau. Reg. Lu. Ant. Cau. Reg. Ant. Cau. Reg. Ant. Cau. Reg. Ant. Cau. Reg. Lu. Ant. Cau. Reg. Ant. Ca. L. G. L. Ins. Ant. G. L. Ant. W. As. Austr. Pac. Is. Ant. G. L. Ant. Cum. Ant. Cau. Reg. Ant. Cau. Reg. Ant. Cau. Reg. Ant. Cau. Reg. Ant. G. L. G. L. G. L. G. L. G. L. G. L. G. | Arnica nudicaulis Ell. | L. | Nar. | N. A. |
| Artemisia Absinthium L. Artemisia arenaria DC. Artemisia maritima L. Artemisia mexicana Willd. Artemisia pontica L. Artemisia trifida Nutt. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centiaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elephantopus glandolusus Less. Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Geneplosis eriocarpa Benth. Grindelia Tournefortii L. G. C. L. Rusby Ant. L. Ant. C. Cau. Reg. Lunch. Ant. Mex. Ant. C. C. L. C. Ant. C. C. L. C. Ant. C. C. L. C. L. Ant. C. C. L. Ant. C. C. L. Ant. C. C. L. C. L. Ant. C. C. L. C. L. C. L. C. L. Ant. C. C. L. Ant. C. C. L. C. L. Ant. C. C. L. C. L. C. L. C. L. Ant. C. C. L. Ant. C. C. L. Ant. C. C. L. C. | Artemisia Abrotanum L. | 1 G. L. | Ineb. | Eur. Temp. A. |
| Artemisia arenaria DC. Artemisia maritima L. Artemisia maritima L. Artemisia montica L. Artemisia pontica L. Artemisia pontica L. Artemisia trifida Nutt. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC, Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum cinerariaefolium Vis. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis C. Dimorphotheca Ecklonis C. Dimorphotheca Ecklonis C. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L, Elptropappus glandolusus Less. Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Genephosis eriocarpa Benth, Grindelia Tournefortii L. Rusby Sm. L. G. Ant. Ant. Cau. Reg. Lant. Ant. Cau. Reg. Lant. Ant. Cau. Ant. Cau. Reg. Lant. Cau. Reg. Lant. Ant. Cau. Reg. Lant. Ant. Cau. Reg. Lant. Cau. Reg. Lant. Cau. Reg. Lant. Ant. Cau. Reg. Lant. Cau. Reg. Ant. Ca. Ca. L. G. L | | G. C. L. | | |
| Artemisia arenaria DC. Artemisia maritima L. Artemisia mexicana Willd. Artemisia pontica L. Artemisia pontica L. Artemisia pontica L. Artemisia vulgaris. Athanasia amara L. Atractylis gunmifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pardalianches L. Clibadium Barbasco DC. | | | | Fur |
| Artemisia maritima L. Artemisia mericana Willd. Artemisia pontica L. Artemisia pontica L. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum Parthenium Bernh. Chrysanthemum Parthenium Bernh. Chrysanthemum Parthenium Bernh. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium cannabinum L. Eurybia Med. Reg. G. C. L. G. C. Cum. Madera Is. S. Afr. Eur. Cum. Madera Is. S. Afr. F. P. G. G. Diur. G. G. Diur. Med. Reg. S. Afr. F. P. S. Am. Mex. Cibiadium surinamense L. G. G. G. G. Diur. Med. Reg. S. Afr. F. P. S. Am. Mex. Cibiadium Surinamense L. G. G. G. Diur. G. G. Diur. Med. Reg. S. Afr. F. P. S. Am. Mex. Cibiadium Med. Reg. C. Cum. Madera Is. S. Afr. F. P. S. Am. Mex. Cibiadium Med. Reg. C. Cum. Madera Is. S. Afr. F. P. S. Am. Mex. Cibiadium Med. Reg. C. Cum. Madera Is. S. Afr. F. P. S. Am. Mex. Cibiadium Med. Reg. C. | Artemisia arenaria DC | | | |
| Artemisia mexicana Willd. Artemisia pontica L. Artemisia trifida Nutt. Artemisia vulgaris. Athanasia amara L. Artactylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum Parthenium Bernh. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium Barbasco DC. Clibadium Burinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elephantopus contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia robusta Nutt. Grindelia Tournefortii L. Grindelia Mutt. Grindelia Mutt. Grindelia Mutt. Grindelia Mutt. Ant. Ant. Ant. Ant. Ant. Ant. Ant. | | | | |
| Artemisia pontica L. Artemisia trifida Nutt. Artemisia trifida Nutt. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium cannabinum L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Genephosis eriocarpa Benth. Grindelia Tournefortii L. Ant. Diaph. Ant. Med. Reg. Med. Reg. Ant. G. C. L. G. L. Diur. G. L. Diur. Cantispas. Ca. Ant. G. L. Diur. Cantispas. Ca. Ant. Ant. Co. Ant. Ant. Co. Ca. L. Diur. Cantispas. Ca. Ca. L. Diur. Cantispas. Ca. Ca. Ca. Ca. Ca. Ca. Ca. Ca. Ca. Ca | | | | |
| Artemisia trifida Nutt. Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Balsamita L. Chrysanthemum parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. Grindelia Tournefortii L. L. Diaph. Antisp. G. C. L. G. C. L. G. | | | 1 . | |
| Artemisia vulgaris. Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum cinerariaefolium Vis. Chrysanthemum cinerariaefolium Vis. Chrysanthemum parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L, Clibadium Barbasco DC. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elytropappus glandolusus Less. Eupatorium cannabinum L. Elytropappus glandolusus Less. Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. Grindelia Tournefortii L. Antispa. G. L. G. | | | | Eur. Cau. Rcg. |
| Athanasia amara L. Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum cinerariaefolium Vis. Chrysanthemum pinnatifidum L. Chrysanthemum | | | Diaph. | W. N. A. |
| Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Cinerariaefolium Vis. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chryscooma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca Pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. C. L. G. Diur. G. G. C. L. G. Diur. G. G. C. L. G. Diur. Med. Reg. Brazil As. Min. Med. Reg. Diur. Feur. N. Af. Eur. N. As. Ant. G. L. Diur. G. L. G. L. G. L. G. L. G. L. Diur. G. L. Ant. G. L. G. | Artemisia vulgaris. | | Antisp. | Tem. Reg. |
| Atractylis gummifera L. Baccharis cordifolia DC. Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum cinerariaefolium Vis. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chryscoma Coma—aurea L. Clibadium Barbasco DC. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. C. L. G. L. Diur. Had. Reg. Brazil As. Min. Med. Pal. Brac. II. II. II. II. III. III. III. III. I | Athanasia amara L. | | Ant. | |
| Baccharis cordifolia DC, Cardopatium corymbosum Pers. Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth, Grindelia Tournefortii L. G. L. G. L. G. L. G. L. G. L. G. L. G. L. G. L. Coum. Madera Is. G. G. L. F. P. G. R. E. G. R. E. G. Diur. Med. Reg. Cum. S. Afr. F. P. G. Diur. Med. Reg. Dalmatia U. Cum. G. G. G. G. Hyd. Sap. S. Afr. Ir. Eur. As. Iv. Iv. Iv. N. An. G. Italy Med. Reg. Italy Med. Reg. Italy Med. Reg. Dalmatia U. Cum. G. G. G. I. Ins. G. Ins. Cal. Ins. C | Atractylis gummifera L. | G. C. L. | | Med. Reg. |
| Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysaoroma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca Ecklonis DC. Dimorphotheca Pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Euyatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. L. Diur. G. L. Diur. As. Min. Med. Reg. Ant. As. Austr. Temp. As. Ant. G. L. Cum. Madera Is. G. R. E. F. P. G. R. E. G. Diur. Med. Reg. S. Afr. F. P. S. Am. G. Diur. Med. Reg. S. Afr. Italy G. Diur. Med. Reg. S. Afr. S. Afr | Baccharis cordifolia DC. | G. L. | Poi. Alk. | |
| Carlina acaulis L. Centaurea Calcitrapa L. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elephantopus tomentosus L. Elephantopus tomentosus L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. L. Diur. G. L. Diur. Heg. Eur. N. As. Austr. Pac. Is. Ant. G. L. Ins. G. C. L. Ins. G. C. L. G. C. L. G. C. L. Ins. G. C. L. G. C. L. G. C. L. G. C. L. Ins. G. C. L. G. C. L. Ins. G. C. L. Ins. G. C. L. G. C. L. G. C. L. Ins. G. C. L. Ins. G. C. L. Ins. G. C. L. G. | | | | |
| Centipeda orbicularis Lour. Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. Ir. Ant. Ant. Ant. Ant. Ant. C. Ant. C. Ant. G. C. Ant. G. C. Ant. G. C. Ant. C. Ant. Cum. Madera Is. Cum. Madera Is. Cum. Madera Is. Cum. Madera Is. Cum. Med. Reg. Diur. G. Diur. G. Bur. C. Ir. Eur. As. N. Am. N. Am. N. Am. N. Am. S. Afr. Eur. As. Ir. Eur. As. Ir. Eur. As. Ir. Eur. As. Ir. Eur. N. As. S. Afr. Eur. Co. C. L. G. C. L. G. C. L. Antispas. Cal. Antispas. As. Austr. Temp. As. As. Austr. Pac. Is. N. Ant. Cum. Madera Is. Cum. Med. Reg. II. II. III. III. III. Antispas. Ant. Antispas. As. Austr. Cal. Antispas. As. Austr. Temp. As. As. Austr. Temp. As. As. Austr. Temp. As. As. Austr. Temp. As. As. Austr. Temp. As. As. Austr. Temp. As. As. As. As. As. As. Austr. Temp. As. As. As. As. As. As. As. A | · | | | |
| Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Cinerariaefolium Vis. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. L. Ant. G. L. Ins. G. L. Ins. G. L. G. Cum. Madera Is. Cum. Madera Is. F. P. S. Am. Mex. Cum. Med. Reg. Diur. G. Diur. G. Diur. Buit. G. G. L. Em. N. Am. N. Am. N. Am. S. Afr. Cum. Madera Is. S. Afr. F. P. S. Am. Mex. Italy Med. Reg. S. Afr. Ir. Eur. As. N. Am. N. Am. N. Am. S. Austr. Pac. Is. Ant. G. Cum. Madera Is. S. Afr. F. P. S. Am. Mex. G. Italy Med. Reg. S. Afr. Ir. Eur. As. N. Am. N. Am. S. Austr. Cal. G. Ant. G. Ant. G. Ant. G. Ant. G. Ant. Cal. Antispas. Cal. Antispas. Cal. Antispas. Cal. Cal. Cal. Cal. Cal. Cal. Cal. Cal | Carlina acaulis L. | G. L. | Diur. | Eur. |
| Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. L. Ant. G. L. Ins. G. L. Ins. G. C. L. G. R. E. F. P. G. R. E. F. P. G. R. E. F. P. S. Am. Mex. G. Diur. Med. Reg. Diur. G. Ir. Eur. As. N. Am. N. Am. N. Am. N. Am. S. Afr. G. L. G. L. G. L. Em. N. Am. G. S. Afr. Eur. As. N. Am. N. Am. S. Afr. Cum. Madera Is. S. Afr. G. Italy G. Diur. Eur. N. As. G. Is. G. J. G. J. J. Antispas. Cal. Ant. Cal. Ant. Cal. Ant. Cal. Cal. Antispas. Cal. Ant. Cal. Antispas. Cal. Antispas. Cal. Antispas. Cal. Antispas. Cal. Antispas. Cal. Cal. Cal. Cal. Chrysanthemum calls. Cal. Antispas. Cal. Cal. Cal. Cal. Cal. Cal. Cal. Cal | Centaurea Calcitrapa L. | L. | Ir. | N. Af. Eur. |
| Centipeda orbicularis Lour. Chrysanthemum Balsamita L. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. L. G. L. G. L. Ins. G. C. L. Ins. G. C. Ins. G. C. Ins. G. Dalmatia Eur. Madera Is. Ccum. Madera Is. G. Cum. G. G. Diur. Med. Reg. Diur. Med. Reg. S. Afr. Ir. Eur. As. Ir. N. Am. N. Am. S. Austr. Pac. Is. Ant. G. L. Dalmatia Eur. S. Afr. G. C. Italy Med. Reg. S. Afr. Ir. Eur. As. N. Am. N. Am. S. Austr. Pac. Is. Ant. G. L. Dalmatia Eur. S. Afr. G. Italy Med. Reg. S. Afr. Ir. Eur. As. N. Am. N. Am. S. Austr. Pac. Is. Ant. G. L. Dalmatia Eur. S. Afr. G. Italy Med. Reg. S. Afr. Ir. Eur. As. N. Am. N. Am. S. Austr. Pac. Is. Ant. G. L. Joiur. Eur. N. As. Cal. G. L. Ant. G. L. Antispas. Cal. Ant. Cal. Cal. Cal. Cal. Cal. Cal. Cal. Cal | | | | Temp. As, |
| Chrysanthemum Balsamita L. Chrysanthemum cinerariaefolium Vis. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elephantopus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. C. L. G. L. Ins. Ant. G. L. Ins. Cum. G. Cum. Madera Is. Cum. Madera Is. Cum. Madera Is. Cum. G. Cum. Madera Is. Cum. Madera Is. Cum. Med. Reg. Italy G. Diur. G. It. Eur. As. Ir. N. Am. S. Afr. Cum. Med. Reg. Italy G. It. Ir. Eur. As. Ir. N. Am. S. Afr. Cum. Med. Reg. Italy Co. Ins. G. It. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir | Centipeda orbicularis Lour. | G. | Ant | |
| Chrysanthemum Balsamita L. Chrysanthemum cinerariaefolium Vis. Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. C | | | | |
| Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. C. L. Ins. Ant. Cum. Madera Is. Cum. Med. Reg. Itr. Eur. As. Iv. Eur. As. N. Am. Cal. Ins. Ant. Cum. Med. Reg. Itr. Eur. As. Iv. Eur. N. As. Cal. Cum. Med. Reg. Italy Cum. Med. Cum. Med. Reg. Italy Cum. Med. Reg. Italy Cum. Med. Reg. Italy C | Chrysanthemum Balsamita L. | G. L. | Ant. | |
| Chrysanthemum Parthenium Bernh. Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. C. L. Ins. Ant. Cum. Madera Is. Cum. Med. Reg. Itr. Eur. As. Iv. Eur. As. N. Am. Cal. Ins. Ant. Cum. Med. Reg. Itr. Eur. As. Iv. Eur. N. As. Cal. Cum. Med. Reg. Italy Cum. Med. Cum. Med. Reg. Italy Cum. Med. Reg. Italy Cum. Med. Reg. Italy C | | G. L. | | |
| Chrysanthemum pinnatifidum L. Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. Grindelia Tournefortii L. G. Cum. G. G. F. P. G. G. F. P. S. Am. Mex. G. Hyd. Sap. G. Hyd. Sap. S. Afr. Ir. Eur. As. N. Am. N. Am. S. Afr. G. Is. G. Ins. G. Joiur. G. Joiur. Eur. N. As. S. Afr. Eur. As. Diur. Eur. N. Am. S. Afr. G. Joiur. G. Joiur. G. Joiur. Joint Eur. Joint | | G C I. | Ins Ant | |
| Chrysocoma Coma—aurea L. Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elephantopus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Guephosis eriocarpa Benth. Grindelia Tournefortii L. G. C. R. E. F. P. S. Am. Med. Reg. Diur. G. Hyd. Sap. S. Afr. G. Ir. Eur. As. Ir. N. Am. G. C. Ins. G. Ins. G. Ins. G. Joiur. G. Ins. G. Ins. G. Joiur. Am. N. Am. S. Afr. Cal. Trop. Am. Austr. Cal. G. Joiur. Antispas. Cal. Cal. Cal. Cal. Cal. Cal. Cal. Cal | | 0. C. 4. | | |
| Clibadium Barbasco DC. Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. Grindelia Tournefortii L. R. E. F. P. G. R. E. F. P. S. Am. Mex. G. Italy Med. Reg. Is. Ir. Ir. Ir. Iv. Ir. Iv. Ir. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv | Chrysocoma Coma—aurea I | Ğ. | Cuii. | |
| Clibadium surinamense L. Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. Grindelia Tournefortii L. GR. E. F. P. G. G. G. Diur. Med. Reg. S. Afr. Eur. As. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Iv. Ir. Ir. Iv. Ir. Iv. Ir. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv | | D E | E D | D. AII. |
| Cosmos sulphureus Cav. Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. G. G. Diur. G. Hyd. Sap. S. Afr. Ir. Eur. As. N. Am. N. Am. S. Afr. G. Ins. G. Ins. G. Ins. G. Joiur. Eur. N. As. Eur. Diur. Eur. N. As. G. Ant. G. Ant. Trop. Am. Austr. G. L. Antispas. G. Em. | | | | C 4 |
| Crepis lacera Ten. Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. Diur. G. Hyd. Sap. S. Afr. Ir. Eur. As. N. Am. S. Hem. N. Am. G. L. Em. G. Ins. G. L. Diur. Eun. Ir. Eur. As. N. Am. S. Afr. G. L. Diur. Eun. Trop. Am. Austr. Cal. Grindelia Tournefortii L. | | | [F. P. | |
| Cynara Cardunculus L. Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. Diur. G. Hyd. Sap. Med. Reg. Med. Reg. S. Afr. G. Ir. Eur. As. N. Am. S. Afr. G. Ir. S. Afr. Ir. N. Am. S. Am. S. Afr. G. Ins. C. Ins. Ins. Ins. Ins. Ins. Ins. Ins. Ins | | | | |
| Dimorphotheca Ecklonis DC. Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. G. Hyd. Sap. S. Afr. S. Afr. Eur. As. It. N. Am. S. Afr. G. Ins. G. Ins. G. Jins. Cal. Cal. Cal. Cal. Cal. Cal. Cal. Cal | | | | |
| Dimorphotheca pluvialis Moench. Doronicum pardalianches L. Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eur. As. Ir. G. Ir. G. Ir. Eur. As. Iv. N. Am. S. Afr. On M. Am. S. Afr. C. Ir. Ir. Ir. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv | Cynara Cardunculus L. | | | |
| Doronicum pardalianches L. Dysodia papposa Hitch. Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia Grindelia Tournefortii L. Grindelia Gri | Dimorphotheca Ecklonis DC. | | Hyd. Sap. | S. Afr. |
| Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium annarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. Grindelia Tournefortii L. G. G. G. G. G. G. Jins. Eur. N. As. Eur. N. As. Trop. Am. Austr. G. G. G. Jins. G. L. Ant. G. Austr. G. G. Jins. Eur. Eur. Ant. Trop. Am. Austr. Cal. Grindelia Tournefortii L. | | G. | | S. Afr. |
| Dysodia papposa Hitch, Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia robusta Nutt. Grindelia Tournefortii L. G. Ir. N. Am. N. Am. S. Afr. G. Ins. G. G. Joiur. G. Ant. G. Austr. G. Austr. G. L. Antispas. G. Em. | Doronicum pardalianches L. | | Ir. | Eur. As. |
| Echinops Ritro L. Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia Tournefortii L. Grindelia Tournefortii L. G. G. G. G. Jins. G. Jins. G. Jins. G. Jins. G. Jins. G. Jins. Eur. N. As. Eur. N. As. G. Ant. G. Austr. G. Jins. G. Jins. Eur. N. As. Eur. Trop. Am. Austr. Cal. G. Ein. | Dysodia papposa Hitch. | i | Ir. | |
| Elephantopus tomentosus L. Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia robusta Nutt. Grindelia Tournefortii L. G. L. Em. N. Am. S. Afr. G. L. Diur. Eur. N. As. G. Ant. Trop. Am. Austr. G. L. Antispas. G. L. Autispas. G. Em. | | G. | | |
| Elytropappus glandolusus Less. Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia robusta Nutt. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia G. G. G. Jins. G. Diur. Eur. N. As. G. Flaveria Cant. Ant. G. Ant. Austr. G. Cal. Antispas. Cal. G. Em. | Elephantopus tomentosus L. | G. L. | Em | |
| Eupatorium amarissimum Eupatorium cannabinum L. Eurybia moschata Elurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia robusta Nutt. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia G. G. Jins. Diur. Eur. N. As. Ca. Ant. Austr. G. Ca. Antispas. Cal. Em. | Elytropappus glandolusus Less | G. | J | IS Afr |
| Eurpatorium cannabinum L. Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia robusta Nutt. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia G. G. G. Joiur. Eur. N. As. Can. Trop. Am. G. Austr. G. L. Antispas. Cal. Em. | Eupatorium amarissimum | | IIne | D. Att. |
| Eurybia moschata Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia robusta Nutt. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia Tournefortii L. | | | 1 | IT's N. A. |
| Flaveria Contrayerba Pers. Gnephosis eriocarpa Benth. Grindelia robusta Nutt. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia Tournefortii L. | | | Diur. | Eur. N. As. |
| Gnephosis eriocarpa Benth. Grindelia robusta Nutt. Grindelia Tournefortii L. Grindelia Tournefortii L. Grindelia Tournefortii L. | | | 1 44 | 1/17 |
| Grindelia Tournefortii L. Grindelia Tournefortii L. G. L. Antispas. Cal. G. Em. | Guenhosis eriocaras Ponth | | Ant. | |
| Grindelia Tournefortii L. G. Em. | Grindelie reducte Nett | | 1 | |
| TT 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 | | | Antispas. | Cal. |
| S. Afr. | Hanlogarpha lurate II- | | Em. | 10 44 |
| | Trapiocarpha lyrata flav. | 1 (• | | S. Afr. |

| NAME OF PLANT | Authority | | Locality |
|--|-------------|----------------|----------------|
| Helenium autumnale L. | Ches. G. L. | Ir. Nar. | N. Am. |
| Helenium tenuifolium Nutt. | Ches. G. L. | Ir. Nar. | N.Am. S.States |
| Helianthus tuberosus L. | | | N. Am. |
| Helichrysum apiculatum D. Don | C. G. | Ant. | Austr. |
| Hieracium virosum Pall. | Ğ. | | N. As. Eur. |
| Ichthyothere Cunabi Mart. | E. G. R. | प भ | Brazil |
| Inula Royleana DC. | G. 7. | 1. 1. | Him, Reg. |
| Lactuca Scariola L. | G.C. Mi.L. | Nar | E. Eur. |
| Lactuca taraxacifolia Schum. | | Nar. | Tr. Afr. |
| Lactuca tatarica C. A. Mey. | Ğ. | 1 | W. As. In. |
| Lactuca virosa L. | G. Mi. L. | Nar | Eur. N. As. |
| Launaea pinnatifida Cass. | G. | l l l | As. Trop. Afr. |
| Montanoa floribunda C. Koch. | Ğ. | | Mex. |
| Montanoa tomentosa Cerv. | G. | | Mex. |
| Mutisia viciaefolia Cav. | G. L. | Sap. Sed. | Chili |
| | G. L. | | |
| Oldenburgia Arbuscula DC. | G. | Sap. | S. Afr. |
| Olearia macrodonta Bak. | | Sap. | N. Zeal. |
| Olearia moschata Hook. | G. | | N. Zeal. |
| Onopordon Acanthium L. | G. | | Eur. N. Afr. |
| | | | Orient. |
| Othonopsis intermedia Boiss. | G. | | Persia |
| Parthenium Hysterophorus L. | L. | } | N. A. Aus. |
| Parthenium integrifolium L. | į G. | Diur. | N. A. |
| Pentzia virgata Less. | G. | | S. Afr. |
| Perezia oxylepis, Sch. | G. | | Mex. |
| Petasites officinalis Moench. | G. | | Eur. N. As. |
| Picris hieracioides L. | L. | Bitter | Eur. Tem. As. |
| | | | Aus. N. Zea. |
| Prenanthes altissima L. | G. L. | Bitter | N. A. |
| Pterigeron adscendens Benth. | G. M. |) | Aus. |
| Pterocaulon pycnostachyum Ell. | G. | Nar. | N. A. |
| Pulicaria dysinterica Gaertn. | L. | Ins. | Eur. |
| Senecio gaudalensis. | Ches. | | Mex. |
| Senecio Grayanus Heimsl | | Åbort. | Mex. |
| Senecio Jacobaea L. | G. L. Ches. | Anodyne | Eur. N. As. N. |
| Senecio toluccanus DC. | G. | , | Mex. |
| Senecio vulgaris L. | G. L. | Astr. | Eur. |
| Senecio vulneraria DC. | G. | | Mex. |
| Silybum Marianum Gaertn. | L. | Exp. | Eur. |
| Solidago Virgaurea L. | G. L. | Diur. | N. Tem. Reg. |
| Spilanthes Acmella Murr. | G. L. | Pung. | Cosm. Trop. & |
| opnanties Henreita Mari. | 0. 2. | i ung. | Sub-trop. |
| Spilanthes oleracea L. | L. | Pung. | S. A. |
| Tanacetum umbelliferum Boiss. | Ğ. | Abort. | Afghan. |
| Tanacetum vulgare L. | G.Rusby L. | | Eur. N. As. |
| Vernonia anthelmintica Willd. | G. L. | Ant. | Him. Reg. |
| Vernonia nigritiana Oliv. and Hiern. | G. L. | Till. | Tr. Afr. |
| Xanthisma texanum DC. | G. | 1520 | Texas |
| Xanthium canadense Mill. | G. Ches. | Sap. Mech. | N. Am. |
| | G. Ches. | MICCH. | Cosm. |
| Xanthium spinosum L. Xanthium Strumarium L. | G. Ches. | | Cosm. |
| Aantinuiti Strumarium L. | G. Ches. | ! | ICOs. |

CONIFERAE

| Cephalotaxus drupacea Sieb and Zucc. | | | Jap. |
|--------------------------------------|------------|--------|--------------------|
| Ginkgo biloba L. | | Acr. | Jap. Eur. N. As |
| Juniperus Sabina L. | C.G. Mi.L. | | |
| Juniperus oxycedrus L. | Sm. | Abort. | S. Eur. Cauc. |
| | 1 | | Reg. |

| Properties Abort. Ir. Ineb. | Locality E. N. A. Eur. Cult. N. Am. S. Eur. Eur. N. As. |
|-----------------------------|--|
| Ir. Ineb. Ineb. | Eur. Cult. N. Am. S. Eur. |
| Ineb. Ineb. | N. Am. S. Eur. |
| Ineb. | S. Eur. |
| Ineb. | |
| | Eur. N. As. |
| r n | |
| | [Cal. |
| F. P. | Eur. |
| Astr. | W. N. A. |
| Astr. | E. N. Am. N. A. |
| ADOFE. | / |
| Actr | Jap. N. N. Am. |
| 1511, | 114. 14. Am. |
| | |
| | test. |
| | Trop. Afr. |
| | Mas. Is. |
| | Mad. Maurit. |
| - | Trop. Afr. |
| C | Trop. Afr. |
| con. | Venez. |
| | |
| Cath. | As. Minor |
| | Eur. N. A. |
| | St. Croix Is. |
| | [N. A. |
| A. P. | Aus. |
| ~ . | |
| Cath. | Eur. N. As. |
| Q .1 | N. Am. |
| Cath. | Eur. Orient. |
| T-4 | N. Am. |
| | Mex. |
| | Mex. |
| | S. E. U. S. |
| | Mex. |
| Ľm. | Am. Calid. |
| | Trop. Reg. |
| | |
| | Med. Reg. |
| | Him. China |
| | Peru |
| | N. Zeal. |
| | Peru |
| | |
| | Cosm. Trop. |
| Ir. | Jamaica |
| Exp. | Japan |
| p. | |
| Exp. | E. N. Am. |
| _ ^ | E. N. Am. N. A. |
| Exp. Astr. Astr. | E. N. Am. |
| Exp. Astr. | E. N. Am. N. A. N. A. E. N. Am. |
| Exp. Astr. Astr. | E. N. Am. N. A. N. A. |
| | Con. Cath. Cath. Em. Em. Pur. Pur. Em. |

CRASSULACEAE

| NAME OF PLANT | Authority | Properties | Locality |
|---|-----------------|------------|--|
| Cotyledon ventricosa Burm. Kalanchoe spathulata DC. Sedum acre L. Sempervivum montanum L. | G. G. Mi. L. G. | | Afr. Austr. Trop. As. Eur. N. As. N. Am. Cent. Eur. |

CRUCIFERAE

| Barbarea vulgaris R. Br. Brassica alba Boiss. Brassica arvensis I. Brassica nigra Koch. Capdamine amara L. Cardamine pratensis L. Cardamine pratensis L. Crambe filiformis Jacq. Diplotaxis erucoides DC. Erysimum cheiranthoides I. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium Draba L. Lepidium Owaihiense Cham, & Schlecht, Lepidium oleraceum Forst. Lepidium piscidium Forst. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus Raphanistrum L. Raphanus Sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium toxophyllum G. A. Mey. Vesicaria gnaphalodes Boiss. Sm. L. G. L. Antisc. Ir. Antisc. Eur. As. N. Am. Eu. As. N. Am. Eu. As. N. Am. Eu. As. N. Am. Ir. Eu. As. N. Am. Ir. Eur. Antisc. G. | CRUCIFERAE | | | |
|--|---------------------------------------|---------|----------|----------------|
| Brassica alba Boiss. Brassica arvensis L. Brassica arvensis L. Brassica nigra Koch. Capdamine amara L. Cardamine pratensis L. Cardinine L. Cardamine pratensis L. Cardamine pratensis L. Cardamine pratensis L. Cardinine L. Car | Barbarea vulgaris R. Br. | G. L. | Antisc. | Eur. Temp. As. |
| Brassica arvensis L. Brassica nigra Koch. Capsella Bursa-pastoris Medic. Cardamine amara L. Cardamine pratensis L. Cardinia Med. Res. N. Am. Eur. Antisc. Cardinia Med. Res. N. Am. Eur. Ant. Cardinia C. Cardinia Med. Res. N. Am. Cardinia C. Cardinia Med. Res. N. Am. Cardinia C. Card | | |) | |
| Brassica nigra Koch. Capsella Bursa-pastoris Medic. Cardamine amara L. Cardamine pratensis L. Crambe filiformis Jacq. Diplotaxis erucoides DC. *Eruca sativa Mill. Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium Owaihiense Cham. & Schlecht. Lepidium oleraceum Forst. Lepidium owaihiense Cham. & Schlecht. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. E. L. G. C. L. G. C. L. G. C. L. G. L. G. C. L. G. C. L. G. R. G. Whi. L. G. G. G. Whi. L. G. G. G. Whi. L. G. G. G. Whisc. G. | | | | |
| Capsella Bursa-pastoris Medic. Cardamine amara L. Cardamine pratensis L. Crambe filiformis Jacq. Diplotaxis erucoides DC. *Eruca sativa Mill. Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium oleraceum Forst. Lepidium owahiense Cham. & Schlecht. Lepidium owahiense Cham. & Schlecht. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus Raphanistrum L. Raphanus Raphanistrum L. Raphanus Sophia L. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. G. G. G. G. J. Wilcox* Ir. Wilcox* Ir. Sm. L. G. G. G. Si. Jr. C. Jr. Sm. Jr. Eur. Anti. Eur. Sm. Jr. Eur. Corient. Eur. Antisc. G. R. G. R. F. P. Pac. Is. Orient. Eur. Antisc. Comop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. Temp, Reg. Eur. Orient. Eur. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. | | | | |
| Cardamine amara L. Cardamine pratensis L. Cardamine pratensis L. Crambe filiformis Jacq. Diplotaxis erucoides DC. Diplotaxis tenuifolia DC. *Eruca sativa Mill. Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium latifolium L, Lepidium oleraceum Forst. Lepidium Owaihiense Cham. & Schlecht. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus Raphanistrum L. Raphanus Sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. G. L. Antisc. C. Mattisc. G. J. Antisc. C. J. Antisc. C. J. Antisc. C. L. Antisc. C. L. Antisc. C. J. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. | | | | |
| Cardamine pratensis L. Crambe filiformis Jacq. Diplotaxis erucoides DC. Diplotaxis tenuifolia DC. *Eruca sativa Mill. Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium Oleraceum Forst. Lepidium Owaihiense Cham. & Schlecht. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus Raphanistrum L. Raphanus Raphanistrum L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. L. G. L. G. L. G. L. G. R. F. P. Eur. Orient. Eur. Antisc. G. R. F. P. Pac. Is. Orient. Egypt, Arab. Eur. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp, Reg. Pat. Chili Med. Reg. As. Minor Ir. Eur. F. P. Eur. Crient. Eur. As.Orient. Egypt, Arab. Eur.As. N. Am. Ir. Eur. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Temp. Reg. Temp. Reg. Temp. Reg. Ear. N. As. | | | Sap. | |
| Crambe filiformis Jacq. Diplotaxis erucoides DC. Diplotaxis tenuifolia DC. *Eruca sativa Mill. Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium oleraceum Forst. Lepidium Owaihiense Cham, & Schlecht. Lepidium piscidium Forst. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L, Raphanus Raphanistrum L, Raphanus Sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. L. G. L. G. R. G. R. F. P. Hawai. Is. F. P. Pac. Is. Orient. Eur. As.Orient. F. P. Pac. Is. Orient. Egypt, Arab. Eur.As, N. Am. Ir. Eur. Antisc. G. C. L. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Temp. Reg. Eur. N. As. | | | | |
| Diplotaxis tenuifolia DC. *Eruca sativa Mill. Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium oleraceum Forst. Lepidium owaihiense Cham, & Schlecht. Lepidium piscidium Forst. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L, Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. L. Unit C. L. Wilcox* Ir. Sm. L. G. G. G. G. G. G. G. F. P. Hawai. Is. F. P. Pac. Is. Orient. Eur. As.Orient. F. P. Pac. Is. Orient. Eur. Antisc. G. L. Antisc. C. L. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. | | | Antisc. | |
| Diplotaxis tenuifolia DC. *Eruca sativa Mill. Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium Intifolium L, Lepidium Owaihiense Cham. & Schlecht. Lepidium piscidium Forst. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus Raphanistrum L. Raphanus Sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Ir. Sm. L. G. G. G. G. G. F. P. Hawai. Is. F. P. Pac. Is. Orient. Egypt, Arab. Eur.As. N. Am. Eur. Antisc. C. L. Ir. C. L. Antisc. G. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. | | | T | |
| Diplotaxis tenuifolia DC. *Eruca sativa Mill. Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium latifolium L, Lepidium oleraceum Forst. Lepidium Owaihiense Cham. & Schlecht. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus Raphanistrum L. Raphanus Sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Ir. Wilcox* Ir. Wilcox* Ir. Wilcox* Ir. Wilcox* Ir. Sm. L. G. G. R. F. P. F. P. G. G. R. G. G. G. R. F. P. Hawai. Is. F. P. Pac. Is. Orient. Egypt, Arab. Ir. Eur. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Temp. Reg. Eur. N. As. | Diplotaxis efucoides DC. | G. 14. | 11. | Minor |
| *Eruca sativa Mill. Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium latifolium L, Lepidium oleraceum Forst. Lepidium Owaihiense Cham. & Schlecht. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L, Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Wilcox* Sm. L. G. R. F. P. G. R. F. P. G. R. F. P. Hawai. Is. F. P. Pac. Is. Orient. Egypt, Arab. Eur. As.Orient. Eur. As.Orient. F. P. C. L. G. Whi. L. G. L. Antisc. C. L. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. | Diplotaxis tenuifolia DC | T. | Tr | |
| Erysimum cheiranthoides L. Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium latifolium L, Lepidium oleraceum Forst. Lepidium owaihiense Cham, & Schlecht. Lepidium piscidium Forst. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L, Raphanus Raphanistrum L, Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Sim. L. G. G. G. G. F. P. Eur. Orient. Eur. As.Orient. F. P. Hawai. Is. Corient. F. P. Pac. Is. Corient. F. P. Hawai. Is. F. P. C. L. C. L. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Temp. Reg. Eur. N. As. | | | 1 | |
| Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium latifolium L. Lepidium oleraceum Forst. Lepidium Owaihiense Cham. & Schlecht. Lepidium sativum L. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus Raphanistrum L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. G. G. G. G. G. G. G. F. P. Hawai. Is. F. P. Pac. Is. Orient. Egypt, Arab. Ir. Eur.As. N. Am. Eur.As. N. Am. Eur.As. N. Am. Eur.As. N. Am. Eur. Orient. Egypt, Arab. Eur. Orient. Eur. Asia Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. | | | | |
| Erysimum crepidifolium Reichb. Isatis corniculata Lepidium Draba L. Lepidium Intifolium L. Lepidium Owaihiense Cham. & Schlecht. Lepidium piscidium Forst. Lepidium sativum L. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium orepidifolium Reichb. G. | Erysimum cheiranthoides L. | Sm. L. | Ir. Ant. | Eur. |
| Lepidium Draba L. Lepidium latifolium L. Lepidium oleraceum Forst. Lepidium Owaihiense Cham. & Schlecht. Lepidium sativum L. Lepidium sativum L. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Sophia L. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. R. G. L. G. R. G. L. G. R. F. P. Ir. G. L. G. Whi. L. G. Antisc. L. Antisc. C. L. Antisc. G. C. G. C. G. C. L. Antisc. Antisc. Cosmop. Eur. Orient. Eur. As.Orient. F. P. Hawai. Is. F. P. Pac. Is. Orient. Egypt, Arab. Lepidium officinale (L.) Scop. Eur. Asia Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. | | G | | Eur. |
| Lepidium latifolium L, Lepidium Oleraceum Forst. Lepidium Owaihiense Cham, & Schlecht. Lepidium piscidium Forst. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L, Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Alliaria Scop. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Sm. G. L. G. R. G. R. G. R. G. Whi. L. G. G. Whi. L. G. L. Antisc. Ir. Eur. As.Orient. N. Zeal. F. P. Hawai. Is, Pac. Is. Corient. Egypt, Arab. Eur.As. N. Am. Ir. Eur. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. | Isatis corniculata | | | |
| Lepidium oleraceum Forst. Lepidium Owaihiense Cham. & Schlecht. Lepidium piscidium Forst. Lepidium sativum L. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Alliaria Scop. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. L. G. R. G. R. G. R. G. Whi. L. G. Whi. L. G. L. L. Antisc. Ir. Leur. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Temp. Reg. Leur. N. As. N. As. | Lepidium Draba L. | G. R. | F. P. | Eur. Orient. |
| Lepidium Owaihiense Cham, & Schlecht, Lepidium piscidium Forst. Lepidium sativum L. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Alliaria Scop. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Lepidium Owaihiense Cham, & Schlecht, G. G. R. G. Whi. L. G. Whi. L. G. Whi. L. C. L. L. C. L. L. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Temp. Reg. Lut. N. As. | Lepidium latifolium L. | Sm. | | |
| Lepidium piscidium Forst. Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Alliaria Scop. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Sigymbrium toxophyllum G. A. Mey. Sigymbrium piscidium Forst. G. R. G. Whi. L. G. Antisc. Ir. L. C. L. L. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. L. Antisc. Sigymbrium toxophyllum G. A. Mey. G. | | | | |
| Lepidium sativum L. Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Alliaria Scop. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. Whi. L. Ir. C. L. Antisc. C. L. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. L. Antisc. L. Antisc. Siymbrium Sophia L. G. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. | Lepidium Owaihiense Cham. & Schlecht. | G | | |
| Matthiola livida DC. Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L. Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Alliaria Scop. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Sisymbrium toxophyllum G. A. Mey. G. C. L. L. Antisc. Ir. Eur. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. L. Antisc. Eur. Asia Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Eur. N. As. | | | | |
| Radicula Armoracia (L.) Robinson. Raphanus Raphanistrum L, Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Alliaria Scop. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Sisymbrium toxophyllum G. A. Mey. Antisc. Ir. L. C. L. L. Antisc. Cosmop. Cu Him. Reg. Temp. Reg. Temp. Reg. L. Antisc. N. As. | | | lr. | |
| Raphanus Raphanistrum L, Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Alliaria Scop. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. C. L. L. Antisc. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. Temp. Reg. L. Antisc. N. As. | | | + | |
| Raphanus sativus L. Senebiera pinnatifida DC. Sisymbrium Alliaria Scop. Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Raphanus sativus L. G. Antisc. Antisc. Cosmop. Eur. Orient. Him. Reg. Temp. Reg. L. Antisc. Sisymbrium toxophyllum G. A. Mey. G. | | Antisc. | | |
| Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. Him. Reg. Temp. Reg. L. Antisc. Eur. N. As. N. As. | | C. 14. | | |
| Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. Him. Reg. Temp. Reg. L. Antisc. Eur. N. As. N. As. | | č. | Millist. | |
| Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. Him. Reg. Temp. Reg. L. Antisc. Eur. N. As. N. As. | | G. | | |
| Sisymbrium Sophia L. Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. G. Antisc. Femp. Reg. Eur. N. As. N. As. | bisymbriam rimaria beop. | Ç. | | |
| Sisymbrium officinale (L.) Scop. Sisymbrium toxophyllum G. A. Mey. Vesicaria gnaphalodes Boiss. L. Antisc. Eur. N. As. N. As. Persia | Sisymbrium Sophia L. | G. | | |
| Sisymbrium toxophyllum G. A. Mey. G. N. As. Vesicaria gnaphalodes Boiss. G. Persia | | L. | Antisc. | |
| Vesicaria gnaphalodes Boiss. G. Persia | | G. | | |
| | | G. | | Persia |

CUCURBITACEAE

| , | | | |
|--------------------------------|------------|------|-----------------|
| Bryonia alba L. | Mi. C. L. | Tr. | |
| Bryonia dioica Jacq. | G.Mi. C.L. | Ir. | Eur. Med. Reg. |
| Cayaponia ficifolia Cogn. | G. | | Brazil |
| Citrullus Colycinthis Schrad. | G. C. | Pur. | Afr. Calid. |
| Corallocarpus epigaeus B. & H. | L. | Ant. | E. Ind. |
| Cucumis africanus Lindl. | L. G. | Em. | S. Afr. |
| Cucumis dipsaceus Ehrenb. | G. | | Arab, Trop.Afr. |
| Cucumis metuliferus E. Mey. | [G. |] | S. Afr. |
| Cucumis myriocarpus Naud. |] L. | Em | S. Afr. |
| Cucumis prophetarum L. | L. | Pur. | Arabia. |

^{*}A case of probable poisoning of sheep by this weed was reported to Dr. Wilcox from Nebraska.

| NAME OF PLANT | Authority | Properties | Locality |
|--|---|---|---|
| Cucumis Sacleuxii Hort. Cucumis trigonus Roxb. Cucurbita maxima Duchesne Ecballium elaterium A. Rich. Echinocystis fabacea Naud. Echinocystis macrocarpa Greenee Fevillea cordifolia L. Lagenaria vulgaris Ser. Luffa acutangula Roxb. Luffa aegyptiaca Mill. Luffa purgans Mart. Melothria scrobiculata Cogn. Momordica Balsamina L. Momordica Charantia L. Momordica cochinchinensis Spreng. | G. L. G. G. L. G. G. L. G. G. L. G. G. G. L. G. G. G. G. G. L. G. | Sap. Ant. Sap. Pur. Sap. Sap. Em. Sap. Cath. Taen. Em. Ant. Em. | Ind. Med. Reg. Calif. Calif. Ind. Trop. Reg. Trop. old world Trop. old world Tr. Am. Abyss. Trop. Reg. Trop. Reg. Trop. Reg. |
| Momordica Cymbalaria Fenzl. Sicydium monospermum Cogn. Telfairia pedata Hook. Trichosanthes Celebica Cogn. Trichosanthes globosa Bl. Trichosanthes pubera Blum. Trichosanthes trifoliata Bl. Trichosanthes wallichiana Cogn. | G. G. G. G. G. G. | Em. Taen. F. P. | Trop. Afr. Brazil Trop. Afr. Celebes. Java Ind. Malay Malay Ind. |
| CYCAD | ACEAE | | |
| Callitris sp. Cycas circinalis L. Cycas media R. Br. Macrozamia sp. Zamia media Jacq. | G. G. G. M. G. M. G. | | Austr. Moluce. Is. Austr. Austr. W. I. |
| CYPER | ACEAE | | |
| Cyperus articulatus L. Kyllingia monocephala Rottb. Schoenus apogon Roem. and Schult. | G. L. G. M. | Ant. | Cosmop. Trop. Trop. old world Austr. N. Zeal. |
| CHAILLE | TIACEAE | | |
| Chailletia cymosa Hook. Chailletia toxicaria Don. Tapura guianensis Aubl. | G. G. R. G. R. | F. P. F. P. | S. Afr. Trop. Afr. Guian. |
| DILLEN | IACEAE | | |
| Davilla rugosa Poir, Doliocarpus Rolandri J. F. Gmelin, Hibbertia glaberrima F. Muell, Hibbertia longifolia F. Muell, Tetracera alnifolia Willd, Tetracera Assa DC, | G. G. G. M. G. G. R. E. | F. P. F. P. | Brazil Guiana, Brazil. Austr. Austr. Tr. Afr. Ind. Malay |
| DIOSCOREACEAE | | | |
| Dioscorea sp. Dioscorea alata L. Dioscorea bulbifera L. Dioscorea deltoidea Wall. | G. G. G. G. | | Tr. As. Tr. As. Ind. |

| NAME OF PLANT | Authority | Properties | Locality |
|---|---|---|---|
| Dioscorea hirsuta Mart. Dioscorea macroura Harm. Dioscorea toxicaria Bojer. Dioscorea triphylla Schimp. Tamus communis L. | G. G. G. G. C. L. | Ir. Diur. | Mex. Tr. Afr. Abyss. Eur. Persia. Afr. |
| DIPSA | CEAE | | |
| Scabiosa Succisa L. | Sm. L. | | Eur. Tr. N. A. & S. A. |
| DIPTEROCA | ARPACEA | E | |
| Ancistrocladus Vahlii Arn. Dryobalanops camphora | G. Sm. L. | Del. | Zeylan. Zeylan |
| DROSER | ACEAE | | |
| Dionaea muscipula Ellis. Drosera binata Labill. Drosera communis S. Hil. Drosera intermedia Hayne. Drosera peltata Sw. Drosera rotundifolia L. Drosera Whittakerii Planch. Drosophyllum lusitanicum Link. | G. G. G. G. M. | Ir. Hyd. Hyd. Hyd. Ir. Hyd. Hyd. | N. Am. Austr. N. Zeal. Brazil Eur. Austr. N. Reg. Austr. Lusitan. Morocco. |
| EBENA | ACEAE | | |
| Diospyros acris Hemsł. Diospyros amara Diospyros Ebeneum Koen. Diospyros embryopteris Pers. Diospyros malacapai A. DC. Diospyros montana Roxb. Diospyros multiflora Blanco. Diospyros obtusifolia Willd. Diospyros samoensis A. Gr. Diospyros tesselaria Poir. Diospyros toxicaria Hiern. Diospyros virginiana L. | G. G. L. R. L. G. | Acr. Astr. F. P. Astr. F. P. Astr. F. P. Astr. F. P. Ant. Astr. | Solomon Is. Ind. Malay Tr.Afr.Philipp. Trop. As. Philipp. N. A. Samoa Is. Nav. Mascar. Is. Madagascar N. A. |
| EQUISE | TACEAE | | |
| Equisetum arvense L. Equisetum hyemale L. Equisetum palustris L. | G. Mi. L. Ches. L. Ches. | Diur. Diur. Diur. | Eur. N. A. Eur. As. N. A. Eur. |
| ERICACEAE | | | |
| Andromeda Polifolia L. Arbutus Andrachne L. Arbutus Unedo L. Arbutus varians Benth. | G. G. L. G. | Nar. Nar. | E. N. Am. Eur. As. Med. Reg. Med. Reg. Mex. |

| Arbutus xalapensis H. B. K. Arctostaphylos polifolia H. B. K. Arctostaphylos Uva-ursi L. Epigaea repens L. Gaultheria microphylla Hook. Arbutus xalapensis H. B. K. G. Diur. Nar. Mex. Sm. L. Ir. Diur. Eur. N. A. E. N. Am. Magellan R. Microphylla R. Microphylla Hook. G. Mex. Sm. L. Ches. Nar. E. N. Am. Magellan R. | eg. |
|--|-------|
| Coultbarro mumulorordos II IIon I I I I III | ava. |
| Gaultheria numularoides D. Don. Gaultheria procumbens. Kalmia angustifolia L. G. G. L. G. Rusby L. Ches. Him. Reg. J E. N. A. E. N. A. | |
| Kalmia glauca Ait. Kalmia hirsuta Walt. Kalmia latifolia L. L. Nar. L. Nar. E. N. A. E. N. A. C. Ches. L. Rusby | |
| Ledum glandulosum Nutt. Ledum latifolium Jacq. Ledum latifolium Jacq. L. Ches. Nar. N. and Arc.I N. N. A. | |
| Ledum palustre L. G. C. Nar. N. & Arc. I Rusby L. | _ |
| Leucothoe Catesbaei A. Gr. Leucothoe racemosa A. Gray. C. L. Ches. Errhine. Ches. Errhine. | ates. |
| Oxydendron arboreum DC. Whi. L. Ir. N. A. | |
| Pieris floribunda Benth. & Hook. Pieris mariana Benth. & Hook. Ches. Ineb. N. A. G. Rusby Ineb. E. N. A. L. Ches. | |
| Pieris nitida Benth. & Hook. Pieris ovalifolia D. Don. L. Ches. G. L. Ineb. E. N. A. Him. Reg. J | ava. |
| Rhododendron arborescens Torr. C. Nar. N. A. | 2 |
| Rhododendron arboreum Sm. Sm. G. Nar. Ind. Him. I Rhododendron californicum Hook. Ches. Nar. N. A. Calif. | keg. |
| Rhododendron campanulatum D. Don. G. Nar. Him. Reg. | |
| Rhododendron catawbiense Mx L. Nar. E. N. A. | |
| Rhododendron caucasicum Pall. G. Nar. Caucas. | |
| Rhododendron chrysanthum Pall. G. Sm. L. Nar. Davur. | |
| Rhododendron cinnebarinum Hook. Rhododendron dauricum L. G. Nar. Him. Reg. G. R. Nar. F. P. Eur. As. | |
| Rhododendron cinnebarinum Hook. Rhododendron dauricum L. Rhododendron ferrugineum L. Rhododendron ferrugineum L. Rhododendron ferrugineum L. G. Nar. Him. Reg. G. R. Nar. F. P. Eur. As. Eur. As. | |
| Rhododendron hirsutum L. G. C. Nar. Eur. Rhododendron indicum Sw. C. Nar. China | |
| | |
| Rhododendron ledifolium G. Don, C. Nar. China | |
| Rhododendron maximum L. G. Ches. C. Nar. E. N. A. Rusby L. | |
| Rhododendron nudiflorum Torr. C. Nar. N. A. | |
| Rhododendron occidentale A. Gray. Ches. Nar. W. N. A. | |
| Rhododendron ponticum L. G. C. Sm. Nar. A. M. | |
| Rhododendron punctatum Andr. C. N. A. | |
| Rhododendron sinense Sw. C. Nar. Japan Rhododendron sublanceolatum Miq. C. Nar. Japan | |
| Rhododendron sublanceolatum Miq. C. Nar. Japan Zenobia speciosa D. Don. G. N. A. | |
| ERYTHROXYLACEAE | |
| Erythroxylon coca Lam. Sm. L. G. Del. Stim. Peru | — |
| Anes. | |
| EUPHORBIACEAE | |
| Acalypha colchica Fisch. & Mey. Acalypha cordifolia Muell. G. Hyd. Caucas. Him. Reg. | |
| Acalypha indica L. G. Em. Tr. As. & A | fr. |
| Acalypha virginica L. L. Exp.Diur.Ir N. Am. | |

| Ealiospermum axillare Bl. Beyeria viscosa Miq. Bridelia retusa Spreng. Chrozophora pilecta A. Juss. Cleistanthus collinus Benth. & Hook. Cnesmone javanica Bl. Croton muricatus Zipp. Croton Tiglium L. Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloecocca verrucosa Eremocarpus setigerus Benth. Euphorbia alsinaeilora Baill. Euphorbia anitquorum L. Euphorbia andiquorum L. Euphorbia caracasana Euphorbia cerebrum L. Euphorbia cerebrum L. Euphorbia cerebrum L. Euphorbia cotinides Miq. Euphorbia betado Cyparissias L. Euphorbia betado Cyparissias L. Euphorbia heptagona L. Euphorbia linearis Retz. Euphorbia phyberna L, | (| | | |
|--|------------------------------|------------|------------|-------------|
| Beyeria viscosa Miq. Bridelia retusa Spreng. Chrozophora plicata A. Juss. Chrozophora tinctoria A. Juss. Cleistanthus collinus Benth. & Hook. Cnesmone javanica Bl. Croton muricatus Zipp. Croton Tiglium L. Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloeococca verrucosa Eremocarpus setigerus Benth. Euphorbia aleppica L. Euphorbia aleppica L. Euphorbia annygdaloides I. Euphorbia annyddaloides I. Euphorbia candelabrum Trem. Euphorbia candelabrum Trem. Euphorbia cerebrum L. Euphorbia comioides Miq. Euphorbia Comioides L. Euphorbia hedroides L. Euphorbia hedroides L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia heterodoxa Muell. Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia plaustris L. Euphorbia pithyusa C. Euphorbia pithyusa C. Euphorbia punicea Sw. G. G. G. C. S.m.R. Pur. F. P. Calif. F. P. Ches. L. C. R. C. R. Ir. Ches. L. C. R. Ir. C. C. R. Ir. C. C. G. R. Ir. C. R. Ir. C. C. Ir. G. R. Ir. F. P. G. Ir. Ches. Ir. G. Ir. G. R. Ir. F. P. Hudustr. Austr. Carlificar. Ind. Malay Austr. Java Carlificar. Ind. Malay Austr. Java Carlificar. Ind. Ma | NAME OF PLANT | Authority | Properties | Locality |
| Bridelia retusa Spreng. Chrozophora plicata A. Juss. Chesophora inctoria A. Juss. Cleistanthus collinus Benth. & Hook. Cnesmone javanica Bl. Croton Tiglium L. Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloeococca verrucosa Eremocarpus setigerus Benth. Euphorbia alsinaeflora Baill. Euphorbia aniquorum L. Euphorbia aniquorum L. Euphorbia anivorea Euphorbia bicolor Engem & Gray. Euphorbia candelabrum Trem. Euphorbia caracasana Euphorbia characias L. Euphorbia cotinioides Miq. Euphorbia cotinioides Miq. Euphorbia Cyparissias L. Euphorbia Drummondii Boiss. Euphorbia betordoxa Muell. Euphorbia helioscopia L. Euphorbia marginata Pursh. Euphorbia pincularia Euphorbia paralias L. Euphorbia pincularia L. Eu | Baliospermum axillare Bl. | G. | | Ind. Malay |
| Chrozophora plicata A. Juss. Chrozophora lincatoria A. Juss. Cleistanthus collinus Benth. & Hook. Cnesmone javanica Bl. Croton muricatus Zipp. Croton Tiglium L. Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloecocca verrucosa Eremocarpus setigerus Benth. Euphorbia aleppica L. Euphorbia anygdaloides L. Euphorbia anygdaloides L. Euphorbia i antorea Euphorbia bicolor Engem & Gray. Euphorbia candelabrum Trem. Euphorbia chanalsyce L. Euphorbia chanalsyce L. Euphorbia corollata L. Euphorbia Cyparissias L. Euphorbia Cyparissias L. Euphorbia Drummondii Boiss. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia ingularia Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia palustris L. Euphorbia palustris L. Euphorbia pinpuase L. Euphorbia palustris L. Euphorbia piphysus L. Euphorbia piphorbia piphysus L. Euphorbia pip | Beyeria viscosa Miq. | | | |
| Chrozophora plicata A. Juss. Chrozophora lincatoria A. Juss. Cleistanthus collinus Benth. & Hook. Cnesmone javanica Bl. Croton muricatus Zipp. Croton Tiglium L. Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloecocca verrucosa Eremocarpus setigerus Benth. Euphorbia aleppica L. Euphorbia anygdaloides L. Euphorbia anygdaloides L. Euphorbia i antorea Euphorbia bicolor Engem & Gray. Euphorbia candelabrum Trem. Euphorbia chanalsyce L. Euphorbia chanalsyce L. Euphorbia corollata L. Euphorbia Cyparissias L. Euphorbia Cyparissias L. Euphorbia Drummondii Boiss. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia ingularia Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia palustris L. Euphorbia palustris L. Euphorbia pinpuase L. Euphorbia palustris L. Euphorbia piphysus L. Euphorbia piphorbia piphysus L. Euphorbia pip | Bridelia retusa Spreng. | G. | | Ind. Malay |
| Cleistanthus collinus Benth. & Hook. Cnesmone javanica Bl. Croton muricatus Zipp. Croton Tiglium L. Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloecocca verrucosa Eremocarpus setigerus Benth. Euphorbia aleppica L. Euphorbia aleppica L. Euphorbia anygdaloides I. Euphorbia anygdaloides I. Euphorbia i arborea Euphorbia bicolor Engem & Gray. Euphorbia bicolor Engem & Gray. Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia caracias L. Euphorbia cotinifolia L. Euphorbia dendroides I. Euphorbia cotinifolia L. Euphorbia ceremophila A. Cunn. Euphorbia prummondii Boiss. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia linearis Retz. Euphorbia ingularia Euphorbia palustris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia piphyusa C. G. E. Ir. F. P. Med. Reg. Ir. F. P. Mader. Is. Mader. Is. Med. Reg. Ir. F. P. Mader. Is. Med. Reg. Ir. F. P. Med. Reg. Ir. F. P. Eur. N. Am. Madera Is. Mi. C. Sm. Ir. G. E. R. Ir. F. P. Mader. Is. Mi. C. Sm. Ir. G. E. R. Ir. F. P. Med. Reg. Ir. F. P. Med | Chrozophora plicata A. Juss. | | | |
| Cresmone javanica Bl. Croton muricatus Zipp. Croton Tiglium L. Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloeococca verrucosa Eremocarpus setigerus Benth. Euphorbia alepica L. Euphorbia amygdaloides L. Euphorbia antiquorum L. Euphorbia antiquorum L. Euphorbia bicolor Engem & Gray. Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia corollata L. Euphorbia cotinioides Miq. Euphorbia cotinioides Miq. Euphorbia Corollata L. Euphorbia Drummondii Boiss. Euphorbia Heloscopia L. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia ingularia Euphorbia ingularia Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia palustris L. Euphorbia palustris L. Euphorbia pidyusa L. Euphorbia preslii Guss. Euphorbia preslii Guss. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Euphorbia punicea Sw. G. C. C. L. A. P. F. P. Calif. F | | | | |
| Croton muricatus Zipp, Croton Tiglium L. Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloeococa verrucosa Eremocarpus setigerus Benth. Euphorbia aleppica L. Euphorbia aleppica L. Euphorbia amygdaloides L. Euphorbia antiquorum L. Euphorbia aintiquorum L. Euphorbia candelabrum Trem. Euphorbia candelabrum Trem. Euphorbia canacias L. Euphorbia chamalsyce L. Euphorbia corollata L. Euphorbia cotinifolia L. Euphorbia dendroides Miq. Euphorbia cotinifolia L. Euphorbia dendroides L. Euphorbia dendroides L. Euphorbia dendroides L. Euphorbia dendroides L. Euphorbia eremophila A. Cunn. Euphorbia eremophila A. Cunn. Euphorbia heptagona L. Euphorbia inearis Retz. Euphorbia ingularia Euphorbia mellifera Ait. Euphorbia mellifera Ait. Euphorbia palustris L. Euphorbia palustris L. Euphorbia pisatoria Ait. Euphorbia pisatoria Ait. Euphorbia pisatoria Ait. Euphorbia preslii Guss Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia punicea Sw. G. C. S. R. G. R. G. R. G. R. G. Ir. G. R. G. Ir. F. P. G. Ir. G. R. G. F. Ir. F. P. | | | | |
| Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloeococca verrucosa Eremocarpus setigerus Benth. Euphorbia aleppica L. Euphorbia alismaeflora Baill. Euphorbia alismaeflora Baill. Euphorbia antiquorum L. Euphorbia arborea Euphorbia candelabrum Trem. Euphorbia caracasana Euphorbia caracasana Euphorbia corollata L. Euphorbia cotinioides Miq. Euphorbia Cotinioides Miq. Euphorbia Cotinioides L. Euphorbia Drummondii Boiss. Euphorbia Gerardiana Jacq. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia helerodoxa Muell. Euphorbia heptagona L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia palustris L. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia punicea Sw. G. S., R. C. L. A. P. F. P. Calif. A. P. F. P. Calif. Ir. Ches. Ir. F. P. G. Ir. Ches. Ir. C. R. Ir. F. P. G. Ir. G. Ir. G. Ir. G. M. Ir. C. R. G. R. Ir. F. P. G. Ir. G. Ir. G. Ir. G. M. Ir. E. P. G. M. Ir. G. R. Ir. F. P. G. Ir. G. Ir. G. Ir. G. M. Ir. E. P. G. M. Ir. Ind. Malay Austr. Java A. P. F. P. Calif. Ir. Austr. Eur. Orient. Ind. N. Am. Ind. Malay Austr. Java Calif. Ir. F. P. Calif. Ir. F. P. Calif. Ir. F. P. Calif. Ir. F. P. Cull. Ind. Malay Austr. Austr. Eur. Ind. N. Am. Ir. Ches. Ir. G. Ir. G. Ir. F. P. G. Ir. G. Ir. G. Ir. F. P. G. Ir. F. P. G. Ir. F. P. G. Ir. G. Ir. F. P. G. Ir. F. P. G. Ir. G. Ir. F. P. G. Ir. G. Ir. F. P. Ind. Med. Reg. Ir. | | | ļ | 1 |
| Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloeococa verrucosa Eremocarpus setigerus Benth. Euphorbia aleppica L. Euphorbia aleppica L. Euphorbia aningdaloides L. Euphorbia ariduorum L. Euphorbia ariduorum L. Euphorbia bicolor Engem & Gray. Euphorbia caracasana Euphorbia caracasana Euphorbia caracias L. Euphorbia cotinifolia L. Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia Drummondii Boiss. Euphorbia Esula L. Euphorbia dendroides L. Euphorbia beterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia palustris L. Euphorbia palustris L. Euphorbia pistyusa L. Euphorbia pistyusa L. Euphorbia pistyusa L. Euphorbia prellus L. Euphorbia pistyusa L. Euphorbia prellus L. Euphorbia pistyptyllos L. Euphorbia pityusa L. Euphorbia punicea Sw. L. C. R. Ir. F. P. Ches. Ir. Ches. Ir. Ches. Ir. Ches. Ir. G. Ir. G. Ir. G. Ir. F. P. Ir. Med. Reg. Ir. F. P. Eur. N. Am. Ir. Austr. Eur. Austr. Eur. N. Am. Ir. R. Mi. C. Ir. Eur. N. Am. Ir. Austr. Eur. N. Am. Ir. R. Mi. C. Ir. Eur. N. Am. Ir. Austr. Eur. N. Am. Ir. R. Mi. C. Ir. G. Ir. F. P. Eur. N. Am. Ir. N. Am. Ir. N. Am. Ir. N. Am. Ir. Austr. Ir. Ches. Ir. Ir. Ches. Ir. Ir. Austr. Ir. Ches. Ir. Ir. Austr. Ir. Austr. Ir. R. Mi. C. Ir. G. Ir. F. P. Eur. N. Am. Ir. N. Am. Ir. N. Am. Ir. N. Am. Ir. Ches. Ir. G. Ir. | | G. | | |
| Croton Verreauxii Baill. Daphniphyllum bancanum Kurz. Eloeococca verrucosa Eremocarpus setigerus Benth. Euphorbia alepica L. Euphorbia alsinaeflora Baill. Euphorbia antiquorum L. Euphorbia bicolor Engem & Gray. Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia carocollata L. Euphorbia cotinioflos Miq. Euphorbia cotinioflos Miq. Euphorbia cotinioflos I. Euphorbia cotinioflos I. Euphorbia prummondii Boiss. Euphorbia prummondii Boiss. Euphorbia beterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia heptagona L. Euphorbia linearis Retz. Euphorbia palustris L. Euphorbia pithyusa L. Euphorbia preslii Guss. Euphorbia preslii Guss. Euphorbia punicea Sw. C. L. A. P. F. P. Austr. A. P. F. P. Austr. Califi. F. P. G. R. Ir. F. P. G. Ir. G. Ir. G. R. Ir. F. P. G. Ir. G. Ir. G. R. Ir. F. P. G. Ir. G. Ir. G. R. Ir. F. P. Guiana G. R. Ir. F. P. Guiana Ind. N. Am. Mi. C. Ir. Mi. C. Ir. Mi. C. Ir. G. R. Ir. F. P. Med. Reg. Austr. Austr. Austr. Eur. N. Am. Mi. C. Ir. Mi. C. Ir. Mi. C. Ir. G. R. Ir. F. P. Med. Reg. Ir. F. P. Min. C. Ir. Ind. Maustr. Austr. Ches. L. Ir. Ir. Ches. L. Ir. Ir. C. C. C. R. Ir. F. P. Min. C. Ir. Ir. N. Am. Min. C. Ir. Ir. Ir. C. R. Ir. F. P. Ind. Austr. Austr. C. C. Ir. Ir. C. | Croton Tighum L. | | Pur. F. P. | |
| Daphniphyllum bancanum Kurz. Eloeococca verrucosa Eremocarpus setigerus Benth. Euphorbia aleppica L. Euphorbia alsinaeflora Baill. Euphorbia amygdaloides L. Euphorbia antiquorum L. Euphorbia antiquorum L. Euphorbia bicolor Engem & Gray. Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia cotinifolia L. Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia Esula L. Euphorbia Eremophila A. Cunn. Euphorbia Euphorbia helioscopia L. Euphorbia phataris L. Euphorbia phataris L. Euphorbia phataris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia platyphyllos L. Euphorbia platyphyllos L. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia punicea Sw. | O . 17 " D " | L. | } | |
| Eloecocca verrucosa Eremocarpus setigerus Benth. Euphorbia aleppica L. Euphorbia alsinaeflora Baill. Euphorbia antiquorum L. Euphorbia arborea Euphorbia bicolor Engem & Gray. Euphorbia canacasana Euphorbia caracasana Euphorbia cerebrum L. Euphorbia caracasana Euphorbia cotolioides Miq. Euphorbia cotinioida L. Euphorbia cotinioida L. Euphorbia cotinioida L. Euphorbia eremophila A. Cunn. Euphorbia eremophila A. Cunn. Euphorbia eremophila A. Cunn. Euphorbia bicolor Engem & Gray. Euphorbia cotinioida L. Euphorbia cotinioida L. Euphorbia cotinioida L. Euphorbia eremophila A. Cunn. Euphorbia eremophila A. Cunn. Euphorbia besula L. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia ingularia Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia plaustris L. Euphorbia pisactoria Ait. | | G. | | _ |
| Eremocarpus setigerus Benth. Euphorbia alepjaca L. Euphorbia alsinaeflora Baill. Euphorbia ansugdaloides L. Euphorbia antiquorum L. Euphorbia arborea Euphorbia bicolor Engem & Gray. Euphorbia candelabrum Trem. Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia caracias L. Euphorbia cotinoides Miq. Euphorbia cotinoides Miq. Euphorbia cotinoides Miq. Euphorbia Cyparissias L. Euphorbia Drummondii Boiss. Euphorbia Gerardiana Jacq. Euphorbia Gerardiana Jacq. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia inpeacuanhae L. Euphorbia inpeacuanhae L. Euphorbia marginata Pursh. Euphorbia palustris L. Euphorbia priscaloria Ait. Euphorbia punicea Sw. | | | } | Java |
| Euphorbia aleppica L. Euphorbia alsinaeflora Baill. Euphorbia antiquorum L. Euphorbia antiquorum L. Euphorbia arborea Euphorbia caracasana Euphorbia cerebrum L. Euphorbia corliata L. Euphorbia cotinifolia L. Euphorbia dendroides Miq. Euphorbia dendroides L, Euphorbia dendroides L, Euphorbia premophila A. Cunn. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia heptagona L. Euphorbia heptagona L. Euphorbia heptagona L. Euphorbia heptagona L. Euphorbia ingularia Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia platyphyllos L. Euphorbia platyphyllos L. Euphorbia platyphyllos L. Euphorbia punicea Sw. Euphorbia punicea Sw. Euphorbia punicea Sw. R. G. R. Ir. F. P. G. M. Ir. G. M. Ir. G. R. Ir. F. P. Eur. Orient. Ind. Austr. Ir. F. P. Eur. Orient. Ind. H. F. P. Eur. Orient. Ind. H. G. R. Ir. F. P. Eur. N. Am. Ir. C. R. Ir. F. P. G. Ir. G. Ir. Med. Reg. Ir. F. P. Eur. Aaustr. Ir. R. M. C. R. G. R. Ir. F. P. Eur. Orient. Ind. H. Ir. Ind. H. Ind. H. Ir. Ind. H. Ind. H. Ir. Ind. H. Ind. H. Ir. Ind. H. Ind. H. Ir. Ind. H. Ind. H. Ir. | | | A D E D | C 115 |
| Euphorbia alsinaeflora Baill. Euphorbia antiquorum L. Euphorbia antiquorum L. Euphorbia arborea Euphorbia candelabrum Trem. Euphorbia caracasana Euphorbia cerebrum L. Euphorbia chamalsyce L. Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia Drunmondii Boiss. Euphorbia eremophila A. Cunn. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia helesdoroja L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia malifera Ait. Euphorbia palustris L. Euphorbia preslii Guss. Euphorbia punicea Sw. G. R. Ir. F. P. G. Ir. Ir. Ches. Ir. Ches. Ir. G. Ir. Med. Reg. Ir. F. P. Eur. N. Am. Ir. F. P. G. Ir. Med. Reg. Ir. F. P. Eur. N. Am. C. R. C. E. Ir. F. P. Eur. N. Am. N. Am. Eur. N. Am. Eur. N. Am. Eur. N. Am. C. Ches. Ir. G. Ir. G. Ir. Med. Reg. Ir. F. P. Ind. Maustr. Ir. N. Am. Ir. Ches. L. Ir. Co. Ir. Mi. C. Ir. Eur. N. Acr. G. Ir. G. Ir. Eur. N. Acr. | | | | Calif. |
| Euphorbia antiquorum L. Euphorbia bicolor Engem & Gray. Euphorbia caracasana Euphorbia cerebrum Trem. Euphorbia cerebrum L. Euphorbia cotinides Miq. Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia Cyparissias L. Euphorbia Esula L. Euphorbia Esula L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia heptagona L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia mellifera Ait. Euphorbia palustris L. Euphorbia Paralias L. Euphorbia pistyusa L. Euphorbia pistyusa L. Euphorbia pistyusa L. Euphorbia pistyusa L. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia pirmulaefolia Baker Euphorbia pinulcea Sw. G. R. Ir. F. P. G. Ir. Ches. Ir. C. R. Ir. F. P. G. Ir. C. R. Ir. C. C. Ir. Med. Reg. N. A. N. Am. Ir. Eur. N. Am. Ir. Austr. Austr. Austr. Austr. Ir. Austr. Austr. Austr. Ir. Austr. Austr. Austr. Austr. Ir. Austr. A. Ac. C. R. Ir. C. A. A. C. R. Ir. A. Austr. Aur. Austr. | | | | Aucta |
| Euphorbia antiquorum L. Euphorbia arborea Euphorbia bicolor Engem & Gray. Euphorbia caracasana Euphorbia caracasana Euphorbia cerebrum L. Euphorbia chamalsyce L. Euphorbia cotinioides Miq. Euphorbia cotinioides Miq. Euphorbia Cyparissias L. Euphorbia Drunmondii Boiss. Euphorbia Drunmondii Boiss. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia malifera Ait. Euphorbia palustris L. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia preslii Guss. Euphorbia pulcherrima Willd. Euphorbia punicea Sw. | | C D | | |
| Euphorbia arborea Euphorbia bicolor Engem & Gray. Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia caracasana Euphorbia characias L. Euphorbia cotoides Miq. Euphorbia cotinifolia L. Euphorbia dendroides L, Euphorbia dendroides L, Euphorbia dendroides L, Euphorbia Gerardiana Jacq. Euphorbia Gerardiana Jacq. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia palustris L. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Euphorbia punicea Sw. Ir. Ches. Ir. Ches. Ir. F. P. G. Ir. F. P. G. Ir. F. P. G. R. Ir. F. P. G. R. Ir. F. P. G. R. Ir. F. P. Med. Reg. N. A. Co. Ir. F. P. Med. Reg. Ir. F. P. Med. Reg. Ir. F. P. Eur. N. Am. L. Eur. Ir. F. P. Eur. Austr. Austr. Austr. Austr. Ir. Eur. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. I | | 1 _ | | |
| Euphorbia bicolor Engem & Gray. Euphorbia candelabrum Trem. Euphorbia caracasana Euphorbia cerebrum L. Euphorbia chamalsyce L. Euphorbia corollata L. Euphorbia cotinides Miq. Euphorbia cotinides Miq. Euphorbia cotinides Miq. Euphorbia cotinides Miq. Euphorbia cotinides L. Euphorbia Cyparissias L. Euphorbia Drunmondii Boiss. Euphorbia Drunmondii Boiss. Euphorbia Gerardiana Jacq. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia helpscopia L. Euphorbia helpscacuanhae L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia palustris L. Euphorbia palustris L. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Ches. L. Ir. G. R. Ir. F. P. Med. Reg. N. A. G. R. Ir. F. P. Med. Reg. Ir. Austr. Austr. Austr. Austr. Austr. Austr. Austr. R. Mi. C. Ir. G. R. Ir. F. P. Eur. N. Am. Med. Reg. N. A. C. R. Ir. F. P. Eur. Austr. Austr. Austr. Austr. Austr. Austr. Austr. Austr. Austr. R. Mi. C. Ir. E. C. R. Ir. F. P. Eur. Austr. Aust | | G. | | 111d. |
| Euphorbia caracasana Euphorbia ceracasana Euphorbia ceracasana Euphorbia chamalsyce L. Euphorbia characias L. Euphorbia cotinioles Miq. Euphorbia cotinioles Miq. Euphorbia cotinioles Miq. Euphorbia Cyparissias L. Euphorbia Cyparissias L. Euphorbia Drummondii Boiss. Euphorbia prummondii Boiss. Euphorbia eremophila A. Cunn. Euphorbia eremophila A. Cunn. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia heptagona L. Euphorbia heptagona L. Euphorbia lipeacauanhae L. Euphorbia Lathyris L. Euphorbia Lathyris L. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia palustris L. Euphorbia palustris L. Euphorbia Peplus L. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia punicea Sw. Euphorbia punicea Sw. Euphorbia punicea Sw. Euphorbia punicea Sw. Ir. G. Ir. G. Ir. Ir. F. P. Med. Reg. Ir. F. P. Guiana Ir. Ir. F. P. Guiana Ir. Ir. F. P. Med. Reg. Ir. F. P. Eur. N. Am. G. R. Ir. F. P. Eur. N. Am. G. R. Ir. F. P. Eur. N. Am. C. R. Ir. F. P. Eur. N. Am. C. R. Ir. F. P. Eur. N. Am. G. R. Ir. F. P. Eur. N. Am. C. R. Ir. F. P. Ind. Austr. Euphorbia pindulaefolia Baker Euphorbia primulaefolia Baker Euphorbia punicea Sw. Ir. F. P. Ind. G. R. Ir. F. P. Ind. G. R. Ir. F. P. Ind. C. R. Ir. F. P. Eur. N. Am. Ir. C. R. Ir. F. P. Ind. C. R. Ir. F. P. Ind. C. R. Ir. F. P. Ind. C. R. Ir. F. P. Eur. N. Am. Ir. C. R. Ir. F. P. Ind. C. R. Ir | | Ches | 1 | N Am |
| Euphorbia caracasana Euphorbia cerebrum L. Euphorbia chamalsyce L. Euphorbia chamalsyce L. Euphorbia coriolata L. Euphorbia cotiniodes Miq. Euphorbia cotiniodes Miq. Euphorbia Cyparissias L. Euphorbia Cyparissias L. Euphorbia dendroides L, Euphorbia dendroides L, Euphorbia eremophila A. Cunn. Euphorbia eremophila A. Cunn. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia heptagona L. Euphorbia heptagona L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia neriifolia L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicae Sw. R. G. Ir. Hed. Reg. Ir. F. P. Guiana Ir. F. P. Guiana Ir. F. P. Guiana Ir. F. P. Had. Reg. Ir. F. P. Had. Reg. Ir. F. P. Eur. N. A. Eur. Eur. N. A. Eur. N. A. Eur. Eur. N. A. Eur. Eur. N. A. Eur. Eur. N. A. Eur. Eur. Eur. N. A. Eur. Eur. Eur. N. A. Eur. Eur. Eur. Eur. N. A. Eur. Eur. Eur. Eur. Eur. Eur. N. A. Eur. Eur. Eur. Eur. Eur. Eur. Eur. Eur | | Circs. | | 114. 23111. |
| Euphorbia cerebrum L. Euphorbia chamalsyce L. Euphorbia characias L. Euphorbia corollata L. Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia dendroides L, Euphorbia dendroides L, Euphorbia eremophila A. Cunn. Euphorbia eremophila A. Cunn. Euphorbia eremophila A. Cunn. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia palustris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia primulaefolia Baker Euphorbia puncherrima Willd. | | RG | | |
| Euphorbia chamalsyce L. Euphorbia characias L. Euphorbia cotinioides Miq. Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia dendroides L, Euphorbia Drummondii Boiss. Euphorbia eremophila A. Cunn. Euphorbia eremophila A. Cunn. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia heterodoxa Muell. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia palustris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia piscatoria Ait. Euphorbia primulaefolia Baker Euphorbia plucherrima Willd. Euphorbia pulnerima Willd. Euphorbia plucherrima Willd. Euphorbia plucherrima Willd. Euphorbia pulneca Sw. G. G | | 1 _ | } = | |
| Euphorbia characias L. Euphorbia corollata L. Euphorbia cotinides Miq. Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia Cyparissias L. Euphorbia Drummondii Boiss. Euphorbia eremophila A. Cunn. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia heptagona L. Euphorbia heptagona L. Euphorbia lipecacuanhae L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia plaustris L. Eupho | | | 1 | |
| Euphorbia cotonides Miq. Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia dendroides L, Euphorbia dendroides L, Euphorbia Drummondii Boiss. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia heptagona L. Euphorbia heptagona L. Euphorbia lipecacuanhae L. Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia meriifolia L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia platyphyllos L. Euphorbia primulaefolia Baker Euphorbia plucherrima Willd. Euphorbia puncea Sw. L. G. R. Ir. F. P. Guiana Ir. F. P. Eur. N. Am. Ir. Austr. A. Austr. Austr. A. Austr. Austr. A. Austr. Austr. A. A. A. Ir. F. P. Eur. Acr. G. R. Ir. F. P. Eur. N. Am. A. C. R. Ir. F. P. Eur. N. Am. A. C. R. Ir. F. P. Ind. Malay Ir. A. Austr. A. Ir. G. R. Ir. F. P. Eur. N. Am. Austr. | | | 1 | Med. Reg. |
| Euphorbia cotinidels Miq. Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia dendroides L, Euphorbia Drunmondii Boiss. Euphorbia Drunmondii Boiss. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia heterodoxa Muell. Euphorbia heterodoxa Muell. Euphorbia Inecacuanhae L. Euphorbia linearis Retz. Euphorbia mellifera Ait. Euphorbia palustris L. Euphorbia Paralias L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia punicea Sw. Euphorbia punicea Sw. | | | | |
| Euphorbia cotinifolia L. Euphorbia Cyparissias L. Euphorbia Drummondii Boiss. Euphorbia eremophila A. Cunn. Euphorbia Esula L. Euphorbia Esula L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia heterodoxa Muell. Euphorbia Ipecacuanhae L. Euphorbia linearis Retz. Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia palustris L. Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Euphorbia punicea Sw. G. E. R. Mi. C. Mi. C. R. Mi. C. Mi. | | G. R. | | |
| Euphorbia Cyparissias L. Euphorbia dendroides L. Euphorbia Drummondii Boiss. Euphorbia eremophila A. Cunn. Euphorbia Esula L. Euphorbia Esula L. Euphorbia helioscopia L. Euphorbia heterodoxa Muell. Euphorbia lipecacuanhae L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia parlustris L. Euphorbia Paralias L. Euphorbia Paralias L. Euphorbia platyphyllos L. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Euphorbia punicea Sw. Mi. C. G. R. Ir. F. P. Eur. N. Am. Med. Reg. Austr. Ir. F. P. Eur. Mi. C. G. R. Ir. F. P. Eur. M. Austr. Acustr. Ir. F. P. Eur. Maustr. Austr. Austr. Austr. Austr. Austr. Austr. Austr. E. Ir. F. P. Eur. Austr. A | | | | Ind. |
| Euphorbia dendroides L. Euphorbia Drummondii Boiss. Euphorbia eremophila A. Cunn. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia helioscopia L. Euphorbia heptagona L. Euphorbia heterodoxa Muell. Euphorbia lipecacuanhae L. Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia palustris L. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Euphorbia punicea Sw. | | Mi. C. | | Eur. N. Am. |
| Euphorbia eremophila A. Cunn. Euphorbia Esula L. Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia heptagona L. Euphorbia heptagona L. Euphorbia hyberna L. Euphorbia Ipecacuanhae L. Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Euphorbia punicea Sw. Mi. C. | | [G, R. | Ir. F. P. | Med. Reg. |
| Euphorbia Esula L. Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia heptagona L. Euphorbia heterodoxa Muell. Euphorbia lipecacuanhae L. Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia pulcherrima Willd. Euphorbia punicea Sw. ER Mi. C. Ir. Mi. C. Ir. C. R. Ir. F. P. Eur. Ir. Eur. R. Mi. C. Ir. Eur. Ir. Eur. Acr. Brazil Eur. Eur. R. Mi. C. Ir. Eur. Acr. Brazil Eur. F. P. Eur. N. A. Eur. Brazil Eur. Brazil Eur. Brazil Eur. Brazil G. R. Ir. C. R. Ir. C. R. Ir. C. R. Ir. G. Ir. M. Am. C. Bur. Bur. Bur. Bur. Bur. Bur. Bur. Bur | |] G. M. | Ir. | Austr. |
| Euphorbia Gerardiana Jacq. Euphorbia helioscopia L. Euphorbia heptagona L. Euphorbia heterodoxa Muell. Euphorbia lipecacuanhae L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia palustris L. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Mi. C. Mi | | | | |
| Euphorbia helioscopia L. Euphorbia heterodoxa Muell. Euphorbia hyberna L. Euphorbia lipecacuanhae L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Mi. C. G. G. Ir. G. R. Ir. G. R. Ir. G. Ir. W. I. G. Ir. W. I. G. Ir. Madeira Is. Ir. N. Afr. Eur. N. As. G. E. Ir. F. P. Mader. Is. Med. Reg. Eur. Afr. Smi. Ir. G. Eur. N. As. Ir. Eur. Med. Reg. Eur. N. As. Ir. G. Eur. N. Afr. Eur. N. As. G. Eur. N. As. Ir. G. Eur. N. Afr. Eur. N. Afr. Eur. N. Afr. Eur. N. Afr. Eur. N. Am. As. G. Eur. N. As. Ir. Eur. N. Am. Afr. Eur. N. Am. As. G. Ir. G. Eur. N. Afr. Eur. N. Afr. Eur. Eur. N. Afr. Eur. N. Afr. Eur. N. Afr. Eur. N. Am. As. G. Eur. N. As. Ir. Eur. N. Am. Afr. Eur. N. Am. Madagas. Eur. F. P. Madagas. Eur. F. P. Madagas. Eur. F. P. Jamaica | | | 1 * | |
| Euphorbia heptagona L. Euphorbia heterodoxa Muell. Euphorbia hyberna L, Euphorbia Ipecacuanhae L. Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia Peplus L. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia palutyphyllos L. Euphorbia primulaefolia Baker Euphorbia punicea Sw. Euphorbia punicea Sw. | | | 1 | |
| Euphorbia heterodoxa Muell. Euphorbia hyberna L. Euphorbia Ipecacuanhae L. Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia palusphyllos L. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. | | | | |
| Euphorbia hyberna L, Euphorbia Ipecacuanhae L. Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. Euphorbia IIr. C. R. M. Rusby Ir. M. Rusby Ir. F. P. Eur. N. Am. C. R. Ir. Madeira Is. Ir. Madeira Is. Ir. Madeira Is. Ir. Eur. N. Afr. Eur. N. Afr. Eur. N. As. Ir. Eur. Med. Reg. Eur. Afr. N. Am. Ir. C. R. Ir. C. R. Ir. Madeira Is. Ir. Eur. Med. Reg. Eur. Mi. C. E. Ir. C. Mi. Ir. C. Mi. C. Eur. Madagas. Eur. Afr. Madagas. Eur. Mex. L. Eur. Madagas. Eur. Mex. L. Eur. Madagas. Eur. Mex. L. Eur. Madagas. Eur. Eur. Mex. L. Eur. Madagas. Eur. Mex. L. Eur. Madagas. Eur. Mex. Eur. Mex. L. Eur. Madagas. Eur. Mex. L. Eur. Madagas. Eur. Mex. Eur. Mex. L. Eur. Mex. Eur. Mex. Eur. Mex. Eur. Mex. Eur. Mex. Eur. Mex. Eur. Eur. Mex. Eur. Eur. Mex. Eur. Eur. Mex. Eur. Eur. Eur. Madagas. Eur. Eur. Eur. Eur. Eur. Eur. Eur. Eur | | | 1 . | |
| Euphorbia Ipecacuanhae L. Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia punicea Sw. M. Rusby Ir. C. R. Ir. F. P. Eur. N. Am. G. Ches. Ir. G. R. Ir. F. P. Ind. Malay N. A. Ir. W. I. Eur. N. Am. Ir. Eur. N. Afr. Eur. N. As. Ir. Eur. N. As. Ir. Eur. N. As. Ir. Eur. F. P. Madeira Is. Ir. Eur. Med. Reg. Ir. Eur. N. As. Ir. Eur. N. As. Ir. Eur. N. As. Ir. Eur. N. As. Ir. Eur. N. Afr. Eur. N. As. Ir. Eur. Afr. Eur. Afr. Eur. Afr. Eur. Afr. Eur. Afr. Eur. Afr. Ir. Ir. Ir. Ir. Ir. Ir. Ir. | | | | |
| Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia pulcherrima Willd. Euphorbia punicea Sw. L. C. R. Ir. F. P. Eur. N. Am. Madeira Is. Ir. N. Afr. Eur. N. Afr. Ir. Med. Reg. Eur. Afr. Smi. Ir. Med. Reg. Eur. Afr. Smi. Ir. N. Am. Med. Reg. Ir. F. P. Madagas. Ir. Mex. Jamaica | | | | |
| Euphorbia Lathyris L. Euphorbia linearis Retz. Euphorbia linearis Retz. Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia Peplus L. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. C. R. Ches. L. Ir. G. Ir. G. Ir. N. Am. Ir. Madeira Is. Ir. F. P. Ind. Malay Ir. Ir. Med. Reg. Eur. Ir. Mi. C. Sm. Ir. G. Eur. Ir. Mi. Ir. Med. Reg. Eur. Afr. Smi. Ir. Mi. Ir. Med. Reg. Eur. Afr. Smi. Ir. Med. Reg. Ir. F. P. Mader. Is. Mi. Ir. Med. Reg. Ir. Ir. Med. Reg. Ir. Med. Reg. Ir. Ir. Med. Reg. Ir. Ir. Med. Reg. Ir. Ir. Med. Reg. Ir. Ir. Ir. Ir. Med. Reg. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir | Euphornia Tpecacuannae L. | | III. | IN. A. |
| Euphorbia linearis Retz. Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia meriifolia L. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia Peplus L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia palutyphyllos L. Euphorbia pithyusa L. Euphorbia primulaefolia Baker Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. Ches. L. Ir. G. Ir. G. Ir. N. Am. Mi. Ir. Med. Reg. Eur. Eur. Mi. Ir. Med. Reg. Eur. Fur. Afr. Smi. Ir. Madagas. Eur. Fur. Madagas. Eur. Eur. Mi. Ir. Med. Reg. Eur. Eur. Mi. Ir. Med. Reg. Eur. Fur. Afr. Smi. Ir. Madagas. Eur. Eur. Madagas. Eur. Eur. Mex. Eur. Mex. Eur. Eur. Mex. Eur. Mex. Eur. | Euphorbia Latheris I | | Ir FP | Fur N Am |
| Euphorbia linearis Retz. Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia platyphyllos L. Euphorbia Preslii Guss. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. G. G. Ir. Ir. Ir. Madeira Is. Ir. K. Ir. Eur. Mi. G. Mi. C. Mi. Ir. Ir. Med. Reg. Ir. Eur. Mi. Ir. Eur. Mi. Ir. Med. Reg. Ir. Mi. Ir. Med. Reg. Ir. Med. Reg. Ir. Med. Reg. Ir. Mi. Ir. Med. Reg. Ir. Ir. Med. Reg. Ir. Med. Reg. Ir. Ir. Ir. Med. Reg. Ir. Ir. Ir. Ir. Med. Reg. Ir. Ir. Ir. Ir. Ir. Med. Reg. Ir. Ir. Ir. Ir. Ir. Med. Reg. Ir. Ir. Ir. Ir. Ir. Ir. Med. Reg. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir | Huphorbia Hathyris H. | | 1 - | 14. 14. 11 |
| Euphorbia lingularia Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia neriifolia L. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia Preslii Guss. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. | Euphorbia linearis Retz. | | 1 | W. I. |
| Euphorbia marginata Pursh. Euphorbia mellifera Ait. Euphorbia neriifolia L. Euphorbia officinarum Euphorbia palustris L. Euphorbia Peplus L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia platyphyllos L. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. G. Ches. G. Ir. G. R. Ir. F. P. Ind. Madeira Is. Ir. N. Am. Madeira Is. Ir. M. Madeira Is. Ir. N. Afr. Ir. M. Madeira Is. Ir. N. Afr. Eur. N. Afr. Eur. N. As. Ir. Eur. Med. Reg. Ir. Mi. Ir. Med. Reg. Ir. Eur. Afr. Smi. Ir. Med. Reg. Ir. Ir. Ir. Ir | | | 1 1 1 1 | |
| Euphorbia mellifera Ait. Euphorbia neriifolia L. Euphorbia officinarum Euphorbia palustris L. Euphorbia Peplus L. Euphorbia pithyusa L. Euphorbia platyphyllos L. Euphorbia platyphyllos L. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. G. R. Ir. F. P. Ind. Madeira Is. Ir. F. P. Ind. Malay Ir. Ir. Med. Reg. Ir. Ir. F. P. Mader. Is. Ir. Med. Reg. Ir. F. P. Mader. Is. Ir. Ir. Med. Reg. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir | Euphorbia marginata Pursh. | | 1 | N. Am. |
| Euphorbia neriifolia L. Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia Peplus L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia platyphyllos L. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. G. R. Ir. F. P. Ind. Malay Ir. Med. Reg. Ir. Smi. Ir. Mi. C. Sm. Ir. Ir. Med. Reg. Ir. F. P. Mader. Is. Mi. Ir. Mi. Ir. Smi. Ir. Smi. Ir. N. Am. Med. Reg. Ir. Afr. Ir. Madagas. Ir. F. P. Mader. Is. Med. Reg. Ir. Med. Reg. Ir. Med. Reg. Ir. Med. Reg. Ir. Madagas. Ir. Madagas. Ir. Mex. Ir. Madagas. Ir. Mex. Ir. Jamaica | | G. | Ir. | |
| Euphorbia officinarum Euphorbia palustris L. Euphorbia Paralias L. Euphorbia Peplus L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia platyphyllos L. Euphorbia Preslii Guss. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. Sm. G. Ir. Ir. Med. Reg. Ir. F. P. Mader. Is. Mi. Ir. Mi. Ir. Med. Reg. Ir. Med. Reg. Ir. Med. Reg. Ir. Med. Reg. Ir. Madagas. Ir. Mex. Ir. Jamaica | | G. R. | Ir. F. P. | |
| Euphorbia Paralias L. Euphorbia Peplus L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia platyphyllos L. Euphorbia Preslii Guss. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. Mi. C. Sm. Ir. Mi. Ir. Mi. Ir. Mi. C. Sm. Ir. Med. Reg. Eur. N. As. Ir. Smi. Ir. N. Am. Madagas. Ir. Madagas. Mi. Ir. Mi. C. Sm. Ir. Mi. C. Sm. Ir. Mi. C. Sm. Ir. Mi. Ir. Mi. C. Sm. Ir. | | Sm. | | |
| Euphorbia Paralias L. Euphorbia Peplus L. Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia pithyusa L. Euphorbia Preslii Guss. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. Mi. C. Sm. Ir. Mi. Ir. Mi. Ir. Mi. C. Sm. Ir. Mi. Ir. Mi. Ir. Mi. C. Sm. Ir. Mi. Ir. Mi. Ir. Mi. C. Sm. Ir. Med. Reg. Eur. N. As. Med. Reg. Eur. N. As. Med. Reg. Eur. N. As. Ir. Mi. Ir. Mi. C. Sm. Ir. Med. Reg. Eur. N. As. Ir. Mi. Ir. Mi. C. Sm. Ir. Med. Reg. Eur. N. As. Ir. Mi. Ir. Mi. Ir. Mi. C. Sm. Ir. Med. Reg. Eur. N. As. Ir. G. Ir. Med. Reg. Eur. N. As. Ir. Med. Reg. Eur. N. As. Ir. Med. Reg. Eur. N. As. Ir. Med. Reg. Eur. N. As. Ir. Med. Reg. Eur. N. As. Ir. Med. Reg. Eur. N. As. Ir. Med. Reg. Eur. N. As. Ir. Iv. Med. Reg. Eur. N. As. Iv. Iv. Med. Reg. Eur. N. As. Iv. Iv. Iv. Med. Reg. Eur. N. As. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv. Iv | Euphorbia palustris L. | | Ir. | Eur. |
| Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia platyphyllos L. Euphorbia Preslii Guss. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. Euphorbia punicea Sw. G. E. R. Ir. F. P. Mader. Is. Med. Reg. Eur. Afr. Smi. Ir. N. Am. Madagas. Ir. Madagas. G. Ir. Mex. G. E. Ir. F. P. Jamaica | Euphorbia Paralias L. | Mi. | Ir. | |
| Euphorbia piscatoria Ait. Euphorbia pithyusa L. Euphorbia platyphyllos L. Euphorbia Preslii Guss. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. Euphorbia punicea Sw. G. E. R. Ir. F. P. Mader. Is. Med. Reg. Eur. Afr. Smi. Ir. N. Am. Madagas. Ir. Madagas. G. Ir. Mex. G. E. Ir. F. P. Jamaica | Euphorbia Peplus L. | Mi. C. Sm. | Ir. | |
| Euphorbia platyphyllos L. Euphorbia Preslii Guss. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. G. Ir. Eur. Afr. N. Am. Madagas. Ir. Mex. G. E. Ir. F. P. Jamaica | Euphorbia piscatoria Ait. | G. E. R. | ltr. F. P. | Mader. Is. |
| Euphorbia Preslii Guss. Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. Smi. Ir. N. Am. Madagas. Ir. G. Ir. Mex. Ir. F. P. Jamaica | |) | | |
| Euphorbia primulaefolia Baker Euphorbia pulcherrima Willd. Euphorbia punicea Sw. G. E. Ir. Madagas. G. Ir. Madagas. Mex. Jamaica | | | | |
| Euphorbia pulcherrima Willd. Euphorbia punicea Sw. G. E. Ir. Mex. Jamaica | | | | |
| Euphorbia punicea Sw. G. E. Ir. F. P. Jamaica | | | | |
| | | G. | | |
| Emphorma pungitormis Boiss. G. Ir. | | | | Jamaica |
| | Emphorma pungitormis Boiss. | I G. | I F. | |

| NAME OF PLANT | Authority | Properties | Locality |
|---------------------------------------|----------------|-------------|----------------|
| Euphorbia Regis-Jubae Webb. & Benth. | I G. | Ir. | Teneriff Is. |
| Euphorbia resinifera Berg. | L. | Ir. | Morocc. |
| Euphorbia Royleana Boiss. | G. R. | Ir. F. P. | Him. Reg. |
| Euphorbia Sibthorpii Boiss. | G. R. | Ir. F. P. | Greece |
| Euphorbia Sieboldiana Moor. & Decue. | | Ir. | Japan |
| Euphorbia thymifolia L. | G. | Ir. | Tropics |
| Euphorbia Tirucalli L. | G. R. | Ir. F. P. | Afr. Ind. |
| Euphorbia venefica Trem. | G. K. | Ir. | Tr. Afr. |
| Euphorbia verrucosa Lam. | | | Eur. |
| | Mi. C. | Ir. | |
| Euphorbia Wulfenii Hoppe. | | Ir. | Dalmatia |
| Excoecaria Agallocha L. | G. R. M. C. | Act. F. F. | Tr. As. Malay |
| Europe d'année 7-11 and Man | L. | | Pac. Is. |
| Excoecaria virgata Zoll and Mor. | G. | r D | Java |
| Fluggea Leucopyrus Willd. | | F. P. | Ind. |
| Fluggea obovata Baill. | G. | | Tr. Afr. |
| Gymnanthes lucida Sw. | Sm. | _ | W. Ind. |
| Hippomane Mancinella | L. G. Sm. | | Ind. |
| Homalanthus crepitans L. | E.&G. C.R. | F. P. | |
| Homalanthus Leschenaultianus A. Juss. | G. M. | | Ind. Malay |
| | | | Austr. |
| Hyoemanche globosa | C. | | |
| Jatropha Curcas L. | G. C. R. L. | F. P. | Tr. Amphig. |
| Jatropha glandulifera Roxb. | G. | | Tr. As. & Afr. |
| Jatropha macrorhiza Benth. | G. | | Mex. |
| Jatropha multifida L. | G. 1 | Ir. | Tr. Amphig. |
| Jatropha urens L. | G. Sm. | Ir. | Tr. Am. |
| , | Ches. | | |
| Joannesia princeps Vell. | G. L. E. | Cath. F. P. | Braz. |
| Macaranga sp. | G. | | Java |
| Mallotus philippinensis Muell. | Sm. L. | Pur. | As. Astr. |
| Manihot Glaziovii Muell. | G. | | 110111 |
| Manihot utilissima Pohl. | | F. P. Hyd. | Braz. |
| 2011 | Wh. Sm. | | Diae. |
| | C L | | |
| Mecurialis annua | C. L. C. L. | Cath. Ir. | Eur. |
| Mecurialis perennis L. | | Ir. Cath. | Eur. |
| Mecurialis tomentosa | C. 2. | li, Catii. | Eur. |
| Pedilanthus tithymaloides Poit. | | Ir. Em. | Mex. S. Am. |
| Petalostigma cordifolia Fv. M. | G. L. | 11. 13 | Mex. S. Alli. |
| Phyllanthus Conami Sw. | E. G. 1 | F. P. | Tr. Am. |
| Thynanthus Conaini Sw. | Sm. Mi. | 1. 1. | II. Am. |
| Phyllanthus Emblica L. | G | | T. A. |
| | G | | Tr. As. |
| Phyllanthus epiphyllanthus L. | G. | | Cuba, |
| Phyllanthus Gasstromi Muell. | G. M. | 1 | Austr. |
| Phyllanthus lacunarius F. M. | G. M. | | Austr. |
| Phyllanthus Niruri L. | G. | | Tr. Reg. |
| Phyllanthus piscatorum H. B. & K. | G. Mi. E. | E D | Venezuela |
| Phyllanthus urinaria L. | | F. P. | Tr. Amphig. |
| Piranhea trifoliata Baill. | G. Mi. | 12 D | Braz. |
| Platygyna urens Mercier | C | F. P. | |
| Putranjiva Roxburghii Wall, | G | | Ind. Burma |
| Ricinus communis L. | G.Sm. C.L. | | Tr. Reg. |
| | Ches. | Pur. | , |
| Sapium indicum Willd. | | F. P. Nar. | Ind. |
| Sapium insigne Trim. | | Ir. | Ind. |
| Sebastiana Palmeri Riley. | G. | | |
| Sebastiana Pavoniana Muell, Arg. | G. | | |
| Securinega ramiflora Muell, Arg. | G. | Hyd. | As. |
| Stillingia lineata Muell, | G. | | Ind. Maurit. |
| | - | | |

| NAME OF PLANT | Authority | Properties | Locality |
|--|-------------------------------|--------------|------------------------------------|
| Stillingia sylvatica L. Toxicodendrum globosum Tragia cordifolia Vahl. Tragia involucrata L. Tragia volubilis L. | L. G. G. G. G. L. | Con. Ves. | U. S. Arabia Tr. As. Ind. |

FAGACEAE

| Castanopsis Tungurrut A. D C. Fagus sylvatica L. Quercus alba L. Quercus Toza Gillet | | | Malay Eur. E. N. A. S. Eur. |
|---|-------------------|-------|--------------------------------------|
| Quercus Toza Gillet Quercus lusitanicus Lam. Quercus sp. | C. L. Ches. | Astr. | Med. Reg. |

FICOIDEAE

| | | ···· | 10 7 46 |
|----------------------------------|----------------|-----------|-----------------|
| Aizoon canariense L. | G. | | Canar. Is. Afr. |
| | _ | J | Orient. |
| Gisekia pharnacioides L. | G. | Taen. | Taen. Ind. |
| Limeum sp. | G. | | S. Afr. |
| Mesembryanthemum anatomicum Haw. | G. G. G. | Nar. | Tr.Reg & Sub. |
| • | ì | ì | Tr. Reg. Afr. |
| Mesembryanthemum crystallinum L. | L. | Diur. | Gr. Can. Is. S. |
| 2.000 | | 1 | Afr. Calf. |
| Mollugo hirta Thunb. | G. M. | ł | Austr. |
| Tetragonia expansa Thunb. | G. M. | Son | Japan |
| | G. G. | Sap. | |
| Trianthema sp. | G. | Sap. | Arab. Afr. |
| | | J | Austr. |
| Trianthema Portulacastrum L. | G. | Sap. Par. | Tr. Am. |
| | | | |

FILICALES

| Adiantum pedatum L. | G. | Astr. | E. As. Jap. |
|------------------------------|----------------------|--------|----------------|
| · | 1 | 1 | N. A. |
| Adiantum peruvianum Kl. | G. | | Peru |
| Adiantum trapeziforme L. | G. | Í | Trop. Am. |
| Anemia oblongifolia Sw. | G. | 1 | Trop. Am. |
| Angiopteris erecta Hoffm. | G. G. G. G. | 1 | · · |
| Aspidium athamanticum Ktze. | G. L. | Ant. | S. Afr. |
| Aspidium Filix-Mas Sw. | G. L. | Ant. | Eur. N. A. |
| Aspidium marginale Ktze. | G. L. | Ant. | Eur. N. A. |
| Aspidium odoratum Willd. | G. | | N. Ind. E. As. |
| Aspidium prestulatum Ten. | G. G. | An. | 1 |
| Aspidium rigidum Sw. | | | |
| Ceterach officinarum Willd. | G. L. | Abort. | Eu. Ind. |
| Cheilanthes fragrans | G. | 1 | Med. Reg. |
| | _ | 1 | Him. Reg. |
| Cryptogramme crispa R. Br. | G. | | N. Eur. N. W. |
| | _ | 1 | Am. |
| Cystopteris alpina Desv. | G. | Hyd. | Eur. Mt. Reg. |
| Cystopteris bulbifera Bernh. | G. | Hyd. | N. Am. |
| Cystopteris fragilis Bernh. | G. L. | Hyd. | Eu. N. Am. |
| a | | | Cosm. |
| Cystopteris montana Bernh. | į. | Hyd. | Cent. Eur. E. |
| 75 111 1 111 1 77 1 | | | Malay, Trop. |
| Davallia brasiliensis Hook. | G. | Hyd. | As. Austr. |
| Davallia elegans Sw. | G. | Hyd. | N. A. |
| | | | IS. A. |

| NAME OF PLANT | Authority | Properties | Locality |
|--|-----------|------------|-------------------------|
| Davallia hirsuta Kaulf. | G. | Hyd. | Japan |
| Davallia majuscula Lowe | Ğ. | Hyd. | Ceylon |
| Davallia pentaphylla Don. | Ğ. | Hyd. | Java & adj. Is. |
| Davallia platyphylla Don. | 0. | Hyd. | Ind. |
| Davallia strigosa Sw. | G. | 1 | N. Ind. Japan, |
| | | | Malay |
| Geniophlebium incanum Sw. | G. | | |
| Gleichenia flabatellata R. Br. | G. | Sap. | |
| Gymnogramme cordata Schl. | G. | | S. Afr. |
| Lindsaea cultrata Bl. | G. | | N. Ind. Malay |
| Polypodium laciniatum Bl. | G. | | Mad. Austr. |
| Polypodium ligulatum Sw. | G. | | Java |
| Polypodium percussum Cav. | G. | | S. A. |
| Polypodium Phymatodes L. | G. | | Malay, Ceylon, |
| Del cedim a sub- a Park | | | Austr. |
| Polypodium scandens Forst. | G. G. | | N. Z. Austr. |
| Polypodium suspensum L. | L. | Exp. Diur. | Trop. Am. Eu. N. Am. |
| Polypodium vulgare L. Polystichum spinulosum DC. | Ğ. | Exp. Diur. | N. Am. |
| Pteris aquilina L. | G. | Hyd. Ant. | Coc.Eu. N.Am. |
| Pteris caudata L. | G. | Ant. | Coc.Eu. N.Am. |
| Trong Character 27 | <u> </u> | 1- 11-01 | <u>'</u> |
| FLAGELLA | ARIACEA | E | |
| Susum anthelminthicum Bl. | G. | | Java |
| FRANKE | TIACEAE | | |
| | | let D | 10 |
| Frankenia ericifolia Chr. | Sm. G. | F. P. | Canary Is. |
| Frankenia grandifolia Cham. & Schlecht. | L. | Astr. | Calif. |
| FUMAR | IACEAE | | |
| Corydalis racemosa Pers. | G. | | Japan |
| Dicentra canadensis Walp. | L. | Diur. | N. A. |
| Fumaria officinalis L. | GL. | Nar. | Temp.old world |
| Fumaria spicata L. | G. | | Lusitan. |
| GENTIA: | NACEAE | | |
| Gentiana verna L. | G. | | Eur. Cauc. Reg. |
| Menyanthes trifoliata L. | G. | | N. Temp. Reg. |
| Tachia guianensis Aubl. | G. | | Guian. |
| GERAN | IACEAE | | |
| Erodium cicutarium | | | Eu. N. Am. S. |
| Pelargonium peltatum Ait. | G. | | Am. S. Afr. |
| • | ACEAE | | , |
| | | | |
| Ephedra L. | G. | 1. | Eur. |
| Ephedra antisyphilitica C. A. Mey. | L. | Astr. | N. A. Mex. |
| Gnetum scandens Roxb. | G. | F. P. | Ind. Malay |
| Gnetum urens Blum. | G. | Ur. | Guian. |
| | IIACEAE | | |
| Goodenia grandiflora Sims. | G. | 14 70 | Austr. |
| Scaevola sp. | G. | A. P. | Austr. |
| Velleia paradoxa R. Br. | G. M. | 1 | Austr. |
| | | | |

GRAMINEAE

| OMMINIMI | | | | |
|---|--------------------|-------------------------------|--|--|
| NAME OF PLANT | Authority | Properties | Locality | |
| Agrostis alba L. Andropogon annulatus Forsk. Andropogon halepensis Anthistiria prostrata Willd. | *Hell. G. | Ur. Hyd. | N. Temp. Reg. Afr.Ind. Austr. Eur. Afr.N.Am. | |
| Anthoxanthum odoratum L. Arrhenatherum avenaceum Beauv. Avena fatua L. Briza sp. | G. G. G. | Cou. Sap. Mech. Hyd. | Eur. As. Afr. Eur. Eur. Ori. As. S. Am. Eur. N. | |
| Bromus catharticus Vahl. Bromus mollis L. Bromus secalinus L. | G. G. G. | | Am. As. Eur. Afr. As. Eur. Med. Reg. As. | |
| Bromus tectorum L. Catabrosa sp. | Pa. G. | Ir. Hyd. | Eu. N. Am. | |
| Cenchrus tribuloides L. Cortaderia argentea Stopf. | G. G. | Mech. | N. Am. | |
| Cortaderia conspicua Cortaderia Kermesiana Deyeuxia Langsdorffii Kunth. | G. G. G. | Hyd. Hyd. Sap. | N. Am. | |
| Elionurus sp. | G. G. | | S. Af. S. Am. Austr. | |
| Elymus sp. Festuca sp. Festuca quadridentata H. B. & K. | G. G. C. | Hyd. Hyd. | Eur. N. Am. Equador | |
| Glyceria sp. Glyceria aquatica Wahleb. | G. G. | Hyd. Hyd. | N. Am. Eur. N. Tem. Reg. Tr. Reg. | |
| Heteropogon hirtus Pers. Hierochloe australis R. & S. Hierochloe borealis Roem. & Schult. | G. | Cou. | Eur. N. Reg. | |
| Hierochloe rariflora Hook, Holcus sp. Lolium Lamarckii L. | G. C. | Cou. Hyd. Hyd. Del. | Eur. N. Am. Eur. As. | |
| Lolium perenne L. Lolium temulentum L. | G. C. Mi. L. Ches. | Hyd. Nar. Intox. | Eur. As. Eur. As. U. S. | |
| Melica sp. Milium effusum L. Molinia caerulea Moench. | G. G. G. C. | Hyd. | N. Temp. Reg. Eur. As. | |
| Panicum Panicum junceum Nees. Panicum sanguinale L. | G. G. G. | Hyd. Hyd. Em. | N. Temp. Reg. Braz. Cosmop. | |
| Paspalum scrobiculatum L. Setaria scandens Schrad. | G. | Nar. | Tr. old world Braz. | |
| Sorghum vulgare Pres. Spartium junceum Stipa capillata L. | G. C. L. G. | Hyd. EmCath. | Tr. Reg. Cosmop. S. Am. As. | |
| Stipa inebrians Hance. Stipa palmata Pall. | G. | Del. | China. | |
| Stipa pennata Hohen. Stipa robusta Nash. Triodia irritans Brown | G. Ches. G. | Mech. Meb. Mech. | Eur. As. N.Ant. N. Am. Ry.Mts. Austr. | |
| Zea Mays L. | G. C. L. | | Paraguay | |

^{*}A case of poisoning by Agrostis alba was reported from West Liberty, la., by Dr. Hell. Probably forage poisoning.

GUTTIFERAE

| | PERAL | | |
|---|---|--|---|
| NAME OF PLANT | Authority | Properties | Locality |
| Calophyllum Calaba Jacq. Calophyllum Inophyllum L, Calophyllum montanum Vieill. Clusia macrocarpa Spreng. Garcinia Cambogia Desr. Garcinia Forsteriana Bl. Mammea americana L. Stalagmitis Mangle | G. G. G. G. Sm. G. G. L. | Sap. F. P. Sap. Ant. | Caith. Is. Tr. old world N. Caled. Guiana Ind. Tr. Am. |
| HAEMODO | ORACEAE | , | • |
| Aletris farinosa L. Gyrotheca majalis Morong. Haemodorum sp. Ophiopogon sp. Sansevieria thyrsiflora Thumb. | G. Ches. G. G. G. | | N. Am. E. N. Am. Austr. Braz. Japan, Ind. S. Afr. |
| HIPPOCAST | TANACEA | E | |
| Aesculus californica Nutt. Aesculus flava Ait. Aesculus glabra Aesculus Hippocastanum L. Aesculus Pavia L. | Ches. G. R. Sm. Ches. G. Ches. L. G. R. Ches. L. | Sap. Sap. F. P. Sap. Sap. Sap. F. P. | W.N.Am, Calif. N. Am. N. Am. Turkey N. Am. |
| HYDROPHY | YLLACEA | E | |
| Eriodictyon glutinosum Benth. | G. L. | Exp. | Calif. |
| HYPERI | CINEAE | | |
| Hypericum crispum L. Hypericum humifusum L. Hypericum maculatum Walt. Hypericum perforatum L. | G. G. Ches. Ches. | Acr. Acr. | Med. Reg. Eur. As. E. N. Am. Eur. N. Am. |
| ILICIN | | | |
| Ilex Aquifolium L. | | Em. Cath. | Eur. As. |
| ILLECEB | | | |
| Paronychia bonariensis DC. Paronychia capitata Lam. | G. G. | Sap. | Argent. Reg. Eur. Orient. |
| IRID | EAE | | |
| Belamcandra punctata Moench. Crocus sativus L. Gladiolus communis L. Gladiolus segetum Homeria collina Vent. Iris Iris florentina Sair. Iris foetidissima L. Iris germanica L. Iris nybrida Retz. Iris neglecta Parl. | C. C. G. M. G. Sm. C. L. Sm. C. C. L. | Ir. Ir. Ir. Ir. Ir. Ir. Ir. Ir. | China. As. Min. Eur. Orient. Med. Reg. S. Afr. N. Am. S. Eur. Eur. Orient. Eur. Orient. |

| NAME OF PLANT | Authority | Properties | Locality |
|---|---------------|--------------------|------------------------------|
| Iris prismatica Pursh. | L. | Ir. | N. A. |
| Iris Pseudoacorus L. | C. Sm. Sm. | Ir. | Eur. As. Ori. |
| Iris reticulata Bieh. | | Ir. | As. Minor |
| Iris sibirica 4. | C. | Ir. | Eur. N. As. |
| Iris variegata L. Iris versicolor | Wh. | Ir. Ir.Em.Cath. | Eur. Orient. |
| Sisyrinchium angustifolium Mill. | L. Rusby | Cath. | N. A. |
| | IDACEAE | 15-1-1-11 | , |
| Juglans cinerea L. | L. | Cath. | N. A. |
| Juglans regia L. | C. L. | Ant. Astr. | Him. Reg. |
| LAB | ATAE | | |
| Achryospermum Gl. | G. | Urt. | Mad. |
| Eremostachys superba Royle. | G. R. | F. P. | Him. Reg. |
| Lagochilus inebrians Bunge. | G. | Nar. | As. N. Tem. Reg. |
| Leonurus Cardiaca L. | G. L. | Card. | N. Tem. Reg. |
| Ocymum viride Willd. | Sm. | | Trop. Air. |
| Salvia amarissima Orteg. | G. | Nar. | Mex. |
| Salvia officinalis L. | L. | Nar. Nar. | Med. Reg. |
| Salvia pratensis L. | G. L. | Sed. | Eur. Caucas. N. Tem. Reg. |
| Scutellaria galericulata L. Scutellaria lateriflora L. | L. | Sed. | N. A. |
| Stachys arvensis L. | G. M. L. | Antisp. | V.Tem. old wld. |
| Stachys palustris L. | L. | Antisp. | Eur. As. N. A. |
| Tetradenia fruticosa Benth | Ğ. | Pur. | Madag. |
| Teucrium Chamaedrys L. Teucrium Marum L. | L. | | Eur. |
| Teucrium Marum L. | G. C. L. | Diaph. | Med. Reg. |
| Teucrium scordium | C. L. | Ant. | Eur. |
| LAUR | ACEAE | | |
| Hernandia sonora L. | I G. | P. | Ind. |
| Laurus nobilis L. | Sm. L. | } | Med. Reg. |
| Lindera Benzoin Muss. | Sm. L. | Ant. | E. N. Am. |
| Umbellularia californica Nutt. | G. C. L. | Hyd. | N. Am. |
| LEGUN | MINOSAE | | |
| Abrus precatorius L. | M. Sm. L | Toxal. Ira. | |
| Abrus pulchellus Wall. | Mi. | | Ind. Afr. |
| Acacia sp. | G. C. | F. P. | Austr. Tr. Reg |
| Acacia arabica Willd. | G. L. | 1. | Afr. As. |
| Acacia catechu Willd. | L. | Astr. | Ind. Ceylon |
| Acacia concinna DC. Acacia Cunninghamii Hook. | G. | Astr. | Chili Austr. |
| Acacia delibrata A. | G. | Sap. | Austr. |
| Acacia falcata Willd | G. G. | Sap. | Austr. |
| Acacia falcata Willd. Acacia Georginae Bailey | G. | | rusti. |
| Acacia Jurema Mart. | L. | Astr. | Brazil |
| Acacia pennata Willd. | Ğ. | | As. Afr. |
| Acacia penninervis Sieber. | Ğ. | | Austr. |
| Acacia pruinescens Kurz. | G. | F. P. | Burm. |
| Acacia pulchella R. Br. Acacia salicina Linal. | G. | | Austr. |
| Acacia salicina Linal. | G. | | Austr. |
| Acacia verniciflua A. Cunn. | G. M. | | Austr. |
| Acacia verticillata Willd. | G. | Sap. | Austr. |
| Adenanthera pavonina L. | G. | 1 | Tr. As. Malay |

| NAME OF PLANT | Authority | Properties | Locality |
|---|-----------------|------------|--------------------------------|
| | | | |
| Albizzia anthelmintica Brongn. | G. L. | Sap. F. P. | Abyss. |
| Albizzia Lebbek Benth. | G. | | Tr. old world |
| Albizzia odoratissima Bentlı. | | | Tr. As. Ind. |
| Albizzia procera Benth. | G. R. E. | lr D | Tr. As. Austr. |
| Albizzia stipulata Boiv. | | F. P. | A *** Car |
| Anagyris foetida | C. G. G. | 1000 | Sm. Cyt. |
| Andira anthelmintica Benth. | L. | Ant. | Braz. |
| Andira araroba Aguiar. | | Ir. | Braz. |
| Andira Aubletia Benth. | L, | Ant. | Guiana |
| Andira inermis H. B. K. | L. | Ir. | W. Ind. Braz. |
| Andira rosea Mart. | _G. | F. P. | Braz. |
| Andira retusa HBK. Andira spinulosa Mart. & Benth. | G. L. L. | Ant. | Guiana Braz. |
| | Mills. | Ant. | Braz. |
| Andira vermifuga Mart. & Benth. | | Ant, | Afr. Austr. |
| Argyrolobium pumilum Eckl. & Zeyh. | G. L. | C | Sic. |
| Astragalus baeticus L. | Ches. | Sap. | W. N. Am. |
| Astragalus Bigelovii A. Gray. | T T | Loco | 1 |
| Astragalus exscapus L. Astragalus galegiformis L. | L. G. | Diur. | Eur. |
| | G. | Sap. | Caucas. Reg. |
| Astragalus garbancillo Cav. | G. L. | D: | Peru |
| Astragalus glycyphyllos L. | G. L. | Diur. | Siber. Eur. |
| A - 4 1 | | | Caucas. Reg. |
| Astragalus hamosus L. | G. | | Caucas. Reg. |
| A . 1 TT . !! A . C | 0 01 | DIT | Med. Reg. Ind. |
| Astragalus Hornii A. Gray. | G. Ches. | Del. Loco. | |
| Astragalus lentiginosus Dougl. | G. L. | Del. Loco. | |
| Astragalus maximus Willd. | G. | Sap. | Armen. Caucas. |
| Astragalus mollissimus Torr. | G. L. Ches | Del. Loco. | Reg. N.Am. Ry. Mts. Col. |
| Astragalus Mortoni Nutt. | G. L. | Del. Loco. | |
| Astragalus ochroleucus Cass. | G. | Del. | Morocco |
| Astragalus occarpus A. Gray. | L. | Del. Loco. | |
| | Ches. | Del. Loco. | W. N. Am. |
| Astragalus Pattersoni A. Gray. | G. | Cyt. | N. Am. |
| Baptisia sp. Baptisia tinctoria R. Br. | L. | Em. Cath. | N. A. |
| | Ğ. | Em. Cath. | Peruv. |
| Barbieria maynensis Poepp. Barbieria polyphylla DC. | G. | | Porto Rico |
| Bauhinia coccinea DC. | G. | | Cochin China |
| Bauhinia guianensis Aubl. | G. | F. P. | Guiana. |
| | G. L. | | China, Burma. |
| Bauhinia variegata L. | 0. 1. | Ant. | Ind. |
| Danidishia similiaidan II D V | R. L. | F. P. | IS. A. |
| Bowdichia virgilioides H. B. K. | | [F. F. | |
| Brachysema undulatum Ker. Gawl. | M. | | Austr. |
| Caesalpinia Bonducella Fleming. | G. L. | A =4 = | Tr. Cosmop. |
| Caesalpinia coriaria Willd. | | Astr. | N. A. |
| Caesalpinia pulcherrima Sw. Calliandra Houstoni Benth. | G. | C | Tr. Cosmop. |
| | G. | Sap. | Mex. |
| Camptosema pinnatum Benth. | G. | | Braz. |
| Canavalia obtusifolia DC. | G. | Descri | Cos. Trop. |
| Cassia acutifolia Deli!. | G. L. | Pur. | Egypt Con Trop |
| Cassia alata L. | G. | F. P. | Cos. Trop. |
| Cassia angustifolia Vahl. | L. | Pur. | Trop. Afr. |
| Cassia didymobotrya Fres. | G. | E É | Tr. Am. Abyss. |
| Cassia hirsuta L. f. | G. R. | F. P. | Tr. Am. |
| Cassia kituiensis Vatke. | G. | F. P. | Tr. Afr. |
| Cassia laevigata Willd. | G. M. | C | Cos. Trop. |
| Cassia marylandica L. | G. L. Sm. L. | Sap. | Tr. m. Cosm. Trop. |
| Cassia occidentalis L. | Sill. L. | Pur. | Cosm. 110p. |

| NAME OF PLANT | Authority | Properties | Locality |
|---|--|------------|-----------------|
| Cassia sophora L. | G. M. | | Tr. old world |
| Cassia stipulacea Ait. | G. | ĺ | Chili |
| Cassia Sturtii R. Br. | G. M. | | Austr. |
| Castanospermum australe A. Cunn. | G. M. | Sap. Ir. | Austr. |
| Centrosema amazonicum Mart. | G. R. | F. P. | Brazil |
| Centrosema Plumieri Benth. | G. R. | F. P. | S. Am. |
| Cercis canadensis L. | G. | Sap. | N. Am. |
| Cercis chinensis Bunge. | G. | Sap. | China |
| Cladrastis amurensis Benth. | G. | J | Amur. Reg. |
| Clitoria amazonum | R. | F. P. | S. Am. |
| Clitoria arborescens Ait. | G. R. | F. P. | Trinidad Is. |
| Clitoria Ternatea L. | L. | Em. | Trop. Reg. |
| Copaifera Langsdorffii Desf. | L. | Diur. | Braz. |
| Copaifera Salikorinda Heck. | G. | Cou. | S. Am. |
| Coronilla Emerus L. | G. C. | | S. Eur. |
| Coronilla juncea L. | G. | | Eur. |
| Coronilla montana Scop. | G. | 10 1 | D. D. M. I |
| Coronilla scorpioides Koch. | | Card. | Per. Reg. Med. |
| Coronilla varia L. Crotalaria alata Hamill. | G. C. | | Eur. |
| Crotalaria Mitchelii Benth. | G. M. | | Tr. As. |
| Crotalaria mitchem Benth. Crotalaria paniculata Willd. | M. G. | | Austr. |
| Crotalaria sagittalis L. | G. | Toursh | Tr. As. |
| Crotalaria sagittans 4. Crotalaria striata Schrank. | | Ineb. | N. Am, Braz. |
| Crotalaria verrucosa L. | G. | | Cosm. Trop. |
| Cytisus Alschingeri Vis. | | | |
| Cytisus austriacus L. | C. | | Dalmatia. |
| Cytisus biflorus Ker. Gawl. | C. C. C. C. C. C. C. | | S. Eur. |
| Cytisus capitatus Scop. | ļ č. | | S. Eur. |
| Cytisus hirsutus L. | ~ | | As. Min. |
| Cytisus nigricans L. | ا <u>۲</u> | | Eur. As. |
| Cytisus proliferus L. | 1 č | } | Teneriffe |
| Cytisus purpureus Scop. | č | 1 | Eur. As. |
| Cytisus scoparius Link. | G. C. L. | Diur. | Eur. Jap.N.Am |
| Cytisus sessilifolius L. | C. C. | | S. Eur. |
| Cytisus supinus L. | C. C. G. | | Eur. |
| Dalbergia lanceolaria L. f. | Ğ. | | Ind. |
| Dalbergia toxicaria Baill. | G. | | Madag. |
| Derris amoena Benth. | G. | | Burm. Malaya |
| Derris elliptica Benth. | G. L. | F. P. Ins. | Burm, Malaya |
| Derris guianensis Benth. | G. R. | F. P. | Guiana. Braz. |
| Derris Negrensis Benth. | | F. P. | Braz. |
| Derris uliginosa Benth. | E.&G. R. L. | F. P. | Tr. old world |
| Detarium Senegalense J. F. Gmel. | G. | | Tr. Afr. |
| Entada polystachya DC. |] G. | | Tr. Am. |
| Entada scandens Benth. | G. L. | Sap. | Tr. Am. |
| Enterolobium cyclocarpum Griseb. | G. | Sap. | Jamaica |
| Enterolobium timboiiva Mart. | G. | Sap. | Braz. |
| Eperua falcata Aubl. | G. | Em. | Guiana |
| Erythrina Corallodendron L. | | Diur. | E. Ind. N. Am. |
| Erythrina Hypahphones Boerl. | G. | | |
| Erythrina Mulungu Mart. | L. | Нру. | Braz. |
| Erythrophleum Comminga Baill. | C. | 4 D 37 | Mad. |
| Erythrophleum guineense G. Don. | C. L. | A. P. Nar. | Trop. Afr. |
| Euchresta Horsfieldii Bena. | 1 | I 75- | Jav. |
| Flemingia congesta Roxb. Galega officinalis L. | CTI | Taen. | Tr. As. |
| Galega orientalis Lam. | G. L. | Sap. | Eur. As. |
| Gastrolobium bilobum R. Br. | G. G. | Sap. | Caucas, Reg. |
| Sacri Stobium Dhobuill R. DI. | G. M. Sm. | Ineb. | Austr. |

| | i | 1 | |
|-----------------------------------|----------------|------------|-----------------|
| NAME OF PLANT | Authority | Properties | Locality |
| Gastrolobium calycinum Benth. | G. M. Sm. | Ineb. | Austr. |
| Gastrolobium Callistachys Meiss. | G. M. Sm. | Ineb. | Austr. |
| Gastrolobium grandiflorum F. M. | G. | Ineb. | Austr. |
| Gastrolobium obovatum Benth. | G. M. Sm. | Ineb. | Austr. |
| Gastrolobium ovalifolium Henfr. | G. | Ineb. | Austr. |
| Gastrolobium oxylobioides Benth. | G. M. Sm. | | Austr. |
| Gastrolobium spinosum Benth. | G. M. Sm. | | Austr. |
| Gastrolobium trilobum Benth. | G. M. Sm. | | Austr. |
| Genista monosperma Lam. | G. | Ineb. | Med. Reg. |
| | G. | Ineb. | Med. Reg. |
| Genista sphaerocarpa Lam. | | | Fran Cara Dan |
| Genista tinctoria L | L. | Sur. | Eur. Cau. Reg. |
| Geoffraea superba | G. | Ant. | S. Am. |
| Gleditschia amorphoides Griseb. | G. | Mech. | N. Am. |
| Gleditschia sinensis Lam. | G. | Sap. | China |
| Gleditschia triacanthos L. | G. | Mech. | N. Am. |
| Gliricidia maculata HBK. | G. | | Guat. |
| Glycine hispida Max. | G. | Em. | Japan |
| Gompholobium uncinatum A. Cunn. | G. M. | Ineb. | Austr. |
| Goodia lotifolia Salisb. | G. | Ineb. | Austr. |
| Gymnocladus canadensis | G. C. | Sap. | N. Am. |
| Gymnocladus chinensis Buill, | G. | Sap. | China |
| Halimodendron argenteum Fisch. | Ğ. | Sap. | Reg. Caus. Per. |
| Translodendron argenteum Pisch. | 0. | Sap. | |
| Hosackia Purshiana Benth. | G. | | N. As. |
| | | A 4 | N. Am. |
| Hymenaea Courbaril L. | G. | Ant. | S. Am. |
| Indigofera anil L. | G. | | S. Am. |
| Indigofera australis Willd. | G. M. | | Austr. |
| Indigofera galegoides DC. | G. | Hyd. | Malay _ |
| Indigofera tinctoria L. | G. | Ins. | Trop. Reg. |
| Isotropis juncea Turcz. | M | | Austr. |
| Laburnum anagyroides Medic. | C. Sm. L. | Depr. Cyt. | Eur. |
| Lathyrus amocnus Fenzl. | C. | | Syria |
| Lathyrus Aphaca L. | C. | Ast. Hyd. | Eur. Orient. |
| Lathyrus Cicera L. | C. C. C. | | Eur. Orient. |
| Lathyrus Clymenum L. | 1 C. | | Med. Reg. |
| Lathyrus odoratus L. | C. | | Italy |
| Lathyrus purpureus Presl. | C. | | Sic. |
| Lathyrus sativus L. | F. Sm. C. | Ent. | Eur. N. Afr. |
| | | | Orient. |
| Lessertia annularis Burch. | M. | | Austr. |
| Leucacna glauca Benth. | G. | | Trop. |
| Lonchocarpus densistorus Benth. | E. G. R. | ਂ ਧ | Guiana |
| Lonchocarpus ichthyoctonus Baill. | G. 1 | 1 | Madag. |
| Lonchocarpus latifolius Kth. | G. R. | F. P. | madag. |
| Lonchocarpus Nicou DC. | | | Tr. Am. |
| Lonchocarpus rariflorus Hart. | | r. r. | Tr. Afr. |
| | R. | r n | Braz. |
| Lonchocarpus violaceus Kunth. | G. Sm. M. | F. P. | Tr. Afr. |
| Lotus australis Andr. | | | Austr. |
| Lotus corniculatus L. | M. | | Tem. old world |
| т • | | | Austr. |
| Lupinus sp. | G. | | N. Temp. Reg. |
| Lupinus albus L. | C. Ches. | | S. Eur. |
| Lupinus angustifolius L. | C. | | Med. Reg. |
| Lupinus densiflorus Benth. | Sm. | | Cal. |
| Lupinus leucophyllus Dougl. | Ches. | | W. N. Am. |
| Lupinus luteus L. | C. | | Med. Reg. |
| Melilotus alba Lam. | L. | Con. | |
| Melilotus indica Desf. | | Con. | Eur. As. |
| Melilotus officinalis Desr. | L. | Con. | Eur. N. As. |
| | | | |

| | | | 1 |
|--|----------------|-------------|------------------------|
| NAME OF PLANT | Authority | Properties | Locality |
| Milletia caffra Meiss. | G. | | S. Afr. |
| Milletia ferruginea Hochst. | | F. P. | S. Afr. |
| Milletia pachycarpa Benth. Milletia rostrata Meig. | G. G. | | Malay |
| Milletia sericea G. & E. | | F. P. | Java Burm. Malay |
| Milletia splendens Wright & Arn. | G. | 1. 1. | Ind. |
| Mucuna capitata Max. | Ğ. | F. P. | Him.Reg.Malay |
| Mucuna gigantea DC. | G. | | Tr. Afr. |
| Mucuna purpureus DC. | Whi. | Ir. | Trop. |
| Mucuna urens DC. | L. | Ir. | S. Am. |
| Mucuna venenosa A. Murr. Muellera moniliformis L. | G. G. E. R. | F. P. | Tr. Afr. Trop. Afr. |
| Muellera Telfairii Baker | G. E. K. | r, I. | Mad. |
| Myroxylon Pereirae Klotzsch. | L. | Stim. | C. A. |
| Myroxylon peruiferum L. | L. | Stim. | Eq. Peru, Braz. |
| Myroxylon toluiferum HBK. | L. | Cou. | Trop. Am. |
| Nissolia fruticosa Jacq. | Ğ. | F. P. | Tr. Am. |
| Ormocarpum glabrum Teis. & Binn. | G. G. | 1 | Celebes. |
| Ormosia coccinea Jacks. Ougeinia dalbergioides Benth. | Ğ. | 1 | Tr. Am. |
| Oxylobium parviflorum Benth. | G. M. | 1 | Austr. |
| Oxytropis foetida DC. | Sm. | | Eur. |
| Oxytropis Lambertii Pursh. | Ches. | Loco | W. N. Am. Ia. |
| | | | Ry Mts. |
| Oxytropis Lapponica Gaud. | G. | Hyd. Sap. | Eur. N. As. |
| Oxytropis sulphurea Fisch. | G. G. | Hyd. | Siber. |
| Pachyrhizus angulatus Rich. Pachyrhizus tuberosa Spreng. | G. | F. P. | Tr. As. Ind. |
| Parkia africana R. Br. | Ğ. | Ir. 1. | Tr. Afr. |
| Pentaclethra macrophylla Benth. | G. | Alk. | Tr. Afr. |
| Phaseolus aconitifolius Jacq. | G. | Hyd. Ast. | Ind. Ar. |
| Phaseolus lunatus L. | Sın. | | S. Am. Cosm. |
| Phaseolus multiflorus Willd. | G. | | Cult. Mex. |
| Phaseolus semierectus L. | G. R. | F. P. | Trop. Reg. |
| Phaseolus vulgaris L. | G. C. | 1. 1. | N. Am. Cosm. |
| 3 | | 1 | Cult. |
| Physostigma venenosum Balf. | G. Sm. L. | Dep. Eser. | Afr. |
| Piptademia peregrina Benth. | C C D I | | Braz. |
| Piscidia Erythrina L. E. Pithecolobium bigeminum Mart. | G.Sm.R.L. | Ineb. P. P | Trop. Am. |
| Pithecolobium fasiculatum Benth. | Ğ. | F. P. | Malay |
| Pithecolobium Minahassae Teis. & Binn. | | 1 | Malay |
| Pithecolobium montanum Benth. | G. | | Him.Reg.Malay |
| Pithecolobium Saman Benth. | _ G · | Sap. | Tr. Am. |
| Pongamia Piscida Steud. | G. E. | [F. P. | Ind. |
| Prosopis juliflora Benth. | G. G. | lc | N. Am. Tex. |
| Prosopis ruscifolia Griseb. Psoralea glandulosa L. | G. L. | Sap. Em. | Argen. Chili. Mex. |
| Psoralea macrostachya DC. | G. | Sap. | Cal. |
| Psoralea pentaphylla L. | G. | | Mex. |
| Psoralea tenuiflora Pursh. | G. | Sap. | N. Am. |
| Pterocarpus esculentus Schw. | [G. | Intox. | Tr. Afr. |
| Pterocarpus Marsupium Roxb. | L. G. | Astr. | Ind. |
| Rhynchosia minima DC. | G. | | Cosmop. Trop. |
| Robinia Pseudo-acacia L. | G. Ches. | Toxal. | States |
| | L. Rusby | | N. A. |
| Robinia viscosa Vent. | I L. Kusby | | 114. A. |

| NAME OF PLANT | Authority | Properties | Locality |
|------------------------------------|----------------------|----------------|--------------------------|
| Sabinea florida DC. | i G. | <u>'</u> | IS. Eur. |
| | G. | | N. Am. S. |
| Securigera Coronilla DC. | | | |
| Sesbania vesicaria Ell. | Ches. | | States |
| Sophora alopecuroides L. | [G. | l D | Him. Reg. |
| Sophora japonica L. | L. | Pur. | Japan, China |
| Sophora mollis Grah. |] G. | } | China, Him. Reg. |
| Sophora secundiflora Lah. | G. Ches. L. | | Mex. |
| Sophora sericea Nutt. | L. Ches. | Nar. | N. A. |
| Sophora tomentosa L. | G. M. | Sop. | Cold Reg. |
| Stryphnodendron Barbatimam Mart. | G. | Sap. | Braz. |
| Stryphnodendron polyphyllum Mart. | L. | Astr. | Braz. |
| Swainsona coronillaefolia Salisb. | G. | Ineb. | Austr. |
| Swainsona Greyana Lindl. | G. M. | Ineb. | Austr. |
| Swainsona Oliverii F. Muell. | M. | Ineb. | Austr. |
| Swainsona phacoides Benthm. | M. | | Austr. |
| | M. | Ineb. | |
| Swainsona procumbens F. Muell. | | | Austr. |
| Swartzia triphylla Willd. | G. M. | | Tr. Am. |
| Templetonia egena Benth. | | | Austr. |
| Templetonia purpurea | M. | | Austr. |
| Templetonia retusa R. Br. | G. | | Austr. |
| Tephrosia astragaloides Benth. | G. | | Austr. |
| Tephrosia candida DC. | G. R. | F. P. F. P. | Tr. As. |
| Tephrosia cinerea Pers. | G. R. | F. P. | |
| Tephrosia coronillaefolia DC. | G. R. | F P | Borbon, Is. |
| Tephrosia densifiora | G. | F. P. F. P. | Tr. Afr. |
| Tephrosia frutescens DC. | Sm. | 1. 1. | Honduras. |
| | G. | F. P. | S. Afr. |
| Tephrosia macropoda Harv. | G. | r. r. | |
| Tephrosia nitens Benth. | | T' D | Tr. Am. |
| Tephrosia periculosa Baker. | G. | F. P. | |
| Tephrosia purpurea Pers. | G. M. R. L. | | Austr. |
| Tephrosia tomentosa Pers. | R. | [F. P. | Arab. |
| Tephrosia toxicaria Pers. | G. R. L. E. | | Jamaica |
| Tephrosia virginiana Pers. | G. | A. P. | N. Am. |
| Tephrosia Vogelii Hook. | G. R. | | Tr. Afr. |
| Teramus labialis Spreng. | G. | Sap. | Cosm. Trop. |
| Tetlapleura caerulea Willd. | Ğ. | Lup. | Tr. Afr. |
| Tetrapleura Thonningii Benth. | Ches. | Diur. | W. N. Am. |
| Thermopsis rhombifolia Rich. | L. | A | |
| | 7. | Tymp. | Egypt |
| Trifolium elegans Sair. | C. | Tymp. | Med. Reg. |
| Trifolium hybridum L. | | 3.6 | S. Eur. As.Min. |
| Trifolium incarnatum L. | G. | Mec. | S. Eur. |
| Trifolium nigrescens Viv. | C. | lan. | S. Eur. N. Temp. Reg. |
| Trifolium repens L. | C. | Tymp. | N. Temp. Reg. |
| Trigonella cretica Boiss. | G. | 1 | As. Min. |
| Vicia Ervilia Willd. | C. C. G. C. G. C. G. | | Eur. N. Afr. |
| Wistaria chinensis DC. | G. | | China |
| Xylia dolabriformis Benth. | G. | Sap. | Burma, Malay |
| LENTIBUI | ARIACEA | | |
| | | | IT Or |
| Pinguicula vulgaris L. | G. | | Eur. Sber. N. Am. |
| | CEAE | | |
| | | , | |
| Agrostocrinum stypandroides Ev. M. | G. M. | | Austr. |
| Allium ascalonicum L. | L. | Ant. | Cult. |
| Allium canadense L. | L. | Ant. | N. A. |
| Allium Cepa L. | L. | Ent. | Per. Bel. |
| Allium Douglasii Hook. | Ī. | Ant. | Calif. |
| | <u> </u> | 1 | Tourit. |

| NAME OF PLANT | Authority | Properties | Locality |
|---|--------------|------------|--------------------------|
| Allium Macleanii Baker | L. | Ant. | Afghan. |
| Allium Moly L. | L. | Ant. | Eur. |
| Allium Porrum L. | L. | Ant. | Eur. |
| | L. | Ir. | Eur. N. A. |
| Allium sativum L. Allium Schoenoprasum L. | L. | Ant. | Eur. As. |
| Allium tricoccum Soland. | | Ir. | N. A. |
| Allium ursinum L. | C. L. | Ant. | |
| Allium Victorialis L. | G. L. | Ant. | Eur. N. As. Eur. Sib. |
| Allium vineale L. | C. | Ir. | Eur. N. A. |
| Aloe abyssinica Lam. | L. | Pur. | Austr. |
| Aloe africana Mill. | Ĭ. | Pur. | S. Af. |
| Aloe arborescens Mill. | ĺ Ĩ. | Pur. | S. Af. |
| Aloe chinensis Steud. | _ Ĩ | Pur. | China |
| Aloe ferox. Mill. | G. L. | Pur. | S. Afr. |
| Aloe saponaria Haw. | G. | Pur. | S. Afr. |
| Aloe spicata L. | L. | Pur. | S. Afr. |
| Aloe succotrina Lam. | Ĩ. | Pur. | S. Afr. |
| Aloe tenuior Haw. | Ğ. | Pur. | Med. Reg. |
| Aloe vera L. | Ğ. | Pur. | Med. Reg. |
| Anemarrhena asphodeloides Bunge. | Ğ. | | China. |
| Asparagus officinalis L. | L. | Ir. | Eur. As. N. A. |
| Asparagus scaber Brign. | L. | Ir. | Med. Reg. |
| Tropuluguo ocubor 21.5 | | 1 | Cauc. |
| Astelia Banksii A. Cunn. | G. | İ | N. Z. |
| Bulbine bulbosa Haw. | G. M. | | Austr. |
| Bulbine semibarbata Haw. | G. | | Austr. |
| Chamaelirium carolinianum Willd. | G. L. | Ant. | E. N. A. |
| Colchicum autumnale L. | G.C.Sm.L. | 1 | Eur. |
| Colchicum neapolitanum Tenore. | C. | Ast. | Italy. |
| Colchicum speciosum Stw. | Sm. | Ast. | Cauc. Reg. |
| Colchicum variegatum L. | C. | Ast. | S. Eur. |
| Convallaria majalis L. | G.Sm.C.L. | | |
| | Ches.Rusby | Car. | N. Tem. Reg. |
| Dianella nemorosa Lam. | G. | | As. Trop. |
| | 1 | | Austr. |
| Dracaena arborea Hort. | G. | | Tr. Afr. |
| Drimia ciliaris Jacq. | G. | Em. | Agr. Austr. |
| Drimia Cowanii Ridl. | G. | | Madag. |
| Erythronium albidum Nutt. | L. | Em. | N. A. \ |
| Erythronium americanum Ker. Gawl. | L. | Em. | N. A. |
| Erythronium Dens-canis L. | L. G. | Em. Ant. | Eur. N. As. |
| Erythronium grandiflorum Pursh. | G. | j | N. Am. |
| Erythronium purpurascens S. Wat. | G. | Sap. | Cal. |
| Fritillaria Imperialis L. | G. C. Mi. | Ir. | Persia. Him. |
| | | j | Reg. |
| Fritillaria Meleagris L. | | Ir. | Eur. Cauc. |
| Fritillaria pudica Spreng. | L. | Ineb. | Reg. |
| Gloriosa simplex L. | G. | Nar. | W. N. Am. |
| Gloriosa superba L. | [<u>G</u> . | Em. | Tr. Afr. |
| Helonias frigida Lindl | G. | | Trop. As. |
| Hyacinthus orientalis L. | G. | | |
| Leucocoryne | M. | | Austr. |
| Leucocrinum montanum Nutt. | ! Ches. | | N. Am. Ry. |
| 26.1.1.1.1. | | 0 | Mts. |
| Medeola virginica L. | G. | Sap. | N. Am. |
| Melanthium cochinchinense Lour. | G | In. D . | Cochin China. |
| Melanthium virginicum L. | G. L. | Fly Poison | |
| Muscaria comosum Mill. | G. | Sap. | Med. Reg. Ori. |

| NAME OF PLANT | Authority | Properties | Locality |
|---|---|---------------------------------------|--|
| Muscaria racemosum Willd. | Sm. | Ir. | Med. Reg. |
| Narthecium ossifragum Huds. | G. C. | | Cauc. Eur. As. Min. |
| Nothoscordum striatum Kunth. | Ches. | | N. Am. N. Am. S. States |
| Ornithogalum sp. Ornithogalum nutans L. Ornithogalum thyrsoides Jacq. Ornithogalum umbellatum L. | Mi. G. | Em. | Eur. As. Afr. Eur. As. Min. S. Afr. N. Afr. As. |
| Paradisea Liliastrum Bertol. Paris obovata Ledeb. Paris quadrifolia L. | G. G. G. C. Mi. Sm. L. | Card. Ir. Nar. | Eur. Siber. Eur. N. As. |
| Polianthes tuberosa L. Polygonatum biflorum Ell. Polygonatum officinale All. Polygonatum verticillatum All. Ruscus aculeatus L. Schoenocaulon officinale A. Gr. Scilla bifolia L. Scilla festalis Salisb. Smilax pseudo-syphilitica Kunth. Stypandra glauca R. Br. Tofieldia calyculata Wahl. Trillium erectum L. Trillium grandiflorum Salisb. Tulipa Gesneriana L. Tulipa sylvestris L. Urginea Scilla Steinh. Veratrum sp. Veratrum album L. E. | Sm. L. G. G. G. G. G. J. G. Sm. J. Sm. G. G. M. G. L. Rusby G. G. G. G. G. G. C. Sm. R. L. C. Sm. R. L. | Ir. Ir. Sap. Ant. Em. Sap. Ast. F. P. | Mex. N. Am. Eur. Siber. Eur. N. As. Eur. Ori. Mex. Eur. As. Min. Eur. Braz. Austr. Eur. N. Am. E. N. Am. N. Am. Eur. Orient Eur. S. Afr. N. Temp. Reg. |
| Veratrum californicum Dur. | | Ir. | W. N. Am. Ry. Mts. |
| Veratrum viride Ait. | G. Ches. L. Rusby | Ir. Em. | E. N. Am. |
| Xanthorrhoea sp. Xerophylla Douglasii S. Wat. | G. | Sap. | Austr. E. N. Am. S. States. |
| Xerophylla setifolium Michx. Yucca filamentosa L. | G. G. | Sap. | E. N. Am. W. N. Am. Ry. |
| Yucca glauca (Nutt.) Carr. | G. | Sap. | Mts. E. N. Am. S. States. |
| Zygadenus angustifolium S. Wats. Zygadenus elegans Pursh. | L. Ches. L. | | N. A. E. N. Am. Ry. Mts. |
| Zygadenus venenosus S. Wat. | Ches. G. L. | | N. W. Am. |

LINACEAE

| Hugonia Mystax L. Linum catharticum L. Linum rigidum Pursh. | G. G. L. Ches. | Pur. Pur. | Ind. Eur. Orient. N. Am. N. M. Tex. |
|---|----------------------|--------------|--|
|---|----------------------|--------------|--|

| NAME OF PLANT | Authority | Properties | Locality | |
|---|---------------------|----------------|----------------------------------|--|
| Linum toxicum Boiss. Linum usitatissimum L. | G. L. | Ir. | Syria. Eur. Orient. | |
| LOGAN | IACEAE | <u> </u> | N. Am. | |
| Anthocleista grandiflora Gilg. | 1 | Alk. | 1 | |
| Buddleia brasiliensis Jacq. | G. | F. P. | Hab.? | |
| Buddleia curviflora Hook. & Arn. |] G. | | Chili. Peru. | |
| Buddleia globosa Hope. Buddleia Lindleyana Fort. | G. | Sap. | China. | |
| | G. G. | F. P. Taen. | Madag. | |
| Buddleia madagascariensis Lam. Buddleia polystachya Friesen. | R. G. | Sap. F. P. | Abyss. Brazil | |
| Buddleia variabilis Hemsl. | | Sap. | 1314211, | |
| Buddleia verticillata HBK. | | F. P. | Mex. | |
| Fagraea ceilanica Thumb. | Sm. G. | Cou. | Zeyl. | |
| Gelsemium sempervirens Ait. | G.C.Sm.L. | Ast. | E. N. Am. S. Sts. | |
| Nicodemia diversifolia Ten. | G. | Sap. | Mascar. Is. | |
| Spigelia Anthelmia L. | G. L. | Ir. Nar. | Trop. Am. | |
| Spigelia glabrata Mart. | G. |] | Brazil, | |
| Spigelia marylandica L. | G. Sm. L. | Ir. Nar. | E. N. Am. | |
| Spigelia pedunculata HBK. Strychnos angustifolia Benth. | G. G. | | N. Old World. China. | |
| Strychnos brachiata Ruiz. & Pav. | Ğ. | Con. | Peru. | |
| Strychnos Castelnaei Wedd. | G. Sm. L. | Con. | India. | |
| Strychnos colubrina L. | Ļ. | A. P. | Guiana | |
| Strychnos cogens Benth. | L. G. L. | A. P. A. P. | Braz. Guiana. | |
| Strychnos Crevauxiana Baill. Strychnos gubleri Planch. | L. | A. P. | Venez. | |
| Strychnos Icaja Baill. | G. L. | Ordeal Poi. | Trop. Afr. | |
| Strychnos Ignatii Berg. | G. Sm. L. | Con. | Philipp. Is. | |
| Strychnos innocua Dcl. | G. | | Nubia. | |
| Strychnos malaccensis Benth. | L. G. | | Bur. Malay. Guiana. | |
| Strychnos Melinoniana Baill. Strychnos Nux-Vomica L. | | Con. F. P. | Burma. | |
| Strychnos paniculata Champ. | G. | | China. | |
| Strychnos potatorum L. | G. | | Burma. | |
| Strychnos Pseudo-quina A. St. Hil. | G. G. | | Braz. | |
| Strychnos Santhierana Pierre. Strychnos spinosa Lam. | G. | | Madag. | |
| Strychnos Tieute Lesch. | G. Sm. L. | Con. A. P. | Java. | |
| Strychnos toxifera Schomb. | G Sm | Con. A. P. | Guian. | |
| Strychnos yapurensis Planch. | L. | A. P. | Ecuador. | |
| LORANT | HACEAE | | | |
| Phoradendron flavesceus Nutt. Viscum album L. | L. Hyams. Sm. L. | Ir. | N. Am. S. Sts. Eur. Temp. As. | |
| LYCOPODIACEAE | | | | |
| Lycopodium clavatum L. Lycopodium Selago | Mi, L. Mi, L. | Diur. Pur. | N. Am. Eur. Eur. | |
| LYTHR | ACEAE | | | |
| Ammannia baccifera L. | G. | Acr. | Tr. Old World | |
| Cuphea viscossissima Jacq. | G. | Card. | N. Am. S. Sts. | |
| Ginora mexicana Lam. | G. | Em. | Mex. | |

| NAME OF PLANT | Authority | Properties | Locality |
|--|-------------|---------------|---------------------------------|
| Lagerstroemia Flos-reginae Retz. | G. | Nar. | Trop. As. |
| Nesaea verticillata HBK. Punica Granatum L. | G. G. L. | Taen. | N. A. S. Am. S. Eur. Maurit. |
| | | Tacii. | O. Eur. Maurit. |
| MAGNOI | JIACEAE | | |
| Drimys aromatica F. Muell. | G. | Hyd. | Austr. |
| Drimys Winteri Forst. Illicium anisatum Gaertn. | G. Sm. L. | Hyd. | S. Am. China. |
| Illicium floridanum Ellis. | G. Ches. L. | | N. Am. S. Sts. |
| Liriodendron chinense Sarg. | G. | | China. |
| Liriodendron tulipifera L. | G. G. | Hyd. Sap. | E. N. Am. N. Am. S. Sts. |
| Magnolia grandiflora L. Manglietia glauca BL. | G. | | Java. |
| Michelia Champaca L. | G. | Alk. | Malay. |
| Michelia fuscata Bl. | G. | Alk. | China. |
| Michelia parvifolia Bl. Talauma macrocarpa Zucc. | G. G. | Alk. | Java. Mex. |
| Talauma ovata A. St. Hil. | G. | | Braz. |
| MALPIGE | HIACEAE | | |
| Byrsonima amazonica Griseb. | G. | | Braz. |
| Byrsonima crassifolia HBK. | G. | | Guiana. |
| Byrsonima spicata Rich. | G. | | Thop. Am. |
| Heteropteris syringaefolia Griseb. Malpighia oxycocca Griseb. | G. G. | | Braz. W. Ind. |
| Sphedamnocarpus angolensis Planch. | Ğ. | | Trop. Afr. |
| MALVA | CEAE | | |
| Althaea rosea L. | G. | Abort. | Ori. Cult. Eur. N. Am. |
| Gossypium herbaceum L. | G. L. | Abort. | Trop. As. |
| Hibiscus diversifolius Jacq. | G. | Abort. | Tr. Old World. |
| Pavonia zeylanica Sida jamaicensis L. | G. G. | Ant. | Тгор. Ат. |
| Sida paniculata L. | G. | Mech. | Trop. Am. |
| Sida urens L. | G. | Ir. | Trop. Am. Afr. |
| MELASTO | MACEAE | | |
| Mouriria rhizophoraefolia Gard. | G. | | Marting. Is. |
| MELIA | CEAE | | |
| Carpa moluccensis Lam. | , G. | | Tr. Old World. |
| Dysoxylum arborescens Miq. Flindersia Schottiana Fv. Muell. | G. | | Borneo. |
| Lansium domesticum Jack. | G. G. | Ant. | Austr. Malay. |
| Melia Azeldarach L. | G. C. | Ant. | Himal. Ch. |
| | Ches. L. | | Cult. S. States |
| Melia birmanica Kurz. | G. G. L. | Em. | Burma. |
| Naregamia alata W. & A. Ptaeroxylon obliquum Rdlk. | G. L. | Elli. | Ind. Afr. |
| Sandoricum indicum Cav. | G. | | Trop. As. |
| Swietinia humilis Zucc. | G. | | Mex. |
| Swietinia Mahagoni Jacq. | L. G. | Astr. | S.A. Mex. W.I. |
| Trichilia sp. Trichilia Moschata Sw. | L. | Sap. Astr. | S. Am. Jamaica. |
| Trichilia trifolia L. | G. | Abort. | Venz. |
| Walsura piscidia Roxb. G. & E. | R. E. | F. P. | India. |

MELIANTHACEAE

| MELIANTHACEAE | | | |
|---|---|---|---|
| NAME OF PLANT | Authority | Properties | Locality |
| Bersama sp. Melianthus comosus Vahl. Melianthus major L. | G. G. G. | | Arab. & Abyss. S. Afr. S. Afr. |
| MENISPE | RMACEAE | , | |
| Anamirta paniculata Colebr. | | Del. F. P. | Ind. |
| Anomospermum japurense Eich. Cissampelos Pareira L. Cocculus Ferrandianus Gaudich. Cocculus glaucescens Bl. Cocculus laurifolius DC, Cocculus toxiferus Wedd. Coscinium Blumeanum Miers. Menispermum canadense L. Pachygone ovata Miers. Pericampylus incarius Miers. Sarcopetalum Harveyanum Fv. M. Stephania discolor Spreng. Stephania aculeata Walp. Tiliacora racemosa Colebr. | R. L. E. G. C. G. G. G. G. G. G. G. G. | F. P. Sap. Sap. Diur. F. P. | Braz. Trop. Reg. Hawai, Is. Bur. Jav. Him. Reg. Jap. Ind. N. A. Malay. Ind. Ind. Malay. Austr. Tr. Old World. Ind. Jav. |
| MONIM | IACEAE | | 1 |
| Kibara angustifolia | I Sm. | ı | 1 |
| MONOTRO | | · | 1 |
| Monotropa uniflora L. | | Androna | E. N. Am. Jap. |
| Pterospora andromedea Nutt. | G. | , and the same | Him. Reg. N. Am. |
| MORA | CEAE | | |
| Morus alba L. Morus nigra L. Morus rubra L. | L,. L,. L,. | Taen. Taen. Taen. | Temp. As. Temp. As. N. A. |
| МУОРО | RACEAE | | |
| Eremophila maculata Fv. M. Myoporum deserti A. Cunn. Pholidia maculata Baill. | M. G. G. M. G. | | Austr. Austr. |
| MYRIC | ACEAE | | |
| Myrica cerifera | L. | Astr. Nar. | N. A. |
| MYRISTI | | | |
| Myristica gibbosa Hook. Myristica philippensis Lam. | L. | Astr. Astr. | India. Phil. Is. |
| MYRSI | NACEAE | | |
| Aegiceras majus Gaertn, Aegiceras minus Gaertn, Anguillaria dioica R. Br. Clavija macrocarpa Ruiz, & Pav, | G. E. R. G. R. M. G. | Sap. F. P. | Tr. Old World. India. Aust. S. Am. |

| NAME OF PLANT | Authority | Properties | Locality |
|---|-------------------|----------------|----------------------------|
| Embelia micrantha A. DC. Embelia Ribes Burm. | G. G. | Ant. | Maurit. Is. Trop. As. |
| Jacquinia arborea Vahl. | G. R. G. E. R. | F. P. F. P. | Trop. Am. |
| Jacquinia armillaris Jacq. Jacquinia obovata Schrad. | G. E. R. | F. P. | W. Ind. |
| Maesa indica Wall. | G. | F. P. | Ind. Malay. |
| Maesa lanceolata Forsk. | G. G. | Taen. | N. Afr. |
| Myrsine africana L. | G. | l aen. | Him. Reg. Tr. |
| | | | Azores W. Ind. |
| Theophrasta americana L. | G. | | |
| MYRTA | ACEAE | | |
| Baeckea frutescens L. | G. | Abort. | Malay. China. |
| Barringtonia acutangulata Gaertn. Barringtonia alba Blume. | G. G. | | Malacca Is. Malacca Is. |
| Barringtonia Butonica Forst. | G. R. | F. P. | Pacific Is. |
| Barringtonia Careya Fv. M. | G. | | Austr. |
| Barringtonia insignis Miq. | G. | T D | Malay. |
| Barringtonia intermedia Viell. Barringtonia neo-caledonica Viell. | G. | F. P. | N. Caled. N. Caled. |
| Barringtonia racemosa Rexb. | | F. P. | Malay, Polyn, |
| Eucalyptus Globulus Labill. | L. | Astr. | Austr. |
| Eucalyptus microtheca F.v. Muell. | Ģ. | F. P. | Austr. |
| Eucalyptus rostrata Schlecht. Eugenia Jambos L. | L. G. | Astr. | Austr. Trop. As. |
| Lecythis amara Aubl. | G. | | Guian. |
| Lecythis lanceolata Poir. | G. | Nar. | Brazil. |
| Melaleuca leucodendron L. | Sm. L. | | Austr. |
| Napoleona Whitfieldii Van Houtte Pimenta acris Kostel. | G. Whi. | Sap. Ir. | Trop. Afr. Ind. |
| Psidium montanum Sw. | G. | | W. Ind. |
| MUSA | CEAE | | |
| Musa sapientum L. | G. | Sap. | Trop. Reg. |
| Nepenthes gracilis Korth. | G. | ļ | Borneo. |
| NYCTAG | INACEAE | | |
| Boerhaavia erecta L. | G. | Em. | N. Am. W. Ind. |
| Boerhaavia hirsuta L. | G. G. | Em. Em. | N. Am. W. Ind. |
| Boerhaavia repens L. Pisonia obtusata Jacq. | G. | Em. | Cosm. Trop. W. Ind. |
| Pisonia tomentosa Casar. | Ğ. | Em. | Braz. |
| NУМРНАІ | EACEAE | | |
| Euryale ferox Salisb. | Sm. | | Ind. China. |
| Nymphaea advena Ait. | L. | Astr. | N. A. |
| Nymphaea alba L. | G. L. | Ante | N. Temp. Reg. |
| Nymphaea lutea L. | | Astr. | Eur. |
| OCHNA | | | |
| Ochna sp. Schreb. | G. | | Afr. Ind. |
| OLAC | | | |
| Villaresia Moorei F. v. M. | G. | | Austr. |

OLEACEAE

| OLEACEAE | | | |
|--|--|--|---|
| NAME OF PLANT | Authority | Properties | Locality |
| Chionanthus picrophloia Fv. Muell. Chionanthus virginica L. Forsythia intermedia Zabel. Forsythia suspensa Vahl. I-orsythia viridissima Lindl. Jasminum floribundum R. Br. Jasminum officinale L. Jasminum Sambac Ait. Ligustrum vulgare L. Olea dioica Roxb. Phillyrea media L. | G. G. G. G. G. G. Sm. G. Sm. G. Sm. Ches. G. G. G. Sm. Ches. G. G. G. G. G. G. | Nar. Sap. Sap. Ast. Ir. Em. Sap. | N. Am. Japan, China. China. Abyss. Trop. As. Eur. As. N.Am. Ind. Med. Reg. |
| ONAGR | ACEAE | | |
| Ludwigia erigata L. Montinia caryophyllacea Thunb. Trapa natans L. | G. | | Ind. S. Afr. Ori. Ind. Eur. |
| ORCHIL | DACEAE | | |
| Angraecum fragans Thou. Catasetum sp. Cymbidium aloifolium Sw. Cypripedium pubescens Willd. Cypripedium spectabile Salisb. Dendrobium nobile Lindl. Eria stellata Lindl. Eulophia virens Spreng. Habenaria nigra R. Br. Neottia Nidus-avis Rich. Orchis coriophora L. Orchis coriophora L. Orchis purpurea Huds. Orchis Simia Lam. Phajus callosus Lindl. Phalaenopsis amabilis Blume. Phalaenopsis Lueddemanniana Reichb. | G. L. G. | Ant. Ir. Ir. Ant. Ant. Cou. Cou. Cou. Cou. | Mascar Is. Ind. Ind. N. Am. N. Am. China. Philipp. Is. Ind. Eur. Eur. N. As. Eur. Orient. Eur. As. Min. Eur. Ori. Malay. Philipp. Is. |
| | DACEAE | | |
| Oxalis Acetosella L. Oxalis amara A, St. Hill. Oxalis Pes-caprae L. Oxalis purpurata Jacq. Oxalis Smithiana Eckl. & Zeyr. | G. L. G. | Ant. Diur. | Eur. As. N. Afr. N. A. Braz. S. Afr. S. Afr. S. Afr. |
| PALM | ACEAE | | |
| Areca Catechu I., var. nigra. Arenga saccharifera Labill. Borassus flabellifera Murr. Cocos amara Jacq. Cocos nucifera I., Corypha umbraculifera I., Hyphaene thebaica Mart. Hyphorbe indica Gaertn. Phoenix dactylifera | G. L., G. G. G. G. G. G. G. G. G. G. G. G. G. | Ver. Taen. | Ind. Malay. Malay. Ind. W. Ind. Trop. Afr. Ind. Trop. Afr. |

| PANDANACEAE | | | |
|---|---|--|---|
| NAME OF PLANT | Authority | Properties | Locality |
| Pandanus odoratissimus L. Pandanus Thomensis Henrig. | G. G. | | Ind. Arab. |
| PAPAVE | RACEAE | | |
| Argemone alba Lestib. Argemone intermedia Sw. Argemone mexicana L. | L. L. Sm.Ches.L. | Nar. Nar. Nar. | S. U. S. Mex. N. Am. Tex. Mex. |
| Bocconia cordata Willd. Bocconia frutescens L. Chelidonium majus L. | L. G. L. C. Rusby G. Sm. Ches. Mi. | Ant. Ant. Ir. Nar. | China. Japan. Mex. Peru. Eur. As. N.Am. |
| Eschscholtzia californica Cham. Glaucium corniculatum Curb. Glaucium flavum Crantz. Hypecoum procumbens L. | G. L. C. Sm. L. G. | Pur. Ir. | Calif. Eur. Eur. Med. Reg. Med. Reg. Arabia. |
| Papaver aculeatum Thumb. Papaver Rhoeas L. | G. G. Sm. Ches. C. L. | Ir. | S. Afr. Austr. Eur. Ori. Afr. |
| Papaver somniferum L. Roemeria violacea Medic. · Sanguinaria canadensis L. Stylophorum diphyllum Nutt. | C. Mi. Ches. L. | Nar. Ir. | Eur. As. Ind. N. Am. Eur. N. Afr. N. Am. N. Am. |
| PASSIFLO | ORACEAE | | |
| Andenia lobata Engl. Carica Papaya L. Carica quercifolia H. St. Hil. Carica spinosa Aubl. Modecca palmata Lam. Modecca trilobata Roxb. Modecca venenata Forsk. Ophiocaulon gummifera Han. Passiflora caerulea L. Passiflora foetida Passiflora Herbertiana Ker-Gawl. Passiflora hispida DC. Passiflora laurifolia L. Passiflora quadrangularis L. Passiflora rubra L. | G. G. G. G. G. G. G. G. G. L. G. L. G. L. G. G. G. C. | Sap. Em. Antisp. Nar. Antisp. Hyd. Nar. Nar. | Trop. Afr. Trop. Amr. Braz. Guiana. Ind. Ind. Trop. Afr. Brazil, Austr. Jamaica. Trop. Amer. Trop. Amer. Trop. Amer. Trop. Amer. W. Ind. |
| PHYTOL | ACCACEA | E | |
| Anisomeria drastica Mey. Gallesia Scorodendron Petiveria alliacea L. Petiveria tetrandra Gomez. Phytolacca abyssinica Hoffm. Phytolacca acinosa Roxb. Phytolacca decandra L. | G. G. L. G. G. G. G. C. L. Ches.Rusby | Ant. Abort. F. P. Sap. Del. Sap. Nar. Em. / Sap. | W. Ind. Brazil. Trop. & S. Afr. Him.Reg. Chin. N. Amer. |

| NAME OF PLANT | Authority | Properties | Locality |
|--|----------------|------------------|---|
| Phytolacca dioica | G. | Sap. | S. Amer. |
| Phytolacca icosandra L. | G. | Sap. | Ind. |
| Pircunia sp. (Phytolacca) | [G. | Sap. | S. Am. |
| Rivina humilis | [G. | 1 | Trop. Am. |
| PIPERA | ACEAE | | |
| Piper Carpunya Ruiz. & Pav. | G. L. | | India. |
| Piper darienense C. DC. | L. R. E. | Sial. | Chili & Peru. |
| Piper methysticum Forst. | R E. | F. P. | Panama |
| Piper nigrum L. | L. | Ineb. | Pacific Is. |
| Piper Palmeri C. DC. | G. | Sap. | /m A |
| Piper peltatum L. | L. | Diur. | Trop. Am. |
| Piper umbellatum L. | L, | Diur. | Cosm. Tr. Reg. |
| PIROL | ACEAE | | |
| Chimaphila maculata Pursh. | G. L. | Astr. | N. Am. |
| Chimaphila umbellata Nutt. | L. | Astr. | Eur. N. As. |
| | _ | | N. A. |
| Monotropa uniflora | Sm. | | Him. Reg. Japan. N. A. |
| D 1 11 2 | _ | 1. | Japan. N. A. |
| Pyrola chlorantha Sw. | Ľ. | Astr. | Eur. N. A. |
| Pyrola elliptica Nutt. | L. | Astr. | N. A. |
| Pyrola rotundifolia L. | G. L. | Em. | N. A. N. Temp. Reg. N. Temp. Reg. |
| Pyrola minor L. | L. | Astr. | IN. Temp. Reg. |
| PITTOSPO | DRACEAE | | |
| Billardiera longiflora Labill. | | Sap. | 1 |
| Pittosporum coriaceum Ait. | G. | Sap. | Madeira Is. |
| Pittosporum cornifolium A. Cunn. | G. | Sap. | N. Zeal. N. Zeal. |
| Pittosporum crassifolium Soland. | G. | Sap. | N. Zeal. |
| Pittosporum eugenioides A. Cunn. | G. | Sap. | N. Zeal. |
| Pittosporum ferrugineum Ait. | | Sap. | Malay. Austr. |
| Pittosporum floribundum W. & A. | G. |] - | Ind. |
| Pittosporum Huttonianum T. Kirk. | G, | Sap. | N. Zeal. |
| Pittosporum javanicum Bl. Pittosporum Moorei F. v. Muell. | G. | F. P. | Java. |
| Pittosporum Moorei F. v. Muell. | G. | Sap. | |
| Pittosporum phillyraeoides DC. | G. | Sap. | Austr. |
| Pittosporum rhombifolium A. Cunn. | |) |] |
| Pittosporum tobira Ait. | G. | | Japan. China. |
| Pittosporum undulatum Vent. | G. | Sap. | Austr. |
| PLANTAG | INACEAE | | |
| PLATAN | | | |
| Platanus occidentalis L. | G. | Hyd. | N. Am. |
| Platanus orientalis L. | G. | Hyd. | S. Eur. Orient. |
| PLUMBAG | INACEAE | | |
| Armeria elongata Hoffm. | L. | Astr. | Eur. |
| Plumbago capensis Thunb. | т (| Em. | S. Eur. Caucas. |
| Plumbago europaea L. | L. | Em. Nar. Acr. | Ind. |
| Plumbago rosea L. Plumbago scandens L. | G. L. G. L. | Pur. | |
| | G. L. | i ui. | Trop. Am. Trop. Afr. |
| Plumbago toxicaria Bertol. Plumbago zeylanica L. | G. | Abort. Ir. | Tr. Old World. |
| | G. | F. P. | |
| Statice pectinata Ait. | U, | A . I . | Canary Is. |

POLEMONIACEAE

| T ODD MONITORING | | | |
|--|-----------|------------|----------------------------|
| NAME OF PLANT | Authority | Properties | Locality |
| Cantua buxifolia Lam. | G. | i | Peru |
| Cantua pyrifolia Juss. | Ğ. | | Peru |
| Cobaea scandens Cav. | G. G. | Sap. | Mex. Ry. Mts. |
| | | | N. Am. |
| Gilia achillaefolia Benth. | G. | Sap. | Calif. |
| Gilia aggregata Spreng. | G. | Sap. | N. A. Ry. Mts. |
| Gilia laciniata Ruiz. & Pav. | G. | Sap. | Peru. Chili. |
| Loeselia caerulea G. Don. | G. L. | Em. | Mex. |
| Phlox L. | G. | | N. Am. |
| Polemonium boreale Adams. | G. | Sap | Siberia. |
| Polemonium caeruleum L. | L. | Sap. | N. Temp. Reg. |
| Polemonium flavum Greene | IG. | San. | N. A. N. Mex. |
| Polemonium gracile Willd. Polemonium humile Willd. | G. | Sap. | |
| Polemonium humile Willd. | G. | Sap. | Arct. Reg. |
| Polemonium panciflorum Ser. Wat. | | | W. N. Am. |
| Polemonium reptans L. | G. L. | Sap. | W. N. Am. N. Am. |
| | LACTATE | · | |
| POLYGA | | | 15 |
| Krameria triandra R. & Pav. | L. | Ast. | Peru |
| Monnina sp. R. & P. | G. | Sap. | Trop. Am. |
| Polygala alba Nutt. | L. | Ir. | N. A. |
| Polygala amara L. Polygala angulata DC. | G. | J | Eur. |
| Polygala angulata DC. | G. | Sap. | Brazil. |
| Polygala aspalatha L. | G. | <u></u> | Braz. |
| Polygala Boykini Nutt. | L. | Ir. | N. A. |
| Polygala Cyparissias A. St. Hil. Polygala glandulosa HBK. | G. | P. | J |
| Polygala glandulosa HBK, | G. | | Mexico. |
| Polygala Javana DC. | G. | Sap. Ins. | Malaga. |
| Polygala Senega L. | G. Sm. L. | | N. Am. |
| Securidaca longepeduculata Fres. | G. | P. | Abyss. |
| POLYGO | NACEAE | | |
| Fagopyrum esculentum Moench. | C. | Ir. | Eur. N. As. |
| Polygonum acre HBK. | R. L. E. | Ir. F. P. | Cosm. |
| Polygonum barbatum L. | G. | F. P. | Tr. Old World. |
| Polygonum Bistorta L. | C. L. | Astr. | N. Reg. N. Temp. Reg. |
| Polygonum Convolvulus L. | C. | | N. Temp. Reg. |
| Polygonum flaccidum Meiss. | G. | Ir. | Ind. Malay. |
| Polygonum Hydropiper L. | G. C. L. | Ir. | Temp. Reg. N. |
| | | | & S. |
| Polygonum hydropiperoides Michx. | G. | Ir. | N. Am. |
| Polygonum orientale L. | G. | Ir | Tr. Old World. |
| Rheum Emodi Wall. | L. | Pur. | Him. Reg. |
| Rheum hybridum Murray | L. | Pur. | Mongolia |
| Rheum officinale Baill. | L, | Pur. | Thibet. |
| Rheum palmatum L. | Sm. | Pur. | Mong. |
| Rheum Rhaponticum Delarb. | L. | Pur. Astr. | Siberia. |
| | Newspaper | | |
| | Death in | | |
| D 1 1 1 - | Iowa | | |
| Rumex abyssinicus Jacq. | G. | Taen. | Abyss. |
| Rumex Acetosa L. | L | Astr. | Eur. N. As. Eur. N. Am. |
| Rumex Acetosella | C. L. | | Eur. N. Am. |
| Rumex Crispus | _ | Oxal. Acid | Eur. N. Am. |
| Rumex Ecklonianus Meiss. | G. ! | Taen. | S. Afr. W. & S. W. N. |
| Rumex hymenosepalus Torr. | G. L. | Astr. | |
| Dumov obtacifolias T | T . | A -4- | Am. |
| Rumex obtusifolius L. | L. | Astr. | Eur. As. N. A. |

| PORTULACEAE | | | |
|---|---|----------------------|--|
| NAME OF PLANT | Authority | Properties | Locality |
| Lewisia rediviva Pursh. Talinum polyandrum Ruiz. & Pav. | G. G. | Acr. | W.N.Am. Mont. Peru. |
| PRIMUI | ACEAE | | |
| Anagallis arvensis L. | G. C. Ches. Mi. L. | Sap. | Eur. Temp. As. |
| Cortusa Matthioli L. Cyclamen europaeum L. | C. | Sap. F. P. | N. As. Eur. Cauc. Reg. |
| Cyclamen graecum Link. Cyclamen hederaefolium Ait. Cyclamen latifolium Sibth. & Sm. Lysimachia Nummularia L. | R, G, G, L, | F. P. Astr. | Greece. S. Eur. Greece, A. Min. Eur. |
| Primula Auricula L. Primula obconica Hance. Primula Parryii A. Gray. Primula reticulata Wall. Primula sinensis | G. G. L. Panı. G. Sm. | Ir. | Eur. China. Ry. Mts. N. A. Him. Reg. China. |
| | ACEAE | Ir. | , ciiiia. |
| Brabejun L. Grevillea mimosoides R. Br. | G. | I-Iyd. | S. Afr. Austr. |
| Helicia robusta Wall, | Ğ. | Tlyd. | Him. Reg. Bur- ma. Malay. |
| Knightia excelsa R. Br. Macadamia ternifolia F. Muell. Protea cynaroides L. Roupala Pohlii Meiss. Roupala vervaineana Hort. Xylomelum pyriforme Knight. | G. G. G. G. | Sap. Hvd. Hyd. | N. Zeal. Austr. S. Afr. Brazil. Austr. Austr. |
| RANUNCI | JLACEAE | | |
| Aconitum Anthora L. Aconitum chinense Sieb. Aconitum ferox Wall. Aconitum columbianum Nutt. | C. L. L. C. Sm. L. Ches. | F. P. Ast. | N. As. Japan. Him. Reg. W. N. Am. Ry. Mts. |
| Aconitum Fischeri Reich. Aconitum heterophyllum Wall. Aconitum japonicum Decne. Aconitum Lycoctonum L. Aconitum Napellus L. | L. C. L. L. Mi.C.G.L. Mi. Rusby C.Sm. Wh. | Ast. | N. As. N. Am. Him., Reg. Japan. Eur. N. As. N. Temp. Reg, Eur. |
| Aconitum noveboracense Gray. Aconitum uncinatum L. Actaea alba (L.) Mill. | L. Rusby | Ast. Em. | E. N. Am. N. A. Japan. N. A. |
| Actaea rubra (Ait.) Willd. Actaea spicata L. Adonis Dill. sp. Adonis aestivalis L. Adonis amurensis Regel & Radde. Adonis vernalis Anemone altaica Fisch. Anemone apennina L. | Ches. Rusby Wh. G. Sm. Mi. G. G. C. Sm. L. G. Mi. | Ir. Ir. | N. Temp. Reg. N. A. Eur. As. Eur. Ori. Manchuria. Eur. N. Am. Siber. Altai. N. Am. |

| NAME OF PLANT | Authority | Properties | Locality |
|--|-------------------|------------------|---------------------------------|
| Anemone coronaria | L, | Ir. | Med. Reg. Ori. |
| Anemone nemorosa L. | Sm. C. G. | Ir. | Eur. N. As. N. |
| Anemone parviflora Michx. | Mi. L. | | N. Am. |
| Anemone patens L. var Wolfgangiana | | Abort. Ir. | N. Cent. St. Ry. |
| Koch. | | | Mts. Br. Am. |
| Anemone Pulsatilla L. | G.C.Mi.Sm. | About To | Eur. |
| Anemone ranunculoides L. | | Ir. | Eur. Can. |
| | | | Siberia. |
| Anemone sylvestris L. | L. | Ir. | Eur. Cauc. Reg. |
| Aquilegia caerulea James. Aquilegia canadensis L. | | Diur. Diur. | W. N. Am. N. A. |
| Aquilegia vulgaris L. | G.C.Mi.L. | | Eur. Ori. |
| Caltha arctica R. Br. | Sm. | Pur. | Arct. Reg. |
| Caltha palustris L. | G. C. Sm. | Acr. | N. Temp. & |
| Cimicifuga foetida L. | C. Sm. | Acr. | Arct. Reg. Eur. Siberia. |
| Cimicifuga racemosa L. | G. Sm. L. | Ast. | N. Am. |
| Clausel's educate falls ///www. | Rusby | | 26 |
| Clematis aethusiaefolia Turcz. Clematis alpina Mill. | G. C. Sm. | Acr. | Mong. Eur. N. Am. |
| Clematis Bergeroni A. Lavall. | G. | Sap. | 1341. 11, 7111. |
| Clematis brachiata Thunb. | G. | Vcr. | S. Afr. |
| Clematis Buchaniana DC. Clematis caripensis HBK. | G. G. | Sap. | Him. Reg. Mex. |
| Clematis cirrhosa L. | G. | Ves. Sap. | Med. Reg. |
| Clematis Flammula L. | C. G. | Sap. | Med. Reg. |
| Clematis florida Thunb. | G. | Ir. Sap. | Japan. |
| Clematis Fremonti S. Wat. Clematis Gouriana Roxb. | G. G. | Sap. Hyd. Ir. | W. N. Am. Ind. Malay. |
| Clematis integrifolia L. | G. C. | Hyd. | S. Eur. N. As. |
| Clematis lanuginosa Lindl. | G. | Ir. Hyd. | China. W. N. |
| Clematis mauritiana L. | G. | | Amer. Ry. Mts. Madag. Maurit |
| Ciematis mauritana 14. | | ł | Is. |
| Clematis orientalis L. | G. | Hyd. | Him. Reg. Ori. |
| Clematis Pitcheri Torr. & Gray. Clematis pseudo-flammula Sch. | G. G. | Sap. Sap. | E. N. Am. |
| Clematis reticulata Walt. | 1 | Sap. | N. Am. |
| Clematis Vitalba L. | G.Sm.C.Mi. | Ir. | Eur. N. Afr. |
| Clematis Wightiana Wall. | L. | Lind | Cauc. Ind. |
| Clematis Williamsii Gray. | G. Sm. | Hyd. | Japan. |
| Delphinium Ajacis L. | M. G. L. | Ir. | Eur. |
| Delphinium Brunonianum Royle | G. | Card. | Him. Reg. |
| Delphinium caeruleum Jacq. | G. | | Him. Reg. N. |
| Delphinium Consolida L. | C. Mi. L. | Ast. | Eur. N. As. |
| Delphinium Geyeri Greene | Ches. L. | Acr. | N. A. Ry. Mts. |
| Delphinium Menziesii DC. | Ches. G. L. Ches. | Car | W. N. Am. |
| Delphinium mauritiana Coss. | G. | | N. Afr. |
| Delphinium peregrinum L. | G. | | S. Eur. Ori. |
| Delphinium recurvatum Greene | Ches. L. | Acr. | W. N. Am. |
| Delphinium Requieni DC. | C. | | Med. Reg. |
| Delphinium scopulorum A. Gray. | L. Ches. | | N. Am. |

| ************************************** | | | |
|---|---------------------|------------|------------------------|
| NAME OF PLANT | Authority | Properties | Locality |
| Delphinium Staphisagria L. | G.Sm.W.C. | Ast.Paras. | Med. Reg. |
| Delphinium tricorne Michx. | G. L. Ches. | Car. | E. N. A. S. Sts. |
| Delphinium trolliifolium Gray. | G. Ches. G. L. | Ir. | W. N. Am. Him. Reg. |
| Delphinium uncinatum Hook. Eranthis hyemalis Salisb. | | | Fun. Reg. |
| Helleborus foetidus L. | Sm. G. C.G.Sm.L. | Pur | Eur. |
| Helleborus niger L. | IG.W.C.Sm. | Pur. | Eur. |
| Helleborus odorus Waldst. | L. Mi. | | Hungary, |
| Helleborus orientalis Lam. | C. Sm. | | A. Min. Greece. |
| Helleborus viridis L. | G. Ches. L. | Pur. | Eu. |
| Hydrastis canadensis L. | L, | Berb. | N. A. |
| Isopyrum fumarioides L. | G. | Hyd. | Eur. N. As. |
| Isopyrum thalictroides L. | G. | | S. Eur. |
| Knowltonia vesicatoria Sims. | G. L. | Acr. Ves. | S. Afr. Cape. Reg. |
| Nigella damascena L. | L | Sap. | Med. Reg. |
| Nigella sativa L. | G. L. | Sap. | Med. Reg. |
| Ranunculus abortivus L. | 000 10 | 1r. | E. N. Am. |
| Ranunculus acris L. | C.G.Sm.Mi. | Acr. | Eur. N. As. |
| Ranunculus alpestris L. | | Acr. | Eur. |
| Ranunculus aquatilis L. | Mi. Sm. | Ir. | Tem. Reg. |
| Ranunculus arvensis L. | G.Sm.C.L. | Acr. | Eur. N. As. |
| Ranunculus, asiaticus L. | | Ir. | Asia. |
| Ranunculus auricomus L. | Mi. Sm. L. | lr. | N. Temp. Reg. |
| Ranunculus bulbosus L. | G.C.Mi.Sm. | lr. | Temp. Reg. |
| Ranunculus Ficaria L. | G.C.Sm.Mi. | lr. | Eur. Caucas. |
| Ranunculus Flammula L. | G.C.Sm.Mi. Rusby | 11. | N. Temp. Reg. |
| Ranunculus hybridus | G. C. Mi. | ī. | S. Eur. |
| Ranunculus lanuginosus L. | Ğ. | | Eur. Cau. Reg. |
| Ranunculus lappaceus | Sm. M. | P. ' | Austr. |
| Ranunculus lingua L. | C. Mi. | | Eur. Siber, |
| Ranunculus polyanthemos L. | Mi. | | Cauc. Reg. |
| Ranunculus repens L. | C. Mi. | | N. Temp Reg. |
| Ranunculus sceleratus L. | G. Mi. Sm. | | Eu. N. Am. |
| Denunculus contentrionalis Dais | RusbyChes. Ir. | Acr. | E. N. Am. |
| Ranunculus septentrionalis Poir. Ranunculus Thora L. | | Ves. | S. Eur. |
| realiuneurus Thora Ly. | Rusby | , | 2, 2,411 |
| Thalictrum flavum L. | C. | | Eur. N. As. |
| Thalictrum foetidum L. | Sm. | | Eur. Siberia |
| Thalictrum macrocarpum Gren. | C. G. | | |
| Trollius asiaticus L. | G. | Sap. | Siberia. |
| Trollius europaeus L. | G. | Sap. | Eur. Cancas. |
| Trollius pumilus D. Don. | G. | Sap. | Him, Reg. |
| Zanthoriza | M. | | Austr. |
| RESED | | | |
| Reseda Tourn. sp. | G. | Hyd. | Eur. |
| Reseda Luteola L. RHAMN | L. | Diur. | Eur. |
| | | 0 | |
| Ceanothus americanus L. | G. L. | Sap. | E. N. Am. |
| Ceanothus azureus Desf. Ceanothus caeruleus Lag. | G. L. | | Mex. Mex. |
| Transitius Cacituicus Lag. | 14. | | MICA. |

| NAME OF PLANT | Authority | Properties | Locality |
|---|----------------|--------------|-------------------|
| Ceanothus integerrimus Hook & Arn. | | Sap. | N. W. Am. |
| Ceanothus ovatus Desf. | G. | Sap. | N. Am. |
| Ceanothus thyrsiflorus Eschw. | G. | Sap. | Calif. |
| Ceanothus velutinus Dougl. | G. L. | Sap. | N. W. Am. Ry. |
| Callatia asimasa Tam | G. | C | Mts. S. Am. |
| Colletia spinosa Lam. | G. | Sap. | Chili. Pat. |
| Discaria serratifolia Benth. & Hook. Gouania Jacq. sp. | G. | Sap. | Cilli. Pat. |
| Karwinskia Humboldtiana Zucc. | G. Coult. | | W. N. Am. Tex. |
| ital williskia Tiulliborditalia Edec. | (Mtchell) | | W. IV. IIII. ICX. |
| Paliurus aculeatus Lam. | G. L. | | S. Eur. W. As. |
| Rhamnus Alaternus L. | C. | | Med. Reg. As. |
| · · | | | Minor. |
| Rhamnus californica Esch. | G. | Ir. | As. Min. |
| Rhamnus caroliniana Walt. | | Lax. | N. A. |
| Rhamnus cathartica L. | C. Sm. L. | Ir. | Eur. As. & N. |
| D. 75 | 0 0 7 | _ | Afr. |
| Rhamnus Frangula L. | G. C. L. | Lax. | Asia. N. Afr. |
| Rhamnus Purshiana DC. | т | T | N. A. |
| | L/. | Lax. | Ind. |
| Rhamnus Wightii W. & A. Zizyphus Joazeira Mart. | L. L. G. | Lax, Sap. | Brazil. |
| Zizyphus Lotus Lam. | G. | ισαμ. } | Med. Reg. |
| Zizyphus sativa Gaertn. | G. | | Med. Reg. |
| Dizyphus sativa Gaerth. | J. | | Temp. As. |
| | I | } | 1pi - 201 |

RHIZOPHORACEAE

| G. Sm. L. | Tan. | Tr. Shores. | | |
|-------------|---|---|--|--|
| ROSACEAE | | | | |
| G. L. | Ant. | N. Tem. | | |
| G. L. | Ant. | Abyss. | | |
| | Sap. | Ry. Mts. W. | | |
| | | N. Am. | | |
| G. | Hyd. | | | |
| G. | Hyd. | N. Temp. Reg. | | |
| | Em. | N. Am. | | |
| | | N. A. | | |
| G. | | Chili. | | |
| G. | Hyd. | Chili. | | |
| | ł | New Gran. | | |
| | | W. N. Am. | | |
| G. | | | | |
| G. | | N. W. Am. | | |
| G. | | N. Am. | | |
| L. | Astr. | Eur. N. Am. N. | | |
| 1 6 | | As. | | |
| Sm. | | Asia | | |
| C. G. Sm. | Hya. | N. Afr. Eur. W. | | |
| 1 0 | TT4 | As. Orient. Mex. | | |
| | | N. A. | | |
| | | Ry. Mts. W. N. | | |
| Cires. | 11 Ly 11. | Am. | | |
| T. | Lav | Eur. Cauc. Reg. | | |
| Sm. Mi.C.L. | | Orient. | | |
| | | S. Eur. Orient. | | |
| | Ast. Hyd. | Temp. As. | | |
| | G. L. G. L. G. G. L. Aut. Sap. G. L. Hyd. Hyd. Hyd. G. L. Em. G. Hyd. G. Hyd. G. Hyd. G. Hyd. G. Hyd. G. Hyd. G. Em. L. Astr. Sm. C. G. Sm. Ast. Ilyd. G. Ches. Ches. Ches. Chyd. Hyd. G. Hyd. G. Em. L. Astr. Sm. L. Astr. Lyd. Ches. Hyd. Hyd. Hyd. Hyd. Hyd. Ches. Ches. Hyd. Hyd. G. Hyd. G. Hyd. Hyd. G. Hyd. Hyd. Hyd. G. Hyd. Hyd. G. Hyd. Hyd. Hyd. G. Hyd. Hyd. Hyd. Hyd. Hyd. Hyd. | | |

| NAME OF PLANT | Authority | Properties | Locality |
|---------------------------------------|----------------------------------|---------------|-----------------------------|
| Prunus serotina Ehrh. | G. Ches. | Hyd. | Temp. Reg. Eur. N. Am. |
| Prunus undulata Ham. | | Hvd. | Him. Reg. |
| | G. C. L. | | E. N. Am. |
| Prunus virginiana L. | | Hyd. | |
| Purshia tridentata DC. | G. | Em. | Ry. Mt. W. N. |
| Dominion of the many II and | | 111 | Tr. Afr. |
| Pygeum africanum Hook. | G. | Hyd. | Java. |
| Pygeum parviflorum Teijsm. & Binn. | G. | Hyd. | |
| Pyrus L. | G. G. G. G. G. | Hyd. | N. Tem. Reg. Eur. N. As. |
| Pyrus Aucuparia Gaertn. | G. | | |
| Pyrus lanata D. Don | G. | | Him. Reg. Eur. N. Afr. |
| Pyrus Sorbus Gaertn. | CI | 10 | Chili. |
| Quillaja saponaria Molina | C. L. | Sap. | Spain. |
| Rhodotypos herricides Sieb, and Zucc. | G. | Sap. Astr. | Eur. Tem. As. |
| Rosa canina L. | L. L. L. L. | Astr. | |
| Rosa gallica L. | L. | | Eur. Cauc. Reg. |
| Rubus chamaemorus L. | L4. | Diur. | N. & Arc. Reg. |
| Rubus cuneifolia E. Merc. | _ L. | | |
| Rubus villosus Ait. | G. L. | Sap. Mech. | E. N. Am. |
| Spiraea Aruncus L. | Ģ. | Sap. | N. Tem. Reg. |
| Spiraea bella Sims. | G. G. G. G. L. G. | Sap. | Him. Reg. |
| Spiraea canescens D. Don. | <u>ا</u> ج | Sap | Him. Reg. |
| Spiraea Filipendula L. | <u>G</u> , | Ant. | N. As. |
| Spiraea Humboldtii Hort. | G. | Sap. | D. M. A. |
| Spiraea hypericifolia L. | L. | Astr. | Rus. Tem. As. |
| Spiraea japonica L. | G. | Sap. | Japan. |
| Spiraea laevigata L. | G. | Sap | Siberia. |
| Spiraea palmata Pall. | G. | Sap. | Siber. China. |
| Spiraea tomentosa L. | L. | Astr. | IN. A. |

RUBIACEAE

| Asperula cynanchica L. | G. | | Eur. As. Min. |
|------------------------------------|----------|----------|----------------|
| Asperula odorata L. | G. L. | Cou. | Eur. |
| Basanacantha tetracantha Hook, | G. | F. P. | Trop. Am. |
| Bothriospora corymbosa Hook. | Ğ. | [| S. A. |
| Cephaëlis toxica A. St. Hil. | G. G. | Dras. | Mex. S. A. |
| ocpinacio toxica II. ot. III. | J | Dias. | Galapagos Is. |
| Cephalanthus occidentalis L. | G. | Sap. | E. N. A. |
| Chiococca P. Br. sp. | G. | | |
| | G, | Dras. | Mex. S. A. |
| Chiococca P. Br. sp. | L. | Dras. | Trop. Am. |
| Chiococca brachiata Ruiz. and Pav. | | Diur. | Trop. Am. |
| Chiococca racemosa L. | L. | Diur. | Ind. |
| Cinchona sp. | G Wh. | Ir. F. P | S. A. |
| Coffea mauritiana Lam. | G. | | Mascar. Is. |
| Coffea odorata Forst. | G. | | Amic. Is. |
| Coprosoma linariifolia Hook. f. | G. | | N. Zea. |
| Coptosapelta flavescens North. | Ğ. | A. P. | Malay. |
| Galium Aparine L. | L. | Diur. | N. Tem. Reg. |
| | | | Magellan. |
| Galium asprellum Mx. | L. | Cou. | N. A. |
| Galium triflorum Mx. | G. L. | Cou. | N. Tem. Reg. |
| Cultum trinorum hix. | G. 14. | Cou. | N. A. Eur. |
| Geophila macropoda DC. | G. | Em. | Peru. |
| Geophila reniformis G. Don. | G. | Em. | Peru. |
| Hamalia antone Ing | | | |
| Hamelia patens Jacq. | G. | e | Tr. Am. |
| Mitchella repens L. | G. L. | Sap. | E. N. Am. Jap. |
| Mitragyna speciosa Korth. | G. | | Borneo. |
| Morelia senegalensis A. Rich. | G. | F. P. | Tr. Afr. |

| NAME OF PLANT | Authority | Properties | Locality |
|---------------------------------|----------------|------------|----------------|
| Mussaenda frondosa L. | G. G. L. | Sap. | Malay. Ind. |
| Oldenlandia senegalensis Hiern, | G. | Ant. | Ind. Tr. Afr. |
| Oldenlandia umbellata L. | L. | Ant. | India |
| Paederia foetida | Sm. | } | Ind. Malay. |
| Palicourea rigida HBK | j | | S. Am. |
| Pavetta reticulata Bl. | G. G. | Em. | Java. |
| Plectronia dicocca Burck. | G. | Hyd. | |
| Psychotria emetica L. | L. | Em. | N. Gran. |
| Psychotria Ipecacnauha Stokes | G. Wh. L. | De.p Em. | Braz. |
| Randia aculeata L. | . Sm. | | W. Ind. |
| Randia dumetorum La. |] G. L. | F. P. | Tr. Old World. |
| Sickungia rubra Mart. | G. | Alk. | Braz. |
| Spermacoce capitata Willd. | G. | | Peru. |
| Spermacoce semierecta Roxb. | G. G. | Cou. | Sumatra. |
| Tricalypsa Sonderiana Hiern. | | P. | Tr. Afr. |
| Vangueria spinosa Roxb. | G. | | Tr. As. |

RUTACEAE

| Acronychia laurifolia Bl. | G. | F. P. | Tr. As. |
|----------------------------------|----------------------------|------------|------------------|
| Casimiroa edulis L. | G. L. | Nar. | Mex. |
| Choisya ternata HBK. | G. | Sap. | Mex. |
| Citrus Aurantium L. | C. |] | Tr. As. |
| Citrus medica L. | G. Wh. | Hyd. | Tr. As. |
| Cusparia febrifuga H. & B. | L. | F. P. | New Gran, |
| Cusparia toxicaria Engl. | L. G. | | Braz. |
| Evodia rutaecarpa Hook. | L. | Pur. | Him. Reg. |
| | | | China. Japan. |
| Lunasia philipinensis Planch. | G. | A. P. | Philipp. |
| Melicope erythrococca Benth. | G. | | Austr. |
| Peganum antidysintericum Kostel. | L. | Astr. | S. Afr. |
| Peganum Harmala L. | G. G. L. G. C. L. | Ant. | Cent, As. |
| Pilocarpus sp. | G. | | W. Ind. S. Am. |
| Pilocarpus officinalis Poehl. | L. | Diaph. | S. A. |
| Pilocarpus pennatifolius Lem. | Sm. Wh. | Dep. Ir. | Brazil. |
| Pilocarpus racemosus Vahl. | L. | Diaph. | W. Ind. |
| Pilocarpus spicatus A. St. Hil. | L. | Diaph. | Brazil. |
| Ptelea trifoliata L. | L. G. | Sap. | E. N. Am. |
| Ruta graveolens L. | G.C.Sm.L. | Abort. Ir. | S. Eur. |
| Ruta montana Mill. | G. | Abort, | Med. Reg. Cau. |
| Skimmia japonica Thunb. | G. | P. | Japan. |
| Thevetia Ahouai A. DC. | | | Braz. |
| Thevetia neriifolia Juss. | | | |
| Zanthoxylum alatum Roxb. | G. G. | F. P. | Him., Reg. Chin. |
| Zanthoxylum caribaeum Lam. | G. | | New Grau, |
| Zanthoxylum Naranjillo Griseb, | G. L. | Diur. | Argent. |
| Zanthoxylum scandens Bl. | G. | 1 | Java. |
| | | | |

SALICACEAE

| Populus balsamitera L. | Halsted Ir. | i. N. Am. Asia. |
|------------------------|--------------|-----------------|
| SA | LVADORACEAE | |
| Salvadora persica L. | G. | Orient, Ind. N. |

SAMYDACEAE

| NAME OF PLANT | Authority | Properties | Locality |
|---|----------------|------------|-----------------------------------|
| Casearia graveolens Dal. Casearia guineensis G. Don. Casearia tomentosa Roxb. | G. G. G. | F. P. | Ind. Tr. As. Tr. As. Austr. |

SANTALACCEAE

| Exocarpus cypressiformis R. Br. | G. M. | | Austr. | |
|--|------------------------------|--|--|--|
| SAPINDACEAE | | | | |
| Alectryon excelsum Gaert. Blighia sapida Kon. | G. | Hyd. | N. Zeal. Afr. W. Ind. | |
| Cardiospermum Halicacabum L. Cupania sp. | G. L. G. | Sap. | Tr. Reg. N. A. S. States | |
| Cupania Pseudorhus A. Rich, Dittelasma Rarak DC. | G. G. | F. P. F. P. | Austr. Malacca | |
| Dodonaea physocarpa Fv. Muell. Dodonaea viscosa Jacq. Ganophyllum falcatum Bl. | G. R. G. G. | F. P. | Austr. Cosmop. Tr. | |
| Harpullia arborea Radlk. Harpullia cupanioides Roxb. | G. R. G. R. | Sap. F. P. F. P. | Malacca. Ind. | |
| Harpullia thanatophora Bl. Koelreuteria paniculata Laxm. | G. G. | | N. Guin. China. | |
| Magonia glabrata A. St. H. Magonia pubescens St. Hil. Nephelium lappaceum L. | G. R. G. R. E. G. | Sap. F. P. Sap. F. P. | Braz. Braz. Malay. | |
| Nephelium Longana Camb. Paullinia costata Schlecht. | G. R. G. | Sap/F. P. | Ind. Burma. Mex. | |
| Paullinia Cupana HBK. Paullinia curassavica Jacq. Paullinia macrophylla Kunth. | G. R. L. R. G. R. | Sap. F. P. Sap. F. P. | Venez. W. Ind. N. Gran. | |
| Paullinia meliaefolia Juss. Paullinia thalictrifolia Juss. | G. G. | Sap. F. P. Sap. F. P. Sap. F. P. Sap. F. P. | Braz. Trop. Amer. | |
| Paullinia trigonia Vell. Sapindus abyssinicus Fres. Sapindus aborescens Aubl. | G. G. | Sap. F. P. Sap. F. P. F. P. | Brazil. Abyss. Guian. | |
| Sapindus marginatus Willd. | Sm. L. | Sap. | N. Am. S. Sts. Tex. | |
| Sapindus Mukorossi Gaertn. Sapindus Saponaria L. Sapindus trifoliatus L. | | F. P. F. P. F. P. | Trop. Asia N. & S. Am. Trop. As. | |
| Schleichera Serjania acuminata Radlk. | G. R. | Hyd. Nar. | Braz. | |
| Serjania curassavica Radlk. Serjania cuspidata Cambess. Serjania erecta Radlk. | R. E. G. E. & G. R. | Sap. F. P. | W. Ind. Brazil | |
| Serjania ichthyoctona Radlk. Serjania inebrians Radlk. | E. R. E. & G. R. | Sap. F. P. Sap. F. P. | Brazil. Costa Rica. | |
| Serjania lethalis A. St. Hil. Serjania mexicana Willd. Serjania nodosa Radlk. | E. G. R. L. L G. R. E. | Sap. F. P. Sap. F. P. Sap. F. P. | Brazil. Mex. W. Ind. | |
| Serjania piscatoria Radlk. Serjania polyphylla Radlk. | E. & G. R. G. R. | Sap. F. P. Sap. F. P. | Brazil. Guian. | |
| Talisia stricta Triana & Planch. Ungnadia speciosa Endl. | G. L. | Sap. Sap. | New Granad. Texas. | |

SAPOTACEAE

| | | | · · · · · · · · · · · · · · · · · · · |
|--|----------------|------------|---------------------------------------|
| NAME OF PLANT | Authority | Properties | Locality |
| Achras sapota L. | G. L. | Sap. | S. Am. |
| Bassia butyracea Roxb. | G. | F. P. | Ind |
| Bassia latifolia Roxb. | Ğ. | | Ind. |
| Rassia langifolia I | l ŭ | Astr. | Ind. |
| Bassia longifolia L. Bassia Mottleyana Miq. | L. G. G. | Hyd. | Malay. |
| Illipe Maclayana | Ğ. | lilyu. | New Guin. |
| | Č. | | |
| Lucuma Bonplandia HBK. | G. G. | 77 4 | Cuba. |
| Lucuma deliciosa Planch & Linden. | G, | Hyd. | New Gran. |
| Lucuma glycyphloca Mart. | L. G. | Astr. | Braz. |
| Lucuma mammosa Gaertn. | G. | Hyd. | S. Am. |
| Lucuma multiflora A. DC. | G. | Hyd. | W. In. |
| Lucuma salicifolia HBK | L. | Sap. | Mex. |
| Omphalocarpum procerum Beauv. | G. | 1 | Tr. As. |
| Payena latifolia Burck. | G. | Sap. | Sunda. Is. |
| Sideroxylon borbonicum A. DC. | G. | | Barbon Is. |
| Sideroxylon dulcificum A. DC. | Ğ. | | Tr. Afr. |
| Sideroxylon toxiferum Thunb. | Ğ. | A. P. | 1 |
| | | 121. 1. | 1 |
| SARRACE | | | |
| Sarracenia flava L. | L. | , | N. A. |
| Sarracenia purpurea L. | G. L. | Astr. | E. N. Am. |
| Sarracenia variolaris Mx. | L. | Astr. | N. A. |
| SAURUE | | | · |
| Saururus cernuus L. | G. L. | Sap. | E. N. Am. |
| | | 1 | <u>'</u> |
| | GACEAE | | |
| Aphanopetalum resinosum Endl. | G. | Alk. | Austr. |
| Callicema serratifolia Andr. | G. | San. | |
| Ceratopetalum apetalum D. Don. | G. | Cou. | Austr. |
| Chrysosplenium alternifolium L. | G. | Poi. | N. As. E. N. |
| | | - 0 | Am. |
| Chrysosplenium americanum Schw. | | P. | N. Am. |
| Deutzia staminea R. Br. | G. | Sap. | Lim Dow |
| Dichroa febrifuga Lour. | G. | Em. | Him. Reg. Him. Reg |
| Diciroa reorriaga Lour. | G. | C,III. | M. h. Cl. |
| Frantiquia muntificidas T. f. | 0 | | Malay China. |
| Escallonia myrtilloides L. f. | G. | | New Gran. |
| Francoa appendiculata Cav. | G. | Alk. | Chili. |
| Hydrangea arborescens L. | G. L. | Sap. Hyd. | N. Am. |
| Hydrangea Hortensia Sieb. | G. | Hyd. | Asia. Orient. |
| Hydrangea involucrata Sieb. | | Hyd. | Japan. |
| Hydrangea Lindleyana | Í G. |) -, | J F |
| Hydrangea Thunbergii Sieb. | G. G. | Hyd. | Japan. |
| Jamesia americana Torr. & Gray. | | Hyd. | Ry. Mt. N. Am. |
| Philadelphus coronarius L. | G. | | C E |
| Philadelphus grandiflorus Willd. | G. | Sap. | S. Eur. |
| Philadelphus I amousi | Ğ. | Sap. | N. Am. |
| Philadelphus Lemonei | Ğ. | Sap. | N. Am. Ry.Mts. |
| Philadelphus Lewisii Pursh. | G. | Sap. | |
| Philadelphus microphyllus A. Gray. | G. | Sap. | N. Am. N.Mex. |
| Ribes aureum Pursh. | [G, | | Ry. Mts. W. |
| | | | N. Am. |
| Ribes cereum Dougl. | G. Sm. | Intox. | W. N. Am. Ry. |
| | 0. 2 | , , , , | Mts. |
| Ribes macrobotrys Ruiz. & Pav. | Sm. | | Peru. |
| Ribes nigrum L. | L. | Diur. | Eur. N. As. |
| Ribes prostratum L. Hcr. | Sm. | | |
| Moes prostratum 12. Tier. | SIII. | | N. Am. |

| NAME OF PLANT | Authority | Properties | Locality |
|---------------------------------------|-----------|------------|----------|
| Saxifraga Andrewsii Harv. | G. | Sap. | Japan, |
| Saxifraga cortusaefolia Sieb. & Zucc. | G. | Sap. | |
| Saxifraga cuneifolia L. | G. | Sap. | Spain. |
| Saxifraga Sibthorpii Boiss. | G. | Sap. | Greece. |

SCITAMINEAE

Tr. As.

Kaempferia rotunda L. Kirtikar

| maempiena rotanda 14. | 1 ILII CINGI | | 121. 210. | | |
|----------------------------------|----------------|------------|----------------|--|--|
| SCROPULARIACEAE | | | | | |
| Beyrichia scutellarioides Benth. | I G. | Ŀm. | Brazil. | | |
| Bonnaya veronicaefolia Spreng. | G. | Ant. | Malay. China. | | |
| Calceolaria scabiosaefolia Sims. | | Em. | Peru. | | |
| Capraria biflora L. | G. | 12777 | Trop. Am. | | |
| Castilleja canescens. Benth. | Ğ. | Car. | Mex. | | |
| Chelone glabra L. | l c i | Ant. | E. N. Am. | | |
| Digitalis ambigua Murr. | G. L. C. L. | Card. | Eur. W. As. | | |
| Digitalis lutea L. | C. C. | Card. | S. Eur. | | |
| Digtalis purpurea L. | G Mi Sm | Card. Ast. | Eur. W. N.Am. | | |
| Digitalis purpurea 14. | Ches. L. | Card. Ast. | Lai. W. N.Aii. | | |
| Digitalis Thapsi L. | E. G. R. C. | E D | Spain. | | |
| Gerardia tenuifolia Vahl | Ches. | 1. 1. | E. N. Am. | | |
| Gratiola officinalis L. | Mi.G.C.Sm. | 1- | Eur. | | |
| Oratiola Omemans 14. | Ches. L. | 11. | Eur. | | |
| Gratiola peruviana L. | G. | | Trop. Am. Aus. | | |
| Oratioia peruviana 14. | } 0. | | | | |
| Limosella aquatica L. | G. | Con | New Zeal. | | |
| Limosena aquanca 4. | J. G. | Sap. | N. & S. Temp. | | |
| Limosella Cymbalaria Mill. | c. | İ | Reg. | | |
| | c. | 1 | Eur. | | |
| Limosella Elatine Mill. | c. | | Eur. Orient | | |
| Limosella spuria Mill. | | - | Eur. N. Afr. | | |
| Linaria vulgaris Mill. | C.Sm.Mi.L. | 1 F. | Eur. N. Am. | | |
| Molemanna amones I | Rusby | 1 | | | |
| Melampyrum arvense L. | C. L. G. L. | D' | Caucasus | | |
| Melampyrum silvaticum L. | | Diur. | Eur. N. As. | | |
| Pedicularis palustris L. | G.Mi.Chest | Ť. | N. Tem. & Arc. | | |
| Dedicates as a fee T |) M: | | Reg. | | |
| Pedicularis sylvatica L. | Mi. | 1 | Eur. | | |
| Rhinanthus major Ehrt. | C. | - | Eur. | | |
| Rhinanthus minor Ehrt. | C. L. | Ins. | Eur. | | |
| Scrophularia aquatica L. | G.Sm.C.L. | Card. | Eur. Cauc. | | |
| Scrophularia nodosa L. | G Sm.C.L. | | N. Temp. Reg. | | |
| Striga euphrasioides Benth. |] G. | P. | India. | | |
| Vandellia crustacea Benth. | G. | Card. | Cosm. Trop. | | |
| Vandellia minuta Miq. | G. | Em. | Java. | | |
| Verbascum Blattaria L. | G. | 1 | Eur. N. As. | | |
| | | | N. Am. | | |
| Verbascum crassifolium Hoffmgg. | G. | | Lusitania. | | |
| Verbascum dubium Roem & Schult. | G | | Lusitania. | | |
| Verbascum orientale Bieh. | E. R. | F. P. | Caucas. | | |
| Verbascum phlomoides L. | E. R. | F. P. | Eur. As. Min. | | |
| Verbascum pulverulentum Vill. | G | l | S. Eur. | | |
| Verbascum simplex Labill. | E. G. R. | F. P. | Med. Reg. Ori. | | |
| Verbascum sinuatum L. | G. | | Syria. | | |
| Verbascum thapsoides L. | G. | 1. | Eur. | | |
| Verbascum Thapsus L. | E.G.R.C.L | . F. P. | Eur. N. Am. | | |
| | | | Ori. Him. Reg. | | |

| NAME OF PLANT | Authority | Properties | Locality |
|--|--------------------------------------|--------------------|--|
| Veronica Beccabunga L. | L. | Diur. | N. Temp. Reg. Eur. As. Min. |
| Veronica officinalis L. Veronica virginica Nutt. | L. G. L. Rusby | Diur. Car. Em. | N. As. E. N. Am. |
| SELAGI | NELLACEAR | <u> </u> | · |
| Globularia Alypum L . | G. L. | Pur. | Med. Reg. |
| SIMAR | UBACEAE | | |
| Ailanthus glandulosa Desf. | G. Rusby L. | Taen. | China. |
| Balanites aegyptiaca Delile. | Ğ. | Sap. | N. Afr. Arab. Pales. |
| Balanites Roxburghii Planch. Brucea sumatrana Roxb. | G. L. | Sap. | Ind. Trop. As. Aust |
| Cneorum tricoccum L. Picraena excelsa Lindl. | C. | Ins. | S. Eur. W. Ind. |
| Picrasma quassioides Benn. | G. L. | Ins. | Him. Reg. China , |
| Samadera indica Gaertn. Simaba Waldinii Planch. Simaruba amara Aubl. | G. L. G. Sm. | | Ind. Trop. Am. |
| SOLA | NACEAE | | |
| Acnistus arborescens Schlecht. | G. | Nar. | Mexico. |
| Anthocercis Lab. sp. Atropa Belladonna L. | G. G.C.Mi.Sm. | | Austr. Eur. Ori. Ind. |
| Brunfelsia Hopeana Benth. Capsicum annuum L. | L. Rusby G. L. Sm. L. | A. P. Ir. | Brazil. Trop. Reg. |
| Capsicum frutescens L. | Sm. L. | Ir. | Mex. S. Am. Tex. Mex. Trop. Reg. |
| Capsicum minimum Blanco. Cestrum aurantiacum Lindl. | Wh. L. Sm. | Ir. Nar. | Philipp. Is. |
| Cestrum auriculatum L'Her. Cestrum macrophyllum Vent. | G. G. | Nar. | S. Am. W. Ind. |
| Cestrum nocturnum L. Cestrum pallidum Lam. | G. G. | | S. Am. Jamaica. |
| Cestrum Parqui L'Her. | G. | | Trop. Amer. |
| Cestrum vespertinum L. Datura arborea L. | G. C. | | W. Ind. S. Amer. |
| Datura fastuosa L. Datura ferox L. | G. L. C. | F. P. | Tr. Old. World |
| Datura Metel L. Datura meteloides DC. | G. C. L. | Nar. | Trop. Am. W. N. A. |
| Datura sanguinea Riz, & Pav. | G. L. | Nar. Nar. P. | S. Ain. |
| Datura Stramonium L. | C.G.Rusby Sm. Mi. Ches. L. | Нур. | Cosm. Trop. |
| Datura suaveolens Datura Tatula L. | C. Ches.Rusby | Hyp. | Cosm. Trop. |
| Duboisia Hopwoodii F. Muell. | Blyth. L. | Нур. | Austr. |
| Duboisia myoporoides R. Br. Hyoscyamus albus L. | G. M. L. G. L. | Card. Hyp. Del. | Austr. Orient. |

| NAME OF PLANT | Authority | Properties | Locality |
|--|-----------------------|------------|-------------------------------|
| Hyoscyamus aureus L. | C. | | A. Min. Mesop. |
| Hyoscyamus Falezlez Coss. | [C. | 1 | W. As. |
| Hyoscyamus muticus L. | C. L. | Nar. | Med. Reg.? |
| Hyoscyamus niger L. | R.Ches.Sm. | Hyp. Del. | Med. Reg.? Eur. W. As. |
| | Mi. Rusby E. G. L. | | Him. Reg. |
| | | | |
| Hyoscyamus physaloides L. | G. | Hyp. Del. | Siberia. |
| Hyoscyamus reticulatus L. | G. | Hyp. Del. | As. Min. Egypt. |
| Latua venenosa Phil. | G. | | Chili, |
| Lycium barbarum L. | G. | | Orient. |
| Lycopersicum esculentum Mill. | G. Mi. Sm. | Sap. | S. Am. Mex. |
| 26 1 2 2 2 | C. L. | | Tex. Tem. Reg. |
| Mandragora autumnalis Bertol. | Sm. | | Med. Reg. |
| Mandragora officinarum L. | C. G. L. | Del. | Med. Reg. |
| Nicandra physaloides Gaertn. | G. | P. berry | Peru. |
| Nicotiana alata Link & Otto. Nicotoana chinensis | C. | | Brazil. China. |
| Nicotoana chinensis Nicotiana glauca Grab. | C. G. M. | į | |
| Nicotiana quadrivalvis Pursh. | C. G. M. | | Arg. W. N. Am. |
| Nicotiana rustica L. | CL | Naus. | Mexico. |
| Nicotiana suaveolens Lehm. | C. L: G. M. | 114443. | Australia. |
| Nicotiana Tabacum L. | RusbyC.R. | Naus F P | S. Am. Temp. |
| Titotiana Labatani 24. | Ches. G. | | Reg. |
| | L. E. | | |
| Nierembergia hippomanica Mers. | I G. | | Arg. Rep. |
| Physalis Alkekengi L. | L. | Diur, | Eur. Japan. |
| Physalis foetens Poir. | G. | Nar. | Trop. Am. |
| Physalis virginiana Mill. | G. | Nar. | E. N. Am. |
| Physochlaina orientalis G. Don. | C. G. | | Orient. |
| Physochlaina praealta Miers. | G. L. G. L. | P. | Him. Reg. |
| Scopolia carnicola Jacq. Scopolia japonica Max. | C. L. | Atropin | Eur. |
| Scopolia japonica Max. | G, L, | Atropa Pi. | Japan. |
| Scopolia Iurida Dun. | G. | P. | Him, Reg. |
| Scopolia physaloides Dunl. | | | Siberia, Altai. |
| Solandra grandiflora Sw. | G. G. | Alk. | Trop. Amer. |
| Solanum aculeatissimum Jacq. | G. | | Tr. As. & Am. Aus. & N. Zeal. |
| Solanum aviculare Forst. | G. | | Venez. |
| Solanum caripense Humb. & Bopl. Solanum carolinense L. | L, | Diur. | N. A. |
| Solanum crispum Ruiz, & Pav. | Sm. | Del. | Chili. |
| Solanum Dulcamara L. | Ches. | Sol. Nar. | Eu. N. Am. |
| Solahani Balcamara 4. | Rusby L. | Con. Trai. | Austr. |
| Solanum ellipticum R. Br. | G. | | Austr. |
| Solanum esuriale Lindl. | | | Austr. |
| Solanum eremophilum Fv. M. | G. M. | | Austr. |
| Solanum grandiflorum R. & P. | G. | | Peru. |
| Solanum mammosum L. | G. | 1 | Trop. Am. |
| Solanum Melongena L. | C. Sm. | | Tr. Old World. |
| | İ | | Austr. N. Am. |
| Solanum nigrum L. | G.M.Rusby | Nar. | Eur. N. Am. |
| | C.Mi.Ches. | | As. |
| | L. | | |
| Solanum saponaceum Dun. | Ğ. | | Peru. |
| Solanum sodemeum L. | G. Sm. | | Trop. Old |
| | | | World. Calid. |
| Solanum Sturtianum F. v. M. | G. M. | | Austrl. |
| Solanum stramonifolium Jacq. | G. | | Cosm. Trop. |
| Solanum torum Sw. | G. | | Cosm. Trop. |
| Dominant torum Div. | | | |

| NAME OF PLANT | Authority | Properties | Locality |
|---|-------------------|--------------------|---|
| Solanum triflorum Nutt. | G. Ches. | | W. N. Am. Ry. |
| Solanum tuberosum L. | M. Rusby Ches. | Sol. | Mts. Neb. S. Am. Cult. Temp. Reg. |
| Solanum villosum Willd. | Mi. | i | Temp. Reg. Eur. As. Afr. |
| Solanum Xanti Gray Triguera ambrosiaca Cav. | G. G. | ļ | Calif. Spain. |
| Vestia lycioides Willd. | G. | | Chili. |
| Withania somnifera Dun. | G | Abort. | Med. Reg. Ori. |
| STERCUI | | | |
| Pterospermum diversifolium Bl. Sterculia alata Roxb. | G. G. | Ir. Nar. | India Malay. India, |
| TERNSTRO | EMIACEA | | |
| Camellia japonica L. | G. | | Japan, China. |
| Camellia Sasangua Thunb. | G. | F. P. F. P. | Japan, China. Japan. |
| Caraipa fasiculata Cambers. | G. | | Guiana. |
| Caryocar glabrum Pers. Llanosia Toquian Blanco. | G. G. | F. P. | Pacific Isl. |
| | J | | Guiana. |
| THYMELA | AEACEAE | | |
| Daphne Gnidium L. | | Ir. | Med. Reg. |
| Dahpne Laureola L. | | Ir. F. P. | Eur. N. As. Eur. N. As. |
| Daphne mezereum L. Daphne striata Tratt. | L. G. | Ir. | Alp. S. Eur. |
| Daphnopsis cestrifolia Meiss. | G. | | N. Granat, |
| Daphnopsis Cneorum L. | G.R.Mi.E. | F. P. F. P. Ir. | Eur. Austr. |
| Daphnopsis Gnidium L. Daphnopsis oleoides Schreb. | | r. P. 1r. Ir. | Med. Reg. Eur. As. Min. |
| Dirca palustris L. | G. L. | Sap. | E. N. Am. |
| Edgeworthia Gardneri Meiss. | G. | F. P. | Him. Reg. Chi |
| Lasiadenia rupestris Benth. | G. | Р. | Japan. Guinea. |
| Lasiosiphon anthylloides Ness. | Ğ. | P. | S. Afr. |
| Lasiosiphon eriocephalus D. C. | G. | P. | India. |
| Thymelaea Tartonraira All. Wikstroemia Chamaedaphne Meiss. | G. G. | Ir. F. P. | S. Eur. China. |
| Wikstroemia viridiflora Meiss. | G. R. | F. P. | Tr. As. Austr. |
| TILIA | CEAE | | |
| Corchorus capsularis L. | G. | | Trop. Cosm. |
| Echinocarpus Sigun Bl. | Ğ. | Hyd. | Java. |
| Grewia asiatica L. | G. R. | F. P. | India. |
| Grewia bracteata Roth. | G. R. | F. P. F. P. | India. Amicor Is. |
| Grewia Malococca L. Grewia orientalis L. | G. K. | r. r. | Tr. As. & Aus. |
| Grewia pilosa Roxb. | Ğ. | | Ind. Trop. Afr. |
| Grewia piscatorum Hance. | | r n | China. |
| Prockia theaeformis Willd. | | F. P. | Mascar Is. |
| TROPAEC | | | |
| Tropaeolum majus | | Ir. | Peru. |
| ТҮРНА | | | 2 42 |
| Typha latifolia L. | G. | P. | N. Temp. Reg. |

ULMACEAE UMBELLIFERAE

| OMBELDITERAE | | | | |
|--|-------------|------------|------------------------|--|
| NAME OF PLANT | Authority | Properties | Locality | |
| Aethusa Cynapium L. | G.C.Mi.Sm. | Ir | Eur. Orient. | |
| Ammi Vienege Lem | G. L. | Diur. | Med. Reg. | |
| Ammi Visnaga Lam. | | | Med. Reg. | |
| Anthriscus cerefolium Hoffm. | L. | Diur. | Eur. N. Asia. | |
| Anthriscus sylvestris Hoffm, | L. | | Eur. N. As. | |
| | | | Can, Reg. | |
| Anthriscus vulgaris Bern. | G. C. L. | | Eur. Orient. | |
| Apium graveolens L. | Mi. L. | Ir. | Eur. Ori. Ind. | |
| Tiplant graveorens 12. | 1411. 14. | 11. | Calif. | |
| | 0.16 | | | |
| Apium leptophyllum F. v. M. | G. M. | ~ | Austr. | |
| Apium nodiflorum Reichb. | Sm. | Ir. | 1 | |
| Azorella Glebaria A. Gray. | G. | P. | 1 | |
| Carum capense Sond, | G. | | S. Afr. | |
| Carum copticum Benth. & Hooker. | G. | | Eur. N. Afr. | |
| Caram copilian Bentin a 2200ner. | | | N. Asia. | |
| Carum Petroselinum Benth & Hooker | C. L. | ſr. | Eur. | |
| | L. Z. | 11. | Eur. Temp. As. | |
| Caucalis daucoides L. | C. | | reur. Temp. As. | |
| Chaerophyllum temulum L. | G. | | Eur. N. Afr. | |
| | | | Caucas. Reg. | |
| Cicuta Bolanderi A. Gray. | Ches. | Con. | Calf. | |
| Cicuta bulbifera L. | Rusby | Con. | N. A. | |
| Cicuta californica A. Gray. | | Con. | Calif. | |
| Cienta manufacta I | Dueby | Con. | Cani. | |
| Cicuta maculata L. | Rusby | Con. | TO NE A | |
| | G. Ches. L. | _ | E. N. Am. | |
| Cicuta occidentalis Greene | Ches. | Con. | W. N. Am. Ry. | |
| | | | Mts. | |
| Cicuta vagans Greene | G. Ches. L. | Con. | W. N. Am. W. N. Am. | |
| Cicuta venenata Nutt. | G. | Con. | W. N. Am. | |
| Cicuta virosa L. | G.Sm.C.Mi. | Con. | Eur. N. Am. | |
| Cicuta viiosa 2. | L. Rusby | 1 | | |
| Conium maculatum L. | | Dep. | Eur.Ori.N.Am. | |
| Comuni maculatum 14. | | Dep. | Lui.Oii.w.mii. | |
| | Rusby G. | | | |
| | Sm.Ches.C. | | l., | |
| Conopodium denudatum Koch. | G. | | Eur. | |
| Coriandrum sativum L. | G. L. | | S. Eur. Ori. | |
| Crithmum maritimum L. | G. | Ant. | Eur. | |
| Daucus Carota L. | C. L. | Diur. | Eur. Ori.N.Am. | |
| Elaeselinum asclepium Boiss | Sm. | Ir. | S. Eur. | |
| Elaeselinum foetidum Boiss | Sm. | Ir. | S. Eur. N. Afr. | |
| | G. | 11. | | |
| Ferula communis L. | 1 | . . | Med. Reg. | |
| Ferula foetida Regel | L. | Antisp. | Turkest. | |
| Ferula galbaniflua Boiss. & Buhse. | Wh. L. | Ir. | Persia. | |
| Heracleum lanatum Michx. | G. L. | Acr. | Eur. N. Am. | |
| | | | W. As. | |
| Heracleum Spondylium L. | C. L. | Acr. | Eur. N. As. | |
| Heteromorpha arborescens Cham. & Schl. | G. | 1 | S. Afr. | |
| | | Nar. | 0. 1111. | |
| Hydrocotyle asiatica | L. | | T | |
| Hydrocotyle javanica Thunb. | [G. | F. P. | Java | |
| Hydrocotyle umbellata L. | G. | Em. | Trop. , Reg. | |
| Hydrocotyle vulgaris | Sm. Mi. | | Eur. | |
| Lichstensteinia Beiliana Eckl. & Zeyh. | G. | | S. Afr. | |
| Lichstensteinia pyrethifolia Cham. & | | | S. Afr. | |
| Schlecht. | | | | |
| Molopospermum cicutarium DC. | G. | Nar. | Eur. | |
| Oenanthe crocata L. | G. C. L. | Ir. | Eur. | |
| Oenanthe fistulosa | C Mi | Ir. | | |
| | C. Mi. | 11. | Eur. Casp. Reg. | |
| Oenanthe Lachenalii G. Gmel. | G. | I | Eur. Casp. Reg. | |

| NAME OF PLANT | Authority | Properties | Locality |
|---|--------------------------------|---------------------|-------------------------------------|
| Oenanthe Phellandrium Lam. | G. Sm. C. | Ir. | Eur. N. As. |
| Oxypolis rigidus Britton. Pastinaca sativa L. Peucedanum ambiguum Nutt. | L. Mi. Ches. C. L. G. | Diur. | E. N. A. Eur. N. A. W. N. A. |
| Peucedanum foeniculaceum Nutt. Peucedanum Ostruthium Koch. | L. L. | Acr. | W. N. A. Eur. |
| Pimpinella Saxifraga L. Sanicula marilandica L. | L. G. L. | Naus. Astr. | Eur. E. N. A. |
| Sium cicutaefolium Schrank. | L. Ches. | | N. Tem. Reg. |
| Sium erectum Huds. Sium latifolium L. | G. Sm. Mi. Mi. C. Ches. | Ir. | Eur. Cauc. Reg. Eur. N. A. |
| Thapsia garganica L. | G. L. | Ir. | Med. Reg. |
| Thapsia villosa L. Trachymene australis Benth. | G. M. | Ir. | S. Eur. Austr. |
| | | <u> </u> | Austi. |
| | ACEAE | | |
| Antiaris toxicaria Lesch. Artocarpus calophylla Z. & M. | Sm. L. | Ast. A. P. F. P. | Ind. Malay Tennasserim |
| Cannabis sativa Lam. | Mi. Sm. L. | | Cent. As. N. |
| Ficus altissima Bl. | Rusby | | Him.Eur.N.Am. |
| Ficus Carica L. | G. G. L. | Lax. | Tr.Asia, Malay S. Eur. Orient |
| | | | Afr. N. & S. |
| Ficus hispida L. | G. | P. | Am. Austr. Trop. As. & Austr. |
| Ficus hypogaea. | G. | | 1 |
| Ficus leucantatomia Poir. Ficus Roxburghii Wall. | G. Sm. | Ast. | Malaya Him. Burma |
| Humulus Lupulus L. | Mi. L. | Sed. | } |
| Laportea canadensis Gaud. Laportea crenulata Gaud. | Wh. L. | Ir. Ir. | N. Am. Trop. As. |
| Laportea Gaudichaudiana Webb. | L. | Ir. | Phil. Is. |
| Laportea stimulans Miq. Trema aspera R. Br. | G. L. M. | Ir. | Java Austr. |
| Urtica dioica L. | Sm. Mi. L. Ches. | Ir. · | N. Temp. Reg. |
| Urtica gracilis Ait. | | Ir. | E. N. Am. Ry. |
| Urtica holosericea Nutt. | Pam. | Ir. | Mts. Ry. Mts. W. N. Am. |
| Urtica pilulifera L. | Sm. L. | Ir. | Orient. Med. |
| Urtica urens L. | Mi. L. | Ir. | Reg. Tem. old world |
| VALERIA | NACEAE | | |
| Valeriana dioica L. Valeriana officinalis L. | C. Sm. C. L. | Te | Eur. Eur. N. As. |
| | | 11. | Lat. IV. AS. |
| | VACEAE | | 137 |
| Callicarpa americana Lam. Callicarpa cana L. | G. G. | | N. Am. Tex. Trop.As. Austr. |
| Callicarpa longifolia Lam. | G. | F. P. | Malay, Austr. |
| Callicarpa tomentosa Willd. Clerodendron infortunatum Gärt. | G. G. | | China Malay |
| Occidention infortunatum Galt. | 1 0. | · | Intalay |

| NAME OF PLANT | Authority | Properties | Locality |
|--|-------------|------------|------------------------|
| Clerodendron Siphonanthus R. Br. | G. | | Ind. & Malay |
| Duranta Plumieri Jacq. | G. | Sap. | Trop. Am. |
| Lantana salviaefolia Jacq. | G. | Abort. | S. Afr. Cosm. Trop. |
| Stachytarpheta indica Vahl. Verbena hastata L. | G. L. L. | Ir. | N. A. |
| Verbena officinalis L. | Sm. L. | Ir. | Amphig. temp. |
| Verbena venosa Gill. & Hook. | Sm. | Tr. | Arg. Reg. |
| Vitex sp. | G. | | S. Am. Afr. A |
| Vitex pteropoda Mig. | G. | | Austr. Malay |
| Vitex vestita Wall. | G. | | Malay |
| The state of the s | | · | |
| VIOLA | ACEAE . | | |
| Anchietea salutaris St. Hil. | G. L. | Em. | Brazil |
| Ionidium Ipecacuanha Vent. | L. | Em. | Brazil S. Am. |
| Noisettia HBK. Viola | G. G. | Em. Em. | N. Temp. Reg. |
| Viola odorata L. | C. L. | Em. | N. Afr. & As. |
| Viola pedata L. | L. | Em. | N. A. |
| Viola sepenicola Jord. | C | <u> </u> | Eur. |
| VITA | CEAE | | |
| Cissus nivea Hochst. | G. | 1 | Abyss. |
| Cissus pruriens Planch. | G. | | |
| Vitis hederacea Ehr. | L. | Ir. | E. N. Am. Ry |
| (Ampelopsis quinquefolia Michx. Psedera quinquefolia (L.) Greene) | Hals. | 1 | Mts. |
| Vitis elongata Wall. | G. | - | Ind. |
| Vitis Minahau. | G. | | |
| Vitis Saponaria Seem. | <u>G</u> . | | Austr. Pac. Is. |
| Vitis sessilifolia Baker. | В. | <u> </u> | Brazil |
| XYRID | ACEAE | | |
| Xyris communis Kunth. | | | Trop. Am. |
| ZINGIBE | CRACEAE | | |
| Globba Beaumetzii Heck. | G. | Taen. | 1 |
| Hedychium longecormatum. | G. | Ant. | |
| Thaunatococcus Daniellii Benth. | G. | т. | Trop. Afr. |
| Zingiber officinale Rose. | Sm. | lr. | India |

ZYGOPHYLLACEAE

| G. L. | Sap. | India |
|-------|---|---------------------------------------|
| G. L. | A. P. | Mex. S.W.U.S. |
| G. | Ineb. | Syria, N. Afr. |
| | | & Tropics |
| G. | | Cosm. Trop. |
| G. | (Sap. | N. Am. Guiana |
| G. | | N. Am. Arab. |
| G. | | Spain, N. Afr. |
| 1 | | W. Asia. |
| G. | | Austr. |
| | G. L. G. G. G. G. G. G. | G. L. A. P. Ineb. G. G. Sap. G. G. G. |

EUTHALLOPHYTA, SCHIZOPHYTA, SCHIZOMYCETES.

| | | PHYTA. ERIA. | SCHIZON | TYCETES. |
|--|--------|--|-------------|--|
| NAME OF PLANT | | Authority | Properties | Locality |
| Bacillus botulinus von Erm. Bacillus piscidus Sieber (i) Schu | m. | Jordan G. | | N. Am. Eur. Eur. |
| For other toxic species see Buch | anan, | Part I of t | his Manual. | |
| EUTHALLOPHYT RHO | | EUPHYCE. ELACEAE | | AE. |
| Alsidium Helminthochortes Ktzg. Chondria vermicularis Hook. Hypnea muscaeformis Rhodomenia palmata Gre. | | G. G. G. G. | | Atlantic Ocean Atlantic Ocean Atlantic Ocean Atlantic Ocean |
| See Parts I and II of this Ma | nual. | | | |
| EUTHALLOPH | YTA. | | MYCETES | • |
| Saprolegnia sp. | | G. | | Cosm. |
| | | YCETES llaceae | | |
| Gyromitra esculenta Fr. | | H. | | Eur. N. Am. |
| I | Туросі | reaceae | | |
| Claviceps purpurea Tul. | | G. Mi. | Abort. | Eur. N. Am. As. Austr. |
| BASI | IDIOI | MYCETES | | |
| | Agario | | | |
| Amanita cothurnata Atk. | | A. | 1 | E. N. Am. |
| Amanita floccocephala Atk. | | A. | | E. N. Am. |
| Amanita Frostiana Pk. | | Cl.A.H.Pk. | | E. N. Am. |
| Amanita magnivelaris Pk. | | H. | | E. N. Am. |
| Amanita mappa Fr. Amanita muscaria L. | | H. | Į | E. N. Am. |
| Amanita muscaria L. | | Cl. G. H. Farl. A. Pk. Fr. | | Eur. N.Am. As. |
| Amanita phalloides Fr. | Ì | Ches. Ph. Cl. Pk. H. Farl. | | Eur. N. Am. |
| Amanita recutita Fr. Amanita rubescens Fr. | 1 | A.Mi. Bull. Cl. A. Fr. Ph. Pk. | | Eur. N. Am. |
| Amanita solitaria Bull. | | Bull. Cl. A. Fr. Ph. Pk. | | Eur. N. Am. |
| Amanita spreta Pk. Amanita strobiliformis Fr. Amanita verna Bull. Amanita virosa Fr. Cantharellus aurantiacus Fr. Clitocybe illudens Schw. | | A. Pk. H. H. Fr. A. H. A. Fr. H. A. G.H.A.Ph. H. A. Cl. | | N. Am. Eur. N. Am. Eur. N. Am. Eur. N. Am. Eur. N. Am. Eur. N. Am. |
| Coprinus narcoticus Batsch. Entoloma clypeatum (L.) Miquel. Entoloma graveolens Pk. Hebeloma crustuliniforme Bull. | | G. Ph. H. Cl. Cl. Cl. | | Eur. Eur. N.Am. Fr. N. Am. N. Am. |

| NAME OF PLANT | Authority | Properties | Locality |
|--|---------------|--------------|----------------------------|
| Hebeloma fastible Fr. | H. Cl. | | Eur. N. Am. |
| Inocybe scaber Müll. | H. | | Eur. N. Am. |
| Lactarius acris Bolt. | G. H. | | Eur. N. Am. |
| Lactarius atroviridis Pk. | Pk. H. | | N. Am. |
| Lactarius camphoratus B. | [H. | | Eur. N. Am. |
| Lactarium chrysorrheus Fr. | Fr. H. | | Eur. N. Am. |
| Lactarius insularis Fr. | Fr. H. | Acr. | Eur. N. Am. |
| Lactarius pergamenus Fr. | Fr. H. | Acr. | Eur. N. Am. |
| Lactarius rubus Scop. Lactarius scrobiculatus Fr. | G | Acr. | Eur. N. Am. |
| Lactarius scrobiculatus Fr. | Fr. H. | Acr. | Eur. N. Am. Eur. N. Am. |
| Lactarius subdulcis Fr. | H. Fr. | Acr. Acr. | Eur. N. Am. |
| Lactarius torminosis Fr. | G. H. Fr. | Acr. Acr. | Eur. N. Am. |
| Lactarius trivialis Fr. | Fr. H. | Acr. | Eur. N. Am. |
| Lactarius turpis Fr. Lactarius vellereus Fr. | Fr. H. | Acr. | Eur. N. Am. |
| Lactarius venereus Fr. Lactarius zonarius B. | G. | Acr. | Eur. |
| Lentinus suavissimus Fr. | G. Fr. | . (С). | Eur. |
| Lentinus stavissinus 11. | G. 11. | | Eur. |
| Lepiota Morgani Pk. | Pk. Či. H. | | N. Am. |
| Marasmias oreades Bolt. | G. | | Eur. N. Am. |
| Marasmias urens Bolt. | G. H. | | Eur. N. Am. |
| Paneolus papilionaceus Fr. | H. | | N. Am. |
| Pholiota caperata | C1. | | N. Am. |
| Pholiota radicosa B. | G. | Acr. | Eur. N. Am. |
| Russula Barlea L. | G | | Eur. N. Am. |
| Russula emetica Sch. Russula foetens Pers. | H.G.A.Cl. | Em. | Eur. N. Am. |
| Russula foetens Pers. | G. Cl. | | Eur. N. Am. Eur. N. Am. |
| Russula fragilis | G. H. Fr. | | Eur. N. Am. |
| Russula rubra Fr. | . H. Fr. | | N. Am. |
| Stropharia aeruginosa Curt. Tricholoma album Schaeff. | H. | Acr. | Eur. N. Am. |
| Tricholoma saponaceum Fr. | Fr. Cl. | Acr. | Eur. N. Ani. |
| Tricholoma sulphureum | Cl. | Acr. | Eur. N. Am. |
| Volvaria volvacea | ı Cl. | | N. Am. |
| HYDN | ACEAE | | |
| Hydnum amarescens L. | G. | | Eur. |
| Hydnum graveolens | G | | Eur. |
| | RDACEAE | | |
| Lycoperdon Bovista Pers. | G. Mi. | | Eur. N. Am. |
| Lycoperdon gemmatum | Mi. | | Eur. N. Am. |
| Lycoperdon giganteum Fr. | Fr. G. | | Eur. N. Am. |
| Lycoperdon saururus Lam. | G. | | Eur. |
| Scleroderma vulgare | G. Cl. | | Eur. N. Am. |
| Secotium acuminatum | | | N. Am. |
| The state of the s | ACEAE | | INT. A Thur |
| Ithyphallus impudicus | | | N. Am. Eur. |
| Lisurus borealis Burt. Mutinus caninus Fr. | H. Fr. H. Cl. | | N. Am. Eur. N. Am. |
| Mutinus elegans Mont. | H. | | N. Am. |
| Phallus duplicatus Bosc. | H. Cl. | | Eur. N. Am. |
| Phallus Ravenellii (B&C) E. Fisch. | H. | | Eur. N. Am. |
| | RIACEAE | | |
| Crucibulum vulgare. | C1. | 1 | Eur. N. Am. |
| Cyathus striatus | Ci. | | Eur. N. Am. |
| Cyathus vernicosus | ci. | | Eur. N. Am. |
| | | ' | 1 |

POLYPORACEAE

| NAME OF PLANT | Authority | Properties | Locality | |
|--|----------------|------------|------------------------------|--|
| Boletus alveolatus B. & C. | H. | | N. Am. | |
| Boletus felleus Bull. | А. Н. | | Eur. N. Am. | |
| Boletus Frostii Russell | H. | | N. Am. Eur. N. Am. | |
| Boletus luridus Schaeff. | М. Н. | | Eur. N. Am. | |
| Boletus satanas Lenz. | H. Cl. | | Eur. N. Am. | |
| Polyporus anthelminticus Berk. | · G. | | Eur. | |
| Polyporus hispidus Bull. | G | | Eur. | |
| Polyporus squamosus Fr. | Fr. G. | | Eur. | |
| UREDI | NEAE | | | |
| Coleosporium Sonchi-arvense (Schw.) | | Ir. | Eur. N. Am. | |
| Thunb. | Ches. | | Austr. Cosmp. | |
| Puccinia coronata Cda. | Pammel | | Eur. N. Am. | |
| D 11 11 D | 77 | | Cosmp. | |
| Puccinia graminis Pers. | Virchow | | Eur. N. Am. | |
| Don't DC | D1 | | Cosmp. | |
| Puccina rubigovera DC. | Pammel | | Eur. N. Am. | |
| Descripto contemptato | Nelson | | Cosmp. | |
| Puccinia xylorrhizae Uromyces trifolii (Hedw.f) Lev | Mohler | 1 | N.Am. Ry.Mts. Eur. N. Am. | |
| Oromyces trifoni (Hedw.1) Lev | Monier | | Cosmp. | |
| | | · | Cosinp. | |
| USTILAG | | | | |
| Ustilago avenae (Pers.) Jens. | Mi. | Ir. | Eur. N. Am. | |
| Ustilago Hordei P) Kell. & Swingle. | Mi. | Ir. | Cosinp. | |
| ostnago izoraci i y izem a piningie. | | | Cosmp. | |
| Ustilago neglecta Niessl. | Pammel | Ir. | Eur. N. Am. | |
| Hatilaga muda (Tanaan) Wall & Swingla | Mi. | Ir. | Cosmp. Eur. N. Am. | |
| Ustilago nuda (Jensen) Kell. & Swingle. | N11. | Ir. | Cosmp. | |
| Ustilago Tritici (Pers.) Jens. | Mi. | Ir. | Eur. N. Am. | |
| Ostnago Tritici (1 cis.) Jens. | 1111. | 11. | Cosmp. | |
| Ustilago utriculosa Tul. | Pammel | Ir. | N. Am. | |
| TILLET | · | | | |
| Tilletia foetens (B.&C.) Trel. | Schaffner | Ir | N. Am. Eur. | |
| Tilletia Tritici (Bjerk.) Wint. | Schaffner | | N. Am. Eur. | |
| FUNGI IMPERFECTI | | | | |
| Cladosporium sp. | l G. | Ir. | Eur. N. Am. | |
| Diplodia zeae Lev. | E. F. Smtih | Pellagra | Eur. N. Am. | |
| Fusarium sp. | G. | Ir. | Eur. N. Am. | |
| Fusarium roseum Lk. | Pammel | Ir. | Eur N. Am. | |
| Giberella sp. | G. | Ir. | Eur. N. Am. | |
| Macrosporium Brassicae B. | Pammel | Ir. | Eur. N. Am. | |
| Polydesmus exitiosus Külin. | Mohler | Ir. | Eur. N. Am. | |
| Polythrincium trifolii Kuntze. | Pammel | Ir. | Eur. N. Anı. | |
| See Part II of this Manual for other sp | ecies. | | | |
| | ENES | | | |
| Cetraria juniperina Ach. | G. | | Eur. N. Am. | |
| Cetraria pinastri Ach. | G. | | Eur. | |
| Evernia vulpina Ach. | G. G. G. | 1 | Eur. N. Am. | |
| Parmelia vulpina Ach. | G. | | Eur. | |
| Peltigera horizontalis L. | G. | | Eur. N. Am. | |
| Variolaria amara Ach. | G | 1 | Eur. | |
| | | | | |

BIBLIOGRAPHY OF POISONOUS PLANTS

BY HARRIETTE S. KELLOGG

In this bibliography we have included such modern works bearing upon the subject of poisonous plants as are more easily accessible to the student, but to make this list more complete many titles of books by older writers have been added. These works are of interest not only from an historical point of view, but also from an artistic standpoint because many of them are hand-somely illustrated. This is particularly true of the works of Brand and Ratzeburg, Berge and Riecke, Godet, Bulliard, Dietrich, Henry, Hegetschweiler, Otto, and Miquel which were published many years ago.

The bibliography also contains the more important articles published by the various experiment stations and by the United States Department of Agriculture besides a long list of rather recent papers appearing in technical chemical journals and in professional journals of medicine, veterinary medicine, and pharmacy. However, this bibliography is by no means complete so far as these technical journals are concerned, but from the papers indicated in the bibliography the student can easily find further literature.

Miss Kellogg has attempted to make a subject catalogue covering a great many different topics such as the sale of poisons, general treatises on poisonous plants, vegetable toxicology, poisons from Abrus, vegetable alkaloids, hydrocyanic acid, immunity, lupinosis, ricinus, poisonous seeds, geographical papers, poisons from a legal standpoint, pellagra. antidotes, etc.

While it would be impossible to indicate a reference to every species, the catalogue of the poisonous plants of the world indicates, to some extent, where the species was discussed. We have had to omit from the catalogue references to many popular treatises such as Lehmann's "Giftpflanzen mit besonderer Berücksichtigung der wirksamen Stoffe," and the works of Godet, Vicat, Henslow's "Poisonous Plants in Field and Garden" and the "Giftpflanzenbuch" by Berge and Riecke. The later as well as the earlier treatises, frequently refer to the literature where the species is mentioned as being poisonous. An example of this is found in Helleborus niger, Berge and Riecke give seventeen references where the plant is mentioned as poisonous; eleven references are cited in regard to Oleander, and five upon Tecoma radicans, showing that many of these plants have passed as poisonous for a considerable length of time.

Of peculiar and local popular treatises Germany has contributed more than any other country. The treatment of the poisonous plants in books such as Goeppert's "Ueber die Giftige Pflanzen Schlesiens," and Krause's "Studies of Poisonous Plants, in the German Colonies," aid materially in determining the distribution of certain plants. The United States has done something along this line, especially through its Department of Agriculture. The publications of Coville and Chesnut, are excellent illustrations of this. Popular accounts in such treatises as that by Miss Huntington are helpful. The subject of poison ivy probably has been treated more exhaustively than any other subject indicated. Dr. Warren has recently published a long list of papers on this subject, and he shows especially how popular impressions get into current literature

without having much foundation in fact. On the subject of the anatomy very little has been published. Attention may be called to the excellent treatise of Collin, "Traite de Toxicologie Vegetale, Application du Microscope a la Recherches des Poisons. Vegetaux." 1907. The German work by Mitlacher, "Toxicologische oder forensische wichtige Pflanzen und vegetabilische Drogen, mit besonderer Berücksichtigung ihrer mikroskopischen verhältnisse", also has an admirable discussion of the subject. There are, however, many pharmocopeal treatises like the work of Flückiger (English translation "Pharmacographia"), "Pharmacognosy" by Powers, Kraemer's "Botany and Pharmacognosy", the "Organic Materia Medica" by Sayre, "Foods and Drugs" by Greenish, Tschirch and Oesterle's "Anatomischer atlas, der Pharmakognosie and Nahrungsmittelkunde," and "Microscopy of Vegetable Foods" by Winton, and the English translation of works of Moeller, which will indicate the sources from which information of this kind can be secured.

We have added some "second hand" references which have been taken from what we believe to be reliable sources. It was quite impossible to obtain all of the original papers referred to in the bibliography. Many of them, however, have been seen in the original, either by Miss Kellogg or myself.

We wish to express our thanks to Dr. Wm. Trelease of the Missouri Botanical Garden, Dr. B. L. Robinson and Dr. W. G. Farlow of Harvard University, Dr. C. S. Sargent of the Arnold Arboretum, Mr. C. G. Lloyd of the Lloyd Library, Cincinnati, to the Surgeon General of the United States Army, and Mr. Johnson Brigham of the Iowa State Library, Des Moines, for courtesies in the loan of books and papers.

L. H. PAMMEL.

PARTIAL CLASSIFICATION OF THE BIBLIOGRAPHY

The numbers following the topics correspond with the numbers in the Bibliography.

ABRUS, 158, 508a, 550 Achorion, 94 Aconite, 186, 263, 302, 366, 474, Aconitum, 503a, 528, 923, 947, 1078 Acorn, 2 Actinomycosis, 88, 123, 138a, 463, 809, 832, 859a, 1072 Aethusa, 147, 302, 1078 Agaricus, 934 Alexipharmics, 679 Alfalfa, 414, 628, 1036 Algae, 33, 34, 70, 74, 126, 285, 286, 676, 728, 742, 991 Alkaloids, Vegetable, 121, 171, 187, 397, 484, 779, 779a, 780, 781, 808, 817, 873a, 938, 1082 Almonds, 688 Amanita, 307, 510, 512, 633a, 660, 797 Anatomy, 236, 237, 763, 887, 997 Andromeda, 695, 799, 801 Andromedotoxin, Anemone, 687, 708 Anhalonium, 193, 593, 599 Anise, see Star anise Anthocercis, 670

Antiarin 662, 916, 917 Antiaris, Antidotes, 157, 307, 319, 376, 410, 478, 507a, 508a, 522, 804, 932, 973 Antitoxins, 561, 718 Arbor toxicaria, 981 Areca, 183, 302 Argemone, 885 Aroideae, 754, 897 Arum. Arrow Poisons, 43, 50, 133, 156, 314, 329, 594, 598, 898, 916, 917 Aspergillosis, 95, 154a, 318, 375, 527, Aspergillus, 5 618, 939 Astragalus, 332, 640, 725 Atractylis, 602 Atropa, 887 Atropin, 151a, 302, 944, 1078 Auto intoxication, 986

BACILLUS, anthracis, 51, 561 oedamatis, 135, 561 suipestifer, 711 tetani, 292, 561 virgula, 1011

Bacterial Poisons, 126a, 144, 321, 544, Corn, 86, 86b, 247a Broom, 427 Kaffir, 767 Cornstalk Disease, 12, 207, 819 1007 Bacterium, diptheriae, 561, 944 tuberculosus, 38, 217, 463, 561, 826 Corydalis, 642 Baneberry, 41 Barium, 209, 482, 487, 984 Beans, 115, 256, 260, 388, 393, 394, 420, Cotoneaster, 460 Coumarin, 440 Cowbane, 732 Creosote, 213 855 Crotalaria, Beech, 316 331, 945 Crotalism, 5 351, 945 Crotin, 508a, 561 Croton, 257, 302, 553, 645 Cryptogams, 75, 714, 778, 805 Curare, 50, 788, 789 Cyanogenesis, 258, 262, 264, 583 Cynoglossum, 246, 883. See Borage Cypripedium, 474, 513, 625, 626, 699, 700, 983 Cytisin. Belladonna, 302, 559, 621 Berberis, 302, 862 Bibliography, 3a, 477, 502a, 561, 579a, 727a, 823, 895, 1025, 1030
Black Tongue, 352 Blastomycosis, 449a, 526a, 776b, 987a Borage Plants, 367, 368 Boraginaceae, Botulism, } 420, 463, 561, 633 Botulismus, Cytisin, 189, 302 Boxwood, 334 Cytisus, CACTACEAE, 193, 444, 599 Calabar Bean, 302, 311, 412 Calandrina, 720 Dalbergia, 199 Damiana, 72 Darnel, 384 Camas, 253 Datura, 159, 239, 302, 337, 835, 887, Camphor, 856 Cannabis, 284, 816 Capsella, 532 Cardiac Poisons, 493 1053 Dermal Mycosis, 659, 675, 680, 713, 741. 783, 784. Dermatitis, J 1043 Dermatomycosis, Cascara, 290 Derris, 727, 763 Cashew, 1063 Dicentra, 35 Digitalis, 270, 302, 563, 669 Cassava, 19, 224, 657, 990, 1058 Castor Oil, See Ricinus Dioscorea, 905 Dioscorin, 905 Catalpa, 1024 Celastrus scandens, 78 Diphtherial Poison, See Bacterium Cerebritis, 128, 247, 247a, 466, 655, diphtheriae 749, 1061a Diplodia, 836a, 930 Cherry, 250, 681 Duboisia, 1078 Chickerinchee, See Ornithogalum Cholera, 1035 Cicuta maculata, 96, 219, 442, 474, ECHITES, 410 803, 929, 1094 Economic Plants, General, 238, 287, virosa, 96, 302, 531, 736, 1078 Cicutoxin, 96. Also see refe 686, 911 Also see references America, 228, 433 Mexico, 911 United States, under Cicuta Claviceps. See Ergot California, 160 Montana, 90 Cleistanthus, 480 Clover, 202, 414 Coca, 921 Porto Rico, 185, 761 Cocaine, 35a, 52a, 579a, 921, 1031a, Vermont, 518 1078 Asia and Adjacent Islands, Japan, 60 Cocos, 271 Coffee, 953 Australia, 282, 638 Colchicum, 6, 6a, 129, 302, 689, 832a, Miscellaneous, Cuba, 655b Guam, 870a 1078 Coniin, 501 Conium maculatum, 336, 501, 538, Entoloma, 464 1078, 1086 Convallaria, 474 Equisetosis, 842, 843, 843a, 961 Equisetum, 1002 Convulsives, 845 Ergot, Ergotism, } Copaifera, 302, 440 52, 231, 423, 530, 632, Ericaceae, 799 Copas, 481

| The 242 E74 | HAY FEVER, 6b, 338, 661, 814, 814a, |
|--|--|
| Erigeron, 243, 574 | |
| Eriobotrya, 458 | 852, 879a. 976 |
| Erythrophloeum, 596 | Haya Poison, 596 |
| Escholtzia, 297 | Heaves, 1062 |
| Eupatorium, 474 | Hemlock, 865 |
| Euphorbiaceae, 474, 925 | Hemlock Water, 501, 572, 573 |
| | Heracleum, 335 |
| Exalgin, 810 | Hippomane, 411, 846 |
| Favus, 94, 191 Ferns, 296, 683 | |
| Ferns, 296, 683 | Honey, Poisonous, 529, 800 |
| Fish Poisons, 277, 278, 279, 371, 480, | Horsetail. See Equisetum. |
| 543, 727, 746, 829 | Hydrocyanic Acid in Plants, 80, 105, |
| Flax, 259 | 125, 196, 224, 261, 349, 369, 372, 385, 386, 387, 388, 389, 390, 391, 392, 393, |
| Flores | 386, 387, 388, 389, 390, 391, 392, 393, |
| Floras, 274 523 | 394, 435, 458, 459, 520, 630, 992, |
| General, 47a, 274, 523 | 1014, 1078 |
| North America, 117, 363 | Hyoscyamus, 887 |
| United States, 116, 117, 155, 359 | |
| Alabama, 673a | IMMUNITY, 229, 252, 306, 502, 561 |
| California, 1030a | Ipoh, 327, 916, 917 |
| Connecticut, 355a | T Canad 900 |
| Iowa, 364a | Ivy, Ground, 890 |
| Maryland, 924 | Ivy Poison. See Rhus |
| | - |
| Minnesota, 631 | JABORANDI, 419 |
| Washington, 786a | Janipha, 409 |
| See also Bulletins from the | Jatropha, 409, 553, 1058 |
| various Experiment Stations | Jequirity, 661 |
| South America, See Medicinal and | J • 1, , |
| Poisonous Plants | KALMIA 204 212 533 695 864 |
| Asia, See Economic, Medicinal and | Kalmia, 204, 212, 533, 695, 864 Kenyah, Dart Poison. See Ipoh |
| Poisonous Plants | Vimenge See Vemange |
| Australia, 685a | Kimanga. See Komanga |
| Europe, | Kirondro Poison, 199 |
| | Kissoumpa. See Menabea |
| England, 70a | Kokra. See Cocos |
| Germany, 554 | Komanga, 439, 439 |
| Switzerland, 554 | Ksopo, 765 |
| Flowers, Poisondus, 22, 23, 853 | • • |
| Foods, 140, 421, 616, 1064 | LAC, Japanese, 948, 998 |
| Forage Poisoning, 125, 291, 465, 469, | Larkspur, 79, 211, 302, 339, 1047 Lathyrus, 267, 396, 900 Laurel. See Kalmia |
| 749, 1061 | Lathyrus 267 396 900 |
| Fungi, Poisonous, 73, 94, 95, 503, | Laurel See Kalmia |
| 509, 610, 611, 629, 903, 1091 | Lauro corneus 1006 |
| Also see Mushrooms | Lauro-cerasus, 1006 |
| | Leguminosae, 465, 557, 684, 761, 857, |
| GALEGA, 682 | 889 |
| Gastro-enteritis, 542 | Lemon, Salts of, 111 |
| Gaultheria, 399, 407, 474, 514, 786, | Lepiota, 161 |
| 820, 859 | Leucomaines, 484, 1008 |
| Gelsemium, 273, 474, 875, 926a, 1078 | Lichens, 553b |
| Genista, 227, 242 | Linamarin, 520 |
| Germicides 1009 | Loco Weeds, 1 89, 209, 210, 233, 234 Locoism, 5 235, 268, 356, 472, 482, 487, 533, 648, 649, 655a, 705, 721 |
| Germicides, 1009 Gift-apple, 232 | Locoism. (235, 268, 356, 472, 482) |
| Cincor 417 400 | 487 533 648 649 6552 705 721 |
| Ginger, 417, 499 | 728a, 743, 810, 876, 946 |
| Gleditschia, 745 | |
| Glucosides, 306, 308, 349, 392, 458, | Loganiaceae, 549 |
| 520, 938 | Lotus, 265 |
| Golden Rod, 910. See Hay Fever. | Lupine 141, 565, 889 |
| Grasses, 729, 731, 739, 740 | Lapinosis |
| Darnel, 384 | Lychnis Githago, 148, 554 |
| Foxtail, 696 | Lymphangitis, 727a, 727d, 749a, 827a |
| Johnson, 214 | |
| Porcupine, 82 | Maclura, 404 |
| Red Top, 423 | Macrosporium, 822 |
| Sleepy 213 | Macrozamia, 591 |
| Sleepy, 213 Sweet, 276 | Manchineal Tree. See Hippomane |
| Sweet, 270 | manenment free, occ mppomane |

| Mangels, 665 | Opium, 229, 302, 639. See Morphine |
|--|---|
| Materia Medica 9 39 55a 57 248 | Orange, 107 |
| 250 265 502 1066 1074 | Orange Foles 106 |
| Materia Medica, 9, 39, 55a, 57, 248, 350, 365, 592, 1066, 1074 Mechanical Injuries, 82, 87, 537, 696, | Orange, False, 106 Ordeal Poisons, 311, 438, 694, 1033, |
| Mechanical Injuries, 82, 87, 537, 090, | |
| 729, 731, 735 | 1057 |
| Medicinal Plants, General Distribu- | Ornithogalum, 495 |
| tion, 71, 169, 170a, 302, 604, 612, | Oxalic Acid, 561, 1078 |
| 620 024 042 1000 | O |
| 620, 834, 943, 1090 | |
| Africa, | Papaveaceae, 747 |
| Soudan, 575 | Parasitic Diseases, 703 |
| America, 84 | Parenia 110 472 720 |
| TILL'4 1 CALAS E7 172 201 020 | Parsnip, 119, 473, 730 |
| United States, 57, 172, 281, 830 | Peas, 507 |
| California, 333 | Pellagra, 29, 86a, 150, 154a, 351a, 581, |
| Massachusetts, 1060 | 661, 756, 825, 836a, 930, 987 |
| Minnesota 1034 | Pollotinio 442 |
| Minnesota, 1034 Missouri, 733 | Pellotinia, 443 |
| Missouri, 755 | Peppermint, 417 |
| North Carolina, 497 | Pharmacognosy 301, 302, 558, 877, Pharmacographia 1046, 1079 |
| Tennessee, 324 | Pharmacographia \$ 1046, 1079 |
| Asia and Adjacent Islands, | Phomosocia 202 450 |
| Afabanistan 26 | Phamacopoeia, 302, 450 |
| Afghanistan, 26 | American, 182 |
| Japan, 60 | American, 182 United States, 774, 776a |
| Malay, 338 | British, 115a |
| Philippines, 44 | |
| Medical Jurisprudence, 65, 155a, 169a, | French, 176a |
| 204 704 004 020 027 047 060 | German, 244a |
| 304a, 586, 804, 828, 837, 967, 968, | Italian, 288 |
| 1078 | Swedish, 775 |
| Menabea, 437, 438, 765 | Phaseolus. See Beans |
| Meningitis, 291, 561, 749 | Dhuasatiana 1 |
| Manines annulatities Con Completitie | Physostigma, { 272, 412 |
| Meningo-encephalitis. See Cerebritis | ~, ocotg) |
| Mesquite, 1063 | Phyto-bezoars, 202, 425, 991a |
| Millet, 219a, 467, 468 | Phytolacca, 474, 1042 Pink-root. See Spigelia Poison Lore, 4, 5a. 92a, 587, 978, 1018 Poison Tree. See Copas |
| Mistletoe, 580 | Pinls root San Spigalia |
| Morphine, 92, 299, 302, 516, 562, 877, | Tillk-100t. See Spigelia |
| 002 027 1045 1074 1079 | Poison Lore, 4, 5a. 92a, 587, 978, 1018 |
| 886, 967, 1045 1074 1078 | Poison Tree, See Copas |
| Mucor, 354 | Poisons, Detection of, 92, 93, 118, 154, |
| Mucorymcoses, 55 | 157, 429, 446, 552, 933 |
| Mulberry, 103, 367 | Poisons in Canaral 4 02 07 130 310 |
| Mushrooms, 36, 173, 175, 201, 203, 205, 308a, 381, 382, 569, 629, 654, 751, 815, 971. See Fungi | Poisons in General, 4, 92, 97, 130, 310, |
| 205 200, 201 202 560 620 654 | 380, 552, 614, 723, 846, 940 |
| 203, 300a, 301, 302, 309, 029, 034, | Mineral Poisons, 723, 963 |
| 751, 815, 9/1. See Fungi | Vegetable Poisons, 30, 31, 197, 475, |
| Mycosis, 145a, 673, 874 | 490 764 827 919 920 1000 1016 |
| | Poisonous Plants General Treatises, 28, 42, 75, 145, 188, 194, 197, 248, 249, 252, 299, 341, 343, 403, 455, 457, 521, 523, 556, 578, 588, 668, 709, 714, 744, 773, 812, 824, 834, 853, 858, 884, 902, 915, 933, 982, 1010, 1018, 1032, 1048, 1077 |
| Nicotine, 93, 302, 758a | 20 42 75 145 100 104 107 240 |
| Nuts, | 28, 42, 75, 145, 188, 194, 197, 248, |
| Areca, 183 Betel, 302 | 249, 252, 299, 341, 343, 403, 455, |
| Retel 302 | 457, 521, 523, 556, 578, 588, 668, |
| Nutrior 1052 | 709 714 744 773 812 824 834 |
| Nutmeg, 1052 | 853 858 884 002 015 033 082 |
| Numa, 873 | 1010 1010 1022- 1040 1077 |
| Nux vomica, 241 | 1010, 1010, 1032a, 1040, 1077 |
| | Geographical Distribution |
| OAK. See Acorn, Quercus | Africa, 839, 851 |
| Oedema. See Bacillus, oedamatis | Soudan, 575 |
| Oenanthe crocata, 61, 1078 | |
| Oils, | South Africa, 374, 1021b, 1059 |
| | America, North, 23 |
| Castor, 230, 302. See Ricinus | Mexico, 911 |
| Croton. See Croton | United States, 27, 162, 163, 166, |
| Gaultheria, 820 | 167 240 500 555 005 |
| Pennyroyal, 1029 | 167, 340, 500, 555, 805 |
| Peppermint, 417 | California, 67, 533, 894, 1005 |
| Savin, 1029 | Colorado, 339 |
| | Idaho, 927 |
| Tansy, 1029 | |
| Vervain, 104 | Indiana, 195 |
| Oleander (892, 1021, 1067 | Iowa, 734, 738 |
| Oleandrin \(\) 692, 1021, 1007 | Massachusetts, 922 |
| | |

| • | |
|--|--|
| Minnesota, 175 Missouri, 733 | Ptomaines, 121, 484, 1008 |
| Montana, 84, 90, 168, 1049 | Quercus, 4, 474, 1021a |
| Nebraska, 82, 83 Nevada, 534, 536 New Jersey, 405, 406 New York, 751, 988 | Ranunculus, 149, 424, 580, 759 |
| New Jersey, 405, 406 | Rape, 49, 469 |
| New York, 751, 988 | Rattle Weed. See Crotalaria |
| New York City, 863 North Carolina, 497, 626 | Raphanus, 606 Rhizopus, 192 |
| North Dakota, 1004 | Rhododendron, 800, 1071 |
| Ohio, 878, 913, 914 | Rhus, 3, 11, 15, 30, 53, 76, 124, 13 |
| Pennsylvania, Erie, 395 | 474, 476, 485, 491, 517, 525, 53 |
| Rhode Island, 47 | 539, 613, 637, 639a, 678, 741, 75 |
| Vermont, 518 | 138, 146, 181, 223a, 422, 426, 43, 474, 476, 485, 491, 517, 525, 53 539, 613, 637, 639a, 678, 741, 75, 771, 772, 783, 784, 790, 868, 90, 949, 951, 955, 965, 969, 972, 974 |
| America, South, Brazil, 143, 571 Chili, 787 | 1025, 1038, 1044, 1056. |
| Chili, 787 | Biblioghaphy on Rhus Poisoning |
| Guiana, 747, 898 Surinam, 747 | 1025, 1026, 1027. Rhus and Bacterial Infection, 136 |
| West Indies, | Ricin, 508a, 561 |
| Cuba, 180 Porto Rico, 185, 761 | Ricinus, 85, 220, 229, 683 |
| Asia, and Adjacent Islands | Robinia, 811 Roots, Poisonous, 836 |
| Afghanistan, 26 | Rosaceae, 386 |
| China, 218 India, 888, 947 | Rumex, 1078 |
| Bombay, 543 | Sakalava, 438, 439, 765 |
| Japan, 948, 998 Malay, 69, 225 | Sale of Poisons, 93, 280, 448 |
| Philippines, 43 | Salikounda, 440 Sambucus, 474 |
| Australia, 46, 282, 634, 635, 636, | Sanguinaria, 474 |
| 684, 996 | Santonin, 345 |
| Victoria, 282 Europe, 445 | Saponin, 45, 372, 896 Savanna Flower. See Echites |
| Austria, 564 | Sawdust, Poisonous, 271 |
| Belgium, 139 | Scarlet Fever Poison, 120 |
| Central Europe, 445 Denmark, 377 | Seeds, Poisonous Beans, 115 |
| England (also Great Britain), | Colchicum, 302 |
| 21, 457, 515 France 131, 344, 973, 1015 | Cotton, 215 |
| France, 131, 344, 973, 1015 Germany, 112, 245, 342, 353, 373, 383, 402, 449, 454, 538, | Croton, 302 Cynoglossum, 883 |
| 373, 383, 402, 449, 454, 538, | Eriobotrya, 458 |
| 556, 778, 792, 881, 904, 1073a, 1087 | Garcia, 152 Larkspur, 79 |
| Colonies, 562 | Leguminosae, 889 |
| Duisberg, 99 | Lupine, 141 |
| Thuringia, 726 Schwarzburg-Rudolstadt, 926 | Lychnis, 589 Nux vomica, 302 |
| Hungary, 393, 566 | Omphalea, 152 |
| Netherlands, North, 666 | Rape, 49 Ricinus, 85, 302 |
| Poland, 73 Saxony, 275 | Sabadilla, 302 |
| Silesia, 941 | Simarubaceae, 185 |
| Switzerland, 446, 447, 470, 1015 Russia, 869 | Sinapis, 302 Vicia, 80 |
| Potato, 617, 627 | Wormwood, 100 |
| Primula, 701, 702, 831 | Septicaemia, 320 |
| Printhogalum. See Ornithogalum Prulaurocerasin, 459, 460 | Shepherd's Purse, 532 Sicyos, 404 |
| Prunus, 459, 4/4, 1014. Also see | Silage, 208 |
| Cherry | Simarubaceae, 199 |
| | |

Sinapis, 860 Sium, 610a, 777 Sneezeweed, 641 Snowdrop, 601 Snowdrop, 601 Solanaceae, 674 Solanum, 283, 474 Sophora, 190, 798 Sorghum, 24, 48, 767, 1022 Spigelia, 295, 315, 846, 954, 979 Sporotrichosis, 3a, 90a, 135a, 145a, 536b, 727a, 727c, 859a, 993a Spruce, 752 Spurge, 568 Staphisagria. See Larkspur Staphisagria. See Larkspur Star-anise, 222, 242, 494 Stellaria media, 150 Stomatitis, 298, 671 Strawberry, 814b Streptococcus, 174, 176, 561 Strophantus, 311, 329 Strychnin, 283, 376, 658, 1045 Strychnos, 133, 241, 436 Stylophorum, 886 Sugar Beets, 665 Suicide, 688 Also see Sumach, 491, 951, 1050. Rhus. Symptoms from Poisoning, 37, 37a, 39, 92, 93, 118, 376, 382, 552, 932. Also see General Treatises upon Poisoning

Tanghin, 311, 437, 438
Tartaric Acid, 1078
Taxus, 150a, 461, 901, 959
Tea, 953
Temperature affecting Toxicity of Poisons, 418, 872, 1088, 1089
Templetonia, 190

Tetanus. See Bacillus tetani Thalictrum, 505, 506 Therapeutics, 39, 55a, 288, 783, 893, **953**, 10**7**0 Thrush, 379, 607, 608, 609 Tobacco. See Nicotin Tonka Bean, 440 Toot Poison, 605 Toxicology, General, 155a, 551, 553a, 567, 597, 651, 724, 796, 828, 918, 937 Vegetable, 173, 177, 197, 553, 561, 851, 107 Veterinary, 592, 706, 715 Toxins, 718 Treatment for Poisons, 39, 595, 646, 689, 693, 722, 804, 933, 956, 962, Trees, Malignant Effect of, 838 Trichophyton, 401, 870, 975 Tropaeolum, 389 Tulip Grass, 109 Tunas, 415 Turpentine, 958

Umbelliferae, 547, 603, 803 Upas, 241 Ustilago, 276

VERATRUM, 58, 114, 221, 302, 313, 760 Verbascum, 669 Verbena, 104 Veronal, 328

WATER BLOOM, 650 Weeds, Poisonous, 734

YEW. See Taxus.

BIBLIOGRAPHY OF POISONOUS PLANTS

BY HARRIETTE S. KELLOGG

| 1. | Abbott, A. C | Principles of Bacteriology; a Practical Manual for Students and Physicians. 641. Phila- delphia and New York. Ed. 8. |
|-------------|---------------------------|--|
| 2. 3. | Acree, S. F. and Syme, W. | Vet. Rev. 13:369. London. |
| 32 | | Some Constituents of the Poison Ivy Plant. Am. Chem. Jour. 36:301-341. Sporttrichosis a Resumé of the Literature |
| Ja. | | Relating to Sporotrichal Infection of the Skin. Brit. Jour. or Derm. 20:296. |
| 4. 5. | Aggio, C | Liber de venenis. 101. Venice, 1492. Two Interesting Outbreaks of Vegetable Poisoning. Am. Vet. Rev. 32:368. |
| 5a. | | Certain Facts Regarding the Poison-lore of the Hindus. Trans. India Med. Cong. 1894:507-510. Calcutta. |
| 6. | Albert | On Colchicum Poisoning. See Zeitsch. f. Staatsarznk. 84:202. 1862. |
| | | On Colchicum Poisoning. See Boll.d.sc.med. |
| 8b. | Alberts | Het Pollen-asthma. Amsterdam. 1903. |
| 8. | Alkaloids in Plants | Botanical Magazine. 16: (Written in Japa- |
| 9. | Allen, Timothy F | nese.) Allen Encyclopedia. The Encyclopedia of Materia Medica. A Record of the Positive Effects of Drugs, etc. 10 vols. Philadelphia. 1877-1879. |
| 10. | Alpers, W. C | The Antidote Cabinet. Phar. Era. 22:136-138. |
| 10a. 11. | Alsberg, C. L | |
| 12. | Alway, F. J. and Peters, | Losses from Cornstalk Disease in Custer Co., |
| | A. T | Nebraska, during the Winter of 1906-1907. Press. Bull. Neb. Agr. Exp. Sta. 27:4. |
| 13. | Anderson, F. W | Poisonous Plants and the Symptoms they Produce. Botanical Gazette. 14:180-181. |
| 14. | Andrews, F. M | The Effect of Alkaloids and other Vegetable-Poisons upon Protoplasm. Ind. Acad. Sci. 1905:195. |
| 14a. 15. | | See Graves, C. B. Chemical Observations on Rhus Toxicoden- |
| 16. | Anonymous | dron. Therapeutic Gazette. 13:676. Acorn Poisoning. Vet. Rec. London. 13:359. |
| 17. 18. | Anon | Alkaloids in Plants. Bot. Mag. 16: Are Birds Affected by Eating Poisonous Food? |
| 19. | | Scientific American 79:424- Cassava Poisoning. Agr. News. Barbadoes. |
| 20. | | 1:5. On Death Ensuing from Rhus Poisoning. See- |
| 20. | | Boston Med. and Sur. Jour. 137:302. |

| 21. | Anon | English Poisonous Plants. Gentleman's Magazine. No. 1755. |
|-------------|----------------------------|--|
| 22. | Anon | Flowers that are Poisonous. Current Litera- |
| 23. | Anon | ture. 33:331-333. Giftigkeit der Amerikablüthen. Pharmaceu- |
| | | tische Centralhalle. 1903:136. |
| 24. | | Green Sorghum Poisoning. West Indian Bulletin 3:326-333. |
| 25. | Anon | Die Immunität von Pflanzen gegen ihr eigenes Gift. Zeitsch.f.ang. Mikr. 13:60-62. Leip- sic, 1907. |
| 26. | | Medical Plants in Afghanistan. Jour. Applied |
| 27. | Anon | Science. 1880 :142. Native Poisonous Plants. Harper's Bazaar. |
| 27a. | Anon | 34:941. Poisoning by Water Hemlock. See Med. |
| | | Times and Gazette 1855:51 |
| 28. | Anon | Our Poisonous Plants. Pathfinder 6:7. |
| 29. | | Pellagra. Boston Evening Transcript. Aug. 18. 1909. |
| 30. | Anon. | Plant Poisons. Sci. Am. 93:361. |
| 31. | Anon. | Plant Poisons, Gardener's Chronicle. 45: 111; 366. |
| 32. | Are Birds Affected by Eat- | |
| | ing Poisonous Food? | Sci. Am. 79 :424. Some Algae of Minnesota Supposed to be Poi- |
| 33. | Arthur, J. C | Some Algae of Minnesota Supposed to be Poi- |
| | | sonous. Bull. Minn. Acad. Nat. Sci. 2:4. |
| | | (Appendix) 3:97. Second Report in Ann. Rept. Minn. Univ. 4: Supplement 1. Also |
| | | Report U. S. Dept. Agr. 1897 :97. |
| 34. | | A Supposed Poisonous Seaweed in the Lakes |
| | | of Minnesota. Proc. Am. Assoc. Adv. Sci. 32:305. (Abs.) |
| 35. | Asahina, Y | Ueber die Alkaloide von Dicentra pusilla. |
| 2 = | | Sieb. et Zucc. Arch. Pharm. 247:201, 1909. |
| 35a. | Ashworth | On Cocain. See Lancet 1889:273. Mushrooms, Edible, Poisonous, etc. 322. Ith- |
| 30. | Atkinson, G. F | aca, N. Y. 1901. Ed. 2. |
| 37. | Atkinson, V. T | Action of Poisons. Special Report. Diseases |
| | | of Cattle and on Cattle Feeding. U. S. Dept. Agr. Bur. An. Ind. 1896 :63-76. |
| 37a. | | Poisons and Poisoning. In Special Report, |
| | | Diseases of Cattle, U. S. Dept. Agr. 1908: |
| 20 | | 53-69. Revised by Leonard Pearson. |
| <i>3</i> 8. | Auclair, J | Les poisons du bacille-tuberculeaux humain et la degenerescence caseuse. Revue de la Tu- |
| | | berculose, 1898, |
| 39. | Aurand, S. Herbert | Botanical Materia Medica and Pharmacology; |
| | | giving the Botany, Pharmacology, Physio- |
| | | logical Action, Therapeutic Range, Treatment of Poisons and Antidotes. 406. Chi- |
| | | cago, 1899. |
| 40. | Avery, Samuel | See Peters, A. T. |
| 41. | Bacon, Alice E | An Experiment with the Fruit of Red Baneberry. Rhodora 5:77-79. |
| 42. | Bacon, C. W | Handbook to Accompany Chart of Poisonous |
| 43. | Bacon, R. F | Plants, 30. London, 1899. Philippine Arrow Poisons. Philippine Jour. |
| 44. | , | Sci. 3-A:41:44. The Physiologically Active Constituents of |
| | | Certain Philippine Medical Plants. Phil. |
| | | Jour. Sci. 1 :1007-1036. |

| 45. | Baccn, R. F. and Marshall, H. I. | The Toxic Action of Saponin. Phil. Jour. |
|------------|--|--|
| 46. | Bailey, F. M. and Gordon, | Sci. 1:1037-1042. |
| | P. R | to Stock. Gov't Printer, Brisbane. 4:112. |
| 47. | Bailey, W. W | Rhode Island's Poisonous Plants. Trans. R. I. Med. Soc. 6:64-72. |
| 47a. | | I. Med. Soc. 6:64-72. Histoire des Plantes. 13 vols. Paris. 1868- 1896. |
| 48. | Ball, C. R | Saccharine Sorghums for Forage, Far. Bull. |
| | | U. S. Dept. Agr. 246:37. On Aconite Poisoning. See Witthaus and Becker's Medical Jurisprudence, 4:648. Abs. of Article in Ann. Univ. d. Med. Milan. 3:635. 1840. |
| | | cases of Poisoning from Rape-Seed Cakes. Ugeskr. Landw. 43:589-591; 604-605, 1897. Abs. in Exp. Sta. Rec. 9:954. |
| 50. 51. | Barbosa, R. IL | Juiraery du curare. 180. 7pl. Brussels. 1903. The Ravages of Bacillus Anthracis in California. South Cailfornia Practitioner. 1894 : 121-153. |
| 52. | Barger, G. and Carr, F. HN | ote on Ergot Alkaloids. Phar. Jour. 23: 257. |
| 52a. | BarkerO | 237. on Cocain. See Am. Jour. Opth. 10:342. 1893. |
| 53. | Barrows, W. BO | In the Non-poisonous Quality of the Ripe- Fruit of Rhus Toxicodendron. See. Bull. |
| 54. | Barth, HermanS | Div. Orn. Mam. U. S. Dept. Agr. 6 :85. tudien über den mikrochemischer Nachweis von Alkaloiden in pharmaceutisch verwendeten Drogen. Botanischer Centralblatt 75 : 225-231; 261-267; 292-300; 326-644; 369-378; 401-409. |
| 55. | | Jucormycoses, Spontanées, Historique. I, Les Mucorinées chez l'homme. Arch. de Para- |
| 55a. | Barthelow, R | sitologie. 1903. Practical Treatise on Materia Medica and Therapeutics. Ed. 7. New York. 1889. |
| 56. | Barton, W. CA | Flora of North America. Philadelphia. |
| 57. | v | 1820. egetable Materia Medica of the United States; or Medical Botany. 2 vols. 1817- |
| 58. | Bastin, E. SSi | 1818. tructure of Veratrum viride. American Jour. |
| 59. 60. | Bastin, E. S. and Trimble. Co. Batchelor, J. and Miyabe, | of Pharmacy. 77:196-203. |
| | K | Medicinally by the Aines. Trans. Asiatic Society of Japan. 21. Abs. in Proc. Am. Phar. Assoc. 44:501-505. |
| 61. | Baume, MPe | Mal. Mentales. 1881. No. 3. Abs. in Ed. Vet. Rev. 4:704. |
| 62. | Baumgarten, PLe | ehrbuch der pathologischen Mycologie. 1 Hfte 220 Braunschweig 1836 |
| 63. | Sı | r les microorganismes morbigènes d'origine végétale. Traduit par Steinen. Jour. de médecine de chirurgie et de pharmacologie. 1886. Mai. |

| 64. | Baumgarten, P. and Müller, | V b. The second dating 7 Taken |
|--------------------|---|---|
| 65. | • | Versuche über accomodative Züchtung von Schimmelpilzen. Berliner klin. Woch, 1882 . The Legal Responsibility of the Pharmacist in the Sale of Poisons and Dangerous Sub- stances. Pharm. Rev. 19 :292-298; 338- |
| 66. 67. | Becker, T. C | 344. See Witthaus, R. A. On the Poisonous Plants indigenous to Cali- |
| 68. | Beilstein, F | fornia. Repr. Handbuch der organischen Chemie. 4 vols. |
| 69. | | Hamburg and Leipzig. 1895-1899. Recherches pharmacologiques sur quelques poisons par les negritos de l'archipel Malais. Archives Italiennes de Biologie. Fasc. 1. 1899. |
| 70. | Bennett, A. W | On Vegetable Growths as Evidence of the Purity or Impurity of Water. St. Thomas Hospital Report. 20:8, |
| 7 0a. | Bentham, George | Illustrated Handbook of the British Flora, 1: 503. 2:505-1076. |
| <i>7</i> 1. | Bentley, Robert and Tri- men, Henry | |
| 72. | Bentley, W. H | in Medicine. 4 vols. London. 1880. Poisoning by Damiana—Recovery. Therap. Gaz. N. S. 4:57, 1883. |
| 73. | Berdan, T. | Tablica grzybon trujaeych w kraju. (Table of poisonous Fungi in Poland.) Warsau. |
| 74. | Berg | 1888. Ueber die Schwimmen bei Kindern, Bremen, 1842. (Clinique des hôpitaux des enfants de Paris, 1842.) |
| 74. | Berge, Fr. and Riecke, V. | Giftpflanzenbuch; oder Allgemeine und be- |
| | Α | sondere Naturgeschichte sämmtlicher in- ländischen so wie der wichtigsten ausländi- schen phanerogamischen und kryptogami- schen Giftgewächse, mit treuen Abbildungen sämmtlicher inländischen und vieler aus- ländischen Gattungen. 11-329. 72pl. Stutt- gart. 1845. |
| 76. 77. | Beringer, G. M Berlman, M | Rhus Poisoning. Am. Jour. Phar. 68:18-20. |
| 7 8. | | Celastrus scandens, L. Amer. Jour. Phar. 4: |
| 79. | Bernon | Empoisonnement par les semences de Staphisaigre. Jour. de Phar. et de Chimie. 1882. |
| 80. | Bertrand, G. | La vicianine, nouveau glucoside cyanhydrique contenu dans les graines de vesce. Bull. Soc. chim. France, 4 ser. 1:151-154. |
| 81. 81a. 82. | Bertuch, F. Besley, F. W. Bessey, C. E. | See Guenther, J. See Shreve, F. Injuriousness of Porcupine Grass. Am. Nat. |
| 82a. | | 18:929. Notes on Poisonous Plants of Nebraska. Proc. |
| 83. | | Soc. Prom. Agr. Sci. 23:34. Poisonous Weeds. Rept. Neb. Agr. Exp. Sta. 16:14-62. |
| 83a. | Besta, C. | |

| 84. | Bigelow, JacobAmerican Medical Botany. A Collection of the Native Medicinal Plants of the United States. 3 vols. Boston. 1817-1820. |
|--------------|--|
| 85. | Bishpam, W. N |
| 85a. 86. | Bissell, C. H |
| | Rept. Ind. Agr. Exp. Sta. 1899 :44-45. Bjerregard, A. P See Hare, R. F. Black, O. F. and Alsberg, |
| | C. L |
| 87. | Blanchard, Raphael Accidents causés par une graminée américaine. |
| 88. | Archives de Parasitologie. 10:187-194Quelques cas anciens d'actinomycose. Arch. |
| 89. | de Parasit. 2:362-376. 1899. Blankinship, J. W The Loco and other Poisonous Plants of Mon- |
| 90. | tana. Bull. Mon. Agr. Exp. Sta. 42:75-104 |
| 90a. | Mon. Agr. Exp. Sta. 56:36. Bloch, BrunoDie Sporotrichose. Mediz. Klinik. 1909. |
| | Block |
| 91a. | Blodgett, F. HSee Shreve, F. |
| 92. | Blyth, A. W |
| | Old and Modern Poison Lore. |
| 93. | W |
| 94. | larged and Rewritten. 772. London. 1906. Bodin, E |
| <i>7</i> 4. | Actiorion Quinckeanum. Archiv. de Parasit. |
| 95. | Bodin, E. and Gautier, L The Toxins Produced by Aspergillus fumigatus. Annales de l'Institut Pasteur, 20:209-224. Abs. in Exp. Sta. Rec. 18:8. |
| 96. | Boehm Cicutoxin. See Archiv. f. exp. Path. p. Phar- |
| 97. | makol. 5:279-310. Böhm, R., Naunyn, B., and |
| | von Boeck, H |
| 98. | Boehmer, G. RCommentatio de plantis segeti infestis: ac- |
| | cidit commentatio de plantis auctoritate publica exstirpandis, custodiendis, et a foro proscribendis. 198. Wittemberg. 1785. |
| 98a | |
| 99. | tur. 12. Wittemberg. 1787. Boeninger, T. K De plantis venenatis et speciatim de plantis venenatis agri Duisburgensis. 133. Duis- |
| 100. | burg. 1790. Bond, A. RPoisoning by Wormwood Seed. Phar, Jour. |
| | Ser. 4. No. 1483. |
| 101. 102. | Boole, L. E |
| 103. | Bouchard, Charles Upon the Role of Poisons of Microbic Origin in Infectious Maladies. Gaz. hebdom. de Med. et de Chim. 1889:120-122. |
| | |

| 104. Bourdier, L | | |
|--|-------|--|
| C. R. Soc, Biol. Paris. Nov. 1. 1907. Bourquelot, E. and Danjon, E. Sur la presence d'un glucoside cyanhydrique dans les fueilles du sureau. C. R. Acad. Sc. Paris. 1905. July 3. 106. Sur un empoisonnement par le fausse oronge, survenu à Bois-le-Roi, Seine-et-Marne, le 6 septembre 1896. Bull. de la Soc. myc. de France. 12:146-150. 107. Boutet. and Clement. Empoisonnement par la fleur Oronze. Bull. de la Soc. des Nat. Sci. de l'Ocust de la France. 1901:45. 108. Boutmy, E. See Brouard, P. 109. Bowbill Tulip-grass Poisoning. Vet. Rec. London.13: 229. 110. Bewer, F. O., Translator. See De Bary, A. 111. Braithwaite, P. F. Suicidal Poisoning with Salts of Lemon: Perforation of the Stomach. Brit. Med. Jour. 1:183. (1905.) 112. Brandt, J. F. and Ratzeburg. Deutschlands phanerogamische Giftgewächse in Abbildungen u. Beschreibungen. Ed. 2. 200. 48 pl. Berlin. 1838. 113. Brebeck. See Fischer, A. 114. Bredeman, G. Alkaloids of Veratrum viride and their Determination. Phar. Jour. 22:283. 115. Bretin, Evesque and Verdier. Haricots toxiques dits de Hongrie. Jour. Phar. et Chim. 26:348. 115a. British Pharmacopoeia. 1898. (Ph. Brit.) 116. Britton, N. L. and Brown, A. Manual of the Flora of the Northern States and Canada. 1080. New York. 1901. 117. Britton, N. L. and Brown, A. Illustrated Flora of the Northern United States and the British Possessions from Newfoundland to the Parallel of the Southern Boundary of Virginia, and from the Atlantic Ocean westward to the 102d Meridian. Vol. 1:612. f. 1:1425; II: 643. f. 1426-2829; III:588 f. 2893-4081. New York. 1896-1898. 118. Brodie, E. C. Further Experiments and Observations on the Arliantic Ocean westward to the 102d Meridian. Vol. 1:612. f. 1:1425; III: 643. f. 1426-2829; III:588 f. 2893-4081. New York. 1896-1898. 119. Brodie, D. A. Poison Parsnip in Western Washington. Bull. Wash. Agr. Exp. Sta. 45:12. 120. Brooke, J. Activity of the Scarlet Fever Poison After a Year. Med. Rec. 2:126. 121. Brown, A. See Britton, N. L. 122. Brown, A. See Britton, N. L. 123. Bro | 104. | Bourdier, L |
| Bourquelot, E. and Danjon, E. Sur la presence d'un glucoside cyanhydrique dans les fueilles du sureau. C. R. Acad. Sc. Paris. 1905. July 3. 106. Sur un empoisonnement par le fausse oronge, survenu à Bois-le-Roi, Seine-et-Marne, le 6 septembre 1896. Bull. de la Soc. myc. de France. 12:146-150. 107. Boutet. and Clement. Empoisonnement par la fleur Oronze. Bull. de la Soc. des Nat. Sci. de l'Ocust de la France. 1901:45. 108. Boutmy, E. See Brouard, P. 109. Bowbill Tulip-grass Poisoning. Vet. Rec. London.13: 229- 110. Braithwaite, P. F. Suicidal Poisoning with Salts of Lemon: Perforation of the Stomach. Brit. Med. Jour. 1:183. (1905.) 112. Brandt, J. F. and Ratzeburg. Deutschlands phanerogamische Giftgewächse in Abbildungen u. Beschreibungen. Ed. 2. 200. 48 pl. Berlin. 1838. 113. Brebeck. See Fischer, A. 114. Bredeman, G. Alkaloids of Veratrum viride and their Determination. Phar. Jour. 22:283. 115. Bretin, Evesque and Verdier. Haricots toxiques dits de Hongrie. Jour. Phar. et Chim. 26:348. 115a. British Pharmacopoeia. 1898. (Ph. Brit.) 116. Britton, N. L. Manual of the Flora of the Northern States and Canada. 1080. New York. 1901. 117. Britton, N. L. and Brown, A. Illustrated Flora of the Northern United States and the British Possessions from Newfoundland to the Parallel of the Southern Boundary of Virginia, and from the Atlantic Ocean westward to the 102d Meridian. Vol. 1:612. f. 1:1425; III: 643. f. 1426-2892; III:588 f. 2893-4081. New York. 1896-1898. 118. Brodie, E. C. Further Experiments and Observations on the Action of Poisons on the Animal System. Phil. Trans. 102:218. 119. Brode, D. A. Poison Parsnip in Western Washington. Bull. Wash. Agr. Exp. Sta. 45:12. 120. Brown, A. See Britton, N. L. 121. Brown, A. See Britton, N. L. 122. Brown, A. See Britton, N. L. 123. Brown, E. L. A Case of Actinomycosis. Chicago Medical Record. 1894:251. 124. Brown, S. A. On Rhus Poisoning. See Med. Rec. 30:221. 125. Brünnich, J. C. Hydrocyanic Acid in Fodder Plants. Jour. Chem. Soc. 1996:788-796. | | C. R. Soc. Biol. Paris. Nov. 1. 1907. |
| E | | |
| 106 | | E |
| 107. Boutet, and Clement Empoisonnement par la fleur Oronze. Bull, de la Soc. des Nat, Sci. de l'Ocust de la France. 1901;45. 108. Boutmy, E | 106. | Sur un empoisonnement par le fausse oronge, survenu à Bois-le-Roi, Seine-et-Marne, le 6 septembre 1896 . Bull. de la Soc. myc. de |
| 108. Boutmy, E | 107. | Boutet. and Clement Empoisonnement par la fleur Oronze. Bull. de la Soc. des Nat. Sci. de l'Oeust de la France. |
| 109. Bowbill | 108. | |
| 111 Braithwaite, P. F | | Bowbill |
| burg Deutschlands phanerogamische Giftgewächse in Abbildungen u. Beschreibungen. Ed. 2. 200. 48 pl. Berlin. 1838. 113. Brebeck See Fischer, A. 114. Bredeman, G Alkaloids of Veratrum viride and their Determination. Phar. Jour. 22:283. 115. Bretin, Evesque and Verdier Haricots toxiques dits de Hongrie. Jour. Phar. et Chim. 26:348. 115a. British Pharmacopoeia 1898. (Ph. Brit.) 116. Britton, N. L Manual of the Flora of the Northern States and Canada. 1080. New York. 1901. 117. Britton, N. L. and Brown, A Illustrated Flora of the Northern United States and the British Possessions from Newfoundland to the Parallel of the Southern Boundary of Virginia, and from the Atlantic Ocean westward to the 102d Meridian. Vol. 1:612. f. 1-1425; II: 643. f. 1426-2892; III:588 f. 2893-4081. New York. 1896-1898. 118. Brodie, E. C Further Experiments and Observations on the Action of Poisons on the Animal System. Phil. Trans. 102:218. 119. Brodie, D. A Poison Parsmip in Western Washington. Bull. Wash. Agr. Exp. Sta. 45:12. 120. Brooke, J Activity of the Scarlet Fever Poison After a Year. Med. Rec. 2:126. 121. Brouard, F. and Boutmy, E. Ptomaines et alcaloides végétaux. Bull. de l'Acad. de Med. 1881:19. 122. Brown, A See Britton, N. L. 123. Brown, E. L A Case of Actinomycosis. Chicago Medical Record. 1894:251. 124. Brown, S. A On Rhus Poisoning. See Med. Rec. 30:221. 125. Brünnich, J. C Hydrocyanic Acid in Fodder Plants. Jour. Chem. Soc. 1903:788-796. 126. Buchanan, R. E Notes on the Algae of Iowa. Proc. Ia. Acad. | | Braithwaite, P. FSuicidal Poisoning with Salts of Lemon: Perforation of the Stomach. Brit. Med. Jour. |
| in Abbildungen u. Beschreibungen. Ed. 2. 200. 48 pl. Berlin. 1838. 113. Brebeck | 112. | Brandt, J. F. and Ratze- |
| 114. Bredeman, G | | in Abbildungen u. Beschreibungen. Ed. 2. 200. 48 pl. Berlin. 1838. |
| termination. Phar. Jour. 22:283. 115. Bretin, Evesque and Verdier | | Brebeck See Fischer, A. |
| 115. Bretin, Evesque and Verdier | 114. | |
| 115a. British Pharmacopoeia 1898. (Ph. Brit.) 116. Britton, N. L Manual of the Flora of the Northern States and Canada. 1080. New York. 1901. 117. Britton, N. L. and Brown, A Illustrated Flora of the Northern United States and the British Possessions from Newfoundland to the Parallel of the Southern Boundary of Virginia, and from the Atlantic Ocean westward to the 102d Meridian. Vol. I:612. f. 1-1425; II: 643. f. 1426-2892; III:588 f. 2893-4081. New York. 1896-1898. 118. Brodie, E. C. Further Experiments and Observations on the Action of Poisons on the Animal System. Phil. Trans. 102:218. 119. Brodie, D. A. Poison Parsnip in Western Washington. Bull. Wash. Agr. Exp. Sta. 45:12. 120. Brooke, J. Activity of the Scarlet Fever Poison After a Year. Med. Rec. 2:126. 121. Brouard, F. and Boutmy, E. Ptomaines et alcaloides végétaux. Bull. de l'Acad. de Med. 1881:19. 122. Brown, A. See Britton, N. L. 123. Brown, E. L. A Case of Actinomycosis. Chicago Medical Record. 1894:251. 124. Brown, S. A. On Rhus Poisoning. See Med. Rec. 30:221. 125. Brünnich, J. C. Hydrocyanic Acid in Fodder Plants. Jour. Chem. Soc. 1903:788-796. 126. Buchanan, R. E. Notes on the Algae of Iowa. Proc. Ia. Acad. | 115. | Bretin, Evesque and Ver- dier |
| 116. Britton, N. L. Manual of the Flora of the Northern States and Canada. 1080. New York. 1901. 117. Britton, N. L. and Brown, A. Illustrated Flora of the Northern United States and the British Possessions from Newfoundland to the Parallel of the Southern Boundary of Virginia, and from the Atlantic Ocean westward to the 102d Meridian. Vol. I:612 f. 1-1425; II: 643. f. 1426-2892; III:588 f. 2893-4081. New York. 1896-1898. 118. Brodie, E. C. Further Experiments and Observations on the Action of Poisons on the Animal System. Phil. Trans. 102:218. 119. Brodie, D. A. Poison Parsnip in Western Washington. Bull. Wash. Agr. Exp. Sta. 45:12. 120. Brooke, J. Activity of the Scarlet Fever Poison After a Year. Med. Rec. 2:126. 121. Brouard, F. and Boutmy, E. Ptomaines et alcaloides végétaux. Bull. de l'Acad. de Med. 1881:19. 122. Brown, A. See Britton, N. L. 123. Brown, E. L. A Case of Actinomycosis. Chicago Medical Record. 1894:251. 124. Brown, S. A. On Rhus Poisoning. See Med. Rec. 30:221. 125. Brünnich, J. C. Hydrocyanic Acid in Fodder Plants. Jour. Chem. Soc. 1903:788-796. 126. Buchanan, R. E. Notes on the Algae of Iowa. Proc. Ia. Acad. | 115a. | British Pharmacopoeia1898. (Ph. Brit.) |
| 117. Britton, N. L. and Brown, A Illustrated Flora of the Northern United States and the British Possessions from Newfoundland to the Parallel of the Southern Boundary of Virginia, and from the Atlantic Ocean westward to the 102d Meridian. Vol. I:612. f. 1-1425; II: 643. f. 1426-2892; III:588 f. 2893-4081. New York. 1896-1898. 118. Brodie, E. C. Further Experiments and Observations on the Action of Poisons on the Animal System. Phil. Trans. 102:218. 119. Brodie, D. A. Poison Parsnip in Western Washington. Bull. Wash. Agr. Exp. Sta. 45:12. 120. Brooke, J. Activity of the Scarlet Fever Poison After a Year. Med. Rec. 2:126. 121. Brouard, F. and Boutmy, E. Ptomaines et alcaloides végétaux. Bull. de l'Acad. de Med. 1881:19. 122. Brown, A. See Britton, N. L. 123. Brown, E. L. A Case of Actinomycosis. Chicago Medical Record. 1894:251. 124. Brown, S. A. On Rhus Poisoning. See Med. Rec. 30:221. 125. Brünnich, J. C. Hydrocyanic Acid in Fodder Plants. Jour. Chem. Soc. 1903:788-796. 126. Buchanan, R. E. Notes on the Algae of Iowa. Proc. Ia. Acad. | | Britton, N. L |
| States and the British Possessions from Newfoundland to the Parallel of the Southern Boundary of Virginia, and from the Atlantic Ocean westward to the 102d Meridian. Vol. I:612. f. 1-1425; II: 643. f. 1426-2892; III:588 f. 2893-4081. New York. 1896-1898. 118. Brodie, E. C. Further Experiments and Observations on the Action of Poisons on the Animal System. Phil. Trans. 102:218. 119. Brodie, D. A. Poison Parsnip in Western Washington. Bull. Wash. Agr. Exp. Sta. 45:12. 120. Brooke, J. Activity of the Scarlet Fever Poison After a Year. Med. Rec. 2:126. 121. Brouard, F. and Boutmy, E. Ptomaines et alcaloides végétaux. Bull. de l'Acad. de Med. 1881:19. 122. Brown, A. See Britton, N. L. 123. Brown, E. L. A Case of Actinomycosis. Chicago Medical Record. 1894:251. 124. Brown, S. A. On Rhus Poisoning. See Med. Rec. 30:221. 125. Brünnich, J. C. Hydrocyanic Acid in Fodder Plants. Jour. Chem. Soc. 1903:788-796. 126. Buchanan, R. E. Notes on the Algae of Iowa. Proc. Ia. Acad. | 117. | Britton, N. L. and Brown, |
| 118. Brodie, E. C. Further Experiments and Observations on the Action of Poisons on the Animal System. Phil. Trans. 102:218. 119. Brodie, D. A. Poison Parsnip in Western Washington. Bull. Wash. Agr. Exp. Sta. 45:12. 120. Brooke, J. Activity of the Scarlet Fever Poison After a Year. Med. Rec. 2:126. 121. Brouard, F. and Boutmy, E. Ptomaines et alcaloides végétaux. Bull. de l'Acad. de Med. 1881:19. 122. Brown, A. See Britton, N. L. 123. Brown, E. L. A Case of Actinomycosis. Chicago Medical Record. 1894:251. 124. Brown, S. A. On Rhus Poisoning. See Med. Rec. 30:221. 125. Brünnich, J. C. Hydrocyanic Acid in Fodder Plants. Jour. Chem. Soc. 1903:788-796. 126. Buchanan, R. E. Notes on the Algae of Iowa. Proc. Ia. Acad. | | States and the British Possessions from Newfoundland to the Parallel of the Southern Boundary of Virginia, and from the Atlantic Ocean westward to the 102d Meridian. Vol. I:612. f. 1-1425; II: 643. f. 1426-2892; III:588 f. 2893-4081. New York. 1896-1898. |
| 119. Brodie, D. A Poison Parsnip in Western Washington. Bull. Wash. Agr. Exp. Sta. 45:12. 120. Brooke, J. Activity of the Scarlet Fever Poison After a Year. Med. Rec. 2:126. 121. Brouard, F. and Boutmy, E. Ptomaines et alcaloides végétaux. Bull. de l'Acad. de Med. 1881:19. 122. Brown, A. See Britton, N. L. 123. Brown, E. L. A Case of Actinomycosis. Chicago Medical Record. 1894:251. 124. Brown, S. A. On Rhus Poisoning. See Med. Rec. 30:221. 125. Brünnich, J. C. Hydrocyanic Acid in Fodder Plants. Jour. Chem. Soc. 1903:788-796. 126. Buchanan, R. E. Notes on the Algae of Iowa. Proc. Ia. Acad. | 118. | Brodie, E. CFurther Experiments and Observations on the Action of Poisons on the Animal System. |
| 120. Brooke, J | 119. | Brodie, D. A |
| 121. Brouard, F. and Boutmy, E. Ptomaines et alcaloides végétaux. Bull. de l' Acad. de Med. 1881:19. 122. Brown, A. See Britton, N. L. 123. Brown, E. L. A Case of Actinomycosis. Chicago Medical Record. 1894:251. 124. Brown, S. A. On Rhus Poisoning. See Med. Rec. 30:221. 125. Brünnich, J. C. Hydrocyanic Acid in Fodder Plants. Jour. Chem. Soc. 1903:788-796. 126. Buchanan, R. E. Notes on the Algae of Iowa. Proc. Ia. Acad. | 120. | Brooke, J |
| Brown, A | 121. | Brouard, F. and Boutmy, E. Ptomaines et alcaloides végétaux. Bull. de l' |
| Brown, S. A | | Brown, A |
| 126. Buchanan, R. E Notes on the Algae of Iowa. Proc. Ia. Acad. | | Brown, S. AOn Rhus Poisoning. See Med. Rec. 30 :221. Brünnich, J. CHydrocyanic Acid in Fodder Plants. Jour. |
| | 126. | Buchanan, R. E Notes on the Algae of Iowa. Proc. Ia. Acad. |

| 126a. Buchner, H | Ueber Bacteriengifte und Gegengifte. Münchener Medicinisch Wochenschrift. 1893 : |
|-----------------------|--|
| 127. Buchner, Max | 449-452; 480-483. Ueber pathogenetische Wirkungen der Pilz- |
| 127a. Buckley, J. S | keimkörper. Jahrb. med. Wissensch. 1868. See Mohler, J. S. |
| 128. Buckley, S. S | A Special Investigation of the So-called "New Horse Disease" in Maryland. Bull. Md. |
| | Probable Poisoning by Colchicum. Lancet. No. 3001. |
| 130. Buechner, J. G | Histoire des plantes vénéneuses et suspectes |
| 133. Bureau, Ed. | de la France. 398. Paris. 1798. Ed. 2. Sur les Strychnos africains et les plantes servent à empoisonner les armes en Afrique. |
| 134. Burgess, T. J. W | Bull. Mus. Hist, nat. Paris. 1901:418-425. The Beneficial and Toxical Effects of the various Species of Rhus. Canadian Jour. Med. |
| 135. Burke, R. W | Sci. (Phar. Jour. and Trans. 1881.) The Bacillus of the Malignant Oedema. Vet. |
| 135a. Burlew, J. M | Jour. 1890 :246-247. Subcutaneous Abcesses Caused by the Sporo- |
| | thrix Schrenckii. Southern California Pract. 24 :1. |
| 136. Burrill, T. J | On Bacterial Infection and Rhus Poisoning. See Am. Mo. Mic. Jour. 3:192; Proc. Am. Assoc. Adv. Sci. 31:515; Am. Nat. 17:319. |
| 137 | Rhus Poisoning. Garden and Forest 8:386-369. |
| 138. Busey, S. C | On Recurrence of Attacks of Rhus Poisoning without Re-exposure. Am. Jour. Med. Sci. 66:436. |
| 138a. Byron, J. M | A Case of Actinomycosis in Man. N. Y. Med. |
| 139. Caels, T. P | Jour. 2:716. (1889.) De Belgii plantis qualitate quadam hominibus caeterisve animalibus nociva seu venenata praeditis symtomatibus ab earum use pro- |
| | ductis nec non antidotis adhibenis. 66. Brussels, 1774. |
| | On Toxins in Foods. See Strasburger-Medi- |
| | Ueber die Alkaloide der Samen von Lupinus angustifolius und von Lupinus perennis, var. polyphyllus. Archiv. der Phar. 237:566- 595. |
| 142. Cambier, Jacob | See Power, F. B. Das plantas toxicas do Brazil. Rio de Janeiro |
| | 1871. Trans. into French by F. Rey. (Catalogue des plantes toxiques de Bresil. Paris, 1880.) |
| 144. Campbell, H. P | Bacterial Poisoning through Medicine. Am. Jour. Phar. 1890:113-118. |
| 145. Card, F. W | Poisonous Plants. Gar. and For. 9:8. Sur une nouvelle mycose sous-cutanée des équides. Madagascar. 1906. Jour. de Méd. vétér. et de Zootechnie. 60:8-22, 75-90, 148-153. Abs. in Centrbl. f. Bakt. Abt. I. |
| 146. Carpenter, C. R | 45 :610-612. Rhus Toxicodendron. Ther. Gaz. 14 :93. |
| | Aethusa Cynapium. Jour. Roy. Agr. Soc. of Eng. 63 :280. |
| 148 | Lychnis Githago. id. 63:279. |

| 149. 150. | | Ranunculus bulbosus. id. 63 :279. Stellaria media. Is it Poisonous? id. 64 : 308. |
|----------------|---|--|
| 150a. | Carruthers, Wm. and | |
| 151. | Others | Yew Poisoning. id. Ser III. 3:698. The Medical Mystery of Today. (Pellagra.) |
| 151a. | | |
| 1511 | a | A Case of Atropin Poisoning. N. Y. Med Jour. Sept. 5, 1903. |
| 1516. | Cash, J. T. | See Albertoni. The Characters and Actions of the Seeds of Omphalea megacarpa (diandra), Omphalea triandra and Garcia nutans (from Trinidad). Phar. Jour. 81:351-352. |
| 153. | Cassava Poisoning | .Agr. News. Barbadoes. 1:5. .Post-Mortem Pathology. A Manual of Post- |
| 154. | Catter, II. W | Mortem Examinations and the Interpretations to be Drawn therefrom. A Practical Treatise for Students and Practitioners. 551. 6 pl. 183f. Ed. 2. 1905. |
| 154a. | Ceni, Carl and Besta, | . Ueber die Toxine von Aspergillus fumigatus |
| | | und flavescens und deren Beziehung zur Pellagra Central f Allgem Path u Path |
| 155. | Chapman, A. W | Anat. Jena. 13:930-941. 1902. Flora of the Southern United States. 698. Ed. 2. New York. 1884. |
| 155a. | Chapman, H. C | A Manual of Medical Jurisprudence and Toxicology. Philadelphia. 1892. |
| 156. | Charteris, F | .On the Active Principles of Poisoned Spears |
| | | Obtained from the Motchi or Monchi Country. Phar. Jour. London. 1901. Ser. 4. 13:43. |
| 157. | Chaussier, H | . Contre-poisons, ou moyens reconnus le plus ef- |
| | | ficaces pour commatre l'effet des diverses especes de poisons; suivis de l'indication des |
| | | secours à donner auz personnes mordues ou piquées par des animaux ou des insectes vé- |
| | | néneux, aux noyes, aux asphyxies, aux enfants nouveaux-nés, etc. Paris. 1818. |
| 158. | Chesnut, V. K | Abrus Poisoning. Asa Gray Bulletin. 5:35-36. Datura Poisoning. id. 5:87. |
| . 159. 160. | ••••• | Datura Poisoning. id. 5:87. |
| 100. | | .Plants Used by the Indians of Mendocino County, California. Cont. U, S. Nat. Herb. 7:295-408. |
| 161. | • | . Poisonous Spores of the Green-spored Lepiota. Asa Gray Bull. 8:87-93. |
| 162. | | . Preliminary Catalogue of Plants Poisonous to |
| | | |
| 163. | ••••• | Principal Poisonous Plants of the United States. Bull. U. S. Dept. Agr. Div. Bot. 20:60. See also Repr. Ill. St. Bd. Health |
| | • | 60 . 1899. |
| 164. | •••••••••• | Problems in the Chemistry and Toxicology of Plant Substances. Science. (N. S.) 15: |
| 165. | | 1016-1028. . Some Common Poisonous Plants. Year Book |
| 166. | | U. S. Dept. Agr. 1896:137-146. Some Poisonous Plants of Northern Stock |
| - 55. | | Ranges. Yr. Bk. U. S. Dept. Agr. 1900 : 305-324. |

| 167. | |
|---|---|
| 168. | Chesnut, V. K. and Wilcox, E. VThe Stock Poisoning Plants of Montana, Bull. |
| 169. | U. S. Dept. Agr. Div. Bot. 26:150. Chevalier, J |
| 169a. | Chevers, N A Manual of Medical Jurisprudence for India. Ed. 3. Calcutta. 1870. |
| 171. | Chrysler, M. A |
| 172. | Clapp, A A Synopsis; or Systematic Catalogue of the Indigenous and Naturalized Medicinal Plants of the United States. Trans. Am. Med. Assoc. 1852. |
| 173. | Clarke, J. F |
| 174. | Claude, H |
| 175. | |
| 176. | Coats, J |
| | 2000.100-112. |
| 176a. | Codex medicamentarius |
| | Gallicus |
| | Gallicus |
| 177a. 178. 179. | Gallicus Pharmacopeé Francaise. 1908. (Ph. Fr. 1908) Collin, Eug Traité de Toxicologie végétale. 210. Paris. 1907. Collins, G. W. See Cook, O. F. Colville, Y. E. Post-mortem Indications of Death from Poisoning. Oklahoma Medical News 15:193-196. |
| 177a. 178. 179. | Gallicus Pharmacopeé Francaise. 1908. (Ph. Fr. 1908) Collin, Eug Traité de Toxicologie végétale. 210. Paris. 1907. Collins, G. W. See Cook, O. F. Colville, Y. E. Post-mortem Indications of Death from Poisoning. Oklahoma Medical News 15:193-196. Combs, Robert Some Cuban Medical Plants. Phar. Rev.12: 87-91; 109-112; 136. Also Cont. Bot. Dept. |
| 177a. 178. 179. | Gallicus Pharmacopeé Francaise. 1908. (Ph. Fr. 1908) Collin, Eug Traité de Toxicologie végétale. 210. Paris. 1907. Collins, G. W. See Cook, O. F. Colville, Y. E Post-mortem Indications of Death from Poisoning. Oklahoma Medical News 15:193-196. Combs, Robert Some Cuban Medical Plants. Phar. Rev.12: 87-91; 109-112; 136. Also Cont. Bot. Dept. Ia. St. Coll. Agr. and Mech. Arts. 5:20. Connor, J. J On Poisoning from the Internal Administration of Tincture of Rhus Toxicodendron. |
| 177a. 178. 179. | Gallicus Pharmacopeé Francaise. 1908. (Ph. Fr. 1908) Collin, Eug. Traité de Toxicologie végétale. 210. Paris. 1907. Collins, G. W. See Cook, O. F. Colville, Y. E. Post-mortem Indications of Death from Poisoning. Oklahoma Medical News 15:193-196. Combs, Robert Some Cuban Medical Plants. Phar. Rev.12: 87-91; 109-112; 136. Also Cont. Bot. Dept. Ia. St. Coll. Agr. and Mech. Arts. 5:20. Connor, J. J. On Poisoning from the Internal Administration of Tincture of Rhus Toxicodendron. See Am. Jour. Der. Gen. Ur. Dis. 11:368. Connor, J. T. American Homeopathic Pharmacopoeia. Phil- |
| 177a. 178. 179. 180. | Gallicus Pharmacopeé Francaise. 1908. (Ph. Fr. 1908) Collin, Eug Traité de Toxicologie végétale. 210. Paris. 1907. Collins, G. W. See Cook, O. F. Colville, Y. E. Post-mortem Indications of Death from Poisoning. Oklahoma Medical News 15:193-196. Combs, Robert Some Cuban Medical Plants. Phar. Rev.12: 87-91; 109-112; 136. Also Cont. Bot. Dept. Ia. St. Coll. Agr. and Mech. Arts. 5:20. Connor, J. J. On Poisoning from the Internal Administration of Tincture of Rhus Toxicodendron. See Am. Jour. Der. Gen. Ur. Dis. 11:368. Connor, J. T. American Homeopathic Pharmacopoeia. Philadelphia. 1893. Ed. 2. Cook, Ernest H. A Case of Poisoning by the Areca Nut. Chem. |
| 177a. 178. 179. 180. 181. | Gallicus Pharmacopeé Francaise. 1908. (Ph. Fr. 1908) Collin, Eug Traité de Toxicologie végétale. 210. Paris. 1907. Collins, G. W. See Cook, O. F. Colville, Y. E. Post-mortem Indications of Death from Poisoning. Oklahoma Medical News 15:193-196. Combs, Robert Some Cuban Medical Plants. Phar. Rev.12: 87-91; 109-112; 136. Also Cont. Bot. Dept. Ia. St. Coll. Agr. and Mech. Arts. 5:20. Connor, J. J. On Poisoning from the Internal Administration of Tincture of Rhus Toxicodendron. See Am. Jour. Der. Gen. Ur. Dis. 11:368. Connor, J. T. American Homeopathic Pharmacopoeia. Philadelphia. 1893. Ed. 2. Cook, Ernest H. A Case of Poisoning by the Areca Nut. Chem. News. No. 1934. 1895. Cook, O. F. Origin of the Hindi Cotton. Cir. Bur. Pl. |
| 177a. 178. 179. 180. 181. 182. 183. | Gallicus Pharmacopeé Francaise. 1908. (Ph. Fr. 1908) Collin, Eug Traité de Toxicologie végétale. 210. Paris. 1907. Collins, G. W. See Cook, O. F. Colville, Y. E Post-mortem Indications of Death from Poisoning. Oklahoma Medical News 15:193-196. Combs, Robert Some Cuban Medical Plants. Phar. Rev.12: 87-91; 109-112; 136. Also Cont. Bot. Dept. Ia. St. Coll. Agr. and Mech. Arts. 5:20. Connor, J. J On Poisoning from the Internal Administration of Tincture of Rhus Toxicodendron. See Am. Jour. Der. Gen. Ur. Dis. 11:368. Connor, J. T American Homeopathic Pharmacopoeia. Philadelphia. 1893. Ed. 2. Cook, Ernest H A Case of Poisoning by the Areca Nut. Chem. News. No. 1934. 1895. Cook, O. F Origin of the Hindi Cotton. Cir. Bur. Pl. Ind. U. S. Dept. Agr. 12:12. Cook, O. F. and Collins, G. N Economic Plants of Porto Rico. Cont. U. S. |
| 177a. 178. 179. 180. 181. 182. 183. 184. | Gallicus Pharmacopeé Francaise. 1908. (Ph. Fr. 1908) Collin, Eug Traité de Toxicologie végétale. 210. Paris. 1907. Collins, G. W. See Cook, O. F. Colville, Y. E Post-mortem Indications of Death from Poisoning. Oklahoma Medical News 15:193-196. Combs, Robert Some Cuban Medical Plants. Phar. Rev.12: 87-91; 109-112; 136. Also Cont. Bot. Dept. Ia. St. Coll. Agr. and Mech. Arts. 5:20. Connor, J. J On Poisoning from the Internal Administration of Tincture of Rhus Toxicodendron. See Am. Jour. Der. Gen. Ur. Dis. 11:368. Connor, J. T American Homeopathic Pharmacopoeia. Philadelphia. 1893. Ed. 2. Cook, Ernest H A Case of Poisoning by the Areca Nut. Chem. News. No. 1934. 1895. Cook, O. F Origin of the Hindi Cotton. Cir. Bur. Pl. Ind. U. S. Dept. Agr. 12:12. Cook, O. F. and Collins, G. N Economic Plants of Porto Rico. Cont. U. S. Nat. Herb. 8:269. Cooke, N. C Oriental Aconite. Phar. Jour. III. 3:563- |
| 177a. 178. 179. 180. 181. 182. 183. 184. 185. | Gallicus Pharmacopeé Francaise. 1908. (Ph. Fr. 1908) Collin, Eug Traité de Toxicologie végétale. 210. Paris. 1907. Collins, G. W. See Cook, O. F. Colville, Y. E Post-mortem Indications of Death from Poisoning. Oklahoma Medical News 15:193-196. Combs, Robert Some Cuban Medical Plants. Phar. Rev.12: 87-91; 109-112; 136. Also Cont. Bot. Dept. Ia. St. Coll. Agr. and Mech. Arts. 5:20. Connor, J. J On Poisoning from the Internal Administration of Tincture of Rhus Toxicodendron. See Am. Jour. Der. Gen. Ur. Dis. 11:368. Connor, J. T American Homeopathic Pharmacopoeia. Philadelphia. 1893. Ed. 2. Cook, Ernest H A Case of Poisoning by the Areca Nut. Chem. News. No. 1934. 1895. Cook, O. F Origin of the Hindi Cotton. Cir. Bur. Pl. Ind. U. S. Dept. Agr. 12:12. Cook, O. F. and Collins, G. N Economic Plants of Porto Rico. Cont. U. S. Nat. Herb. 8:269. |

| 189. | |
|--------------|--|
| 190. | Acad. des Sci. de Paris. 1886:13. Vénenosite de quelques légumineuses exotiques appartenant aux genres Templetonia et Sophora. Compte. Rend. hebdom. de la Soc. de Biol. à Paris. 1693. 29 Avril. |
| 191. | Costantin, J |
| 192. | la Soc. de Myc. de France. 1893:166. Costantin, J. and Lucet, A. Sur un Rhizopus pathogene. Bull. de la Soc. |
| 193. | de Myc. de France. 19:200. Coulter, J. M |
| 194. | Coulter, S. MPlants Poisonous when Taken into the System. Ann. Rept. Dept. Geol. and Nat. Re- |
| 195. | sources of Indiana. 1899:553-1074, |
| 196. | Acad. Nat. Sci. 1904:51-63. Couperot, |
| 197. | Jour. Phar. et Chim. 27:542. Coupin, H |
| 198. 199. | Sci. 30:414-415. 1902 |
| | icaria, as stated by Baillon, but from one of the Simarubaceae. See Ann. de l'Inst. Co- lon. de Marseille, 1905:195-247. (Rev. in Phar. Jour. 1906:463. |
| 200. | Court, G See Pictet, A. |
| 201. | Courtet, A |
| 202. | Coville, F. V |
| 203. | Agr. Div. Bot. 8:4Observations on Cases of Mushroom Poisoning in the District of Columbia. Cir. U. S. |
| 204. | Dept. Agr. Div. Bot. 13:24. |
| 201. | latifolia). Proc. Am. Assoc. Adv. Sci. |
| 205. | Coville, F. V. and Funston, |
| 20.6 | Coville, F. V. and Funston, F |
| 206. 207. | Cownley, A. H |
| 208. | Sta. 3:10. Silage Poisoning. Supp. to Bull. Pur. Univ. |
| 209. | Agr. Exp. Sta. 147:628. Crawford, Albert C Barium, a Cause of the Loco Weed Disease. |
| 210. | Bull. Pur. Pl. Ind. 129:87Laboratory Work on Loco-weed Investigation. Bull. U. S. Dept. Agr. Bur. Pl. Ind. 121:39-48. |
| 211. | Larkspurs as Poisonous Plants. Bull. U. S. |
| 212. | Dept. Agr. Bur. Pl. Ind. 111:1-12 |
| 213. | U. S. Dept. Agr. Bur. Pl. Ind. 121:21-35 |
| | Bush, Phar, Rev. 26:230-235. Repr. |

| 214. | |
|---------------|--|
| 215 | Bull. U. S. Dept. Agr. Bur. Pl. Ind. 90:31-34. |
| 215. | |
| 216. | Crook, E. and Powell, R. D |
| 217. | Crookshank, E. MOn the Morphology, Cultivation and Toxic Products of the Tubercle Bacillus. Lancet, 1:296-300. 1891. |
| 218. | Crow, W. E |
| 219. | Crozier, A. A |
| 219a. 220. | |
| 221. | Czapek, Dr. FBiochemie der Pflanzen. I:584; II:1026. |
| 222. | Jena. 1905. Da Gama, J. A |
| 223. | Dakin, ROn Rhus Toxicodendron. See Am. Jour. |
| 223a. | Med. Sci. 4:98. Damen, CarlDie Gesundheitspflege der landwirtschaftlichen |
| 224. | Haussäugetiere. Ed. 3. 873. Berlin. 1902. Daniell, F. W |
| 225. | Daniels, C. W |
| 226. | 3, 5. Danjon, Em |
| 227. | Weeds Philadelphia 1847 |
| | American Weeds and Useful Plants, Being a Second Edition of the Above. New York. 1859. |
| 228a. 229. | Dassonville, C See Matruchot, L. Daugherty, C. M The Castor-oil Industry. Yr. Bk. U. S. Dept. |
| 230. | Agr. 1904:287-298. Davenport, C. B. and Neal, |
| | H. VOn the Acclimatization of Organisms to Poisonous Chemical Substances. Arch. f. Entwcklngsmechn. d. Organ., Leipz. 2:564- 583. 1895-1896. |
| | Davidson, |
| 232. | Davies, D. H |
| 233. | Day, M. G Experimental Demonstrations of the Toxicity of the "Loco Weed." N. Y. Med. Jour. 49:237. |
| 234. | Loco Weed. Ref. Book of Prac. Ther. by F. P. Foster. 1:587. |
| 235. | |

| 236 | . De Bary, A | retative Or- |
|--------------|--|---|
| 200. | gans of the Phanerogams and F lated and Annotated by F. O. Bo H. Scott. 659. Oxford, 1884. | erns. Trans- |
| 237. | | logy of the |
| | Fungi, Mycetozoa and Bacteria. H. E. F. Garnsey. Rev. by I. 525. Oxford, 1887. | Trans. by |
| 238. | De Candolle, AlphonseOrigin of Cultivated Plants. En 468. New York. 1892. | |
| 239. | . Deffernez, EdmondTriple empoisonnement par le D monium. Extr. du Bull. med. d 6. 1895 . | atura Stra- e Charleroi. |
| 240. | | |
| 241. | Java appelé Upas tieuté, et s vomique, la fève de St. Ignace, l potatorum et la pomme de Vont du même genre de plantes, o tieuté. 48. Paris. 1809. | ur la noix e Strychnos ac, qui sont ui l' Upas |
| 242. | Delotte | med., Lim- |
| 243. | De Puy, C. E | tory, Chem- Qualities of |
| 244. | | |
| | a. Deutsches Arzneibuch Ed. 4. 1900. (Ph. Germ. IV.). | |
| 245. | Familien aufgestellt mit Abbilo | lungen. 64. |
| 246. | See Eckhard's Beiträge zur A Physiologie | sburg, 1868. Anatomie u. |
| 246a. | a. Dinsmore, S. CSee Kennedy, P. B. | |
| | '. Dinwiddie, R. ROn the Toxic Properties of Molds Agr. Exp. Sta. 40:35-36. | |
| 247a. | a Unsound Corn and Forage as a C ease in Live Stock. Bull. Ark Sta. 25:1-9. | |
| 248. | | 73. 4 4 |
| | zarbeus | ano Antonio |
| 249. | Doeltz, J. CNeue Versuche und Erfahrung Planzengifte, Herausgegeben v Ackermann. 53. Nuremberg, 18 | on J. C. G. |
| 250. | Dohine, A. R. L. and En- | |
| | gelhardt, HFurther Experiments with Wild C Phar. Rev. 14 :13-15. See also 13 :260. | |
| 251. 252. | | ftgemischen, St. Peters- |
| 252a. | 2a Die Heilpflanzen der verschiede und Zeiten; ihre Anwendung | nen Völker |
| 252Ъ. | Bestandheile und Geschichte. 18 2b. Dungern, E. VBeiträge zur Immunitätslehre. M Woch. 1900. Nos. 20 and 28. | 396. Iünch, med. |
| | | |

| 253. | Dunglison, RobleyA Dictionary of Medical Science, New Edition by R. J. Dunglison. Camas Poisoning. 820-829. Philadelphia. 1874. |
|--------------|---|
| 254. 254a | New Remedies. Ed. 4. Philadelphia. 1874. |
| 255. | Dunglison, R. J |
| 256. | Dunstan, W. RA Report on the Chemical Examination of the Beans of Phaseolus lunatus. Agricultural Ledger. 1905. No. 2:11-16. |
| 257. | Dunstan, W. R. and Boole. |
| | L. E |
| 258. | Dunstan, W. R. and Henry, T. A |
| | Rept. Brit. Assoc. Adv. Sci. 1906:145-157. |
| 259. | Le glucoside cyanogénétique du lin. Bull. de |
| | l'Acad. roy. de Belgique. (Classe des Sci- ences). No. 7:790-793. |
| 260. | The Poisonous Properties of the Beans of |
| | Phaseolus lunatus. Jour. Board of Agr. Great Britain and Ireland. XIV. 12 :722- 731. |
| 261. | Sur la formation de l'acide cyanhydrique dans |
| | les végétaux. Ann. Chi., et de Physique. 8 ser. 10:118-128. |
| 262. | Dunstan, W. R., Henry, T. |
| | A. and Auld, S. J. M Cyanogenesis in Plants. Proc. Royal Soc. London, 78. |
| 263. | Dunstan, W. R. and Um- |
| | ney, C |
| | the true Aconitum Napellus, III. The For- |
| | mation and Properties of Aconine and its Conversion into Aconitine, Jour. of the |
| | Chem. Soc. 51 : 1892. |
| 264. | Dunstan, W. R. Wyndham, R., and Henry, T. ACyanogenesis in Plants. Pt. II. Proc. Roy. |
| | Soc. 70 :153-154. Part III (by Dunstan and |
| 265 | Henry). Proc. Roy. Soc. 72:285-294. |
| 265. | The Nature and Origin of the Poison of Lotus arabicus. Philos. Trans. Roy. Soc. |
| 0.00 | London, 1943:515-533. |
| 266. 267. | Dupony See De Nabias. Duvernoy, G. D De Lathyri quadam venenata specie in com- |
| 207. | itatu Montbelgardensi culta. 19. Basle. |
| 267a | 1770. Eames, E. H |
| 268. | Eastwood, A |
| 269. | Eccles, R. G |
| 270. | Eden, R. T Digitalis and its Cumulative Action. Ther. Gaz. 6:89. |
| 271. | Elborne, Wm |
| | of the Sawdust. Chem. and Drug. 189:85- |
| 272. | 86. Abs. in Proc. Am. Phar. Assoc. 44:642. Elphinck, Edward Case of Physostigmine Poisoning. Am. Vet. |
| 272 | Rev. 34:50. Elwert, C. PGelsemium sempervirens. Proc. Cal. Phar. |
| 273. | Soc. 1881. (abs.). |
| 273a. | Soc. 1881. (abs.). Engelhardt, HSee Dohme, A. R. L. |

| 274 | The Law Advice with The edit |
|-------|--|
| 274. | Engler, Adolf and Prantl, Karl |
| | Gattungen und wichtigeren Arten insbeson- |
| | dere den Nutzpflanzen. Leipsic. 1887-1909. |
| 275. | Erdmann, K. G Sammlung und Beschreibung der Giftpflanzen, |
| | die in Sachsen wild wachsen. 9 parts. Dresden, 1797. |
| 276. | Eriksson, J Giftiges Süssgras, Glyceria spectabilis, von Us- |
| 270. | tilago longissima befallen. Zeitsch. f. |
| | Pflanzenkr. Stuttgart 10:15. |
| 277. | Ernst, A Enumeracion de les especes de plantas em- |
| | pleadas en los diferentes paises para el |
| | metodo de pesca conocido bajo el nombre de "embarbascar." Univ. Centr. Venezuela |
| | Rev. Cientit. 1:318-32/. 1888. |
| 278. | Memoria Botanica sobre el Embarbascar o sea |
| | La Pesca por medio de Plantas venenosas. |
| 270 | 16. Caracas. 1881 |
| 217. | turf. Freunde. Berlin, Sitz. vom Juni, |
| | 1888:111-118. (Abs. Bot. Centrbl. 41:232- |
| 200 | 233.) |
| 280. | Evans, J. F. and Bose, C. L |
| | Sale of Poisons in Bengal. Indian Med |
| | Chir. Rev. Bombay, 3:289-336. |
| 281. | Evans, J. RMedicinal Plants of the Cherokees. Proc. |
| 281a | Ain. Phar. Assoc. (abs.) 23:154-156. EvesqueSee Bretin. |
| 282. | Ewart A I assisted by |
| | Tovey, J. RThe Weeds, Poison Plants and Naturalized |
| | Aliens of Victoria. Dept. Agr. Victoria. |
| 283 | 8:110. Melbourne. 1909. Faber, J. M |
| 200. | tiquorum vel Solani funosi recentiorum his- |
| | toriae monumentum, indolis nocumentum, |
| | antidoti documentum. 107. 13 pl. An- |
| 2832 | gustae Vind, 1877. Falck, F. ALehrbuch der praktischen Toxicologie. Stutt- |
| 2004. | gart. 1880. |
| 284. | |
| | Famulener, L. W. and Lyons, A. BThe Physiological Assay of Cannabis indica. |
| | Paper read before the Am. Phar. Assoc. |
| . 205 | 1903. Abs. in Phar. Rev. 21:370-371. |
| 203. | Farlow, W. GRelation of Certain Forms of Algae to Disagreeable Tastes and Odors. Proc. Am. |
| | Assoc. Adv. Sci. 32:306. (abs.) |
| 286. | Remarks on some Algae found in the Water |
| | Supplies of the City of Boston, Bull. Bus- |
| 207 | sey Inst. Jan. 1, 1877. |
| 287. | Some Edible and Poisonous Fungi. Bull. U. S. Dept. Agr. Div. Veg. Path. and Phys. |
| | 15:453-470. |
| 288. | Farmacopea italianaOssis Dizionario popolare di farmacia e ter- |
| | epeurica, con l'aggiunta delle sostaze ali- |
| | mentari, preparato e metodi industriali piu |
| | commune compilato de scelti farmacisti e dottori in chimica e medicina, sotto la dire- |
| | zione del prof. Guiseppe Gallo. Fasc. |
| | 1.8:1-448. Turin, 1880. |
| 239. | Farr, E See Wright, R. |
| | |

| <i>2</i> 90. | Fenn, C. M | Untoward Effects of Cascara Sagrada. Ther. |
|--------------|---------------------------|---|
| 291. | Ferguson, A. T | Gaz. 4:522. Cerebro-spinal Meningitis or Forage Poison- |
| 292. | Fermi C and Pernossi I. | ing. Am, Vet. Rev. 34:383. Ueber das Tetanusgift Vergleichende Studien |
| 2,2. | | mit Berücksichtigung andere Gifte und der Enzym. Zeitsch. f. Hygien. 16 :385-444. |
| 293. | Fernald, M. L | See Gray's Manual. |
| 294. | | Toxicite de l'acide filicique. 75. Toulouse. 1897. |
| 295. | · | Vorlesungen über Bakterien. Ed. 2. 374. 69f. 1903. |
| 296. | | Zur Morphologie, Biologie and Systematik der Kahmpilze, der Monilia candida und der Sooreregers Jena 1804 |
| 297. | Fischer, R | The Alkaloids of Escholtzia californica. Proc. |
| 298. | Fischer, T. | Am. Phar. Assoc. 49:443-453. Ein Fall von Stomatitis, Bacterium stomato |
| | | foetidum, ein aërober Faülnisserreger. Zeitsch f. Hyg 49 :329. |
| 299. | Flandin, C | De l'empoisonnement par les végétaux ou leurs principes immediats ou général, par |
| | | l'opium ou la morphine en particulier. |
| | • | Compt. Rend. Acad. Sci. Paris. 25:150-154. 1847. |
| 300. | Fleming, Geo | See Neumann, J. G. |
| 301. | Flückiger, F. A | Pharmakognosie des Pflanzenreiche. 2 Aufl. Berlin. 1881-1882. |
| 302. | Flückiger, F. A. and Han- | |
| | bury, David | Pharmacographia. A History of the Principle Drugs of Vegetable Origin, Met with in |
| | • | Great Britain and British India. London, 1879. |
| | | Fermenta und Mikroparasiten. 308. Leipsic. 1883. |
| 303a. | | Die Mikroorganismen mit besonderer Berück- |
| | | sichtigung der Aetiologie der Infektions- krankheiten. 692. Leipsic. 1886. |
| 304. | Fodéré, F. E | Voyage aux alpes maritimes, ou Histoire nat- |
| | | urelle agraire, civile et médicale du Comté de Nice et pays limitrophes. 2 v. 376+426. Paris. 1821. |
| 304a. | | Traité de médecine legale. 6v. Paris. 1813. |
| | Fogel, Estelle D | See Pammel, L. H. |
| 306. | Ford, W. W | Artificial Immunity to Glucosides. Proc. Am. Assoc. Adv. Sci. 1908. Abs. in Science, 27:652-657. |
| 307. | | The Toxins and Antitoxins of Poisonous |
| | | Mushrooms, Amanita phalloides. Jour. Inf. Dis. 3:191. |
| 308. | ••••• | Antibodies to Glucosides with especial Reference to Rhus Toxicodendron, Jour. Inf. |
| 308a | | Dis. 4:541-551. The Distribution of Poisons in Mushrooms. |
| | | Science. 30 :97-108. |
| 309. | rorrest | On Hydrocyanic Poisoning from Kidney Bean. See Glasgow Med. Jour. 18 :68. 1882. |
| 310. | Forster, E. L | Poisons: their Source, Use and Antidotes. Brit. Jour. Nursing. London. 30:164; 185; |
| | | 225; 246; 309; 353; 397; 429; 469; 508. |
| | | 31 :25; 64; 104;144; 185. |

| 311. | Fraissenet, J. F. F Poisons d'epreuve tanguin, M'Boundou, strophantus, féve de Calabar. Bordeaux. 1896. | |
|---------------|--|----|
| 312. 313. | Francis, George Poisonous Australian Lake. Nature. 18:11. Frankforter, G. B The Alkaloids of Veratrum. Minn. Bot. Studies. 1897:1024-1043. | , |
| 314. | Frazer, T. B. and Tillie, Joseph | |
| | 1895: 76-77. Free, J. E | |
| 315a. | er. 70 :633. Freeman | |
| 315Ъ. | Friedberg, L. H. Trans- | |
| 316. | lator See Kobert, R. Friedberger, F. and Fröh- | |
| | ner, E | |
| 316a. | Fröhner, EugenLehrbuch der Toxicologie für Tierärzte. Ed 3. 391. Stuttgart. 1910. | |
| 317. | Fröhner, E See Friedberger, F. | |
| 317a. 318. | Frothingham, L See Page, C. G. Fuchs, F. A Keratom, Aspergill. Wiener Klin. Wochen- | |
| 319. | schr. Leber Keratom als Ursache der Hypopyon- | |
| | keratis. Grafes Arch. 25 abt. 2 p. 285. 1879 Gaffky, George | |
| 320. | sicht auf progressive Virulenz u. akkomoda tive Zuchtung. Mittb. d. Kais. Ges. Amt | - |
| 321. | Gamaleia, N | |
| 322. | Gardiner, W. W Poisoning from Eating Locust-tree Bark | |
| 323. 324. | Garnsey, H. E. F See De Bary, A. Gattinger, A The Medicinal Plants of Tennessee. 128 | |
| | Nashville. 1894. Gautier, ALes toxines microbiennes et animales. Paris | |
| 325. | Soc. d'edit. scientif. 1896. | |
| 326. 327. | Gautier, L. and Bodin, E. See Bodin, E. Geiger, Paul Beitrag sur Kenntniss der Ipoh-Pfeilgifte. Mi | t |
| | einem Anhang; Pharmakognostiche Mitteil ungen über einige zur Heestellung von | - |
| | Ipohyerwendete Giftpflausen, 103. Basle. | |
| 328. | German, JohnOn Veronal Poisoning. See Jour. Am. Med Assoc. 1906 : (No. 26) 1999. | |
| 329. | Gerrard, A. W"Wangka," a new African Arrow-poison; it | s |
| | Composition and Properties (Strophanthus) Phar. Jour. 111. 11:833-834. 1889. Se | ė |
| 330. | also pp. 849-850. Gibbes, H | t. |
| 331. | Gibbons, H | |
| 332. | Jour. 21:496 Poisonous Species of Astragalus. Am, Journ | ١. |
| 333. | Phar. 51:237-240. Gibbons, W. PThe Medicinal Plants of California. Proc | ·. |
| | Am. Phar. Assoc. 19:297-308. (abs.) | |

| 334. | in | the Occurrence of a poisonous Alkaloid "West African Boxwood." Quart. Jour. st. Comm. Research. Topics, 1:32-40. |
|--------------|---------------------------|--|
| 335. | L | iverpool. e sur l'action vesicante de Heracleum. |
| 336. | Λ | nn. Soc. Bot. Lyons. 26 :133-150, ium maculatum. Jour. belge d'homoe- |
| 337. | 01 | pathie. 6:254-261. (1899). Datura Poisoning. See Brit. Med. Jour. |
| 338. | 1 | 903:1137. eral Species in the Malay Peninsula Used |
| 339. | 10 | or Asthma. Brit. Med. Jour. 1903:1137. kspur and other Poisonous Plants. Bull. |
| 340. | | ol. Agr. Exp. Sta. 113:24. Poison Weed Problem of the Arid West. |
| | A | m. Vet. Rev. 34:493-502 |
| 341. | 13 | sonous Weeds. Bull. Col. Agr. Exp. Sta. 37:5-8. |
| 342. | w S ti | andlung von den giftigen Gewächsen, elche in Deutschland und vornehmlich in chwaben wild wachsen. 228. Ulm. Stetn. 1775. |
| 343. | N | remeine Geschichte der Pflanzengifte, 525. uremberg. 1775. |
| 344. | cl | Plantes vénéneuses du Canton de Neu- natel, avec 26 planches coloriees. 59. Neu- |
| 345. | Godfrey, BSan | tonine Poisoning. (Of two Dogs). The |
| 346. 347. | Godfrey, GeorgeSee | eterinarian. 73:150-151. Michael, L. |
| 348. | Goeppert, H. R Uet | per die giftige Pflanzen Schlesiens. Bres- u. 1832. |
| 349. | Gola, GL'ac | cido cianidrico e i glucosido nel regno veg- |
| 350. | Good, Peter A I | ale. Suppl. Encicl. Chimica, 23:43-79. Materia Medica Botany. New York. 1846. |
| 351. | Gordon, P. R See | Bailey, F. M. |
| 351a. | te d | erche Batteriologiche e Chimiche sulle Al- erazione del Mais. Contributo all' etiologia ella Pellagra. (Memoria 2a). Rivista d' giene e Sainto Pubblica, 7:825. 1896. |
| 352. | GottheilBla | ck Tongue: its Etiology. Archives of ediatrics. April. 1889. |
| 353. 354. | GözeUns | sere Giftpflanzen. cor corymbifer in the External Auditory |
| 355. | У | Ieatus. Lancet. 2:1379. 1890. träge zur systematischen Botanik der |
| <i>333.</i> | p | flanzlichen Parasiten. Virsch. Archiv. 887:546. |
| 355a | a. Graves, C. B. et alCat | alogue of the Flowering Plants and Ferns f Connecticut growing without Cultivation. |
| 356. | 5- | 69. Hartford, 1910. Animal Poison of the Far West—"Loco- |
| 357. | 0 | r Crazy Weed." Am. Agr. 1878:380-381. |
| | C | Botanical Text-Book, an Introduction to cientific Botany. New York, 1850. |
| 358. | ic | Contribution to the Flora of North America. American Academy of Arts and Scinces. 1883, |
| 359. | | nera of the Plants of the U.S. 2 vols. New York. 1848. |

| 360 | |
|-------------|--|
| 361 | Physiology. New York. 1874 |
| | |
| 362 | United States. 710. Boston. 1848. Ed. 7 of Gray's Manual. A Handbook of the Flowering Plants and Ferns of the Control West of the Flowering Plants and Ferns of the Flowering Plants and Ferns of the Control West of the Flowering Plants and Ferns of the Flowering Plants and Ferns of the Flowering Plants and Ferns of the Flowering Plants and Flo |
| | Central and Northeastern United States and |
| | Adjacent Canada, Rearranged and Ex- |
| | tensively Revised by B. L. Robinson and M. L. Fernald. 926. New York. 1908. |
| 363 | M. L. Fernald. 926. New York. 1908 |
| | vols New Vork 1878-1884 |
| 364. Gree | en. I. ReynoldsThe Soluble Ferments and Fermentations. |
| 3642 Grad | 512. Cambridge. 1901. en, Wesley |
| | Lowa 1907 |
| 365. Gree | eenish, H. GAn Introduction to the Study of Materia |
| | Medica, being a Short Account of the more Important Crude Drugs of Vegetable and |
| | Animal Origin, 571, London, 1899. |
| 366. Gree | enleaf, R. W Case of Aconite Poisoning. Rept. Boston |
| 367. Grei | Med. Surg. Jour. imer, KarlUeber giftige Boragineen: Alkaloids. Phar. |
| | Zeitung 43: No 20 |
| 368 | Ueber giftig wirkende Alkaloide einiger |
| 369. Gres | Boragineen. 1900. Shoff, M |
| | etable Kingdom, Rept. Colonial Museum |
| 270 | Haarlem, Holland. 1906. (Reprint) |
| 370 371 | |
| 0,1, | plantem bij de vischvangst in gebruik. 2 |
| 37 2 | vols. completed; v. 3 incomplete. 1893-1910, |
| 3/2 | Misc. Inf. Roy. Bot. Gar. Kew. 1909:397- |
| | 418. |
| 373. Gres | ssler, F. G. LDeutschlands Giftpflanzen mit zahlreichen Il- lustrationen und 8 Tafeln in Fabendruck. |
| | 16 Anfl. 40. Langensalza, 1897. |
| 374. Grey | y, George |
| 375. Grig | Phar. Jour. III. 5:248-250. (1879). gorescu, GAction des substances toxiques sur l'excitabil- |
| 0,0. 0116 | ite des nerfs et des muscles periphériques; |
| | un antidote de la strychnine. Arch de phy- |
| | siol nom norm, et Path. Paris, 1894, ser. 56 :32:39. |
| 376. Gro | he Experiments über die Injektion der Pilzsporen |
| | von Aspergillus glaucus u. Penicillium |
| 377. Grö | glaucum, Berlin, Klin, Wochenschr. 1870. nlund, Chr |
| 378. Gros | ss, HermanLocust Bark Poisoning Horses. Breeder's Ga- |
| 379. Grui | zette. 55 :455. |
| Ji J. Giul | game qui constitue le vrai muguet des en- |
| | fants. Compte Rend. des Séan. de l'Acad. |
| 380. Guai | de Paris. 14. 1842. rdia, J |
| | Phar. 7:542-548. 1893. |
| 381. Gues | guen, J Les champignons vénéneux et leurs carac- |
| 382 | tères. Revue scientifique. 1908 :321-328 |
| | causes et traitement des empoisonnements. |
| | Rev. sci. 1908 :361-366, |

| <i>3</i> 83. | Guenther, J. and Bartuch, | Disclosing to the form of the City's confidence of the |
|--------------|---------------------------|--|
| | F | Pinakothek der deutschen Giftigewächse. 16. Jena. 1840. |
| 384. | Guerin, P | Jena. 1840. The Probable Cause of the Poisonous Effect of the Darnel, Lolium temulentum. Bot. Gaz. 28:136-137. |
| 385. | Guignard, L | Le haricot à acide cyanhydrique (Phaseolus |
| 386. | | Nouveaux exemples de Rosacées à acide cy- anhydrique. C. R. Acad. Sci. Paris. 143 : 461. |
| 387. | | Recherches physiologiques sur la greffe des plantes à acide cyanhydrique. Ann. sc. Nat. |
| 388. | | ser. 9. Bot. 6 :261-305. A Study of Beans yielding Hydrocyanic Acid. Rev. Vit. 26 :5-9; pl. 1.; 33-37; 341-350. f.9; 543-547; 573-576; 626-631; 663-687; 689-694; |
| 389. | | 715-721. Sur la localisation des principes actifs chez |
| | | les Tropéolées, Compt. Rend, des. Séan de l'Acad. des sci. de Paris. 117 :587-590. Abs. in Beihefte zum Bot. Centrbl. 4:220- 221. |
| 390. | | Sur l'existence, dans le sureau noir d'un com- posé fournisseant de l'acide cyanhydrique. |
| 391. | | Bull. Sci. Pharmacol. 12 and 13. Sur la greffe des plantes à acide cyanhydrique. C. R. Acad. Sc. Paris. 30 Dec. 1907 . |
| 392. | | Sur la metamorphose des glucosides cyanhy- driques pendant la germination. C. R. Acad. |
| 393. | | Sc. Paris. 147:1023. Sur la prétendue toxicité des Haricots de Hongrie. C. R. Acad. Sc. Paris. 9 Dec. |
| 394. | | 1907. Sur les quantities d'acide cyandydrique four- nier par le Phaseolus lunatus. L., cultivé sous le climat de Paris. Bull. Sc. Phar. 14:565. |
| 395. | Guttenberg, Gustave | Poisonous Plants which Grow in and around Erie. (Read before the Nat. Hist. Soc. Sept. 5, 1879 . 21-25. |
| 396. | Hager, Hermann | Ueber die giftige Wirkung einiger Lathyrus- Arten. Monatliche Mittheiligungen aus dem Gesammt-Gebiete der Naturwissenschaften. 6:153. |
| 397. | Haines, W. S | Alkaloidal and other Organic Poisons. In: Hamilton and Godkin's System of Legal Medicine, 1:415-473. |
| 398. | | |
| 399. 400. | Halderman | On Gaultheria Poisoning. See Lancet. 1886. The special Symptomology of the New Remedies. Philadelphia. 1877. |
| 401. | | Etude sur les trichophytes de la barbe. Ar- |
| 402. | | Die deutsche Giftpflanzen. 2 v. 119+126. 24 col. pl. 1784-1795. |
| 403. | Halsted, B. D | Notes on Poisonous Plants. Gar. and For. 8: |
| 404. | | Other Poisonous Plants. (Maclura aurantiaca |
| 405. | | The Poisonous Plants of New Jersey. Bull. N. J. Agr. Eyn. Stp. 135-28 |

| 406. | |
|------|--|
| 407. | HamiltonOn Recovery from Gaultheria Poisoning. See N. Y. Med. Jour. 1875 . |
| 408. | Hamilton, A. M. and God- kin |
| 400 | on Poisons by C. E. Pellew, W. S. Haines, and V. C. Vaughan. |
| 409. | Hamilton, W On the Genus Jatropha and the Janipha Manihot and J. Loeflingii. Phar. Jour. 5:27-31; 31-33. |
| 410. | On the Noxious Properties of the Echites suberecta or Savanna flower and their Anti-dotes. Phar. Jour. 6 :25-28. (1846.) |
| 411. | On the Properties of the Hippomane Mancinella or Manchineel Tree. Phar. Jour. 5: |
| 412. | Hanbury, David Note on the Ordeal Bean of Calabar. (Physostigma venenosum Balf.) Phar. Jour. II. 4: 559-561. (1863.) |
| 413. | See Flückiger, F. A. |
| 414. | Hansen, N. E The Wild Alfalfas and Clovers of Siberia, with a Perspective View of the Alfalfas of the World. Bull. Bur. Pl. Ind. 156:31. |
| 415. | Hare, R. F., Mitchell, S. R. |
| | and Bjerregard, A. P Denatured Alcohol from Tunas and other |
| | Sources. Bull. N. M. Agr. Exp. Sta. 72: 57. |
| | Harger, E. B See Graves, C. B. |
| 416. | Harknis, W. D. and Swain, R. E The Chronic Arsenical Poisoning of Herbivo- |
| | rous Animals. Jour, Am. Chem. Soc. 30: 928. |
| 417. | Harlan, HerbertBlindness and Death from Drinking Essences of Ginger, Peppermint, etc., due to Methyl |
| 418. | Alcohol. Phar. Rev. 19:51-53. |
| 410. | Harnack, E. and Hochheim, W |
| • | krampferregender Gifte. Ztschr. klin. Med. Berlin. 25:16-45 . |
| | Harnack, E. and Meyer, H Untersuchungen über die Alkaloide der Jaborandiblätter. Lic. Ann. d. Chem. 204 : 67-84. |
| 420. | Harrington, COn Poisoning by Canned Beans, Caused by |
| | Toxins rather than by Living Bacteria, Bot- ulism suggested. Rev. in Am. Jour. Med. |
| | Sci. 133 :811. From Article by Rolly in |
| | Münch. Med. Wochenschr. 1906. No. 37. |
| 421. | 1798 On Poisoning from Foods. Rev. in Am. Jour. |
| 421. | Med. Sci. 132:810. From Art, by Dr. W. G. Willoughby in Pub. Health. 1906:626. |
| 422. | Harrison, W. HThe Poison Ivy. Gar. and For. 8:68. |
| 423. | Harshberger, J. W Ergotism. (Account of an Outbreak at Scran- |
| | ton, Pa., due to Eating Red Top, Agrostis alba vulgaris with Claviceps microcephala, (Wallr.) Tul.) Rept. Pa. Dept. Agr. 1902: |
| 424. | 160 |
| 424. | ous Plant. Bot. Gaz. 19:159. Gar. and For. 7:170. |
| | |

| 425. | | A Review of our Knowledge of Phytobezoars. Jour. of Com. Med. and Vet. Arch. 19: 143. |
|-------|--------------------------|---|
| 426. | | When is Rhus Toxicodendron most Active? Gar. and For. 8:239. |
| 427. | Hartley, C. P | Broom Corn. Far. Bull. U. S. Dept. Agr. 174:30. |
| 428. | Hartman, G | Mechanische Vergiftungen. Berlin. 1896. |
| 429. | Harvey, P. F | Observations on the Toxic Effects of some Common Drugs. N. Y. Med. Jour. 73: 148-150. |
| 430. | Hartzell, Stella | |
| 431. | Harz, C. D | Landwirthschaftliche Samenkunde. 2:1315. |
| 431a. | Haslam, Thos. P | Meningo-Encephalitis. Bull. Kan. Agr. Exp. |
| 432. | Hatch W G | Sta. 173:235-251. (Contains Bibliography.) On Loss of Limb from Rhus Poisoning. See |
| 432. | | Cal. State Med. Jour. 1:465. |
| 433. | | Drink Plants of the North American Indians. Bull. Torr. Bot. Club. 23:33-46. |
| 434. | Heald, E. D | |
| 435. | Hebert, A | Recherches sur la presence de l'acide cyanhydrique chez diverses plantes. Bull. de la Soc, chim. de Paris. 1896. No. 74:310. |
| 436. | | Sur les princeps actifs des fruits d'un Strychnos africain. Jour. Phar. et Chem. 27:151. |
| 437. | Heckel E | Nouvelles sur le Tanghin du Ménabé (Mena- |
| | | bea venenata, Baill.) et sur sa racine toxique et médicamenteuse. C. R. Acad. Sc. Paris. 134 :441-443. |
| 438. | | Sur la Menabea venenata, Baillon, qui four- nit, par les racines, le Tanghin de Ménabé ou des Sakalaves (poison d'épreuve), en- core denommé Kissoumpa ou Kimanga, à Madagascar, sa patrie. C. R. Acad. Sc. Paris. 134:364-366. |
| 439. | | Sur un autre poison des Sakalaves appelé Komanga ou Kimanga et fourni par l'Ery- throphloeum Coumingo Baillon. Rev. des cult. Coll. 11:129-134. |
| 440. | Heckel, E. and Schlagden | |
| | hauffen, Fr | Sur la Copaifera Salikounda Heckel de l' Afrique tropicale et sur ses graines à cou- marine (Salikounda des peuples Sousous) au point de vue botanique et chimique. Com- paraison avec la fève de Tonka. Ann. de la fac. des sci. de Marseille. III. Sep-abd. 1-18. |
| 441. | Hedges, Florence | |
| | | Cicuta, a Plant that Poisons Cattle. Bull. Ore. Agr. Exp. Sta. 46:12. |
| 443. | Heffter, A | Pellotinia, a new Hypnotic. Phar. Rev. 114: 153-154. |
| 444. | | . Ueber Cacteen Alkaloide. Ber. deutsch, chem. |
| 445, | Hefti, J. J | Gesell. 34 :3004. Eein Beitrag zur Kenntniss der speziell in |
| | | central Europa. Giftpflanzen und Pflanzen- gifte. Schwanden. 1889. |

| 446. | Hegetschweiler, JFlor | e des plantes vénéneuses de la Suisse ntenant leur description l'époque de leur |
|--------------|--------------------------------|--|
| | | raison, les lieux, l'indication de celles qui |
| | | nt employées en médecine, les symptomes i elles produissent et les premiers soins ci |
| | | nner dans les empoisonnements. Desti- |
| 447. | Hegetschweiler, J. and La- | e à l'usage compagne. 117. 23 pl. 1849. |
| 4.40 | bram, J. D | Giftpflanzen der Schweiz. 46. Zürich. |
| 448. | Do | Sale of Poisons in India. Ind. Med. Chirv. 3:351-360. Bombay. |
| 449. | Hein, Heinrich Deut | schlands Giftpflanzen. Eine kurze Be- ireibung der giftigen und verdächtigen |
| | . scl | reibung der giftigen und verdächtigen heimischen Pflanzen nebst Hinweis auf |
| | de | ren Verwendung in der Pharmacie, zu- |
| | gle | ren Verwendung in der Pharmacie, zu- ich als Text für die vom Verwasser, he- |
| | Le | isgegebenen Gift-pflanzenherbarien für hrer, Schüler, und zum Hausgebrauche. |
| 440- | 74. | Hamburg, 1880. |
| 449a | a. Hektoen, Ludwig Systo Gr | ematische Blastomycose und coccidioidales anulon, Chiari-Festschr, herauseg, von |
| | Pr | anulon. Chiari-Festschr. herauseg. von of. Paul Dittrich in Prag. Wien. u. Leip. |
| 450. | Hempel, C. J New | 108:116-142. Homoeopathic Pharmacopoeia of Jahr, |
| | Bu | chner and Gruner New York 1850 |
| 451. | Inc | rican Medicinal Barks. Bull. Bur. Pl. 1. U. S. Dept. Agr. 139:49. |
| 452. | Wee | ls Used in Medicine. Far. Bull. U. S. |
| 453. | Wild | Medicinal Barks of the United States. |
| 454. | Henry, AimeDie | ll. Bur. Pl. Ind. U. S. Dept. Agr. 89:76. Giftpflanzen Deutschlands zum Schulge- |
| | bra | uch und Selbstunterricht durch Abbild- |
| | 10 | gen und Bescreibungen erläutert. Bonn. 32 col. pl. 1836. |
| 455. | Henry, JLes | principales plantes vénéneuses. Brussels. Dunstan, W. R. |
| 456. 457. | Henry, T. A See | Junstan, W. R. Donous Plants in Field and Garden. 189. |
| | Lo | ndon. 1897. |
| 458. | | r das Blausäure liefernde Glykosid der men von Eriobotrya japonica. Arch. der |
| | Ph | ar. 245 :469. |
| 459. | Uebe | ar. 245 :469. r das Prulaurocerasin, das Blausäure fernne Glukosid der Blätter von Prunus ro-cerasus. Arch. der Phar. 245 :463- |
| | lau | ro-cerasus. Arch. der Phar. 245 :463- |
| 460 | 460 | |
| 460. | in | Cotoneaster microphylla, Wall. Arch. |
| | der | Phar. 245 :473. |
| 461. | Herissey, H. and Lefebre, | r das Vorkommen der Raffinose in Taxus |
| C | bac | cata. Arch. der Phar. 245:481-486. |
| 462. | ***** | rsuchungen zur Physiologie der Muskeln 1 Nerven. Heft 3. |
| 463. | - Vo | ise Producing Microörganisms, 644. New rk, 1910. |
| 464. | Hetier, FrEmpo | pisonnement par l'Entoloma lividum. Bull. |
| 465. | Heyer, FUebe | r die Eirgenthümliche giftige Wirkung |
| | | er zu Leguminosen gehörigen Futter- nze. Landw. Post. Berlin. 1887. April |
| | 26. | |

| 466. | Hickman, R. W | . Epizoötic Cerebro-spiual Meningitis of Horses. Rept, Bur. Am. Ind. U. S. Dept. Agr. 1906 . 165-122. |
|------|-------------------------|--|
| 466a | . Hilger, A | |
| 467. | Hinebauch, T. D | Feeding of Millet to Horses. Bull. N. D. |
| 468. | | . Rheumatism in Horses. Bull. N. D. Agr. Exp. Sta. 7:15 |
| 469. | | .Rape as a Forage Crop. Far. Bull. U. S. Dept. Agr. 164:16. |
| 470. | | Die Giftgewächse Deutschlands und der Schweiz. Esslingen, 1884 |
| 471. | | Lehrbuch der gerichtlichen Medicin. Ed. 5. |
| 472. | Hoffman, Fr | .Loco Weeds. Phar. Rund. 1889:168. |
| 473. | Hoffman, Kalph | . Virulence of the Wild Parsnip. Rhodora. 4:188. |
| 474. | Holm, Theo | Medicinal Plants of the United States. |
| 475. | Halman E M | Merck's Report. 1907—1910 (not yet complete). |
| 4/3. | | Agricultural and Horticultural Poisons. Chem. and Drug, London. 63:247. |
| 476. | ••••• | Rhus Toxicodendron. Phar. Jour. 81:231- |
| 477. | ••••• | Some Books of Medicinal and Toxicological Interest. Phar. Jour. Ser. IV. 16:5-7. |
| 478. | | Vegetable Poisons and their Antidotes. Phar. Jour. III. 4:769-770. |
| 479. | Homen, E. A | De l'action du streptocoque et de ses toxins |
| | | sur les nerfs, les ganglions spinaux et la moelle épinière. C. R. de la Soc. de Biol. 1896 :518-523. |
| 480. | | The Bark of the Cleistanthus collinus as a Fish Poison. Phar, Jour, IV. 1465. |
| 481. | | An Essay on the Copas or Poison Tree of Java, Addressed to the Hon. Stamford Rof-fles, Lieut. Gov. |
| 482. | | Occurrence of Barium in the Ohio Valley Brines and its Relation to Stock Poisoning. Bull. W. Va. Agr. Exp. Sta. 103:281-295. |
| 483. | | . See Morse, F. W. |
| 484. | Huart, Louis | Les alkaloides, les leucomaines, et les pto- maines. Ann. de Phar. 1899 . No. 5. |
| 485. | Hubbard, R | On Poisoning from Internal Administration of Rhus Preparations. See Med. Rec. |
| 486. | Hugemeyer | 30:601. Ueber Abschwachung pathogener Schimmel- |
| 487. | Hughes, D. 'A | pilze. Bonn. 1888. Loco-weed Poisoning and Barium Salts. Am. |
| 488. | | Vet. Rev. 33:613-617. Poisonous Plants Destructive to Live Stock |
| 1001 | | with Special Reference to a Recent Book on the Subject. Am. Vet. Rev. 36:672-679. |
| 489. | Hughes, Richard | A Manual of Pharmacodynamics. Ed. 3. |
| 490. | Hunt R and Iones W | London, 1876. Non-alkaloidal Organic Poisons. In: Text- |
| 170. | Train, it and joines, w | book Leg. Medicine (Peterson and Haines). 2:532-643. Philadelphia, 1904. |
| 491. | Huntington, Annie O | Poison Ivy and Swamp Sumach. 58. 7 pl. Jamaica Plain. Mass. 1908. |

| 492. | Huseman, A., Hilger, A. and Huseman, ThDie Pflanzenstoffe in chemischer, physiolog- |
|---------------|--|
| | ischer und toxicologischer Hinsicht. 320. Berlin, 1883. |
| 493. | Huseman, Th The Distribution of Cardiac Poisons in the Vegetable Kingdom. (abs.) Phar. Jour. III. 7:795-796;834-835. |
| 494. | |
| 495. | Hutcheon, DPoisoning of Horses by Printhogalum thyrsoides or "Chickerinchee" (Ornithogalum thyrsoides). Agr. Jour. Cape of Good Hope. 28:165-172. |
| 496. | Hutyra, Franz, and Marek, Specielle Pathologie und Therapie der Haus- |
| 497. | Hyams, C. W |
| 498. 499. | Hyde, J. N |
| 500. | Illinois State Board of Health |
| 501. | Imbert-Gourbeyre For the Identity of Hemlock and the Athenian Poison, see "De la mort de Socrate." Paris, 1875. |
| 502. | Immunitat (Die) von Pflanzen gegen ihr eigenes Gift. Zeitsch. f. ang. Mikr. 13:60-62. Leipsic. 1907. |
| 502a. | Index CatalogueOf the Surgeon General's Office, U. S. Army. Ser. 2. 13:594-603. (Bibliography on Poisons and Allied Subjects). See also Cor- responding Volume of Ser. 1. There are many cross references to other volumes that are helpful. |
| 503. | Inoko, YZur Kenntniss der Pilzvergiftung. Fort- schritte der Medicin. 1893 :444-449. |
| 503a. | IrvingOn Aconite. See Med. Times and Gaz. 1:63. |
| 504. | Israel, O., Klingman, Th., Kahlenberg, L., True, R. H., Heald, E. D. and Cou- pin, H |
| 505. | 188. Sur l'existence dans le Thalictrum aquile- legifolium d'un composé fournissent de l' acide cyanhydrique. Jour. de Phar. et de Chimie. Oct. 1905. |
| 506. | |
| 507. | Jackson, J. R |
| 507a. 508. | Jacob, J Poisons et Contrepoisons. Paris. 1877. Jacobj, C Definition des Wortes Gift. Deutsch. med. Wochenschr. 26:2-4. |

| 508a. Jacoby, M |
|--|
| 509. Jahn, E |
| 510 Jeanmire De la pocivité relative et temporaire de |
| l'Amanita junquilles, Bull. Soc. myc. de |
| 511. Jeliffe, S. M |
| 512. Jennings, O. E |
| Jour. Myc. 13:187. 513. Jesup, H. G |
| Touch? Bot. Gaz. 18:142. 514. Jewett |
| Gaz. 1867 :68. 515. Johnson, CharlesBritish Poisonous Plants, 59. London. 1856. |
| 515. Johnson, Charles British Poisonous Plants, 59. London, 1856. 516. Johnson, Lawrence A Manual of the Medicinal Botany of North |
| America 202 Now Vorte 1994 |
| 517 On Toxicity of Tincture of Rhus Toxicoden- dron which had been Kept Thirty Years. |
| See Med. Dot. 119. |
| 374 A TO - C4. 145 56 100 |
| Vt. Agr. Exp. Sta. 145:50-199. 518a See Rich, F. A. |
| 519. Jones, W |
| 520. Jorissen, ALa linamarine, glucoside générateur d'acide |
| cyanhydrique. Réponse à la Note de MM. |
| Dunstan et Henry. (Bull. de l'Acad. Roy. de Belgique-Classe de Sci. 7:773-778. Abs. in Bot, Centrbl. 107:242-243. |
| 521. Juch, K. W Die Giftpflanzen. Zur Belehrung für Jeder- |
| mann. Augsburg. 1817. 522. Judkins, E. H |
| 523. Kabsch, W Das Pflanzenleben der Erde. Eine Pflanzen- |
| geographie für Laien und Naturforscher. |
| Nach dem Tode des Verfassers mit einem |
| Vorwort verschen von H. A. Berlepsch. |
| 642. Hanover, 1855. |
| 524. Kahlenberg, L See Israel, O. 525. Kalm, P On Poisoning from Rhus Toxicodendron. See |
| Travels 1:79 1770 |
| 526. Karsten, GeorgSee Strasburger, E |
| 526a. Kartulis, S |
| schr. f. Hy. u. Infektkr. 64:285. |
| 527. Kaufmann, Paul Nouvelle experiences sur l'injection le spores de Aspergillus glaucus, Lyon Med. No. 1832. |
| No. 10. |
| 528. Kaupp, B. F |
| 529. Kebler, L. F |
| 530. Keeler, D. D |
| 531. Kellerman, W. A Poison Ivy and Ivy Poisoning. Ohio Naturalist. 2:227. |
| 532 Poisoning by Shepherd's Purse, Bot. Gaz. 20:325-326. |
| 533. Kellogg, A |
| Acad. Sci. 6:3. |
| 533a. Kennedy, G. WKalınia latifolia. Am. Jour. Phar. 47:5. |
| 534. Kennedy, P. BSummer Ranges of Eastern Nevada Sheep. Bull. Nev. Agr. Exp. Sta. 55:53. |
| Dam Field Fight Like October |

| 535. | Kennedy, P. B. and Dinsmore, S. C |
|------------------------------|---|
| 536. | Nev. Agr. Exp. Sta. 71:20-32 Kennedy, P. B. and Doten, S. B |
| 537. | Sta. 51:56. Kerner, Anton von and Oli-Poison from Stinging Hairs. See The Natural ver, F. F |
| 538. 539. | Kerner, J. S Deutschlands Giftpflanzen. Hanover, 1798. Khittel, J On Rhus Toxicodendron. See Abs. Am. |
| 540. 541. | Jour. Phar. 30:542. King, John |
| 542. 542a. 543. | Cincinnati. 1880. Kingsley, A. T |
| 544. 545. 546. 547. | 317; 487-493. Kitt, Theo. Bakterienkunde. 1899. Klebs. Handbuch der pathologische Anatomie. 1869. Klingman, Th. See Israel, O. Knolle, Plantae venenatae umbelliferae. Leipsic. 1771. |
| 548. 549. 550. | Knowles, M. E |
| 551. | 9:114-117. Kobert, E. RCompendium der praktischen Toxicologie. |
| 552. | 1903 Lehrbuch der Intoxicationen. 816. Stuttgart. |
| 553. | 1893. |
| | Practical Toxicology for Physicians and Stu- dents. Translated and Edited by L. H. Friedburg, 201. New York, 1897. |
| 553b. | Ueber Giftstoffe der Flechten. Sep. Abd. aus Sitzungsber. der Dorpat Naturf. Gesell. 1892:156-186 |
| 554. | Koch, D. G. D. J Synopsis Florae Germanicae et Helveticae, etc. Ed. 3. 875. Leipsic, 1877. |
| 555. | Koenig, A |
| 556. | Kohlhaus, J. J |
| 557. | schreibungen. Regenburg. 1805. Kohn-Abrest, E Sur les legumes toxiques. Rev. internat. d. |
| 558. | falsific. 20 :25. Paris. 1907. Kraemer, Henry A Textbook of Botany and Pharmacognosy. |
| 559. | 850. Ed. 3. Philadelphia, 1905. Some of the distinguishing Morphological Characters of Belladonna Scopplia, Proc. |
| 560. | Am. Phar. Assoc. 56 :819-824. f. 77-79. Krapf, K. von Experimenta de Ranunculorum venenata qualitate. Vienna. 1838. |
| 561. | Kraus, R. V. and Levaditi. |
| | C |

| 561a. | Krause, A See Mueller, G. |
|---------------|--|
| | Krause, M |
| 563a. | |
| 563ს. | Kren and SchramekUeber Sporotrichose. Wiener klin. Wochen- schr. 1909. No. 44. |
| 564. | Kreutzer, K. JOestereiches Giftgewächse. Vienna. 1838. |
| 565. | KrockerZur Lupinenkrankheit der Schafe. Landw. Jahrb. 9:27-35. |
| 566. | Kubingi, A Plantae venenosae Hungariae. Merges novenyik, etc. Buda. 1842. |
| 567. | Kunkel, A. J |
| 568. | Kyle, H. M On the Action of the Spurge on Salmonoid Fishes. Proc. Roy. Soc. 70:48-66. |
| 569. | Labesse |
| 570. | Labram, J. D See Hegetschweiler, S. |
| 571. | Lacerda, J. B. deExperiencias physiologicas cornalgumas plantas toxicas do Brazil. Rio de Janiero. 1890. |
| 572. | Ladd, E. F |
| 573. 574. | See also Am. Chem. Jour. 20:862. Lafitte, Henry Erigeron canadense. Phar. Post. 1887:802- |
| | 803. |
| 575. | La Glesch, Louis, and Vuil- let, Jean |
| | cais. Schweiz. Wochenschr. Chem. Pharm. 40:546-548. |
| 576. | Lambert, LSee Noel, C. |
| 577. 578. | Lang, W. H., Translator See Strasberger, E. Langguth, G. A Programma de plantarum venenatarum ar- |
| 370. | cendo scelere. 12. Wittemberg. 1770. Larsen, C |
| 579. | Larsen, C See Michael, L. G. |
| | Latte |
| 580. | Laurent, EmileSur l'existence d'un principe toxique pour le |
| | Pourier, dans les baies, les graines et les plantules du Gui. C. R. Acad. Sci. Paris. 1901. |
| 581. | Lavinder, C. H Pellagra à Précis. Pub. Treas. Dept. Marine Hospital Service. U. S. A. 22. 1 pl. Wash- |
| F00 | ington, 1908. |
| 582. 582a. | Law, James |
| 583. | Leather, J. W |
| 584. | Leclerc, N |
| | doit employer pour constater. Strasburg. 1803. |
| 585. | Lefebvre, G See Herissey, Ch. |

| 586. | Legrand du Saulle, H., Berryer, G., and Pouchet, |
|--------------|--|
| | GTraité de médicine légale, de jurisprudence médicale et de toxicologie, etc. Ed. 2. Paris. 1886. |
| 587. | Legue, G |
| 588. | Lehmann, OttoGiftpflanzen mit besonderer Berücksichtigung der wirksamen Stoffe und zahlreichen in den Text gedruckten Illustrationen. 126. Hamburg. 1882. |
| 589. | Lehmans, K. B., and Mori, R |
| | men von Agrostemma Githago. Archiv. f. Hyg. 9:257. (Bot. Gaz. 14:233). L'Engle, E. MSee Price, T. W. |
| 590. 591. | Leuterer, Dr Investigations of Poisonous Characters of |
| 591a. | Macrozamia. Chem. and Drug. 1896:171. Levaditi, A |
| | italiana de veterinaria. 388. Milan. 1899. Lewandowski, F |
| 592a. | Beiträgen zur Kenntnis der Staphylo-und |
| 593. 594. | f. Dermatol. u. Syphilis. 94. Heft. 2 u. 3. Lewin, L |
| 571. | Arrow Poisons. Review of Different Kinds and their Sources. Phar. Centrbl. 1895 : 599-600. See also Naturw. Runds. 1895 : 386. (Abs. in Proc. Am. Phar. Assoc. 44 : |
| 595. | 508-509) Erste Hilfeleistung bei Vergiftungen. Cong. internat. d'hyg. et de demog. C. r. 1894, |
| 596. | Budapest, 1896. 8:286-294. pt. 5 |
| 597. | Gaz. 4:145 Lehrbuch der Toxicologie. Ed. 2. x+509. 7f. |
| 598. | I plDie Pfeilgifte. Historische und experimentelle |
| 599. | Untersuchungen, 152. Berlin, 1894, Ueber Anhalonium Lewinii and andere giftige |
| 600 | Cacteen. Ber. der Deutsch. bot. Gesell. 1894:283-290. |
| 600. | |
| 601. | Liautard, A Poisoning of Horses by Snow Drop. Am. Vet. Rev. 30:298. |
| 602. | Ligouzat Empoisonnement par la racine d'un chardon (Atractylis gummifera, el heddad des Arabes). Arch de med. et phar. mil., Paris. 37:164-168. 1901. |
| 603. | Lilienfeld, Fr Dissertatio circa Phytotoxicologiam cechicam, plantas venenatas Cechiae indigenas Umbelli- |
| 603a. | feras exhibens, 108. Prague, 1834. Lindenberg, AdolphUn nouvean mycétome. Arch. de Parasit. |
| 604. | Lindley, JohnFlora Medica. A Botanical Account of all |
| | the more Important Poisonous Plants Used in Medicine in different Parts of the World. 656. London. 1838. |
| 605. | Lindsay, W. LThe Toot-poison of New Zealand. Phar. Jour. II. 5:371-373. 1864. (Proc. Brit. Assoc.) |

| | | D D 1 1 21 11 1 1771 |
|--|----------------------------|--|
| 606. | Linné, Carl v | De Raphanis. 21. Upsala. 1763. Recherches morphologiques sur le champignon. |
| 607. | Linossier, G. and Roux, G. | du muguet. Arch. exp. med. et d' anat. |
| | | pathol. 1890 :63-87. |
| 608. | | Sur la morphologie et la biologie du Cham- |
| | | pignon du muguet. Compte Rend. Acad. |
| 609. | | Sci. 1889. Sur la mycose expérimentale due au cham- |
| 009. | | nignon du muguet Lyon medical 1889. |
| 610. | List, A | . Untersuchungen uber die in und auf dem |
| | | Körper des gesunden Schafes vorkommen- |
| | | den niederen Pilze. Dissertation. Leipsic. 1885. |
| 6102 | Little | On Poisoning by Sium sp. Clinic. Cincinnati. |
| | | 7 :49. 1874. |
| 611. | Lloyd, C. G | . Mycological Notes. Cincinnati, Series of Pa- |
| | | pers covering many of the important fleshy |
| 6112 | Lloyd, J. U | fungi. See King John |
| 612. | Lloyd, J. U. and C. G | Drugs and Medicines of North America. I. |
| | | Ranunculaceae 304. Cincinnati, 1884-1885. |
| 613. | Lodeman, E. G | On Recurrence of Attacks of Rhus Poisoning |
| | | without Reëxposure. See Gar. and For. 8:398. |
| 614. | Loew. O | Ein natürliches System der Gift-Wirkungen. |
| | | Miinch 1893 |
| 615. | Loudon, J. C | An Encyclopedia of Plants. 1574. London, |
| (16 | T b. amaza. | 1886. On Poisoning from Foods. See Centrbl. f. |
| 616. | Lubeman | On Poisoning from Foods. See Centrbl. f. Bakt. 1906. I. Abt: 433. |
| 617. | Lucas, E | . De solano tuberoso ejusque principio nartico. |
| | | 32 Hirschherg 1846 |
| 618. | Lucet, A | De l'aspergillus fumigatus chez les animaux |
| | | domestiques et dans les oeufs en incubation. (etude clinique et expérimentale). 108. |
| | | Paris. 1899. |
| 619. | | . Also see Constantin, J. |
| 620. | Luerssen, Ch | Medicinisch-pharmaceutisch Botanik. Leipsic. |
| 620a. | Lyons A R | . Assay of Third Extract of Belladona Leaves. |
| 021. | | |
| 622. | | . Plant Names, Scientific and Popular. Ed. 2. |
| 622 | | |
| 623. | M-C-wiles Con-14 | 630. Detroit, 1907. |
| | McCarthy, Gerald | .The Poisonous Plants of North Carolina. |
| | McCarthy, Gerald | The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). |
| 624. | McClintock, C. F | .The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. .See Vaughan, V. C. |
| 624. 625. | McClintock, C. F | .The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. .See Vaughan, V. C. .On the Poisonous Influence of Cypripedium |
| | McClintock, C. F | The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. See Vaughan, V. C. On the Poisonous Influence of Cypripedium spectabile and C. pubescens, Minn. Bot. |
| 625. | McClintock, C. F | .The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. .See Vaughan, V. C. .On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894 :32-36. |
| | McClintock, C. F | The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. See Vaughan, V. C. On the Poisonous Influence of Cypripedium spectabile and C. pubescens, Minn. Bot. |
| 625. 626. | McClintock, C. F | . The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. . See Vaughan, V. C On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894:32-36. . Poisonous Influence of various Species of Cypripedium, Minn. Bot. Studies. 1895: 455-451. |
| 625. 626. | McClintock, C. F | . The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48 See Vaughan, V. C On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894:32-36 Poisonous Influence of various Species of Cypripedium, Minn. Bot. Studies. 1895: 455-451 Poisoning Horses by Spoiled Potatoes. Jour. |
| 625. 626. | McClintock, C. F | The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. See Vaughan, V. C. On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894:32-36. Poisonous Influence of various Species of Cypripedium, Minn. Bot. Studies. 1895: 455-451. Poisoning Horses by Spoiled Potatoes. Jour. Roy. Soc. England. 1897. Ser. III. (No. |
| 625. 626. | McClintock, C. F | . The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48 See Vaughan, V. C On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894:32-36 Poisonous Influence of various Species of Cypripedium, Minn. Bot. Studies. 1895: 455-451 Poisoning Horses by Spoiled Potatoes. Jour. |
| 625.626.627.628. | McClintock, C. F | . The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. . See Vaughan, V. C On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894:32-36. . Poisonous Influence of various Species of Cypripedium, Minn. Bot. Studies. 1895: 455-451. . Poisoning Horses by Spoiled Potatoes. Jour. Roy. Soc. England. 1897. Ser. III. (No. 29). 115-135. Abs. in Exp. Sta. Rec. 9:389. Impaction from Alfalfa Hay. Am. Vet. Rev. 38:238-239. |
| 625. 626. 627. | McClintock, C. F | . The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. . See Vaughan, V. C On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894:32-36. . Poisonous Influence of various Species of Cypripedium, Minn. Bot. Studies. 1895: 455-451. . Poisoning Horses by Spoiled Potatoes, Jour. Roy. Soc. England. 1897. Ser. III. (No. 29). 115-135. Abs. in Exp. Sta. Rec. 9:389. Impaction from Alfalfa Hay. Am. Vet. Rev. 38:238-239. . Fungi, Edible and Poisonous. Am. Jour. |
| 625.626.627.628.629. | McClintock, C. F | . The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48 See Vaughan, V. C On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894:32-36 Poisonous Influence of various Species of Cypripedium, Minn. Bot. Studies. 1895: 455-451 Poisoning Horses by Spoiled Potatoes. Jour. Roy. Soc. England. 1897. Ser. III. (No. 29). 115-135. Abs. in Exp. Sta. Rec. 9:389. Impaction from Alfalfa Hay. Am. Vet. Rev. 38:238-239 Fungi, Edible and Poisonous. Am. Jour. Phar. 68:651. |
| 625.626.627.628. | McClintock, C. F | . The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48. . See Vaughan, V. C On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894:32-36. . Poisonous Influence of various Species of Cypripedium, Minn. Bot. Studies. 1895: 455-451. . Poisoning Horses by Spoiled Potatoes. Jour. Roy. Soc. England. 1897. Ser. III. (No. 29). 115-135. Abs. in Exp. Sta. Rec. 9:389. Impaction from Alfalfa Hay. Am. Vet. Rev. 38:238-239. . Fungi, Edible and Poisonous. Am. Jour. Phar. 68:651. . Three Cases of Poisoning by Potassium Cyanide. Am. Jour. Med. Sci. 129:684. |
| 625.626.627.628.629. | McClintock, C. F | . The Poisonous Plants of North Carolina. Bull. N. C. Agr. Exp. Sta. (St. Bd. Agr.). 22:48 See Vaughan, V. C On the Poisonous Influence of Cypripedium spectabile and C. pubescens. Minn. Bot. Studies. 1894:32-36 Poisonous Influence of various Species of Cypripedium, Minn. Bot. Studies. 1895: 455-451 Poisoning Horses by Spoiled Potatoes. Jour. Roy. Soc. England. 1897. Ser. III. (No. 29). 115-135. Abs. in Exp. Sta. Rec. 9:389. Impaction from Alfalfa Hay. Am. Vet. Rev. 38:238-239 Fungi, Edible and Poisonous. Am. Jour. Phar. 68:651. |

| L. H. The Danger of Feeding Hay that Contains Ergot. Press Bull. Ia. Agr. Exp. Sta. 1908. 632a. MacQuean, James See Neumann. 632b. Madsen, B. A. See Michael, L. G. 633. Madsen, Thorwald Botulismustoxin. Chapter in von Kraus and Leviditi's Handbuch der Tech. u. Meth. etc. 1:137-143. (Contains Bibliography). 633a. Magnin, L. Un cas d'empoisonnement par l'Amanita muscaria. Bull. de la Soc. myc. de France. 19:173-175. 634. Maiden, J. H. Native Plants Poisonous to Stock. Agr. Gaz. 4:679. Plants Injurious to Stock. Agr. Gaz. 4:679. Plants Injurious to Stock. Agr. Gaz. 4:679. Plants Reported to be Poisonous to Stock in Australia. N. S. W. Dept. Agr. Misc. Pub. No. 477. 1901. 637. Poison lvy. Roy. Soc. N. S. Wales, 1908:3. Useful Native Plants of Australia including Tasmania. 693. London. 1889. 639. Maish, J. M. California Opium. Am. Jour. Phar. 46:106. 640. Poisonous Species of Astragalus. Am. Jour. Phar. Assoc. 13:166. 640. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. 641. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi Useful Native Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 643a. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 645a. Soc. Marchand, L. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 645a. See Numan, A. Poisons and their Treatment. Internat. Clinic. 4:48. Philadelphia, 1893. 645. Marchand, L. Drosons Mants. Jour. Dept. Agr. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 650a. Marshall, H. T. See Bacon, R. T. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1842. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de phagmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | | | |
|--|-------|-----------------------------------|--|
| 632b. Madsen, B. A. See Michael, L. G. 633. Madsen, B. A. See Michael, L. G. 633. Madsen, Thorwald Botulismustoxin. Chapter in von Kraus and Leviditi's Handbuch der Tech. u. Meth. etc. 1:137-143. (Contains Bibliography). 633a. Magnin, L. Un cas d'empoisonnement par l'Amanita muscaria. Bull. de la Soc. myc. de France. 634. Maiden, J. H. Native Plants Poisonous to Stock. Agr. Gaz. 635. Notes on Some American and Australian Plants Injurious to Stock. Agr. Gaz. 636. Some American and Australian Plants Injurious to Stock. Agr. Gaz. 637. Plants Reported to be Poisonous to Stock in Australia. N. S. W. Dept. Agr. Misc. Pub. No. 477. 1901. 638. Useful Native Plants of Australia including Tasmania. 693. London. 1889. 639. Maish, J. M. California Opium. Am. Jour. Phar. 46:106. 630a. Phars. Assoc. 13:166. 640. Poisonous Species of Astragalus. Am. Jour. Phar. 75:1239. 641. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi Ueher Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 24G:381. 643. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 644. Manuheim On Cocain. See Zeitsch. f. Klin. Med. 1890. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques, 94. 2 pl. Paris. 1861. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:43. Philadelphia. 1893. 659. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 12:33-338. 660. Marshall, H. T. See Bacon, R. T. Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 12:33-338. 671. Maschka, J. V. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Scidel, Th. Husemann, and A. Schauenstein.) 672. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de phagmacod. 673. 11:465-481. Brussels and Paris. 1903. 674. Maschka, J. V. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Scidel, Th. Huse | 632. | McNeil, J. H. and Pammel L. H. | The Danger of Feeding Hay that Contains |
| 633. Madsen, Thorwald Leviditi's Handbuch der Tech. u, Meth. etc. 1:137-143. (Contains Bibliography). 634. Maiden, J. H | 6322 | MacQuean James | Ligot. Press Bull. 1a. Agr. Exp. Sta. 1908. |
| 633. Madsen, Thorwald Leviditi's Handbuch der Tech. u, Meth. etc. 1:137-143. (Contains Bibliography). 634. Maiden, J. H | 632b. | Madsen, B. A | See Michael, L. G. |
| Leviditi's Handbuch der Tech. u, Meth. etc. 1:137-143. (Contains Bibliography). | 633. | Madsen, Thorwald | Botulismustoxin. Chapter in you Kraus and |
| 1:137-143. (Contains Bibliography). Un cas d'empoisonnement par l'Amanita muscaria. Bull. de la Soc. myc. de France. 19:173-175. | | | Leviditi's Handbuch der Tech. u. Meth. etc. |
| caria. Bull. de la Soc. myc. de France. 19:173-175. 634. Maiden, J. H Native Plants Poisonous to Stock. Agr. Caz. 6:57-58. New South Wales. Notes on Some American and Australian Plants Injurious to Stock. Agr. Gaz. 4:679. 636. Plants Reported to be Poisonous to Stock. Notes on Some American and Australian Plants Injurious to Stock. Agr. Gaz. 4:679. 637. Plants Reported to be Poisonous to Stock. No. 477. 1901. 638. Useful Native Plants of Australia including Tasumania. 693. London. 1889. 639. Maish, J. M. California Opium. Am. Jour. Phar. 46:106. 639a. On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. 640. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. 641. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi Ueher Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 246:381. 643. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550:561. 643a. Mann, J. D. Forensic Medecine and Toxicology. London and Philadelphia. 1893. 644. Mannheim On Cocain. See Zeitsch. f. Klin. Med. 1890. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. 94. 2 pl. Paris. 1861. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia. 1898. 647. Marek, J. See Huyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bnll. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 12:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:336. 662. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 663. Massey, W. F. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) | | | 1:137-143 (Contains Ribliography) |
| 634. Maiden, J. H. Native Plants Poisonous to Stock. Agr. Gaz. 6:57-58. New South Wales. 635. Notes on Some American and Australian Plants Injurious to Stock. Agr. Gaz. 4:679. 636. Plants Reported to be Poisonous to Stock in Australia. N. S. W. Dept. Agr. Misc. Pub. No. 477. 1901. 637. Poison Ivy. Roy. Soc. N. S. Wales, 1908:3. 638. Useful Native Plants of Australia including Tasmania. 693. London. 1889. 639. Maish, J. M. California Opium. Am. Jour. Phar. 46:106. 639a. On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. See Proc. Am. Phar. Assoc. 20:233-236. 640. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. Phar. 51:239. 641. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi Ueber Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 246:381. 643. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 643a. Mann, J. D. Forensic Medecine and Toxicology. London and Philadelphia. 1893. 644. Mamnheim On Cocain. See Zeitsch. f. Klin, Med. 1890. 380. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. 94. 2 pl. Paris. 1861. See Numan. A. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 4:48. Philadelphia. 1898. | 633a. | | caria. Bull. de la Soc. myc. de France. |
| 635. Notes on Some American and Australian Plants Injurious to Stock. Agr. Gaz. 4:679. 636. Plants Reported to be Poisonous to Stock in Australia. N. S. W. Dept. Agr. Misc. Pub. No. 477. 1901. 637. Poison Ivy. Roy. Soc. N. S. Wales. 1908;3. 638. Useful Native Plants of Australia including Tasmania. 693. London. 1889. 639. Maish, J. M. California Opium. Am. Jour. Phar. 46:106. 639a. On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. 640. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. 641. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi Ueber Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 246:381. 643. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 644. Mannheim On Cocain. See Zeitsch. f. Klin. Med. 1890. 380. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. 94. 2 pl. Paris. 1861. 645a. See Numan, A. See Numan, A. See Numan, A. See Numan, A. See Hutyra, Fr. Gew. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 37:38. 647. Marek, J. See Hutyra, Fr. Gew. Marsh, C. W. The Loco Weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 12:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:33-38. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Hussemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de phatpmacod. 11:465-481. Brussels and Paris. 1903. | 634. | Maiden, J. H | Native Plants Poisonous to Stock. Agr. Gaz. |
| Plants Injurious to Stock. Agr. Gaz. 4:679. Plants Reported to be Poisonous to Stock in Australia. N. S. W. Dept. Agr. Misc. Pub. No. 477. 1901. Associated to be Poisonous to Stock in Australia. N. S. W. Dept. Agr. Misc. Pub. No. 477. 1901. Poison Ivy. Roy. Soc. N. S. Wales. 1908;3. Useful Native Plants of Australia including Tasmania. 693. London. 1889. Galifornia Opium. Am. Jour. Phar. 46:106. On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. Poisonous Species of Astragalus. Am. Jour. Phar. Assoc. 13:166. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. G42. Makosi. Ucher Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 246:381. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. G43a. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. G444. Mannheim. On Cocain. See Zeitsch. f. Klin. Med. 1890. G45. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. G45. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. G45. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. G46. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. G47. Marck, J. See Hutyra, Fr. G48. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. G49. Results of Loco-weed Investigation in the Field. Bull. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 12:37-38. G50. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. G50. Marshall, H. T. See Bacon, R. T. G51. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) G52. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. Massey, W. F. | 625 | | 6:57-58. New South Wales. |
| 636. Plants Reported to be Poisonous to Stock in Australia. N. S. W. Dept, Agr. Misc. Pub. No. 477. 1901. 637. Poison Ivy. Roy. Soc. N. S. Wales, 1908;3. 638. Useful Native Plants of Australia including Tasmania. 693. London. 1889. 639a. On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. 640. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. 641. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi Ueber Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 2-16:381. 643. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 644a. Mannheim On Coccain. See Zeitsch. f. Klin. Med. 1890. 380. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques, 94. 2 pl. Paris. 1861. 645a. See Numan, A. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia. 1898. 647. Marck, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 12:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauen. 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. | 035. | | Plants Injurious to Stools Asia Con 4.670 |
| No. 477. 1901. Poison Ivy. Roy. Soc. N. S. Wales, 1908:3. Useful Native Plants of Australia including Tasmania. 693. London. 1889. California Opium. Am. Jour. Phar. 46:106. On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. Phar. 51:239. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 236. Ueber Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 2-46:381. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-565. Mann, J. D. Forensic Medecine and Toxicology. London and Philadelphia. 1893. Forensic Medecine and Toxicology. London and Philadelphia. 1893. See Zeitsch. f. Klin. Med. 1890. 380. See Numan, A. See Numan, A. See Numan, A. See Numan, A. See Hutyra, Fr. See Bacon, R. T. See See See See See See See See See S | 636. | | Plants Reported to be Poisonous to Stock in |
| No. 477. 1901. Poison Ivy. Roy. Soc. N. S. Wales, 1908:3. Useful Native Plants of Australia including Tasmania. 693. London. 1889. California Opium. Am. Jour. Phar. 46:106. On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. Phar. 51:239. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 236. Ueber Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 2-46:381. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-565. Mann, J. D. Forensic Medecine and Toxicology. London and Philadelphia. 1893. Forensic Medecine and Toxicology. London and Philadelphia. 1893. See Zeitsch. f. Klin. Med. 1890. 380. See Numan, A. See Numan, A. See Numan, A. See Numan, A. See Hutyra, Fr. See Bacon, R. T. See See See See See See See See See S | 000. | | Australia, N. S. W. Dept. Agr. Misc. Pub. |
| Useful Native Plants of Australia including Tasmania. 693. London. 1889. 639. Maish, J. M. California Opium. Am. Jour. Phar. 46:106. 640. On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. 640. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. 641. See Ramination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 643. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 644. Mannheim On Cocain. See Zeitsch. f. Klin. Med. 1890. 380. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. 94. 2 pl. Paris. 1861. 645. See Numan, A. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia. 1898. 647. Marek, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | | | No. 477. 1901. |
| Tasmania. 693. London. 1889. 639a | | | Poison Ivy. Roy. Soc. N. S. Wales. 1908:3. |
| On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. 640. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. 641. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi Ueber Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 246:381. 643. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 643a. Mann, J. D. Forensic Medecine and Toxicology. London and Philadelphia. 1893. 644. Mannheim On Cocain. See Zeitsch. f. Klin. Med. 1890. 380. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. 94. 2 pl. Paris. 1861. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia. 1898. 647. Marck, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 12:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Massey, W. F. Massey and Used in North Carolina. Bull. N. C. | 638. | | Useful Native Plants of Australia including |
| On Rhus Toxicodendron. See Proc. Am. Phar. Assoc. 13:166. 640. Poisonous Species of Astragalus. Am. Jour. Phar. 51:239. 641. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi Ueber Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 246:381. 643. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 643a. Mann, J. D. Forensic Medecine and Toxicology. London and Philadelphia. 1893. 644. Mannheim On Cocain. See Zeitsch. f. Klin. Med. 1890. 380. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. 94. 2 pl. Paris. 1861. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia. 1898. 647. Marck, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 12:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Massey, W. F. Massey and Used in North Carolina. Bull. N. C. | 630 | Maich I M | California Ocium Am Jour Phor 46:106 |
| Phar, Assoc. 13:166. Poisonous Species of Astragalus. Am. Jour. Phar, 51:239. 641 | 639a | Maisii, J. M | On Rhus Toxicodendron See Proc Am |
| Phar. 51:239. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi | 0074. | | Dhar Assas 12.166 |
| Phar. 51:239. Sneezeweed. Proc. Am. Phar. Assoc. 20:233-236. 642. Makosi | 640. | | Poisonous Species of Astragalus. Am. Jour. |
| 642. Makosi Ueber Alkaloide der chinesischen Corydalis Knollen. Arch. der Phar. 246:381. 643. Mann, E. A. Examination of the Western Australian Poison Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 643a. Mann, J. D. Forensic Medecine and Toxicology. London and Philadelphia. 1893. 644. Mannheim On Cocain. See Zeitsch. f. Klin. Med. 1890. 380. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. 94. 2 pl. Paris. 1861. 645a. See Numan, A. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia. 1898. 647. Marck, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | | | Phar. 51 :239. |
| Son Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 643a. Mann, J. D | 641. | | 226 |
| Son Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 643a. Mann, J. D | 642. | Makosi | Ueber Alkaloide der chinesischen Corydalis |
| son Plants. Jour. Dept. Agr. W. Austr. 12:550-561. 643a. Mann, J. D | 642 | 3. F T ⁵ . A | Knollen, Arch, der Phar. 2-46:381. |
| and Philadelphia. 1893. 644. Mannheim On Cocain. See Zeitsch. f. Klin. Med. 1890. 380. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. 94. 2 pl. Paris. 1861. 645a. See Numan, A. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia. 1898. 647. Marck, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | | Mann, E. A | son Plants. Jour. Dept. Agr. W. Austr. |
| 644. Mannheim On Cocain. See Zeitsch, f. Klin, Med. 1890. 380. 645. Marchand, L. Du Croton Tiglium recherches botaniques et therapeutiques. 94. 2 pl. Paris. 1861. See Numan, A. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia. 1898. 647. Marek, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | 643a. | | and Dhiladalahia 1902 |
| 380. Du Croton Tiglium recherches botaniques et therapeutiques, 94. 2 pl. Paris, 1861. See Numan, A. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia, 1898. 647. Marek, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | 644 | Manuhaim | On Cocain See Zeitsch f Klin Med 1890 |
| therapeutiques, 94. 2 pl. Paris, 1861. Sec Numan, A. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia, 1898. 647. Marek, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | 011. | Mannicini | |
| therapeutiques, 94. 2 pl. Paris, 1861. Sec Numan, A. 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia, 1898. 647. Marek, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | 645. | Marchand, L | Du Croton Tiglium recherches botaniques et |
| 646. Marcus, H. D. Poisons and their Treatment. Internat. Clinic. 1:48. Philadelphia. 1898. 647. Marck, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | - 45 | | therapeutiques 94 2 nl Paris 1861 |
| 4:48. Philadelphia. 1898. 647. Marsk, J. See Hutyra, Fr. 648. Marsh, C. W. The Loco Weed Disease. Far. Bull. U. S. Dept. Agr. 380:16. 649. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | | M II D | See Numan, A. |
| 647. Marek, J | 040. | | 4.48 Philadelphia 1808 |
| Dept. Agr. 380:16. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | 647 | Marek I | See Hutyra, Fr. |
| Dept. Agr. 380:16. Results of Loco-weed Investigation in the Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | | Marsh. C. W | The Loco Weed Disease, Far. Bull. U. S. |
| Field. Bull. Bur. Pl. Ind. U. S. Dept. Agr. 121:37-38. 650. Marsh, D. C. Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | | | Dept. Agr. 380:16. |
| 121:37-38. 650. Marsh, D. C Water Bloom. Bull. Wis. Geol. and Nat. Hist. Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | 649. | ••••• | |
| Sur. 12:36. 650a. Marshall, H. T. See Bacon, R. T. 651. Maschka, J. v. Handbuch der gerichtlichen Medicin, 4 v. Tübingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | | | 121 - 37 - 38 |
| 650a. Marshall, H. T | 650. | Marsh, D. C | Water Bloom. Bull. Wis. Geol. and Nat. Hist. |
| 651. Maschka, J. v | 650a. | Marshall H. T | See Bacon, R. T. |
| bingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauenstein.) 652. Masoin, P. De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F. Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | 651. | Maschka, J. v | Handbuch der gerichtlichen Medicin, 4 v. Tü- |
| 652. Masoin, P De la rapidité d'absorption des poisons par l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | | , , | bingen, 1831-1832. (The Toxicology is contained in Vol. 2, and is by B. Schuchardt, M. Seidel, Th. Husemann, and A. Schauen- |
| l'organism. Arch. Internat. de pharmacod. 11:465-481. Brussels and Paris. 1903. 653. Massey, W. F Medicinal Plants which have been Collected and Used in North Carolina. Bull. N. C. | 652 | Masoin P | De la rapidité d'absorption des poisons par |
| 653. Massey, W. F | 052. | masom, 1 | l'organism. Arch. Internat, de pharmacod. |
| and Used in North Carolina, Bull. N. C. | | | 11:465-481. Brussels and Paris. 1903. |
| and Used in North Carolina, Bull. N. C. | 653. | Massey, W. F | Medicinal Plants which have been Collected |
| 5.1 -41-1/1 | | | and Used in North Carolina, Bull. N. C. Agr. Exp. Sta. 150:331-409. |

| 653a | Matruchot, L. and Dasson- | |
|-------|--|--|
| ooda. | ville, C. | On Dermatomycosis of Fowls. Rev. Gen. Bot. 11:429-444. Abs. in Exp. Sta. Rec. 12:94. |
| 654. | Maurizio, AdamL | es Maladies causes aux poisons et aux ocufs |
| 655. | Mayo, N. S | par les champignons. Revue Myc. 1897 (No. 75): 79-85. pl. 170-172. Enzoötic Cerebritis, or "Staggers" of Horses. |
| 655a. | L C | Bull. Kan. Agr. Exp. Sta. 24:107-116. Some Observations upon Loco. Bull. Kan. |
| | | Agr. Exp. Sta. 35:113-119. Flora Habanera. Fanerogams. 1:268; 2:269-597. |
| 656. | Mead, Richard | Mechanical Account of Poisons. London. |
| 657. | Mee, George | 174. Poisonous Principle of the Bitter Cassava |
| 658. | | Root. Phar. Jour. II. 6 :322-333. On the Absorption of Strychnine and Hydro- |
| | | cyanic Acid from the Mucous Membrane of the Stomach; an Experimental Study on Rabbits. Trans. Assoc. Am. Phys. 11 :221- 228. Philadelphia. 1896. |
| 659. | Melvin, A. D. and Mohler, | Dermal Mycosis Associated with Sarcoptic |
| | | Mange in Horses. Proc. Am. Vet. Med. Assoc. 1907 :175. 2 pl. See also Rept. Bur. An. Ind. 24 :259-275. |
| 660. | Meiner | Empoisonnement par l'Amanita phalloides à Noir-Montier (Vendée). Bol. Soc. Myc. de France. 24 :68-71. |
| 661. | Merck, E | Annual Report of Recent Advances in Pharmaceutical Chemistry and Therapeutics. 23. See also earlier Volumes. See Merck's Weekly Report. |
| 662. | Merrill, E. D | The Occurrence of Antiaris in the Philippines. |
| 663. | Mever, Arthur | Phil. Jour. Sci. Sect. C. 2:111. Beiträge zur Kentniss pharmaceutisch wichti- |
| 664. | Meyer, H. | C A1 J Dl |
| 665. | Michael, L. G., with the Collaboration of E. T. Robbins, Carl Larsen, B. A. Madson, Stella Hart- zell, Claude Kinnebrew. | |
| | and George Godfrey | The Influence of Feeding Sugar Beets and Mangels to Breeding Animals with Refer- |
| | | ence to the Formation of Renal and Urinary |
| 666. | Miquel, F. A. W | Calculi. Bull. Ia. Agr. Exp. Sta. 112:35-77. De Nord-Nederlandsche vergiftige Gewassen. 220. Amsterdam, 1838. |
| 667. | Millspaugh, C. F | American Medical Plants. An Illustrated and Descriptive Guide to the American Plants Used as Homeopathic Remedies, their History, Preparation, Chemistry, and Physiological Effects. 1:99. pl. 1-99; 2:100-190. pl. 100-180. |
| | Mitchell, S. R | See Hare, R. F. |
| 668. | Mitlacher, W | Toxicologisch oder forensisch wichtige Pflanzen und vegetabilische Drogen, mit besonderer Berücksichtigung ihrer mikroskopischen Verhältnisse. Berlin and Vienna. 1904. |
| 669. | Moeller, JI | Digitalis and Verbascum. Phar. Post. 37: 677-680. |

| 669a. 670. | Moeller, Josef |
|---------------|--|
| 671. | Mohler, J. R |
| 672. 673. | Mohler, J. R See Melvin, A. D. |
| 0,01 | Mohler, J. R. and Buckley, J. S |
| 673a. | U. S. Dept. Agr. 20:122-138. pl. 6-10. Mohr, Ch |
| 674. | Alabama, 1901. Molle, PhLa localization des alkaloides dans les So- |
| 675 | lancées. Bull. de la Société belge de microscopie. 21:8-20. 1894-1895. |
| 675. | Hyde, J. N |
| 676. | Moore, G. T |
| 677 | Mori, R See Lehmann, K. B. |
| 678. | Moorman, J. W On Poisoning from Internal Administration of Rhus Preparations. See Am. Jour. Med. Sci. 51:570. |
| 679. | mics or Snake-bite Antidotes. Annals of |
| (00 | Botany, 1:153-161. |
| 680. 681. | Morrow, P. A |
| | C. D. Poisonous Properties of Wild Cherry Leaves. Bull. N. H. Agr. Exp. Sta. 56 :113-123. |
| 682. | Moussu and Desaint Poisoning of Sheep by Galega officinalis, Rec. Med. Vet. 84:562-569. Abs. in Exp. Sta. Rec. 19:785. |
| 683. | Moutet |
| | tion de la fougère mâle et de l'huile de ricin. Bull. med. de l'Algérie. 1 :263, ser. 2. Algiers. 1903. |
| 684. | Mueller, Ferdinand Von Notes on Leguminous Poisonous Plants of Southwest Australia. Aus. Chem. and Drug. 1884. |
| 685. | |
| 685a. | Second Systematic Census of Australian Plants, Pt. 1, 224, Melbourne 1889 |
| 686. | Select Extra-Tropical Plants readily Eligible |
| | for Industrial Culture or Naturalization. With Indications of their Native Countries and Some of their Uses, 449. Detroit, 1884. |
| 687. | Mueller, Georg and Krause. |
| | C |
| 688. | Thierheilkunde. 23:326-335. Mueller, O |
| 690 | 1907. 20 :37. |
| 689. 690. | Mueller, Karl |
| 691. | Murrell, WWhat to do in Cases of Poisoning. Ed. 10. London. 1907. |
| | |

| 692. | Muter, John |
|--------------|---|
| 693. | Mutual Life Insurance |
| (04 | Company of New York. Poisons: Remedies, 45. New York. 1903. De Nabias and Dupony Sur un poison d'epreuve du Haut. Oubanghi. |
| 694. | Jour, de med. de Bordeaux. 33:273. 1903. |
| 695. | Jour. de med. de Bordeaux. 33:273. 1903. Nagelvoort, J. B |
| 695a. | Neal, H. VSee Davenport, C. B. |
| 696. | Nelson, A |
| 697. | Nelson, S. B Feeding of Wild Plants to Sheep. Rept. U. S. |
| 698. | Dept. Agr. Bur. An. 1nd. 15:425Feeding Wild Plants to Sheep. Bull. Wash. |
| 700 | Agr. Exp. Sta. 73:64. Nestler, A |
| 699. | Festsch. 1908 :200-206. |
| 700. | Das Sekret der Drüsenhaare der Gattung Cyp- |
| | ripedium mit besonderer Berücksichtigung seinerhautreizenden Wirkung. Ber. der |
| | Deutsch, Bot, Gesell, 25:554-567, 1 bl |
| 701. | Die hautreisende Wirkung der Primula Obcon- |
| | ica Hance and Primula sinensis Lindl, Sep. Ber, der Deutsch. Bot. Gesell. 18 :189-202, |
| *** | pl. 7-8. |
| 702. 703 | |
| 700. | Diseases of the Domesticated Animals. |
| | Trans, by Fleming. Ed. 2. Rev. and Rev. by Jas. MacQueen. 697. New York. 1907. |
| 703a. | Nichols, H. J See Phalen, J. M. |
| 704. 705. | Nobecourt |
| | 20 :570. |
| 706. | Noe, J |
| | bernants (herissons); variabilité et specif- icité des effets des substance toxiques. |
| | Archiv. internat. de pharmacod. 12:153- |
| 707. | 158. Brussels and Paris. 1903. Noel, CharlesContributions à l'étude médicale de l'Anemone |
| | Pulsatilla. 81. Nancy. 1897. |
| 708. | Noel, Ch. and Lambert, L. Recherches experimentales sur l'Anemone Pul- satilla. Arch, inter. de phar. 1897 :Fasc. |
| | 1-2 |
| 709. | Nossrey, EPlantes vénéneuses croissant dans les prairies et dans les artificiels. Agriculture rationelle. |
| | 1898. No. 15. |
| 710. 711. | Noll, Fritz See Strasberger, E. Novy, F. G The Toxic Effects of the Bacillus of Hog |
| | Cholera Med News 2:231-237 |
| 712. | See also McClintock, C. F. |
| 713. | Nuckolds, Coleman Mycotic Dermatitis. Am. Vet. Rev. 25:635-659. |
| 714. | Numan, A. and Marchaud, |
| | I |
| | domestiques par les productions crypto- gamiques, 115. 5 pl. Gröningen, 1830. |
| 715. | Numb. Joshua Vetermary Toxicology. New York 1901-1907 |
| 716. | Oesterle See Tschirsch, A. Oliver, F. W See Kerner, Anton von. |
| 717. | Oliver, F. WSee Kerner, Anton von. |

| 718. | Oppenheim, Carl |
|--------------|---|
| 719. 720. | Orcutt, C. R The Calandrina Plant. West American Sci- |
| 721. | entist. 7 :190 The Loco Weed. do. 7 . |
| 722. | Orfila, M. J |
| 723. | Traité des poisons tirés des règnes minéral, végetal, et animal ou Toxicologie générale. Paris. 1813. |
| 724. | Traité de Toxicologie. Ed. 5. Reviewed, Corrected and Considerably Enlarged. 2 v. |
| 725. | Paris. 1852. Ott, J |
| | Otto, K |
| | Pabisch, H |
| 727 a | Page, C. GSporothrix and Epizootic Lymphangitis. Jour. Inf. Dis. 23:137-149. (Contains Bibliography). |
| 727b 727c | Paige, J. B |
| | de cultures jeunes de sporotrichose, broyées, dilués dans du sérum et stérilisées. Compt. |
| | Pallin, W. A A Treatise on Epizootic Lymphangitis. Lon- |
| | Pammel, L. HBeggiotoa alba and the Dying of Fish in Iowa. Abs. in Proc. Ia. Acad. Sci. 1889 :90-91. (Vol. 1.) |
| 728a 729. | Loco Weeds. Vis. Medicatrix. 1:40. |
| 730. 731. | Mechanical Injuries and Obstructions from Grasses. See Bull, Ia. Geol. Sur. 1:176-182. Is Parsnip Poisonous? Gar. and For. 8:228. Poisonous Effects of Grasses. See Bull. Ia. |
| 732. | Geol. Sur. 1:171-176 |
| 733. | Sta. 28:215-228. Poisonous and Medical Plants of Missouri. |
| 734. | Rull Mo St Ed Hort 14:46 |
| , DT. | Poisonous Weeds. See pp. 423-448 in "Some Weeds of Iowa." Bull, Ia. Agr. Exp. Sta. |
| 735. | Squirrel-tail Grass or Wild Barley. Bull, Ia. |
| 736. | Agr. Exp. Sta. 30:302-319. Vergiftung durch Wasserschierling. (Cicuta virosa, var. maculata.) Phar. Rund. 13: 102-103. 4 f. |
| 737. 738. | Pammel, L. H. and Fogel, |
| | E. D A Catalogue of the Poisonous Plants of Iowa. Proc. Ia, Acad. Sci. 14 :147-172. |

| 739. | Pannuel, L. H., Ball, C. R., and Scribner, F. L Grasses of Iowa Pt. 2. 436. Bull. Ia. Geol. |
|-----------------------|---|
| 740. | and Scribner, F. L Grasses of Iowa Pt. 2. 436. Bull. Ia. Geol. Sur. Suppl. Rept. 1904. Pannicl, L. H., Weems, J. B., and Scribner, F. L Grasses of Iowa. Bull. Ia. Geol. Sur. 1:525. |
| 741. | Park, R Dermatitis venenata, or Rhus Toxicodendron |
| 742. | found in the Waters of the State, July, 1887- June, 1889. Report. St. Bd. Health of Mass. 1890:581-620. |
| 743. 744. 745. | Parker, W. TLoco-Weed. Science. 23:101. Patek, JohannDie Giftpflanzen. Prague. 1866-1867. Paul, B. H. and Cownley, |
| 746. | A. H |
| 747. | giften. 87. Leyden. 1903. Pavesi, VStudi comparativi su tre specie si papaveri |
| 748. | Payne, G. T |
| 749. | Am. Pharm. Assoc. 42:135. Pearson, L |
| 749a. | |
| 750. 751. | Peck, Charles H Annual Report of the State Botanist of New York. 43 colored Plates of Edible and Poisonous Fungi with Full Descriptive Text. |
| 752. 753. | 241. Albany. 1897 |
| 754. | Pedler, A. and Warder, H.On the Nature of the Toxic Principle of the Aroideae. Jour. Asiatic Soc. of Bengal, Calcutta. 1890:106-117. |
| 755. | Pellagra See Boston Evening Transcript. "The Clinic" |
| 756. 756a. 757. | |
| 758. 758a. | Penhallow, D. PRhus Poisoning. Gar. and For. 8:359. Penn, W. AThe Soveraine Herb. A History of Tobacco. 326. London. 1901. |
| 759. | Percival, C. H |
| 760. | Brit, and Foreign. 32:184. Percy, S. RAn Inquiry into the Properties of Veratrum |
| 761. | viride. Trans. Am. Med. Assoc. Perkins, J The Leguminosae of Porto Rico. Cont. U. S. |
| 762. | Nat. Her. 10:220, Pernossi, L |
| 763. | Perredes, P. E. F The Anatomy of the Stem of Derris uliginosa, Benth. Bull. Wellcome Chem. Res. Lab. 33:10. |
| 764. | Perret, A. HRecherches des poisons pruritants dans les végétaux. C. R. Soc. de Biol. Paris. 59:602- |
| 765. | Perrot, Em |

| • | |
|--------------|---|
| 766. 767. | Peters, A. T |
| | Kaffir Corn. Bull. Neb. Agr. Exp. Sta. 77: |
| 768. | Peterson, F. and Haines, |
| 700. | A. FAmerican Text-Book of Legal Medicine and |
| 769. | Peterson, Maude G How to Know Wild Fruits. A Guide to Plants when not in Flower, by Means of Fruit and Leaf 340, 1905 |
| 770. 771. | Peyre, A |
| | Dot Com V 1009.15 16 |
| 772. | On the Active Principle of Rhus Toxicodendron and Rhus venenata, Jour. Exp. Med. |
| 773. | Poisonous Plants, Phar, Rund, 9:8, 1891. |
| 774. | |
| 775. | Pharmacopoeia Swedica, Ed. 7. 290. Stock- |
| 776. | holm. 1899. Review. Pfaff, Fr. and Orr, S. SOn Rhus Toxicodendron. See Science n. s. |
| 776a. | 1:119 Pharmacopoeia of the United States of Amer- |
| 776h | ica. 8th. Dec. Rcv. 1905. (U. S. P. VIII.) Phalen, J. M., and Nichols, |
| | H. J |
| 777. | The Military Surgeon. 242. 1909. Phillips |
| 77 8. | See Trans. Me. Med. Assoc. 10:147, 1889. Phoebus, Philipp Deutschlands kryptogannische Giftgewächse in Abbildungen und Beschreibungen. 114. 9 |
| 779. | Abbildungen und Beschreibungen. 114. 9 col. pl. Berlin. 1838. Pictet, ALa constitution chimique des alcaloides végétaux. 1897. |
| 779a. | Quelques considérations sur la genèse des alca- |
| | loides dans les plantes. Archiv. des Sci. |
| 780. | |
| 781. | Pictet, A. and Court, G Sur quelques nouveaux alcaloides végétaux. Bull. Soc. Chim. France. IV. 1:1001-1016. |
| 782. | Pictet, A. and Wolffenstein, |
| 702 | R See Wolffenstein, R. |
| 783. | Piffard, H. GMateria Medica and Therapeutics of the Skin. 97. New York. 1881. |
| 7 84. | |
| 785. 786. | Pilzmerkblatt Berlin. 1906. Pinkham On Gaultheria Poisoning. See Trans. Mass. |
| 786a. | MedLeg. Soc. 1878:87. Boston. Piper, C. VFlora of Washington. Cont. U. S. Nat. Herb. |
| 787. | Pishek, A Die Giftpflanzen in der Umgebung von Cilli. Program der K. R. Staats-Gymnasiums in |
| 788. | Cilli. 1884 :513. Planchon, GustavePlantes qui fournissent Curare. Jour. de Phar. |
| 789. | 1880. Paris Sur les plantes qui servent de base aux di- |
| | vers Curares. Compte. Rend. 90:133. Paris. |

| 790. | Planchon, Louis | Accidents causes par le contact du Rhus Toxicodendron. Montpellier Médical. 1887: Ju. and Sept. |
|--------------|---|--|
| 791. 792. | Plant Poisons | |
| 7 93. | | 1840. Beitrag zur systematischen Stellung des Soorpilzes in der Botanik. Leipsic. 1885. |
| 7 94. | | Das organisirte Contagium der Schafpocken und die Mitigation desselben nach Tous- saint's Manier. Leipsic. 1882. |
| 7 95 | | Die Hyphenpilze oder Eumyceten in W. Kolle u. A. Wasserman Handbuch der pathogenen Mikroorganismen. 1:526-660. f. 55. Jena. 1903. |
| 7 96. | Plenck, J. J | Toxicologia, seu doctrina de venenis ct anti- |
| 797. | • | dotis. Vienna. 1785. Six Fatal Cases of Poisoning by Amanita phalloides (Vaill) at Ipswich in September, 1907. Trans. Brit. Myc. Soc. 1908:25-26. |
| 798. | Plugge, P. C | Das Alkaloid von Sophora tomentosa, L. Archiv. der Pharm. 229:561-165. |
| 799. | | Andromedotoxin haltige Ericaceen. Arch. der |
| 800. | | Phar. 229:552-554. Giftiger Honig von Rhododendron ponticum. |
| 801. | | Arch, der Pharm. 229: 554-556. Ucber Andromedotoxin den giftigen Bestandheil der Andromeda Japonica. Phar. Zeitsch. |
| 802. | | f. Russland. 1883:609-613; 625-631. Die Wichtigiren Heilmittel in ihrer Wechselnden chemischen Zusammensetzung und |
| 803. | Pohl, J. E | Pharmakodynamischen Wirkung, Jena. 1866. De plantis venenatis umbelliferis. 4. Leipsic. |
| 203- | | 1771. See also Arch. f. Exp. Path. u. Ph. 34 :265. |
| | | 1894. |
| 804. | Poison Register, Universal | Containing also Poison Acts of the various States, Lists of Poisons in Groups, with Antidote Treatment, Maximum Doses, etc. Chicago. 1898. |
| 805. | Porcher, F. P | The Medical, Poisonous and Dietic Properties of the Cryptogamic Plants of the United States. 126. New York. 1854. From |
| 806. | | the Trans. Am. Med. Assoc. 7. Report on the Indigenous Medicinal Plants of South Carolina. Trans. Am. Med. Assoc. |
| 807. | ••••• | 1849. Resources of the Southern Fields and Forests. |
| 808. | Pouchet, G | Richmond. 1863. . Importance de l'action exercée par certains |
| | , | albuminoides sur la variation de toxicite des alcaloides et des principes actifs. Nouv. remedes. 15 :289-293, Paris, 1899, |
| 809. | Powell, R. D., Godlee, R. S and Taylor, H. H | |
| 0 | | tinomycosis hominis. Medchir. London. 1889:175-210. |
| 810. | Power, F. B | Notes on the So-called Loco-weeds. Phar. Rund. 8:29-38. |

| | 811. | Power, F. B. and Cambier, Jacob |
|---|----------------|--|
| | 812. | Praag, L. van, and Opwyrda, R. JLeerbuch voor practische giftleer. 3 Pts. |
| | 813. | Utrecht. 1871. Praschil, W. WPlantae venenatae in territorio Vindobonense |
| | | sponte crescentes. 40. Vienna. 1840. Prausnitz, C Die Heusebergifte. von Kraus and Levaditi's |
| | | Handbuch der Technik und Meth. etc. 1: 317-329. (Contains a Bibliography.) |
| | 814a. 814b. | Prentiss, A. N |
| | 815. | Prentiss, D. WFive Cases of Mushroom Poisoning, three of which Proved Fatal. Treatment of the Poisoning. Sep. Phil. Med. Jour. Sept. 24. |
| | 816. | 1898:16. 5 f |
| | 817. | Ther. Gaz. 1892:103-106. Prescott, A. BAlkaloidal Poisons. In: Text-book, Legal Medicine (Peterson and Haines). 2:447- |
| | 818. | 531. Laboratory Notes,—A Partial Analysis of the Oxytropis Lamberti. Am. Jour. Phar. 50: 564. |
| | 819. | Price, T. M Enzymes in Corn Stalks and their Relation to Cornstalk Disease. Cir. Bur. An. Ind. U. S. |
| | 820. | Dept. Agr. 84:75. Price, T. W. and L'Engle, E. M |
| | 821. 822. | 1904. Price, W., TranslatorSee Orfila, M. J. B. Prillieux, E. and Delacroix, |
| | | G Maladie de l'oeil produite par le Macrospo- rium parasiticum, Thum. Bull. de la Soc. Myc. de France. 1893 :201. |
| | 823. | Pritzel, G. A Thesaurus literaturae botanicae omnium gentium inde a rerum botanicarum initiis ad nostra usque tempora, quindecim milia op- |
| | 824. | erum recensens. 547. Leipsic. 1851. Proctor |
| | 825. 826. | Procupin, G |
| | 827. | 284. Puihn, J. GMateria veneraria regni vegetabilis. 196. Leip- |
| | | sic. 1785. Quain, E |
| | | Tubercular Lymphangitis of the Upper Extremity. St. Paul Med. 1904. (Aug.). Rabuteau, A Elémens de Toxocologie et de Médicine Lé- |
| | 828. | Rabuteau, A Elémens de Toxocologie et de Médicine Légale, applikuées à l'Empoisonnement. Paris. 1873. Ed. 2. Edited by E. Bourgoing. 1888 |
| | 829. | Radlkofer, L |
| , | 830. | Rafinesque, C. SMedical Flora: or Manual of the Medical Botany of the United States of North America. 1:268; 2:276. 48 pl. |
| | | |

| 831. | Rambusch, W. TAre the Leaves of the Primula obconica Poisonous? Am. Gar. 17:215. |
|-------|--|
| 832. | Ransom, W. B |
| 832a. | Ratti On Colchicum Poisoning. See Phar. Jour. and |
| 833. | Trans. III. 6:47. Ratzeberg, J. F. C See Brandt, J. F. |
| 834. | Rawton, Oliver deVegetales que curan y vegetales que natan. 348. Paris. 1888. |
| 835. | Read, Alex On the Use of Datura Stramonium in Certain Cases. 1816. |
| 836. | Reed, H. S |
| 836a. | 1908 The Fungus, Diplodia, as a Possible Factor in the Aetiology of Pellagra. Repr. N. Y. Med. |
| £37. | Jour. Jan. 22. 1910:18. Reese, J. J |
| 838. | icology. Ed. 2. Phila. 1889 The Malignant Effect of certain Trees upon |
| Coo. | Surrounding Plants Plant World, 10:279. |
| 839. | Renner Native Poisons. West Africa. Jour. Afr. |
| 840. | Soc. 1904:109-111. Ribbert, M. H. W Ueber den Untergang pathogen Schimmel- |
| | pilzener organismus. 59. Vers. D. Naturf. |
| 841. | Rice, W. S Suspicious Character of the Woods. Outing. |
| 842. | 40 :551-554. Equisetum Poisoning. Am. Vet. Rev. 26 :944- |
| 843. | 951. Rich, F. A Poisonous Equisetum. Proc. Am. Vet. Assoc. 1902:178. |
| 843a. | Rich, F. A. and Jones, L. R. A. Poisonous Plant: the Common Horsetail. |
| 844. | Bull. Vt. Agr. Exp. Sta. 95:187-192. Richardson |
| 845 | Am. Vet. Rev. 33:566-571. Pichet, C Les poisons convulsivants. Arch. internat. de |
| | pharmacod. 4:293-309. Paris. 1897-1898. |
| 846. | Ricord-Madianna, J. BRecherches et expériences sur les poisons d' Amérique, tirés des trois règnes de la na- ture, etc. (Du Brinvilliers, Spigelia anthel- |
| | mintica, L. II Du Mancenillier, Hippomane |
| 847. | Mancinella, L.) 169. Bordeaux. 1826. Riecke, V. A See Berge, F. E. |
| 848. | Robin, C Histoire naturelle des vegetaux parasites qui |
| | croissent sur l'homme et sur les animaux |
| 849. | vivants. Paris. 1853. Robinson, B. LSee Gray's Manual. |
| 850. | Robbins, E. L See Michael, L. G. |
| 851. | Rochebrune, A. T. de Toxicologie africaine. Etude botanique, his- |
| | torique, ethnographique, chimique, physiolo- gique, therapeutique, pharmacologique, po- |
| | sologique, etc., sur les végétaux toxiques et suspects propres au continent africain et |
| | aux iles adjacentes. Avec figures dans le texte par Charles Ricard. Bull. de la Soc. |
| | de Paris. Fas. 1-5. 768. 1896-1898. (Not |
| 852. | completed.) Rochussen, F |
| | 26 :167-176. |

| 853. | Rodigras, EmFleurs vénéneuses. Bull. d'arboricult. et de floricult. potagere. 1889 :281. |
|---------------|--|
| 854. | Rodin, HLes plantes médicinales et usuelles, des |
| 855. | Champs Jardins Forets. Paris. 1882. Rolly |
| | Wochenschr. 1906 . No. 37. 1798. |
| 856. | Phar. Assoc. (abst.) 24 :129. |
| 857. | Rosenthal, D. AUeber Giftpflanzen aus der Klasse der Leguninosen. Schles. Gesellsch. 1864 :57. |
| 858. | Rossi, P De nonnullis plantis, quae pro venenatis ha- |
| | bentus, observationes et experimenta a Pe- tro Rossi Florentiae instituta. 66. Pisa. |
| 859. | 1762. Rotenbiller vonOn Gaultheria Poisoning. See Klinisch. Ther. |
| | Wochenschr. May. 1900. |
| 859a. | Rothe, L |
| 860. | Roub, J. F |
| 861. | Vet. Rev. 26 :437-439. Roux, GSee Linossier, G. |
| 862. | Rüdel, C Beiträge zur Kenntniss der Alkaloide von Ber- |
| | beris aquifolium und Berberis vulgaris. Arch. der Pharm. 229:631-649; 649-660. |
| 863. | Abs. in Bot. Centrbl. 2:294. Rusby, H. H The Poisonous Plants of the Vicinity of New |
| | York City, Repr. Lecture before the Alum- |
| | ni of the College of Pharmacy of the City of New York. 1895. 18. Also Med. and |
| 864. | Sur. Jour. 72:839-844 Poisonous Properties of the Mountain Laurel. |
| 865. | Drug. Cir. 46:27. Ruschhaupt, Fr. and L. F. Poisoning by a Hemlock Species. Rept. Wis. St. Bd. Health. 18:99. |
| 866. | Russell, W |
| | végétaux. Compte. Rend. Acad. Sci. Paris. |
| 867. | Sur la siège de quelques principes actifs des végétaux pendent le repos hibernal. Revue Gen, de Bot. 15:160. |
| 86 8. | Ryon, Angie MPoison Ivy and its Effects. Amer. Bot. 4:87- |
| 869. | Sabanajeff, N. and Berl- |
| | mann, MDie Giftpflanzen des Gouvernements Jaroslaw, unit Angabe ihres Schadens für die Haus- |
| 070 | thiere. Jaroslaw, 8-16, 1886. |
| 870. 870a. | Safford, W. E |
| 871. | Comet M D Deuberghee des plantes très vénérouses par |
| | l'essai sur les tetrads des Batriciens. Sep. |
| 872. | Saint Hilaire Influence de la temperature organique sur l' |
| | action de quelques substances toxiques. In: Richet, C. Physiol. Trav. du lab. 1:390-430. Paris. 1893. |
| 873. | Saunders, C. FPoisonous Effects of a California Shrub. (Nu- |
| 873a | ma Parryi). Plant World. 6: 245-246. Savoure, CamilleEtude sur les alcaloides d'origine microbienne. |
| 0,00 | 97. Rennes. 1896. |

| 874. | |
|---------------|--|
| 875. | Sayre, L. E further Study in the Alkaloids of Gelsemi- |
| 876. | um. Proc. Åm. Phar. Assoc. 26Loco Weed. Am. Vet. Rev. 11:555. Also see |
| 877. | Proc. Am. Phar. Assoc. 38:107-108. |
| 878. | Schaffner, J. H Poisonous and other Injurious Plants of |
| 0.0. | Ohio. Ohio Naturalist. 4 :16-19; 32-35; 69-73. |
| 879. 879a. | Schenck, H See Strasburger, E. Scherer On Hay Fever. See Jour. Mich. St. Med. |
| 880. | Schimper, A. F. W See Strasburger, E. |
| 881. | Schimpfky, RichardDeutschlands wichigste Giftgewächse in Wort und Bild, nebst einer Abhandlung über Pflanzengift. Für den Schulgebrauch und |
| | zum Selbstumterrichte. Gera-Unterhaus verlag von Er Engen Kohler, 1893. |
| 882. 883. | Schlagdenhauffen, Fr See Heckel, E. Schlagdenhauffen, Fr., and |
| | Rieb |
| 884. | Schlitzberger, S Die Gift-und Heilpflanzen. Eine Anleitung, die giftige und heilkraftigen wirkungen |
| | kennen zu lernen und Anweisung, wie man sie gegen allerlei Krankheiten Benutzen kan. |
| 885. | Leipsic. 1899. Schlotterbeck, J. O Does Argenn de Morphine? |
| 886. | Phar. Rev. 19:458-461. Schlotterbeck, J. O., and |
| 887. | Watkins, H. CContribution to the Chemistry of Stylophorum diphyllum. Phar. Rev. 19:453-455. Schlotterbeck, J. O., and |
| 007. | Zwaluwenberg, A. von Comparative Structure of the leaves of Datura stramonium, Atropa Belladonna, and Hyos- |
| | cyamus niger. Pharm. Arch. I. 1898. No. |
| 888. | Schmelz, J. D. EZur Kenntniss indischer Gifte. Internat. Arch. |
| 889. | f. Ethnog. 15:54. Leyden. 1902. Schmidt, Ernst Ueber die Alkaloide der Lupinosensamen. Archiv. der Pharmacie. 235:Heft. 3. Schmiedeberg, O Grund Arz. 167. 1887. |
| 890. 891. | On Digitalin. See Archiv. Exp. Path. 16: |
| 892. | 149 On Oleandrin. See Ber. Chem. Gesell. 16: |
| 893. | Schneidemuhl, Georg Specielle Pathologie u. Therapie der Haus- |
| 894. | thiere fur Thierarzte, Aerzte und Studic- rende. Berlin. 1908. 799. f. 53. Schneider, ANative and Introduced Poisonous Plants of |
| 895. | California. Pacific Phar. 1907. Literature on Medicinal Plants and Drug |
| 2, 2, | Plant Culture. Phar. Rev. 26 :201-214; 236-245. |
| 896. | Schneider, MaxUeber Saponine. Zeitsch. des allgen. österreischlichen Apotheker-Vereins. 43:893-898; |
| 897. | Schniegans, MDer Giftstoff von Arum maculatum. J. d. Oh. f. Illsaes-Lohr. 62. |
| | |

| 898. | Schomburgh, R. H The Urari or Arrow Poison of the Indians of |
|--------------|---|
| 889. | Guiana. Phar. Jour. 16:500-507. (1857). Schorlemmer, C |
| | Compounds or Organic Chemistry. London. |
| 8802 | SchramekSee Kren. |
| 900. | Schribaux, E The Poisoning of Cattle by Eating the Green |
| | Plants of Lathyrus Clymenum. Jour. Agr. |
| 901. | Prat. 59 : No. 28. 32-55. Schuetz, K. E De Taxo baccata ejusque veneno. 41. Vrat- |
| 902. | islavia. 1840. Schulze, J. E. FToxicologia veterum plantas venenatas exhi- |
| 902. | bens Theophrasti, Galeni Dioscoridis, Plinii, |
| | alorium que auctoritate ad deleteria venena |
| | relatas. Loca ex veterum monimentis eruta |
| 903. | imenta et observata adjecit. 78. Halle, 1788. |
| 905. | Schummel, T. EUeber die giftige Pilze, mit besonderer Rücksicht auf Schlesien. 28. Breslau. 1840. |
| 904. | Schünemann, H Die Pflanzen-Vergiftungen. Ihre Erschei- |
| | nungen und das Vorzunehmende Heilver- fahren geschildert an den Deutschland heim- |
| 905. | schen Giftpflanzen, 88. Braunschweig, 1891. |
| 203. | · Alcaloide uit de Knollen van Dioscorea hir- |
| | suta Bl. Nederlandsch Tijdschrift voor |
| | Pharmacie, Chemie en Toxicologie. 9:Mai. 1897. |
| 906. | Schutz, Josef |
| | dadurch bedingten Erkrankungen der Augen, etc. Mitth. a. d. Kais. Gesundh. ante 1894. |
| 907. | 208. Schwalbe, K On Rhus Toxicodendron. See Münch. med. |
| | Woch 49:1 616: Med Rec 63:855 |
| 908. | Schwarz, A Giftpflanzen, Heilpflanzen, und Nährpflanzen. Bilder aus der heimschen Pflanzenwelt mit |
| | erlauternden Text, 277, Fürth, 1899. |
| 909. 910. | Scott, D. H., TranslatorSee De Bary, A. Scott, J. LGolden-rod Killing Horses. Gar. and For. 8: |
| | 477-478. |
| 911. | Segura, Jose CEl Maguey. Memoria sobre el cultivo y beneficio de sus productos. 407. Mexico. 1901. |
| 913. | Selby, A. D A First Ohio Weed Manual. Bull. Ohio Agr. |
| 914. | Exp. Sta. 83:249-500 A Second Ohio Weed Manual. Bull. Ohio |
| 015 | Agr. Exp. Sta. 175:291-383 |
| 915. | 1896 :119-128 |
| 916. | Seligman, C. GOn the Kenyah Dart Poison and its Active |
| | Principle, Antiarin. See Brit. Med. Jour. 1903:1131. |
| 917. | On the Physiological Action of the Kenyah |
| | Dart Poison "Ipoh" and its Active Principle, Antiarin. Jour. Phys. 1903 :39-58. |
| 918. | London. |
| 919. | Selmi, F |
| 920. | 30. 1842. Phar. Jour. 2:429-442 Lecture on Vegetable Poisons. Delivered at |
| | the Establishment of the Pharmaceutical So- |
| 920a | ciety, June 22, 1842. Phar. Jour. 2:17-31. SeymourOn Gelsemium. See Boston Med. and Sur. |
| >=\u0. | Jour. 105:590. |

| 921. | Sharp, G |
|--------------|---|
| 922. | Jour. 82:28-30; 117-118; 184-186. Shenstone, J. C |
| 923. | Shionoyana, J |
| 924. | Other Plants. Bot. Mag. 7:441. Shreve, F., and OthersThe Plant Life of Maryland. Maryland |
| 925. | Weather Service 3:533. Siegel, A |
| 925a. | aug. Dissert. Dorpat. 55. 1895. Simon, J. F See Sobernheim. |
| | Simpson, WilliamThe Names of Medical Plants of Commercial Value that are Gathered in North Carolina: their Value and the Relative Amount Sold in this Country and Exported. Bull. Phar. 8:448-450. |
| 926a. | SinklerOn Gelsemium. Sec Phil. Mcd. Times. 8: 150. 1877-1878. |
| 927. | ho. Bull. Id. Agr. Exp. Sta. 37:159-590. |
| 928. 929. | |
| 930. | Smith, E. F. and Hedges, FlorenceDiplodia Disease of Maize. (A Suspected |
| 931. | Cause of Pellagra). Science 30:60-61. Smith, FrederickManual of Veterinary Hygiene. Ed. 5. 1035. |
| | New York. Smith, J. BAntidotes to Alkaloidal Poisons. Med. Times |
| 932. | and Hosp Gaz London 5:22-24 |
| 933. | Smith, S. Bernhard Poisonous Plants of all Countries. 88. Bristol. 1905. |
| 934. | Smith, W. G |
| 935. | Smith, W. G. as Translator Sce Tubeuf. |
| 936. 937. | Smithsonian Institution See various Publications, especially No. 258. Sobernheim, J. F. and Si- |
| | mon, J. F |
| | Alkaloids; Bitter Principles; Glucosides. their Sources, Nature, and Char- acteristics, with Tabular Summary, Classifi- cation of Reactions, and full Botanical and General Indexes. 194. London. 1894. |
| | Soir and AlexandresenSur la toxicité d'un type d'Aspergillus fumigatus isolé du mais avarie. Compte. Rend. Soc. Biol. 64 :288-289. |
| 940. | Sommerfeld, T Entwurf einer Liste der gewerblichen Gifte ein Auftrage der internationalen Verein- |
| | igung für gesetzlichen Arbeitenschutz; unter Mitwirkung, von Thomas Oliver und Felix Putseys. Jena. 1908. |
| 941. | Spieler, A. J. T De plantis venenatis Silesiae. 42. Vratislavia. 184. |
| 942. | |
| 943. | |
| | burgh, and Dublin Pharmacopocia. 2 vols. London. 1829. |

| 944. | Sprouch, C. H. E Le poison diptherique considére principalement au point de vue de son action sur le rein. Compte. Rend. d l'Acad. des Sci. de Paris. 109:260-262. |
|--------------|--|
| 944. | Stalberg, S |
| 945. | Rept. Ia. Agr. Coll. Dept. of Bot. 1884: |
| 946. | 114-115. The Loco Plant and its Effect upon Animals. |
| 947. | Rept. Bur. An. Ind. U. S. Dept. Agr. 3:272. Stapf, O. The Aconites of India: a Monograph. Ann. Roy. Bot. Gar. Calcutta 10:115-179. 1905. |
| 948. 949. | Stevens, A. B |
| 950. 951. | Phar. Assoc. 1906:61-64 |
| | Phas Assoc 55.423-443 |
| 952. 953. | Also See Am. Phar. Jour. 79:518. Stewart, F. E |
| 954. | Stockberger, W. W The Drug Known as Pink-root. (Spigelia marilandica). Bull. Bur. Pl. Ind. U. S. Dept. Agr. 100:41-44. |
| 955. | Stokes, J. SOn Poisoning by the Internal Administration of Tineture of Rhus Toxicodendron. See |
| 956. | Med. Sur. Rept. 17:373. Stowe, W |
| 954. | Strasburger, E., Noll, F., Schenck, H., and Karsten, G |
| | tion. Revised by W. H. Lang. 746. Lon- |
| 958. | don. 1908. Strattuard, R |
| 959. | tine. Abs. in Am. Vet. Rev. 34:58. Strickland, C. W |
| 960. | Strong, A. B The American Flora. 2 vols. New York. 1848. |
| 961. | Sturdevant, L. B. and Peters, A. T |
| 962. | Sutton, R. L The Emergency Treatment in Cases of Acute |
| 963. | Swain, R. E. and Harkins, W. DArsenic in Vegetation Exposed to Smelt or |
| 0 | Smoke, Iour, Am. Chem. Soc. 30:915 |
| 964. 965. | Syme, R. L |
| 966. | Hopkins University. 1906Also see Acree, S. F. |

| 967. | Taylor, A. S | On Poisons in Relation to Medical Juris- |
|--------------|-----------------|---|
| | • • | prudence and Medicine. Third Am. and Eng. Ed. Philadelphia. 1875. |
| 968. | | Principles and Practice of Medical Juris- |
| 969. | | prudence. 3 vols. London. 1873. On Rhus Toxicodendron. See Jour. Cut. and |
| | | Ven. 'Dis. 1 :434. |
| 970. 971. | Taylor, H. H | Edible and Poisonous Mushrooms. |
| 972. | Ten Bosch, L | Does Poison Ivy Discriminate? Gar. and |
| | | For. 9:239. Essai sur les poisons végétaux la plus com- |
| 973. | | mun en France et leurs antidotes. 19. Mont- |
| 974. | Thacher, J | .On Loss of Hair and Nails from Rhus Poi- |
| 975. | Thin, G | soning. See Am. New Disp. 334. 1817. Experimental Researches concerning Tricho- |
| | | phyton tonsurans. Brit. Med. Jour. 1469: |
| 976. | Thomas, R. H | .A Contribution to the Causes of the So-called Hay Fever, Nasal Asthma, and Allied Af- |
| | | fections: Considered from a Chemical |
| | | Standpoint. Med. Rec. 1887:305-306. |
| 978. | | . Poison Romance and Poison Mysteries. London. 1899. |
| 979. | Thompson, Hedge | .An Experimental Dissertation on the Spigelia |
| | | Marilandica or India Pink. Philadelphia, 1802. |
| 980. | Thomson, Thomas | Chemistry of Organic Bodies. 183. London. |
| 981. | Thunberg, C. P | . Arbor toxicaria Macassariensis. Upsala. 1788. |
| 982. 983. | Thurber Coo | De plantis venenatis. 15. Upsala. 1788. Poisoning by Cypripedium. Bull. Torr. Bot. |
| 900. | | Club. 6:1 5. |
| 984. | Tidy, C. H | On Poisoning by Nitrate of Baryta. Med. Pres and Cir. 6:448. |
| 985. | Tillic, T. B | See Frazer, T. B. |
| 986. | Tirclli, V | . Auto intossicazioni secondarie all'azione dei |
| | | caustici e del calore ricerche tossicologiche. Ann. di freniat—Turin. 8:253-273. |
| 987. | Tizzioni, Guido | Neue bakteriologische Untersuchungen über |
| | | die Pellagra. Centrbl. f. Bakt. u. Parasit. |
| 987a | . Tokishige, H | . Ucber pathogene Blastomyceten. Centbl. f. |
| | | Bakt. Abt. I. 19 :105-113. 1896. |
| 988. | Torrey, John | Natural History of the State of New York. |
| 989. | Tovcy, J. R | Pt. 2. Botany. 2 vols. 1843. |
| 990. | Tracy, S. M | . Cassava. Far. Bull. U. S. Dept. Agr. 167:31. |
| 991. | Trelease, Wm | The Working of the Madison Lakes. Wis. |
| 001. | | Acad. Sci. Arts and Letters. 7:121-129. An Unusual Phyto-bezoar. Trans. St. Louis |
| | | Acad Sci 7:493-497, 1 bl |
| 992. | Treub, M | Nouvelles recherches sur le role de l'acide |
| | | cyanhydrique dans les plantes vertes. Ann. Jardin bot. Buitzenborg. Ser. II. 6:79-106. |
| 993. | Trimble | See Bastin, E. S. |
| 993a | . Trimble, W. K | A Case of Sporotrichal Infection. Jour. Kan. |
| 004 | 77-i II | Med. Soc. 9:305. |
| 994. 995. | Trinen, H | See Israel O. |
| 995. | Tryon, H. | Plants Poisonous to Stock. Brisbane Courier. |
| ,,,,,, | | May 24, 1887. |

| 997. | Tschirch, A. and Oesterle. Anatomischer Atlas der Pharmakognosie und Nahrungsmittelkunde. 334. Leipsic. 1900. |
|-----------------|---|
| 998. | Tschirch, A. and Stevens, |
| | A. BOn Japanese Lac, the milk sap of Rhus vernicifera. Arch. Phar. 243:504. Also Am. Phar. Jour. 78:53. |
| 999. | Tubeuf, Carl, and Smith, W. G |
| 1000. | Tuczek, FBehandlung der Vergiftungen mit Pflanzen- stoffen (excl. der chron. Morphinismus und Cocäinismus des Ergotismus, der Pellagra, und der Lathyrismus). Handb. d. Spec. Therap. innerer Krankh., 2:234-334. Jena. 1894. |
| 1001. 1002. | Umney, C |
| | (Abs. in Phar. Rev. 25 :323-324). Valentin, G. CEin Fall von Soor des Mittelohres. Arch. f. Ohrenheilkunde. 26 . |
| 1004. | Van Es, L. and Waldron, L. R |
| | ta. Bull. N. D. Agr. Exp. Sta. 58:321-352. Vasey, George |
| 1006. | Vater, A |
| 1007. | Vaughan, V. C Some New Bacterial Poisons, their Causal Relation to Disease and the Changes in our Theories Suggested by their Action. Med. News. 2:158-162. |
| 1007a. 1008. | See Hamilton A M |
| 1009. | Vaughan, V. C. and Novy, F. G. Ptomaines and Leucomaines. Vaughan, V. C., Novy, F. G., and McClintock, C. T.The Germicidal Properties of Nucleins. Med. |
| | de Vaulabelle, ALes plantes qui tuent. Jour. d. Hyg. 25:153: |
| 1011. | 169; 185. Paris, 1900. Verderau, L Le toxine du Bacillus virgula. Compt. Rend. |
| 1012. | Soc Biol Paris 1908:803-835 |
| 1013. | Verdier See Bretin. Verschaffelt, E. Bepaling der werking van vergiften op planten. Versl. d—wis-en-natuurk. Afd. d. k. Akad. v. Welensch. Amst. 12:ged. 855-859. 1903-1904. |
| 1014. | les bourgeons de Prunus en voie d'ecclosien. Arch. Neerland Sc. exact. nat. Ser. 27. 497-509. 1902 . |
| 1015. | Vicat, P. R Histoire des plantes vénéneuses de la Suisse, contenant leur description, leurs mauvais effets sur les hommes et sur les animaux, avec leurs antidotes, rédigee, d'apres ce qu'on à de mieux sur sette matière, et surtout d'après l'Histoire des plantes helvétiques de M. le baron de Haller, mise à la portée de tout le monde avec le lieu natal de chaque plante pour la France, les figures nécessaires, et plusiers observations nouvelles. 29+392; 112+8. Versailles. 1776. |
| 1016. | Villiers, ARecherches des poisons végétaux et animaux. 132. Paris. 1882. |

| 1017 | Volkens, G |
|----------------|---|
| 1017. | Königl, bot. Gar. u. Mus. 24 Berlin-Dahlem. 5:42-64. Ap. 22. |
| 1018. | Wainwright, J. WSecret Poisons and their Use in Ancient |
| 1019. | Waldron, C. APoisoning from Locust Bark. Am. Vet. Rev. 3:456-459. |
| 1020. | Waldron, L. RSee Van Es. L. |
| 1021. | Waleff, S Poisoning from Oleander. Deut. med. Wochenschr. 17:801. Abs. Am. Jour. Med. Sci. |
| 1021- | Wallace's FarmerClipping on Killing Horses by Oak-leaf diet. |
| 1021a. | (Based on Rept. For. Ser. U. S. Dept. Agr.) |
| 1021b. | Walsh, L. H South African Poisonous Plants. Being Notes |
| | on South African Plants Poisonous to Stock, with Particulars of Symptoms and Treat- |
| 1022. | ment. 56. Cape Town. 1909. Warburton, C. WThe Non-saccharine Sorghums. Far. Bull. |
| 1023. | U. S. Dept. Agr. 288:28. Warder, H See Pedler, A. |
| 1024. | Warder, J. A The Western Catalpa: a Memoir of the Shav- |
| 1025 | anon Tour Am Agr Assoc 1881 |
| 1025. | Warren, L. E |
| | Phar. Rev. 44:149-150: 218-219: 288-290. |
| 1026. | Phar. Rev. 44 :149-150; 218-219; 288-290. The Poisonous Species of Rhus. Phar. Jour. |
| 1027. | Oct. and Nov. 1909. Repr |
| 1020 | Repr. Am. Jour. Phar. 1910 :499-506. |
| 1028. 1029. | See Stevens, A. B. Watkins, W. HCase of Poisoning by the Essential Oils of |
| 102). | Savine, Pennyroyal, and Tansy. New Or- |
| | leans Med. and Sur. Jour. 5:230. |
| | Watson, SerenoBibliographic Index to North American Botany. Polypetalae. Washington. 1878. |
| 1030a. | Botany of California Geol. Sur. of Cal. 2:559. |
| | |
| 1031. | WayOn Cocain. See Med. News. Phil. 1:486. |
| | 1887. |
| 1032. | Weatherby, C. A See Graves, C. B. Webb, Paul C Plants Injurious to Animals. Alumni Report |
| 10024. | Associated Distriction of District April 27 |
| 1033. | Wellman, F. C The African Poison Test as Observed in the |
| 1034. | Portugeese Colony of Angola. 1906. Weschke, CarlThe Enumeration of the Medicinal Plants of |
| 1057. | Minnesota, Phar Rund 8:155-157 |
| 1035. | Westbrook, F. FContributions à l'étude des toxines du cholera. Ann. de l'institut Pasteur. 1894:318-337. |
| 1036. | Westgate, J. MAlfalfa. Far. Bull. U. S. Dept. Agr. 339:48. (Alfalfa Bloat p. 37). |
| 1037. | Wettstein, R. von |
| 1038. | Whelpley, H. MRhus Toxicodendron. Bull. of Phar. 8:552- |
| 1039. | Whipple, G. C |
| 1040. | York. Ed. 2. White, E. A A Preliminary Report on the Hymeniales of |
| | Connecticut. Bull, Geol. and Nat. Hist. Sur. of Conn. 3:80. |
| | |

| 1041. | ••••• | Second Report of the Hymeniales of Connecticut. Bull. Geol. and Nat. Hist. Sur. of |
|-------|----------------------------|---|
| 1042. | White, G. R | Conn. 15:70. Phytolacca 20320 in Cattle. Am. Vet. |
| 1043. | White, J. C | Rev. 26 :339-340. Dermatitis venenata: an Account of the Ac- |
| | | tion of External Irritants on the Skin. 216. Boston. 1887. |
| 1044. | ••••• | Death from Rhus Poisoning. N. Y. Med. Jour. 17:225. |
| 1045. | Widal, F. and Nobecourt | Recherches sur l'action antitoxiques des cen- tres nerveux pour la strychnine et la mor- |
| | | phine. Bull. et mém. Soc. méd. d. hôp. de Paris. Ser. 3. 15 :182-184. |
| 1046. | Wigand, Albert | Lehrbuch der Pharmacognosie, Berlin, 1879. |
| 1047. | Wilcox, E. V | Larkspur Poisoning of Sheep, Bull, Mon. |
| | | Agr. Exp. Sta. 15:37-51. Abs. in Ann. Rept. Bur. Ind. U. S. Dept. Agr. 15:473-479. |
| 1048. | | Plant Poisoning of Stock Ann. Rept. Bur. |
| 1049. | | An. Ind. U. S. Dept. Agr. 17:91-121. |
| 1042. | | Poisonous Plants of Montana. Rept. Mon. Agr. Exp. Sta. 1899:51. |
| 1050. | | Poisonous Sumach once more. Gar. and For. 8:478. |
| 1051. | Wilcox, E. V | See Chesnut, V. K. |
| 1052. | Wilcox, R. W | On Nutmeg Poisoning. See Lancet. 1895: 50. Rev. in Am. Jour. Med. Sci. 110:90. |
| 1053. | Wilcox, R. W. and Jeliffe, | |
| | S. M | On Datura Poisoning. See Brit. Med. Jour. 1903:1137. (Article by Dr. J. D. Gim- |
| | | lette). Rev. in Am. Jour. Med. Sci. 126: |
| 1054. | | 348. On the Kenyah Dart Poison and its Active |
| 1054. | ••••• | Principle, Antiarin. (Article by Dr. C. G. |
| | | Principle, Antiarin. (Article by Dr. C. G. Seligmann in Brit. Med. Jour. 1903 :1131). Rev. in Am. Jour. Med. Sci. 126 :347. |
| 1055. | Wilcox, R. W. and Pellon, | Review of Article by John Germann on Ver- |
| | Н. Н | Review of Article by John Germann on Ver- onal Poison in Jour. Am. Med. Assoc. 1906: |
| | | (No. 26) 1999. Am. Jour. Med. Sci. 133: |
| 1056. | Wilcox T E | 795. On Recurring Attacks of Rhus Poisoning |
| 1030. | ΨΠΟΧ, Ι. Ε | without Re-exposure. See Gar. and For. |
| 1057. | Wildoman E da | 8:478. Poisons d'epreuve de l'Afrique occidentale. |
| 1037. | | Assoc. Iranc. Avanc. Sci. 1905:/30-/43. |
| 1058. | Wiley, H. W | Sweet Cassava, Jatropha Manihot. Bot Gaz. 14:71-76. |
| 1059. | Williams, A. J | Poisonous Plants in South Africa. Vet. Rec. |
| 1060. | Williams, S. W | London, 16 :421. Report on the Indigenous Medical Botany of |
| | | Massachusetts. Trans. Am. Med. Assoc. 1849. |
| 1061. | Williams, T. A | A Report on the Grasses and Forage Plants |
| | | and Forage Conditions of the Eastern Rocky Mountain Region. Bull. Div. Agros. |
| 104 | | U. S. Dept. Agr. 12:78. |
| 1061a | . Williams, W. L | Enzoötic Cerebrospinal Meningitis of Horses in Idaho. Rept. U. S. Dept. Agr. Bur. An. |
| | | Ind. 1897:179-187. |
| 1062. | Williams, W. T | The Etiology of Heaves. Am. Vet. Rev. 26: 955-957. |

| 1063. | | shew (Mesquite) Poisoning. Jour. Jamaica |
|--------|---------------------------------|---|
| 1064. | Willis, G. S. V | Agr. Soc. 1:319-321. 1897. Manual of Vegetable Materia Medica. 480. |
| 1065. | | London, 1889. Ed. 11. Poisoning from Foods. See Public |
| 1066. | | Health 1906:626. bservations on the Poisonous Vegetables |
| 1000. | | which are either Indigenous in Great Britain or Cultivated for Ornament. 103. Lon- |
| 1067. | Wilson E W | don. 1781. eander Poisoning of Live Stock. Bull. |
| 1000 | Wil I N | don. 1781. leander Poisoning of Live Stock. Bull. Ariz. Agr. Exp. Sta. 59:381-397. 6f. www. Remedy for Sheep Poisoning on the |
| 1068. | | Stock Ranges. Yearbook. U. S. Dept. Agr. 1902;22. |
| 1069. | Wilson, James and Coville, F. V | Cavitta E V |
| 1070. | | |
| | | peutics for the Use of Practitioners and Students, 1326. Philadelphia 1898 |
| 1071. | Wilson, PRI | nododendron Poisoning in a Goat. The |
| 1073. | Wingate, J. BAc | Veterinarian. 70 :639-640. ctinomycosis. Jour. Comp. Med. 17 :276- 281. |
| 1073. | Winkler, LSä | mmtliche Giftgewächse Deutschlands nat- |
| | | urgetrau dargestellt und allgemein fasslich beschreiben. Mit einer Vorrede von Fried- lich Schwalgrichen. 119. 96 col. pl. Ber- lin. 1831. |
| 1074. | Winslow, KenelmVe | eterinary Materia Medica and Therapeutics. 755. New York, 1902. Ed. 2. |
| 1075. | Winton, A. L., and Moeller, | |
| 1076. | J | icroscopy of Vegetable Foods. 701. 1906. |
| 1077. | To | oxicology. (Vol. 4. of next book named) Vegetable poisons. 600-834. |
| 1078. | Witthaus, R. A. and Beck- | |
| | er, T. CMe | edical Jurisprudence, Forensic Medicine and |
| 1079. | Wittstein, G. C | Toxicology. 4:892. 35f. New York, 1896. andwörterbuch der Pharmakognosie des Pflanzenreichs. 464. Breslau. 1882. 1st |
| 1080. | Tı | Hälfte). 1e Organic Constituents of Plants and Veg- |
| 1000. | | etable Substances and their Chemical Analysis. Melbourne. 1878. |
| 1082. | Wolffenstein, R. U. and | January 11-1-1-1-1 124 TM 2 1000 |
| 1082a. | Wood, AlphonsoCl | lanzenalkaloide. 134. Ed. 3. 1900. ass Book of Botany. 829. New York. 1867. |
| 1083. | Wood, H. C | ext-book of Therapeutics. Ed. 11. |
| 1084. | | edical Botany. Ed. 2. 4 vols. London. 1810. |
| 1085. | | otanical Products of the United States Pharmacopoeia. 1890. Proc. Ind. Acad. Sci. 1894:108-119. |
| 1086. | Wright, R. and Farr, E. H.Tl | ne Pharmacy of Conium maculatum. Chem. and Drug. 49. |
| 1087. | Wunchmann, F De | eutschlands Giftpflanzen. Berlin. 1833. |
| 1088. | | e Beeinflussung der Giftswirkung durch die Temperatur, sowie durch das Zusammen- |
| | | greifen von zwei Giften. Zeitsch. f. allgem. |
| | | Physiol. 8 :140-190. Abs. in Bot. Centrbl. 108 :328-329. |
| | | |

| 1089. | Zoethout, W. DOn some Analogies between the Physiological Effects of High Temperature, Lack of Oxy- |
|-------|---|
| | gen, and Certain Poisons. Am. Jour. Phy- |
| | siol. 2:220-242. 1899. Abs. in Science 8: |
| | 776-778. (n. s.). |
| 1090. | Zoon, J |
| | Abbildungen von Arzny gewächsen. Nurem- |
| | berg. 1784. |
| 1091. | Zopf, Wilhelm Die Pilze in morphologischer, physiologischer, |
| | biologischer, und systematischer Beziehung. |
| | 500. Breslau. 1890. |
| 1092. | Die Pilzthiere. 1890. |
| 1093. | Zwaluwenberg, A. vanSee Schlotterbeck, J. O. |
| 1094. | Glenk, Robert Cicuta maculata, Linne. Am. Jour. Phar. |
| | 63 :328-332, |
| 1095. | Maish, J. M Notes on some North American plants. Am. |
| | Iour. Phar. 63:321-328. |
| 1096. | Mearns, E. A Two new species of poisonous sumachs from |
| | the states of Rhode Island and Florida. |
| | Proc. Biol. Soc. Washington, 15:147-149. |
| 1097. | Nelson, A., and Others Woody Aster. Bull. Wyo. Agr. Exp. Sta. |
| | 88:20 |

| Abbott, A. C., 869 | Acer macrophyllum, 615 | Meconic, 144 |
|--|--|---|
| Abbreviations | Negundo, 615 | Mineral, 6 |
| For poisonous qualities, | nigrum, 615 | Myristic, 389, 593 |
| 803-804 | rubrum, 615 | Myronic, 7 |
| For literature, 804 | sacchorinum, 615 | Nitric, 78 |
| For literature, 804 Abel, J. J., 35, 242 | soccharum, 615 | Ophelic, 689 |
| Abelia unifloro, 812 | .\CERACEAE, 607, 615 | Oxalic, 6, 72, 150 |
| Abies bolsamea, 327 | Aceros, 90 | Oxy- 144 |
| concolor, 327 | Acetanilid, 147 | Palmitic, 593 Phytolaccic, 435 |
| Abortion 78 100 101 123 | Acetoxin, 666 Acetyl, 147 | Pierie, 72 |
| Abortifacient, 803 Abortion, 78, 100, 101, 123 Abortives, Poisoning by, 76 | Achillea, 757, 758, 785-787 | l'oisonous, 72 |
| Symptoms, 76 | Millefolium, 140, 149, 756. | Polygalic, 62, 536 |
| Treatment, 76 | f. 786, 786-787 | Polygonic, 107 |
| Abrin, 11, 55, 72, 534, 536, | Millefolium, 140, 149, 756, f. 786, 786-787 moschata, 787 | Pumic, 679 |
| Symptoms, 76 Treatment, 76 Abrin, 11, 55, 72, 534, 536, 595 | Ptarmica, 756, 786, 787, | Pyrogallic, 608 |
| Abroma angusto, 622 | 814 | Quillaic, 7, 62, 505, 506 |
| Abrotanin, 755, 791 | Achillein, 140, 149, 756, 786, | Quinic, 144 |
| Abrotin, 140 Abrus, 52, 55, 149, 862, 863 precotorius, 1, 527, 531, 534, 595, 828 | 787 | Rheotannic, 419 Ricinoleic, 594 Salicylic, 505 |
| hrecotorius 1 527 531 | Achlya prolifera, 209 | Salicylic 505 |
| 534 505 828 | Achorion, 12, 13, 863 okromegalicum, 296 | Sphacelenic, 278, 279 |
| pulchellus, 828 | cysticum, 296 | Stearic, 593 |
| Abscesses, in rabbits, 179 | demergens, 296 | Sulphuric, 6 |
| Metastatic, 170 | didikroon 296 | Tannic, 144, 403, 503, 608 |
| Absinth, 793 | didikroon, 296 entythrix, 296 | Tartario 520 |
| Absinthe, 755, 756, 791, 793 | moniliforme, 296 | Tiglinic, 593, 789 |
| Absinthe, 755, 756, 791, 793 "Absinthe epilepsy," 793 | radians, 296 | Tropic, 716 |
| Absinthiin, 140, 793 Absinthin, 140, 755 | Schoenleinii, 17, 294, 295 | Tiglinic, 593, 789 Tropic, 716 Uric, 146 Acne, 78 |
| Absinthin, 140, 755 | tarsiferon, 296 | Acne, 78 |
| Absintlinum, 1. 141, 756, 793 | Achras, 89 | Achistiis, 89 |
| Absinthium, *f. 141, 756, 793 Absinthol, 756 Abutilon, 624, 626-627 indicus, 623 Theobreati 124, 627 | Sapota, 128, 679, 681, 851 | orborescens, 853 |
| indicus 622 | Achyranthes bidentato, 805 | Acokonthero Lamorkii, 806 |
| Theophrosti, 124, 627 | Achryospermum, 828 | venenoto, 806 Aconin, 109, 450 |
| Acacia, 89, 523, 533 | Acids, 6, 72, 144 | Aconine group of 450 |
| Acacia, 89, 523, 533 arabico, 529, 828 Catechn, 529, 533, 828 | Aconitic, 144 | Aconite, 2, 4, 72, 75, 85, 87, |
| Catechn, 529, 533, 828 | Amido fatty, 73 Anemonic, 111, 454 Angelic, 573, 648, 789 | 109, 381, 446, 449-453, |
| concinua, 828 | Angelic, 573, 648, 789 | 863 |
| Cunuinghamii, 828 | Arsenious, 2 | Columbia, 46 |
| delibrata, 828 | Benzoic, 144, 683 | European, 46, 109, 450, |
| falcota, 828 | Carbolic, 6, 77, 78, 79 | 452, 453 |
| False, 559, 560. f. 559 | Arsenious, 2 Benzoic, 144, 683 Carbolic, 6, 77, 78, 79 Carbonic, 77 | Group of alkaloids, 148 |
| Fornesiana, 523 Georginae, 828 Gerrordi, 523 | Catechutannic, 530 Cathartic, 418, 526 | Indian, 449, 450 |
| Gerrordi 523 | Cathartic, 418, 526 | Poisoning by. 6, 75, 450 |
| Juremo, 828 | Chrysophanic, 418 | Seed, 83, 450 Statutes regarding, 6 |
| pallens, 534 | Cinnamic, 683 | Toxicity of 109 450 |
| pennata, 828 | Citric, 144, 505, 512, 529, 572, 583 Copaibic, 529 | Toxicity of, 109, 450 Treatment for poisoning |
| penninervis, 828 | Consibir, 529 | by, 79 |
| pruinescens, 828 | Crassulacic, 501 | Western, 450, 452-453, /. |
| pulchello, 828 | Crotinic, 588 | |
| salicino, 828 | Crotinic, 588 Cygnic, 533 | Aconitic Acid, 144 |
| Senegal, 529 | Embelic, 679 | Aconitin, 73, 77, 78, 109, 144, |
| tencrima, 530 | Ergotinic, 7, 28, 277 | Aconitin, 73, 77, 78, 109, 144, 145, 148, 450, 452 |
| vernicistua, 828 verticistata, 828 | Eriodictyonic, 703 | Pseudo, 450 |
| Acalypha, 590, 603, 604 | Fatty, 73 | Aconitism, 44, 45, 46 Aconitum, 46, 448, 449-453, |
| colchica, 821 | Filicic, 77, 319 Formic, 106, 412 | 863 |
| cordifolia, 821 | Gallotannic, 533 | altigaleatum, 4 |
| gracilens, 604 | Gastrolobic, 533 | Anthoro, 4, 844 |
| indico, 821 | Gentisic, 689 | commarum, 4 |
| Ostryaefolia, 604 | Helvellic, 78, 238, 253 | chosmanthum, 4, 450 |
| virginico, 122, 604, f. 605, | Hydrocyanic, see Hydro- | chinense, 844 |
| 821 | cyanic Acid | columbianum, 46, 109, 450, |
| ACANTHACEAE, 90, 692, 698, | Inglandic, 401 | 1. 452, 844 |
| Acarthophyllum 20 412 | Lauric, 593 | deinorrhysum, 450 |
| Acanthophyllnin, 89, 813 | 1,00enc. 734 | ferox, 449, 450, 844 |
| Aconthosicyos horrido, 750 | Loco, 565 | Fischeri, 449, 844 heterophyllum, 148, 449, |
| The reference is to figure | Lupulic, 106 | heterophyllum, 148, 449, 450, 844 |
| on page 141. | Maizenic, 342 Malic, 144, 501, 505, 512 | japonicum, 844 |
| | ,,,,, | ,,, o |
| | | |

Afzelia, 1, 534 Agallocha, 588 Aganosma Salycina, 806 Agaric, Deadly, 238-239, 240 Fly, 10, 31, 74, 99, 236-238, f. 236 Lycoctonum, 4, 449, 844
Napellus, 46, 75, 87, 109,
145, 148, 446, 449, f.
451, 453, 844 Alexipharmic, 863 Alfalfa, 118, 525, 552-553, f. 554, 863 Dodder, 130, 701 Yellow-flowered, 526 109, noveboracense, Alfilaria, 578, f. 579 Algae, 90-95, 98, 184-195, 859, 863 844 AGARICACEAE, 99, f. 235, 859-860 Agaricus, 863 99. palnatum, 450
reclinatum, 109, 449
spicatum, 450
nucinatum, f. 45, 109, 449,
f. 452, 453, 844
variabile, 4
Acorin, 371
Acorn, 105, 403, 863
Acorns Calanus, 371
Acquetta di Napoli, 2
ACRASIEAE, 160
Acrec, S. F., 869
Acrid poison, 803
Acrocomia, 370 palmatum, 450 235-245. Blue-green, 91, 97, 160, 185, 187 Agariens, 863
arvensis, 235
bulbosus, 78
cumpestris, f. 233, 235
oreades, 503
Agave, 63, 89
americana, 105, 386
heteracantha, 386
rivida 386 Brown, 154, 194 Green, 153, 160 Green, 153, 160 In dynamite, 188 In water supplies, 91-94 Prevention of growth of, 94 Red, 154, 194-195 Violet, 154, 194-195 Algarrobe, 523 Algicides, Use of, 94 Alhogi maurorum, 527 Alisma Plantago, 335, f. 335, 804 rigida, 386 Schotli, 386 virginica, f. 384 Agave saponin, 386 Age of plant affecting activity Acrid poison, 803 Acrocomia, 370 Acronychia laurifolia, 849 Actaea, 447, 467-468 alba, 108, 844 rubra, 108, 467, 844 spicata, 844 Actinella, 757 Actinonycosis, 863 Age of plant affecting action of poison, 83
Agelaea emetica, 817
Algeratum, 90, 756
Conyzoides, 756
Aggio, C., 869
Agglutinin, Bacterial, 16:
Agrimouia eupatoria, 847
gryposepala, 505
Agropyron, 340
occidentale, 364
Ergot on, 276 804 Plantago-aquatica, 102 ALISMACEAE, 102, 335, 804 Alizarin, 742 Actinomycosis, 863 Adamson, II. G., 869 Adausonia digitata, 621 Adder's Tongue, 313 Adenanthera paronina, 828 Adenin 146 Differs from alkaloid how? 143 Caustic, 72 Corrosive, 72 Salts, 144 Adenin, 146 Adenium Bochmianum, 806 Ergot on, 276 repeus, 103, 364-365, f. 365 Ergot on, 276 Alkaloids lkaloids
Aconite group, 148
Chemistry of, 142, 150
Classification of, 145-146
Composition of, 143, 144
Defined, 143
Gluco, 149
Lso-quinoliu group, 145
Liquid-volatile group, 147
Mydriatic group, 148
Occurrence of, 144-145
Opinm group, 147
Physiological action of, 146, obesum, 806 ovesum, 806
Somaleuse, 806
Adiantum, 90, 316, f. 316
capillus-Veneris, 316, 317
pedutum, 101, 316-317, f.
317, 824
peruvianum, 824 teuerum, 364 feuerum, 364
Agrostemma, 436, 439-441
Githaga, 62, 63, 107, 439441, f. 440, 813
sapotoxin, 62
Agrostemmin, 108
Agrostis alba, 826
Ergot on, 276
Agrosterium, stybandroides trapeziforme, 824 Adlunia, 485 Adlumia, 485
cirrhosa, 480
Adonidin, 148, 446
Adonis, 78
aestivalis, 446, 844
auntrensis, 446, 844
Adoyuis, 5antes de, 869
Adoxa, 741
moschatellina, 741
Adoxaceae, 741, 804
Accidium berberidis, 226
columnare, 222 Agrostocrinum stypaudroides, 833
Ailanthus, 583-584
Family, 583-584
glandulosa, 121, 583-584,
f. 584, 853
Air Potato, 374
Airyar, N. S., 869
Aizoaceak, 423
Aizoon canarieuse, 824
Akebia, 629
quinata, 629
Alabama, Flora of, 865
Alban, 681
Albert, 869
Albertoni, 869 833 147 Properties of, 143, 144 Purin group, 146 Pyridin group, 145 Pyridin group, 145 Quinolin group, 145 Solid, 144 Solubility of, 143 Strychnin group, 147 Tetanus-producing group, Accidium berberiats, 2 columniare, 222 Rhauni, 229 Aegle Marmelos, 582 sepiaria, 582 Aegiceras, 52 majus, 838 Vegetable, 862, 863, 869 Abrotaniu, 755, 791 Abrotin, 140 Achillein, 756, 786, 787 Aconitin, 73, 77, 78, 109, 144, 145, 148, 450, Albertoni, 869 Alberts, 869 Albizzia, 1, 52, 89 anthelmintica, 829 minus, 838 144, 452 anthelminitica, 829
Lebbek, 829
Incida, 530
odoratissima, 829
procera, 829
stipulata, 829
stipulata, 829
AlbuginAceae, 204, 208
Albugin, 195
canida, f. 204, 206
Albumin, Poisonous, 73
Alcohol, 728
Methyl, 144
Poisoning by, 6, 73, 77, 78
Alcoholism, 77
Treatment, 78 Aegolethron, 64
Aeschynomene aspera, 524
Aesculetin, 617
Aesculin, 123, 617, 642, 683
Aesculins, 89, 616-617
californica, 123, 827
flava, 827
glabra, 123, 617, f. 619, 827
Hippocastannun, 63, 123,
617, f. 618, 827
octandra, 123
parviflora, 123
parviflora, 617
Pavia, 63, 123, 617, f. 619,
827 Aegolethrou, 64 Agrostemmin, 108 Alstonamin, 692 Amaryllin, 386 Anhalonidin, 637 Anhalonin, 637 Arccaidin, 370 Arccolin, 370 Arccolin, 78, 370 Argyraescin, Aricin, 742 123, 617 Aristolochinin, 41 Aspidosamin, 692 Aspidosamin, 692 Aspidospermatin, 692 Aspidospermin, 147, 692 Atisin, 148, 450 Atropamin, 715 Atropin, 3, 60, 61, 88, 145, 148, 149, 238, 584, 706, 715, 716, 726, 728, 730, 731, 732, 827 827
Aethusa, 650, 659, 863
Cynapium, 4, 76, 126, 659,
' /. 660, 856
Afghanistan, Poisonous plants
of, 867
Africa, Medicinal plants of, Treatment, 78 Alder, 404 Alder, 404
Aldrovanda, Leaf of, 499
Alectryon excelsum, 850
Aletris farinosa, 104, 827
Alcurites Fordii, 587
moluccana, 587
trileba, 587 Africa, M 866 Poisonous plants of, 866 African Marigold, 755-756 732 Alexandresen, 869 Baccharin, 755

| Baptitoxin, 530 Belladonnin, 386, 715 Benzoylecognin, 575 Berbamin, 472 |
|---|
| Berbamin, 472 Berberin, 88, 90, 108. 112, 146, 468, 470, 472, 473, 478, 483 Bikhaconitin, 450 Brucin, 145, 146, 147, |
| Brucin, 145, 146, 147, 686 Bryonicin, 751 Caffeidin, 628 |
| 614, 628, 742 Calabarin, 148, 528, 531 Calycanthin, 112, 476 Carpain, 627 |
| 686 Bryonicin, 751 Caffeidin, 628 Caffein, 144, 146, 606, 614, 628, 742 Calabarin, 148, 528, 531 Calycanthin, 112, 476 Carpain, 627 Cathin, 614 Celastriu, 614 Cephaelin, 742 Chelerythrin, 113, 480, 485 |
| 485 Chelidonin, 480, 485 Chenopodin, 429 Chrysthanthemin, 789 Cicutin, 656 Cinchonidin, 145 Cinchonin, 145, 742 Cinnamyleoccain, 575 Cocain, 6, 73, 77, 79, 144, 147, 575, 576, 614 |
| Cinchonidin, 145 Cinchonin, 145, 742 Cinnamylococain, 575 Cocain, 6, 73, 77, 79, |
| 614, 147, 373, 376, 614 Codamin, 147, 481 Codein, 59, 77, 112, 113, 146, 147, 479, 481 Colchicin, 77, 78, 79, 146, 148, 375, 479 Columbamin, 473 |
| Colchicin, 77, 78, 79, 146, 148, 375, 479 Columbamin, 473 Conicein, 651 |
| 146, 148, 375, 479 Columbamin, 473 Conicein, 651 Coniferin, 382 Coniin, 3, 73, 77, 79, 126, 135, 144, 147, 648, 651, 652, 653, 688, 726, 728, 747 |
| Cornutin, 28, 77, 78, 277, |
| Corydalin, 147, 480 Cryptopin, 481 Curarin, 73, 77, 687 Curin, 148, 687 Cusobygrin, 575 |
| 278, 279 Corybulbin, 147, 480 Corydalin, 147, 480 Cryptopin, 481 Curarin, 73, 77, 687 Curin, 148, 687 Cusohygrin, 575 Cusparin, 582 Cynapin, 126, 659 Cynoglossin, 706 Cytisin, 77, 78, 79, 90, 118, 148, 530, 537, 543, 551, Damascenin, 446 |
| 551, Damascenin, 446 Delphinin, 108, 446, 464 Delphinoidin, 108, 464 Delphisin, 108, 464 Dispinin, 108, 464 Dispinin, 692 Drumin, 588 Duhoisin, 716 Ecbolin, 149, 277 Echitamin, 692 Echitamin, 692 Ergotin, 277, 278 Ergotinin, 277 Ergotoxin, 278 |
| Disinchonin, 742 Ditamin, 692 Drumin, 588 Duhoisin, 716 |
| Ecbolin, 149, 277 Echitamin, 692 Echitemin, 692 Ergotin, 277, 278 Ergotinin, 277 |
| Erythropholoein, 48, 534 Eserin, 82, 531, 729 Eseridin, 528 |
| 147, 689 Gelseminin, 147, 689 |
| Gnoscopin, 481 Gratiolin, 734 |

| Guvacin, 370 |
|---|
| |
| Guvacin, 370 Heliotropin, 705 Hydrastin, 108, 146, 468 Hydrastium, 146 Hydrococotarnin, 481 Hydroquinon, 505, 742 Hygrin, 147, 575 Hyoscin, 148, 716, 726, 727, 730 Hyoscin-pseudocyanim, 148 |
| Hydrastin, 108, 146, 468 |
| Hydrastium, 146 |
| Hydrococotarnin, 481 |
| Hydroquinon, 505, 742 |
| 11ygrin, 147, 575 |
| Hyoscin, 148, 716, 726, |
| 727, 730 |
| Hyoscin-pseudocyanim, |
| 148 11yoscamin, 3, 60, 61, 77, 133, 148, 716, 726, 727, 730, 732, 756 Imperialin, 375 Inaein, 692 Indaconitin, 450 Isopilocarpin, 148 Isopyrin, 446 Jaborin, 148, 450 Jervin, 103, 148, 381 Juruhehin, 715 Kisanin, 386 Lanthopin, 481 Lappakonitin, 450 |
| 11yoscamin, 3, 60, 61, 77, 133, 148, 716, 726, |
| 77, 133, 148, 716, 726, |
| 727, 730, 732, 756 |
| Imperialin, 375 |
| Inacin, 692 |
| Indaconitin, 450 |
| Isopilocarpin, 148 |
| Isopyrin, 446 |
| Jaborin, 148, 582 |
| Japaconitin, 148, 450 |
| Jervin, 103, 148, 381 |
| Juruhehin, 715 |
| Kisanin, 386 |
| Lanthopin, 481 |
| Lappakonitin, 450 |
| Laudanin, 147, 481 |
| Laudanosin, 481 |
| Lobelin, 78, 752 |
| Lophophorin, 637 |
| Lupanin, 530 |
| Lycotin, 386 |
| Lanthopin, 481 Lappakonitin, 450 Laudanin, 147, 481 Laudanosin, 481 Lobelin, 78, 752 Lophophorin, 637 Lupanin, 530 Lycorin, 386 Manacin, 715 Mandragorin, 715 |
| Mandragorin, 715 |
| Matrin, 530 |
| Meconidin, 481 |
| Menispermin, 112, 149, |
| 473 |
| Mescalin, 637 Mcthylconiin, 126, 651 Morphin, 3, 59, 73, 78, 80, 82, 85, 90, 112, 113, 143, 144, 146, 147, 479, |
| Methylconiin, 126, 651 |
| Morphin, 3, 59, 73, 78 |
| 80, 82, 85, 90, 112, 113 |
| 143, 144, 146, 147, 479 |
| 489 |
| Moschatin, 140, 787 |
| Muscarin, 10, 32, 73, 74, |
| 77. 78. 80. 82. 148. 150 |
| 237. 238. 242 |
| Myriocarpin, 750 |
| Narcein, 146, 481 |
| 1.01.001111 7.101 |
| Narcotin 146 147 481 |
| Narcotin, 146, 147, 481 Neurin, 237 |
| Narcotin, 146, 147, 481 Neurin, 237 Nicotin, 7, 73 77, 78, |
| Narcotin, 146, 147, 481 Neurin, 237 Nicotin, 7, 73, 77, 78, 80, 85, 87, 133, 144. |
| Narcotin, 146, 147, 481 Neurin, 237 Nicotin, 7, 73, 77, 78, 80, 85, 87, 133, 144, 147, 551, 716, 728, 729 |
| Narcotin, 146, 147, 481 Neurin, 237 Nicotin, 7, 73, 77, 78, 80, 85, 87, 133, 144, 147, 551, 716, 728, 729 Nigellin, 446 |
| Narcotin, 146, 147, 481 Neurin, 237 Nicotin, 7, 73, 77, 78, 80, 85, 87, 133, 144, 147, 551, 716, 728, 729 Nigellin, 446 Oxyganthin, 112, 472 |
| Narcotin, 146, 147, 481 Neurin, 237 Nicotin, 7, 73, 77, 78, 80, 85, 87, 133, 144, 147, 551, 716, 728, 729 Nigellin, 446 Oxycanthin, 112, 472 Palmatin, 450 |
| Narcotin, 146, 147, 481 Neurin, 237 Nicotin, 7, 73, 77, 78, 80, 85, 87, 133, 144, 147, 551, 716, 728, 729 Nigellin, 446 Oxycanthin, 112, 472 Palmatin, 450 Papaveramin, 113, 147 |
| 489 Moschatin, 140, 787 Muscarin, 10, 32, 73, 74, 77, 78, 80, 82, 148, 150, 237, 238, 242 Myriocarpin, 750 Narcein, 146, 481 Narcotin, 146, 147, 481 Neurin, 237 Nicotin, 7, 73, 77, 78, 80, 85, 87, 133, 144, 147, 551, 716, 728, 729 Nigellin, 446 Oxycanthin, 112, 472 Palmatin, 450, Papaveramin, 113, 147 Papavarin, 59, 113, 146, 729 Nigellin, 446 |
| l'apavarin, 59, 113, 146, |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 522 |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 522 |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 522 |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 522 |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, 531 Pilocerein, 637 Piperin, 396 Piturie, 5 Piturin, 147, 716 Protopin, 113, 480, 481, 485 |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, 531 Pilocerein, 637 Piperin, 396 Piturie, 5 Piturin, 147, 716 Protocurin, 148, 687 Protopin, 113, 480, 481, 485 Protoveratridin, 381 |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, 531 Pilocerein, 637 Piperin, 396 Piturie, 5 Piturin, 147, 716 Protocurin, 148, 687 Protopin, 113, 480, 481, 485 Protoveratridin, 381 |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Pilocarpidin, 582 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, 531 Pilocerein, 637 Piperin, 396 Piturie, 5 Piturin, 147, 716 Protocurin, 148, 687 Protopin, 113, 480, 481, 485 Protoveratridin, 381 Pseudohyoscyanin, 148, 715, 726 |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Pilocarpidin, 582 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, 531 Pilocerein, 637 Piperin, 396 Piturie, 5 Piturin, 147, 716 Protocurin, 148, 687 Protopin, 113, 480, 481, 485 Protoveratridin, 381 Pseudohyoscyanin, 148, 715, 726 |
| Papavarin, 59, 113, 146, 147, 481 Pectenin, 637 Pellotin, 637 Pelosin, 413, 478 Pereirin, 147 Phytolaccin, 107, 435 Physostigmin, 146, 148, 582 Picolin, 728 Pilocarpidin, 582 Pilocarpidin, 582 Pilocarpin, 146, 148, 528, 531 Pilocerein, 637 Piperin, 396 Piturie, 5 Piturin, 147, 716 Protocurin, 148, 687 Protopin, 113, 480, 481, 485 Protoveratridin, 381 |

Pseudomorphin, 481 Pyrethrin, 754
Pyridin, 145, 146, 728
Quebrachin, 692
Quebrachinamin, 692 Quebracho, 147 Quercitrin, 506, 510, 584, 666 Quinamin, 742 Quinin, 77, 78, 87, 145, 147, 741, 742 Rhoeadin, 481 Rhoeadin, 481 Rubijervin, 381 Sambucin, 747 Sanguinarin, 113, 480, 484, 485 Sarracenin, 114, 497 Scopalamin, 60, 77, 78, 148, 716, 732 Secalin, 278 Septentrionalin, 450 Solanein, 718 Solanidin, 132, 148, 718 Solanidin, 132, 148, 718 Solanidin, 132, 148, 718, 721, 722, 724 Sophorin, 530, 543 144, 145, 149, 716, 721, 722, 724
Sophorin, 530, 543
Snartcin, 147, 551
Spigelin, 688
Staphisagrin, 108, 464
Strychnin, 1, 72, 73, 82, 129, 144, 145, 147, 628, 686, 688, 728, 729, 736
Stylopin, 480
Synaktonin, 450
Taxin, 101, 148, 328
Thalictrin, 446
Thebain, 146, 629, 742
Theobromin, 144, 146, 621, 628
Theo-phyllin, 146, 628
Trigonellin, 531, 648
Tropa-cocain, 575
Truxillin, 575
Trubo-curarin, 148
Illerin, 530 Tubo-curarin, 148 Ulexin, 530 Vicin, 120 Xanthopuccin, 108, 468 Nanthopuccin, 108, 468
Veratrum group, 148
Volatile, 144
Alkona tinetoria, 704
Alkanet, 704
Allonanda cathartico, 806
Alleghany Thermopsis, 538S39
Allen. Dr. 440 Allen, Dr., 440 Allen, H. M., 608 Allen, Timothy F., 869 Alligator Pear, 478 Allium, 104, 377, 383 ascalonicum, 375, 383-384, 833 Cepa, 383, 833 Douglosii, 833 Moleonii, 834 Moly, 383, 834 Porrum, 373, 383, 834 sotium, 375, 383, 834 Schoenoprasum, 375, 383, 834 833 834 tricoccum, 104, 383, 834 unifolium, 104 ursinum, 834 Victoriolis, 834 rineole, 834 Allspice, 639 Almacigo, 575 Almond, 505, 863 Bitter, 88 Oil, 315

Essence, 55 Pulp, 517 Seeds, 505 Flowering, 505, 514 Indian, 638 Oil, 608 Aluus glutinasa, 404 Alacosia indico, 808 montano, 808 Aloe, 375 obyssinico, 834 ofricana, 834 American, 386 orbarescens, 834 chinensis, 834 False, f. 384 ferax, 834 saponario. 834 spicato, 834 succotrino, 377, 834 tenuiar, 834 vero, 834 Aloin, 377 Aloin, 377
Aloinum, 375
Alpers, W. C., 869
Alpinia Colonga, 392
officinarum, 392
striato, 391
Alsberg, Dr. C. L., 343, 869
Alsidium
Helminthochartes, 859 Alsike Clover, 525, 553, 554 Alsine crassifalia, 436 Alstonamin, 692 Alstania, 691
canstricta, 806
Schularis, 806
spectabilis, 692
venenoto, 806
Alstonamin, 692 Alstonamin, 692 Altenoria brassicoe, 283 Althaea officinalis, 623 rosco, 623, 837 Altinarans, 128 Alum Root, 500 Alumbaugh, W. E., 869 Alyssum moritimum, 486 Sweet, 486 Alyxia, 90 buxifalia, 806 daphuaides, 807 Alway, F. J., 869 Amanito, 99, 235-243, 863 chlorinasmo, 36, 241, 242 cathurnato, 241, 859 flaccocephala, 241, 859 Deadly, 35, 36 Fly, 236-238, f. 236 Frastiano, 99, 238, 242, 859 mognivelaris, 859 moppa, 859 muscaria, 31, f. 22, 35, 74 daphnaides, 807 morpa, 859
muscaria, 31, f. 33, 35, 74, 99, 148, 236-238, 859
pantherina, 237
pholloides, 31, f. 32, 35-36, 79, 99, 237, 238-243, f. 239, 859
parphyrio, 241, 242
rodicata, 36, 241, 242
recutio, 859
rubesceus, 36, 99, 241, 242 rubesceus, 36, 99, 241, 242, 859 solitaria, 36, 99, 242, 859 spreto, 36, 99, 241, 242, 859 strobiliformis, 36, 241, 242, 859 veruo, 36, 99, 241, 859 viitadini, 242 viroso, 36, 99, 241, 242, 859 White, 35 Amanita-hemolysin, 35, 242 Amanitin, 10, 238

Amanita-toxin, 35, 36, 242, 243 Amaranth Family, 430-433 Spiny, 432, f. 433 AMARANTHACEAE, 423, 430-433, 805 Amaranthus, 431-433 blitoides, 431, f. 432 gangeticus, 805 gangericins, 431 hybridus, 432, 805 hypochoudriacus, 107, 805 retroflexus, 107, 432, f. 433 spinosus, 107, 432-433, 1 433 viridis, 805 Amaryllidaceaf, 63, 89, 104, 375, 386-388, 805 Amaryllin, 386 Amaryllis, 386 Amaryllis, 386 Belladouna, 386, 805 Family, 386 Ambrasio, 757, 764-767, f. 755 Ambrasio, 757, 754-767, 755, Ambrasio, 757, 764-767, f. 795 ortemisiae folio, 137, 755, f. 766, 766-767 trifido, 136, 137, 755, 765-766, f. 766 Ambrisiaceae, 748, 753 AMBROSIEAE, 757 Amelonchier canndensis, 504 spicota, 504 spicota, 504
vulgaris, 53
America, Economic plants of, 864 Medicinal plants of, 866 Medichiai plants of, 866-867
North, 866
South, 867
American, Aloe, 386
Beech, 403
Celandine, 480
Centaury, 689
Chestnut, f. 492, 403
Columbo, 689
Elder, 747
Elm, 405, 406-408
Hellebore, White, 85, 381
Holly, 614
Nelumbo, 108
Pawpaw, 111, f. 476, 476-477
Pennyroyal, 711
Savin, 102
Vetch, 570
Yew, 101, 328
Amianthemium muscaetaxicum, 104, 376 Poisonous plants of, 866-867 Amianthenum muscaetaxi cum, 104, 376 Amianthus, 16 Amianthus, 13, 143, 145, 149-150 Ammania boccifera, 638, 836 Ammi Visnogo, 856 Ammonia, 72, 145 Poisoning by, 6 Ammoniac, 650 Amanis caryophyllata, 639 Amonum moximum, 391 Amonium moximum, 391 Amonum moximum, 391
subulatum, 391
subulatum, 391
santhoides, 391
Amorphophallus, 371
viridis, 808
Amplelopsis quinaucfalia, 858
Amphicorpaea monaica, 204
Amsonia Tobernoemantona, 807 807 Amygdalin, 7, 54, 83, 116, 117, 144, 503, 504, 505, 506, 512, 517, 518, 519, 803

Amygdalus persica, 117 Amyris balsamifera, 817 Anabaeno, 91, 93, 94, 184 flos-aquae, 92, 98 Hossalii, 186 macraspermo, f. 185 stagnalis, 98

Anacaroiceae, 53, 122-12 607, 614, 805-806 Anacardium accidentale, 607 Anacyclus afficinorum, 814 122-123, Anacyclus afficinorum, 814
pedunculotus, 814
Pyrethrum, 754, 814
Anagalis, 89, 676-677
orvensis, 128, 676-677, f.
678, 844
Anagyrin, 530
Anogyris faetida, 90, 530, 551, 829 Analgen, 147 Anamirta, 52 poniculata, 1, 472, 838 Anonas sativus, 373, 811 Anostatica hierachuntica, 486 Anatomy, 863 Anatomy, 863
Anchietea solutaris, 858
Anchietea solutaris, 228, 404
Ancistracladus Vohlii, 820
Andenia labata, 841
Anderson, Dr. A. P., 561
Anderson, F. W., 869
Andiro anthelmintico, 829
arorabo, 528, 829
incrmis, 829
roseo, 829
retuso, 829
spinuloso, 829
vermifuga, 829 rermifuga, 829
Andrachue cordifalio, 590
Andrec, G., 348
Andrews, F. M., 869
Andrews, L., 869
Andromeda, 667, 673-674, 863
calyculota, 65
Cotesbaci, 65
floribundo, 127, 674
glaucaphylla, 127
joponica, 65, 666
palifolio, 65, 127, 666, 673-674, f. 674, 820
Andromedotoxin, 64, 65, 127, 666, 667, 668, 669, 803, 863 vermifuga, 829 863 Audrapogau, 339, 344-348 anuulatus, 826 citratus, 344-345 halapensis, f. 344, 345, 826 Ischaeman, 276 unricotus, 345 unricotus, 345 Nardus, 345 Schoenauthus, 344 sorghum, 102, 345-348, f. 345 Androsoce, 89 Anemorrheno osphadeloides, 834 Anemia ablangifolia, 824 Anemare, 109, 447, 453-456, 863 altaico, 844 altaico, 844
openiun, 844
conodcusis, 455
caronorio, 844
European, f. 454
Meadow, 455
uenoroso, 4, 77, 109, 844
Pasque, f. 454, 454
potens, 76, 454
potens, 76, 454
potens, 7. 11/alfgangiana,
109, 446, 453-455, f. 451,
844 844 pratensis, 844 Pulsatilla, 453, 454, f. 454, 844 aninquefolia, f. 455, 455-456 ranunculaides, 844 sylvestris, 844 Wood, 77, 109

| Anemonic aeid, 111, 454 | Anthrax, 8, 177 | Japanese, 647 |
|--|--|--|
| Anemonin, 109, 446, 454, | Baeillus, f. 178 | undicaulis, 647, 808 |
| 459 | Symptomatic, 175 | racemosa, 647, 808 |
| Anemonol, 111 | Anthriscus cerefolium, 856 | spinosa, 808 |
| Anesthetic, 803 | sylvestris, 856 | ARALIACEAE, 89, 125, 646-647, |
| Anethol, 648 | vnlgaris, 856 | . 808 |
| Anethum graveolens, 648 | Anthurium Ferrierense, 808 | Aralien, 647 |
| | Authorna 245 | Auntin 647 |
| Anenrosis, 77 | Anthurus, 245 Antiarin, 148, 406, 863 | Araliin, 647 |
| Angelie acid, 593, 648, 789 | Antiarin, 148, 4061 863 | Araroba, 528 |
| Angelica, 650, 660-661 | Antiaris, 148, 863 | Arancaria brasiliana, 327 |
| atropurpurea, 126, 660-661 | As Arrow poison, 1, 52 | excelsa, 327 |
| Great, 660-661 | As Fish poison, 52 | Aranjia sericifera, 809 |
| Purple-stemmed, 126 | toxicaria, 1, 52, 85, 148, 406, f. 408, 587, 857 Antidotes, 2, 72, 73-77, 79-81, | Arbar toxicaria, 863 |
| Angiapteris erecta, 824 | 406, f. 408, 587, 857 | vitae, 328 |
| ANGIOSPERMAE, 102, 155 | Antidotes, 2, 72, 73,77, 79,81 | Arbutin 505 665 668 |
| | 862 | Arbutin, 505, 665, 668 Arbutus, 665 |
| Ang-quae, 247 | | Andreaking 920 |
| Angraecum, 90, 392 | Antifebrin, 78, 147 Antipyrin, 78, 147 | Andracline, 820 |
| fragraus, 629, 840 | Antipyrin, 78, 147 | Trailing, 127, 665 |
| Anguillaria diaica, 838 | Antirrhinum majus, 733 | Mentiesii, 005 |
| Angustura Bark, 582 | Antiscorbutic, 803 | Unedo, 665, 820 |
| l'alse, 688 | Antispasmodie, 803 | varians, 820 |
| | Antitoxin, 164, 165, 863 | xalapensis, 821 |
| Angusturin, 582 | Apeiba, 621 | Arcenthabium, 106, 415 |
| Anbalonidin, 637 | APETALAE, 156, 397 | ARCHICHLAMYDEAE, |
| Anhalonin, 637 | Aphanes Braunii, f. 208 | 156 305 |
| Anhalanium, 88, 837, 863 | Aphanizomenon, 93 | 156, 395 Arctium, 757, 758, f. 796, 797 |
| fissuratum, 637 | | I abbo 141 755 (n=/ 70%) |
| Jaurdanianum, 637 | Aphanomyees stellatus, f. | Lappa, 141, 755, f. 796, 797 |
| Lewinii, 125, 637 | 208 | major, 68 |
| brianaticum 627 | Aphonopetalum resinosum, | minus, 756 |
| prismaticum, 637 | 851 | Arctastaphylas, 629 |
| Williamsii, 637 | Aphrodaesein, 617 | palifolia, 821 |
| Anide, John, 869 | Apium gravealens. 126, f. | Ura-nrsi, 77, 665, 666, 821 |
| Anide, John, 869 Anilin, 72, 147 | 647, 648, 856 | Areca, 863 |
| Colors, 78 | leptophyllnm, 650, 856 | Catechn, 370, 849 |
| 1)yes, 72 | nadiflorum, 856 | Catechia v. nigra, 840 |
| Animals, action of poisons | Apoguniaria 52 00 120 130 | |
| on, 6, 7 | APOCYNACEAE, 52, 90, 129-130, | Areenidin, 370 |
| Auica 474 967 | 147, 683, 691-695, 806- | Areenin, 370 |
| Anise, 474, 863 Oil, 648 | 807 | Areeolin, 78, 370 |
| Conta 640 | Apoeynein, 692 | Arena labata, 623 |
| Seeds, 648 | Apoeynin, 129, 148, 692 | Arcuaria, 89 |
| Star, 474, 863 | Apacynum, 692-693 | serpyllifolia, 436, 813 |
| Tree, 112 | andrasaemifalium, 129, 692- | Arenga pinnatus, 370 |
| Anisette, 474 | 693, 807 | saccharifera, 840 |
| Anisomeria drastica, 434, 841 | cannabinum, 129, 692, 6 93, | Argania Sideroxylon, 681 |
| Annatto, 627 | f. 693, 807 | Argemone, 480, 483, 863 |
| Annual, Mercury, 603, f. 603 | renetum, 807 | alba, 841 |
| Annatto, 627 Annual, Mercury, 603, f. 603 Sow Thistle, 759 | sp., 148 | intermedia, 113, 483, 841 |
| Anomospermum japurense, | Apomorphin, 59, 78, 147 | mexicana, 88, 90, 113, 483, |
| 838 | Apple, 504 | 841 |
| Anana | Balsam, Wild, 751 | Arginin, 548 |
| amara, 806 | Bark, 506 | Argyrolabinum pumilum, 829 |
| Chevimalia, 806 | Commou, 116, 512, f. 512 | Argyraesein, 123, 617 |
| | Crab | |
| muricata, 806 | | Ariein, 742 |
| palustris, 806 | Eastern, Wild, 512 | Arisaema, 372 |
| reliculata, 806 | Iowa, 512. f. 513 Old World, 512 | curratum, 808 |
| spinescens, 806 | Old World, 512 | Dracontium, 103, 372 |
| squamosa, 806 | Wild, 512 | tartuasum, 808 |
| Anonaceae, 111, 444, 446, | Wild, 512 Custard Family, 111 | triphyllnm, 103, 372, f. 372, |
| 476-477, 806 | Mammey, 627 | 808 |
| Austie, Dr., 471 | May, 75, 112, 469-471, f. | culgare, 808 |
| Antennaria, 757 | 170 | Aniatida 220 252 252 |
| Anthelminsia 202 | of Peru, 131, 726 | Aristida, 339, 352-353 |
| Anthelmintic, 803 ANTHEMIDEAE, 757, 758 | Rose, 639 | basiramea, f. 353 |
| ANTHE MIDEAE, 757, 758 | | hygrometrica, 352, 354 |
| Authemidin, 754 Authemin, 789 Anthemis, 757, 758, 787-789 | Star, 679, 681 | tuberculosa, 352-353 |
| Authemin, 789 | Thorn, 715, 729-733 | Purple, 352, f. 353 |
| Anthemis, 757, 758, 787-789 | | Aristalachia, 106, 417 |
| acincusis, 814 | twigs, 150 | /1/13141RCHIQ, 100, 417 |
| altissima, 814 | Apricot, 505, 514 | anguicida 808 |
| arvensis, 140, 788-789, 815 | Apricot, 505, 514 Hydrocyanic acid in, 88 | anguicida, 808 |
| quatriana 015 | Apricot, 505, 514 Hydrocyanic acid in, 88 | anguicida, 808 antihysterica, 808 |
| | Apricot, 505, 514 Hydrocyanic acid in, 88 | anguicida, 808 antihysterica, 808 argentina, 808 |
| avstriaca, 815 Blancheana, 815 | Apricot, 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 8 | anguicida, 808 antihysterica, 808 argentina, 808 |
| Blancheana, 815 | Aprient, 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduet, Claudian, 8 Aquifoliaceae, 123, 607, 614, | anguicida, 808 antihysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418 _{b.} |
| Blancheana, 815 chia, 815 | Apricot, 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 8 AQUIFOLIACEAE, 123, 607, 614, 807 | anguicida, 808 antihysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 |
| Blancheana, 815 chia, 815 Cata, 815 | Aprient, 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 8 AQUIFOLIACEAE, 123, 607, 614, 807 Aquilegia caerulea, 446, 844 | anguicida, 808 antihysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418 _b . 809 elegans, 808 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 170, 140, 756, 787 | Apricot, 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 8 AQUIFELIACEAE, 123, 607, 614, 807 Aquilegia caerulea, 446, 844 canadensis, 446, 844 | anguicida, 808 antihysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 European, 106, 417, f. 418 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 130, 140, 756, 787- 788, 815 | Apricot, 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 8 AQUIFOLIACEAE, 123, 607, 614, 807 Aquilegia caerulea, 446, 844 canadensis, 446, 844 vulgaris, 53, 446, f, 447, 844 | anguicida, 808 antihysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 Enropean, 106, 417, f. 418 grandiflora, 417, 809 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 170, 140, 756, 787- 788, 815 chuncis, 815 | Apricot. 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 80, 807 Aquilegia caerulea, 446, 844 canadensis, 446, 844 vulgaris, 53, 446, f. 447, 844 Arabin, 530 | anguicida, 808 antilhysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 European, 106, 417, f. 418 grandiflora, 417, 809 indica, 417, 809 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 170, 140, 756, 787- 788, 815 chuncis, 815 | Apricot. 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 80, 807 Aquilegia caerulea, 446, 844 canadensis, 446, 844 vulgaris, 53, 446, f. 447, 844 Arabin, 530 | anguicida, 808 antihysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 European, 106, 417, f. 418 grandiflora, 417, 809 indica, 417, 809 Kaempferi, 809 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 170, 140, 756, 787- 788, 815 elhuensis, 815 montana, 815 nabilis, 754, f. 788, 789, 814 | Apricot, 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 8 AQUIFOLIACEAE, 123, 607, 614, 807 Aquilegia caerulea, 446, 844 canadensis, 446, 844 rulgaris, 53, 446, f. 447, 844 Arabin, 530 ARACEAE, 53, 89, 103, 370- 372, 808 | anguicida, 808 antihysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 European, 106, 417, f. 418 grandiflora, 417, 809 indica, 417, 809 Kaempferi, 809 longa, 809 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 170, 140, 756, 787- 788, 815 elhuensis, 815 montana, 815 nabilis, 754, f. 788, 789, 814 Anthistiria prastrata, 826 | Apricot. 505. 514 Hydrocyanic acid in, 88 Apthanhyta. 302 Aqueduct. Claudian, 8 Aquifeulacear, 123, 607, 614, 807 Aquilegia caerulea, 446, 844 canadensis, 446, 844 rulgaris, 53, 446, f. 447, 844 Arabin, 530 Araceae, 53, 89, 103, 370- 372, 808 Arachis Hypagaca, 521, f. | anguicida, 808 antilhysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 European, 106, 417, f. 418 grandiflora, 417, 809 indica, 417, 809 Kaempferi, 809 longa, 809 macraphylla, 417, f. 417 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 1.30, 140, 756, 787- 788, 815 elhucnsis, 815 montana, 815 nabilis, 754, f. 788, 789, 814 Anthistiria prastrata, 826 Anthoccrcis, 853, 863 | Apricot, 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 8 AQUIFOLIACEAE, 123, 607, 614, 807 Aquilegia cacrulca, 446, 844 canadensis, 446, 844 rulgaris, 53, 446, f, 447, 844 Arabin, 530 ARACEAE, 53, 89, 103, 370- 372, 808 Arachis Hypagaca, 521, f, 522 | anguicida, 808 antihysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 European, 106, 417, f. 418 grandiflora, 417, 809 indica, 417, 809 kaempferi, 809 longa, 809 macraphylla, 417, f. 417 inexicana, 417 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 170, 140, 756, 787- 788, 815 elbucnsis, 815 montana, 815 nabilis, 754, f. 788, 789, 814 Anthistiria prastrata, 826 Anthoccrcis, 853, 863 Anthacleista grandiflara, 836 | Apricot. 505. 514 Hydrocyanic acid in, 88 Apthanhyta. 302 Aqueduct. Claudian, 807 Aquilegia caerulea. 446, 844 canadensis. 446. 844 rulgaris. 53. 446, f. 447, 844 Arabin, 530 Araceae, 53. 89, 103. 370- 372. 808 Arachis Hypagaca, 521, f. 522 Aragallus spicatus. 569 | anguicida, 808 antilhysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 European, 106, 417, f. 418 grandiflora, 417, 809 indica, 417, 809 Kaempferi, 809 longa, 809 macraphylla, 417, f. 417 mexicana, 417 pallida, 809 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 130, 140, 756, 787- 788, 815 elbuensis, 815 montana, 815 nabilis, 754, f. 788, 789, 814 Anthistiria prastrata, 826 Anthocercis, 853, 863 Anthaclesta grandiflara, 836 Anthacanthum, 90 | Apricot. 505. 514 Hydrocyanic acid in, 88 Apthanhyta. 302 Aqueduct. Claudian, 807 Aquilegia caerulea. 446, 844 canadensis. 446. 844 rulgaris. 53. 446, f. 447, 844 Arabin, 530 Araceae, 53. 89, 103. 370- 372. 808 Arachis Hypagaca, 521, f. 522 Aragallus spicatus. 569 | anguicida, 808 antihysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 European, 106, 417, f. 418 grandiflora, 417, 809 indica, 417, 809 kaempferi, 809 longa, 809 macraphylla, 417, f. 417 inexicana, 417 |
| Blancheana, 815 chia, 815 Cata, 815 Cotula, f. 170, 140, 756, 787- 788, 815 elbucnsis, 815 montana, 815 nabilis, 754, f. 788, 789, 814 Anthistiria prastrata, 826 Anthoccrcis, 853, 863 Anthacleista grandiflara, 836 | Apricot, 505, 514 Hydrocyanic acid in, 88 Apthanhyta, 302 Aqueduct, Claudian, 8 AQUIFOLIACEAE, 123, 607, 614, 807 Aquilegia cacrulca, 446, 844 canadensis, 446, 844 rulgaris, 53, 446, f, 447, 844 Arabin, 530 ARACEAE, 53, 89, 103, 370- 372, 808 Arachis Hypagaca, 521, f, 522 | anguicida, 808 antilhysterica, 808 argentina, 808 brasiliensis, 809 Clematitis, 106, 417, f. 418, 809 elegans, 808 European, 106, 417, f. 418 grandiflora, 417, 809 indica, 417, 809 Kaempferi, 809 longa, 809 macraphylla, 417, f. 417 mexicana, 417 pallida, 809 |

| sempervirens, 417 | European, f. 371 | Oryzae, 261, f. 261 |
|--|---|---|
| Serpentaria, 417, 809 | European, f. 371 Family, 372 | repens, f. 257 |
| sp., 809 Aristolochiaceae, 52, 106, | italicum, 371, 808 | subfuscus, 266 sulphureus, 257 |
| ARISTOLOCHIACEAE, 52, 106, 416-417, 808-809 ARISTOLOCHIALES, 156, | maculatum, 4, 53, 77, 371, f. 371, 808 | sp., 82 |
| ARISTOLOCHIALES, 156, | iriphymum, ouo | Asperugo procumbens, 811 Asperula, 90, 741 |
| 416-417 | 21 unuinui 10. 41/ | Asperula, 90, 741 |
| Aristolochin, 417 Aristolochinin, 417 | Asafoetida, 648, 650 Asahina, Y, 870 Asamin, 62 | cynanchica, 848 odorata, 744, 848 |
| Aristotelia Macqui, 622 | Asamin, 62 | Aspidin, 319 |
| Armeria elongata, 842 Arnica, 89, 140, 758, 794-795 | Asarın, 416, 417 | Aspidium, 101, 315 |
| alpina, 795, 815 | Asarum albivenum, 809 arifolium, 809 | Eilirmas f 200 f 212 315 |
| Chamissonis, 815 | canadense, f. 415, 416, 809 | Filix-mas, f. 309, f. 312, 315, f. 318, 319, 824 fragrans, 319 |
| cordifolia, 140, 794-795 | caudatum, 809 | fragrans, 319 |
| European, 140 | europaeum, 416, 809 virginicum, 809 | marginale, 101, 315, 319- 320, f. 320, 824 odoratum, 824 |
| mollis, 795 monocephala, 795 | ASCLEPIADACEAE 53 54 90 | odoratum, 824 |
| montana, 140, 755, 795, | ASCLEPIADACEAE, 53, 54, 90, 130, 683, 695, 697, 809- | prestulatum, 824 |
| 815 | 810 | rigidum, 824 |
| nudicaulis, 815 Root, 755 | Asclepiadin, 696 Asclepias, 695, 696-697 | Aspidosamin, 692 |
| Tincture of, 791 | campestris, 130 | Aspidosperma, 52 |
| Arnicin, 755, 794 Arnold, 548 | curassavica, 695, 809 eriocarpa, 130, 696, 809 | quebracho blanco, 147, 692, |
| Arnold, 548 AROIDEAE, 863 | eriocarpa, 130, 696, 809 | 807 |
| Aroids, 150, 371 | incarnata, 130, 696, 809 mexicana, 130, 696, 809 | sessilifolium, 807 Aspidospermatin, 692 |
| Arracacia, 648 | mexicana, 130, 696, 809 speciosa, 130, 696-697, f. | Aspidospermin, 147, 692 |
| xanthorrhiza, 648 | 607 | Asplenium, 315, 318-319 |
| Arrack, 198 Arrhenatherum avenaceum, | stellifera, 696 syriaca, 130, 697, 809 | Filix-foemina, 318-319 Asprella hystrix, 276 |
| 826 | tuberosa, 130, 695, 696, 809 | Astelia Banksii, 834 |
| elatius, f. 340 | vestita, 696 | Aster. 757, 776 |
| Arrow Arum, 372 | Asclepion, 130, 697 | Chinese, 756 |
| Poisons, Plants furnishing, 1, 2, 52-58, 386, 406, | Ascogonium, f. 21 ASCOLICIENES, 154, 307 | Chinese, 756 laevis, 776 multiflorus, 776 |
| 587, 606, 640, 687, 692, | ASCOIDEACEAE, 247 | New England, 776 |
| 803, 863 | ASCOMYCETES, 12, 100, | novae-angliae, 776 |
| Arrow-head, 335, f. 336 Large, 102 | 154, 222, 224, 247-281, 307, 859 | Parry's, 776 Parryi, 138, 776 |
| Arrow-root, 77, 391 | Ascospore, 12, f. 21 | White, Small, 776 |
| West Indian, 391 | Asebogenin, 666 | Woody, 138, 776 |
| Arsenic, 2 Poisoning by, 6 | Asebotin, 666 | ASTEREAE, 757 Asterionella, 188 |
| Statutes regarding, 6 | Ash, 629 Black, 683, f. 687 | Asthenia, 177 |
| Arsenious acid, 2 | Green, 683 | Asthenics, 803 |
| Artault, Stephen, 198 Artemisia, 756, 758, 790-794, | Mountain, 115 | Poisoning by, 75 Symptoms, 75 |
| | European, 629 Prickly, 581 | Treatment, 75 |
| Abrotanum, 755, 791, 815 Absinthium, 74, 140, 141. 755, 756, 791, 793, 815 arbuscula, 792 | White, 683 | Astier, 573 |
| Absinthium, 74, 140, 141. | Ashton, 607 | Actilha iahonica 500 |
| arbuscula, 792 | Ashworth, 870 Asia, Economic plants of, | Astragalus, 119, 531, 534, 535, 562-567, 863 |
| arenaria, 815 | 864, 865 | arizonicus, 567 |
| Barrelieri, 791 | Flora of, 865 | baeticus, 527, 829 |
| biennis, 140, 755, 791.792, | Medicinal plants of, 866 | Bigelovii, 564, 567, 829 caryocarpus, 558, 562, f. |
| f. 792 cana, 791, 792 | Poisonous plants of, 867 Asimin, 477 | 563 |
| Cina, 754 | Asminia, 476-477 | Coffee, 527 |
| Dracunculus, 756 maritima, 755, 791, 815 | triloba, 111, 476-477, f. 476, 806 | Crotalaria, 564 diphysus, 567 |
| maritima v. Stechmanniana, | Asparagin, 623 | exscapus, 829 |
| 791 | Asparagus, 375 | galegiformis, 829 |
| mexicana, 815 | medeoloides, 375 officianalis, 104, 375, 834 | garbancillo, 829 |
| pontica, 755, 815 tridentata, 140, 755, 791, | scaber, 834 | glycyphyllos, 829 gummifer, 530 |
| 792-793 | ASPERGILLACEAE, 100, 256-266 | hamosus, 829 Hornii, 119, 564, 829 |
| trifida, 792, 815 | Appareillania 20 20 20 963 | Hornii, 119, 564, 829 |
| vulgaris, 793-794, 815 Artemisin, 791 | Aspergillus, f. 21, 79, 199, 237-266, f. 258, 261, f. 262, 287, 863 | Lamberti, 566 lentiginosus, 564, 829 |
| Millianis, 107, 171 | 287. 863 | maximus, 829 |
| Arthur, J. C., 187, 227, 230, | flavescens, 29 | mollissimus, f. 30, 119, 534, 544, 562, 563, 564, 565, |
| 870 Artichoke, 656 | flavus, 100, 199, 262, 264, f. 261, 265, 595 fumigatus, 29, f. 20, 100, | 566 567 829 |
| European, 756 | fumigatus, 29, f. 20, 100, | 566, 567, 829 Martoni, 829 |
| Jerusalem, 710, 756 | 199, f. 261, 262, 265. | ochroleucus, 829 |
| Artocarpus calophylla, 857 | 595 | oocarpus, 564, 829 Pattersoni, 564, 829 |
| incisa, 405 integrifolia, 405 | glaucus, 100, f. 257, 257, 261, 287 | Thurberi, 567 |
| Arum, 89, 149, 863 | malignum, 261 | Pattersoni, 564, 829 Thurberi, 567 Astringent, 803 Atamasco Lily, 105, 387-388, |
| Arrow, 372 Dioscoridis, 808 | nidulans, 199, 266, f. 267 | Atamasco 1,11y, 105, 387-388, |
| | nigger 20 f 20 100 100 | 1. 388 |
| Dracontium, 808 | niger, 29, f. 29, 100, 199, 266, 505 | f. 388 Athamantin, 648 |

| Athanasia omara, 815 | Bacbelor's Button, 756 | Balloon Vine, 604 |
|---|---|--|
| Athyrocorpus persicorioefal- | BACILLARIALES, 153, 188- | Balm, Sweet, 710 |
| inni, 814 Atisin, 148, 450 | 189 Bacillus, 171, 863 | Balm-of-Gilead, 397, <i>f. 399</i> Balmony, 134 |
| Group, 450 | oceticus, 163 | Balsam, Apple, 751 |
| Group, 450 Atkinson, G. F., 96, 234, 237, 238, 240, 803, 870 Atkinson, V. T., 870 | ocidi-lactici, 250 alvei, 174 | Calaba, 627 |
| Atkinson, V. T., 870 | omylovorus, 163 | Family, 618-619 of Copaiba, 78, 529, f. 530 |
| Atractylin, 83 Atroctylis, 863 | anthracis, 97, f. 160, 775, 863 | of Tolu, 528 |
| gummifera, 83 | onthrocis v. symptomatici, | BALSAMINACEAE, 123, 607, 618- |
| Atribler, 426 | 175 | 620, 811 Bamboo, 338 |
| canescens, f. 425 halimus, 107, 813 hortensis, 107, 424, 813 | batulinus, f. 174, 174, 859 clooceoe, f. 164 | Banana, 391, f. 301 |
| hortensis, 107, 424, 813 | cali, 172 | Baneberry, 467-468, 864 Red, 108, 467-468 |
| laciniata, 107, 813 Nuttollii, 107, 813 | diptheriae, 97 enteridis, 172-173 | White, 108 |
| rosea, 813 | lorvoe, 175 | Bang, B., 871 |
| totarica, 813 vesicaria, 813 | muriscpticus, 176 oedemotis, 175, 863 | Banyan Tree, 405-406 Baptisio, 118, 530, 533, 534, |
| Atriplicismus, 107, 426 | oedemotis v. maligni, f. 175 | 535, 539-542 |
| Atropa, 863 | piscidus ogilis, 172, 859 | olbo, 90 |
| Belladonno, 4, 60, 133, 148, 150, 715, f. 716, 853 | prodigiosus, 163 psitticosis, 176 | oustrolis, 90, 540 brocteato, 117, f. 540, 541- |
| Atropamin, 715 | psitticosis, 176 Sorghi, f. 164 | 542 |
| Atropin, 3, 60, 61, 82, 148, 149, 238, 584, 715, 715, | subtilis, f. 160 suipester, 171-172, 863 | leucontho, 90, 117, 541, f. |
| 726, 728, 730, 731, 732, | tetoni, 174, f. 174, 863 | 541 perfolioto, 90 |
| 863 | typhi, f. 161 | tinctorio, 90, 540-541, 829 |
| Poisoning by, 61, 77, 78 Treatment for poisoning, | typhosis, 173, f. 173 zirgula, 863 | ziersicolor, 90 |
| 79 | Bacamacae rocave f 202 | Baptisin, 533, 540 Baptitoxin, 530 |
| Attalus Philometer, 2 | Bacon, Alice E., 870 Bacon, C. W., 870 Bacon, R. F., 52, 62, 870, 871 Bacteremia, 170, 173, 176 | Barborea vulgoris, 818 |
| Attar of Rose. 699 Auclair, J., 870 Aucuba japonica, 664, 817 | Bacon, R. F., 52, 62, 870, 871 | Barbarer, 468 6 460 |
| Aucuba japonica, 664, 817 | Bacteremia, 170, 173, 176 | Barberry, 468, f. 469 Family, 112, 469-472 |
| Aucubin, 664 Auld, S. J. M., 881 Aurand, S. H., 870 | Bacteria, 10-11, 97, 160, f. 161, 161-163, 775, 859 | Borbierio maynensis, 829 |
| Aurand, S. H., 870 | Biology of, 161-163 | polyphyllo, 829 Barbosa, R. J., 871 |
| Aurantia, 78 Aurantiamaric acid, 583 | Nitrifying, 163, f. 163 Poisonous properties, 10, | Bare, C. L., 871 |
| Auriculario, f. 202 | 163-165 | Barger, Geo., 278, 871 |
| Australia, Economic plants of, 864 | Saprophytic, 10, 97 Soil, 163 | Barilla, 424 Barium, 864 |
| Flora of, 865 | BACTERIACEAE, 165, 171-182 | in Loco Weeds, 40, 566, 567 |
| Poisonous plants of, 867 Australian, Gastrolobiums, | Bacterial poisons, 864 Classified, 10, 11 | Bark, Acacia, 533 |
| Australian, Gastrolobiums, | Bacteriolysin, 165, 177, 182 | Angostura, 582 Calisaya, 741, 742 |
| Indigo, 534 | Bacterium, 864 | Canella, 62/ |
| Salt-bush, 107, 425 Wattles, 523 | onthracis, 177-179, f. 178, f. | Cascarilla, 588 Cherry, Wild, 518 |
| Austria, l'oisonous plants 91, | ostheniae, 177 | Cinchona, 78, 87, 145, 146, 741, 742 |
| 867 Austrian, Pine, 329 | avium, 177 bovisepticum, 177 | 741, 742 Clove, 478 |
| Turpentine, 329 | cancrosi, 176 | Cotton Root, 624-626 |
| AUTOBASIDOMYCETES, 220, 233-247 | cancrosi, 176 choleroe, 177 | Cuprea, 742 Dita, 691-692 |
| Auto-intoxication, 10, 863 | coli commune, 172 diphtheriae, 179, 864 | Dogwood, 664 |
| Auto-poisoning, 73 | leproc, 180 | Funifera, 642 |
| Aveno, 339, 358-359 fotna, 102, 215, f. 358, 359, | mollei, 180 necrophorus, 179 | Hemlock, 327 Logwood, 528 |
| 826 | pestis, 181 | Maple, 615 |
| sativa, 102, 215, 359, f. | pneumonioe, 176 sanguinarium, 177 | Mulberry, Paper, 406 Oak, 403 |
| Avery, S., 96, 348, 870, 904 | suicida, 177 | Paper, 642 |
| Avocado, 478 Oil of, 478 | tuberculasis, 180, f. 180, 864 | Pereira, 147 Peruvian, 144, 741, 742 |
| Awned Brome Grass, 359-369, | Boeckeo frutescens, 839 | Quillaja, 78 St. Ignatius, 688 |
| f. 361 Axwort, f. 523 | Boeckeo frutescens, 839 Bailey, F. M., 871 Bailey, W. W., 871 | St. Ignatius, 688 Simaruba, 583 |
| Azaleo, 666, 667 | Baillon, H., 871 | Soap, 505 |
| California, f. 668, 668-669 javanica, 666 | Bolonites, 89 | Walnut, 401 |
| Azallo, 315 | oegyptiaca. 853 Roxburghii, 853 | Barker, 248, 871 Borlerio cristato, 692 |
| Azorella gleboria, 856 | BALANOPHORACEAE, 416 | Inpulino, 692 |
| В | Balanaps, 399 BALANOPSIDACEAE, 399 | Prionitis, 698 Barley, 366-367 |
| | BALANOPSIDALES, 156, | 2-rowed, 366 4-rowed, 366 |
| Babcock, H. H., 393 Baccharin, 755 | 399 Balfour, A., 346 | Little, 367, f. 368 |
| Bacchoris cordifolio, 138, 755. | Boliospermum axillare, 822 | Little, 367, f. 368 Smut, 98, 217 |
| 815 holimifolio, 138 | Ball, C. R., 345, 871, 903 Ballardini, 871 | Wild, 66, f. 67, 103, 338, f. 368 |
| , , , , | | 300 |

| Yellow leaf disease of, 281, | Sacred, 444 | Berberis, 89, 469, 471-472. |
|---|---|--|
| _ f. 281, 282 | St. Ignatius's, 74, 145 | 864 |
| Barasma crenulata, 583 Barrett, J. T., 100, 288 | Seeds, 867 | Aquifalium, 112, 472, f. |
| Barringtania, 2, 52, 89, 638, | Scarlet Runner, 119, 520, 527, 574 | aristata, 469, 810 |
| 647 | Sieva, 574 | 472, 810 aristata, 469, 810 repens, 112, 472 vulgaris, 469, f. 469, 810 |
| acutaugula, 839 | Soja, f. 520 | vulgaris, 469, f. 469, 810 Berchemia scandens, 620 |
| alba, 839 Butanica, 839 | Soja, <i>f. 52</i> 0 Soy, 520, 548 String, 520 | Berdan, T., 872 |
| Careya, 839 | Sword, 521 Tonka, 523, 552, 868 | Berensprung, 301 |
| insignis, 839 | Tonka, 523, 552, 868 Tree, 533 | Beresem, 525 Berg, 302, 303, 872 |
| intermedia, 839 nea-caledonia, 839 | Wax, 520 | |
| racemasa, 839 | White Indian, 574 Yam, 521 | Oil of, 86, 583 Berge, Fr., 862, 847, 872 Beringer, G. M., 872 |
| Barringtonin, 647 Barrows, W. B., 871 | Liearberry 665 | Beringer, G. M., 872 |
| Barth, Hermann, 871 | Beard Grass, 344-348 | Berlmann, M., 872, 908 Berlese, A. N., 206 |
| Barthelat, C. J.: 198, 871 Barthelcmy, 290 Bartholow, Dr. R., 453 Barton, W. C., 65, 871 | Beard Grass, 344-348 Beard-tongue, 733 Bccker, T. C., 872 Bedstraw, 771 | Bernegau, Ludwig, 85 |
| Bartholow, Dr. R., 453 | Bedstraw, 771 | Bernhard, II., 872 |
| Barton, W. C., 65, 871 | Sweet-scented, 744 | Bernon, 872 Berry, Indian, 1, 149 |
| Bartuch, F., 887 Barry, A. de, 257, 874, 880, 910 | Bee Plant, Rocky Mountain, | Fish, 1 Oso, 115 |
| | Beech, 864 | Oso, 115 Poison 123 |
| Basanacantha, 89, 90 | American, 403 European, 403 | Poison, 123 Berryer, G., 895 Bersama, 89, 838 |
| tetracautha, 848 Basellaceae, 424 | Family, 105 | Bersama, 89, 838 |
| Basidiabalus ranarum, 204 | Nut, 105 Beefsteak, Vegetable, 234 | Berthollet, 348 Berthollet, 3 |
| Basideal lichens, 307 BASIDIO-LICHENES, 307 | Becr, Bitter flavor of, 406 | Berthalletia excelsa, 638. j. |
| BASIDIOMYCETES, 98, | Spruce, 328 | 038 nabilis 638 |
| 154, 209·247, 307, 859 | Beet, Common, 424 Sugar, 107, 424-427, 868 | nabilis, 638 Bertrand, G., 872 Bertuch, F., 872 |
| Basidiaphara entospara, f. 205 Basil, 771 | Beggar's Lice, 707 | Bertuch, F., 872 |
| Sweet, 709 | Beggar-ticks, 779-781 | |
| Basiner, 459 Bassia, 89 | Black, 139 Small, 139 | Besley, F. W., 872 |
| butyracea, 680, 681, 851 | Beggar-weed, 525 | erecta, 650, f. 650 Besley, F. W., 872 Bessey, C. E., 96, 357, 415, 544, 593, 724, 872 Besta, C., 265, 595, 872, 876 |
| latifalia, 851 | Beggiataa, 91 | Besta, C., 265, 595, 872, 876 |
| langifolia, 851 Mattleyana, 851 | BEGGIATOACEAE, 166 | Deta vargaris, 107, 424 |
| Bassorin, 623 | Begania, 89, 150, 627 | Betain, 150 Betel Nut, 87, 396 |
| Basswood, 621 Bastard, Cardamon, 391 | gracilis, 810 Begoniaceae, 89, 627, 810 | raini, 370 |
| Toadflax, 416 Bastin, E. S., 871 | Begoniaceae, 89, 627, 810 Behr, H. M., 872 | Pepper, 396 Betula alba, 404 |
| Bastin, E. S., 871 | Behrend, 300 Behring, 534, 594 | alba v. papyrifera, 404 |
| Batchelor, J., 871 Bauberlein, 715 | Belladonna, 6, 72, 77, 148, 560, 718, 727, 731 732 | lenta, 404, 810 lutea 404 f 405 810 |
| Baubin, C., 275 | 560, 718, 727, 731 732 Source of drug, 87, 715 | lutea, 404. f. 405, 810 nigra, 404 |
| Bauhinia, 1, 52 coccinea, 829 | Statutes regarding, 6 | BETULACEAE, 810 |
| guianeusis, 829 | Beilstein, F., 872 | Beyeria viscasa, 822 Beyrichia scutellariaides, 852 |
| variegata, 829 Bann, 300 | Belamcanda chineusis, 105 punctata, 827 | Bezoars, 68, 359 |
| Baume, M., 871 | Belgium, Poisonous plants of. | Bhang, 411 Bibliography on poisonous |
| Baumgarten, P., 871, 872 Bay, 478 | 867 Belladonna, 864 | plants, 863-918 |
| Bay, 478 Bayberry, 639 | Belladonnin, 386, 715 | plants, 863-918 Bideus, 757, 779-783 bipinnata, 780-781 |
| Wax, 399 | Bellis perennis, 756 Bengal Cardaman, 391 | #15C014ea, 78U |
| Baycurn, 675 Bay rum, 399, 639 | | frondosa, 67, f, 68, 139, f 780, 781 Biennial Wormwood, 140, /. |
| Bazin, 297, 602 | Bennett, A. A., 143 Bennett, A. W., 872 Bennett, Hughes, 295 Bennett J. T., 662 BENNETTIALES, 326 | Biennial Wormwood, 140, /. |
| Beal Fruit, 582 Beal, W. J., 213, 359, 872 | Bennett, Hughes, 295 | 702 |
| Bean, 574, 864 | Bennett J. T., 662 | Bigelavia, 757 Bigelow, Dr. J., 601, 602, 665, 671, 873 |
| Adzuki, 520, 526 Broad <i>f</i> 522, 523 | Bentham, Geo., 872 | 665, 671, 873 |
| Broad, f. 522, 523 Burma, 574 | Bentley, R., 872, 913 Bentley, W. H., 872 | Bignania, 52 |
| Calabar, 72, 85, 148, 528, | Bentley, W. H., 872 Benzaldehyde, 517 | cuprealate, 135, 739, 810 Family, 134-135, 738-739 |
| 864 Cape, 574 | Benzene, 145 | venusta, 739 |
| Castor, 75 Coffee, f. 116, 117 | Benzoic acid, 144, 683 | BIGNONIACEAE, 52, 53, 134- 135, 698, 738-739, 810 |
| Common, 520, 574 | aldehyde, 505 Benzoin, 682 | Bikhaconitin, 450 |
| Common, 520, 574 Coral, 117, 542, 543 Java, 574 | aestivalis, 477 | Billardiera langiflara, 842 Bindweed, 701 |
| Java, 574 Kentucky Coffee, 116, 117 | Siam. 683 Benzoylecgonin, 575 | Bindweed, 701 Black, 421 |
| Kentucky Coffee, 116, 117 Kidney, 'Three-lobed, 520 Lima, 75, 87, 119, 520, 574 | Berbamin, 472 | European, 702 |
| Lima, 75, 87, 119, 520, 574 | Berberidaceae, 53, 63, 89, 112, 444, 469-472, 810 Berberin, 88, 90, 108, 112, 146, 468, 470, 472, 473, 473, 473, 473, 473, 473, 473, 473 | Hedge, 701-702, f. 702 Birch, 404 |
| Mescal, 88, 125 637 | Berberin, 88. 90, 108, 112. | Black, 404 |
| Moth, 526 | 146, 468, 470, 472, 473, | Cherry, 404 Paper, 404 |
| Ordeal, 528 | 478, 483, 803 | aper, to |

| Tar, 404 | Blastomycosis, 100. 303-306, | satanns, 235, 861 |
|--|--|---|
| White, 404 | f. 304, 864 Blazing Star, 104, 773 | scaber, 234 |
| Witches Broom on, 253 | Blazing Star, 104, 773 | Bitter, 234 Bolley, 11, L., 227 |
| Yellow, 404, f. 405 Bird-foot Violet, 631 | Blechnum exaltata, f. 312 | ROMBACEAE, 621 |
| Birthwort, 385 | occidentale, f. 312 Bleeding Heart, 480 | Bond, A. R., 873 |
| Bispham, W. N., 595, 873 | Blennorrhea, 170 | Bone Oil, 145 |
| Bispham, W. N., 595, 873 Bissell, C. 11., 873 | Blighia, 89 | Boneset, 755, f. 763, f. 77a, 770-772 |
| Bitter Almond, 88 | safida, 607, 850 | Oil of, 770 |
| Essence, 505 | Blight, Fire, 163 Sorghum, 163, f. 164 | Bonnaya veronicnefolia, 852 |
| Oil, 315 Pulp, 517 | Stem, 267, f. 268 | Boole, L. E., 873 |
| Seeds, 505 | Blite, Strawberry, 424 | Boorsnis, 392 |
| Seeds, 505 Boletus, 234 | Bloat, 68, 107, 118, 419, 424 | Boot Jack, f. 68, f. 786 Borage, 68, 704 |
| Cassava, 503, 587 Orange, 582 | Symptoms of, 554-556 Block, B., 873 | Family, 131, 227, 704-707 Plants, 864 |
| Bitternut, 402 | Block, 873 | Plants, 864 |
| Bitter-root, 423 | Blodgett, F. 11., 873 | BORAGINACEAE, 131, 698, 704- |
| Bitter-sick Trec, 628 | Blood-berry, 434 | 707, 811, 864 Borago officinalis, 704, 811 |
| Bittersweet, 60, 132, 614-615, | Bloodroot, 113, 479, 480, 483- 484, f. 484 | Barassus flabellifer, 840 |
| f. 615, 715, 718-719, f. | Bloodwort, 385 | Borna's disease, 26 Borneol, 417, 709, 710 |
| 7/9 Climbing, 123 | Blue, Cohosh, 112, 446, 469, | Borneol, 417, 709, 710 |
| Family, 123 | 485 | Borreria, 90 Boscia urens, 812 |
| Bitter-weed, 766 | Flag, f. 388 | Bose, C. L., 873, 882 |
| Yellow-flowered, 539 | Carolina, 389 Exotic, 389 | Bose, C. L., 873, 882 Boston Ivy, 620 Botrychium Lunaria, 313 |
| Bitting, A. W., 260, 290, 343, 769, 772, 873 | Large, 105 | Botrychium Lunaria, 313 |
| Bixa Orellana, 627 | Large, 105 Grape, Wild, 620 Grass, 127, 271, 272, 276, | virginianum, 313, 692 |
| BIXACEAE, 52, 53, 627, 810 | Grass, 127, 271, 272, 276, | Bothriospora corymbosa, 848 Bothriospora, 191 |
| Bjerregard, A. P., 873, 888 | 277, 338, f. 339 Indigo, 524 | Botryodiplodia, 286 |
| Black, O. F., 869, 873 | | Botrydium granulatum, f. 92. |
| Black, Ash, f. 682, 683 Beggar-ticks, 139 | Larkspur, 460, 466 Lettuce, 761-762 | 191, f. 191 |
| Birch, 404 | Lobelia, 136 Lupine, 118 | Bottytis, 297 Bottle Grass, Black spot on, |
| Snakeroot, 692 | Mould 82 / 266 256.257 | 280 |
| Bully Tree, 680 | Mould, 82, f. 255, 256-257 Passion Flower, 633 | 280 Botulism 11, 77, 174, 772, Botulismus 864 Bouchard C., 873 |
| Bottle Grass, 280 | Spiderwort, 373 | Botulismus (864 |
| Byrony, 149 Currant, 498 | Vervain, 708 | Bouchard, C., 873 Bouley, 290 |
| Drink, 614 | Violet, 125 Bluebell Family, 136 | |
| Elder, 746 | Blueberry, 665 | Bouncing Bet 62, 77, 103, |
| Fir, 327 | Dwarf, 665 | Bouncing Betty 6. 436, 441- |
| Hellebore, 446, f. 448, 449 Henbane, 133, 726-727, f. | Mountain, 665 | Bourgoing F 874 |
| 727 . | Tall, 665 | Bourgoing, E., 874 Bourlier, 572, 573 |
| I.ocust, 52, 55, f. 57, 64, f. 66, 118, 527, f. 559, 560 | Blue-bottle, 802 Blue-flowered Water Lily, 445 | Bourquelot, Em., 747, 874, |
| 00, 118, 527, f. 559, 560 | Blue-Green Algae, 91, 97, 160, | 879 |
| Medick, 553 Mold, 98, 195-200, f. 196, | 185, 187 Blueweed, 707 | Bourreria havanensis, 811 |
| 200 | Blueweed, 707 | Boussingaultia basellaides, 424 Boutet, 874 |
| Mustard, 77, 486, 488, f. | Blumenbachia, 633 grandiflora, 633 | |
| 489, 490 Nightehada 60 61 74 131 | Blyth, A. W., 2, 4, 6, 54, 55, | Boutiny, E., 874 Boutron, L., 505 |
| Nightshade, 60, 61, 74, 131, f. 132, 719-721, f. 720, 771 | Blyth, A. W., 2, 4, 6, 54, 55, 59, 60, 61, 62, 63, 72, | Borista plumbea, f. 246 |
| 771 | 85, 147, 149, 450, 480, | Bowbill, 874 Bowdichia, 52 |
| Oats, 66 | 551, 588, 607, 649, 651, 689, 727, 873 | virgilioides, 829 |
| Pepper, 77, 87, 396, /. 396 | 689, 727, 873 Blyth, M. W., 873 | Bower, F. O., 874 |
| Scours, 533 | Bocconia, 485 | Box, 122, 604, f. 606 Elder, 615, 629 |
| Snakeroot, 121, f. 585, 585- | cordata, 841 | Poison 533 |
| 586 | frutescens, 841 | Poison, 533 Boxwood, 864 |
| Tea, 628 | Bodin, F., 15, 265, 873, 884 Bocck, II. vnn, 4, 873 | Brabejun, 844 |
| Valuat / 400 401 | Boehm, 873 | Brachysema undulatum, 829 Bracket Fungus, f. 233, 234 |
| Walnut, f. 400, 401 Blackberry, 507 High-bush, 508, f. 508 | Boehmer, G. R., 873 | Bragantia tomentoea 900 |
| High-bush, 508, /. 508 | Boehmeria, 409 | Bragantia tomentosa, 809 Brain, Poisons acting on, 73 |
| 1411Y, 103 | nivea, 406 Boeninger, T. K., 873 | Braithwaite, P. F., 874 |
| Black-cap Raspberry, 508 | Baerhannia crecta, 839 | Brake, Common, 100-101, 313, 315, 317-318, f. 317 |
| Black-leg, 175 Blackman, V. II., 222 | hirsuta, 839 | 315, 317-318, /. 317 |
| Black-purple Vetch, 526 | repens, 839 | Pranch, Ivy, /. 672 |
| Black-spot of Grasses, 280 | Böhm. R., 4, 653, 873 Bog, Rosemary, 127 | Brandt, J. F., 4, 5, 874, 907 Brassica, 486-490 |
| Black-stem Rust, 227 | Bog-bean, See Buck bean | alba, 77, 486, 818 |
| Bladder Fern, 101, 319-320 | Boils, 171 | Brassica, 486-490 alba, 77, 486, 818 arvensis, 488, f. 489, 818 |
| Nut, 604 | Boleria, 135 | Besseriana, 490 campestris, 486 |
| Bladderwrack, f. 192, 193, 194 Bladderwort, 698 | Boletus, 234-235 | juncea, 490 |
| Blakeslee, A. F., 195 | alveolatus, 861 edulis, 234 | Napus v. dichotoma, 490 |
| Blanchard, R., 354, 873 | felleus, 99, 234, 861 | nigra, 77, 486, 488, f. 18). |
| Blanche, Dr. G. W., 380 Blankinship, J. W., 873 | felleus, 99, 234, 861 Frostil, 861 | 818 |
| Blankinship, J. W., 873 | luridus, 234, 235, 237, 861 | aleracea, 486 |

Pe-Tsai, 486
Rapa, 490
Brayera anthelmintica, 505,
f. 506, 595, 847
Brazil Nut, 638, f. 638
Poisonous plants of, 867
Passilian, Inggae, 631 Poisonous plants of, 867
Brazilian Ipecae, 631
Magonia, 64, 607
Stinging Nut, 596
Breadfruit, 405
Breadnut Tree, 405
Bready, G. W., 630
Brebcck, C., 320, 874, 883
Bredeman, G., 874
Breder's Gazette, 561
Brefeld, O., 213, 256
Bremia Lactucae, 205
Bretin, 874, 882, 914
Bridetia ovata, 53
retusa, 822 Bridelia ovata, 53
retusa, 822
Brewer, W. H., 564
Brier, Green, 104
Brigham, Johnson, 863
Brintball, 25
Bristle Fern, 313
Bristly Foxtail, f. 339
Britton, N. L., 874
Brida, 90, 826
Broad Bean, f. 522, 523
Brodie, D. A., 874
Brodie, E. C., 874
Broken-back, 575
Brome Grass, 66
Awned, 359-360, f. 361
Brannelia, 89
Pinguin, 373, 811 Bramelia, 89
Pinguin, 373, 811
BROMELIACEAE, 89, 372, 811
Bromelin, 373
Bromin, 72
Branus, 226, 340, 359-360
catharticus, 826
mollis, 826
secalinus, 826
Septoria on, 286
tectarum, 66, 359-360, f. 361
Bronchitis, 170, 171
Membranous, 263
Purulent, 176
Bronchomycosis, 264 Bronchomycosis, 264 Bronchomycosis, 264
Bronchopneumomycosis, 262
Brooke, J., 874
Broom, 527
Common, 147
Corn, 345, 864
Rape, 698
Scotch, 118, 551 Scotch, 118, 551
Broom-weed, 621
Brosimum Alicastrum, 405
Brouard, F., 874
Brown, Crum, 146
Brown, E. L., 874
Brown, Dr. J. J., 662
Brown, S. A., 874
Brown Mustard, 490
Brucea sumatrana, 853 Brown, Mustard, 490
Brucea sumatrana, 853
Brucin, 145, 146, 147, 686
Brunfelsia Hopeana, 715, 853
latifolia, 715
Brunnich, J. C., 54, 874
Brunton, T. L., 518
Brussels Sprouts, 486
Bryonia alba, 751, 818
diaica, 135, 751, f. 799, 818
Bryonicn, 751
Bryonin, 751
Bryonin, 751
Bryony, f. 749, 751
Black, 149
BRYOPHIYTA, 155, 308, 312
Bryophytes, 310 Bryophytes, 310 Bryum, 310 Bubonic Plague, 181

Buchanan, R. E., 24, 243, 720, 859, 874

Buchanania, 805

Buchner, H., 250, 302, 875

Buchner, M., 875

Buchu, 87, 583

Buckbean, 129, 690-691, f. 691 Buckeye, 123, 616-617 California, 123, 617-618 Family, 123, 616-618 Ohio, 123, 617, f. 619 Red, 123, 617, f. 619 Buckley, J. S., 25, 263, 265, 875 Ruckley, S. S. 875 875
Buckley, S. S., 875
Buckthorn, 123, 124, 229, 620
Family, 620-621
Sea, f. 640, 641
Southern, 679
Buckwheat, 12, 78, 106, 419420, 421
Family, 418-423
Flour, poisonous, 419
Budd, Charles, 875
Buddleia, 52, 89
brasiliensis, 836
curviflara, 836
elobasa, 836 curviflara, 836 globasa, 836 Lindleyana, 836 836 madagascariensis, madagascariensis, 836
palystachya, 836
variabilis, 836
verticillata, 836
Buechner, J. G., 875
Buffalo Berry, f. 639, 641
Bur, 718, f. 722, 722-724
Bugloss, Bristles of, f. 71
Viper's, 131
Bulb-bearing Hemlock, 126
Bulbine bulbosa, 376, 834
semibarbata, 834
Bulbocin, 237 Bulbocin, 237
Bulbous Crowfoot, 110, 459
Water Hemlock, f. 658, 659
Bull Nettle, 72, 122, 596, 724-725 Thistle, 141, 798, f. 800 Bulliard Pierre, 3, 803, 852. 875 Bully Tree, Black, 679 Bumelia lanuginasa, 679 lyciaides, 679 Tyciaides, 679
Bunch flower, 103, 379
Common, 379-380, f. 380
Bunt, 219
Stinking f. 219, 219-220
Wheat f. 219, 386, 805 Buphane disticha, 386, 805
Bur, Clover, 552
Cuember, 135
Marigold, 779
Oak, 403
Bur-reed, 332, f. 334
Burdock, 67, f. 68, 141, 756
Root, 755
Source of drug 87 Root, 755
Source of drug, 87
Bureau, E., 875
Burgess, T. J. W., 875
Burgundy Pitch, 328
Burke, R. W., 875
Burlew, J. M., 875
Burlew, J. M., 875
Burlay, Bean, 574 BURMANNIACEAE, 811 BURMANNIACEAE, 811
Burnet, 116
Burning Bush, 123
Burrill, T. J., 100, 288, 875
Bursera simaruba, 575
BURSERACAE, 575, 811
Buschke, 304
Busey, S. C., 875 Bush, Burning, 123
Calico, 669
Caustic, 696
Honeysuckle, 135
Busse, Otto, 249, 304
Busses, Bus

С

Cabhage, 486
Chinese, 486
Skunk, 371
Cacalia luberasa, 417
Cacao, 621, 624
Tree, f. 623
CACTACEAR, 125, 634-637, 811, 864
Cacti Spines cause phytobezoars, 68
Cactin, 637
Family, 634-637
Giant, 635
Night-blooming, 730
Old Man, 635
Cadaba indica, 812
Cadaverin, 10, 149, 172
Cadinenc, 648
Cactis, 71. P., 875
Cacsalpinia, 89
Banducella, 829
cariaria, 533, 829
pulcherrima, 829
Caeama, 221
Caffeidin, 628, 742
Caffeidin, 628, 742
Caffeid, 144, 146, 606, 614, 628, 742
Cahn, A., 875
Caisimon, 396
Cajanus indicus, 521
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Cajaputol, 639
Ca

Calandrina, 864

| Calceolaria, 733 | CAMPANULATAE, 158, | Carapa, Oil of, 575 |
|---|---|--|
| crenatiflora, 733 | 748-802 | procera, 575 |
| scabiosaefolia, 852 | Campbell, H. P., 875 Campbell, J. R., 379 Camphor, 72, 77, 87, 477, 864 | procera, 575 Caraven-Cachin, A, 602 Caraway, Oil of, 648 |
| Calcitripin, 464 | Campbell, J. R., 379 | Caraway, Oil of, 648 |
| Calculi, Renal, 107, 424-425 | Camphor, 72, 77, 87, 477, 861 | Source of drug, 87 Carbolic acid, 6, 77, 78, 79 |
| Calendula officinalis, 756 | Camphorin, 478 | Carbolism, 78 |
| Calicium, f. 308 | Campion, 439 | Carbon disulphial, 72 |
| Callicema serratifolia, 851 | Bladder, 437 Starry, 437 | in alkaloids, 143 |
| Calico Bush, 669 | Camptosema, 52 | monoxid, 72, 728 |
| California | pinnatum, 829 | Carbonic acid, Poisoning by |
| Azalea, 668-669, f. 668 | Canada Moonseed, 112 | 77 |
| Buckeye, 123, 617-618 | Pitch, 327 | Card, F. W., 875 |
| Economic Plants of, 864 | Thistle, 141, 798, f. 799 Violet, 631 Canadian Cocklebur, 763 | Cardamine amara, 818 |
| Flora of, 865 | Canadian Captalahur 763 | Cardamon Rastard 301 |
| Hellehore, 103, 104, 381 Laurel, 112, 478 | Canadin, 468 | Bengal, 391 |
| Lily, 375 | Canaigre, 418 | pratensis, 818 Cardamon, Bastard, 391 Bengal, 391 Java, 391 |
| Medicinal Plants of, 866 | Canarium commune, 811 | Malabar, 391 Round, 391 Cardiac Poisons, 73, 74-75, 148, 692, 803, 864 |
| Poison Ivy, 123, 609-613, f. | sp., 811 | Round, 391 |
| Poissoners Plants of 966 | Canavalia ensiforme, 521 | Cardiac Poisons, 73, 74-75 |
| Poisonous Plants of, 866 Poppy, 479, f. 480 | obtusifolia, 521, 829 | non-poisonous to insects |
| Rhododendron, 127, 667 | Candlenut Tree, 587 CANDOLLACEAE, 748 Candolla A de 275 732 | 7 |
| Walnut, 401 | Candolle, A. de, 275, 732 | Stimulant, 637 |
| Calla palustris, 371 | Candytuft, 486 | Carilinal-flower, 136 |
| Calliandra, 89 | Cane, Frosted, 346 | Cardinehermum Halicacahum |
| Houstoni, 829 | Canella alba, 627, 812 | 606, 850 |
| Callicarpa americana, 131, 708, 857 | Bark, 627 | 606, 850 Cardol, 78, 395, 608 Cardoon, 756 Cardopatium corymbosum |
| cana. 857 | CANELLACEAE, 627, 812 | Cardopatium corymbosum |
| cana, 857 longifolia, 857 tomentosa, 857 | Canna edulis, 391 | 815 |
| tomentosa, 857 | flaccida, 391, f. 391 | Carduus Flodmannii, 801 |
| Camrnoe involucrata, 624 | indica, 391 Cannabin, 106, 411 | natans, 65 |
| triangulata, 624 | Cannabin hybrid, 411 | Careya, 89 |
| Callistephus hortensis, 756 Callitris, 819 | Cannabinol, 411 | Careya, 89 Carex, 368, 369 arenaria, 360, f. 369 |
| quadrivalvis, 328 | Cannabis, 72, 410, 864 | hirta, f. 369 |
| Callsen, Jacob, 875 | Cannabis, 72, 410, 864 indica, 411, 445, 637 sativa, 74, 106, 410-411, f. | Carica, 89 |
| Calochortus, 375 | sativa, 74, 106, 410-411, f. | Papaya, 627, 841 |
| Calluna, 666 | 410, 857 | quercifolia. 627, 841 |
| rulgaris, 665 | CANNACEAE, 391 | spinosa, 841 Caricaceae, 627 |
| Callunatannic acid, 666 | Canoe Gum, 405 | Caricin, 627 |
| Calomel, 78 Sweet, 371 Vegetable, 371 | Cantharellus aurantiacus, 859 Cantharides, 2, 6, 7 | Carissa ovata, 807 |
| Vegetable, 371 | Cantharidin, 72 | Xylopicron, 807 |
| Calaphyllum, 627 | Cantua, 89 | Carleton, M. A., 227, 230 |
| Calaba, 827 | buxifolia, 843 | Carlina acaulis, 815 |
| Inophyllum, 827 | pyrifolia, 843 | Carnation, 436 |
| mantanum, 827 Calotratis gigantea 809 | Caoutchouc, 588 | Carniolica, 148 |
| procera, 65, 695, 809 | Cape Bean, 574 | Caroba, 134, 739 |
| Calotropis gigantea, 809 procera, 65, 695, 809 Caltha, 447, 448-449 | Gooseberry, 715 | Carob Tree, f. 526, 527 Caroba, 134, 739 Carobin, 134 |
| arctica, 845 | Jasmine, 742 Myrtle, 637 Saffron 1 287 | Carolina Blue-nag, 389 |
| palustris, 4, 75, 381, 448- 449, 845 | Saffron. f. 387 | Jasmine, 147 |
| Caltrop, 120 | Caper, 495 | Larkspur. 108, f. 461 Moonseed, 472 |
| CALYCANTHACEAE, 112, 444, | Family, 111, 495-497 Spurge, 121, f. 590, 602 | Carougeau, M., 875 |
| 445. 475-476, 811 | Spurge, 121, f. 590, 602 | Carpa moluccensis, 837 |
| Calveanthin, 112, 476 Calveanthus, 475 | Capillitium, 160 | Carpain, 627 |
| Calveanthus, 475 | CAPPARIDACEAE, 52, 114, 479, 486, 495-497, 812 | Carpenter, C. R., 875 |
| florishs, 112, 475, 476, f. | Capparis Cynophallophora, | Carr, F. 11., 278, 871 |
| 475 glaucus, 811 | 812 | Carrageen, 194 |
| sp. 112 | ferrnginea, 812 | Carrion Flower, 64 |
| Calyptospora Gaeppertiana, | glabifera, 812 | Carrot, f. 68, 127, 647, 663 Family, 647-664 |
| 222 Common 277 270 864 | odoratissima, 812 spinosa, 495 | Wild, 663-664 |
| Camas. 377-379. 864 Death, 103. 375, f. 376, 377- | Capraria biflora, 629, 852 | Carruthers, Wm., 875, 876 |
| 378. 466 | CAPRIPOLIACEAE, 53, 135, 741. | Carter, Marion H., 876 |
| Poison, 378 | CAPRIFOLIACEAE, 53, 135, 741, 744-748, 812-813 | Carthamus tinctorius, 756 |
| 378, 466 Poison, 378 Swamp, 103 Cambier, Jacob, 875 Cameling, 486, 403 | Capsacutin, 726 Capsaicin, 726 | Cartwright, C. B., 876 |
| Cambier, Jacob, 875 | | Carum capense, 856 |
| Camelina, 486, 493 sativa, 493, f. 494 | Capsella, 487, 493, 864 | Carri, 648 |
| sativa, 493, f. 494 Camellia, 89, 629 | Bursa-pastoris, f. 492, 493, | Copticum, 856 Petroselinum, 648, 856 |
| Hongkongensis, 629 | 818 Capiseum 717 725-726 | Carvacrol, 709 |
| japonica, 855 | Capiseum, 717, 725-726 annunm, 77, 395, 714, 725- | Carver, Geo. W., 272 |
| sasanqna, 855 | 726, f. 725, 853 frutescens, 714, 725, 853 minimum, 853 | Carver, Geo. W., 272 Carvol, 648, 709 |
| sasanqna, 855 Cameraria latifalia, 807 | frutescens, 714, 725, 853 | Carya cordiformis, 402 |
| Caminhoa, J. M., 875 | | |
| | minimum, 853 | glabra, 402 |
| CAMPANULACEAE, 52, 136, 811- 812 | minimum, 853 Caragana, 527, 534 Caraipa fasciculata, 855 | glabra, 402 illinoiensis, 401 laciniata, 401 |

ovata, 401 tomentosa, 402 Caryocar, 52, 89 glabrum, 855 CARYOCARACEAE, 627 CARYOPHYLLACEAE, 63, 89, 107-108, 423, 424, 435-444, Caryophyllin, 639 Casali, 869, 876 Cascara, 864 sagrada, 620 Cascarilla, 588 sagrada, 620
Cascarilla, 588
Cascarillin, 588
Cascarillin, 588
Cascarillin, 588
Cascinium fenestratum, 473
Caslew, 607-608, 864
Family, 607-614
Nut, 608
Oil, 608
Casimiroa edulsi, 849
Cascaria esculenta, 627
graveolens, 850
guineensis, 850
tomentosa, 850
Cassava, 87, 587, 864
Bitter, 503, 587
Bread, 587
Meal, 587
Cassia, 477, 528, 535-536
acutifolia, 528, 536, 829
aluta, 829
angustifolia, 528, 536, 829
Clamaechrista, 117, 529,
536, f. 536
didymobotrya, 829
Fistula, 529, f. 532
hirsuta, 829
lispidula, 534, 536, 829
ktmiensis, 829
lacrigata, 829
marilandica, 117, 535, 536,
f. 537, 829
occidentalis, 523, 829
Oil of, 86, 523
Pomade, 523
Purging, 529, f. 532
Sophera, 830
Sturtii, 830 stipulacea, 830 Sturtii, 830 Castalia odovata, 445 tuberosa, 445 Castanea umericana, f. 4a2, dentata, 105, 403 japonica, 403 sativa, 403 Castanopsis Tungurrut, 824 Castanospermum australe, 527, 533, 830 Castillcia canescens, 852 Castilleia cauescens, 852
coccinea, 734
Catabrosa, 90, 826
Castillou clustica, 405, f. 406
Castor Oil. 594, 595, 864
Renn, 55, 56, 594-595
Plant, f. 56, 121, 594-595, f. 595
Secd, 149, 594
Casuarinaceae, 395 CASUARINACEAE, 395 Casuarina equisetifolia, 395 Cat-tail. Common, 102, 332. Catalogue of Poisone Plants, 803-861
Plants, 803-861
Catalpa, 739, 864
bignonioides, 134, 739
Common, 134, f. 740
Hardy, 134, f. 740
speciosa, 134, 739, f. 740
Catalpin, 739
Catarrii, 170
Castrojutestical 20, 278 Poisonous Gastro-intestinal, 20, 279

Catasetum, 392, 840
Catch-fly, 436, 437
Night-flowering, f. 437, 438439, f. 439
Sleepy, 108, 438, f. 438
Catechin, 533
Catechol, 530
Catechu, 529
Catechuranic acid, 530 Catechutannic acid, 530 Catechutannic acid, 530
Catesby, 470
Catyut, 558
Catlus edulis, 501, 614, 813
Cathartic, 803
Cathartic acid, 408, 528
Cathin, 614
Catnip, 710
Cattell, II. W., 6, 876
Caucalis daucoides, 856
Cauliflower, 486
Caulophyllum, 89
thalictroides, 63, 112, 469, 484, 810 484, 810 Caustic Bush, 696 Cavantou, 3
Cavara, F., 285
Cayapouia ficifolia, 818
Cayenne Pepper, 77, 87, 133, 714, 1, 725, 725-726
Ceanothus, 124
americanus, 124, 621, 629, 846 846 azureus, 124, 846 azureus, 124, 846
caernleus, 846
integerrimus, 847
ovatus, 124, 621, 847
thyssillarus, 124, 847
velutinus, 124, 847
Cecropia, 405
Cedar, Oil of, 330
Platte, 330, 331
Red, 101, 102, 330-331, 1.
331, 332
West Indian, 575
White, 327, 328
Cedren, 101
Cedren-camphor, 101 Cedren-camphor, 101 Cedrene, 332 Cedrela odorata, 575 Cedrol, 332 Cedrol, 332 Cedron, 583 Ceir, 370 Celandine, 113, 479, 485 American, 480 Common, f. 485 CELASTRACEAE, 53, 123, 607, 614-615, 813 Celastrin, 614 Celastrus, 614-615, 864 articulatus, 615 scandeus, 123, 615, f. 615, scandens, 123, 615, f. 615 813 Celeriac, See Celery Celery, 126 Turnip-rooted, f. 647, 648 Celosia authelmintica, 805 Cetosia authelminica, 805 cristata, 431 trigyna, 805 Celtis occidentalis, 408 Cenchrus, 339, 352 Tribuloides, 67, f. 68, f. 59, f. 349, 352, 826 Ceni, C., 265, 595, 872, 876 Ceniin, 688 Centaurez, 757, 758, 801,802 Ceniin, 688
Centaurea, 757, 758, 801-802,
f, 802
calcitrapa, 815
Cyanus, 756, 758
scabiosa, 65
solstitialis, 141, 802
Centaury, American, 689
Centella asiatica, 648
Centipedo orbicularis, 815
Centrosema, 52 Centrosema, 52

amazonicum, 830 Plumieri, 830 CENTROSPERMAE, 156, 423-444 Century Plant, 105, 386 Cephaëlin, 742 Cephaëlis toxica, 848 Cephalanthin, 135, 744 Cepialanthus, 89, 744 occidentalis, 135, 744, 848 Cephalanthus, 89, 744 Cephalin, 78 CEPHALOTACEAE, 498 Cephalotaxns drupucea, 816 Ceratonia siligna, f. 526, 527 Ceratopetalum, 90 apetalum, 851 apetatiin, 851 demersum, 445 Ceratophyllaceae, 444, 445 Cerbera, 52 Odollam, 807 Tanghin, 692, 807 Cercis canadensis, 830 chinensis, 830 Silignastrum, 527 Cercocarpus ledifolius, 117 parviflorus, 89, 117, 503, 847 Cercospora angulata, 281 Cerebritis, 864 Enzootic, 259 Cerebro-spinal System, Poisons affecting, 73
Cereus, 125
Bouplandii, 811
flagelliformis, 811 giganteus, 635 grandiflorus, 125, 635, 637, 811 gummosus, 637 McDonaldii, 811 Night-blooming, 125, 635 speciosissinus, 635 senilis, 635 Ceropegia bulbosa, 809 Cestrum, 133 aurantiacum, 853 auriculatum, 853 cauliflorum, 133 elegans, 715-716 macrophyllum, 853 macrophyllum, 853
nocturum, 133, 853
pallidum, 716, 853
Parqui, 853
vespertinum, 853
Ceterach officinarum, 824
Cetraria islandica, f. 309
juniperini, 861
pinastri, 861
Cevadillin, 377
Cevadin, 103, 148, 377
Cevadina, 103
Chaerophyllum temulum, 659,
856 856 Chaetocladium Jonesii, 196 Chaetodiplodia, 286 Chaetomiaceae, 280-281 Chaetomium chartarum, 281 Chaia Resin, 627 Chailletia cymosa, 52, 575, 819 Tapura, 52 toxicaria, 575, 819 CHAILLETIACEAE, 52, 575, 819 Chamaemeles, 53 japonica, 503 Chamaelirium, 89 carolinianum, 834 Chamonuile, 756
Corn, 140, 788-789
German, 754
Oil of, 751 Roman, 754

| Chancre, Soft, 176 | 453, 462, 463, 466, 467. | Hog, 8, 171 |
|--|--|--|
| Chancroid, 176 | 453, 462, 463, 466, 467, 476, 479, 517, 518, 519, 537, 543, 548, 549, 561, 564, 565, 568, 581, 593, 594, 612, 617, 626, 628, | Hog, 8, 171 Asiatic, 181-182 |
| Chantemesse, A., 262, 263 Chapman, A. W., 876 Chapman, H. C., 876 | 537, 543, 548, 549, 561, | Organism of, 182 Cholesterin, 62, 506 |
| Chapman, H. C., 876 | 594, 612, 617, 626, 628, | Cholesterin, 62, 506 Cholin, 106, 237, 238, 648 As a remedy for ptomainc |
| Chara, f. 92, 194 | 030, 031, 033, 030, 007, | As a remedy for ptomainc |
| Chara, f. 92, 194 CHARALES, 154, 193-194 | 009, 0//, 084, 094, /20, | poisoning, 10 Chondria vermicularis, 859 |
| Chardinia xeronthemoides, 53 Charlock, 114 | 722, 730, 733, 762, 769, 775, 782, 784, 793, 803, | Chondrodendrum tomento- |
| English, 488, f. 480 | 804, 876, 877, 892, 916 | sum, 473 Chondromyces pedunculata, |
| Charras, 411 | Chestnut, 105 | Chondromyces pedunculata, f. 167 |
| Charteris, F., 876 Chaussier, H., 876 | American, f. 402, 403 Burs, 105 | Chondrus crispus, 194 |
| Chavicol, 648 Chayote, 750 | European, 403 | CHORIPETALAE, 156 Chorisia, 624 |
| Chayote, 750 Cheat, 66 | Oak, 403 Spanish, 403 | Christian, Robert, 746 |
| Septoria on, 285 | Water, 640 | Choristigma Stieckertianum. |
| Smut, f. 218 | Chevalier, J., 877 | 809 Christman, A. H., 222 |
| Cheeses, 624 Cheilanthes, 90 | Chevers, N., 877 Chibon, 575 | CHROOCCACEAE, 184-185 |
| frogrous, 824 | Chick Pea. f. 110, 120, 521. | Chrococcus turgidus, 185 |
| Cheiranthin, 78, 148 | Chick Pea, f. 119, 120, 521, f. 573, 573-574 | Chrozophoro plicata, 822 tinctoria, 822 |
| Cheiri, 486 | Chickasaw Pium, 505, 514 | Chrysanthemin, 789 |
| Cheiratin, 689 | Chickerinchee, 377, 864 Chickweed, 107, 442, 444 | Chrysanthennum, 90, 756, 753, |
| Chelcrythrin, 113, 485 Chelidonin, 480, 485 | Chickweed, 107, 442, 444 Cominon, 442, 444, f. 441 | 767, 789 Bolsomita, 815 |
| Chelidonium, 480, 485 | Unicorin, 137, 759 | cinerariifolium, 789, 815 |
| Chelidonium, 480, 485 majus, 113, 479, 485, f. 485, 841 | Chicory, 137, 755, 756, 758- | coccineum, 789 coronarium, 756 |
| Chelone globra, 134, 733, 852 | Root, 759 | coronarium, 756 frutescens, 756 indicum, 756 |
| Cheney, M., 464, 485, 503, | Chile Poisonous plants of | indicum, 756 |
| 602 | Chili, Poisonous plants of, 867 | Leuconthemum, 789 Marshallii, 789 |
| CHENOPODIACEAE, 52, 89, 107, 423, 424-430, 813-814 | Chimabbila maculota 842 | Parthenium, 756, 789, 815 |
| Poison in reduced by heat, | umbellato, 665, 842 Chimaphin, 665 | pinnotifidum, 815 sinense, 756 |
| 83 Chenopodin, 429 | Cilina, Poisonous plants of, | Summer, 756 Chrysler, M. A., 877 |
| Chenopodium, 52, 89, 107, | 867 Chincona, 848 | Chrysler, M. A., 877 Chrysobalanus Icaco, 575 |
| 427-429 | Red, 741 | Chrysocoma Coma-aureo, 315 |
| album, f. 426, 427 ambrosioides, 107, 424, 423, | Chinese Aster, 756 | Chrysoinyxa Rhododendri, |
| f. 428, 629, 813 | Cabbage, 486 Ginseng, 647 | 222 Chrysophan, 518, 528 |
| ambrosioides v. onthelmin- | Indigo, 421 | Chrysophanic acid, 418 |
| ambrosioides v. onthelmin- ticum, 107, 428, 429 anthelminticum, 814 | Jujube, 621 | Chrysophyllum Cainito, 679, 681 |
| Bouns-Henricus, 427, f. 427, | Sumac, 583, f. 584 Tea, 614 | Chrysosplenium alternifolium, |
| 814 Botrys, 428 | Umhrella Tree, 121 | 851 |
| californicum, 814 | Yam, 699 Wood, Oil of, 587 | omericanum, 851 oppositifolium, 115 |
| capitatum, 424 | Yeast, 198 | tetandros, 115 |
| hybridum, 428, 814 mexiconum, 107, 814 | Chinquapin, Water, 444 | Chrysotoxin, 277 |
| polyspermum, 814 | Chiococca brachiata, 848 _rocemosa, 848 | Churchill, J., 877 CHYTRIDIACEAE, 204 |
| Quinoa, 427 rubrum, 814 | Chionanthin, 683 | Cicer, 535, 573-574 |
| Vulvaria, 814 | Chiononthus, 89 picrophloia, 840 | arietinum, f. 110, 120, 521, f. 573, 573-574 |
| Chequen, 640 | virginica, 69, 129, 683, 840 | CICHORIACEAE, 748, 753, 762 |
| Cherry, 505, 514-519, 864 Birch, 404 | Chios Turpentine, 608 Chirata, 689 | CICHOREAE, 757 Cichorium, 757, 758-759 |
| Choke, 40, 116, 503, 514- | Chiratin, 689 | Cicliorium, 757, 758-759 Endivia, 756, 758 Intybus, 137, 755, 756, 758- |
| 515 Fuglish 505 | Chiratin, 689 Chives, 375, 383 | Intybus, 137, 755, 756, 758- 759 |
| English, 505 Ground, 715, f. 715 | Chlamydobacteriaceae, 165, | Cicuta, 44, 50, 651-659, 746 |
| Ground, 715, f. 715 Laurel, 85, 117, 503, 504 | f. 166, 183-184 | Cicuta, 44, 50, 651-659, 746 Bolanderi, 126, 658, 856 bulbifero, 49, 126, f. 658, 659, 856 |
| Leaves, 517, 629 Powdery Mildew on, 269, | Chlamydospores, f. 17 Chloral, 73 | 659 856 |
| f. 269 | Chloral hydrate, 6 | conformica, 830 |
| Sand, 116, 505 | Chlorocodon Whiteii, 809 | Douglasii, f. 655, 656 |
| Sour, 505, 514 Sweet, 514 | Chlorophora excelsa, 408 Chloroform, 6, 73 | moculota, 49, 79, 83, 126, 648, 651, f. 652, 652- |
| 317:11 60 | Clilarogalum, 89 | 030, 000, 001 |
| f, 515, 515-516 | pomeridianum, 63, 104, 389 Спропорнуста, 98, 154, 190- | occidentalis, 6, f. 48, 126, 856 |
| red, 116, f. 504, 505, f. | 193 | vagans, 49, 126, 665-658, f. |
| 516, 516-519 | Biology, 190, 191 | 657, 856 |
| Chesnut, V. K., 5, 31, 44, 64, | Choke Cherry, 40, 116, 503. | virosa, 4, 79, 83, 126, 652, |
| Wild, 60 black, 64, 117, 503, 505, f. 515, 515-516 red, 116, f. 504, 505, f. 516, 516-519 Western, 515 Chesnut, V. K., 5, 31, 44, 64, 65, 84, 96, 107, 111, | Choisya ternata, 849 Choke Cherry, 40, 116, 503, 514, 515 Choking Distemper, 21 | venenata, 856 virosa, 4, 79, 83, 126, 652, f. 653, 656, 864 |
| . 112, 121, 120, 130, 133, | Choking Distemper, 21 Chlorocystitis, 172 | Cicutin, 656 |
| 241, 328, 378, 379, 381, 383, 385, 386, 388, 394, | Cholera, 864 | Cicutoxin, 50, 77, 79, 126, 149, 435, 475, 648, 652, |
| 403, 430, 434, 440, 441, | Chicken, 177 | 653, 656, 864 |
| | | |

Cienkowsky, L., 305 Clarkia elegans, 644 CLATHRACEAE, 245 Clathrocystis, 91, 93, 184-185 aeruginosa, 184 Cimicifuga, 467, 468 foetida, 845 racemosa, 75, 446, 845
Cinchona, 741, 742, f. 742
Bark, 78, 87, 145, 146
lancifolia, 741, f. 742
Ledgeriana, 741
officinalis, 741 Kützingiana, 185 roseo-persicina, 185
Clathrus columnatus, 245
Claude, H., 877
Clavaria argillacea, f. 232
aurea, f. 232
Claviceps, f. 27, f. 274, 275-279, 864
microcephala, 276
purpurea, 28, 31, 76, 100, 275-279, 859
pusilla, 276
setulosa, 276
Clavija macrocarpa, 838
Clayton's Fern, 313, 322
Clearweed, 771
Clegg, 184
Cleistanthus, 864
collinus, 822
Clement, 874
Clematis, 109-110, 447, 456roseo.persicina, 185 succiruba, 741 Cinchonamin, 742 Cinchonidin, 145 Cinchonin, 145, 742 Cincol, 640, 710, 791 Cineral, 756 Cinerol, 754 Cinnabar, 2 Cinnamic acid, 683 Cinnamomum, 478
Camphora, 477
Cassia, 477
zeylanicum, f. 477, 477
Cinnamon Fern, 313, 322 Cinnamon Fern, 313, 322
Oil, 86
Tree, f. 477
Wild, 627
Cinnamylcocain, 575
Cinquefoil, Shrubby, 88
Cirsium, 757, 758, 797-801
Cirsium, 757, 758, 797-810
altissimum, f. 754
arvense, 141, 798-799, f. 709
canescens, f. 708, 801
discolor, 801
Drummondii, 798
criocephalum, 798
criocephalum, 798
tanceolatum, 141, f. 142,
798, f. 800
ochrocentrum, 801
nndulatum, 798, 801 Clement, 874
Clematis, 109-110, 447, 456457
aethusiaefolia, 845
alpina, 845
Bergeroni, 845
brachiata, 845
Buchaniana, 845
caripensis, 845
cirrhosa, 845
erecta, 110
Flanmula, 845
florida, 845 florida, 845 Fremonti, 110, 447 Gouriana, 845 Gouriana, 845
integrifolia, 447
Jackmanin, 446
lanuginosa, 447, 845
ligusticifolia, 110, 456
mauritiana, 845
orientalis, 447, 845
Pitcheri, 110, 447, 456, f.
457, 845
Pseudo-flammula, 447, 845
recta, 110, 447, 456
reticulata, 845
Viorna, 456
zirginiana, 109, 446, 456, f. nndulatum, 798, 801 undulatum v. megacephalum, 801 Cissampelos Pareira, 838 Cissus nivea, 124, 858 pruriens, 124, 858 CISTACEAE, 627, 814 Cistus polymorphus, 627 Citral, 708 Citric acid, 144, 505, 512, 529, 572, 583 Citron, 582, 750 Oil, 582 zirginiana, 109, 446, 456, f. Vitalba, 845 Wightiana, 845 Williamsii, 845 Williamsii, 845 Clement XIV, Poisoning of, 2 Clements, F. E., 224, 803, Citronella Oil, 345 Ins Colocynthis, 818 Citrullns 749, vulgaris, f. 748, 750 Citrus, 582, 583 Cleome, 52, 495-496, 497 Chelidonii, 812 frntescens, 812 gigantea, 812 Aurantium, f. 582, 582, 849 Aurantium v. vulgaris, 582 Bigardia, 582 Bigardia, 582
decnmana, 582, 583
japonica, 582, 583
medica, 53, 582, 583, 849
trifoliata, 582, 587
Cladonia furcata, f. 307
rangiferina, 307
CLADONIACEAE, 307
CLADONIACEAE, 307
Cladophora, 93, 190
fracta, 190
Cladosprium, 284, 861 graveolens, 812 lutea, 114, 496, f. 496 psoraleaefolia, 812 rosea, 812 rosed, 612 scriulata, 114, 496, f. 496 spinosa, 812 Western, 114 Yellow, 496 Cleomella, 495, 497 angustifolia, 497 Tracta, 190 Cladosporium, 284, 861 herbarum, 284, 286 Cladothrix boris, 183, f. 183 farcinica, 184 madureae, 184 Cleopatra, 2 Clerodendron Bhramaramari, 65 infortunatum, 857 serratum, 65 Siphonanthus, 858 Thompsonae, 708 Cladrastis amurensis, 830 Clammy Locust, 119, 560 Weed, 497 Clibadium, 52 Clantrian, G., 877 Clapp, A., 877 Clarke, J. F., 877 Barbasco, 815 surinamense, 815 Cliffe, 679

Climate, Toxicity of pla affected by, 83, 85 Climbing Bittersweet, 1 615, 1.79 Fern, 315 Fumitory, 480 Clinton Grape, 620 Chitorybe illudens, 99, 859 Clitoria, 1, 52, 534 amazonum, 830 arborescens. 830 arborescens, 830 Ternatea, 529, 830 Ternatea, 529, 830
CILLOROPHYCEAE, 307
Closed Gentian, 690
Closterium, f, 92, 189, f. 190
Clotbur, 137, 767.768
Spiny, f. 137
Thorny, f. 768
Clove, 89, 639
Bark, 478
Tree, f. 638
Clover, 150, 553-557, 864
Alsike, 525, 553, 554, 557
Bur, 552
Crimson, 68, 118, 556-557, Crimson, 68, 118, 556-557, 1. 556
Dodder, 130, 701
Hop, 552
Low, 553
Yellow, 553
Italian, 118
Japan, 525
Red, 279, 525, f. 553, f. 555
Rust, 20, 99, 230-231, f. 231 Crimson, 68, 118, 556-557, Rust, 20, 99, 230-231, f.
231
Sickness, 556
Stinking, 114
Stone, 553
Sweet, 552
White, 118, f. 551, 552
Yellow, 118, 552
White, 118, 525, 553, 557, f. 557
Club-root of Cabbage, f. 159, 160 160 Clusia macrocarpa, 827 Cluster-cup Fungus on Bar-berry, 226 Cneorum tricoccum, 853 Cnesmon tricoccum, 853 Cnesmone javanica, 822 Cnestis corniculata, 817 glabra, 817 polyphylla, 817 Cnicin, 756, 798 Cnicus, 757 Cnicus, 757
benedictus, 798
Coats, J., 877
Cobb, N. A., 284, 285
Cobaea scandens, 843
Coca, 575, f. 576, 576, 864
Cocain, 6, 73, 77, 79, 144, 147, 575-576, 614, 864
Cocainism, 77
Coccolacillus, 343 Cocco-bacillus, 343 Coccognin, 642 Cocculin, 112 Cocculus, 89 carolinus, 472 Ferrandianus, 838 rerrandianus, 838 glaucescens, 838 indicus, 112, 149 laurifolius, 838 toxiferus, 838 Coccus, Golden-pus, 167 White-pus, 167 Cochineal Plant, 635 Cockerell, V., 466 Cockle, Corn, 107-108, 439, f. 440 Meal, 441 Secds, 440 Cow, 108

| Cocklebur, 67, 137, 767-770 Canadian, 763, f. 769 |
|---|
| Canadian, 763, f. 769 Cock's-comb, Gangrene of, |
| 278 |
| Cocoa, 742 Cocoa-nut Palm, 370 Cocoon Antidote, 751 |
| Cocos amara, 840 |
| Cocos amara, 840 nucifera, 370, 840 Oil, 627 |
| Codamin, 147, 481 |
| Oil, 627 Codamin, 147, 481 Codein, 59, 77, 112, 113, 146, 479, 481 Codinene, 330 Coffee arabica, 629, 742, f. |
| Coffee arabica, 629, 742, f. |
| Coffea arabica, 629, 742, f. 743 matrifiana, 848 coffee, 629, 741, 742-744, 864 Adulterant of, 759 Bean, f. 116, 117 Mocha, 744 Plant, 742, f. 743, 744 Seeds, 629 Substitutes for, 523, 525, 527, 614 Tree Kentucky, f. 116, 117 |
| odorata, 848 |
| Adulterant of, 759 |
| Bean, f. 116, 117 Mocha, 744 |
| Plant, 742, f. 743, 744 |
| Substitutes for, 523, 525, |
| Substitutes for, 523, 525, 527, 614 Tree, Kentucky, f. 116, 117 Wild, 745 Cohagen, S. L., 552 Cohosh, 75 Black, 446 Blue, 112, 469 Coix Lacryma, 339 |
| Wild, 745 Cobagen S. L., 552 |
| Cohosh, 75 |
| Blue, 112, 469 |
| Cala a |
| Habit, 621 |
| Colchiccin, 375 |
| Nut, 621 Nut, 621 Nut, 621 Colchicein, 375 Colchicin, 77, 78, 79, 146, 143, 375, 479 Colchicum, 6, 72, 83, 85, 148, |
| 375. 864 |
| autumnale, 4, 148, f. 376, 834 |
| ncabolitanum 834 |
| Seeds, 867 speciosnm, 834 |
| voriegatum, 834 Coleman, 65 |
| Coleosphaerium, 185 |
| vortegatitit, 634 Coleosphaerium, 185 Coleosphaerium, 222 solidagiuis, 98, 222, 775 Sonchi-arrense, 861 Colic Root, 104 Colletia, 124 sbinosa, 621, 847 |
| Colic Root, 104 |
| Colletia, 124 spinosa, 621, 847 |
| Collin, Eng., 863, 877 |
| spinosa, 621, 847 Collin, Eng., 863, 877 Collins, G. W., 877 Collonia gracilis, 698 Colloturin 681 |
| Colocasia antiquorum, 808 |
| |
| riposa, 808 Colocynth, 749, f. 750 Colocynthin, 78, 749, 750 Colomia Discase, 279 Color of animals, 386 Colorado, Poisonous plants of, 866 |
| Colombia Disease, 279 |
| Color of animals, 386 Colorado, Poisonous plants of. |
| |
| Coltsfoot, 755 Colubrina fermenta, 621 |
| Columba Root, 473 False, 473 |
| Columbia Aconite, 46 |
| Columbina Fretzen 446 |
| European, 446, f. 447 Rocky Mountain, 446 Colombo, American, 689 Columella, 160 Colutea, 534 |
| Columbia, American, 689 |
| Colutea, 534 |
| arborescens, 534 crncuta, 551 Colville, G. W., 877 |
| Colville, G. W., 877 |

Colza, Indian, 490 Coma, 375 Comandra umbellata, 416 COMBRETACEAE, 53, 640, 814 Combretum brocteosum, 814 constrictum, 53 erythrophyllum, 814 phaneropetolum, 814 filaneropetolum, 814
racemosum, 814
trifoliatum, 814
Combs, R., 396, 587, 877
Comfrey, 704
Wild, 131, 706
Comma bacillus, 149 Commelina deficiens, 814 nudiflora, 814 tuberoso, 814 COMMELINACEAE, 89, 373, 814 COMMELINACEAE, 89, 373, 814
Commiphora abyssinica, 575
Comocladia, 608
glabra, 805
COMPOSITAE, 52, 53, 89, 90,
137-141, 222, 692, 748,
753-802, 814-816
Composite Family, 137
Concord Grape, 620
Condurango, 696
Cone-flower, 139, 779
Tall, 779
Western, 779
Conferva, 190
CONFERVALES, 190
CONFERVALES, 190
Conhydrin, 147 Conhydrin, 147 Conicein, 651 Conidiobolus utriculosus, f. Conidiophore of Aspergillus, f. 21 CONIFERAP, 155, 326, 327-332, 651, 816-817 Coniferin, 392 Conifers, 101, 327 Conicein, 651 Coniin, 3, 73, 77, 79, 126, 135, 144, 147, 648, 651, 652, 653, 726, 728, 747, 864 Conydrin, 651 Conium, 2, 6, 44, 650, 651 Alkaloids of, 147 machlatum, 4, f. 50, 51, 75, 126, 648, 651, 652, 855, CONJUGATAE. 154. 189-190 Conjunctivitis, 77, 169, 170 Connaraceae, 498, 817 Connaras africanus, 817 Connarus 1812 of 865 Connecticut, Flora of, 865 Connor, J. T., 877 Conopodium denudatum, 856 Consolidin, 706 Consumption, 180 CONTORTAE, 158, 683-697 Convallamarin, 104, 148, 385 Convallaria, 78, 377, 384-385, 864 majalis, 64, 104, f. 384, 384-385, 834 Convallarin, 104, 148, 385 Convicin, 572 Convolvent, 572
Convolvulaceae, 53, 89, 130, 698-702, 817
Convolvulin, 130, 699, 702
Convolvulus, 701-703 Convolvulus, 701-703 arvensis, 702 Family, 130, 698-702 floridus, 699 Scammonia, 699, 817 scoporius, 699 sepium, 130, 701-702, f. 702, 817 renenatus, 817 Convulsive Poisons, 803, 864

Conydrin, 120, 147, 651 Cook, Dr., 607 Cook, E. II., 877 Cook, O. F., 877 Cooke, N. C., 877 Copiaba, Balsam of, 78, 529, Copaibic acid, 529 Copoifera Langsdorfii, 830,... officinalis, 529, Salikorinda, 830 Copas, 864 Copernicia cerifera, 370 Copper sulphate as an algi-cide, 94-95 Coprinus, 234 comatus, 235
narcoticus, 859
Coprosoma linariifolia, 848
Copostella flavescens, 848
Coral Bean, 117, 542, 543
Fungi, f. 237
Tree, 527
Coral-berry, 135, 748
Corallocarpus epigaeus, 818
Corchorus capsularis, 621, f. 622, 855
Cord Grass. Fresh-water. f. comatus, 235 Cord Grass, Fresh-water, f ... CORDAITALES, 326 CORDAITALES, 326
Cordia alba, 704
Cordier, 877
Cordyceps, 273
cinerea, f. 273
militaris, f. 273
nophioglossoides, f. 273
Ravenelli, 273
Taylori, f. 273
Cordyline terminalis, 375
Corean Foxtail Millet, f. 349
Coreopsis, 756, 757
discoidea, 139 discoidea, 139 tinctorio, 756 Coriaceae, 607, Coriamyrtin, 607 Coriander, 648, f. 649 Coriandrol, 648 Coriandrum sativum, 648, f. Coriandrum sativum, 648, 4 649, 856 Coriario, 607 myrtifolia, 607, 817 naepaleusus, 817 ruscifolia, 607, 817 sarmentosa, 149, 607, 817 thymifolia, 607, 817 Tutu, 607 sarmentosa, 149, 607, 817
thymifolia, 607, 817
Tutu, 607
Cork, 403
Tree, 581
Wood, 6, 401, f. 717
Corkwood, 621, f. 717
Cormus foliosa, 503, 847
Corn, 102, f. 337, 338, 864
Broom, 345, 864
Chamomile, 140
Cockle, 62, 107-108, 439, f. 440
Meal, 441
Seeds, 440
Crop, Statistics of, 342
Dent, 341
Diplodia on, f. 287
Flusarium on, 287
Flint, 341
Kaffir, 345, 346, 347, 343, 864 Kaffir, 345, 346, 347, 343, Leaf-browning of, 281, f. 282 Oil, 342 Pod, 341 Pop, 341 Poppy, 112, 483 Rust, f. 230

Salad, 741 Silk, 342 Catyledan ventricosa, 501, Mitchelii, 830 818
Coulter, J. M., 878
Coulter, S. M., 878
Counarin. See Cumarin
Couperot, 877
Courbaril, 527
Courchet, M. L., 878
Court, G., 878, 904
Courtet, A., 878
Coville, F. V., 68, 96, 118, 237, 448, 557, 746, 878, 917
Cow Cockle, 108 paniculata, 830 sagittalis, 118, 543-546, 4. 818 Smut, 98, 210-215, f. 211, f. Soft, 341 Starchy sweet, 341 Sweet, 341 Wilt, 163 543 striata, 830 verrucosa, 830 Crotalism, 118, 543, 544-546, Wilt, 163
**CORNACEAE, 127, 646, 664, 817
**CORNACEAE, 127, 646, 664, 817
**CORNACEAE, 127, 648, 85, 83, 96, 803, 804, 877, 878
**CORNACEAE, 802
**CORNACEAE, 802
**CORNACEAE, 802
**CORNACEAE, 803, 804, 877, 878
**CORNACEAE, 804
**CORNACEAE, 817
**CORNACEAE, 817
**STATEMENT AND THE STATEMENT AND THE 864 Crotin, 595, 864 Crotinic acid, 588 Craton, 52, 121, f. 589, 599, 591, 593, 864 capitatus, 121, 591, f. 59t, 237, 448, 557, 746, 878, 917

Cow Cockle, 108

Oak, 403

Parsnip, 126, 663

Pea, 520, f. 532

Poison, 109

California, 463, f. 464

Wheat, 134, 734

Cowhage, f. 48, 49-51, 79, 33, 126, 652-656, 661, 364

Cowhage, 527

Cow-lierb, 442

Cownley, A. H., 878, 903

Cowslip, American, 676

English, 676

Coyotillo, 621

Crab-apple, Eastern wild, 512

Iowa, 512, f. 513

Old World, 512

Wild, 512

Crab's Eye, 527

Craig, R. A., 261, 291, 878

Cramber maritima, 486, 818

Cranberry, 665

Small, 665

Crandall Currant, 500

CRASSULACEAE, 114, 498, 500, 501-503, 818

Crassulacic acid, 501

Crataegus, 506, 513-514

mollis, 116, 504, 513-514, f. 514

orientalis, 503

Oxycantha, 53

punctata, 504, 513

Crawford, A. C., 40, 65, 84, 96, 124, 346, 465, 563, 566, 567, 626, 772, 878

Creeping Buttercup, 110

Crowfoot, 457

Water Parsnip, f. 650

Crenothrix palyspora, f. 166

Creosote, 6, 864

Oil, 330

Crepis lacera, 815

Crescentic cucurbitina, 810

Crujete, 810

Cress Garden, 113 593 593
cluteria, 588, 593, 595
muricatus, 822
Oil, 6, 72, 149, 588, 589
Seeds, 867
Texas, 121, 592, f. 592, 593
texensis, 121, 592, f. 592
Tiglium, 149, 588, f. 580,
592, 822
Verrauxii, 822
rotonilic acid. 7 Cow Cockle, 108 Crotonilic acid, 7 Crotonol, 588, 593 Crotonolglyccrid, Toxicity of, 279 Carauillo, 78 Emerus, 830 Crow ,W. E., 879 .
Crow ,W. E., 879 .
Crow Poison, 104
Crowberry, 604
Crowfoot, 4, 110, 457
Bulbous, 111, 457
Common, f. 110
Creeping, 457
Cursed, 77, 111, 457, f. 458
Ditch, 457
Family, 148, 446-468
Small-flowered, 458, f. 459
Tall, 458
Water, 446
sp., 446
Crowned Rust, f. 223, f. 225, juncea, 830 juncea, 830
montauna, 830
scorpioides, 524, 830
varia, 119, f. 523, 830
Coronillin, 148
Corozo Palm, 370
Corrosive sublimate, 6
Corrothers, Wm., 444
Cortaderia argentea, 826
conspicua, 826
Karmeiona, 926 Kermesiana, 826 Corticium amorphum, f. 232 Corticium anorphum, f. 2 Cortusa Matthiali, 844 Corybulbin, 147, 480 Corydalin, 147, 480 Carydalis, 564, 864 aurea, 113 aurea v. accidentalis, 564 cava, 147 Golden, 113 lutea, 480 racemosa, 825 tuberosa, 480 Tuherons-rooted, 480 Corylus americana, 403 sp., 440 Crowned Rust, f. 223, f. 225, f. 228, 228, 229 Crozier, A. A., 654, 879 Crucibulum vulgare, f. 246, 247, 860 CRUCIPERAE, 52, 53, 113-114, 485-495, 818 Cruel Plant, 600 Crustaceous Lichens, 307 Cryptocoryne spiralis, 808 Cryptogans, Poisonous, Corylus americana, 403 Avellana, 402 Corynocarpus laevigata, 53, 864 805 Cryptogramme crispa, 824 Corypha, 370 Cryptopin, 481 Cryptopin, 481
Cryptostegia grandiflora, 809
Cuba Bast, 623
Economic plants of, 864
Poisonous plants of, 867 umbracnlifera, 840 Cascinium, 89 Blumcannm, 838 Cosin, 505 Crescentia cucurbitina, 810 Cujete, 810 Cress, Garden, 113 Casmarium, f. 92, 189 Cosmos bipinnatus, 756, 815 Cuban Physic Nut, 596 Cubcbene, 78 Cubcbin, 396 Cubcbs, 396 Cujete, 810
Cress, Garden, 113
Penny, 114
Criminal Schools, 2
Crimson Clover, 68, 118, f.
556, 556-557
Crinum asiaticum, 805
zeylanicum, 805
zeylanicum, 805
Crithnum maritimum, 856
Crocin, 742
Crocodile Poison, 2
Cracasmia aurea, 389
Cracus, 109, 389, 446, 454
sativus, f. 387, 827
vernus, 389
Cronyn, J., 278
Crook, E., 879, 905
Crookshank, E. M., 879
Cross-vine, 135
Crotalaria, 524, 533, 535, 543546, 565, 864
alata, 830
juncea, 524 Cosmostigma racemosu, 809 Cosmostigma racemosu, 809 Costantin, J., 200, 202, 203, 878, 897 Costoxin, 595 Costus, 755 Cuckoo pint, f. 371
Cucumber, 750, f. 751
Bur, 135
Root, Indian, 104
Squirting, 749
Star, 751
Tree, 474
Wild, 751 Costus, 755
Catoneaster, 864
integerrina, 53, 503, 847
Cotton, 124, 622-623, 624-626,
f. 625
Crop, 625
Fiber, 623
"Frenching" of, 287
Gun, 623
Root, 625-626
Sea Island, 623-624
Seed, 124, 623, 625, 867
Meal, 124
Silk, 621
'Tree, 623, 624
Upland, 625-627
Cottonwood, f. 398 Wila, Cucumis, f. 751 africanus, 818 Anguria, 750 Anguria, 750 dipsacens, 818 Mela, 750, f. 751 metuliferus, 750, 818 myriacarpus, 750, 818 fraphetarum, 818 Sacleuxii, 819 sativus, 750 'Cottonwood, f. 398 juncea, 524 trigonus, 819

| C The test that a pri | | |
|--|--|--|
| | Cuathus strictus 1 216 242 | cabitatus 930 |
| Cucurbito foetidissima, 751 | Cyathus striotus, f. 246, 247, 860 | capitatus, 830 |
| moximo, 750, 819 moschoto, 751 | | formosissimus, 90 |
| Para 250 251 | vernicosus, 860 | hirsutus, 90, 830 |
| Pepo, 750, 751 | CYCADACEAE, 819 | Laburnum, 65, 85, 90, 148 |
| CUCURBITACEAE, 89, 135, 748, 749-751, 818-819 | CYCADALES, 325 | monspeliensis, 90 |
| | Cycas circinnalis, 325, 326, | nigricans, 90, 830 |
| Culture, affects toxicity of | 819 | polytrichus, 90 proliferus, 90, 534, 830 |
| plant, 87 | media, 325, 819 | proliferus, 90, 534, 830 |
| Culver's-physic, 733, 736 Culver's-root, 134, 733, 735 | Cyclamen, 52, 89, 128, 676 | Ruthenicus, 90 |
| Culver's-root, 134, 733, 735 | сигораент, 4, 128, 844 | scoparins, 90, 527, 551, 830 |
| Cumarin, 90, 523, 551, 552, | Furopean, 676 groecum, 844 | sessilifolins, 830 |
| 773, 803 | groecum, 844 | supinus, 830 |
| Cumin Oil, 648 Cuminum Cyminum, 648 | hederacjouum, 844 | Weldeni, 90 |
| Cuminani Cyminam, 648 | latifolium, 844 | Cytomycosis, 264 |
| sotivum, 648 | latifolium, 844 persicum, 128 | Cytomycosis, 264 Czapek, F., 543, 879 |
| sotivum, 648 Cuminol, 583 Cummin Seeds, 648 | Persian, f. 677 Cyclamin, 128, 676, 677 | * * * |
| Cummin Seeds, 648 | Cyclamin, 128, 676, 677 | D |
| Cupania, 53 | Cycloloma atriplicifolium, 424 | |
| Pseudorlius, 850 | CYCLOSPOREAE, 194 | DACRYOMYCETINEAE, |
| Cuphea, 637 | Cygnic acid, 533 | 233 |
| viscosa, 638 | Cygnin, 533 | Dactylis glomerato, 229 |
| viscosissima, 638 | | Daedalea quercina, 1. 232 |
| Cupraceus Laurenniana 222 | Cylindropuntia, 636 | Daemonorops Draco, 370 |
| Cupressus Lawsoniana, 327 | Cymbidium aloifolium, 840 | Daffodil, 386 |
| noolkatensis, 327 | Cymene, 648, 709 Cymol, 710 | Da Cama I A 970 |
| Constant Services | Cymol, 710 | Da Gama, J. A., 879 |
| thyoides, 327 Curare, 72, 687, 706, 788, 789, 803, 864 | Cynonclium acutum, 809 | Dahlia, 756 |
| 789, 803, 864 | sarcostenimoides, 809 | variabilis, 756 |
| Alkaloids, 148 | Vincetoxicum, 4 | Daisy, 776 |
| Plants, 148, f. 685 | Cynapin, 126, 659 Cynara Cordunculus, 756, 815 CYNAREAE, 757, 758 Cynoglossin, 706 | Garden, 756 |
| Curarie. See Curare | Cynara Cordunculus, 756, 815 | Ox-eye, 789 |
| Curarin, 73, 77, 687 | CYNAREAE, 757, 758 | Dalbergia, 864 |
| Curcas Oil, 78 | Cynoglossin, 706 | lanceoloria, 830 |
| Curarie. See Curare Curarin, 73, 77, 687 Curcas Oil, 78 Curcin, 595 | Cynoglossum, f. 68, 705, 706, | taxicorio, 830 |
| Curculigo scorzonerocfolia, | 864 | Dakin, R., 879 Dale, E., 278 |
| 805 | officinale, 131, 705-706, f. | Dale, E., 278 |
| Curcuma lencorhizo, 391 | 706, 811 | Damascenin, 446 |
| longa, 391 | Secds, 867 | Damen, C., 879 |
| Zedoraia, 392 | virginionum, 706 | Damiana, 864 |
| Curin, 148, 687 | virginicum, 131 | Damiana, 864 Daminar, 627 |
| Curled Dock, 419, f. 420 | Cyphomandra betacco, 716 | Dandelion 755, 756 f 750 |
| Curly Grass, Small, 315 | CYPERACEAE, 336, 367-369, | Root. 755 |
| Manle 615 | 819 | Root, 755 Daniell, F. W., 879 |
| Maple, 615 Currant, Black, 184, 498 | Cuparus 168 160 | Daniells, C. W., 879 |
| Crandall, 500 | Cyperus, 368, 369 articulatus, 819 | Danjon, Em., 747, 874, 879 |
| Indian, 744, 748 | Determination, 819 | Dapline, 52, 642-643 |
| Microusi 400 / too | Papyrus, 369 | cannabina, 642 |
| Missouri, 498, f. 500 | Cypress Plant, 130 | ('nggrum 642 |
| Red, 498 | Spurge, 121, 600, f. 601 | Cucorum, 642 Gnidinu, 855 |
| Septoria on, 284 | | |
| | Summer, 107 | I |
| Spot Disease of, 281 | Vine, 699 | Laurcola, 855 |
| Spot Disease of, 281 Tomato, 715 | Vine, 699 | Laurcola, 855 mezereum, 4, 125, 149, 642- |
| Curry Powder, 392, 527 | Vine, 699 Cypripedium, 105, 392-395, | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, | Vine, 699 Cypripedium, 105, 392-395, 864 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 | Laurcola, 855 mezereum, 4, 125, 149, 642- |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cuscus Grass, 345 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidum, 105, 393, 395 hirsutum, 393-395, f. 394 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, 1. 458 Cuscotoxin, 505 Cuscus Grass. 345 Cuscuta americana, 817 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsutum, 393-395, f. 394 macrauthum, 395 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cuscus Grass, 345 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidnm, 105, 393, 395 hirsulum, 393-395, f. 394 macranthum, 395 mantanum, 395 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cuscus Grass, 345 Cuscuta americana, 817 orvensis, 701 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnm, 105, 393, 395 hirsutum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 parviforum v. pubescens. | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta americana, 817 orvensis, 701 australis, 817 Dorycnium, 817 Epilinum, 701 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsulum, 393-395, f. 394 macrauthum, 395 mantanum, 395 parviforum r. pubescens, 392-393, f. 393 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis cestrifolia, 855 Cneorum, 855 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457. f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta americana, 817 orvensis, 701 australis, 817 Dorycnium, 817 Epilinum, 701 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsutum, 393-395, f. 394 macranthum, 395 mantanum, 395 parviflorum 7. pubescens, 392-393, f. 393 pubescens, 105, 395, 840 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, I. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum bancanum, 822 Daphnopsis cestrifolia, 855 Cneorum, 855 Gnidium, 855 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass, 345 Cuscuta antericana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epilinum, 701 Epilinum, 701 Epilinum, 130, 901, 817 evropea, 817 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidum, 105, 393, 395 hirsutum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 parviflorum v. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 oleoides, 855 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass, 345 Cuscuta antericana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epilinum, 701 Epilinum, 701 Epilinum, 130, 901, 817 evropea, 817 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsulum, 393-395, f. 394 macranthum, 395 parviflorum 7: pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum bancanum, 822 Daphnopsis cestrifolia, 855 Cneorum, 855 Gnidium, 855 Olcoides, 855 Darlington, Wm., 879 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass, 345 Cuscuta antericana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epilinum, 701 Epilinum, 701 Epilinum, 130, 901, 817 evropea, 817 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsulum, 393-395, f. 394 macranthum, 395 parviflorum 7: pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 | Laureola, 855 mezereum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cucorum, 855 Gidium, 855 oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass, 345 Cuscuta antericana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epilinum, 701 Epilinum, 701 Epilinum, 130, 901, 817 evropea, 817 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsulum, 393-395, f. 394 macranthum, 395 parviflorum 7: pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 Oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass, 345 Cuscuta antericana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epilinum, 701 Epilinum, 701 Epilinum, 130, 901, 817 evropea, 817 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidnu, 105, 393, 395 hirsulum, 393-395, f. 394 macrauthum, 395 parviflorum r. pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 | Laureola, 855 mezereum, 4, 125, 149, 642- 643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum bancanum, 822 Daphnopsis cestrifolia, 855 Cneorum, 855 Gnidium, 855 Gleides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361- |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta aniericana, 817 orvensis, 701 australis, 817 Dorycnium, 817 Epilinum, 701 Epilhymum, 130, 901, 817 eurofaco, 817 Cuscutin, 701 Cushaw, 751 Cuslny, R., 445, 879 Cusick, W. C., 465 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnun, 105, 393, 395 hirsutum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 pareiflorum 7, pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystitis, 172 | Laureola, 855 mezereum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cucorum, 855 Gindium, 855 oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, 1, 458 Cuscotoxin, 505 Cnscus Grass, 345 Cuscuta aniericana, 817 orvensis, 701 australis, 817 Dorycnium, 817 Epilinum, 701 Epiliymum, 130, 901, 817 europaeo, 817 Cuscutin, 701 Cushaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidnu, 105, 393, 395 hirsulum, 393-395, f. 394 macrauthum, 395 parviflorum r. pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 | Laureola, 855 mezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 Olcoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta aniericana, 817 orvensis, 701 australis, 817 Dorycnium, 817 Epithymum, 130, 901, 817 europaeo, 817 Cuscutin, 701 Cuschaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cusparia febrifuga, 582, 849 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsulum, 393-395, f. 394 macranthum, 395 parviflorum r. pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 Cyrtospermo, 53 laxioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 | Laureola, 855 meezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 825 Cneorum, 855 Cneorum, 855 Olcoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta aniericana, 817 orvensis, 701 australis, 817 Dorycnium, 817 Epilinum, 701 Epilhymum, 130, 901, 817 eurofaco, 817 Cuscutin, 701 Cuschin, 701 Cuschin, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cusparia febrifuga, 582, 849 toxicaria, 582-583, 849 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnm, 105, 393, 395 hirsntum, 393-395, f. 394 macranthum, 395 mantanum, 395 parviflorum r. pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 Cyrtospermo, 53 laxioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbi[ero, 320, 824 | Laureola, 855 mezereum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphninhyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cucorum, 855 Gudium, 855 Oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Dartre crouteuse, 16 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cuscus Grass, 345 Cuscuta antericana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epilinum, 701 Epilinum, 701 Epithymum, 130, 901, 817 europaco, 817 Cuscutin, 701 Cuschaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cusparia febrifuga, 582, 849 toxicaria, 582-583, 849 Cusparin, 582 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsulum, 393-395, f. 394 macrauthum, 395 partiflorum r. pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 Cyrtospermo, 53 latioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 | Laureola, 855 mezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis costrifolia, 855 Cneorum, 855 Gnidium, 855 Olcoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Dartre crouteuse, 16 Darwinia, 713 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta aniericana, 817 orvensis, 701 anistralis, 817 Dorycnium, 817 Epithumm, 701 Epithymum, 130, 901, 817 europaco, 817 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 705 Cuscohygrin, 875 Cuscohygrin, 575 Cuscohygrin, 575 Cuscaria, 582-583, 849 toxicaria, 582-583, 849 Cusparin, 582 Custard Apple, 110 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsulum, 393-395, f. 394 macrauthum, 395 partiflorum r. pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 Cyrtospermo, 53 latioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 | Laureola, 855 meezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 Gnidium, 855 olcoides, 855 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Neal, 363 Dartre crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, I. 458 Cuscotoxin, 505 Cuscus Grass, 345 Cuscuta americana, 817 orvensis, 701 australis, 817 Dorycnium, 817 Epilinum, 701 Epilinum, 701 Epilinum, 701 Cuscutin, 701 Cuscutin, 701 Cushaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 576 Cusohygrin, 576 C | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnun, 105, 393, 395 hirsutum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 pareiflorum r. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 montaua, 824 | Laureola, 855 mezereum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 Oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 493 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Datte crouteuse, 16 Darwinia, 713 Dassouville, Ch., 13, 15, 252, 879 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, 1, 458 Cuscotoxin, 505 Cuscotoxin, 505 Cuscus Grass. 345 Cuscuta americana, 817 orvensis, 701 australis, 817 Dorycnium, 817 Epithymum, 130, 901, 817 europaeo, 817 Cuscutin, 701 Cuscotoxin, 575 Cuscotoxin, 575 Cuscotoxin, 575 Cuscotoxin, 582-583, 849 toxicaria, 582-583, 849 Cusparia febrilaga, 582, 849 toxicaria, 582-583, 849 Cusparia, 582 Custard Apple, 110 Family, 476-477 Cut-leaved Water Parsnip, | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidum, 105, 393, 395 hirsutum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 parviflorum r. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 laxioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbi[cro, 320, 824 fragilis, 315, 320, 824 unontana, 824 Cystopus, 206 | Laureola, 855 meezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 Gnidium, 855 olcoides, 855 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Neal, 363 Dartre crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta aniericana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epithymum, 130, 901, 817 europaco, 817 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 705 Cuscutin, 705 Cuscutin, 705 Cuscutin, 706 Cuscutin, 707 Cuscutin, 708 Cuscutin, 708 Cuscutin, 708 Cuscutin, 708 Cuscutin, 709 Cuscutin, 709 Cuscutin, 709 Cuscutin, 709 Cuscutin, 709 Cuscutin, 709 Cuscutin, 709 Cuscutin, 709 Cuscutin, 701 Cuscutin, | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, f. 393, 395 candidum, 105, 393, 395 hirsulum, 393-395, f. 394 macrauthum, 395 parviflerum r. pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 montana, 824 Cystopus, 206 Cytisin, 77, 78, 79, 90, 113, | Laureola, 855 mezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis costrifolia, 855 Cneorum, 855 Gnidium, 855 oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Darte crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cuscus Grass, 345 Cuscuta americana, 817 Orvensis, 701 australis, 817 Dorvenium, 817 Epilinum, 701 Epilinum, 701 Epilinum, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 705 Cusck, W. C., 465 Cuschygrin, 575 Cusparia febrifuga, 582, 849 toxicaria, 582-583, 849 Cusparin, 582 Custard Apple, 110 Family, 476-477 Cut-leaved Water Parsnip, 76 Cutch, 530 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnm, 105, 393, 395 hirsntum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 pareiflorum 7, pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 montaua, 824 Cystopus, 206 Cytisin, 77, 78, 79, 90, 113, 148, 530, 537, 551, 803, | Laureola, 855 meezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 Gnidium, 855 Olcoides, 855 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Neal, 363 Dartre crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 Datisea cannobina, 628 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta aniericana, 817 orvensis, 701 anstralis, 817 Dorycnium, 817 Epithymum, 130, 901, 817 europaeo, 817 Cuscutin, 701 Cuschaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cuschygrin, 575 Cuscaria, 582-583, 849 toxicaria, 582-583, 849 Cusparin, 582 Custard Apple, 110 Family, 476-477 Cut-leaved Water Parsnip, 76 Cutch, 530 Cyamopsis tetragonolobo, 526 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnin, 105, 393, 395 hirsitum, 393-395, 1, 394 macranthum, 395 mantanim, 395 parviflorum r. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 laxioides, 53 Merkusti, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 montana, 824 Cystopus, 206 Cytisin, 77, 78, 79, 90, 113, 148, 530, 537, 551, 803, 864 | Laureola, 855 mezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 493 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Datte crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Datisca cannobina, 628 glomerota, 125, 628 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass, 345 Cuscuta aniericana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epithymum, 130, 901, 817 europaco, 817 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 705 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 705 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 702 Cuscutin, 702 Cuscutin, 703 Cuscutin, 703 Cuscutin, 703 Cuscutin, 706 Cutch, 530 Cyaniopsis tetragonolobo, 526 Cyanide of Potassium, 6 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnun, 105, 393, 395 hirsutum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 paraifforum r. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 montana, 824 Cystopus, 206 Cytisin, 77, 78, 79, 90, 113, 148, 530, 537, 551, 803, 864 Cystins, 535, 551, 864 | Laureola, 855 meezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 825 Cneorum, 855 Cneorum, 855 olcoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Dartre crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 Datisca cannobina, 628 glomerota, 125, 628 Datiscaceae, 125, 628 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass, 345 Cuscuta aniericana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epithymum, 130, 901, 817 europaco, 817 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 705 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 705 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 701 Cuscutin, 702 Cuscutin, 702 Cuscutin, 703 Cuscutin, 703 Cuscutin, 703 Cuscutin, 706 Cutch, 530 Cyaniopsis tetragonolobo, 526 Cyanide of Potassium, 6 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnm, 105, 393, 395 hirsntum, 393-395, 1, 394 macranthum, 395 mantanum, 395 parviflorum v. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystiis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbi[cro, 320, 824 fragilis, 315, 320, 824 unontana, 824 Cystopus, 206 Cytisin, 77, 78, 79, 90, 113, 864 Cytisns, 535, 551, 864 Adomi, 90 | Laureola, 855 mezereum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphninhyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cucorum, 855 Gindium, 855 Oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Datte crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 Datisca caunobina, 628 glomerota, 125, 628 Datiscaceae, 125, 628 Datiscaceae, 628 Datiscaceae |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cuscus Grass, 345 Cuscuta americana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epilinum, 701 Epilinum, 701 Curopaco, 817 Cuscutin, 701 Cushaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cusparia febrifuga, 582, 849 toxicaria, 582-583, 849 Cusparia, 582 Custard Apple, 110 Family, 476-477 Cut-leaved Water Parsnip, 76 Cutch, 530 Cyamides, 72, 73 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calccolum, f. 393, 395 candidnm, 105, 393, 395 hirsulum, 393-395, f. 394 macrauthum, 395 parviflorum r. pubescens, 392-393, f. 393 pubescens, 105, 395, 840 regina, f. 393 spectabile, 105, f. 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 montana, 824 Cystopus, 206 Cytisin, 77, 78, 79, 90, 113, 148, 530, 537, 551, 803, 864 Cytisns, 535, 551, 864 Adomi, 90 olpinus, 90 | Laureola, 855 mezereum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphninhyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cucorum, 855 Gindium, 855 Oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Datte crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 Datisca caunobina, 628 glomerota, 125, 628 Datiscaceae, 125, 628 Datiscaceae, 628 Datiscaceae |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta aniericana, 817 orvensis, 701 anistralis, 817 Dorycnium, 817 Epithymum, 130, 901, 817 europaco, 817 Cuscutin, 701 Cuschaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cuschygrin, 575 Cuschamin, 582-583, 849 toxicaria, 582-583, 849 Cusparia febrilinga, 582, 849 toxicaria, 582-583, 849 Cusparia, 582 Custard Apple, 110 Family, 476-477 Cutch, 530 Cyanidesis tetragonalobo, 526 Cyanides, 72, 73 Cyanogenesis, 53, 348, 864 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnm, 105, 393, 395 hirsntum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 mantanum, 395 pareiflorum 7, pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 montaua, 824 Cystopus, 206 Cytisin, 77, 78, 79, 90, 113, 148, 530, 537, 551, 803, 864 Cytisms, 535, 551, 864 Adami, 90 alpinus, 90 alschingeri, 90, 830 | Laureola, 855 mezereum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphninhyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cucorum, 855 Gindium, 855 Oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Datte crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 Datisca caunobina, 628 glomerota, 125, 628 Datiscaceae, 125, 628 Datiscaceae, 628 Datiscaceae |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass, 345 Cuscuta aniericana, 817 orvensis, 701 anistralis, 817 Dorycnium, 817 Epithumum, 701 Epithymum, 130, 901, 817 europaco, 817 Cuscutin, 701 Cushaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cusparia [cbrifuga, 582, 849 toxicaria, 582-583, 849 Cusparin, 582 Custard Apple, 110 Family, 476-477 Cut-leaved Water Parsnip, 76 Cutch, 530 Cyaniopsis tetragonalobo, 526 Cyanide of Potassium, 6 Cyanides, 72, 73 Cyanogenesis, 53, 348, 864 Cyanogenesis, 53, 348, 864 Cyanogenetic species, 315 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidum, 105, 393, 395 hirsntum, 393-395, 1, 394 macranthum, 395 mantanum, 395 parviflorum r. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 laxioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbi[cro, 320, 824 fragilis, 315, 320, 824 unontana, 824 Cystopus, 206 Cytisin, 77, 78, 79, 90, 113, 148, 530, 537, 551, 803, 864 Cytisns, 535, 551, 864 Adami, 90 alpinus, 90 Alschingeri, 90, 830 Attleanus, 90 | Laureola, 855 mezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cucorum, 855 Gidinm, 855 oleoides, 855 Darlington, Wm., 879 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Marte crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 Datisca caunobina, 628 glomerota, 125, 628 Datiscin, 628, 769 Datura, 88, 148, 717, 727, 729-733, 864 alba, 88, 133, 729 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cuscus Grass, 345 Cuscuta americana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epilinum, 701 Epilinum, 701 Epilinum, 751 Cuscutin, 751 Cuslaw, 751 Cuslaw, 751 Cuslaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cusparia febrifuga, 582, 849 toxicaria, 582-583, 849 Cusparin, 582 Custard Apple, 110 Family, 476-477 Cutch, 530 Cyamopsis tetragonalobo, 526 Cyanide of Potassium, 6 Cyanides, 72, 73 Cyanogenesis, 53, 348, 864 Cyanogenetic species, 315 Cyanoperical Cyanoperical Cyanoperical Securicans, 307 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnun, 105, 393, 395 hirsutum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 pareiflorum r. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 montana, 824 Cystopus, 206 Cytisn, 77, 78, 79, 90, 113, 148, 530, 537, 551, 803, 864 Cytisns, 535, 551, 864 Adami, 90 olpinus, 90 Alschingeri, 90, 830 Attleanus, 90 austriacus, 830 | Laureola, 855 mezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphninhyilum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 Oleoides, 855 Darlingtoni californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Dartre crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 Datisca cannobina, 628 glomerota, 125, 628 Datiscin, 628, 769 Datura, 88, 148, 717, 727, 729-733, 864 alba, 88, 133, 729 arborea, 853 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cnscus Grass. 345 Cuscuta antericana, 817 orvensis, 701 australis, 817 Dorycnium, 817 Epithum, 701 Epithymum, 130, 901, 817 europaeo, 817 Cuscutin, 701 Cushaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cusohygrin, 575 Cusparia febrifuga, 582, 849 toxicaria, 582-583, 849 Cusparia, 582 Custard Apple, 110 Family, 476-477 Cut-leaved Water Parsnip, 76 Cutch, 530 Cyanides, 72, 73 Cyanogenesis, 53, 348, 864 Cyanogenetic species, 315 Cyanopsis, 78, 241 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnm, 105, 393, 395 hirsntum, 393-395, 1, 394 macranthum, 395 mantanum, 395 parviflorum v. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystiis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbi[cro, 320, 824 fragilis, 315, 320, 824 montanua, 824 Cystopus, 206 Cytisin, 77, 78, 79, 90, 113, 148, 530, 537, 551, 803, 864 Cytisns, 535, 551, 864 Adomi, 90 alpinus, 90 Alschingeri, 90, 830 Attleanus, 90 anstriacus, 830 billorus, 90, 830 | Laureola, 855 meezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphniphyllum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 Gnidium, 855 Olcoides, 855 Darlingtonia californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Dartre crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 Datisca cannobina, 628 glomerota, 125, 628 DATISCACEAE, 125, 628 DATISCACEAE, 125, 628 DATISCACEAE, 125, 628 Daticin, 628, 769 Datura, 88, 148, 717, 727, 729-733, 864 alba, 88, 133, 729 arborea, 853 atrox, 729 |
| Curry Powder, 392, 527 Cursed Crowfoot, 77, 110, 457, f. 458 Cuscotoxin, 505 Cuscus Grass, 345 Cuscuta americana, 817 orvensis, 701 australis, 817 Dorycuium, 817 Epilinum, 701 Epilinum, 701 Epilinum, 751 Cuscutin, 751 Cuslaw, 751 Cuslaw, 751 Cuslaw, 751 Cusliny, R., 445, 879 Cusick, W. C., 465 Cusohygrin, 575 Cusparia febrifuga, 582, 849 toxicaria, 582-583, 849 Cusparin, 582 Custard Apple, 110 Family, 476-477 Cutch, 530 Cyamopsis tetragonalobo, 526 Cyanide of Potassium, 6 Cyanides, 72, 73 Cyanogenesis, 53, 348, 864 Cyanogenetic species, 315 Cyanoperical Cyanoperical Cyanoperical Securicans, 307 | Vine, 699 Cypripedium, 105, 392-395, 864 acoule, 395 Calceolum, 1, 393, 395 candidnun, 105, 393, 395 hirsutum, 393-395, 1, 394 macrauthum, 395 mantanum, 395 pareiflorum r. pubescens, 392-393, 1, 393 pubescens, 105, 395, 840 regina, 1, 393 spectabile, 105, 1, 393, 840 Cyrtospermo, 53 lasioides, 53 Merkusii, 808 Cystitis, 172 Cystopteris, 315, 319-320 alpina, 824 bulbifero, 320, 824 fragilis, 315, 320, 824 montana, 824 Cystopus, 206 Cytisn, 77, 78, 79, 90, 113, 148, 530, 537, 551, 803, 864 Cytisns, 535, 551, 864 Adami, 90 olpinus, 90 Alschingeri, 90, 830 Attleanus, 90 austriacus, 830 | Laureola, 855 mezerum, 4, 125, 149, 642-643, f. 643, 855 pontico, 65 striata, 855 Daphnetin, 125, 642 Daphnin, 125, 642 Daphnin, 125, 642 Daphninhyilum baucanum, 822 Daphnopsis ccstrifolia, 855 Cneorum, 855 Gnidium, 855 Oleoides, 855 Darlingtoni californica, 497, f. 498 Darnel, 4, 72, 74, 103, 361-364, 864 Poison, 361-364, f. 362, 363 Meal, 363 Dartre crouteuse, 16 Darwinia, 713 Dassonville, Ch., 13, 15, 252, 879 Date Palm, 370 Datisca cannobina, 628 glomerota, 125, 628 Datiscin, 628, 769 Datura, 88, 148, 717, 727, 729-733, 864 alba, 88, 133, 729 arborea, 853 |

nudicaule, 109 occidentale, 463 Penardi, 460-461, f. 461, 467 peregrinum, 845 recurvatum, 109 ferox, 853 Metel, 64, 715, 729, 733, 853 Meteloides, 715, 729, 732-733, 853 sanguinea, 853 Stramonium, 4, 60, f. 61, 74, 88, 133, f. 730, 732. 853 Requieni, 845 Scopulorum, 109, 463, 464, 845 853
Suaveolens, 729, 853
Tatula, 133, 729-732, 853
Wrightis, 64, 133
Wrights, 133, 732-733
Daturin, 730
Daucus, 650, 663-64
Carota, f. 68, 127, 647, 663-664, 856
Dongherty, C. M., 879
Davidson, 879
Davies, D. H., 879
Davillia brasilieusis, 315, 824
elegans, 824 simplex, 462 staplusagria, 108, 446, 448, 460, 463, 846 tricorne f. 43, 108, 463, 846 trolliifolium, 109, 463, f. 464, 846 uncinatum, 846 Delphinoidin, 108, 464 Delphinosis, 43, 44-45, 47, 79, 463 Delphisin, 108, 464 Deming, Dr., 244 Demosthenes, 2 Dendrobium nobile, 840 simplex, 462 Dendrobium nobile, 840 Denmark, Poisonous plants of, 867 elegans, 824 elegantissima, 315 elegantissima, 315
litrsuta, 825
majuscula, 825
pentaphylla, 315, 825
platyphylla, 825
Strigosa, 825
Davenport, C. B., 879, 901
Davilla rugosa, 819
Davil, B. M., 169, 209
Davy, G. Burtt, 133, 696
Day, Dr. C. M., 140, 795
Day, Dr. C. M., 140, 795
Day, Dr. C. M., 564, 879
Day, Lily, 375
De Bary, A. See Bary, A. de
De Caudolle, A. See Caudolle, A. See Caudolle, A. See Caudolle, A. See Puy, C.
E. de
De Nabias. See Nahias, De
De Puy, C. E. See Puy, C.
Le de
Dead Nettle, 131, 712, f. 712 Denmark, Poisonous plants of, 867
Depre, 290
Depressant poisoning, 74-75
Deptford Pink, f. 437
Dermal Mycosis, 864
Dermatitis, 12-19, 78, 79, 104, 106, 120, 121, 122, 126, 131, 371, 393, 510, 584, 588, 704, 864
Acute, 464, 583, 663, 755
Necrotic, 179
Dermatomycosis, 12, 80, 295, Dermatomycosis, 12, 80, 295, Dermatomycosis, 12, 80, 295, 864 Dermatophytes, 13 Derosne, 3, 143 Derris, 1, 52, 864 amoena, 830 amoena, 830 elliptica, 830 guianesis, 830 Negrensis, 830 nliginosa, 830 Desaint, 534, 880, 900 Desenne, 299 Desmids, f. 92 DESMIDIAGEAF, 189 Dead Nettle, 131, 712, f. 712 Deadly Agaric, 238-239, 240 Amanita, 34. See A. phalloides phalloides
Nightshade, 133
Death Camas, 103, 375, 376, 377-378, 466
Cup, 238-243, f. 239
Decringia celosioides, 805
Defleroix, G., 880, 906
Delafoy 686
Delite A. R. 880
Delite A. R. 880 375, *. DESMIDIACEAE, 189 Desmodium tortuosum, 525 triflorum, 525 Despranches, 572 Detarium senegalense, 830
Deutzia, 115
scabra, 500
staminea, 115, 500, 851
De Vecchj, Count Achilles Deliriant poisoning 73-74, 803 Delirium a symptom of poisoning, 72 237
Devergie, A., 301
Devil's Apron, 194
Dewberry, 505
Dewey, I. II., 430, 625
Deyeuxia Langsdorfii, 826
Dhourin, 54, 348
Diamins, 10, 73, 149
Diantlus, 89 oning, 72
Delotte, 880
Delphinin, 108, 446, 464
Delphinium, 446, 448, 460-467
.ijacis, 460, 845
.azureum, 460
Barbeyi, 467
bicolor, 6, f. 461, 462-463,
466 Honorianum, 845
caerulcum, 845
camporum, 84
Consolida, 84, 108, 446, 460, 463, 464
decorum v. nevadense, 465
elatum, 460 Dianella neinorosa, 834
Diantlus, 89
barbatus, 436
caryophyllus, 439
chineusis, 813
Diaphoretic, 803
Diarrhoea, 78
Diatoms, 92, 188-189, f. 189
Diceutra, 864
spectabilis, 480, 825
DICHOPETALACEAE, 53
Dichopsis Gutta, 680, f. 680, 681
Dichora febrifuga, 851 elatum, 460 elongatum, 467 exaltatum, 108 Gcycri, 109, 461, 467, 845 glaucum, 6, f. 47, 84, f. 464, 466, 467 grandiflorum, 108 Menziesii, 44, 97, 109, 461-462, 466, 845 mauritiana, 845 Nelsoni, 467 Dichroa febrifuga, 851 Dicksonia, 313 DICOTYLEDONEAE, 155, 395, 802 105-Dictamnus albus, 581

DICTYOTACEAE, 194
DICTYOTALES, 154, 194
Dictyuchus clavatus, 208
Dicypellium caryophyllatum, Dicypellium caryophyllatum,
478
Diedra, 299
Diervilla japonica, 135, 812
Lonicera, 135
trifida, 812
Dietrich, D. N. F., 862, 880
Dieudülin, J., 880
Dieulafoy, 263
Digitalein, 736
Digitalein, 736
Digitalin, 78, 130, 134, 144, 148, 385, 441, 534, 692, 736, 737
Digitalis, 6, 7, 52, 72, 73, 77, 80, 87, 89, 637, 633, 692, 694, 734, 736-737.
864
ambigua, 852 864
ambigua, 852
lutea, 852
purpurea, 4, 75, 134, 148,
441, 733, 736-737, f.
737, 852
Digitogenin, 148
Digitonin, 134, 148, 736
Digitonin, 134, 148, 736
Digitonin, 134, 148, 736
Digitonin, 134, 148, 736
Digitonin, 134, 377, 736
Dill, 648
Oil, 648
DILLENIACEAE, 52, 628, 819
Dimorphotheca Ecklonis, 815
pluvialis, 815
Dinsmore, S. C., 880, 894
Dinwiddie, R. R., 290, 880
Diodia, 90
Dionaea, f. 420 ambigua, 852 Dionaea, f. 429 muscipula, 498, f. 499, 820 Dioon, 326 Dioscorea, 89, 105, 374, 864 aculeata, 374 alata, 374, 819 bulbifera, 374, 819 deltoidea, 819 divaricata, 374 hirsuta, 820 japonica, 374 macroura, 82 820 macroura, 820 sativa, 374 toxicaria, 820 triphylla, 820 villosa, 374, f. 374 DIOSCORRACEAE, 89, 105, 374. 819-820 Dioscorides, P., 2, 601, 880 Dioscorin, 864
Diosmin, 583
Diospyros, 52, 681
acris, 820
amara, 820
Cargillia, 681
chloroxylon, 681
discolor, 681
Ebenum, 681, 820
embryopteris, 681, 820
Kaki, 681
malaccapai, 820 Dioscorin, 864 malaccapai, 820 melanoxylon, 681 moutana, 820 multiflora, 820 obtusiflora, 820 pentamera, 681 quesita, 68 rubra, 681 ruora, 631 samoensis, 820 tesselaria, 820 toxicaria, 820 zirginiana, 128, 681, 820 Dipholis salicifolia, 680

| Diptheria, 179 | Dracunculus 2 |
|--|--|
| Diptheria, 179 Avian, 179 | Dragendorff, (|
| Diphtherial poison, 864 Diplococcus, intracellulus | Dragon-tree, 3 |
| Diplococcus, intracellulus | Dragon-tree, 3 Dragon's Bloo Dragon's Head |
| meningitidis, 169 | Drabernaldia |
| Diblodia, 77, 286, 861, 864 | Drapernaldia, Drimia ciliari, |
| Zeae, 100, f. 286, 286-287, | Cowanii, 83 |
| of Neisser, 170 Diplodia, 77, 286, 861, 864 Zeae, 100, f. 286, 286-287, f. 287 Diplodiella, 286 Diplodia, 286 | Drimvs aroma |
| Diplodiella, 286 | Winteri, 47 |
| Diplodiella, 286 Diplodina, 286 Diplotaxis erucoides, 818 | Drosera, 497 binata, 498, communis, 8 |
| tenuifolia, 818 | communic. |
| Debadium bunctatum 811 | intermedia, |
| Dipper Gourd, 751 | peltata, 820 |
| DIPSACEAE, 741, 820 | Pigments in |
| Dipper Gourd, 751 Dipsaceae, 741, 820 Dipsacus fullonum, 741 sylvestris, f. 741 | rotundifolia, |
| DIRTEROCARRACTAE 627 820 | Whittakerii, Droseraceae, |
| Dibterocarbus, 627 | Drosophyllum |
| Dipteryx, 90, 523 | 498, 82 |
| DIPTEROCARPACEAE, 627, 820 Dipterocarpus, 627 Dipteryx, 90, 523 odorata, 523 Dirca, 643 | 498, 82 Drug plants, 3 Drumin, 588 |
| Dirca, 643 | Drumin, 588 |
| palustris, 125, 642, 643, f. 644, 855 | Drummond, 5. |
| Discaria serratifolia, 847 | Dry-rot Fungi |
| Disinchonin, 742 | Duboisia, 148, |
| Disophenol, 583 | Hopwoodii, |
| Disinchonin, 742 Disophenol, 583 Dita Bark, 691-692 Ditamin, 692 | Leicharain, |
| Ditamin, 692 | Myoporoide. |
| Ditamin, 692 Ditch Crowfoot, 457 Dittelasma Rarak, 850 | 853 Duboisin, 716 |
| Diuretic, 803 | Duchesnea ind |
| Dock, Curled, 419, f. 420, 421 | Duckweeds, 32 Duckweeds, 32 Duges, A., 34 |
| Great Water, 421 | Duges, A., 34 |
| Pale, 419, 420-421 | Duckweeds, 33 Duges, A., 34 Duisberg, Po of, 867 |
| Dodecatheny Meadia 676 | Dulcamara, 7 |
| Dodder, 130, 701 | Dulcamarin, 6 |
| Alfalfa, 130, 701 | Dunbar, 755, |
| Clover, 130, 701 | Dugern, E. \ |
| Ditteidsma Karak, 850 Ditteitic, 803 Dock, Curled, 419, f. 420, 421 Great Water, 421 Pale, 419, 420-421 Sour, 106, f. 430 Dodecatheon Meadia, 676 Dodder, 130, 701 Alfalfa, 130, 701 Clover, 130, 701 Exotic, 130 Flax 701 | Dunbar, 755, Dugern, E. V Dunglison, R. Dunglison, R. Dunning, H. Dunstan, W. |
| Dodonaea. 52, 89 | |
| Dodonaea, 52, 89 physocarpa, 850 | Dunstan, W. 450, 53 |
| paysocarpa, 850 viscosa, 850 Doeltz, J. C., 880 Doerr, Dr. John, 725 Dog Fennel, f. 139, f. 785, 787-788 Dog's-Tooth Violet, 104 | 430, 33 |
| Doeltz, J. C., 880 | 890, 914 |
| Dog Fennel, f. 139, f. 785, | Dupony, 881 Duranta Plun |
| 787-788 | Duris zihethin |
| | Durra, 345, 3 Dusty Miller, Dutchman's F Duvernoy, G. Dwarf, 665 |
| Dogbane, 692-693 | Dusty Miller, |
| Common, 148 Family, 129-130, 148, 691- | Dutchman's I |
| 695 | Dwarf, 665 |
| Spreading, 129, 692-693 Dogwood, 127, 609, 664 Family, 127, 664 | Blueberry, |
| Dogwood, 127, 609, 664 | Blueberry, 62 Mallow, 62 |
| Family, 127, 664 | Dyer's Broom Dyer's Weed, Dyer's-woad, |
| Flowering, 664 Jamaica, 74 | Dyer's weed, |
| Panicled, 664 | Dyes, among l |
| Round-leaved, 664 | Dysodia, 758, |
| Western, 664 | chrysanthen |
| Dohme, A. R. L., 88, 880, | papposa, 78 Dysoxylum a Dyspagia, 25 |
| Doir, 572 | Dysoxylum a |
| Dolichandrona falcata 910 | Dyspnoea, 55 |
| Dolichos bistorus, 526 | |
| Dolichos bistorus, 526 Lablab, 53, 521 Doliocarpus Rolandri, 819 Door-yard Knotweed, 423 | |
| Donocarpus Rolandri, 819 | |
| | Eames, E. H. |
| Doronicum pardalianches, 815 | Earth-star, 24 |
| Dorpat, Laboratory at, 62 | East Tennesse |
| Dotten, S. B., 880, 894 | Eastern Colu |
| Doronicum pardalianches, 815 Dorpat, Laboratory at, 62 Doten, S. B., 880, 894 DOTHIDEACEAE, 279-280 DOTHIDEALES, 279-280 DOUGLES, Fig. 327 | Easterfield, 6 |
| Douglas Fir, 327 | ERENACELE E |
| Dow, Dr., 25 | Easterfield, 6 Eastwood, A. EBENACEAE, 5 681, 82 |
| Dracaena arborea, 834 | EBENALES, |
| Dracontium asperum, 808 | Ebony, 681 |
| Dracontomelon mangiferum, | Black, 681 |
| 608 | Ceylon, 68 |
| | |

us vulgaris, 808 ff, G., 4, 894 ee, 375 Blood, 370 Head, 103, 372 dia, f. 92 liaris, 834 , 834 omatica, 837 474, 837 197 198, 820 is, 820 lia, 498, 820 820 820 olia, 498, f. 500, 820 erii, 820 AE, 497-498, 820 lum lusitanicum, 820 ts, Sources of, 87 1, 533 ungus, f. 232, 234 ungus, f. 232, 234 ops camphora, 820 148, 864 dii, 147, 716, 853 dtii, 716 oides, 716, f. 717, indica, 505, 509 s, 372 , 342 Poisonous plants 867 , 718 a, 718 iin, 62 755, 767 E. V., 880 i, R., 881 i, R. J., 881 H. A. B., 881, 892 W. R., 54, 88, 348, i, 534, 754, 873, 881, Plumieri, 858 thinus, 621 5, 346 ller, 756 l's Pipe, 417, f. 4 , G. D., 572, 881 oom, 524-525, f. 525 eed, 479 eed, 479 ad, 486 ng legumes. 524-525 758, 784-785 themoides, f. 139 , 785, f. 785, 815 n. arborescens, 837 55 E H., 881, 885 245 essee Pink-root, 688 d, 607, A., 881, E, 52, 128, 679, 679, 820 ES, 158, 679-683

681

Family, 128, 681 Green, 681 Indian, 681 Red, 681 Striped, 681 White, 681 Ecballium Elaterium, 749, 819 officinale, 749 Ecbolin, 149, 277 Eccles, R. G., 881 Ecgonin, 144 Echinocactus, 635, 811 Ottonis, 635 texensis, 635 Williamsii, 811 Wislizeni, 635 Wislizeni, 635
Echinocarpus, 53
Sigun, 622
Echinocereus stramineus, 635
Echinocystis, 89
fabacea, 819
lobata, 751
macrocarpa, 751, 819
Echinops Ritro, 815
Echitamin, 692
Echitamin, 692 Echitemin, 692 Echites, 864 alexacaca, 807 macniata, 807 venenosa, 807 Echinm, 707 Echium, 707
vulgare, 131, 228, 707, 1.
708, 811
Economic plants, Among Legumes, 520-527
In America, 864
Australia. 864
Cuba, 864
Cuba, 864
Cuam, 684
Japan, 864
United States, 864
Eczema, 301 Eczema, 301 marginatum, 300, 301 Eden, R. T., 881 Edgeworthia Gardneri, 855 Edward, 684 Edward, 684
Fel Grass, Fresh-water, 332
Effront, J., 486
Egg Plant, 714, 718
Egyptian Lotus, 445
Wheat, 345
Eichornia speciosa, 374
Eidamella, 13
spinosa, 12, 13 252
Eisendrath, 306
ELAEAGNACEAE, 125, 640-641
Elaeagenus apprustifolia, 641 640-642 ELARGNACEAR, 125. 640-6: Elaeagnus angustifolia, 641 hortensis, 641 uniltiflora, 641, 6, 641 orientalis, 641 Elaeis guineensis, 370 ELAEOCARPACEAR, 622 Elagagaga varrusan 822 Elaeococca verrucosa, 822 Elaeodendron australe, 614 orientale, 813 Elaeoselinum asclepium, 856 foetidum, 853
Elaphomyces granulatus, f. 273 Elaterin, 749 Elateriospermum Tapos, 53 Elaterium, 749
Elborne, Wm., 881
Elder, 135, 744, 745-747
American, 747 Marsh, 137, 763-764, 767

Marsh, 137, 763-764, 767

Small-flowered, 763 Rcd-herried, 135 Elephantopus tomentosus, 815 Elettaria cardamomum, 391

644

Epipactis pubescens, 692 Epiphyllum truncatum, 635 Epipremum mirabile, 808 Equisetaceae, 101, 323-325, Elionurns, 826 Ellertonia Rheedi, 807 Elm, 405, 406-408 Elm, 405, 406-408 American, 406-408, f. 407 Rock, 408 Scotch, 408 Slippery, 408, f. 409 Elodica conadensis, 332 Elphinck, E., 881 Elpmrs, 90, 826 canadensis, 276 robustus, 276 EQUISETACEAE, 101, 323-323, 825 EQUISETALES, 322 Equisetosis, 20, 37, 38, 80, 323, 864 Equisetum, 323-325, 864 arvensee, 37, f. 38, 101, 323, 820 hyemale, 323, 820 hyemale v. robustum, 101, 323 robustus, 276 striatus, 276 virginicus, 276 virginicus v. submuticus, palustris, 820 Poisoning, 324-325 Eragrostis major, f. 339 megastachya, 692 Elytropappus glandulosus, 815
Einbelia micrantha, 839
Ribes, 839
Embelic acid, 679
EMBRYOPHYTA, 100, 101, 154, 325-802
Emery, Z. P., Dr., 560
Emetic, 804
Drastic, 804
Emetin, 78, 633, 742
Emodin, 377
EMPETRACE, 604
Empetrnm nigrum, 604 megastachya, 692
Franthis hyemalis, 846
Frbe, 291
Frdmann, K. G., 882
Frect Wake-robin, 104
Fremoscus, 252
Fremostarpus settigerus, 822
Fremophila maculata, 838
Fremostachys, 52 Eremocarpus setigerus, 8.22
Eremophila maculata, 838
Eremostachys, 52
superba, 828
Ergochrysin, 277
Ergot, 2. 6. 28. 31, 72. 79.
275-279, 546, 864
Chemical composition f. 28. 277-278
Toxicology of, 278-279
Ergot on Agrostis alba, 216
Blue Grass, f. 27, 276
Blue Joint, f. 27
Bottle Grass, f. 27
Calamagrostis, 276
Darnel, 364
Glyceria, 276
Koeleria cristato, f. 27
Manna Grass, f. 27
Onack Grass, 363
Reed Canary Grass, f. 27
Timothy, f. 27, 276
Western Wheat Grass, 365
Wild Rice, 276
Wild Rye, f. 27, 149, 546
Ergotin, 277-278
Ergotinic acid, 7, 28, 277
Ergotinic acid, 7, 28, 277
Ergotinic, 277, 278 EMPETRACEAC, 604
Empetram nigrum, 604
Empasa, 195
Grylli, 204
Mnscoc, f. 203, 204
sphaerosperma, f. 203, 204
Enulsin, 83, 505, 517
Encephalitis, 290
Endive, 756, 758
Endocarditis, 169
Ulcerative, 167, 171
Metastatic, 177
Endoconidium temulentum Endoconidium temulentum, 364 Endometritis, 171 ENDOPHYLLACEAE, 221 Endo-toxin, 10 Engelhardt, II., 88, 880, 831 England, Poisonous plants or, 867 Engler, A., 882 English, Charlock, 488, f. Ergotin, 277-278
Ergotinic acid, 7, 28, 277
Ergotinic, 277, 278
Ergotism, 28-29, 31, 77, 100 276-279, 864
Epidemics of, 276-277
Effects of, f. 278
Nervous, 279
Gangrenous, 28
Soasmodic, 28
Ergotoxin, 278
Eria stelluta, 840 489 Cherry, 5 Oak, 403 Oak, 403

Walnut, 401

Entada, 89

polystachys, 830

scandeus, 63, 525, 751, 839

Enteritis, 105, 171, 804

Haemorrhagic, 490

Enterolobium, 1, 63, 89

cvelocarpum, 830

Timboiru, 830

Futoloma, 864 77, 80 Eseridin, 528 Eria stellata, 840 Erico, 666 Entoloma, 864 clypeatum, 859 groveoleus, 859 ERICALES, 52, 64, 127-128 664-675, 820-821, 864 ERICALES, 157, 664-675 127-128, ENTOMOPHTHORACEAE, f. 202, 204 Ericinol, 666 Entyloma ronunculi, 219 Environment. Toxicity plant dne to, 83 Enzootic Cercbritis, 20 Enzymes, 77 Ericolin, 127, 665, 666, 669 Erigorou, 757, 776-779, 865 annus, 138, 777, 778, f. 830 animis, 50., 778, 778, canodensis, 138, 755, 777-778, f. 777 divaricatus, f. 777 philadelphicus, 777 ramosus, 138, 778 Eperna falenta, 830 Ephedru, 326 autisyphilitico, 825 nevodense, f. 326 sp., 825 Ephedrin, 77 ramosus, 138, Eriksson, J., 882 Eriobotrya, 865 japonica, 53 Epichloe typhina, 273 Eriocaulon septangulare, 372 Eriocolin, 703 Epigaeo, 666 repens, 127, 665, 821 Epilobium angustifolium, 629, Eriodendron anfractuosum, 621, 624

Eriodictyon crassifolium, 703 glutinosum, 827 Friodictyonic acid, 703 Eriogomm umbellatum, 419 Eritrichium gnaphaloides, 629-Frnst, A. 52, 803, 804, 882 Frodium, 578-579 Erndium, 578-579 cicutarium, 67, 120, 578, f. moschutum, 120, 578, 579, moschutum, 120, 578, 5 f. 579 Musk, f. 579 Eruca şativa, 818 Eryngium maritimum, 648 ynccaefolium, 648, 692 Erkystbaceae, 268-273 Erysimum, 486 asperum, 486 cheirauthoides, 818 Erysipelas, 171 Erysipelas, 171
Erysipelas, 171
Erysiphe, 269
("choraccarum, 269
communis, 269, 272
graudiis, f. 270, f. 271, 272273 Erythema, 421, 611, 731 Scarlatiniform, 78 Erythrasma, 298
Erythrina Corollodendron, 527, 830
Hypaphanes, 830 Mulungn, 830 Erythrocentaurin, 689 Erythronium albidnm, 834 americanum, 834 Deus-canis, 834 grandiflorum, 834 purpurascens, 104, 834 Erythrophloein, 148, 534 Erythrophloenn, 1, 865 Comminga, 830 gnineense, 534, 830 Erythroxylaceae, 575, 821 Erythrroxylon Coca, 147, 575, f. 576, 821, 835 Escallonia myrtilloides, 851 Escharotic, 805
Eschaltzia, 865
californica, 479, f. 480, 841
Douglasii, 113
Escriu, 82, 531, 729, 804 Ester, 500 Ester, 500 Ether, 73 EUASCALES, 248-281 EUASCI, 248-281 EUBASIDH, 209, 220-247 Eucalyptol, 640 Encalypts, 640 Encalyptus Globulus, 640, 839 microtheca, 839 Oil of, 640 rostvata, 640, 839 Euchlaena mexicana, 342 Enchresta Horsfieldii, 90, 551, Fudoriaa, 93, 193 liugenia caryophyllata, f. 638, 639 Chequen, 640 Jambos, 839 Eugenin, 639 Engenol, 392, 478, 583 liuglena viridis, 188 EUGLENACEAE, 188 EUGLENALES, 188 Eulophia virens, 840 EUMYCETES, 98-100, 154, 195-308 Enonymin, 148, 615

| Enomymus, 615 | resinifero, 121, 588, 601, | betulinus, 253 |
|--|--|--|
| atropurpurens, 123, 148, | 823 | Cerosi 253 |
| | | communis, f. 253 deformons, 253 Pruni, 252, f. 252 EXOBASIDIINEAE, 233 |
| 614, f. 615, 813 | Royleona, 823 | deformance 253 |
| europaens, 813 latifolius, 813 | Sibthorpii, 823 | aejormons, 255 |
| latifolius, 813 | Sicholdiono, 823 | Pruni, 252, 1. 252 |
| Function 772 | splendens, 588 | EXOBASIDIINEAE, 233 |
| Euparin, 772 | thymifolio, 823 | Exocorpus cupressiformis, 850 |
| EUPATORIEAE, 757 | Time calli 923 | Exotic Blue-flag, 389 |
| Eupatorin, 138, 755, 772 | Tirucolli, 823 | |
| Eupotorium, 90, 757, 770-772, | venefico, 823 | Dodder, 130 |
| | verrucoso, 823 | Expectorant, 804 |
| 865 | verrucoso, 823 Wulfenii, 823 | Extra-cellular Toxin, 10 |
| altissimum, f. 763 | EURHOPRIACEAE 52 53 89. | |
| amorissimum, 815 connabinum, 138, 815 | EUPHORBIACEAE, 52, 53, 89, 121-122, 575, 577, 586- | F |
| counabinum, 138, 815 | 121-122, 3/3, 3/7, 300- | · |
| capillifolium 770 | 604, 821-824, 865 | E-L T M 002 |
| copillifolium, 770 | Euphorbon, 121, 588, 602 | Faber, J. M., 882 |
| perfoliotum, 138, 755, 771, | Euphorbon, 121, 588, 602 EUPHYCEAE, 98, 153, 188- | Fobiano imbricoto, 716 |
| 772 | 104 950 | Fadong, 326 |
| purpureum, 771, 772 | 194, 859 Europe, Poisonous plants of, | FAGACEAE, 105, 824 FAGALES, 156, 402, 404 |
| serotinum, 771 | Europe, Foisonous plants of, | EACATES 156 402 404 |
| urticoefolium, 138, f. 770. | 867 | Facouriem 90 |
| | European Aconite, 46, 109, | Fagopyrism, 80 |
| 771-772 | 450, 452, 453 | Fagopyrismus, 12 |
| Enphorbio, 52, 65, 588, 589, 590, 597, 603 | Anemone, f. 454 | Fogopyrum, 419.420 |
| 590, 597, 603 | Anemone, f. 454 Aristolochia, f. 418 | esculentum, 106, 419, 843 totoricum, 106, 419-420 Fograeo ceilonico, 836 |
| oleppico, 822 | A -4'-1-1- 756 | totoricum, 106, 419-420 |
| alsinaefloro, 589, 822 | Artichoke, 756 | Fourges coilouico 836 |
| | Arum, f. 371 Beech, 403 | Farmer Communico, 650 |
| amygdoloides, 822 antiquorum, 589, 822 | Beech, 403 | Fogus ferrugineo, 403 |
| antiquorum, 589, 822 | Bindweed, 702 | sylvotico, 403, 824 Fairchild, Dr. D. S., 544 |
| arboreo, 822 bicolor, 822 | Chestnut 403 | Fairchild, Dr. D. S., 544 |
| hicolor 822 | Chestnut, 403 | Falck, C. P., 4 Falck, F. A., 882 |
| canariancie 580 | Columbine, 446, f. 447 | Palale T A 992 |
| canariensis, 589 | Cyclamen, 676 | Faick, F. A., 002 |
| condclobrum, 822 | Grape, 620 | Falk, 688 |
| carocosono, 822 | Heliotrope, 705 | False Acacia, f. 559, 559.560 |
| cercbrnm, 822 | | Aloe, f. 384 |
| Chamaesyce, 822 | Holly, 614 | Columba Root, 473 |
| Clauries for 022 | Horse-chestnut, 617 | |
| Charocios, 589, 822 | Licorice, 527, f. 528 | Flax, 493, f. 404 |
| corolloto, 121, J. 122, 398, T. | Lupine, 118 | Hellebore, 103, 375 |
| 598, 601, 602, 822 | | Tasmine, 129 |
| cotinoides, 822 | White, 547 | Jasmine, 129 Mallow, 626 |
| antinfalla 022 | Mistletoe, 106 | Mailow, 020 |
| cotinfolia, 822 | Mountain Ash, 629 | Mistietoe, 100 |
| (yporissios, 4, 121, 122, | Oak, 105 | Mistletoe, 106 Ragweed, f. 764 Truffle, 247 |
| 589, 600, 601, f. 601, | Pennergrass 486-493 | Truffle, 247 |
| 822 | Peppergrass, 486-493 Plum, 500, 514 | Famulener, L. W., 882 |
| dendroides, 822 | Fidili, 300, 314 | Farcinica organism, 184 |
| | Prickly Lettuce, 756, 760 | Tarenica organism, 101 |
| Drummondii, 11, 588, 589, | 761, f. 761 | FARINOSAE, 155, 372-374 |
| 822 | Saint John's wort, 631 | FARINOSAE, 155, 3/2-3/4 |
| cremophilo, 589, 822 | Sandwort, 436 | Farkleberry, 665 Farlow, W. G., 91, 96, 186, 205, 217, 236, 238, 239, 245, 803, 863, 882 Farr, E. H., 882, 917 Fotsio horrido, 125, 647 |
| Esnlo, 589, 822 | | Farlow, W. G., 91, 96, 186, |
| criano 580 | Strawberry, 115 | 205 217 236 238 239 |
| exigno, 589 Gerordiana, 822 | Sumac, 608 | 245 002 063 002 |
| Geroraiana, 822 | Vetch, 87 | 243, 803, 803, 802 |
| Helioscopio, 589, 822 | Violet, 83 | Farr, E. H., 882, 917 |
| Hebtogona, 589, 822 | Water Hemlock, f. 653 | Fotsio horrido, 125, 647 |
| Helioscopio, 589, 822 Heptogona, 589, 822 hcterodoxo, 597, 822 hcterophyllo, 121, 588, 600 | Vater Hemiock, 7, 033 | Fatty acids, 73 Faville, G. C., 277 Favus, 12, 14, 17-19, 100, 294- 297, 865 of cats, 295 dogs, 295 |
| heterophyllo 121 588 600 | Yew, 101, 328 | Faville G C 277 |
| h.h 922 | Eurotia cerotoides, 107, 424, | Taville, 0. C., 277 |
| hyberno, 822 | 814 | ravus, 12, 14, 17-19, 100, 254- |
| Ipecachuonha, 599, f. 600, | lanoto, 107 | 297, 865 |
| 602, 822 | Euryale ferox, 445, 839 | of cats, 295 |
| Lothyris, 1, 121, 589, f. 590 599, 602, 822 | Europia machata 915 | dogs, 295 |
| 599, 602, 822 | Eurybio moschato, 815 | guinea pigs, 295 |
| lineoris, 822 | Eustoma Russellianum, 690 | mouse, f. 294, 295 |
| | EUTHALLOPHYTA, 97, | on hair of human beings, |
| lingulorio, 822 | 153, 160-308, 859 | |
| maculato, 121, 122, f. 597, | EUTUBERACEAE, 253, f. 254 | 295 |
| 598 | Evans, I. F., 873, 882 | onchomycosis, 295 |
| morginata, 64, 121, 598, f. | Evans, J. F., 873, 882 Evans, J. R., 882 | Fawcett, H. S., 121 |
| | 1. Valle, 1. IV., 004 | T1 1 207 |
| soo. 600. 822 | 42 1 Total 12 420 | |
| 500, 600, 822 | Evening Lychnis, 439 | Fehr, 297 |
| 599, 600, 822 incllitero, 822 | Evening Lychnis, 439 | Fenchone, 648 |
| 599, 600, 822 incllitero, 822 | Evening Lychnis, 439 | |
| \$99, 600, 822 mcIlifero, 822 neriifolia, 822 obtusota, 121 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 | Fenchone, 648 Fenn, C. M., 883 |
| \$99, 600, 822 mcIlifero, 822 neriifolia, 822 obtusota, 121 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 |
| \$99, 600, 822 mcIlifero, 822 neriifolia, 822 obtusota, 121 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 |
| \$90, 600, 822 ncliliero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 folustris, 4, 589, 822 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 |
| 50, 600, 822 mcllifero, 822 nerifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Porolios, 822 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodu rutoecorpo, 849 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 |
| 50, 600, 822 mcIlifero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Peplus, 589, 822 Peplus, 589, 822 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodu rutoecorpo, 849 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Seeds, 648 Fenshoe, 328 |
| 50, 600, 822 mcllifero, 822 nerifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Porolios, 822 | Evening Lychnis, 439 Primose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Seeds, 648 Fenshoe, 328 |
| \$00, 600, 822 mellifero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 folustris, 4, 589, 822 Porolios, 822 Peplus, 589, 822 pilnifero, 601 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 Fenshoe, 328 Fenugreek, 527 |
| \$90, 600, 822 nncllitero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 folustris, 4, 589, 822 Porolios, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Seeds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 |
| \$90, 600, 822 mcIlifero, 822 neriifolia, 822 obtusoia, 121 officinarum, 822 polustris, 4, 589, 822 Peplus, 589, 822 Pilnifero, 601 piscatoria, 822 Pithyuso, 822 | Evening Lychnis, 439 Primose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoccaria, 52 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Seeds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 |
| \$90, 600, 822 nncllitero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 folustris, 4, 589, 822 Porolios, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoccaria, 52 Apollocho, 588, 823 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Seeds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 |
| \$60, 600, 822 mellifero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Porolios, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoccaria, 52 Apollocho, 588, 823 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Seeds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 |
| \$90, 600, 822 mellifero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Porolios, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 Preslii, 121, 597, f. 602, 603, | Evening Lychnis, 439 Primose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evadia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoccaria, 52 Agollocho, 588, 823 glandulosa, 588 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Seeds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 |
| \$60, 600, 822 mcIliero, 822 neriifolia, 822 obtusoia, 121 officinarum, 822 polustris, 4, 589, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 Preslii, 121, 597, f. 602, 603, 822 | Evening Lychnis, 439 Primose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Ercoecaria, 52 Agollocho, 588, 823 glandulosa, 588irgoto, 823 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 Fungi in, 249 Fermi, C., 883, 903 Fernald, M. L., 883 |
| \$60, 600, 822 mellifero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Porolios, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 Preslii, 121, 597, f. 602, 603, 822 primulacfolio, 822 | Evening Lychnis, 439 Primose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evadia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoccaria, 52 Agollocho, 588, 823 glandulosa, 588 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 Fungi in, 249 Fermi, C., 883, 903 Fernald, M. L., 883 |
| \$60, 600, 822 mellifero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Porolios, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 Preslii, 121, 597, f. 602, 603, 822 primulacfolio, 822 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoccaria, 52 Agollocho, 588, 823 glandulosa, 588irgoto, 823 Exoccarin, 588 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 Fungi in, 249 Fermi, C., 883, 903 Fernald, M. L., 883 |
| \$90, 600, 822 mcIlifero, 822 neriifolia, 822 neriifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Popolios, 822 Popolios, 822 Pinifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 Preslii, 121, 597, f. 602, 603, 822 primulacfolio, 822 pulcherrimo, 679, 822 | Evening Lychnis, 439 Primose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evadia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoecaria, 52 Agollocho, 588, 823 glandulosa, 588 . rirgoto, 823 Exoecarin, 588 Exidio truncato, f. 220 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 Fungi in, 249 Fermi, C., 883, 903 Fernald, M. L., 883 |
| \$69, 600, 822 mcIliero, 822 neriifolia, 822 obtusoia, 121 officinarum, 822 polustris, 4, 589, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 Preslii, 121, 597, f. 602, 603, 822 primulaefolio, 822 pulcherrimo, 679, 822 puniceo, 822 | Evening Lychnis, 439 Primrose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoccaria, 52 Agollocho, 588, 823 glandulosa, 588 . rirgoto, 823 Exoccarin, 588 Exidio truncato, f. 220 EXOASCACEAE, f. 252, | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 Fungi in, 249 Fermi, C., 883, 903 Fernald, M. L., 883 Ferns, 100-101, f. 309, f. 311, f. 312, 312-322, 865 Bladder, 101, 319-320 |
| \$60, 600, 822 mellifero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 Preslii, 121, 597, f. 602, 603, 822 primulacfolio, 822 pulcherrimo, 679, 822 puniceo, 822 puniceo, 822 punicy 822 | Evening Lychnis, 439 Primose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evalar rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoccaria, 52 Agollocho, 588, 823 glandulosa, 588irgoto, 823 Exoecarin, 588 Exidio truncato, f. 220 EXOASCACEAE, f. 252, 252-253 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 Fungi in, 249 Fermi, C., 883, 903 Fernald, M. L., 883 Ferns, 100-101, 1, 300, 1, 311, 1, 312, 312, 322, 865 Bladder, 101, 319-320 Bristle, 313 |
| \$90, 600, 822 merlifero, 822 neriifolia, 822 neriifolia, 822 obtusola, 121 officinarum, 822 polustris, 4, 589, 822 Perplus, 589, 822 Pilnifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 preslii, 121, 597, f. 602, 603, 822 primulacfolio, 822 pulcherrimo, 679, 822 punico, 822 pungiformis, 822 Regis-Jubae, 823 | Evening Lychnis, 439 Primose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evodia rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoecaria, 52 Agollocho, 588, 823 glandulosa, 588 . rirgoto, 823 Exoecaria, 588 Exidio truncato, f. 220 EXOASCACEAE, f. 252, 252-253 Exooscus Alni-inconoe, f. 252 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 Fungi in, 249 Fermi, C., 883, 903 Fernald, M. L., 883 Ferns, 100-101, f. 309, f. 311, 312-322, 865 Bladder, 101, 319-320 Bristle, 313 Cinnamon, 313, 322 |
| \$60, 600, 822 mellifero, 822 neriifolia, 822 obtusota, 121 officinarum, 822 polustris, 4, 589, 822 Peplus, 589, 822 pilnifero, 601 piscatoria, 822 Pithyuso, 822 platyphyllos, 589, 822 Preslii, 121, 597, f. 602, 603, 822 primulacfolio, 822 pulcherrimo, 679, 822 puniceo, 822 puniceo, 822 punicy 822 | Evening Lychnis, 439 Primose Family, 643-645 Everlasting Pea, 572-573 Evergreen Oak, 403 Evernia vulpina, 861 Evesque, 874, 882, 914 Evalar rutoecorpo, 849 Ewart, A. J., 882, 913 Exalgin, 78, 865 Exanthemata, 170 Excoccaria, 52 Agollocho, 588, 823 glandulosa, 588irgoto, 823 Exoecarin, 588 Exidio truncato, f. 220 EXOASCACEAE, f. 252, 252-253 | Fenchone, 648 Fenn, C. M., 883 Fennel, 648 Oil, 86 Source of drug, 87 Secds, 648 Fenshoe, 328 Fenugreek, 527 Ferguson, A. T., 25, 883 Fermentation, 250 Fungi in, 249 Fermi, C., 883, 903 Fernald, M. L., 883 Ferns, 100-101, 1, 300, 1, 311, 1, 312, 312, 322, 865 Bladder, 101, 319-320 Bristle, 313 |

Fischer, A., 874, 883
Fischer, B., 302
Fischer, E., 146
Fischer, R., 883
Fischer, T., 883
Fischer, T., 883
Fisch, 612
Fish, Poisoning from, 77, 78
Poison, 1, 2, 52, 53, 54, 120, 121, 131, 472, 606, 621, 804, 865
California, 125
Scanwart, 503 Forage Plants, 338, 525-526 Poisoning, 20-24, 80, 287, 343, 865 Ford, W. W., Prof., 31, 35, 36, 235, 238, 241, 242, 243, 613, 883 Forget-me-not, 704 Formalin, 78 Climbing, 315 Filmy, f. 309, 313 Flowering, f. 321 Maiden Hair, 101, 313, f. 316, 316-317, f. 317 Male Shield, 149, f. 309, 315 Forget-me-not, 704
Formin, 78
Formic Acid, 80, 106, 412
Forrest, 883
Forster, E. I., 883
Forster, E. I., 883
Forsythia, 128
internedia, 840
suspensa, 128, 683, 840
viridissima, 128, 840
Foss, B., 347
Foster, 687
Foul Brood of Bees, 163
American, 175
European, 174-175
Fouquiera splendens, 627
Fourcroya, See Furcraea
Fourie, 16
Fowl Meadow Grass, Powdery Mildew on, 272
Foxglove, 75, 736-737
Common, 385
Purple, 134, 736, 737, f. Extract of, 77 Formalin, 78 Extract of, // Ostrich, 321 Royal, f. 309, 313, 322 Sensitive, 320-321 Shield, f. 320 Sweet, 399 Tree, 313 California, 125 Soapwort, 593 Fistulina hepatica, 234 PLACOURTIACEAE, 627 FLACOURTIACEAE, 627
Flag, Blue, f. 388
Carolina, 389
Exotic, 389
Large, 105
Sweet, 105
FLAGELLARIACEAE, 825
FLAGELLATAE, 153, 188
Flandin, C., 883
Flaveria Contrayerba, 815
Flax, 120, 580-581, 865
Annual Blue-flowered, 580
Dodder, 701 Fertility, Toxicity of plant affected by, 83 Ferula, 650 communis, 856 foetida, 856 galbaniflua, 648, 856 Narthex, 648 Sumbul, 648 Festuca quadridentata, 826 sylvatica, 229 Fetid Marigold, 139, 140, 784-Dodder, 701 False, 493, f. 494 Family, 120, 580, 581 Fiber, 580-581 785, f. 785 Fetterbush, 673-674 Mountain, 674, 675 Mountain, 674, 675
Fever, Relapsing, 182
Rheumatic, 171
Scarlet, 171
Splenic, Bacillus of, f. 173
Typhoid, 173
Bacillus of, f. 173
Feverwort, 135, 744, 745
Fevillea cordifolia, 751, 819
Feydel, Paul, 883
Fiber, Ceir, 370
Cotton, 623
Jute, 621
Plants, 345, 386, 580-581, 621, 622, 623
Among Leguminosae, 524
Ramie, 406
Raphia, 370
Sisal, 386
Ficcaria, 89
FICOIDEAE, 89, 824
Ficus, 89 Family, 180, 581 Fiber, 580, 581 Large-flowered Yellow, 120, 581 Red Garden, 580 Red Garden, 300 Seed, 120, 581 Fleabane, 776-777, 778-779 Fleming, George, 13, 277, 883 Fleming, Luke, 746 Flemingia congesta, 830 Flickinger, Dr. P. W., 720 Flindersia Schottiana, 837 629 virginiana var. illinoiensis, Floras, Sec page 865 FLORIDEAE, 194 Flower-de-luce, 389 505 Fragrant Golden-rod, 774 Violet, 631 Fraissenet, J. F. F., 884 France, Poisonous plants of, Flowering Almond, 505, 514 Dogwood, 664 Fern, f. 321, 322 Plants, 101 867
Francis, Dr., 25, 54
Francis, Geo., 91, 97, 884
Francoa appendiculata, 851
Frangulin, 620
Frank, A. B., 288, 296
Frankenia ericifolia, 825
granulifolia, 825
Frankenia ericifolia, 825 Spurge, 121, 122, 598, f. Ficus, 89 altissima, 406 Tobacco, 133, 715 Flowers, Poisoning from, 64 benghalense, 406 Carica, 405, f. 407, 857 elastica, 405 hispida, 857 Poisonous, 865 Fluavil, 681 Flückiger, F. A., 59, 86, 449, 481, 482, 594, 718, 863. 883, 888 Flügge, C., 883 Fluggea Leucopyrus, 823 FRANKENIACEAE, 825 hypogaea, 857 Frankforter, G. B., 884 Frasera carolinensis, 689 hypogaea, 857 leucantatomia, 857 religiosa, 406 Roxburghii, 857 Sycamorus, 405 Field Larkspur, 108, 460 Marigold, f. 139, 140 Mushroom, 235 Pea, 521 speciosa, 689 Fraser's Honeysuckle, 744 Fingga Leucopyrus, 823 obovata, 823 Fly Agaric, 10, 31, 74, 148, f. 236, 236-238 Fungus, 195, f. 203, 204 Killer, 34 Frazer, 146 Frazer, T. B., 884, 913 Fraxinus, 629 americana, 683 nigra, f. 682, 683 ornus, 683 pennsylvanica v. lanceolata, Pea, 521 Penny Cress, 495, f. 495 Sow Thistle, 759, 760 Fig. 405 Smyrna, 405 Riler, 34
Poison, 104, 131, 376, 726
Fodder Pea, 45
Fodere, F. E., 3, 5, 883
Foeniculum vulgare, 648
Fogel, Estelle D., 96, 883, 683 Sycamore, 405 Tree, 407 Figwort Family, 134, 733-738 Filbert, European, 402 FILICALES, 312, 313-322, Frederick, Dr. II. J., 176 Free, J. E., 884 Free, J. E., 884
Freeman, 884
Freeman, E. M., 364
Freesia, 389
French Honeysuckle, 525
Mulberry, 708
Pharmacopoeia, 866
Sorrel, 419
Turpentine, 330
"Frenching" of cotton, 287
Fresh-waterCord-grass, f. Fog-fruit, 708 Foliaceous Lichens, 307, f. 824 FILICES, 89, 90, 417 Filicic acid, 77, 319 Filicin, 319 Folliculitis, 16 Fomes applanatus, 234 Foods, Poisonous, 865 Fool's Parsley, 76, 126, 659, Filipendula Ulmaria, 115, 629 f. 660 Filmy Fern, f. 309, 313
Fine-leaved Sneezeweed, f. Foot and Mouth Disease, 277 Foot-rot, of cattle and sheep, Fir, Balsam, 327 Black, 327 179 Eel Grass, 332 Friedherg, L. H., 4, 884, 894 of orange and lemon trees, Douglas, 327 287

| Friedberger, F., 20, 46, 324, | Galangal, 392 | Gouro, 644-645 |
|--|---|--|
| Friedberger, F., 20, 46, 324, 328, 330, 377, 383, 403, 441, 449, 483, 548, 552, 560, 574, 581, 589, 626, 642, 649, 696, 707, 721, 723, 751 | Golonthus nivalis, 386, 805 | biennis. 644-645 |
| 441, 449, 483, 548, 552, 560, 574, 581, 589, 626 | Golega, 534, 865 | coccinea, 645 |
| 642, 649, 696, 707, 721, | officinalis, 534, 535, 830 orientalis, 830 | parvifloro, 645 Scarlet, 645 |
| 748, 731, 884 | GALEGEAE, 504 | Gautier, L., 265, 873, 884 |
| Fries, 803 | Galen, 276 | Goutiero morchollaeformis, f. |
| Frijolillo, 543 Fringe Tree, 129, 683 | Galeopsis tetrolit, 131 Golium, 90, 741 | Gaylussacia baccato, 665 |
| Fringed Gentian, 690 | Golium, 90, 741 Aparine, 848 | Genster 245 |
| Fritilloria, 377 | osprellum, 848 | fimbrialus, f. 246 Geiger, P. L., 3, 884 Geissospermum Vellosii, 147, |
| Imperialis, 4, 834 Meleogris, 834 | triflorum, 744, 848 Gallesio Scorododendron, 841 | Geissospermum Vellovii 147 |
| pudica, 834 Fröhner, E., See Friedberger Frothingham, L., 884 Frothingham, L., 884 | Gallic acid, 503, 608, 612 Gallotannic acid, 533 Galloway B T 565 | 807 |
| Fröhner, E., See Friedberger | Gallotannic acid, 533 | Gelsemin, 77, 80, 129, 147, |
| Frollingham, L., 884 | | 689 Gelseminin, 147, 689 |
| Frullonio, 310 Fruticose Lichens, 307 | Galls, Aleppo, 403 Gamaleia, N., 884 | Gelsemium, 688-689, 865 |
| FUCACEAE, 194 Fuchs, F. A., 265, 884 | Gamaleia, N., 884 Gambier, 742 | sempervirens, 129, 147, f. |
| Fuchsio, 644 | Gambier, J., 565, 567 Gamboge, 627 | 686, 689, 836 Geniophlebium incanum, 825 |
| Fucus vesiculosus, f. 102, i. | American, 629 | Genista, 865 |
| 193, 194 | Gamgee, J., 213, 261, 290 | ephedroides, 90 |
| Fuligo, 160 Fuller's Teasel, 741 | American, 629 Gamgee, J., 213, 261, 290 Gammer, 65 GAMOPETALAE, 157 | florido, 90 |
| Funaria officinolis, 113, 825 | Gangrene, Dry. 28, 100 | germanico, 90 monospermo, 90, 831 |
| spicata, 825 | Gangrene, Dry, 28, 100 Erysipelatous, 600, 601 | monospermo, 90, 831 ramosissimo, 90 |
| FUMARIACEAE, 113, 824 | Gonophyllum, 89 | sphaerocorpo, 531, 831 |
| Fumarin, 483 Fumitory, 113 | folcotum, 850 Garcinio Cambogio, 827 | spicota, 90 tinctoria, 90, 524-525, f. |
| Climbing, 480 | Forsteriano, 827 | tinctoria, 90, 524-525, f. 525, 831 GENISTEAE, 530, 551 |
| Family, 113 | Hanburyi, 627 | GENISTEAE, 530, 551 |
| Funaria, 310 hygrometrica, f. 211 | Mongostana, 627 | Gentian, Closed, 690 Family, 129, 689-691 |
| hygrometrica, f. 311 Fungi, 53. 90, 98, 195-306 | Garden Cress, 113 Leek, 383 | Fringed, 690 |
| Cup, 253 | Lettuce, /30, /00, /02 | Horse, 135 |
| Edible and poisonous, Dif- ference between, 239- | Pea, 521, f. 521 Poppy, f. 60, 73, 111, f. 487, 481-482 Purclay 423 | White, 690, f. 690 Yellowish, 690 |
| _ 240 | 481. 481-482 | Gentiana Andrewsii, 689, 690 |
| Gelatinous, 220 | 1 ussicy, 423 | crinito, 690 flavido, 690, f. 690 puberulo, 689 |
| Parasitic, 281 Poisoning by, 31-36, 78 | Raspberry, 505 | buherulo. 689 |
| Poisonous, 2, 5, 72, 803, | Strawberry, 505 Thyme, 709 | verno, 825 |
| 865 | Gardenia josminioides, 742 | GENTIANACEAE, 129, 683, 689- |
| Trichophytic, 295, 300 Fungi Imperfecti, 100, 154, | Gardiner, W. W., 884 | 691, 825 Gentiopicrin, 689 |
| 281-306, 861 | Garget, 434 | Gentistic acid, 689 |
| Cause of disease, 12 Fungus, Bird's nest, f. 246 | Garlic, 375, 383 Field, 383 | Geoffraea superba, 831 |
| Cat-tail, 273 | Golden, 383 | Geophila mocropodo, 848 |
| Corn-silage, f. 248 | Wild, 383 Garner, W. W., 86 | reniforemis, 848 GERANIACEAE, 120, 575, 577- |
| Corn-silage, f. 248 Death-cup, 241 | Garner, W. W., 86 Garnsey, H. E., 884 | 579, 825 |
| Dry-rot, f. 232 Erythrasma, 298 | Gorrya Fremontii, 127, 817 | GERANIALES, 157, 574-604 Geraniol, 577, 709 |
| Fly, 195, 204 Mold, 293 | Gas (illuminating), poisoning | Geraniol, 577, 709 |
| Mold, 293 | by, 6 | Geronium, Cultivated, f. 578 Family, 67, 120 |
| Prickle, f. 232 Rape, 100, 283 | Gas Plant, 581 GASTEROMYCETES, f. 246 | Oil, 506 |
| Funifero utilis, 642 | Gastro-enteritis, 10, 20, 319, | Robertionum, 578 |
| Fürbringer, R., 199 | 328, 330, 371, 375, 377, | sp., 150 Geranoil, 88 |
| Furcroeo, 89, 386 gigantea, 805 | 386, 421, 581, 643, 652, 694, 725, 726, 804, 865 | Geranoil, 88 Gerordia, 734, 737-738 grondifloro, 134, 738 |
| Furcroyo, See Furcroea Fusarium, 260, 287-294, f. | Gastrolobic acid, 533 | grondifloro, 134, 738 |
| 19 18 18 18 18 18 18 18 18 18 18 18 18 18 | Gastrolobin, 531 | |
| culmorum, 290 | Gostrolobium, Australian, 533 | tenuifolio, 134, 738, 852 Slender, 134, 738 Gerhardt, 145 |
| equinum, 100, f. 292, 293 | bilobum, 531, 830 colycinum, 533, 831 | Gerhardt, 145 |
| net crosporum, 288, 1. 289 | collistochys, 831 | Gerlach, M., 16, 295, 301 German, John, 884 German Chamomile, 754 |
| linonis, 287 lycopersici, 287 | grondiflorum, 533, 831 | German Chamomile, 754 |
| mouiliforme, 290 | obovatum, 831 ovalifolium, 831 | Millet, 102, 7. 349, 350 |
| roseum, 100, 273, 288, 364, 861 | oxylobioides, 831 | Pharmacopocia, 866 Vetch, 572 |
| vasinfectum, 287 | Poisonous to Stock, 37, 533 polystochyuni, 533 | Germander, 709 |
| sp., 291, 293 | spinosum, 831 | Germany, Poisonous plants of, 867 |
| Fusicladium destruens, 284 | trilobum, 533, 831 | of, 867 |
| Fustic, 406 | Gattinger, A., 884 | Germicides, 865 Gerrard, A. W., 884 |
| G | Goultheria, 666, 669, 865 nicrophylla, 821 | Gesneriaceae, 698 |
| | | Geum nivole, 505 |
| Gaffky, George, 884 Gaillard, A., 237 | Oil of, 665, 866 procumbens, 404, 665, 821 Shallon, 665 | urbonum, 505 Geyer, C. A., 801 |
| Gaillardio pulchello, 756 | Shallon, 665 | Gherkins, West Indian, 750 |
| | , 555 | The state of the s |
| | | |

Giant Cactus, 635
Puff-hall, 99
Gibbeellina cercalis, 267, f. 268
Gibberella, 273, 861
Sanbenetii, 273, f. 288, 288
Gibbes, H., 884
Gibbons, H., 884
Gibson, R. J. H., 885
Gid, 569
Giesecke, 3 (11d, 569) Giesecke, 3 Gift Apple, 865 Gilchrist, W. D., 556 Gilia achillacfolia, 843 aggregata, 130, 698, 843 lacinitata, 843 Gillenia stipulacea, 505, 847 Gillenia stipulacea, 505, 8 stipulata, 115, 847 Gillot, Victor, 244, 245 Gillot, X., 885 Gilman, J. F., 885 Giltay, E., 284 Giltner, Ward, 344 Gimlette, J. D., 729, 885 Gin, Juniper used in manufacture of. 3. Ginger, 77, f. 390, 391, 865
Wild, f. 415, 416
Gingerol, 391
Ginkgo biloba, 326, 815
Seed, 395
GINKGOACEAR GINKGOACEAE, 326 GINKGOALES, 326 Ginora mexicana, 836 Ginseng, f. 646, 647 Chiuese, 647 Family, 125, 646-647 Gipsies, Poison used by, 6 Gisekia pharnacioides, 824 Githagin, 440 Githagin, 440
Githagism, 441
Gladiolus, 389
communis, 827
segetum, 827
Glanders, 180
Bacillus of, f. 181
Farcy, 180
Glaucin, 113
Claucin corriculatum Glaucium corniculatum, 841 flavum, 841 navam, 841 Gleditschia, 63, 89, 865 amorphoides, 831 sinensis, 831 triacanthos, 527, 537, 83! Gleichenia, 315 flabellata, 825 GLEICHENIACEAE, 315 Glenk, R., 918 Gleocapsa, 184, 187, f. 309 Gliricidia maculata, 831 Globba Beaumetsii, 858 Globularia Alypum, 853 Gloiotrichia, 93, 186-187 Pisum, 187 Gloiopeltis coliformis, 194 Gloiopeltis coliformis, 194
Gloriosa simplex, 834
superba, 376, 834
Glover, G. II., 96, 346, 347, 467, 885
Glozinia, 698
Gluco-alkaloids, 149
Achillein, 149
Solanin, 149
Glucose, 489, 505
Glucosides, 865
Absinthin, 140, 755
Achillein, 140, 149, 756
Adonidin, 148, 446
Aesculetin, 617
Aesculin, 123, 617, 642, 683

Amygdalin, 7, 54, 83, 116, 117, 144, 503, 504, 505, 506, 512, 517, 518, 519
Antiarin, 148, 406
Apocynein, 692
Apocynin, 129, 148, 692
Arbutin, 505, 506, 665, 666
Asclepiadin, 696
Asclepiadin, 696
Asclepiadin, 694
Baptisin, 533, 540
Bryonin, 751
Caricin, 627
Catalpin, 739
Cephalanthin, 135, 744
Cheiranthin, 78, 148
Cheiratin, 689
Cinceria 127, 750 Cheiratin, 689 Chicorin, 137, 759 Colocynthin, 77, 749, 750 Convallamarin, 104, 148, Convallamarin, 104, 148, 385
Convallarin, 104, 148, 385
Convolvulin, 130, 699, 702
Coronillin, 148
Crocin, 742
Cyclamin, 52, 89, 128, 676
Daphnetin, 125, 642
Daphnetin, 125, 642
Dhurrin, 54, 348
Digitalin, 78, 80, 130, 134, 144, 148, 385, 441, 534, 692 736, 737
Digitonein, 148
Digitonin, 134, 148, 736
Digitonin, 134, 148, 736
Digitonin, 134, 17, 736
Diosmin, 583
Elaterin, 749
Ericolin, 127, 665, 666, 669
Erythrocentaurin, 689
Erythrophloein, 418, 534
Eunayinin, 148, 615
Euparin, 772
Eupatorin, 138, 755, 772
Frangulin, 620
Gastrolobin, 531
Centionicin, 689 385 Gastrolobin, 531 Gentiopicrin, 689 Glycyrretin, 531 Glycyrrhizin. 527, 531 Gratiolin, 734
Gynocardin, 54
Helleborein, 449
Helleborein, 73, 148, 449
Helleborin, 73, 148, 449
Hesperidin, 583 Hydrochinon, 506 Ibo, 683 Indican, 106, 419 Ipomoein, 130, 701 Irigenin, 389 Jalapin, 702 Kolanin, 621 Lappin, 755 Leptandrin, 733, 736 Linariin, 735 Loganin, 687 Lollin, 103 Lotusin, 54, 534 Lupinin, 118, 531, 548 Lupinin, 118, 531, Melanthin, 62 Nerin, 130, 148 Oleandrin, 148 Ononid, 531 Ononin, 531 Oxycoccin, 666 Phaseo-lunatin, 54 Phillyrin, 683 Pinicrin, 328 Populin, 397 Prophetin, 749 Prophetin, 749 Purshianin, 621 Queratrin, 560 Quercitrin, 506, 510, 584, 666

Rhamnetin, 124, 620 Rhamnin, 124, 620 Rhamno-cathartin, 124 Rhamno, cathartin, 124
Rhinanthin, 738
Robinin, 119, 560
Salicin, 397, 505
Scillain, 148, 377
Sinalbin, 489
Sinigrin, 489
Solauin, 60, 83, 132, 133, 144, 148, 149, 718, 721, 722, 724
Tampicin, 702
Tephrosin, 533, 558
'Thevetin, 148, 692
Turpethin, 702
Tutin, 149, 607
Veratramarin, 381
Villosin, 508
Violin, 632, 633
Wisterin, 534
Xanthostrumarin, 137, 769 Wisterin, 534
Xanthostrumarin, 137, 769
Glucotropaeolum, 575
Glue, 529
GLUMIFLORAE, 336-369
Gluta Benghas, 805
Glyceria, 90, 826
aquatica, 53
canadensis, 692
fluitans, Ergot on, 276
Glycine hispida, 520, f. 540, 831
Soia, 520 831
Soja, 520
Glycosuria, 73
Glycyrretin, 531
Glycyrriiza, 534
glabra, 527, f. 528
glabra v. glandulifera, 527
Glycyrriizin, 527, 531
Gmelin, J. F., 3, 885
Gnephosis eriocarpa, 815
GNETALES, 826
GNETALES, 326
GNETALES, 326
GRETUN SCANGERS, 825 Guetum scandens, 825 urens, 825 Gnidia carinata, 642 Mrens, 825
Graidia carinata, 642
Gnoscopin, 481
Goat's Death. 64
Goat's Rue. 118, 558
Godet, C., 862, 885
Godfrey, B., 885
Godfrey, B., 885
Godfrey, George, 885, 899
Godlee, R. S., 885, 905
Goeppert, II. R., 885
Gola, G., 885
Godkin, 888
Golden Corydalis, 113
Glow, 756, 779
Seal, 448, f. 468, 688
Golden-rod, 138, 222, 755, 767, 774-776, 865
Fragrant, 774
Riigid, 775, 776
Rust, 222, 775
Goldsmith, W. W., 383, 384
Gompholobium, 534, 831
uncinatum, 533
Goniothalamus macrophyllus, Goniothalamus macrophyllus, 806 Gonolobus laevis, 809 obliquus, 809 Gonorrhea, 170 Good King Henry, 427, f. Good, Peter, 885 Goodale, G. L., 633 Goodenia grandistora, 825 GOODENIACEAE, 825 Goodia, 533 lotifolia, 533, 831 medicaginea, 533

| Gooseberry, Cape, 715 Cultivated, 500 Missouri, 500 Septoria on, 284 Goosefoot, 427 Gooseplant, 417 Family, 424-430 Gordon, P. R., 871, 885 Gardania, 629 | Orchard Powdery Mildew | Tea, 628 Greene, E. R., 746 Greenish, H. G., 886 Greenleaf, R. W., 886 Greimer, Karl, 886 Greshoff, M., 54, 65, 84, 88, 89, 90, 96, 101, 110, 114, 115, 124, 128, 130, 135, 141, 315, 426, 447, 498, 500, 503, 535, 567, 590, 621, 698, 750, 803, 804, 886 Gressler F. G. L., 886 Grewia, 2, 52, 89, 621 asiatica, 855 bracteata, 855 |
|---|--|---|
| Cultivated 500 | Orchard, Powdery Mildew on, 272 Rust on, 229 | Greene, E. R., 746 |
| Missouri, 500 | Rust on, 229 | Greenish, H. G., 886 |
| Septoria on, 284 | Spot-disease of, 282, 71 | Greenleaf, R. W., 886 |
| Goosefoot, 427 | 283 | Greimer, Karl, 886 |
| Gooseplant, 417 | Pigeon, 98 | Greshoff, M., 54, 65, 84, 88, |
| ramily, 424-430 | Porcupine, 102, 355-556, f. 356, 865 Smut on, f. 216 | 114 115 124 128 130 |
| Gordon, P. R., 8/1, 883 | Smut on f. 216 | 135, 141, 315, 426, 447, |
| Gardania, 629 Gosio, B., 885 | Poverty | 498, 500, 503, 535, 567, |
| Gossypium, 624, 626 | Long-awned, 352-353, f. | 590, 621, 698, 750, 803, |
| arharousu 623 | 353 | 804, 886 |
| barbadeuse, 623, 625 herbaceum, 76, 124, 622, f. 625, 625-626, 837 hirsutum, 625 | Short-awned, f. 353 Quack, 103, 364-365 Black-spot Disease of, f. 280, 280 Ergot on, 276 | Grevillea wimoenides 844 |
| herbaceum, 76, 124, 622, | Quack, 103, 364-365 | Grenia 2. 52. 89 621 |
| f. 625, 625.626, 837 | 280 280 | asiatica, 855 |
| hirsitum, 625 | Ergot on, 276 | bracteata, 855 |
| Gottneil, 803 | Quaking, 90 | Malacocca, 855 orientalis, 855 |
| Gotti, 263 | Quaking, 90 Ramie, 406 | orientalis, 855 |
| Gouania, 2, 52, 89, 621, 847 tomentosa, 621 Goumi, 641, f.641 | | pilosa, 855 |
| Goumi. 641. f.641 | Red Top, 338, 865 | Grev Geo 886 |
| Gourd, Dipper, 751 Family, 135, 749-751 Missouri, 751 Nest-egg, 750 Sponge, 750 Towel, 750 | Red Top, 338, 865 Ergot on, 277 Rice-cut, f. 338, 339 Rust, f. 223, 224-227, f. | piscatorum, 855 Grey, Geo., 886 Griffiths, David, 635 Grohe, 886 |
| Family, 135, 749-751 | Rust, f. 223, 224-227, f. | Grohe, 886 |
| Missouri, 751 | | Gromwell, 629 |
| Nest-egg, 750 | Rye, 361-364 | Grigorescu, G., 886 |
| Sponge, 750 | Rye, 361-364 Common, 361 | Grindella, 89, 757, 774 |
| C=-0 995 | | rahusta 774 815 |
| Göze, 885 Gracilaria lichenoides, 194 | Salt, 90 | Grigorescu, G., 886 Grindelia, 89, 757, 774 lanceolata, 138 robusta, 774, 815 squarrosa, 138, 755, 774 Tourneforti, 815 Grindelin, 755, 774 Grönlund, C., 886 |
| Graham H 885 | Sleepy, 102, 355, 357-358 865 | Tournefortii, 815 |
| Graham, H., 885 GRAMINEAE, 53, 89, 90, 102- 103, 336-367, 692, 825 Graininol, 767 | | Grindelin, 755, 774 |
| 103, 336-367, 692, 825 | Smut, 98, 212, 214-219 Snake, 692 | |
| Graminol, 767 | Sprouting Smut, f. 214 | Gross, Herman, 886 Ground Cherry, 715, f. 715 Lvy, 131, 710, f. 711 |
| Grand Traverse Disease, 311 | Squirrel-tail, 66, 103, 360, | Ground Cherry, 715, f. 715 |
| Grandilla, 633 | 366-367, f. 368 | Plum f. 662 |
| Grandilla, 633 Grape, Clinton, 620 | Powdery Mildey on 272 | Plum, f. 563 Groundsel, 795-797 |
| Concord, 620 | Smut, 98, 212, 214-219 Snake, 692 Sprouting Smut, f. 214 Squirrel-tail, 66, 103, 360, 366-367, f. 368 Injuries from, 66, 360 Powdery Mildew on, 272 Staggers, 21 Sweet, 865 Tickle Rust on, f. 223 | Salt, 138 Grouse poisoned by Laurel, 669 |
| European, 620 Family, 124 | Sweet, 865 | Grouse poisoned by Laurel, |
| Fox | Staggers, 21 Sweet, 865 Tickle, Rust on. f. 223 Triple-awned, 352 | 669 |
| Northern, 620 | Triple-awned, 352 | "Grub-in-the-head," 138, 776 Gruby, 13, 295, 301, 302, 886 Grünfeld, 279 |
| Southern 620 | Tulip, 868 Vanilla, f. 341, 552 Vernal, 524 Sweet, 552 Wheat, 364 | Grünfeld 279 |
| Fruit, 582 | Vanilla, 1. 341, 552 | Guaço, 417 |
| Janesville, 620 | Sweet 552 | Guaicum officinale, 575, 853 |
| Fruit, 582 Janesville, 620 Mustang, 124, 620 Oregon, 112, 472, f. 472 Powdery Mildew on, 269 | Wheat, 364 | Guanin, 146 |
| Powdery Mildey on 260 | Western, Ergot on, 276 | Guapacum, 89 |
| Small 620 | Wild, Ergot on, 276 | Guar, 526 |
| Small, 620 Wild Blue, 620 | Wire, 374 Yellow-eyed, 372 | Guarana, 606 |
| Worden, 620 | Yellow-eyed, 372 | Guardia, J., 886 Guatteria veneficiorum, 806 Guava, 525, 639 Gueguen, J., 886 Guenther, J., 887 Guerin, P., 363, 364, 887 Guiana, Poisonous plants of, |
| Graphis scripta, f. 308 | Gratiola officinalis, 4, 134, 734, 852 | Guava. 525. 639 |
| | peruviana, 852 | Gueguen, J., 886 |
| Beard, 344-348 Bent, 225 Blue, 338, f. 339 Ergot on, f. 27, 276 Powdery Mildew on, f. 27, 277, 272 Blue-joint, Ergot on, f. 27 Bottle Front on, f. 27 | sp., 134 | Guenther, J., 887 |
| Bent, 225 | sp., 134 Gratiolin, 734 | Guerin, P., 363, 364, 887 |
| Frant on f 27 276 | Gravel-root, 772 Graves, C. B., 881, 885 Grawitz, Paul, 260, 295, 298, 302, 305, 885 Gray, Dr. Asa, 533, 602, 885, 885 | Guiana, l'oisonous plants of, |
| Powdery Mildew on, f. | Graves, C. B., 881, 885 | |
| 271, 272 | Grawitz, Paul, 260, 295, 298, | Guignard, L., 119, 574, 747, 887 |
| Blue-joint, Ergot on, f. 27 | Cross Dr. Acr. 522 602 895 | Guinea Pepper, 725 |
| Bottle, Ergot on, f. 27 Bristly Foxtail, f. 339 | 886 | Guizotia abyssinica, 755 |
| Bristly Foxtail, J. 339 | Greasewood, 68, 107, 429, 430 | Gum, Canoe, 405 |
| China 406 | Great Angelica, 660 | Elastic, 588 |
| Brome, 338 China, 406 Cord, Fresh-water, f. 69 | Lobelia, 753 Ragweed, f. 136, 137, 765- 766 | Galbanum, 648 |
| Curly, Small, 315 | Ragweed, f. 136, 137, 765- | Kino, 523, 640 Sandarac, 328 |
| Curly, Small, 315 Cuscus, 345 | 766 | Sandarac, 328 |
| Eel, Fresh-water, 332 Family, 89. 90, 336-337 Foxtail, 207, 349, 865 Fowl Meadow, Powdery Mildew on, 272 | St. John's-wort, 125, 629. 630-631, f. 631 | Sour, 664 Tragacanth, 530 |
| Family, 89, 90, 336-337 | Willow-herb 644 | Tupelo, 664 |
| Fowl Meadon Powdery | Great-flowcred Larkspur, 460 | Gumbo, 623 |
| Mildew on 272 | Magnolia, 473, f. 474 | Gumweed, 755, 774 |
| Holy, 339 | 630-631, f. 641 Willow-herb, 644 Great-flowcred Larkspur, 460 Magnolia, 473, f. 474 Great Britain, Poisonous plants of, 867 | Gun Cotton, 623 |
| Hungarian, 102, 350 | plants of, 867 | Gustavia, 52 |
| Holy, 339 Hungarian, 102, 350 Johnson, 1. 344, 345, 346, | Green, J. R., 250, 486, 886 Green, Wesley, 886 Green Algae, 160 Ash, 683 | Augusta, 814 brasiliana, 814 |
| 863 | Green, Wesley, 886 | Gutta, 681 |
| Knot, 421 Lemon, 344 | Green Algae, 160 | Gutta-percha, 680, 681 |
| Meadow-oat Tall f 240 | Asn, 683 Brier, 104 | Tree, f. 680 |
| Mexican, 342 Needle, 66, f. 70, 102, 338, f. 355, 355-357 Western, f. 66, 102 | Hellebore, f. 45, 75, 109, | Guttenberg, G., 96, 435, 887 |
| Needle, 66, f. 70, 102, 338, | 449 | Guttenberg, G., 96, 435, 887 GUTTIFERAE, 827 |
| f. 355, 355-357 | Mold, f. 257, 257, 261 Pigweed, 107 | Guvacin, 370 |
| Western & 66 102 | l'igweed, 107 | Gymnauthes Incida, 823 |

Gymnema latifolium, 53, 809 sylvestre, 809 GYMNOASCEAF, 252 Gymnoascus, 252 Gymnoascus, 252 Gymnoascus, 89, 534, 535, 536-538 canadensis, 527, 831 chinensis, 831 dioica, 63, f. 116, 117, 537, f. 538 Gymnogramme cordata, 825 GYMNOSPIERMAE, 101-102, 155, 325-332 Gymnosperms. See Gymnospermae Gymnosporangium macropus, 222 Gymosporangium macropus, 627, 810 Gynocardia odorata, 53, 54, 627, 810 Gynocardin, 54 Gynopogon, 691 Gypsophila, 89, 436 Gynsophila, 89, 436 Gynsophila, 89, 436 Gynocardiata, 436-437 Struthium, 63, 436, 437, 813 Gypsophila-sapotoxin, 62 Gyromitra esculenta, 253, 859 Gyrotheca, 385 majalis, 827

Habenaria nigra, 840 Hackberry, 408 Hackel, E., 362 Haemanthus, 805
coccineus, 805
Haematein, 528
Haematoxylin, 528
Haematoxylon campechiannm, 528 Haematuria, Caused by Marsin Marigold, 499 Caused by Ferula, 650 HAEMODRACEAE, 104, 375, 385-386, 827 Haemodorum, 827 385-386, 827
Haemodorum, 827
Haemodorum, 827
Haemolysis, 107
Hager, Hermann, 887
Hahn, G., 301
Haines, W. S., 887, 888, 904
Hair-balls. See Phyto-bezoars
Hairy Phacelia, f. 703, 704
Trilisa, 773
Vetch, 527, 570
Halesia carolina, 682
Halderman, 887 Halesia carolina, 682
Halderman, 887
Hale, E. M., 887
Half-breed Weed, 137
Halgand, 887
Halle, J. S., 887
Haller, 362, 721
Halcanemum frulicosum, 814
HALORHAGIDACEAE, 640
Halsted, B. D., 105, 135, 276, 381, 386, 415, 575, 583, 601, 634, 663, 732, 751. 887, 888
Hamamelis virginiana, 500
Hamburger, W. W., 305
Halimodendron argenteum, 831 831 Hamelia patens, 848 Hamilton, 411, 888 Hamilton, A. M., 888, 903, 914 Hamilton, W., 888 Hanbury, D., 59, 86, 449, 481, 482, 593, 718, 883, 914 Honcornia, 691 Hannibal, 2

Hanriot, Maurice, 558
Hansen, N. E., 526, 641, 888
Haplocarpha lyrata, 815
Hapliphyton cimicidum, 807
Harcourt, A. M., 553
Hard, M. E., 234, 803
Hard Maple, 615, f. 616
Hard-shell Hickory, 402
Hardin, M. B., 626
Hardy Catalpa, 134, f. 740
Pink, 436
Hare, R. F., 635, 873, 888.
899
Harger, E. B., 888 899 Harger, E. B., 888 Harknis, W. D., 888, 912 Harlan, H., 888 Harnack, E., 888, 899 Harper, R. A., 222, 267 Harpullia, 52, 607 arborea, 850 cupanioides, 850 thanatophora, 850 thanatophora, 850
Harrington, C., 888
Harris, N. M., 305
Harrison, F. C., 25
Harrison, W. H., 888
Harshberger, J. W., 277, 342, 888, 889
Hartley, C. P., 342, 889
Harttanan, G., 889
Hartzell, Stella, 422.423, 889
Harz, C. O., 247, 248, 359, 889
Hash. See Hashish Hash, See Hashish Hashish, 7, 411, 445 Hashishin, 411 Haslam, T. P., 889 Hatch, 199 Hatch, W. G., 889 Havard, Dr. V., 357, 889 Haw, 116, 506, 513-514 Red, f. 514 Hawthorn, 513-514 Hay, Moldy, as cause of disease, 259 Hay-fever, 136, 137, 764, 767, 865 Serum, 767 865
Serum, 767
Haya Poison, 865
Hayes, Capt., 626
Hazelnut, 403
Hazeltine, Dr. R., 49, 655
Heald, E. D., 889, 892
Heald, F. D., 96, 100
Heart, Poisons acting on the,
74, 75
Also see Cardiac Poisons Also see Cardiac Poisons Heart's ease, 632-633 Heath Family, 127-128, 664-675 Heather, 65 Scotch, 665 Heaves, 865 Heboloma crustuliniforme, 859 fastibile, 860 Hebert, A., 889 Hecate, 2 Heckel, E., 889, 909 Hedeoma, 710, 711 Oil, 711 pulegiodes, 131, 711 Hedeomol, 711 Hedera Helix, 808 Hedge Bindweed, 701-702, f. 703 Hyssop, 134 Mustard, 114, 487-488 Hedges, Florence, 287, 889, 911 Hedrick, U. F., 3, 656, 658,

Hedychinm longecormatum, 858 Hedysarum coronarium, 525 Heffter, A., 889 Hefti, J. J., 889 Hegetschweiler, J., 862, 890, 895 Hehir, P., 890 11ein, Heinrich, 890 Hein, Heinrich, 890 HEKLOR, L., 890 HELENHEAE, 757, 758 Helenium, 754 Antunnale, 139, 756, 781-783, f. 782, 816 autunnale v. grandiflorum, 781 781 Bigelovii, 781 Hoopesii, 140, 781 nudiflorum, 781 quadridentatum, 783 tenvifolium, 139, f. 783-784, f. 784, 816 Heliamphora, 497 HELIANTHEAE, 757 Helianthemum canadense, 620 Helianthus, 757 annuus, 756 Maximiliani, 763 tuberosus, 756, 816 Helichrysum apiculatum, 755, 816 Helicia robusta, 844 Helicin, 505 282 inconspicnum, 281-282 turcicum, 281, f. 282 HELOBIAE, 332-336 HELOBIAE, 332-336

Helconias, 89
frigida, 834

Helvella, 253
esculenta, 237, 238
suspecta, 253

HELVELLACEAE, 253, 859

Helvellic acid, 78, 238, 253

HELVELLINEAE, 253-255

HEMIASCALES, 247-248

HEMIASCALES, 247-248 HEMIBASIDIALES, 210-220

| HEMIBASIDH, 209, 210-220 | Hibiscus abelmoschus, 624 |
|---|--|
| Hemidesmus, 90 | cannabinus, 623 |
| indicus, 695, 809 Hemileia vastatrix, 222 | diversifolius, 837 clatus, 623 |
| Hemizonia macradenia, 138 | csculentus, 623 |
| Hemlock, 327, f. 327, 651-659, 865 | csculentus, 623 ficulneus, 623 ficulneus, 623, 624 tiliaceus, 623, 624 tiliaceus, 623, 624 Hickman, R. W., 25, 891 Hickory, 402, 403 Hiard-shell, 402 |
| Bulb-bearing, 126 | tiliaceus, 623 |
| l'itch, 327 | Hickman, R. W., 25, 891 |
| Poison, f. 50, 51, 72, 75, 648, 651 | Hard-shell 402 |
| Purple-stemmed, f. 655 | Missouri, 401 Shell-bark, 401 Hieracium, 757 venosum, 692, 755 virosum, 816 |
| Water, 126, 648, 651-659, 865 | Shell-bark, 401 |
| Bulbous, f. 658, 659 | venosum, 692, 755 |
| European, f. 653 | virosum, 816 |
| Bulbous, f. 658, 659 European, f. 653 Oregon, 656-658, f. 657 Hemolysins, 35, 36, 242 Hemorrhagic Septicemia, 177 | Hierochloc, 90 australis, 826 |
| Hemorrhagic Septicemia, 177 | barcalis, 826 |
| | odorata, 339, 1. 341 varidora 826 |
| Indian, 72, 129, 411, 692, | odorata, 339, f. 341 rariflora, 826 High-bush Blackberry, 508, f. |
| Common, f. 41a Indian, 72, 129, 411, 692, 693, f. 693 Manila, 391 | 508 Hilger, A., 891, 892 Hillman, F. II., 378 Hiltner, R. S., 348 Hinebauch, T. D., 350, 351, |
| Mauritius, 386 | Hillman, F. II., 378 |
| Queensland, 624 | Hiltner, R. S., 348 |
| Henbane, 2, 6, 718, 726,727 | Hinebauch, T. D., 350, 351, 891 |
| Mauritius, 386 Queensland, 624 Hempel, C. J., 890 Henbane, 2, 6, 718, 726-727 Black, 133, 726-727, f. 727 Henderson, 278 | Hippcastrum equestre, 805 |
| Henderson, 278 Henkel, Alice, 85, 732, 890 | rutilum, 805 HIPPOCASTANACEAE, 123, 607, |
| | HIPPOCASTANACEAE, 123, 607, 616-618, 827 |
| Hennings, P., 229 | 616-618, 827 HIPPOCASTANEAE, 52. |
| Henry, J., 890 | 63, 89 Hippocrates, 572 |
| Henry, T. A., 54, 88, 348, | Hippocrates, 572 Hippomane, 865 |
| 450, 534, 881, 890 Henry W. A. 213 | Mancinella, f. 586, 587, 823 Hippophae rhamnoides, 125, |
| Hennings, P., 229 Henry, Aimé, 862, 890 Henry, J., 890 Henry, T. A., 54, 88, 348, 450, 534, 881, 890 Henry, W. A., 213 Henslow, G., 862, 890 Hepaticae, 310 Hectafleurum emarginatum. | f. 640, 641 |
| Hepaticae, 310 Heptapleurum emarginatum, | Hippurus vulgaris, 640 Hirtz, 721 |
| Heptapleurum emarginatum, 808 | Hitchcock, A. S., 891 |
| scandens, 808 | Hitchcock, A. S., 891 Hoary Pea, 558 Hochsetter, C. F., 891 Hoffman, E. R., 891 Hoffman, F. R., 891 |
| venulosum, 808 Heraclea, Poisonous honey | Hoffman, E. R., 891 |
| in. 64 | Hoffman, F. R., 891 |
| Heraeleum, 650, 663, 865 lanatum, 126, 648, 663, 865 | Hog Pea, 204 |
| spondylium, 865 | |
| spondylium, 865 Heraclin, 126, 663 Herb of Grace, 76 Herbarium Mold, 257 | Hogweed, 766 Hogwort, 121, 591, f. 591 Holcus, 826 |
| Herbarium Mold, 257 | Holcus, 826 |
| nerus, poisonous, necate | lanatus, 90 Holden, R. J., 222 |
| Herissey, J., 890, 895 | Holigarna caustica 805 |
| Herman, L., 890 | ferruginea, 608 |
| Herniaria, 63, 89, 436 | ferruginea, 608 longifolia, 805 Holland, J. W., 10 Holly, American, 614 |
| rici maria-saponini, 02 | Holly, American, 614 |
| Hernandia sonora, 828 Heröin, 147 | European, 614 Family, 123, 614 Sea, 648 |
| Herpes, 15, 17 tansurans, 301 | Sea, 648 |
| tausurans, 301 | Hollyhock, 623 Holm, T., 656, 772, 891 Holmes, E. M., 587, 891 Holy Grass, 339 Hamalanthus crepitans, 823 |
| Herr, Bertha D., 803 Herzog, M., 890 | Holmes, E. M., 587, 891 |
| Herzog, M., 890 Hesperidin, 583 | Holy Grass, 339 |
| Hester, Dr., 655 Heteramarpha arbarescens, | Leschenaultianus, 588, 823 |
| 856 | Homalium, 53 |
| Heteropagon cantartus, 354 hirtus, 826 Heteropteris syringaefolia, | Hamalomena aramatica, 808 Homen, E. A., 891 |
| Heteropteris syringaefolia, | Hameria callina 300 827 |
| 837 Hetier, Fr., 890 | Homeria collina, 390, 827 Homochelidonin, 485 Honey Locust, 527 Plant, Simpson, 134 Poisonous, 64, 121, 601, |
| Henchera americana, 500 | Plant, Simpson, 134 |
| Hensinger, H., 262, 295 | Poisonous, 64, 121, 601, |
| Hevea, 587 brasiliensis, 53, 588 | 606, 865 Honeycomb Ringworm, 294 |
| Heyer, F., 890 | Honev-dew. 275 |
| Heyl, 637 | Honeysuckle, Bush, 135 |
| Hibbertia glaberrima 819 | Francis, 135, 744-748 |
| Hibbertia glaberrima, 819 longifolia, 819 | Honeysuckle, Bush, 135 Family, 135, 744-748 Fraser's, 744 French, 525 |
| | |

Rocky Mountain Bush, 135 Sullivant's, 744 Trumpet, 744 Western, 744 Western, 744 Honeywort, 676 Hoof-ail, 277 Hooper, D., 891 Hop Clover, 552 Low, 553 Yellow, 553 Tree, 581 Hops, Common, 106, 406 Oil of, 406 Horand, 14 Hordeum, 340, 366-367 caespitosum, 103 distichum, 366
Gussanianum, 103
jubatum, 66, f. 67, 103,
366-367, f. 368
Ergot on, 276
murinum, 36, 367
nodosum, f. 67
secatinum, 103, 367, f. 368
spontanum, 366
vulgare, 103, 366
Horehound, 709
Horse Gentian, 135
Nettle, 60, 132, 718, f. 723,
724
Horse-chestnut, 123, 617, f. distichum, 366 Horse-chestnut, 123, 617, f. Horse-chestnut, 123, 617, f.
618
European, 617
Horseradish, 113, 435, 486,
490,491, f. 491, 656
Horsetail, 37, f. 28, 101, 312,
322-325, 865
Horse-weed, 724, 755, 777778, f. 778
Small, 777
Horsfield, Thomas, 891
Hosackia Purshiana, 525, 831
Houghton, Dr., 411
Hound's Tongue, 131, 705706, f. 706
Houseleek, 221
Howard, C. D., 517, 891, 900
Howell, Miss J. K., 231
Hoya caruosa, 695
Huart, L., 891
Hubbard, R., 891
Huckleberry, 665, 719
Huff, Elisha, 771
Hugemeyer, 891
Hughes, D. A., 541, 891
Hughes, D. A., 541, 891
Hughes, Richard, 891
Hugnia Mystax, 835
Hume, 90
Hunnulus Lupulus, 106, 406, 857 618 Humulus Lupulus, 106, 406, 857 857
Oil of, 106, 406
Hungarian Grass, 102
Hungary, Poisonous plants of, 867
Hunt, Reid, 686, 891, 893
Huntington, Annie Oakes, 612, 862, 891
Hura crepitans, 587, f. 587
Husemann, A., 4, 892
Husemann, Th., 4, 891, 892
Hutcheon, D., 892
Huttpa, F., 13, 26, 892, 893
Hyacinth, 375
Hyaciuthus arientalis, 375 Hyacinthus arientalis, 375, 834 Hyaena Poison, 588 Hyams, C. W., 380, 415, 541, 580, 663, 677, 688, 747, 892 Hydatt, Virgil, Case of, 49 Hybanthus Ipecachuanha, 632 Hyde, J. N., 13, 19, 295, 298, 299, 302, 892, 900

| Hydnocarbus, 52 | Hyoscyamus, 2, 52, 60, 72, | Inaein, 692 |
|---|--|--|
| Hydnocarpus, 52 anthelminticum, 810 | Hyoscyamus, 2, 52, 60, 72, 78, 148, 716, 717, 726-727, 865 | Indaconitin, 450 |
| castanea, 810 | 727. 865 | India, Poisonous plants of, |
| heterophylla, 811 | albus, 853 | 867 |
| inebrians, 54 | | Wheat, 419 |
| Kursii, 811 | Falezlez, 854 muticus, 854 miger, 4, 60, 133, 726-727, f. 727, 854 | India-rubber Tree, 405 Indian Aconite, 449, 450 |
| piscidia, 811 | muticus 854 | Indian Aconite 449 450 |
| venenata, 53, 627, 811 | niger, 4, 60, 133, 726-727, | Almond 638 |
| Wrightiana, 811 | f. 727. 854 | Almond, 638 Bean, White, 574 |
| Hydnum amarescens 860 | physaloides, 854 | Berry, 1, 149 |
| graveolens 860 | reticulatus, 854 | Colza, 490 |
| imbricatum f 222 | Hypecoum procumbens, 841 | Cucumber 104 |
| graveolens, 860 inibricatum, f. 232 Hydrangea, 84, 89, 500, 629 arborescens, 115, 500, 851 Cultivated, 500 | HYPERICACEAE, 124-125, 629- | Berry, 1, 149 Colza, 490 Cucumber, 104 Currant, 744, 748 |
| arborescens 115 500 851 | 631 | Heliotrope, 705 |
| Cultivated, 500 | HVPERICINEAE 827 | Hemp, 72, 129, 411, 692, |
| Cardon 500 | Hypericum 629.631 | Hemp, 72, 129, 411, 692, 693, f. 693 |
| Garden, 500 Hortensia, 84, 115, 500, | Hypericum, 629-631 Ascyron, 125, 629, 630-631, | Ipecacuanha, 695 |
| Hortensia, 84, 115, 500, 629, 851 | f. 631 | Laurel, 478 |
| angiolograta 500 X51 | crispum, 827 | Mallow, 124, 626-627 |
| involucrata, 500, 851 Lindleyana, 500, 851 paniculata, 500 Thunbergii, 500, 851 Wild, 115, 500 | crispum, 827 humifusum, 827 | Mallow, 124, 626-627 Mustard, 490 |
| haniculata 500 | Kalmianini 629 | Opium, 85 |
| Thunharai 500 851 | maculatum, 124 630 827 | Pear, 635 |
| Wild 115 500 | maculatum, 124, 630, 827 Oil of, 630, 631 | Pennywort, 648 |
| 77 1 1 100 146 469 | perforatum, 125, 629-630, | Persimmon 681 |
| Hydrastin, 108, 140, 400 | f 620 631 827 | Persimmon, 681 Pink, 129, 688 |
| Hydrastis, 447, 408 | perforatum, 125, 629-630, f. 630, 631, 827 prolificum, 629 | Sarsaparilla, 695 |
| Hydrastin, 108, 146, 468 Hydrastis, 447, 468 canadensis, 88, 108, 446, 468, f. 468, f. 468, 846 Hydrastium, 146 | punctatuni, 630 | Shot 301 |
| 468, 1. 408, 840 | Hypha f 17 | Strawberry, 505, 509 Teak Tree, 708 Tobacco, 75, 136, 752 Turnip, 372, f. 372 Indian Poissons of |
| Hydrastium, 146 | Hypha, f. 17 Hyphaene thebaica, 840 HYPHOMYCETES, 249 | Teak Tree 708 |
| Hydric Cyamu, 75 | HVPHOMVCETES 240 | Tobacco 75 136 752 |
| Hydrochinon, 506 | Hyphorbe indica, 840 | Turnin 372 f 272 |
| Hydrocotarnin, 481 | Hypnaga muscaetromic 850 | Indiana, Poisonous plants of. |
| Hydrocotyle asiatica, 856 javanica, 856 umbellata, 856 . vulgaris, 856 | Hypnaea muscaefromis, 859 Hypnotic Poison, 804 | 866 |
| javanica, 856 | Hypocratic facies, 35 | Indians, Food plants of, 10d Indican, 106, 419 Indies, East, Fish-poison of, 1 |
| umbellata, 856 . | IIVPOCREACEAR 100 273-278 | Indican 106 419 |
| vulgaris, 856 | IIYPOCREACEAE, 100, 273-278, 859 | Indies East Fish-poison of 1 |
| Hydrocyanic acid, 4, 6, 53- | HYPOCREALES, 273-279 | Indigo, 424 |
| 55, 77, 80, 82, 83, 89, | Ilyssop, Hedge, 134 | Australian, 534 |
| Tydrocyanic acid, 4, 6, 53- 15, 77, 80, 82, 83, 89, 90, 101, 102, 105, 115, 116, 119, 120, 128, 140, 141, 315, 346, 348, 447, 498, 500, 503, 512, 517, | 11) 350p, 11cage, 154 | Blue, 524 |
| 116 119 120, 128, 140, | 1 | Chinasa 421 |
| 141, 315, 346, 348, 447, | • | Chinese, 421 Large White Wild, 117, |
| 498, 500, 503, 512, 517, | Iberis, 486 | 541, f. 541 |
| 498, 500, 503, 512, 517, 518, 519, 567, 574, 590, 622, 627, 633, 681, 747, 804, 862, 865 Distribution in Vegetable | Ibo, 683 | Large-bracted Wild, 541 |
| 622, 627, 633, 681, 747, | Ibotu, 683 | |
| 804, 862, 865 | Ibotu, 683 Icaco Plum, 505 Ice Plant, 423 | Indigo-eaters, 533 |
| Distribution in Vegetable | Ice Plant, 423 | Indigofera, 534 Anii, 524, 831 australis, 534, 831 galegioides, 53, 747, 831 tinctaria, 831 |
| Kingdom, 53-55, 89-90 | Ichthvothere 57 | australia 521 921 |
| Effect of, 54 | Cunabi 816 | galagicidan 53 747 921 |
| Effect of, 54 Toxicity of, 7 | Ictrogen, 78, 149, 548 | tinetaria 921 |
| Hydrodictyon reticulatum, | Idaho, Poisonous plants of. | *************************************** |
| 190 | Cunabi, 816 Ictrogen, 78, 149, 548 Idaho, Poisonous plants of, 866 | Inebriants, 74, 804 |
| | Idaho Pea, 120 Ikeno, S., 248 Ilex Aquifolium, 123, 614 Cassine, 123, 614, 742, 807 glabra, 807 apaca, 614 | Inflammation of internal or- gans, 172 |
| Hydrogen, Peroxid of, 72 Hydrogen in Alkaloids, 144 | Ikeno, S., 248 | gans, 1/2 |
| Hydrogen potassium sulphate, | Ilex Aquifolium, 123, 614 | respitory passages, 170 serous membrane, 171 |
| 489 | Cassine, 123, 614, 742, 807 | Serous memorane, 1/1 |
| Hyppopuyutaceae 131 698 | glabra, 807 | 11111attin, 752 |
| HYDROPHYLLACEAE, 131, 698, | apaca, 614 | Influenza, 176 |
| 703-704, 827 Hydroquinon, 505, 742 Hygrin, 147, 575 | puraguensis, 014, 742 | Inga vera, 525 Ink Tree, 608 Inocybe, 244 |
| Hygrin 147 575 | theezans, 614 | 1пк 1гее, 608 |
| Hygrophila hispida, 804 | verticillata, 807 | 11100000, 244 |
| Hygrophorus agathosmus, 53 | Ilicin, 614 ILICINEAE, 827 | ecumata, 244 |
| Hymenaea courbaril, 523, 527, | ILICINEAE, 827 | 111144, 244 |
| 831 | ILLECEBRACEAE, 89, 827 | Juste V 227 000 |
| | Illicium, 89 | 1поко, т., 237, 892 |
| Hymenocallis, 805 | anisatum, 149, 474, 837 floridanum, 112, 474, 837 | echinata, 244 infida, 244 scaber, 860 Inoko, Y., 237, 892 Insect Powders, 789 |
| HYMENOLICHENES, 154 | floridanum, 112, 474, 837 | |
| Hymenogaster tener, f. 246 HYMENOGASTRINEAE, | vernum, 474 Illinois State Board of | Persian, 789 Traps, f. 499 Insecticide, 804 |
| HYMENOGASTRINEAE, | Illinois State Board of | Traps, f. 499 |
| 245-247 | Health, 892 | Insecticide, 804 |
| HYMENOGASTRACEAE, 245 | Illipe Maclayana, 681, 851 | Hydrocyanic acid as, 54 |
| HYMENOMYCETES, f. 232, | | |
| | Imbert-Gourbeyre, 892 | Insectivorous plants, 498 |
| 281 | Imbert-Gourbeyre, 892 | Intoxicant, 804 |
| HYMENOMYCETINEAE, | Imbert-Gourbeyre, 892 | Intoxicant, 804 |
| HYMENOMYCETINEAE, 233-245 | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 | Intoxicant, 804 |
| HYMENOMYCETINEAE, 233-245 Hymenophilaceae 313 | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 Impatiens, 618, 619-620 | Intoxicant, 804 Inula, 757 Helenium, 754 Royleana, 816 |
| HYMENOMYCETINEAE, 233-245 Hymenophilaceae 313 | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 Impatiens, 618, 619-620 ampharata, 810 | Intoxicant, 804 Inula, 757 Helenium, 754 Royleana, 816 INULEAE, 757 |
| HYMENOMYCETINEAE, 233-245 Hymenophilaceae, 313 Hynson, H. P., 881, 892 Hypenanche, globasa, 823 | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 Impatiens, 618, 619-620 ampharata, 810 Balsamina, 619 | Intoxicant, 804 |
| HYMENOMYCETINEAE, 233-245 Hymenophilaceae, 313 Hynson, H. P., 881, 892 Hypenanche, globasa, 823 | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 Impatiens, 618, 619-620 ampharata, 810 Balsamina, 619 | Intoxicant, 804 Inula, 757 Helenium, 754 Royleana, 816 INULEAE, 757 Inulin, 753, 754, 756 Ionidium Ibecacuanha, 858 |
| HYMENOMYCETINEAE, 233-245 Hymenophulaceae, 313 Hynson, H. P., 881, 892 Hyoemanche globasa, 823 Ilyoscin, 148, 716, 726, 727, 730 | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 Impatiens, 618, 619-620 ampharata, 810 aurea, 810 Balsamina, 619 biflora, 620, 810 fulra, 123 | Intoxicant, 804 Inula, 757 Helenium, 754 Royleana, 816 INULEAE, 757 Inulin, 753, 754, 756 Ionidium Ibecacuanha, 858 |
| HYMENOMYCETINEAE, 233-245 Hymenophulaceae, 313 Hynson, H. P., 881, 892 Hyoemanche globasa, 823 Ilyoscin, 148, 716, 726, 727, 730 | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 Impatiens, 618, 619-620 ampharata, 810 aurea, 810 Balsamina, 619 biflora, 620, 810 fulra, 123 | Intoxicant, 804 Inula, 757 Helenium, 754 Royleana, 816 INULEAE, 757 Inulin, 753, 754, 756 Ionidium Ipecacuanha, 858 Iowa, Poisonous plants of, 866 |
| HYMENOMYCETINEAE, 233-245 Hymenophulaceae, 313 Hynson, H. P., 881, 892 Hyoemanche globasa, 823 Ilyoscin, 148, 716, 726, 727, 730 | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 Impatiens, 618, 619-620 ampharata, 810 aurea, 810 Balsamina, 619 billora, 620, 810 fulra, 123 Nali-tangere, 123, 810 Royalii, 123 | Intoxicant, 804 Inula, 757 Helenium, 754 Royleana, 816 INULEAE, 757 Inulin, 753, 754, 756 Ionidium Ipecacuanha, 858 Iowa, Poisonous plants of, 866 |
| HYMENOMYCETINEAE, 233-245 Hymenophulaceae, 313 Hynson, H. P., 881, 892 Hyoemanche globasa, 823 Ilyoscin, 148, 716, 726, 727, 730 | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 Impatiens, 618, 619-620 ampharata, 810 Balsamina, 619 | Intoxicant, 804 Inula, 757 Helenium, 754 Royleana, 816 INULEAE, 757 Inulin, 753, 754, 756 Ionidium Ipecacuanha, 858 Iowa, Poisonous plants of, 866 |
| HYMENOMYCETINEAE, 23.245 HYMENOPHLLACEAE, 313 Hynson, H. P., 881, 892 Hyoemanche globasa, 823 Hyosein, 148, 716, 726, 727, | Imbert-Gourbeyre, 892 Immunity, 165, 862, 865 Impaction, 9, 68, 102, 342 Impatiens, 618, 619-620 ampharata, 810 aurea, 810 Balsamina, 619 billora, 620, 810 fulra, 123 Nali-tangere, 123, 810 Royalii, 123 | Intoxicant, 804 Inula, 757 Helenium, 754 Royleana, 816 INULEAE, 757 Inulin, 753, 754, 756 Ionidium Ibecacuanha, 858 |

INDEX 947

| Roots of, 78, 742 Wild, 599, 1, 600 Ipreacutanha, Source of, 87 Ipreacutanha, Source of, 87 Ipreacutanha, Source of, 87 Ipreacutanha, Source of, 87 Ipreacutanha, Source of, 87 Ipreacutanha, Source of, 87 Ipreacutanha, Source of, 87 Ipreacutanha, Source of, 87 Ipreacutanha, Source of, 87 Ipreacutanha, Source of, 87 Ipreacutanha, Source, 89 Ipreacutanha, Source, 89 Ipreacutanha, Source, 89 Ipreacutanha, Source, 89 Ipreacutanha, Source, 99 Ipreacutant, 10, 817 Ipreacutanha, Source, 99 Ipreacutant, 10, 817 Ipreacutant, 82 Ipreacutant, 10, 817 Ipreacutant, 82 Ipreacutant, 99 Ipreacut | | | |
|--|---------------------------------|---------------------------------|---------------------------------|
| Wild, 599, 1, 600 precacatank, Source of, 87 pob, 865 pob, 860 pob, 865 pob | Roots of, 78, 742 | Ivy, Boston, 620 | |
| Input | Wild, 599, f. 600 | Branch, f. 672 | |
| Input | Ipecacuanha, Source of, 87 | | Jenersonia aipnyna, 88, 409 |
| Bona.nox, 699 congesta, 699 dissecto, 591 dispecto, 599 dissecto, 591 dispecto, 599 dispecto, 591 dispecto, 599, 817 partiguia, 699, 4, 700, 701 Jalapa, 699, 817 partiguia, 699, 4, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 817 plepiophylid, 701 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 817 plepiophylid, 701 partiguia, 699, 817 plepiophylid, 701 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 190 point and 181, 190 point and | Indian, 695 | 865 Japan 620 | Jenking E H 130 701 |
| Bona.nox, 699 congesta, 699 dissecto, 591 dispecto, 599 dissecto, 591 dispecto, 599 dispecto, 591 dispecto, 599, 817 partiguia, 699, 4, 700, 701 Jalapa, 699, 817 partiguia, 699, 4, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 817 plepiophylid, 701 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 817 plepiophylid, 701 partiguia, 699, 817 plepiophylid, 701 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 190 point and 181, 190 point and | Ipob, 865 | Poison 77 122 123 394. | Jennings, O. E., 893 |
| Bona.nox, 699 congesta, 699 dissecto, 591 dispecto, 599 dissecto, 591 dispecto, 599 dispecto, 591 dispecto, 599, 817 partiguia, 699, 4, 700, 701 Jalapa, 699, 817 partiguia, 699, 4, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 817 plepiophylid, 701 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 817 plepiophylid, 701 partiguia, 699, 817 plepiophylid, 701 partiguia, 699, 1, 700, 701, 817 partiguia, 699, 1, 700, 701, 817 partiguia, 190 point and 181, 190 point and | Patatas 600 f 600 | 600, 608, 613, 676, 679, | Jequirity, 534, 865 |
| dissecto, 537 fastigiata, 699, 617 Jalopa, 699, 817 leptophylla, 701 pandurata, 130, 817 pargarese, 130, 817 pargarese, 1699 sinuata, 817 tuberesa, 1809 sinuata, 817 tuberesa, 1809 sinuata, 817 tuberesa, 1809 sinuata, 817 tuberesa, 1809 sinuata, 817 tuberesa, 1809 sinuata, 817 tuberesa, 1809 Dwarf Garden, 180, 827 Jalopin, 180, 827 Jalopin, 180, 827 Jalopin, 180, 827 Jalopin, 180, 827 Jalopin, 180, 827 Jalopin, 180, 827 Jalopin, 180, 827 Jalopin, 180, 827 Jalopin, 180, 827 Jalopin, 180, 827 Jalopin, 180, 828 Jalopin, 180, 829 Jaris, 180, 827 Jalopin, 180, 828 Jalopin, 180, 829 Jalo | Bong. noz. 699 | 865 | Ophthalmia, 56 |
| fastiguida, 699, 1, 700, 701 Talajaq, 699, 810 Talajaq, 699, 811 Talajaq, 699, 817 Talajaq, 699, 690, 700, 701, 817 Turgar, 699 | congesta, 699 | California, 123, 609 | Plant, 11, 52 |
| fastiguida, 699, 1, 700, 701 Talajaq, 699, 810 Talajaq, 699, 811 Talajaq, 699, 817 Talajaq, 699, 690, 700, 701, 817 Turgar, 699 | dissecto, 53 | Three-leaved, 609-610 | Seed, 55, 527 |
| fastiguida, 699, 1, 700, 701 Talajaq, 699, 810 Talajaq, 699, 811 Talajaq, 699, 817 Talajaq, 699, 690, 700, 701, 817 Turgar, 699 | | | Jerusalem Artichoke, 710, 756 |
| rimberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timber | fastigiata, 699, f. 700, 701 | J | Jervin, 103, 148, 381 |
| rimberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timber | Jalapa, 699, 817 | Taborandi 149 592 865 | Jessamine, See Jasmine |
| rimberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timber | leptophylla, 701 | Taborin 148 582 | lew Wandering 374 |
| rimberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timber | panahrata, 130, 617 | Jacaranda, 52 | Jewel Weed, 619 |
| rimberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timberosa, 818 timberosa, 817 timber | Parga, 699, J. 700, 701, 817 | procera, 739, 810 | Tewett, 893 |
| simate, 817 timberesa, 817 timberesa, 130, 701 legenin, 30, 701 legenin, 310, 375, 388-390 lixid, 180, 287 lixid, 180, 287 Carolina, 389 Dwarf Garden, 389 Pamily, 388-390 florentina, 389, 827 footidistian, 827 germanica, 389, 827 doctidistian, 827 germanica, 389, 827 hybrida, 827 missourients, 390 prismatica, 828 presudacearens, 828 prismila, 390 prismatica, 828 presudacearens, 828 prismila, 390 prismatica, 828 presudacearens, 828 prismila, 390 prismatica, 828 presudacearens, 828 prismila, 390 prismatica, 828 prismila, 390 prismil | Ouomockit 699 | Jack Fruit, 405 | Jimson Weed, 6, 60, 61, 133, |
| Indian 130, 701 Iregenin, 389 IRIDEAE, 827-828 IRIDEAE, 827-829 IrideAmy-Lantern, 93 IRIDEAE, 827-829 IrideAmy-Lantern, 93 IRIDEAE, 827-829 IrideAmy-Lantern, 93 IRIDEAE, 827-829 IrideAmy-Lantern, 93 IRIDEAE, 827-829 IrideAmy-Lantern, 93 IrideAmy-Lantern, 92 Irid | sinuata, 817 | Pea, 521 | 7. 133, 729-733, 7. 730 |
| Jack-my-Lantern, 99 JRIDEAE, 827-828 823 Jackson, J. K., 892 Jackson, J. K., 893 Jackson, J. K., 893 Jach, J. S., 993 Jackson, J. S., 893 Jackson, J. K., 893 Jackson, | | Pine, 330 | Purple, 133, 729, 732-733 |
| RIDACER, 105, 375, 388.390 RIDEAE, 827-828 RIDEAE, 827-828 RIDEAE, 827-828 RIDEAE, 827-828 RIDEAE, 827-828 RIDEAE, 827-828 RIDEAE, 827-828 RIDEAE, 827-828 RIDEAE, 827-829 RIDEAE, 827-828 | Ipomoein, 130, 701 | Jack-in-the-Pulpit, 103 | Joannesia, 52 |
| RIDBARE 827-828 Iridin, 105 1875, 388-390 Tris, i., 380, 827 Carolina, 389 Reveal 1805, j. 388-390 Reveal 1805, j. 389-390 | Iregenin, 389 | Jack-my-Lantern, 99 | princeps, 823 |
| Jacoby, M., 893 | IRIDACEAE, 105, 375, 388-390 | Jackson, J. R., 692 | Ion Due Weed 771 |
| Jacoby, M., 893 | IRIDEAE, 827-828 | Jacobi 278 | Joe 1 ye Weed, 771 |
| foreitidissima, \$27 germanica, 389, 826 foreitidissima, \$27 germanica, 389, 827 germanica, 389, 827 germanica, 389, 827 germanica, 328 foreitidida, 390 foreiti | Iridin, 105 | Tacobi C., 892 | Johne, L. 26 |
| foreitidissima, \$27 germanica, 389, 826 foreitidissima, \$27 germanica, 389, 827 germanica, 389, 827 germanica, 389, 827 germanica, 328 foreitidida, 390 foreiti | Carolina 380 | Jacoby, M., 893 | Johnson, C., 893 |
| foreitidissima, \$27 germanica, 389, 826 foreitidissima, \$27 germanica, 389, 827 germanica, 389, 827 germanica, 389, 827 germanica, 328 foreitidida, 390 foreiti | Dwarf Garden, 389 | Jacquinia, 52 | Johnson, F. M., 460 |
| foreitidissima, \$27 germanica, 389, 826 foreitidissima, \$27 germanica, 389, 827 germanica, 389, 827 germanica, 389, 827 germanica, 328 foreitidida, 390 foreiti | Family, 388-390 | arborea, 839 | Johnson, L., 96, 114, 390, 419, |
| foetidistima, 827 germanica, 389, 827 hybrida, 827 germanica, 389, 827 hybrida, 827 missouriensis, 390 meglecta, 827 pallida, 390 perismatica, 828 proudacorris, 828 proudacorris, 828 primila, 390 perismatica, 828 sibirica, 390 meglecta, 827 pollida, 390 perismatica, 828 sibirica, 390 meglecta, 827 pollida, 390 perismatica, 828 sibirica, 390 meglecta, 827 pollida, 390 perismatica, 828 proudacorris, 639 primila, 390 perismatica, 828 primila, 390 perismatica, 828 primila, 390 perismatica, 828 primila, 390 perismatica, 828 primila, 190 primila, 829 primila, 190 primila, 829 primila, 190 primila, 829 primila, 190 primila, 829 primila, 829 primila, 820 primila, 820 primila, 820 primila, 820 primila, 821 primila, 821 primila, 821 primila, 821 primila, 821 primila, 821 primila, 821 primila, 821 primila, 821 primila, 821 primila, 831 primila, 831 primila, 831 primila, 831 primila, 831 primila, 831 primila, 831 primila, 831 primila, 831 primila, 831 primila, 832 primila, 832 primila, 832 primila, 832 primila, 833 primila, 834 primila, 834 primila, 835 | norenting, 389, 041 | arminuris, os | 421, 433, 433, 400, 404, |
| Instruction | foetidissima, 827 | obovata, 839 | 491, 560, 625, 669, 719, |
| Instruction | germanica, 389, 827 | Jahn, E., 893 | 736, 746, 767, 772, 773, |
| Prismatica, 828 Pseudoacorns, 828 psumila, 390 reticulata, 828 sibirica, 390 variegata, 828 versicolor, 105, f. 388, 389- 390, 828 Irish, H. C., 714, 725 Irisin, 389 Iron, Salts of, 7 Ironweed, 765 Iron-wood, 395, 681 Irvitants, 76, 804 Irving, 572, 892 Isaria farinosa, 273 Isatis corniculata, 818 sinctoria, 846 Isoportlerin, 886 Isoportlerin, 886 Isoportlerin, 887 Isoportlerin, 889 Isop | hybrida, 827 | Jaiap, 130, 699, 7. 700, 702 | 7/4, /9/, 093 |
| Prismatica, 828 Pseudoacorns, 828 psumila, 390 reticulata, 828 sibirica, 390 variegata, 828 versicolor, 105, f. 388, 389- 390, 828 Irish, H. C., 714, 725 Irisin, 389 Iron, Salts of, 7 Ironweed, 765 Iron-wood, 395, 681 Irvitants, 76, 804 Irving, 572, 892 Isaria farinosa, 273 Isatis corniculata, 818 sinctoria, 846 Isoportlerin, 886 Isoportlerin, 886 Isoportlerin, 887 Isoportlerin, 889 Isop | | | Johnson T V 654 |
| Prismatica, 828 Pseudoacorns, 828 psumila, 390 reticulata, 828 sibirica, 390 variegata, 828 versicolor, 105, f. 388, 389- 390, 828 Irish, H. C., 714, 725 Irisin, 389 Iron, Salts of, 7 Ironweed, 765 Iron-wood, 395, 681 Irvitants, 76, 804 Irving, 572, 892 Isaria farinosa, 273 Isatis corniculata, 818 sinctoria, 846 Isoportlerin, 886 Isoportlerin, 886 Isoportlerin, 887 Isoportlerin, 889 Isop | negiccia, 827 | Locust Tree, 523 | Johnson Grass, f. 244 345. |
| Pseudoacorus, 828 | pattiali, 390 | Sorrel, 623 | 340 |
| reticulata, 828 sibirica, 390 variegata, 828 versicolor, 105, f. 388, 389- 390, 828 Irish, H. C., 714, 725 Irisin, 389 Iron, Salts of, 7 Ironweed, 765 Iron-wood, 395, 681 Irritants, 76, 804 Irving, 572, 892 Isaria farinosa, 273 Isatis corniculata, 818 tinctoria, 846 isonandra, 53 Isopilocarpin, 148 Isopyrin, 446 Isopyrin, 446 Isopyrin, 446 Isopyrin, 446 Isopyrin, 446 Isorottlerin, 589 Istotma axillaris, 812 Brownii, 812 longifora, 812 Isotaropis juncea, 831 Israel, O., 878, 889, 892, 893, 894, 915 Millet, 350 Pharmacopoeia, 666 Rye Grass, 361 Italia, L. van, 892 Itch, Barber's, 100, 300 Phobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 79, 860 Iva, 757, 753-764 axillaris, 763 xanthiifola, 137, 763-764, f. 764 Total of the control of | Pseudoacorus, 828 | Jambosa malaccensis, 639 | Tointill, 179 |
| versicolor, 105, f. 388, 389- 390, 828 Irish, H. C., 714, 725 Irisin, 389 Iron, Saits of, 7 Ironweod, 395, 681 Irritants, 76, 804 Irving, 572, 892 Isaria farinsa, 273 Isatis corniculata, 818 tinctoria, 846 isonandra, 53 Isopilocarpin, 148 Isopyrun finnarioides, 846 thalictroides, 446, 846 Isoprin, 446 Isopyrun finnarioides, 846 thalictroides, 446, 846 Isopolicarpin, 181 Israel, O., 878, 889, 892, 893, 894 Istalian Clover, 118 Millet, 350 Pharmacopocia, 866 Rye Grass, 361 Italia, L. van, 892 Disease of Horses, 293-294 Ithyophallus inpudicus, 79, Disease of Horses, 293-294 Ithyophallus inpudicus, 79, 860 Iva, 757, 763-764 avillaris, 763 xonthiifolia, 137, 763-764, f. Iristin, 389 Janczewski, 286 Janpaconitin, 148, 450 Janpan Clover, 525 Japaconitin, 148, 450 Japan Clover, 525 Iapaconitin, 148, 450 Japan Clover, 525 Iapaconitin, 148, 450 Japan Clover, 525 Inpun, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 867 Ispace, 742 Carolina, Alkaloids of, 147 False, 129 Large-flowered, f. 684 Oil, 86 Cape, 742 Carolina, Alkaloids of, 147 False, 129 Large-flowered, f. 684 Oil, 86 Yellow, 688-689 Jaminium floribundum, 840 grandiflorum, 683, f. 684 officinale, 840 Ispace, 722 Judkins, E. II., 893 Judkins, E. II., 893 Judkins, E. II., 893 Judkins, E. II., 893 Juck, K. W., 893 Juck, K. W., 893 Judkins, E. II., 893 Juck, K. W., 894 Juck, Medicinal plants of, 866 Plum, 505, 514 Poisonous plant | | nulgaris, 639 | Jolliffe, J. J., 215 |
| versicolor, 105, f. 388, 389- 390, 828 Irish, H. C., 714, 725 Irisin, 389 Iron, Saits of, 7 Ironweod, 395, 681 Irritants, 76, 804 Irving, 572, 892 Isaria farinsa, 273 Isatis corniculata, 818 tinctoria, 846 isonandra, 53 Isopilocarpin, 148 Isopyrun finnarioides, 846 thalictroides, 446, 846 Isoprin, 446 Isopyrun finnarioides, 846 thalictroides, 446, 846 Isopolicarpin, 181 Israel, O., 878, 889, 892, 893, 894 Istalian Clover, 118 Millet, 350 Pharmacopocia, 866 Rye Grass, 361 Italia, L. van, 892 Disease of Horses, 293-294 Ithyophallus inpudicus, 79, Disease of Horses, 293-294 Ithyophallus inpudicus, 79, 860 Iva, 757, 763-764 avillaris, 763 xonthiifolia, 137, 763-764, f. Iristin, 389 Janczewski, 286 Janpaconitin, 148, 450 Janpan Clover, 525 Japaconitin, 148, 450 Japan Clover, 525 Iapaconitin, 148, 450 Japan Clover, 525 Iapaconitin, 148, 450 Japan Clover, 525 Inpun, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 867 Ispace, 742 Carolina, Alkaloids of, 147 False, 129 Large-flowered, f. 684 Oil, 86 Cape, 742 Carolina, Alkaloids of, 147 False, 129 Large-flowered, f. 684 Oil, 86 Yellow, 688-689 Jaminium floribundum, 840 grandiflorum, 683, f. 684 officinale, 840 Ispace, 722 Judkins, E. II., 893 Judkins, E. II., 893 Judkins, E. II., 893 Judkins, E. II., 893 Juck, K. W., 893 Juck, K. W., 893 Judkins, E. II., 893 Juck, K. W., 894 Juck, Medicinal plants of, 866 Plum, 505, 514 Poisonous plant | | Jambuse Berries, 639 | Jones, L. R., 101, 324, 893 |
| versicolor, 105, f. 388, 389- 390, 828 Irish, H. C., 714, 725 Irisin, 389 Iron, Saits of, 7 Ironweod, 395, 681 Irritants, 76, 804 Irving, 572, 892 Isaria farinsa, 273 Isatis corniculata, 818 tinctoria, 846 isonandra, 53 Isopilocarpin, 148 Isopyrun finnarioides, 846 thalictroides, 446, 846 Isoprin, 446 Isopyrun finnarioides, 846 thalictroides, 446, 846 Isopolicarpin, 181 Israel, O., 878, 889, 892, 893, 894 Istalian Clover, 118 Millet, 350 Pharmacopocia, 866 Rye Grass, 361 Italia, L. van, 892 Disease of Horses, 293-294 Ithyophallus inpudicus, 79, Disease of Horses, 293-294 Ithyophallus inpudicus, 79, 860 Iva, 757, 763-764 avillaris, 763 xonthiifolia, 137, 763-764, f. Iristin, 389 Janczewski, 286 Janpaconitin, 148, 450 Janpan Clover, 525 Japaconitin, 148, 450 Japan Clover, 525 Iapaconitin, 148, 450 Japan Clover, 525 Iapaconitin, 148, 450 Japan Clover, 525 Inpun, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 867 Ispace, 742 Carolina, Alkaloids of, 147 False, 129 Large-flowered, f. 684 Oil, 86 Cape, 742 Carolina, Alkaloids of, 147 False, 129 Large-flowered, f. 684 Oil, 86 Yellow, 688-689 Jaminium floribundum, 840 grandiflorum, 683, f. 684 officinale, 840 Ispace, 722 Judkins, E. II., 893 Judkins, E. II., 893 Judkins, E. II., 893 Judkins, E. II., 893 Juck, K. W., 893 Juck, K. W., 893 Judkins, E. II., 893 Juck, K. W., 894 Juck, Medicinal plants of, 866 Plum, 505, 514 Poisonous plant | sibirica, 390 | Jamesia, 89, 115 | Jones, W., 891, 893 |
| Janczewski, 260 Janczeski, | variegata, 828 | americana, 115, 500, 851 | Jonquii, 386 |
| Iron, Salts of, 7 Ironweed, 765 Ironweod, 395, 681 Irritants, 76, 804 Irving, 572, 892 Isaria farinosa, 273 Isatis corniculata, 818 tinctoria, 846 Isonandra, 53 Isopilocarpin, 148 Isopyrum fumarioides, 846 thalictroides, 446, 846 Isorotterin, 589 Isotoma axillaris, 812 Brownii, 812 Brownii, 812 Isostopis juncea, 831 Israel, O. 878, 889, 892, 893, 894, 913 Italian Clover, 118 Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 29, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 764 Ironweod, 755 Ivy, 620 Medicinal plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 867 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Pepsermint, 681 Potato, 710 Sumac, 608 Walnut, 401 Yam, 374 Isase, 683 Cape, 742 Carolina, Alkaloids of, 147 False, 129 Large-flowered, f. 684 Oil, 86 Yellow, 688-689 Isonoteria, 846 Isopulrin, 464 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum funce, 831 Israel, O. 878, 889, 892, 893, 894, 913 Italien Clover, 118 Millet, 350 Pharmacopoeia, 866 Ry Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Pium, 505, 514 Poisonous plants of, 867 Ispusiona callents, 67 Iuglandic acid, 401 Iuglandic acid, 401 Iuglandic acid, 401 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglan | versicolor, 105, f. 388, 389- | Jamestown Weed, 732 | Jorissen, A., 893 |
| Iron, Salts of, 7 Ironweed, 765 Ironweod, 395, 681 Irritants, 76, 804 Irving, 572, 892 Isaria farinosa, 273 Isatis corniculata, 818 tinctoria, 846 Isonandra, 53 Isopilocarpin, 148 Isopyrum fumarioides, 846 thalictroides, 446, 846 Isorotterin, 589 Isotoma axillaris, 812 Brownii, 812 Brownii, 812 Isostopis juncea, 831 Israel, O. 878, 889, 892, 893, 894, 913 Italian Clover, 118 Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 29, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 764 Ironweod, 755 Ivy, 620 Medicinal plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 867 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Pepsermint, 681 Potato, 710 Sumac, 608 Walnut, 401 Yam, 374 Isase, 683 Cape, 742 Carolina, Alkaloids of, 147 False, 129 Large-flowered, f. 684 Oil, 86 Yellow, 688-689 Isonoteria, 846 Isopulrin, 464 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum funce, 831 Israel, O. 878, 889, 892, 893, 894, 913 Italien Clover, 118 Millet, 350 Pharmacopoeia, 866 Ry Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Pium, 505, 514 Poisonous plants of, 867 Ispusiona callents, 67 Iuglandic acid, 401 Iuglandic acid, 401 Iuglandic acid, 401 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglan | 390, 828 | | Juch, K. W., 893 |
| Iron, Salts of, 7 Ironweed, 765 Ironweod, 395, 681 Irritants, 76, 804 Irving, 572, 892 Isaria farinosa, 273 Isatis corniculata, 818 tinctoria, 846 Isonandra, 53 Isopilocarpin, 148 Isopyrum fumarioides, 846 thalictroides, 446, 846 Isorotterin, 589 Isotoma axillaris, 812 Brownii, 812 Brownii, 812 Isostopis juncea, 831 Israel, O. 878, 889, 892, 893, 894, 913 Italian Clover, 118 Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 29, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 764 Ironweod, 755 Ivy, 620 Medicinal plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 867 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Peppermint, 709 Pepsermint, 681 Potato, 710 Sumac, 608 Walnut, 401 Yam, 374 Isase, 683 Cape, 742 Carolina, Alkaloids of, 147 False, 129 Large-flowered, f. 684 Oil, 86 Yellow, 688-689 Isonoteria, 846 Isopulrin, 464 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum fumarioides, 846 Isopyrum funce, 831 Israel, O. 878, 889, 892, 893, 894, 913 Italien Clover, 118 Millet, 350 Pharmacopoeia, 866 Ry Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Pium, 505, 514 Poisonous plants of, 867 Ispusiona callents, 67 Iuglandic acid, 401 Iuglandic acid, 401 Iuglandic acid, 401 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglandic, 105 Iuglan | Irish, H. C., 714, 725 | | Judas Tree, 527 |
| Ironweed, 765 Ironweod, 395, 681 Irritants, 76, 804 Irving, 572, 892 Isaria farinosa, 273 Isatis corniculata, 818 tinctoria, 846 isonandra, 53 Isopilocarpin, 148 Isopyrum fumarioides, 846 thalictroides, 446, 846 Isorotterin, 589 Isotoma axillaris, 812 Brownii, 812 Isotoma axillaris, 812 Brownii, 812 Isostropis juncea, 831 Israel, O. 878, 889, 892, 893, 894, 913 Italian Clover, 118 Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 29, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 219 Medicinal plants of, 866 Plum, 505, 514 Poisonous plants of, 866 Plum, 505, 514 Poisonous plants of, 867 Ispame, 687 Peppermint, 709 Peppermint, | Trisin, 389 | Japan Clover, 525 | |
| Irving | Tron, Saits of, 7 | Ivv. 620 | |
| Irving | Iron-wood 305 681 | Medicinal plants of, 866 | |
| Irving | Irritants, 76, 804 | Plum, 505, 514 | |
| Isatia farinosa, 273 | Irving, 572, 892 | Poisonous plants of, our | |
| Isatis corniculata, 818 | Isaria farinosa, 273 | Japanese Aralia, 647 | |
| tinctoria, 846 | | Peppermint, 709 | |
| Sopilocarpin, 148 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Walnut, 401 Sumac, 608 Sieboldiana, 401 Suite, 621 Chinese, 621 Suncace, 812 Sumac, 608 Sumac, 608 Sumac, 608 Sumac, 608 Sumac, 608 Sumac, 608 Walnut, 401 Sumac, 608 Sumac, 608 Sumac, 608 Sumac, 608 Sumac, 608 Sumac, 608 Walnut, 401 Suite, 621 Chinese, 621 Suncace, | | | nigra, 105, f. 400 |
| Wainut, 401 Yam, 374 Junube, 621 Chinese, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 Juncate In Junibe, 621 | | | regia, 401, 828 |
| Yam, 374 Jasmine, 683 Chinese, 621 Juncaceae, 374 Juncus tennis, f. 373, 374 Juncus tennis, f. 4 Juncus tennis, f. 4 Juncus tennis, f. 4 Juncus tennis, f. 373, 374 Juncus tennis, f. 4 Juncus tennis, | | Walnut 401 | Sieboldiana, 401 |
| Jasmine, 683 | Isopyrin, 446 | | Jujube, 621 |
| Solution | Isopyrum finarioides, 840 | Jasmine, 683 | Chinese, 621 |
| Solution | | Cape, 742 · | |
| Brownii, 812 Iongiflora, 812 Israel, O., 878, 889, 892, 893, 894, 913 Italian Clover, 118 Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 99, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 764 Targe-flowered, f. 684 Oil, 86 Valley, 688-689 Jasminium floribundum, 840 grandiflorum, 683, f. 684 officinale, 840 Sambac, 840 Jateorrhiza Columba, 473 Jatropha, 52, 590, 596-597, 865 Italia, L. van, 892 Itch, Barber's, 100, 300 Grandiflorum, 683, f. 684 Oil, 86 Sambac, 840 Jateorrhiza Columba, 473 Jatropha, 52, 590, 596-597, 865 Italian, L. van, 892 Itch, Barber's, 100, 300 Jaminium floribundum, 840 Sambac, 840 Jateorrhiza Columba, 473 Jatropha, 52, 590, 596-597, 865 Italian, L. van, 892 Itch, Barber's, 100, 300 Jaminium floribundum, 840 Sambac, 840 Jateorrhiza Columba, 473 Jatropha, 53 Curcas, 587, 595, 596, 823 gossypifolia, 587 macrorhiza, 823 Monihot, 75 multifida, 823 stimuloso, 77, 122, 596, f. 331, 817 Jurubebin, 715 Justicia Gendarussa, 804 | Isorottierin, 589 | Carolina, Alkaloids of, 147 | Juncus tennis, f. 373, 374 |
| longiflora, 812 Oil, 86 S8-689 S84, 913 Istalian Clover, 118 Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Italie, L. van, 892 Italie, L. van, 892 Italie, L. van, 892 Italie, L. van, 892 Italie, L. van, 892 Italie, L. van, 892 Italie, L. van, 894 Italie, L. van, 895 Italie, L. van, 895 Italie, L. van, 896 Monihot, 75 M | | False, 129 | Juniper, 330-332 |
| Isotropis juncea, 831 Israel, O., 878, 889, 892, 893, 894, 913 Italian Clover, 118 Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 29, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 2010, 68-689 Jaminium floribundum, 840 grandiforum, 683, f. 684 officinale, 840 Sambac, 840 Jateorrhiza Columba, 473 Jatropha, 52, 590, 596-597, 865 angustidens, 53 Curcas, 587, 595, 596, 823 glandulifera, 823 glandulifera, 823 macrorhiza, 823 Ithyophallus inpudicus, 29, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. Tarabara Angustidens, 77 multifida, 823 stimuloso, 77, 122, 596, f. sopulorum, 101, 330 virginiana, 101, 330-331, f. Jurubebin, 715 Justicia Gendarussa, 804 | | | Common 101 330-331 |
| Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 99, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 764 Ava Bean, 574 | | Vellow 688-680 | Low 332 |
| Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 99, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 764 Ava Bean, 574 | Israel, O., 878, 889, 892, 893, | | Oil of 101 |
| Millet, 350 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 99, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 764 Ava Bean, 574 | 894, 913 | | Savin, f. 331, 332 |
| Name C, 840 Pharmacopoeia, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f, 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 99, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. To be a marked by the contains a communis v. alpina, 330 horizontalis, 102 Knighti, 332 Knighti, 332 Curcas, 587, 595, 596, 823 gossypifolia, 587 moltifida, 823 stimuloso, 77, 122, 596, f. axillaris, 763 xanthiifolia, 137, 763-764, f. To be a marked by the communis v. alpina, 330 horizontalis, 102, 331 Oxycedrus, 330, 816 Sabina, 4, 102, 149, f. 33t, 332, 816 scopulorum, 101, 330 virginiana, 101, 330-331, f. 332, 816 Sabina, 4, 102, 149, f. 33t, 32, 816 scopulorum, 101, 330 virginiana, 101, 330-331, f. 331, 817 Jurubebin, 715 Justicia Gendarussa, 804 | Italian Clover, 118 | | Juniperus, 4, 102, 149, 330-332 |
| Pharmacopoeta, 866 Rye Grass, 361 Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f, 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 99, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 764 Ideorrhiza Columba, 473 Autropha, 52, 590, 596-597, 865 angustidens, 53, 596-597, 865 Alerors, 596, 597, 596, 823 Curcas, 587, 595, 596, 823 glandulifera, 823 gossypifolia, 587 macrorhiza, 823 Monihot, 75 multifida, 823 stimuloso, 77, 122, 596, f. 331, 817 Jurubebin, 715 Justicia Gendarussa, 804 Lava Bean, 574 Communis v. alpina, 330 Arientha, 473 Arientha, | | | communis, 101, 330, 332 |
| Italie, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 99, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. Italie, L. van, 892 Ingustidens, 53 Curcas, 587, 595, 596, 823 gossypifolia, 587 macrorhiza, 823 Monihot, 75 multifida, 823 stimuloso, 77, 122, 596, f. 331, 817 Jurubebin, 715 Justicia Gendarussa, 804 Lava Bean, 574 Knightii, 332 occidentalis, 102, 331 Oxycedrus, 330, 816 Sabina, 4, 102, 149, f. 33t, 33, 816 scopulorum, 101, 330 virginiana, 101, 330 virginiana, 101, 330-331, f. Jurubebin, 715 Justicia Gendarussa, 804 | | Jateorrhiza Columba, 473 | communis v. alpina, 330 |
| Tatle, L. van, 892 Itch, Barber's, 100, 300 Fungus causing, f. 300 Dhobic, 299 Disease of Horses, 293-294 Ithyophallus inpudicus, 99, 860 Iva, 757, 763-764 axillaris, 763 xanthiifolia, 137, 763-764, f. 2064 Iva Bean, 574 angustidens, 53 Curcas, 587, 595, 596, 823 glandulifera, 823 gossypifolia, 587 macrorhiza, 823 Monihot, 75 multifida, 823 stimuloso, 77, 122, 596, f. 331, 817 Jurubebin, 715 Justicia Gendarussa, 804 Iva Bean, 574 Iurube 1, 332 Angustidens, 53 Curcas, 587, 595, 596, 823 gossypifolia, 587 macrorhiza, 823 stimuloso, 77, 122, 596, f. 331, 817 Jurubebin, 715 Justicia Gendarussa, 804 | | Jatropha, 52, 590, 596-597, 865 | |
| Disease of Horses, 293-294 gossypt ofta, 387 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 817 330, 331, 61 330, 33 | Italie, L. van, 892 | angustidens, 53 | Knightii, 332 |
| Disease of Horses, 293-294 gossypt ofta, 387 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 817 330, 331, 61 330, 33 | Fungus couring f | Curcas, 587, 595, 596, 823 | Ornedrus 220 916 |
| Disease of Horses, 293-294 gossypt ofta, 387 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 816 332, 817 330, 331, 61 330, 33 | Dhobic 200 | gianaulijera, 823 | Sahina 4 102 149 f 221 |
| 1tty optialtus in pudicus, 194, Monifict, 75 Scopulorum, 101, 330 Wirginiana, 101, 330 Wirgin | Disease of Horses 203,204 | gossypijona, 587 | 332. 816 |
| axillaris, 763 596 Jurubebin, 715 xanthiifolia, 137, 763-764, f. urens, 596, 823 Justicia Gendarussa, 804 764 Java Bean, 574 Jute, 621 | Ithyophallus inbudicus, 29. | Monihot. 75 | scopulorum, 101, 330 |
| axillaris, 763 596 Jurubebin, 715 xanthiifolia, 137, 763-764, f. urens, 596, 823 Justicia Gendarussa, 804 764 Java Bean, 574 Jute, 621 | 860 | multifida, 823 | virginiana, 101, 330-331, f. |
| axillaris, 763 596 Jurubebin, 715 xanthiifolia, 137, 763-764, f. urens, 596, 823 Justicia Gendarussa, 804 764 Java Bean, 574 Jute, 621 | Iva, 757, 763-764 | stimuloso, 77, 122, 596, f. | 331, 817 |
| xanthiifolia, 137, 763-764, f. urens, 596, 823 Justicia Gendarussa, 804 764 Java Bean, 574 Jute, 621 | axillaris, 763 | 596 | Jurubebin, 715 |
| Tvory, Vegetable, 370 Java Bean, 574 Cardamon, 391 Jute, 621 Plant, f. 622 | xanthiifolia, 137, 763-764, f. | urens, 596, 823 | |
| Ivory, Vegetable, 5/0 Cardamon, 391 Plant, f. 022 | 764 | | Jute, 621 |
| | Ivory, vegetable, 3/0 | Cardamon, 391 | riant, J. 022 |

Kabsch, W., 893 Kaempferia Galanga, 391 rotunda, 391, 852 Kaffir Corn, 345, 346, 347, 348, 864 Kageneckia angustifolia, 503, Kageneckia angustifolia, 503, 847
oblanga, 847
Kahlenberg, L., 893
Kairin, 147
Kairolin, 147
Kairolin, 147
Kalanchae spathulata, 818
Kalm, P., 893
Kalmia, 666, 667, 669-672, 865
angustifolia, 127, 669, 670, f. 671-672, 821
glanca, 821
hirsnta, 821
latifolia, 64, 65, 127, 665, 669-671, f. 670, 821
polifalia, 127, 672
Kalm's I,obelia, 136
St. John's-wort, 629
Kalo Nut, 587
Kamala, 589
Kamas, 589
Kamas, 589
Kapok, 695
Oil, 624
Seed, 621
Tree, 621
Karatas Plumieri, 811
Karsten, G., 893, 912
Kartulis, 893
Karwinskia Humbaldtiana, 124, 621, 847
Kassner, 724 847 Karsten, G., 893, 912
Kartulis, 893
Karwinskia Humbaldtiana, 124, 621, 847
Kassner, 724
Kauffman, C. H., 209, 260
Kaufmann, P., 893
Kauter, 637
Kava-kava, 396
Kebler, L. F., 64, 65, 893
Kedzie, R. C., 213
Keeler, D. D., 893
Kellerman, K. F., 94, 893
Kellogg, A., 893
Kellogg, Harriette S., 803, 862, 869
Ketty, W. A., 219
Kennan, George, 34
Kennedy, G. W., 893
Kennedy, P. B., 96, 467, 880, 893, 894
Kennedy, W. J., 424
Kentucky Coffee Tree, f. 116, 117, 527, 537, f. 538
Kenyah Dart Poison, 865
Keratiris, 354
Purulent, 263
Keratorwocosis, 265, 266 Keratitis, 354
Purulent, 263
Reratomycosis, 265, 266
Rerner, Anton von, 894, 901
Kerner, I. S., 894
Kew Garden, 501
Kerria japanica, 504
Khittel, J., 894
Kibara angustifalia, 838
Kicksia, 691
Kidney Bean, Three-lobed, 520 Kicks. Kidney 1: 520 520
Kiggelaria africana, 53, 627
Kilborne, F. L., 215
Kimanga, 865
King, F. H., 357
King, John, 894, 897
Kingsery, Mrs. A., 654
Kingsbury, Capt., 358
Kingsley, Charles, 587
Kingsley, Dr., 363
Kingsley, A. T., 894
Kinnebrew, C., 894, 899
Kinnikinnik, 77

Kino, 523, 640 Kirondro Poison, 865 Kirtikar, K. R., 372, 376, 391, 698, 751, 804, 894 Kisanin, 386 Kissenia, 633 Kissoumpa, 865 Kitt, Theo., 894 Klebahn, H., 229 Klebs, 894 Klebahn, H., 229
Klebs, 894
Klemperer, F., 302
Klingman, Th., 894
Knapweed, 141, 802
Knightia excelsa, 844
Knolle, 894
Knotweed, Dooryard, 423
Saghalen, 421
Knowles, M. E., 894
Knowlton, F. H., 894
Knowlton, F. H., 894
Knawltania vesicataria, 846
Knotgrass, 421
Kobert, R., 1, 4, 7, 28, 35, 62, 72, 77, 79-81, 96, 237, 238, 241, 242, 277, 278, 279, 505, 572, 889, 894
Köbner, 298
Köbner, 298
Kobus, 621
Koch, D. G. D. J., 894
Kochia arenaria, 426, 814
scaparia, 107, 426, 814
trichaphylla, 814
Koda Millet, 349
Koelrenteria, 89
panicialata, 607, 850
Koenig, A., 894
Kohln-Abrest, F., 894
Kokra, 865
Kolmin, 621 Kohn-Abrest, F., 894
Kokra, 865
Kolanin, 621
Kalarca, 621
Koltjar, E., 298
Komanga, 865
Koochia Tree, 685
Kooco, 505, f. 506
Koto, P. O., 721
Kraemer, IL, 2, 724, 755, 863, 894
Kraemer, It, 12, 724, 755, 863, 894
Kraemeria triandra 843 894
Krameria triandra, 843
Krapf, K. von, 894
Krasheninnikoff, 33
Kraus, R. V., 894, 896
Krause, A., 862, 895, 900
Kremers, Edward, 86, 88, 662
Kren, 895, 910
Kreutzer, K. J., 895
Krocker, 895
Krusterstern M. de, 341 Krocker, 895
Krustenstern, M. de, 341
Kruskal, 440
Ksopo, 865
Kubach, Oscar, 771
Kubingi, 895
Kudzu, 526
Kulti, 526
Kumquat, 582
Kunkel, A. J., 895
Kurrinia zeylanica, 53, 813
Kyle, H. M., 895
Kyllingia monocephala, 819

La Calve, 16
La Glesch, L., 895
Labesse, 895
Labrata, 52, 131, 698, 708-713, 828
Labrann, J. D., 890, 895
Labrador Tea, 127, 629, 666
Laburnum, 72, 148, 527
anagyraides, 527, 551, 831
Lac, Japanese, 865
Lace-bark, 623, 642
Lacerda, J. B. de., 895

Lacuanthes, 385 tinctaria, 104, 386 Lacquer, Source of, 608 Lactarius, 90 actarins, 90 acris, 860 atraviridis, 860 campharatus, 860 insularis, 860 pergamenus, 860 rubus, 860 scrabiculatus, 860 scrabiculatus, 860 subdulcis, 860 tarminasis, 860 trivialis, 860 turpis, 860 vellereus, 860 Lactone, 149 Lactuca, 755, 757, 760-762 canadensis, 761, 762 pulchella, 761-762 sativa, 756, 760 Scariala, 137, 760-761, f. sativa, 76, 760
Scariala, 137, 760-761, f.
76t
Scariala v. integra, 137, 816
taraxacifalia, 816
taitarica, 816
virosa, 5, 762, 861
Lactucarium, 755, 756, 762
Lactucopicrin, 755, 762
Lactucopicrin, 755, 762
Lactucorin, 762
Lactucin, 762
Lactucin, 762
Ladd, E. F., 352, 895
Lady Laurel, 642-643, f. 643
Lady's Slipper
Showy, 393-395, f. 304
Small White, 105, 393
Small Yellow, 105, 393
Thistle, 802
Thumb, 421, f. 422
"La face vulteuse," 35
Lefitte, H., 895
Laguaria vulgaris, 750, 751, Lagenaria vulgaris, 750, 751, 819 Flas-reginae, Lagerstroemia 837 indica, 637
Lagetta lintearia, 642
Lagachilus inebrians, 828
Lake Shore Disease, 344
Lakes Working of, 91-93 I,ake Shore Disease, 344
Lakes, Working of, 91-93
Lambert, L., 895, 901
Lambkill, 127, 669, 671
Lamb's Quarter, 427, 433
Laminn, 710, 712
album, 712, f. 712
amplextcaule, 712, f. 7
Laminaria digitata, 194
Landalphia, 691
Lane, 714
Lance-leaved Sage, 710
Lang, W. H., 895, 912
Langebeck, 302
Langebeck, 302
Langebeck, 302 Langenbeck, Langsdorff, F. E. L., 34 Lansium domesticum, 837 Lantana, 708 salviaefolia, 858 sattaefola, 658 Lanthopin, 481 Laportea, 410, 413-414 canadensis, 77, 106, 414, f. 414, 857 crenulata, 414, 857 Gandichaudiana, 857 gigas, 414 stimulans, 2, 857 stimulasa, 857

Lappa majar, 67

| Lappakonitin, 450 | Cherry, 85, 117, 503, 504 | LEITNERIA, 400 |
|--|---|---|
| Lappin, 755 | Cherry, 85, 117, 503, 504 Family, 112 Indian, 478 | floridana, 400, f. 401 |
| Lappin, 755 Lappula, 706 | Ludian, 478 | LEITNERIACEAE, 400 |
| echinata, 707 | 1 adv 642.643 f 642 | Leitneriales, 156, 400 |
| floribunda, 707 | 1.ady, 642-643, f. 643 Mountain, 64, 84, 665, 669- | Lejolisia mediterranea, f. 194 |
| officinalis 131 | 671 f 620 | I.cinna, 372 |
| officinalis, 131 | 671, f. 670 Great, f. 666, 667 | |
| virginiana, 707 | Spurge, 125, 642-643, f. | I,EMNACEAE, 372 |
| Large Arrow-head, 102 | | Lemon, 582 |
| Blue Flag, 105 | 643 | Grass Oil, 344 |
| Cranberry, 665 | Swamp, 672 Laurent, E., 895 | Oil of, 583 |
| Peppergrass, 494 | Laurent, E., 893 | Salts of, 865 |
| Rhododendron, 127 | Laurie aeid, 593 | Verbena, 708 |
| Spotted Spurge, 597, f. 602 Tupelo, 664 White Wild Indigo, 117, | Laurocerasin, 503 | Lens esculenta, 521 |
| Tupelo, 664 | Lauro-eerasus, 865 | LENTIBULARIACEAE, 698, 833 |
| White Wild Indigo, 117, | Laurotetanin, 478 | Lentil, 521 |
| 541, f. 541 Large-bracted Wild Indigo. | Laurus canariensis, 478 | Lentinus stypticus, 860 |
| Large-bracted Wild Indigo. | nobilis, 478, 828 | suavissimus, 860 |
| 541 | Lavandula Spica, 709 | Leonia glycycarpa, 632 |
| Large-flowered Jasmine, f. | vera, 86, 709 Lavender, 709 Oil of, 709 | Leontice, 89 |
| 684 | Lavender, 709 | Leontopetalum, 810 Leonurus, 710, 711-712 |
| Trillium, 104 | Oil of, 709 | Leonurus, 710, 711-712 |
| Wake-robin, 385 | I.avinder, C. II., 343, 895 | Cardiaca, 131, 711-712, 828 Lepidogathis Wightiana, 804 |
| I.arkspur, 43, f. 47, 108-109, | Lavoisier, 3 | Lepidogathis Wightiana, 804 |
| I.arkspur, 43, f. 47, 108-109, 865, 868 | Law, James, 259, 277, 296, 293, 895 | Lepidium, 52, 487, 493-494 |
| Blue, 460, 466 | 293, 895 | apetalum, 113, 487, f. 404, |
| Carolina, 108, f. 461 | Lawrence, G. W., 245 | 494 |
| Field, 108, 460 | Lawrence, H., 324 | campestre, 493 |
| Great-flowered, 460 | Lawsonia inermis, 638 | Draba, 818 |
| Mountain, Tall, 109 Poisoning, 44-45, 465-467 | Laxative, 804 | latifolium, 818 |
| Poisoning 44.45 465.467 | Lazell, F. L. 895 | oleraceum, 818 |
| | Le Count E. R. 305 306 | Owaihiense, 818 |
| Proirie 460 461 | L'Engle Dr 665 896 906 | hiscidium 919 |
| Prairie, 460, 461 Purple, 44, 84, f. 461, 462- 463, 466, 467 Reset 460 | Lazell, F. J., 895 Le Count, E. R., 305, 306 L'Engle, Dr., 665, 896, 906 Le Grand, Dr., 793 | piscidium, 818 sativum, 53, 113, 818 virginicum, 487, 494 Lepidoptera, Fungi upon, 273 |
| 1 di pie, 44, 64, 1. 401, 402- | Lead Poisoning 6 72 | sirrainiaum 497 404 |
| Pooles 460 | Lead Poisoning, 6, 72 | Legidenters Euroi was 272 |
| MOCKEL, 400 | Leather I W 005 | Lepidoptera, Fungi upon, 273 |
| sp., 446 | Toothorwood 125 C42 6 C44 | Lepiota, 243, 865 |
| Tall, 108, 462, 463, f. 464, | Lead-color Puffball, f. 246 Leather, J. W., 895 Leatherwood, 125, 643, f. 644 Leaves, Wilted, 102, 116, 119 Leber, Th., 263, 265 | Morgani, 99, 243, 860 |
| 405, 400, 40/ | Leaves, Writed, 102, 116, 119 | nausina, 240 |
| Toxicity of, 84, 467 | 1,eber, 1 n., 263, 265 | procera, 235, 243 |
| Western Purple, 109 | Lecanora, 307 | Smooth, 240 |
| Wyoming, 109 | esculenta, f. 308 | Toadstool, 243 |
| Larrea mexicana, 858 | subfusca, f. 308 | Leprosy, 180, 181 |
| Larsen, C., 895, 899 Laserpitin, 648 | LECANORACEAE, 307 | Leptandra, 89 |
| Laserpitin, 648 | Lechea minor, 627 | Leptandrin, 733, 736 Leptomitus lacteus, f. 208 Leptosphaeria tritici, 286 |
| Lascrpitium latifolium, 648 | Leelere, N., 895 | Leptomitus lacteus, f. 208 |
| Lasia aculeata, 808 | LECYTHIDACEAE, 638 | Leptosphaeria tritici, 286 |
| Lasiadenia rupestris, 855 | Fish poisons among, 2 | Lessertia, 533, 534 |
| Lasiosiphon anthylloides, 355 | Lecythis, 638 | Lessertia, 533, 534 |
| eriocepholus 255 | allaria, 638 amara, 839 | annularis, 831 |
| Latex, Poison elaborated in, | amara, 839 | Lettuee, 756, 760-762 Blue, 761-762 |
| 9.2 | lanceolata, 839 | Blue. 761-762 |
| 82 | *************************************** | |
| | Ledum, 666 | Cultivated, 756, 762 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 | Ledum, 666 | Cultivated, 756, 762 European Priekly, 756 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 | Ledum, 666 Catesbaei, 127 | Cultivated, 756, 762 European Priekly, 756 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus, See Lathyrism | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76t |
| Lathyrism, 44, 45-46, 80, 534, 572-573 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76t |
| Lathyrism 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrus. 87, 149, 535, 572-573 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1, 52 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrus, 87, 149, 535, 572-573 amoenus, 831 Arhaca, 831, 865 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrus, 87, 149, 535, 572-573 amoenus, 831 Arhaca, 831, 865 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 laifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1, 52 glauca, 831 Leuein, 277 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrus, 87, 149, 535, 572-573 amoenus, 831 Arhaca, 831, 865 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Versia lenticularie f 238 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Versia lenticularie f 238 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 761 Wild, 761, 762 Leucaena, 1, 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocryue, 834 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrus, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicra, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 laifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 761 Wild, 761, 762 Leucaena, 1, 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocryue, 834 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrus, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicra, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 laifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1, 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocrinum, 834 Leucocrinum, 104 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 hirsutus, 572 orhrolencus, 572 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leucin, 277 Leucocidin, 168 Leucocoryuc, 834 Leucocriuum, 104 montanum, 104, 834 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Afhaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochrolencus, 572 adoratus, 831 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 76r Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocoryne, 834 Leucocrinum, 104 montanum, 104 1.eucocytes, 179 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 hirsulus, 572 orhrolencus, 572 adoratus, 831 oryatus, 572 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 Legumes, See Leguminosae | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leucocidin, 168 Leucocorinum, 168 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucocytes, 179 Leucocytosis, 169, 179 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochrobeneus, 572 adoratus, 831 ornatus, 831 ornatus, 572 palustris, 572 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 Legumes, See Leguminosae | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76r- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocoryue, 834 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucojum aestivum, 805 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris. 87, 149, 535, 572-573 amoenus, 831 Athaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochroleucus, 572 adoratus, 831 ornatus, 572 palustris, 572 polymarphus, 572 polymarphus, 572 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 Legumes, See Leguminosae | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 761 Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leucin, 277 Leucocidin, 168 Leucocoryuc, 834 Leucocryuc, 834 Leucocriuum, 104 montanum, 104, 834 Leucocytes, 179 Leucocytes, 179 Leucocytes, 179 Leucoimn, 805 vernum, 805 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicra, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 palymarphus, 572 purpureus, 831 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 Legumes, See Leguminosae Leguminosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76r Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocoryue, 834 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucojum aestivum, 805 vernum, 805 Leucona, 265 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrism See Lathyrism Lathyrism See Lathyrism Lathyrism See Lathyrism Lathyrism See Lathyrism Lathyrism See Lathyrism Lathyrism See Lathyrism Lathyrism See Lathyrism See Lathyrism Lathyrism See Lathyrism Lathyrism Medical See Lathyrism See Lathyrism Lathyrism Lathyrism See Lathyrism Lathyrism Lathyrism See Lathyrism Lathyrism Lathyrism Lathyrism See Lathyrism Lathyrism Lathyrism See Lathyrism Lathyrism Lathyrism Lathyrism Lathyrism See Lathyrism Lathyrism Lathyrism Lathyrism See Lathyrism See Lathyrism See Lathyrism See Lathyrism Lathyrism See | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Leguand du Saulle, II., 895 Legue, G., 896 Legue, G., 896 Legumes, See Leguminosae Leguminosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76r Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocoryue, 834 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucojum aestivum, 805 vernum, 805 Leucona, 265 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrism See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 hirsuitus, 572 ochroleucus, 572 aldoratus, 831 ornatus, 572 palymarphus, 572 polymarphus, 572 purpureus, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sylvestris, 572 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 laifolium, 629, 821 ralustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legrand du Saulle, II., 895 Legumes, See Leguminosae Legumnosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 Fconomie plants of, 520-527 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucocytes, 179 Leucocytes, 179 Leucojum aestivum, 805 vernum, 805 vernum, 805 Leucoma, 265 Leucoma, 265 Leucoma, 10, 865 LEUCOSPOREAE, 235 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochroleneus, 572 adoratus, 831 ornatus, 572 palustris, 572 polymarphus, 572 purpureus, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 556 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Leguand du Saulle, II., 895 Legue, G., 896 Legumes, See Leguminosae Leguminosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 Economie plants of, 520-527 Fish poisons of, 1 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76r Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocryne, 834 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucojum aestivum, 805 vernum, 805 Leucoma, 265 Leucomains, 10, 865 LeuCOSPOREAE, 235 Leucothoe, 667, 672-673 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrism See Lathyrism Lathyris. 87, 149, 535, 572-573 amoenus, 831 Athaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochroleucus, 572 adoratus, 831 ornatus, 572 palystris, 572 polymarphus, 572 purpureus, 831 sativus, 45, 534, 831 sylvestris, 572 tingitanus, 526 yenosus, 525, 572 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 laifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legurand du Saulle, II., 895 Legure, G., 896 Legumes, See Leguminosae Legumnosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 863 Arrow poisons among, 1 Feonomie plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 761- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocoryne, 834 Leucocryne, 834 Leucocrinum, 104 montanum, 104 montanum, 104, 834 Leucocytes, 179 Leucocytes, 169, 179 Leucojum aestivum, 805 vernum, 805 Leucoma, 265 Leucoma, 265 Leucomains, 10, 865 LEUCOSPOREAE, 235 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrism See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 palymarphus, 572 purpurcus, 831 salicus, 45, 534, 831 salicus, 45, 534, 831 salicus, 45, 532 tingitanus, 526 venosus, 525, 572 Latte, 895 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Lees, E, H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 Legumes, See Leguminosae Legumnosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 Economie plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leucin, 277 Leucocidin, 168 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucocytosis, 169, 179 Leucojum aestivum, 805 vernum, 805 Leucoma, 265 Leucoma, 265 Leucoma, 265 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, 821 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Afhaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jhirsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 831 saticus, 45, 534, 831 sylvestris, 572 tingitanus, 526 venosus, 525, 572 Latte, 895 Lutia veneuosa, 854 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Leguand du Saulle, II., 895 Legue, G., 896 Legumes, See Leguminosae Leguminosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 Economie plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 76r Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocryne, 834 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucojum aestirum, 805 ternum, 805 Leucoma, 265 Leucoma, 265 Leucomains, 10, 865 LEUCOSPOREAE, 235 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, 821 rocemosa, 128, 673, f. 673, |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrism See Lathyrism Lathyris. 87, 149, 535, 572-573 amoenus, 831. 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 birsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 831 sativus, 45, 534, 831 sativus, 47, 481 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 laifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 Legumes, See Leguminosae Leguminosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 863 Arrow poisons among, 1 Economic plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 Ornamental plants of, 527 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 761 Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leucin, 277 Leucocidin, 168 Leucocrynic, 834 Leucocrynic, 834 Leucocrynic, 834 Leucocytes, 179 Leucocytes, 179 Leucocytes, 169, 179 Leucojum aestivum, 805 vernim, 805 Leucoma, 265 Leucoma, 265 Leucoma, 265 Leucomains, 10, 865 LEUCOSPOREAE, 235 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, 821 rocemosa, 128, 673, f. 673, 821 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 572 purpureus, 831 sativus, 45, 534, 831 sylvestris, 572 tingitanus, 526 venosus, 525, 572 Latte, 895 Lutua venenosa, 854 Laudanin, 147, 481 Laudanosin, 481 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 Legumes, See Leguminosae Legumnosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 Feonomie plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 Ornamental plants of, 527 Poisonous plants of, 530- | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76r Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucocytes, 169, 179 Leucojum aestivum, 805 rernum, 805 Leucoma, 265 Leucomains, 10, 865 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, 821 rocemasa, 128, 673, f. 673, 821 Swamp, 673, f. 673 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris, 87, 149, 535, 572-573 amoenus, 831 Arhaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochroleneus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 572 purpureus, 831 sativus, 45, 534, 831 sylvestris, 572 tingitanus, 556 venosus, 525, 572 Latte, 895 Lutta veneuosa, 854 Laudanin, 147, 481 Laudanosin, 481 Laudanosin, 481 Laudanosin, 481 Laudanosin, 481 Laudanosin, 427 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legurand du Saulle, II., 895 Legure, G., 896 Legumes, See Leguminosae Legumnosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 863 Arrow poisons among, 1 Economic plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 Ornamental plants of, 527 Poisonous plants of, 530- 535 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 761 Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocoryne, 834 Leucocryne, 834 Leucocryne, 834 Leucocytess, 179 Leucocytess, 179 Leucocytess, 169, 179 Leucojum aestivum, 805 vernum, 805 Leucoma, 265 Leucoma, 265 Leucoma, 10, 865 LEUCOSPOREAE, 235 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, 821 rocenosa, 128, 673, f. 673, 821 Swamp, 673, f. 673 Leukemia, Infectious, 177 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyrism See Lathyrism Lathyris. 87, 149, 535, 572-573 amoenus, 831. 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 572 polymarphus, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 544 Laudanin, 526 Lutua veneuosa, 854 Laudanin, 147, 481 Laudanosin, 481 Laudanosin, 481 Laudanoum, Poisoning, 6 | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 ralustre, 4, 666, 821 Leek, Garden, 383 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legrand du Saulle, II., 895 Legumes, See Leguminosae Legumnosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 Feonomie plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 Ornamental plants of, 527 Poisonous plants of, 530- 535 LEGUMINOSAE - PAPI- | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 761 Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 277 Leueocidin, 168 Leucocoryne, 834 Leucocryne, 834 Leucocryne, 834 Leucocytess, 179 Leucocytess, 179 Leucocytess, 169, 179 Leucojum aestivum, 805 vernum, 805 Leucoma, 265 Leucoma, 265 Leucoma, 10, 865 LEUCOSPOREAE, 235 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, 821 rocenosa, 128, 673, f. 673, 821 Swamp, 673, f. 673 Leukemia, Infectious, 177 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris. 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 572 purpureus, 831 saticus, 45, 534, 831 saticus, 45, 854, 854 saticus, 45, 854, 854 saticus, 45, 854, 854 sa | Ledum, 666 Catesbaei, 127 glandulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legue, G., 896 Legue, G., 896 Legumes, See Leguminosae Legumnsosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 F.conomic plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 Ornamental plants of, 530- 535 LEGUMINOSAE - PAPI- LIONACEAE, 53, 89 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76r Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leucin, 27 Leucocidin, 168 Leucocrinum, 104 montanum, 104, 834 Leucocytesis, 169, 179 Leucojum aestivum, 805 vernum, 805 Leucomains, 10, 865 Leuconsolum, 805 Leucomains, 10, 865 Leucomains, 10, 865 Leucomains, 10, 872 Catesbaei, 127, f. 672, 673, 821 Swamp, 673, f. 673 Leukemia, Infectious, 177 Leuterer, Dr., 896 Levaditi, A., 894, 896 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris. 87, 149, 535, 572-573 amoenus, 831 Athaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 orbrokeicus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 572 purpureus, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 sulvestris, 572 tingitanus, 526 venosus, 525, 572 Latte, 895 Lutta vencuosa, 854 Laudanosin, 481 Laudanoum, 627 Laudanum, 627 Laudanum, 627 Laudanum, 627 Laudanum, 627 Laudanosin, 481 Laudanosin, 481 Laudanosin, 481 Laudanoum, 627 Laudanoum, 627 Laudanoum, 627 Laudanoum, 627 Laudanoum, 412, 444, 477-473, | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 laifolium, 629, 821 palustre, 4, 666, 821 Leek, Garden, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legrand du Saulle, II., 895 Legumes, See Leguminosae Leguminosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 Economic plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 Ornamental plants of, 527 Poisonous plants of, 530- 535 LEGUMINOSAE - PAPI LIONACEAE, 53, 89 Lehmann, Otto, 328, 416, 446. | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 76r Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leucin, 277 Leucocidin, 168 Leucocryne, 834 Leucocryne, 834 Leucocryne, 834 Leucocryne, 834 Leucocytes, 179 Leucojum aestivum, 805 rernun, 805 Leucoma, 265 Leucoma 10, 865 LEUCOSPOREAE, 235 Leucomains, 10, 865 LEUCOSPOREAE, 235 Leucoma, 267 Catesbaei, 127, f. 672, 673, 821 rocunosa, 128, 673, f. 673, 821 Swamp, 673, f. 673 Leukemia, Infectious, 177 Leuterer, Dr., 896 Levaditi, A., 894, 896 Levaditi, A., 894, 896 Levant Wormseed, Oil of, 791 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris. 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 572 purpureus, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 47, 481 sativus, 47, 481 sativus, 47, 481 sativus, 47, 481 sativus, 47, 481 sativus, 48, 48, 48, 48, 48, 48, 48, 48, 48, 48 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Leers, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 Legums, See Leguminosae Leguminosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 Economic plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 Ornamental plants of, 527- Poisonous plants of, 530- 535 LEGUMINOSAE - PAPI LIONACEAE, 53, 89 Lehmann, Otto, 328, 416, 446, 589, 590, 606, 650, 666, | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leucin, 277 Leucocidin, 168 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucocytosis, 169, 179 Leucojum aestivum, 805 vernum, 805 Leucoma, 265 Leucoma, 265 Leucoma, 265 Leucoma, 265 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, 821 rocemosa, 128, 673, f. 673 Leukemia, Infectious, 177 Leuterer, Dr., 896 Levaditi, A., 894, 896 Levant Wormseed, Oil of, 791 Levi, G., 896 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrism See Lathyrism Lathyrism 87, 149, 535, 572-573 amoenus, 831 Afhaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jhirsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 572 polymarphus, 572 purpurcus, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534, 831 saticus, 45, 534 Laudanin, 147, 481 Laudanosin, 481 Laudanosin, 481 Laudanum, 627 Laudanum Poisoning, 6 Laurace pinnatifida, 816 Lauraceae, 112, 444, 477-478, 828 Laurel, 127, 478, 865 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Lees, F. H., 54 Lefebre, C., 890, 895 Legue, G., 896 Legumes, See Leguminosae Legumnosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 863 Arrow poisons among, 1 Economie plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 Ornamental plants of, 527 Poisonous plants of, 530- 535 LEGUMINOSAE - PAPI LIONACEAE, 53, 89 Lehmann, Otto, 328, 416, 446, 589, 590, 606, 650, 666, 719, 721, 756, 862, 896 | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Prickly, 137, 760-761, f. 76r Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leuein, 27 Leueocidin, 168 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucojum aestivum, 805 vernum, 805 Leucoma, 265 Leucoma, 265 Leucomains, 10, 865 LEUCOSPOREAE, 235 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, 821 rocemasa, 128, 673, f. 673, 821 Swamp, 673, f. 673 Leukemia, Infectious, 177 Leuterer, Dr., 896 Levant Wormseed, Oil of, 791 Levi, G., 896 Levant Wormseed, Oil of, 791 Levi, G., 896 Levaiticum officinale, 648 |
| Lathyrism, 44, 45-46, 80, 534, 572-573 Lathyrismus. See Lathyrism Lathyris. 87, 149, 535, 572-573 amoenus, 831 Aphaca, 831, 865 Cicera, 45, 534, 831 Clymenum, 45, 534, 572, 831 Jirsutus, 572 ochrolencus, 572 adoratus, 831 ornatus, 572 polymarphus, 572 polymarphus, 572 purpureus, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 45, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 46, 534, 831 sativus, 47, 534, 831 sativus, 46, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 534, 831 sativus, 47, 47, 481 sativus, 47, 481 sativus, 47, 481 sativus, 47, 481 sativus, 47, 481 sativus, 48, 48, 48, 48, 48, 48, 48, 48, 48, 48 | Ledum, 666 Catesbaei, 127 glaudulosum, 127, 667, 821 groenlandicum, 127, 666 latifolium, 629, 821 palustre, 4, 666, 821 Leek. Garden, 383 Wild, 104, 383 Leersia lenticularis, f. 338 Leers, F. H., 54 Lefebre, C., 890, 895 Legrand du Saulle, II., 895 Legue, G., 896 Legums, See Leguminosae Leguminosae, 52, 63, 89, 117- 120, 148, 395, 498, 500, 519-574, 828-833, 865 Arrow poisons among, 1 Economic plants of, 520-527 Fish poisons of, 1 Medicinal plants of, 527- 530 Mildews on, 269 Ornamental plants of, 527- Poisonous plants of, 530- 535 LEGUMINOSAE - PAPI LIONACEAE, 53, 89 Lehmann, Otto, 328, 416, 446, 589, 590, 606, 650, 666, | Cultivated, 756, 762 European Priekly, 756 Garden, 760 Priekly, 137, 760-761, f. 76t- Wild, 761, 762 Leucaena, 1. 52 glauca, 831 Leucin, 277 Leucocidin, 168 Leucocrinum, 104 montanum, 104, 834 Leucocytes, 179 Leucocytosis, 169, 179 Leucojum aestivum, 805 vernum, 805 Leucoma, 265 Leucoma, 265 Leucoma, 265 Leucoma, 265 Leucothoe, 667, 672-673 Catesbaei, 127, f. 672, 673, 821 rocemosa, 128, 673, f. 673 Leukemia, Infectious, 177 Leuterer, Dr., 896 Levaditi, A., 894, 896 Levant Wormseed, Oil of, 791 Levi, G., 896 |

Lewin, L., 637, 896
Lewis, L. L., 9
Lewisia rediviva, 423, 844
Liatris, 90, 757, 773-774
punctata, 773-774
pycnostachya, 773
spicata, 755, 773
squarrosa, 417
Liautard, A., 377, 383, 896
Licania hypolenca, 847
LICHENES, 154, 307, 861
ASCO-, 307
BASIDIO-, 307
Lichens, f. 307, 308, 309, 865 Lichens, f. 307, 308, 309, 865 Basideal, 307 Basideal, 307 Crustaceous, 307 Foliaceous, 307, f. 309 Fruticose, 307 Reindeer, 307 Lichtheim, 199 Lichtensteinia Beiliana, 856 pyrethifolia, 856 Licorice, 531 European, 527, f. ev Licorice, 531 European, 527, f. 528 Extract of, 527 Russian, 527 Wild, 531 Liebig, H. V., 505 Light, its effect upon toxicity of plants, 83 Lignum-vitae, 575 Ligouzat, 896 LIGULIFLORAE, 757, 758-762 Ligusticum canadense, 649 Ligusticum canadense, 649 Ligustrum, 683-684 robustum, 683, 684 vulgare, 128, 684, 840 Lilac, 683 Powdery mildew on, 267, 269 Liliaceae, 52, 63, 89, 103-104, 374, 375, 386, 417, 833-835 Lilienfeld, Fr., 896 LILIIFLORAE, 155, 374-390 Lilium superbum, 104 tigrinum, 375 Lily, 375 Atamasco, 105, 387-388, f. 388 Blackberry, 105 California, 375 California, 375 Day, 375 Family, 375-385 Mariposa, 375 Sacred, 444 Texas, 690 Tiger, 375 Turk's-cap, 104 Water, 445 Blue-flowered, 445 Vellow f. 445 Yellow, f. 445 Lily-of-the-Valley, 64, 104, 148, 375, 384-385, f. 384 Lima Bean, 75, 87, 119, 520, Lime, 582 Limettin, 583 Limeum, 824 LIMNANTHACEAE, 486 Limonium carolinianum, 128 Limonium carolintanium, 128 Limosella, 89 aquatica, 852 cymbalaria, 852 Elatine, 852 spuria, 852 LINACEAR, 53, 120, 575, 577, 580-581, 835-836 Linacol, 713 Linalool, 709 Linamarin, 120, 865

Linaria, 734, 735 vulgaris, 735, f. 735, 738, 865 865
Linariin, 735
Lindenberg, A., 896
Lindely, J., 5, 362, 385, 533, 738, 896
Lindau, Gustave, 301, 302
Lindaa Benzoin, 828
Lindsae acultrata, 825
Lindsae, W. L., 896
Lingon Berry, 665
Linin, 120
Linn Tree, 621
Linnaeus, See Linne, C. v.
Linne, Carl v., 277, 642, 643
Linossier, G., 302, 897, 908
Linoleic acid, 581
Linoxyn, 581
Linseed, 580, 621
Cake, 54
Linseed Oil, 581
Linum, 580-5581
catlarricum, 120, 835
grandiflorum, 580 Linariin, 735 grandiflorum, 580 perenne, 580 rigidum, 120, 581, 835 toxicum, 836 usitatissimum, 53, 120, 580
581, 836
Linuttin, 584
Lipicrre, 169
Lippia citriodora, 708
mexicana, 708
nodiflora, 708
Liquidanbar orientalis, 500
Styraciflua, 500, f. 501
Liriodendrin, 474
Liriodendrin, 474
Liriodendrin chinense, 837
tulipifera, 474, 837
List, A., 897
Lisurus borealis, 860
Litchi, 607
Lithosperamum, 704 usitatissimum, 53, 120, 580-Lithosperamum, 704 arvense, 228 officinale, 629 omeinate, 029 Lithraea venenosa, 805 Litmus, 307 Little, 897 Little Barley, 367, f. 368 Live Oak, 403 Live-for-ever, 501, 502, f. Liverwort, f. 309, 310, f. 311 Lizard's Tail, 396 Llanosia Tognian, 855 Lloyd, C. G., 863, 897 Lloyd, J. U., 96, 484, 894, Lloyds, J. U. and C. J., 452, 454, 456, 460, 465, 468, 471, 897 471, 897
Loasaceae, 125, 628, 633-634
Lobelacin, 752
Lobelia, 378, 752-753
Blue, 136
Breynii, 752
cardinalis, 136, 753, 812
erinus, 752
Family, 751-753
fulgens, 812
Great, 753
inflata, 72, 75, 136, 464,
752, 753, 812
Kalmii, 136, 812
Kalmii, 136, 812
Kalmis, 136
nicotianaefolia, 752, 812
Pale Spiked, 136
pratioides, 812
purpurascens, 752 purpurascens, 752 rhynchopetalum, 812 siphilitica, 136, 753, 812 spicata, 753, 812

Spiked, 753
Tupa, 812
Lurens, 812
LubeLinceAr, 748, 751-753
"Lobelia acid, 752
Lobelin, 136
Lobelin, 78, 752
Lobloliny Pine, 330
Lock-jaw, 174
Lockbart, Dr., 25
Loco acid, 565
Loco Disease, 501
Loco Disease, 501
Loco pisoning, 804 Loco Disease, 501 Loco poisoning, 804 Loco Weed, 6, 37, 39, 97, 533, 544, 562-570, 865 Distribution of, 563, 570 Loss from poisoning by, Purple, 563, 567-570, f. Purple, 563, 567-570, f.
568
Stemless, 39, 119, 566, 567-570, f. 568
Woolly, 39, 562, 567, 568
Locoism, 37-40, 80, 865
Locust, 526, 746,
559-560
Bark of, 55
Clammy, 119, 560
Flowers, 64, f. 66
Honey, 527
Poisoning, 56-58
Tree, 118, 527, 558, 559-560
of Jamaica, 523
of West Indies, 527
Lodcman, F. G., 897
Lodgepole Pine, 330
Loeselia coerulea, 843
Loew, O., 897
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania Family, 129
Logania, 587
Loganoid, 528 148, 683, 684-689, 836, 865
Loganin, 687
Logwood, 528
Lollin, 103
Lollin arvense, 363
festucaceum, 364
titalicum, 363, 364
Lamarckii, 826
linicolum, 363
multiflorum, 364
perenne, 364, 826
remotum, 364
temulentum, 4, 72, 74, 103, 361-364, 826
Lombroso, C., 290
Lonchocarpus, 1, 52
densiflorus, 831
ichthyoctonus, 831
latifolius, 831
rariflorus, 831
rariflorus, 831
rariflorus, 831
cong-smooth-fruited Poppy, 113
Long-awed Pine, 329
Long-leaved Pine, 329
Long-leaved Pine, 329
Long-leaved Pine, 329
Long-smooth-fruited Poppy, 113 Long-leaved Pine, 329 Lonicer, A., 275 Lonicera, 744 chrysantha, 812 flava, 744 nava, 744 fragrantissima, 744 involucrata, 135, 744, 812 japonica, 744, 812 Periclymenum, 744 sempervirens, 744 Standishii, 812

| Sullisantii 744 | hireutus 140 548 | McIlyaine C M 243 807 |
|--|---|---|
| Sullivantii, 744 tatarica, 744, 812 | lursutus, 149, 548 holosericeus, 548 | McIlvaine, C. M., 243, 897 McKelway, J. I., 897 MacMillan, Conway, 897 McMullen, Dr., 25 McNeil, Dr. J. H., 279, 546, 898, 902 MacQuean, 534, 552 MacQuean, James, 898 McWilliams, J. W., 782 Macadamia termifolia, 844 |
| tamentella, 812 | leucophyllus, 118, 548, 550, | MacMillan, Conway, 897 |
| Xylosteum, 812 | 831 | McMullen, Dr., 25 |
| Lephopetalum pallidum, 813 | linifolius, 548 | McNeil, Dr. J. H., 279, 546, |
| tatarica, 144, 812 tamentella, 812 Xylosteum, 812 Lophoptelum fallidum, 813 Lophophorin, 637 Lophophyton, 13 anliume, 14 | lutens, 149, 531, 546, 548, 831 | MacOwen 534 552 |
| gallinac, 14 | arnatus, 550 | MacQuean, James, 898 |
| LORANTHACEAE, 106, 415, 836 | perennis, 118, 546 | McWilliams, J. W., 782 |
| Lotase, 534 | perennis, 118, 546 Plattensis, 546, 550 | |
| Loturin, 681 | pusillus, 550 | Macaranga, 823 |
| Loturdin, 681 Lotus, 54, 90, 865 | sericeus, 550 Lupulic acid, 106 | Mace, 445 Maclura, 410, 865 |
| Bark, 681 | Lupulin, 406 | Maclura, 410, 865 pomifera, 406, 414-415 tinctoria, 406 Macrocystis pyrifera, 194 |
| arabiscus, 53, 534 australis, 534, 831 | Lutembacher, 897, 902 | tinctoria, 406 |
| australis, 534, 831 | Luteolin, 479, 736 Luzula, 374 | Macrocystis pyrifera, 194 |
| corniculatus, 534, 831 Egyptian, 445 | Luzuia, 374 Lychnidin, 439 | Macrodiplodia, 286 Macrospora cicadina, 204 |
| Sugreolens, 90, 551 | Lychnis, 63, 89, 436, 439, 865 | Macrosporium Brassicae, 861, |
| Stateolens, 90, 551 Tree, 681 Lotusin, 54, 534 Loudon, J. C., 587, 601, 642, 897 | Lychnis, 63, 89, 436, 439, 865 chalcedonica, 439, 813 | 865 |
| Lotusin, 54, 534 | dioica, 439, 813 | Macrozamia, 325, 819, 865 Madder, 524, 742 Family, 135, 741-744 Maderia Vine, 424 |
| Loudon, J. C., 587, 601, 642, | Evening, 439 | Madder, 524, 742 |
| Lousewort, 134, 734, 738 | Flos-cuculi, 439, f. 440, 813 indica, 813 | Maderia Vine. 424 |
| Mountain, 134 | Meadow, f. 440 | Madia glamerata, 138 |
| Mountain, 134 Swamp, 738 Lovage, 648, 649 Low Hop Clover, 553 | Meadow, f. 440 Scarlet, 439 Lycium, 717, 733 | Madia glamerata, 138 Oil, 755 |
| Lovage, 648, 649 | Lycium, 717, 733 | sativa. 755 |
| Low Hop Clover, 553 | barbarum, 854 chinense, 716 | Madrona, 003 |
| Juniper, 332 Lupine, 550 | halimifolium, 133, 716, 733 | Madrona, 665 Madsen, B. A., 898, 899 Madsen, Thorwald, 898 |
| Lubeman, 897 | Lycogola epidendron, f. 159, | Madura-foot, 184 |
| Lucas, E., 897 | 160 | Maerua angolensis, 812 |
| Lucet, A., 200, 202, 203, 877, 897 | LYCOPERDACEAE, 99, 245-247, | Maesa indica, 839 |
| Luco-encephalitis, 20 | 860 LYCOPERDINEΛE, 245-247 | lanceolata, 839 Magar, 623 |
| Epizootic, 25 | Lycaperdon Bovista, 99, 247, | Magnan, Dr., 793 |
| Lucuma, 89 | 860 | Magnan, Dr., 793 Magnin, L., 898 Mognolia, 111 |
| Banplandia, 53, 128, 851 | cyathiforme, 247 | Mognolia, 111 |
| deliciosa, 851 | excipuliforme, 246 | acuminata, 474 |
| mammasa 128 679, 851 | gemmatum, 860 giganteum, 245, 860 | Family, 111-112, 149, 473- |
| multiflora, 851 | saururus, 860 | grandiflora, 64, 111, 474, f. |
| glycyphlaea, 851 mammosa, 128, 679, 851 multiflora, 851 abovata, 679 | sp., 246 | grandiñora, 64, 111, 474, f. 474, 837 Great-flowered, 473, f. 474 Large-leaved, 474 |
| salicifora, 681, 851 Ludwigia erigata, 840 Luerssen, Ch., 897 Luffa, 89 | Lycopersicum, 89 esculentum, 132, f. 713, 714 | Great-flowered, 473, f. 474 |
| Luawigia erigata, 840 Luerssen Ch 807 | esculentum, 132, f. 713, 714 | Large-leaved, 4/4 |
| Luffa, 89 | pimpinellifolium, 715 Lycopodiaceae, 836 | macrophylla, 474 obovata, 474 |
| acutangula, 819 | LYCOPODIALES, 312 | obovata, 474 Purple, 474 |
| acutangula, 819 acgyptiaca, 750, 819 | Lycohodium 312 | tripetala, 474 |
| purgans, 819 | clavatum, f. 313, 836 Selogo, 836 Lycorin, 386 Lycoris radiata, 805 | virginiana, 474 |
| Lumpy Jaw, f. 183 | Selogo, 836 | Yulan, 474 Magnoliaceae, 89, 90, 111, |
| Lunasia amara, 583 philippensis, 849 | Lycorii, 300 Lycoris radiata 805 | Magnoliaceae, 89, 90, 111, 444, 446, 473.475, 837 Magnolin, 474 Magonia, 52, 63, 89, 607 Brazilian, 64, 607 glabrata, 850 |
| Lungwort, 704 | sp., 386 | Magnolin, 474 |
| Lupanin, 530 Lupigin, 531 | Lygodesmia, 757, 762 juncea, 762 | Magonia, 52, 63, 89, 607 |
| Lupigin, 531 | juncea, 762 | Brazilian, 64, 607 |
| Lupin Oil, 548 Lupine, 6, 118, 149, 525, 546- | Lygodium falmatum, 315 | glabrata, 850 pubescens, 850 |
| 550, 865 | Lymphadenis, 184 Lymphangitis, 171, 184, 865 | Mahogany, 575 |
| Blue, 118 | Lyngbya, 93, 188 | Mountain, 89, 503 Nigeria, 587 |
| European, 118 White, 547 | aestuarii, f. 185 | Nigeria, 587 |
| Hairy 546 | Lynghya, 93, 188 aestuarii, f. 185 Lyonia, 667, 674-675 inariana, 128, 674-675, f. | Mahonia Trailing, 112, 472 Maiden, J. H., 4, 37, 96, 390, 533, 534, 588, 650, 716, 724, 729, 734, 755, 803, |
| Hairy, 546 Low, 550 | 675 | 533, 534, 588, 650, 716, |
| Nebraska, 546, 550 | Lyons, A. B., 803, 804, 897 | 724, 729, 734, 755, 803, |
| Poisoning in Montana, 548- | Lyperia atropurpurea, 134 | 804, 898 |
| 550 Silvery, 550 | Lysimachia Nummularia, 676, | Maiden-hair Fern, 101, 313, f. 316, 316, 317, f. 317 Maidismus, 11, 80 Maish, J. M., 898, 918 "Mai-trunk," 744 Maize, See Corn |
| Western, 118 | 844 Tusin 10 | Maidismus. 11, 80 |
| Wild 546 | Lysin, 10 Lythraceae, 637, 836-837 | Maish, J. M., 898, 918 |
| Lupinidin, 118, 548 | Lythrum Salicaria, 638 | "Mai-trunk," 744 |
| Lupinin, 118, 531, 548 | | Maize, See Corn |
| Lupinidin, 118, 548 Lupinin, 118, 531, 548 Lupinin II, 548 Lupinin III, 548 | M , | maizeme acid, ora |
| Lupinosis, 20, 37, 40-43, 80. | Mahala 681 | Majun, 411 Makosi, 898 |
| Lupinosis, 20, 37, 40-43, 80, 548-550, 862, 865 | Mahola, 681 McCallum, 25 | Malabar Cardamon, 391 |
| Lubinotoxin, 78, 548 | McCallum, 25 McCarroll, Dr., 25 | Malassezia Furfur, 298 |
| Lupinus, 535, 546-550, 831, 865 | McCarthy, Gerald, 23, 245, | Maldi-gomma of orange and |
| albus, 118. 392, 525, 546, | 897 | lemon trees, 287 Male Shield Fern, 149, f. |
| 548, 831 | McCord, Dr. E. S., 50 MacDougal, Dr. D. T., 386, | Male Shield Fern, 149, f. 309, 315 |
| angustifolius, 546, 548, 831 | 394, 395, 897 | Extract of, 77 |
| argenteus, 118, 546, 550 | McCord, Dr. E. S., 50 MacDougal, Dr. D. T., 386, 394, 395, 897 McFarland, Dr., 184 McFaydean, J., 897 McGinnis, J. W., 897 | Malic acid, 144, 501, 505, 512 |
| argophyllus, 118 densiflorus, 831 | McFaydean, J., 897 | Mallotus phillippensis, 589, 823 |
| Genamorna, Gal | MeGinnis, J. W., 897 | 023 |
| | | |

Mallow, Dwarf, 624, f. 624
False, 626
Family, 564, 622-627
Iudian, 124, 626-627
Marsh, 623
Poppy, 624
Malmsten, 13
Malanetia niida, 807
Malpighia axycacca, 837
MALPIGHIACEAE, 575, 837
Malva moschata, 624
rotundifolia, 624, f. 624
MALVACEAE, 124, 621, 622-627, 837 MALVACEAE, 124, 621, 622-627, 837
MALVALES, 157, 621-627
Malvastrum, 626
coccincum, 564, 626
Mannillaria, 635, 811
Mammea americana, 627, 827
Manney Apple, 627
Man-of-the-earth, 699, f. 700, 701
Manacin, 715
Manchineal Tree, f. 586, 587, 865 865 Mandarin, 582 Mandragora, 715 autumnalis, 854 officinarum, 715, 854 autiminalis, 854
officinarium, 715, 854
vernalis, 4
Mandragorin, 715
Mandrake, 88, 112, 469-471,
f. 470, 715
Manganese, Salts of, 7
Mange, Doughy, 301
Sarcoptic, f. 292
Umatilla Horse, 100
Mangels, 425, 866
Mangifera indica, 608, 805
Mangifera indica, 608, 805
Manglotic glauca, 837
Mango, 608
Mango, 608
Mangorier, 627, 681
Mangrove, 638
Mangorier, 575
Manihet, 52, 586, 587
Glaziovii, 823
utilissima, 53, 503, 587, 823
Manila Hemp, 391
Mann, J. D., 898
Manna, J. D., 898
Manna, 527, 683
Manna Grass, 90
Ergot on, f. 27
Mannetol, 683
Mannatelo, 683
Mannite, 506
Maple, Curly, 615
Family, 615
Family, 615
Fard, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615
Silver, 615 Maranta arundinaceae, 391, 587 MARANTACEAE, 391
Marasnius oreades, 53, 860
tenerrimus, f. 232
urens, 860
Marattia, 315
MARATTYALES, 315
Marchand, L... 898
Marchantia, 310
polymorpha, f. 309, 311
Maregravia umbellata, 628
MARCGRAVIACEAE, 628
MARCUS, SI. D., 898
Marek, J., 13, 26, 892, 898
Mare's-tail, 640, f. 777
Marguerite, 756 MARANTACEAE, 391 Marguerite, 756 Marie, 572 Marigold, 756 African, 755-756

Burr, 779
Fetid, f. 139, 140, 784-785, f. 785
Flowers, 756
Marsh, 75, 381, 448-449
Pot, 756
Marimbo, 751
Mariposa Lily, 375
Marjoram, 710
Marka vitiensis, 127, 817
Marmalade, 679
Marubiun, 709
Marubiun vulgare, 709
Marsdenia Cundurango, 696, 809 Marrubium vulgare, 709
Marsdenia Cundurango, 696, 809
erecta, 809
Marsh, C. Dwight, 40, 93, 96, 467, 565, 566, 567, 569, 898
Marsh Cess, 490
Elder, 137, 763-764, 767
Small-flowered, 763
Mallow, 623
Marigold, 75, 381, 448-449
Skullcap, 131
Vetchling, 572
Marshall, C. E., 344
Marshall, H. T., 62, 871, 898
Marsilia, 315
Quadrifolia, f, 313
MARSILIACEAE, 315
Martin, 290
Maschka, J. v., 241, 898
Masson, P., 687, 898
Mason, W. P., 8
Massachusetts, Poisonous
plants of, 866
Report of State Board of
Health of, 95
Massey, W. F., 898
Mash, Bitter, 403
Sweet, 403
Mastic, 608
Maté, 614, 742
Materia Medica, Literature
of, 866
Matico, 396
Matico de Peru, 396
Matico de Peru, 396
Matricaria Chamomilla, 754
Matrimony Vine, 133, 733
Matrin, 530
Matruchot, L., 13, 15, 252,
879, 899
Mathhold incana, 486
livida, 818
Mauritius Hemp, 386
Maurizio, A., 899
May Apple, 469-471, f. 470
Maybole, Record of injury
from polluted water in,
Mayday Trcc, 504, f. 519
Mayday Trcc, 504, f. 519
Mayday Trcc, 504, f. 519
Maydism, See Maidismus 809 Mayday Tree, 504, f. 519
Maydism, See Maidismus
Mayo, N. S., 215, 259, 290,
565, 899
Maypop, 633
Mayweed, 140, 756, 787-788
Maximilian's Sunflower, f. Maza, M. G. de la, 899
Mead, Richard, 3, 899
Meadow Anemone, 455
Lychnis, f. 440
Rue, 88, 111
Saffron, 148
Sweet, 629
Meadowoot, Grass, Ta Meadow-oat Grass, Tall, f. Mease, James, 277
Mease, James, 277
Meat poisoning, 172, 174
Mechanical injuries, 64, 66,
804, 866 Meconic acid, 144

Meconidin, 481 Medea, 2 Medeola, 89 virginiana, 104 virginiana, 104 virginica, 834 Medicago, f. 535, 552-553 falcata, 526-552 hispida, 552 lupulina, 553 blatvacrae, 526 Inpulina, 553
platycarpa, 526
ruthenica, 526
sativa, 118, 525, 526, 553
Medical Jurisprudence, 866
Medicinal plants among l
gumes, 527-535
Distribution of, 866
Medick, 552
Black, 553
Mee, George, 899
Meehan, 679
Megnin, 14
Mehlthau, 272
Meiner, 899
Meissner, 300 Meissner, 300 Melaleuca leucodendron, 639, 839 Melambsora betulina, f. 221 populina, 222 MELAMPSORACEAE, 98-99, 221-222 Melampyrum arvense, 734, 852 lineare, 134 silvaticum, 134 Melandryum, 89 Melanopsichium, 210 MELANOSPOREAE, 235 MELANTHACEAE, 379 Melanthin, 62 Melanthiun, 52 cochinense, 834 latifalium, 380 parviflarum, 380 virginicum, 103, 379-380, f. 380, 834 MELASTOMACEAE, 53, 640, 837 Melia Azedarach, 121, 575, 837 MELIACEAE, 52, 121, 575, 837 MELIANTHACEAE, 89, 838 Melianthus comosus, 838 major, 838 Melica, 826 Melicope erythrococca, 849 Melilotus, 90, 523, 535, 551-552 alba, 831, 118, f. 551, 552 indica, 552, 831 officinalis, 118, 552, 831 Melissa officinalis, 710 Melassa omerinais, 710
Meladinus, 52
monogynus, 807
Melon, Nutmeg, 750
Sugar, 750
Melothria scrobiculata, 819
Meltzer, S. J., 899
Melvin, A. D., 293, 294, 899,
900
Mensester, 52 Memecylan, 53 Menabea venenata, 809, 866 Mendis, 575 Menes, 2 Menes, 2 Meningitis, 169, 171, 172, 866 Cerchro-spinal, 260 Epidemic, 169 Epizootic, 20, 259 Infectious, 21 Spinal, 291 Meningo-encephalitis, 866 MENISPERMACEAE, 52, 89, 112, 444, 472-473, 838 Arrow poisons among, 1, 52 Menispermin, 112, 149, 473

953

| Menisperuum canadense, 112, | Micromeria punctata, 710 | Mimusaps Kauki, 681 |
|---|--|--|
| 472-473, f. 473, 838 | Micracaccus caprinus, 168 | Mineral poisons, Testing for, |
| cacculus 149 | catarrhalis, 170 | 3 |
| cacculus, 149 Menispin, 112, 472 Mentha, 78, 710, 713 | ganarrheae, 170 | Mint, 713 |
| Mentha 78 710 713 | lancealatus, 169-170 | Family, 131, 708-713 Horse, 709, f. 709 Minteer, J. A., 653 Mio Mio, 755 Miquel, F. A. W., 214, 247, 287, 803, 804, 862, 899 Mirbelia racenosa, 533 Miscouri Currant 498, f. 509 |
| arvensis v. piperascens, 709 | meningitidis, 169 | Horse 709 f 700 |
| arvensis v. canadensis, 713 | huggange 171 | Minteer I A 653 |
| arvensis v. canadensis, 713 | pyagenes, 171 | Min Min 755 |
| crispa, 713 | pyogenes v. albus, 167 | Minust E A W 214 247 |
| piperita, 709, 713, f. 713 | pyogenes v. anrens, 107 | 207 002 004 862 800 |
| Pulegium, 709 | pyogenes v. aureus, 167 tetragenus, 170 MICROSPERMAE, 155, 392 Micrasphaera Alni, 267, 269 | 207, 003, 004, 002, 077 |
| spicata, 709 Menthol, 709 Mentzela, 125, 633-634 | MICKOSPERMAE, 133, 392 | Mirvetta faceinosa, 555 |
| Menthol, 709 | Micrasphaera Anni, 267, 269 | Missouri Currant, 470, 7. Joe |
| 272 | Micraspira comma, 181-182 Micrasparon, 13, 299, 302 | Gaoseberry, 500 |
| Family, 633-634 | Micrasparon, 13, 299, 302 | Gourd, 751 |
| granoviaefalia, 633 | adouini, 301, 302 | Hickory, 401 |
| aligosperma, 633, 634 | canis, 299 | Hickory, 401 Missouri-Bottom Disease, 118, |
| arnata, 633-634 avata, 125 | adouini, 301, 302 canis, 299 equi, 299 Furfur, 298 | 546 |
| avata, 125 | Furfur, 298 | Mistletoe, 866 |
| Showy, 633-634, f. 634 | tigris, 299 | European, 106 |
| Mcnyanthes, 690-691 | Microsporosis, 14 | False, 106 |
| trifaliata, 129, 690-691, f. | Microthyrium microscapicum, | Southern, 415, f. 416 |
| 601, 825 | | Southern, 415, f. 416 Mitcham Oil, 86 |
| Manuanthin 129 691 | Mignapotta 479 736 | Mitchell A 124 621 |
| Marhana Oil of 6 | Mignoficite, 477, 750 | Mitchell, A., 124, 621 |
| Menals E 900 | Migula, W., 101, 103 | Mitchell, S. R., 899 |
| Menyanthin, 129, 691 Merbanc, Oil of, 6 Merck, F., 899 | Mignoncite, 479, 736 Migula, W., 161, 165 Mildew, Downy, 98 On Bean, 208 Clover, 205, 206 Grape, 205 Lettuce, 205 | Mitchella, 89 repens, 742, 848 |
| Merck & Report, 050 | Clause 205 206 | repens, 742, 848 |
| Mercurialin, 83 | Clover, 203, 206 | Mitlacher, W., 863, 899 |
| Mercurialis, 590 | Grape, 205 | Mitracarpum, 90 |
| annua, 83, 603, f. 603, 823 | Lettuce, 205 Millet, 205 | Mitragyne speciasa, 848 |
| ferennis, 603, 823 | Millet, 205 | Mnium, 310 |
| tamentasa, 823 | Union, 205 | harmum, f. 311 Moccasin Flower, f. 393 |
| Mercury, Annual, 603, f. 603 | Sunflower, 205 | Moccasin Flower, f. 393 |
| Biniodide of, 6 | Powdery | Showy, 105 Mocha Coffec, 744 |
| Three-seeded, 603-604 | On Cherry, f. 269 Grape, 269 | Mocha Coffee, 744 |
| Mericourt, 572 | Grape, 269 | Mock Orange, 500 |
| Merismapedia, 184, 188 | Grasses, f. 267, 268, | Madecca, 89 |
| Mermaid-weed, 640 | Grasses, f. 267, 268, 269-272, f. 271 | palmata, 841 |
| Merrill, E. D., 52, 899 | Hop, f. 269 | trilabata, 841 |
| Mertensia sibirica, 704 | Legumes, 267 | |
| virginica, 704 | Lilacs, 267 | Moeller T. 899 |
| Merulius lacrhrymans, f. 232, | Milfoil Water 640 | Moeller Tosef 900 917 |
| 234 | Milfoil, Water, 640 Milium, 102 | Most TE 900 |
| Mescal bean, 637 | A 926 | Tenenata, 841 Moeller, J., 899 Moeller, Josef, 900, 917 Moffit, J. E., 900 Mohler, J. R., 26, 28, 231, 263, 265, 283, 284, 293, 294, 875, 899, 900 Mohr, Ch., 900 Molinia caerulea, 826 |
| Button See Mescal beam | effusum, 826 Milk Plant, 589 | 265 292 294 203 204 |
| Button, See Mescal bean Mescalin, 637 | Milk Flant, 309 | 203, 203, 204, 273, 274, |
| Mescain, 657 | Vetch, 562-567 | 8/5, 899, 900 |
| Mesembryanthemum anatami- | Milk-fever caused by Eupa- | Monr, Ch., 900 |
| cuin, 824 | torium, 138 | |
| | | |
| Crystallinum, 424, 824 | Milk-sickness, 771, 772 | Molle, Ph., 900 |
| Mesquit Tree, 120, 527, 866 | torium, 138 Milk-sickness, 771, 772 Milkweed, 130, 386, 598, 696- | Mollugo hirta, 824 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, | 697 | Mollugo hirta, 824 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 | 697 | Mollugo hirta, 824 Molopospermum cicutarium, |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 | 697 Family, 130, 695-697 | Mollugo hirta, 824 Molopospermum cicutarium, 856 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 | Milkweed, 130, 386, 598, 696- 697 Family, 130, 695-697 Narrow-leaved, 130 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 | 697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsanina, 819 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, 12., 249 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Monardica Balsannina, 819 charantia, 819 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stauntani, 809 Meta-phosphoric acid, 124 Methyl xanthin, 146 Methyl xanthin, 146 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, 12, 249 Methyl xanthin, 146 Methylconiin, 126, 651 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Momarda citradara, 710 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Momarda citradara, 710 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, 15., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Momarda citradara, 710 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Momarda citradara, 710 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, /. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Momarda citradara, 710 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 combalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methyamin, 10, 149 Methysticin, 396 Metritis, 179 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 | Mollugo hirta, 824 Molopospermum cicutarium, 836 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metrylan Rumphii, 370 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 | Mollugo hirta, 824 Molopospermum cicutarium, 836 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metraxylan Rumphii, 370 Mexican Mulberry, 131 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 349, 350 Italian, 350 Koda, 349 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 colarantia, 819 cochinchinensis, 819 cymbalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 functata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 leterasparus, 247 furpureus, 247, f. 248 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metraxylan Rumphii, 370 Mexican Mulberry, 131 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T. 546 Miller, W. H. 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 didyma, 629, 709 fistulasa, 709, f. 709 functata, 709, 7. 709 functata, 709, 7. 709 MONASCACEAE, 247-248 Manascus, 24 leterasparus, 247, f. 248 Monlia candida, 302 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metraylan Rumphii, 370 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 349 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 244, f. 248 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 colarantia, 819 cochinchinensis, 819 cymbalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 functata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 leterasparus, 247 furpureus, 247, f. 248 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanntani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metris, 179 Mctraxylan Rumphii, 370 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 349 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 244, f. 248 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 colarantia, 819 cochinchinensis, 819 cymbalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 functata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 leterasparus, 247 furfureus, 247, f. 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey Flower, 733 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Metricaylan Rumphii, 370 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey Flower, 733 Monkey-broad Tree, 621 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, 12, 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metraxylan Rumphii, 370 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Coren, 350 Corean Foxtail, f. 349 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferryiginea, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey Flower, 733 Monkey-broad Tree, 621 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, 12, 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metraxylan Rumphii, 370 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 349 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 ferruginea, 832 ferruginea, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey Flower, 733 Monkey-broad Tree, 621 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Metricaylan Rumphii, 370 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A, 899 Meyer, 11, 888, 899 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 340, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 pachycarfa, 832 pachycarfa, 832 pastrata, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey Flower, 733 Monkey-broad Tree, 621 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 349 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferriginea, 832 fachycarpa, 832 rastrata, 832 restrata, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey Flower, 733 Monkey-broad Tree, 621 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 340, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 pachycarfa, 832 rastrata, 832 sericea, 832 sericea, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 combalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 740 Monascus, 24 lielerasparus, 247 lielerasparus, 247, f. 248 Monilia candida, 302 Monimaceare, 838 Monkey-Flower, 733 Monkey-Flower, 733 Monkey-pot Tree, 621 Monkey-pot Tree, 621 Monkey-pot Tree, 638 Monkshood, Wild, 109, 453 Minnina, 89, 843 MONOCOTYLEDONEAE, |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 340, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 pachycarfa, 832 rastrata, 832 sericea, 832 sericea, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 functata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 leterasparus, 247, f. 248 Monilia candida, 302 Monimiaceae, 838 Monkey Flower, 733 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkshood, Wild, 109, 453 Minnina, 89, 843 MONOCOTYLEDONEAE, 102, 155 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 340, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 pachycarfa, 832 rastrata, 832 sericea, 832 sericea, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 danarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 Monimaceae, 838 Monkey Flower, 733 Monkey-bread Tree, 621 Monkshood, Wild, 109, 453 Minnina, 89, 843 MONOCOTYLEDONEAE, 102, 155 Manaspara bicuspidata, 249 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 340, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 pachycarfa, 832 rastrata, 832 sericea, 832 sericea, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 Monilmaceare, 838 Monkey Flower, 733 Monkey-pot Tree, 621 Monkey-pot Tree, 621 Monkey-pot Tree, 638 Monkshood, Wild, 109, 453 Minnina, 89, 843 MONOCOTYLEDONEAE, 102, 155 Manaspara bicuspidata, 249 Manotrapa unifara, 65, 838, |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 340, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 pachycarfa, 832 rastrata, 832 sericea, 832 sericea, 832 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 danarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 punctata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 Monimaceae, 838 Monkey Flower, 733 Monkey-bread Tree, 621 Monkshood, Wild, 109, 453 Minnina, 89, 843 MONOCOTYLEDONEAE, 102, 155 Manaspara bicuspidata, 249 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 Mezerein, 642 Mezereon, 149 Mezereum, 642 Family, 125, 642-643 Michael, Helen A., 375 Michael, L. G., 424, 885, 889, 894, 895, 898, 899, 907 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 ferruginea, 832 splendeus, 832 Millspaugh, C. F., 96, 134, 449, 449, 454, 459, 471, 484, 489, 509, 552, 560, 586, | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 combalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 10 MONASCACEAE, 247-248 Manascus, 24 lulerasparus, 247, f. 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey-Flower, 733 Monkey-Flower, 733 Monkey-Pot Tree, 631 Monkey-pot Tree, 638 Monkey-pot |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 Mezerein, 642 Mezereon, 149 Mezereum, 642 Family, 125, 642-643 Michael, Helen A., 375 Michael, L. G., 424, 885, 889, 894, 895, 898, 899, 907 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferringinea, 832 ferringinea, 832 pachycarpa, 832 rastrata, 832 sericea, 832 splendeus, 832 Millspaugh, C. F., 96, 134, 411, 419, 429, 435, 440, 449, 454, 459, 471, 484, 489, 509, 552, 560, 586, 602, 617, 632, 655, 659, | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 combalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 10 MONASCACEAE, 247-248 Manascus, 24 lulerasparus, 247, f. 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey-Flower, 733 Monkey-Flower, 733 Monkey-Pot Tree, 631 Monkey-pot Tree, 638 Monkey-pot |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, 15., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metrysticin, 396 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 Mezerein, 642 Mezereon, 149 Mezereon, 149 Mezereum, 642 Family, 125, 642-643 Michael, Helen A., 375 Michael, L. C., 424, 885, 889, 894, 895, 898, 899, 907 Michelia Champaca, 837 fuscala, 837 | Milkweed, 130, 386, 598, 696- Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 349 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 ferruginea, 832 ferruginea, 832 sericea, 832 splendeus, 832 Millspaugh, C. F., 96. 134, 441, 419, 429, 435, 440, 444, 454, 471, 484, 489, 509, 552, 560, 586, 602, 617, 632, 655, 659, 661, 671, 728, 739, 747. | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 danarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 10 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 Monimaceae, 838 Monkey-Flower, 733 Monkey-Flower, 733 Monkey-pt Tree, 638 Monkey-pt Tree, 638 Monkshood, Wild, 109, 453 Minnina, 89, 843 MONOCOTYLEDONEAE, 102, 155 Manaspara bicuspidata, 249 Manotrapa uniflara, 65, 838, 842 Monotropaceae, 838 Manstera deliciosa, 371 bertusa, 808 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, 15., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metrysticin, 396 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 Mezerein, 642 Mezereon, 149 Mezereon, 149 Mezereum, 642 Family, 125, 642-643 Michael, Helen A., 375 Michael, L. C., 424, 885, 889, 894, 895, 898, 899, 907 Michelia Champaca, 837 fuscala, 837 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 349 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 ferruginea, 832 splendens, 832 splendens, 832 splendens, 832 splendens, 832 Millspaugh, C. F., 96, 134, 440, 449, 454, 459, 471, 484, 489, 509, 552, 560, 586, 602, 617, 632, 655, 659, 661, 671, 728, 739, 747, 752, 762, 790, 793, 899 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 danarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 10 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 Monimaceae, 838 Monkey-Flower, 733 Monkey-Flower, 733 Monkey-pt Tree, 638 Monkey-pt Tree, 638 Monkshood, Wild, 109, 453 Minnina, 89, 843 MONOCOTYLEDONEAE, 102, 155 Manaspara bicuspidata, 249 Manotrapa uniflara, 65, 838, 842 Monotropaceae, 838 Manstera deliciosa, 371 bertusa, 808 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, 15., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metrysticin, 396 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 Mezerein, 642 Mezereon, 149 Mezereon, 149 Mezereum, 642 Family, 125, 642-643 Michael, Helen A., 375 Michael, L. C., 424, 885, 889, 894, 895, 898, 899, 907 Michelia Champaca, 837 fuscala, 837 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 ferruginea, 832 pachycarpa, 832 splendeus, 832 Millspaugh, C. F., 96, 134, 410, 449, 454, 459, 471, 484, 489, 509, 552, 560, 586, 602, 617, 632, 655, 659, 661, 671, 728, 739, 747, 752, 762, 790, 793, 899 Milnes, Dr., 544 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 danarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 10 MONASCACEAE, 247-248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 Monimaceae, 838 Monkey-Flower, 733 Monkey-Flower, 733 Monkey-pt Tree, 638 Monkey-pt Tree, 638 Monkshood, Wild, 109, 453 Minnina, 89, 843 MONOCOTYLEDONEAE, 102, 155 Manaspara bicuspidata, 249 Manotrapa uniflara, 65, 838, 842 Monotropaceae, 838 Manstera deliciosa, 371 bertusa, 808 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, 15., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metrysticin, 396 Metritis, 179 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 Mezerein, 642 Mezereon, 149 Mezereon, 149 Mezereum, 642 Family, 125, 642-643 Michael, Helen A., 375 Michael, L. C., 424, 885, 889, 894, 895, 898, 899, 907 Michelia Champaca, 837 fuscala, 837 | Milkweed, 130, 386, 598, 696- Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 340, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 ferruginea, 832 splendeus, 832 Millspaugh, C. F., 96, 134, 440, 449, 454, 459, 471, 484, 489, 509, 552, 560, 586, 602, 617, 632, 655, 659, 661, 671, 728, 739, 747, 752, 762, 790, 793, 899 Milnes, Dr., 544 Minasa. 89 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f, 709 functata, 709, 710 MONASCACEAE, 247-248 Manascus, 24 leterasparus, 247, f, 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey Flower, 733 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkey-bot Tree, 638 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkshood, Wild, 109, 453 Minnina, 89, 843 MONOCOTYLEDONEAE, 102, 155 Manaspara bicuspidata, 249 Manotrapa uniflara, 65, 838, 842 Montana, Economic plants of, 864 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metrazylan Rumphii, 370 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 Mezerein, 642 Mezereon, 149 Mezereum, 642 Family, 125, 642-643 Michael, Helen A., 375 Michael, L. G., 424, 885, 889, 894, 895, 898, 899, 907 Michelia Champaca, 837 nilagirica, 474 parvifalia, 837 Michener, C. B., 259 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 349 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 ferruginea, 832 splendeus, 832 splendeus, 832 splendeus, 832 splendeus, 832 Millspaugh, C. F., 96, 134, 449, 454, 459, 471, 484, 489, 509, 552, 560, 586, 602, 617, 632, 655, 659, 661, 671, 728, 739, 747, 752, 762, 790, 793, 899 Milnes, Dr., 544 Mimasa, 89 MIMOSEAE, 530 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 danarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 fistulasa, 709, f. 248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 Monimaceare, 838 Monkey-Flower, 733 Monkey-bread Tree, 621 Monkey-bread Tree, 621 Monkey-bread Tree, 621 Monkey-bread Tree, 638 Monke |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metraylan Rumphii, 370 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 Meyer, H., 888, 899 Mezerein, 642 Family, 125, 642-643 Michael, L. G., 424, 885, 889, 894, 895, 898, 899, 907 Michelia Champaca, 837 fuscata, 837 nilagirica, 474 parvifalia, 837 Michener, C. B., 259 Michands, 587 | Milkweed, 130, 386, 598, 696- Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 340 German, 102, f. 340, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 pachycarpa, 832 rastrata, 832 sericea, 832 splendeus, 832 Millspaugh, C. F., 96. 134, 440, 454, 459, 471, 484, 459, 471, 484, 489, 509, 552, 560, 586, 602, 617, 632, 655, 659, 661, 671, 728, 739, 747, 752, 762, 790, 793, 899 Milmose, Dr., 544 Mimasa, 89 MilMOSEA E, 530 Mimulus Inteus, 733 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cymbalaria, 819 Manarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 functata, 709, f. 709 functata, 709, f. 709 functata, 709, f. 709 functata, 709, f. 709 MONASCACEAE, 247-248 Manascus, 24 leterasparus, 247, f. 248 Monilia candida, 302 MONIMIACEAE, 838 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkey-bread Tree, 638 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkey-bread Tree, 621 Monkey-pot Tree, 638 Monkey-bread Tree, 638 Monkey-bread Tree, 631 Monkey-bread Tree, 638 |
| Mesquit Tree, 120, 527, 866 Metabolism, Products of, 82, 548 Metaplexis Stanutani, 809 Meta-phosphoric acid, 124 Metchnikoff, E., 249 Methyl xanthin, 146 Methylconiin, 126, 651 METACHLAMYDEAE, 157 Methaemoglobin, 72 Methoxy, 147 Methylamin, 10, 149 Methysticin, 396 Metritis, 179 Metrazylan Rumphii, 370 Mexican Mulberry, 131 Prickly Poppy, 113, 483 Tea, 107, 629 Mexico, Economic plants of, 864 Meyer, 203 Meyer, A., 899 Meyer, H., 888, 899 Mezerein, 642 Mezereon, 149 Mezereum, 642 Family, 125, 642-643 Michael, Helen A., 375 Michael, L. G., 424, 885, 889, 894, 895, 898, 899, 907 Michelia Champaca, 837 nilagirica, 474 parvifalia, 837 Michener, C. B., 259 | Milkweed, 130, 386, 598, 696-697 Family, 130, 695-697 Narrow-leaved, 130 Showy, 130, 696-697, f. 697 Swamp, 130 Milkwort, 585-586 Family, 584-586 Miller, S. T., 546 Miller, W. H., 454 Millet, 102, 350-352, 866 Broom Corn, 350 Corean Foxtail, f. 349 German, 102, f. 349, 350 Italian, 350 Koda, 349 Poisoning, 350-351 Smut, f. 214, f. 218 Milletia, 1, 52, 89 caffra, 832 ferruginea, 832 ferruginea, 832 splendeus, 832 splendeus, 832 splendeus, 832 splendeus, 832 Millspaugh, C. F., 96, 134, 449, 454, 459, 471, 484, 489, 509, 552, 560, 586, 602, 617, 632, 655, 659, 661, 671, 728, 739, 747, 752, 762, 790, 793, 899 Milnes, Dr., 544 Mimasa, 89 MIMOSEAE, 530 | Mollugo hirta, 824 Molopospermum cicutarium, 856 Molteno Cattle-disease, 795 Momardica Balsamina, 819 clarantia, 819 cochinchinensis, 819 cochinchinensis, 819 danarda citriadara, 710 didyma, 629, 709 fistulasa, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 punctata, 709, f. 709 fistulasa, 709, f. 248 Manascus, 24 luterasparus, 247, f. 248 Monilia candida, 302 Monimaceare, 838 Monkey-Flower, 733 Monkey-bread Tree, 621 Monkey-bread Tree, 621 Monkey-bread Tree, 621 Monkey-bread Tree, 638 Monke |

199, 200, 201
equinus, 202
fusiger, 198
inacquaelis, 200
melittophtorus, 198
mucedo, 98, 195, f. 196, Montgomery, F. II., 13, 19, 295, 298, 299, 302, 892, 900 295, 298, 299, 302, 892, 900

900

Montinia caryophyllacea, 840

Moon-flower, 699

Moonseed, 472-473, f. 473

Canada, 112

Carolina, 472

Family, 472-473

Moon, Dr., 482

Moore, G. T., 94, 95, 188, 900

Moore, Dr. V. A., 22, 214

Mosse-wood, 125, 642

Morchella esculenta, 253

Morelia senegalensis, 848

Morbitz, 726

Morchella esculenta, 253

Morelia senegalensis, 848

Morgan, 637

Mori, R., 896, 960

Morman, J. W., 900

Morindin, 742

Morindin, 742

Morindin, 742

Morindin, 742

Morindin, 742

Morindin, 742

Morindin, 742

Morphin, 3, 59, 73, 77, 78, 80, 82, 85, 90, 112, 113, 143, 144, 146, 147, 479, 489, 866

Group, 147

Morphinism, 77

Morphinism, 77

Morphinism, 77

Morphinism, 77

Morphinism, 77

Morphinism, 77

Morphinism, 77

Morphinism, 77

Morphinism, 77 nigricans, 19 Oryzae, 198 parasiticus, 2 pusillus, 201 202 racemosus, 98, 198, 200-201 ramosus, 201 Regnieri, 200 rhizopodiformis, 98, 200 Rouxii, f. 197, 198 stolonifer, 98, 195, f. 198, 200 septatus, 199, 202 Trichisi, 200 sp., 287 Mucoraceae, 12, 13, 98, 195, 202 Mucormycosis, 203, 866
Mucormycosis, 203, 866
Mucuna capitata, 832
gigantea, 524, 832
pruriens, 524, 527-528
purpureus, 832
urens, 832
venenosa, 832
Mudar, 695
Mudarin, 695
Muchlenbeckia platyclados,
419
Mueller, C., 85 143, 144, 146, 147, 479, 489, 866
Group, 147
Morphinism, 77
Morphinism, 77
Morris, Daniel, 900
Morris, Daniel, 900
Morris, Malcolm, 300
Morrisonia americana, 812
Morrow, P. A., 900
Mortierella Rostafinskii, 196
Morus alba, 406, 629, 838
rubra, 838
Moschatin, 140, 187
Mosely, E. L., 771-772
Moses, Jacob, 553
Moss, f. 300
Pink, 423
Spanish, 372
Mosses, 310
Moss, f. 311
Club, f. 312
Iceland, 309
Inniper, f. 311
Sphagnum, 310
Moth Bean, 526
Mullein, 134, 734-735
Motherwort, 711, 712
Common, 131
Moulds, 16, 22, 195-203, 209, 206
Blue, 82, f. 255, 256-257 419 Mueller, C., 85 Mueller, F. v., 752, 900 Mueller, G., 895, 900 Mueller, C., 900 Mueller, K., 900 Mueller, R., 872, 900 Mueller, R., 872, 900 Muellera moniliformis, 832 Telfairiii, 832 Mugwort, 796 Mugwort, 796
Common, 793-794
Muir, R., 305
Mulberry, 405, 406, 629, 866
French, 708
Mexican, 131
Paper, 406
White, 406
Mullein, 733, 734-735
Common, 733, 734, f. 734
Moth, 134, 734-735
Müller, 25
Mundulea, 1 Müller, 25 Mundulea, 1 suberosa, 533 Muntingia, 622 Murphy, 1f. S., 56 Murphy, J. B., 183 Murrain, Dry, 9 Murray, 459 Murrell, W., 900 Murrill, W. A., 244 Black, 96, 193-200, 7, 196,
Blue, 82, f. 255, 256-257
Green, f. 257, 257-261
Herbarium, 257
Pale, f. 261, 262
Slime, f. 158, 159, 167
Water, f. 208
Mountain Ash, 115, 629
Blackberry, 665
Fetter Bush, 674
Larkspur, Tall, 109
Laurel, 64, 84, 665, 669-671, f. 670
Great, f. 666, 667
Lousewort, 134
Rose-bay, 127, 667-668
Sneezeweed, 140
Mouriria rhizophoraefolia,
837
Mouse Septicemia, 176 200 Musa sapientum, 391, f. 391, 839 textilis, 391 MUSACEAE, 391, 839 Muscaria, 89 comosum, 834 тасетовин, 834

Миссатіп, 10, 32, 73, 74, 77, 78, 80, 82, 148, 150, 237, 238, 242 Musci, 310 Musgrove, 184 Mushrooms, 99, 160, 220, f. 233, 866 Common, 239, 240 Cultivated, 235 Field, 235 837 Mouse Septicemia, 176 Moussu, 534, 880, 900 Moutet, 900 Mucor, 195-203, 208-866 Biology of, 195-198 corymbifer, 98, 198-199, f. Poisonous, 235-242 Shaggy-mane, 235

Musk Flower, 733 Seed, 624 Tree, 755 Tree, 755
Muskmelon, 750, f. 751
Musk-root, 741
Musquash Root, 83
Mussaenda, 89
frondosa, 849
Mustang Grape, 124, 620
Mustard, 460, 488, 491
Black, 77, 486, 488, f. 489, Biack, 77, 490

490

Brown, 490

Family, 113, 485-495

Hedge, 114, 487-488

Indian, 490

Myronic acid of, 7

Oil of, Essential, 489, 575

Ethereal, 7, 77

Volatile, 114

Sarepta, 490

Tumbling, 114, f. 487, 488

White, 77, 486, 488, f. 489, 496

Muter, John, 692

Mutinus caninus, 245, 860

elegans, 860 Mutinus caninus, 245, 860
elegans, 860
Mutisia, 89
viciaefolia, 816
Mutual Life Insurance Co., of
New York, 901
Mycetoma, 184
Mycetoza, 158
Mycorrhiza, 154
Mycose in fungi, 277
Mycosis, 199, 301, 866
Dermal, f. 292
Pulmonary, 263
Mycotic stomatitis, 99, 100 Mycotic stomatitis, 99, 100 Mydalein, 150 Mydatein, 150 Mydatoxin, 150 Myelin, 395 Myers, J., 305, 30 Mykosyrinx, 210 Myoporaceae, 838 MYOPORACEAE, 838
Myoporum deserti, 838
Myoporum deserti, 838
Myosotis scorpioides, 704
Myrobalons, 640
Myrica, Oil of, 640
Myrica, f. 398
acris, 399
asplenifolia, 399
cerifera, 399, 838
Nagi, 399
MYRICACEAE, 397-399, 838
Myricales, 156
Myricaria germanica, 627
Myricetin, 399
Myriocarpin, 750
Myriocarpin, 750
Myriophyllum, 93
spicatum, 640 Myriocarpin, 750
Myriophyllum, 93
spicatum, 640
Myristic acid, 389, 593
Myristica fatua, 445
fragrans, 445
gibbosa, 838
succedana, 445
Myristicaceae, 444, 445, 838
Myristicin, 445
Myristicin, 445
Myristin, 594
Myronic acid of Mustard,
Toxicity of, 7
Myrosin, 486, 489, 491, 633
Myroxylon, 523, 627
Pereirae, 832
peruiferum, 832
toluiferum, 528, 832
Myrsine africana, 839
Myrsine africana, 839
MYRSINEAF, 52
MYRACEAE, 52, 53, 89, 629, 639, 839
MYRTIFLORAE, 157, 637-645 645

INDEX

| Myrtle, 575, 637-645 | Nephritis, 613 | Spreading, f. 721 Three-flowered, f. 721, 722 |
|--|---|--|
| Cape, 637 | Suppurative, 172 | Three-flowered, f. 721, 722 |
| Spurge, 121, 599 | Nephrolegis exaltata, f. 312 | Yellow-flowered, 692 |
| Myrtus communis, 640 Ugui, 629 | Neriauthin, 130 | Nine-bark, 504 |
| Ugui, 629 | Neriin, 130, 148 | Nipa fruticaus, 370 |
| MYXOBACTERIACEAE, 166-167 | Neriin, 130, 148 Nerium, 419, 693-695 | Nissolia fruticosa, 832 |
| MYXOBACTERIACEAE, 166-167 MYXOGASTERES, 160 MYXOMYCETES, 167 | odorum 691 807 | Nitraria tridentata, 858 |
| MYXOMYCETES, 167 | oleander, 4, 65, 75, 129, 130, | Nitric aeid, 78 |
| MYXOTHALLOPHYA, 153, | 691, f. 694, 694-695, 807 | Nitrobenzene, 72 |
| 158-160 | oleander, 4, 65, 75, 129, 130, 691, f. 694, 694-695, 807 Nervin, 10, 78 | Nitrogen in alkaloids, 144, |
| | Nervous system, Poisons af- | 145 |
| N | fecting, 72 | Nitroug avid 73 |
| | Nesaea verticillata, 837 | Nobecourt, 901,916 Nocard, Ed., 184 Nockolds, C., 901 Nodularia, 97 |
| Nabias, De, 901, 881 | Nest-egg Gourd, 750 | Nocard, Ed., 184 |
| Nandina domestica, 53, 810 | Nestler, A., 364, 395, 901 | Nockolds, C., 901 |
| Napaleona Whitfieldii, 839 | Nest-egg Gourd, 750 Nestler, A., 364, 395, 901 Netherlands, Poisonous plants | Nodularia, 97 |
| Nara, 750 | of the, 867 | spunigera, 91 |
| Naranja, 582 | Nettle, 76, 77, 405, 594 | Noel, Ch., 895, 901 |
| Narcein, 146-481 | of the, 867 Nettle, 76, 77, 405, 594 Bull, 72, 122, 596, 724-725 | Noffrey, J., 901 |
| Marciacus 104 396 | Common, 594 | spunigera, 91 Noel, Ch., 895, 901 Noffrey, F., 901 Noll, F., 901 |
| Janguilla, 386 foeticus, 104, 386, 805 Poet's, 386 | Dead, 131, 712, f. 712 | Naisettia, 858 |
| roeticus, 104, 386, 805 | Family, 409-415 | Nobalca caccinellifera, 635 |
| Poet's, 386 | Ilandling of, 78 | Norfolk Pine, 327 |
| Pseuda-Narcissus, 4, 386 | Slender, 106 | Norgaard, V. A., 293 |
| 805 | Handling of, 78 Slender, 106 Small Stinging, f. 412, 412- | Norfolk Pine, 327 Norgaard, V. A., 293 Northern Plum, Wild, 505 |
| Tazetta, 380, 805 | 413 | Nosebleed, 787 |
| Narcosis, 125 | Spurge, 122, f. 593, 596, f. | Nastoc, 184, 186 |
| Narcotic, 804 | 596 | caeruleum, 97 |
| Narcotic poisoning, 73, 78 | Stinging, 106, 413 | cammune, 97 |
| Narcotics, 73 | Western, 106 | muscarum, 97 |
| Narcotin, 3, 112, 146, 147, | Western, 106 Wood, 77, 106, 414 | paludosum, f. 92 |
| Narcotin, 3, 112, 146, 147, 481, 482 | Neumann, L. G., 13, 14, 16, | verrucosum, 186 |
| Naregamia alata, 837 | 198. 901 | NOSTOCACEAE, 97, 186 |
| Naringin, 583 | Neumayer, I., 249 | Nothoscordum bivalve, 104 |
| Narrow-leaved Milkweed, 130 | Neuridin, 149, 150 | striatum, 835 |
| Sanaramand 702 70. | 198, 901 Ncumayer, J., 249 Neuridin, 149, 150 Neurin, 150, 237 | Novy, F. G., 171, 901 |
| Narthecium assifragum, 835 Nasturtium, 120, 575 Family, 120 Garden, 575, f. 577 officinale, 729 falustre, 490 | New England Aster, 776 New Jersey Tea, 124, 621, 629 Newbauldia laevis, 739 | Nuein, 105 |
| Nasturtium, 120, 575 | New Jersey Tea, 124, 621, 629 | Nuckolds, C., 901 |
| Family, 120 | Newbauldia laevis, 739 | Numa 866 |
| Garden, 575, f. 577 | Nicander, 2 | Numan, A., 898, 901 |
| officinale, 729 | Nicander, 2 Nicandra, 717, 726 fhysaloides, 131, 726, 854 Nichols, II. J., 901, 904 Nicholson, I. T. Dr., 9 Nicodemia diversifalia, 836 Nicotelin, 728 Nicotellin, 728 | Numan, A., 898, 901 Nunn, J., 901 Nut Areca, 866 |
| talustre, 490 | thysaloides, 131, 726, 854 | Nut Areca, 866 |
| Native Plum, 614 | Niehols: II. I., 901, 904 | Becch, 105 Betel, 87, 396, 866 Brazil, 638, f. 638 |
| Naunyn, B., 873 | Nicholson, I. T. Dr., 9 | Betel, 87, 396, 866 |
| Nauseant, 804 Neal, H. V., 879, 901 Nebraska Lupine, 546, 550 | Nicodemia diversifalia, 836 | Brazil, 638, f. 638 |
| Neal, H. V., 879, 901 | Nicotein, 728 | Cashew, 608 |
| Nebraska Lupine, 546, 550 | Nicotellin, 728 | Cola, 621 |
| Neckweed, 735-736 | Nicotemin, 728 | Hickory, 401 |
| Necomospora, 287 | Nicotia, 752 | Kalo, 587 |
| Necrobacillosis, 179 | Nicotiana, 52, 717, 727-729 | Levant, 1 |
| Necrobacillosis, 179 Necrosis, 179, 265 | Nicotiana, 52, 717, 727-729 alata, 133, 715, 729, 854 | Pecan, 401 |
| Nectandra Radiaei, 478 | attenuata, 729 | Physic, Cuhan ,596 |
| Nectar, Poisonous, 64 | chinensis, 854 | Pistaehia 608 |
| Nectria, 273 Neebe, C. H., 296 Needle Grass, 66, 102, 355-357 | glauca, 133, 854 | Sapucaya, 638 Stinging, Brazilian, 596 Water, 640 |
| Neebe, C. H., 296 | glutinosa, 728 | Stinging, Brazilian, 596 |
| Needle Grass, 66, 102, 355-357 | | Water, 640 |
| Western, f. 70, 102, 355, f. | quadrivalvis, 729, 854 | Water-chestnut, 640 |
| 35 5 | rustica, 728, 729, 854 | Nutmeg, 445, 866 Oil of, 445 |
| Negunda acevoides, 615, 629 | suaveolens, 729, 854 | Oil of, 445 |
| Neisser, 168 | Tabacuin, 75, 133, 714, f. | Poisoning, 445 |
| Nekoe, 52 | 714. 727-729 | Nuttall, Thomas, 801 |
| Nekoeid, 52, 53 | matropyna, 729, 854 quadrivalvis, 729, 854 rustica, 728, 729, 854 suaveolens, 729, 854 Tabacum, 75, 133, 714, f. 714, 727-729 Nicotin, 7, 73, 77, 78, 80, 85, 87, 133, 144, 147, 551, 716, 728, 729, 866, 868 | Nuttalia cerasiformis, 53, 115, |
| Nelson, Aven. 138, 332, 367, | 85, 87, 133, 144, 147, | 503, 847 |
| 776, 901, 918 | 551, 716, 728, 729, 866, | Nux vanica, 6, 866 |
| Nelson, S. B., 96, 97, 109, 378, 462, 795, 901 | | Family, 648-689 Tree, 74 |
| 378, 462, 795, 901 | Nicoulin, 531 | 1 ree, /4 |
| Nelumbo, American, 108 hitea, 108, 444 | NIDULARIACEAE, 247, 860 | NYCTAGINACEAE, 423, 839 |
| hitea, 108, 444 | Nierembergia Trippomanica, | Nymphaca advena, 839 |
| nucifera, 444 | 854 | alba, 839 |
| Nemalion multifidum f 104 | Nigella, 89, 446 | lotus, 445 Intea, 839 |
| "Nenta" Disease, 501, 533 "Nenta Lessertia" Disease, 37 | damascena, 446, 846 | inted, 839 |
| "Nenta Lessertia" Disease, 37 | satizia, 846 | polysepala. 444, f. 445 |
| Neottia Nidus-avis, 840 | Nigellin, 446 | stellata, 445 |
| Neovossia iowensis, 219 | Migel On, 733 | Nумрнаеаселе, 108, 414, 839 |
| Nepenthes gracilis, 839 | Nigger-head, 779 | Nyssa aquatica, 664 |
| villosa, f. 498 | Night-blooming Cactus, 730 | sylvatica, 666 |
| Nepeuthaceae, 497 | Cereus, 125, 635 | |
| Nepeta, 710 | Night-flowering Catchfly, f. | 0 |
| cataria, 710 | | |
| cururu, /IU | A37 | |
| | Nightshade, Black, 60, 61, 74, | Oak, 866 |
| hederacea, 131, 710, f. 711 | Nightshade, Black, 60, 61, 74, | Oak, 866 Bark, 403 |
| hederacea, 131, 710, f. 711 | Nightshade, Black, 60, 61, 74, 131, f. 132, 719-721, f. 720, 771 | Bark, 403 Bur, 403 |
| hederacea, 131, 710, f. 711 Nephelium, 89, 607 lappaceum, 850 | Nightshade, Blaek, 60, 61, 74, 131, f. 132, 719-721, f. 720, 771 Common, 132, 719-721 | Bark, 403 Bur, 403 Chestnut, 403 |
| hederacea, 131, 710, f. 711 Nephelium, 89, 607 lappaceum, 850 Litchi, 607 | Nightshade, Black, 60, 61, 74, 131, f. 132, 719-721, f. 720, 771 Common, 132, 719-721 Deadly, 133 | Bark, 403 Bur, 403 Chestnut, 403 Cow, 403 |
| hederacea, 131, 710, f. 711 Nephelium, 89, 607 lappaceum, 850 | Nightshade, Blaek, 60, 61, 74, 131, f. 132, 719-721, f. 720, 771 Common, 132, 719-721 | Bark, 403 Bur, 403 Chestnut, 403 |

Castor, 52, 56, 594, 595, 864, 866
Cedar, 101, 330
Chamomile, 751
Cheken, 640
Chinese Wood, 587
Cinnamon, 86
Citron, 582
Citronella, 345
Clove, 93, 639
Cocos, 627
Corn, 342
Creosote, 330
Croton, 6, 72, 78, 149, 588, 589, 866
Cumin, 648
Curcas, 78
Dill, 648 Oleander, 75, f. 129, 130, 148, 691, f. 694, 695, 866 Oleandrin, 148, 866 Olearia argophylla, 755 macrodonta, 816 moschata, 816 Oleaster Family, 125, 640-642 Russian, 641 Olein, 683 European, 105
Evergreen, 403
Galls, 403
Live, 403
Pin, 403
Pin, 403
Poison, 609, 610, f. 611, 612
Red, 403, f. 404
Scarlet, 403
White, 403
Oat, Rust, 99
Smut, 215-217, f. 216
Oats, 215, 225, 338-339, 538
Black, 66
Common, 102, 359, f. 360
Cultivated, 359
Rust of, 225
Smut, 98, 215-217, f. 216,
Wild, 102, 215, f. 358, 359
Obigenin, 560
Ochna, 839
Ochnaceae, 339 European, 105 Oleoresin of Male Feru, 419 Oleson, 654 Olive, E. W., 222, 248 Olive, 683 Oleson, 654
Olive, E. W., 222, 248
Olive, 683
Family, 128, 683, 684
Oli, 608, 683
Wild, 641
Oliver, F. F., 894, 901
Omphalocarpum, 89
procerum, 63, 681, 851
Omphalophlebitis, 179
ONAGRACEAE, 643-647, 840
Oncocarpus witiensis, 805
Onion, 150, 375, 383
Poisoning, 384
Wild, 104
Onoclea, 315, 320-321
sensibilis, 320
Struthiopteris, 321
Ononid, 531
Ononis, 531
Ononis spinosa, 531
Onopordon Acanthium, 816
OOMYCETES, 154, 204-209
Oospora, 13, 294
porriginis, 17, f. 294
pholic acid, 689
Ophiocaulon gummifera, 841
OPHIOGLOSSACEAE, 313, 692
Ophiopogon, 827
Ophthalmia Jequirity, 56
Ophuls, 306
Opium, 2, 6, 7, 72, 84, 112, 144, 145, 395, 479, 481, 482, 866
Alkaloids of, 145, 146, 431
Consumption of, 59
Indian, 85
Poisoning by, 6, 59, 77, 78
Poppy, 112
Properties of, 482
Smyrna, 85
Statistics of deaths from, 59
Toxicology of, 482
Turkey, 481
Openhice, 402 Curcas, 78 Dill, 648 Eucalyptus, 640 Eupatorium, 770 Eupatorium, 770
Fennel, 86
Gaultheria, 665, 866
Geranium, 506
Hedeoma, 711
Hops, 406
Horseweed, 778
Jasmine, 86
Juniper, 101
Kapok, 624
Lavender, 86, 709
Lemon, 583
Lemon Grass, 344
Linseed, 581
Lupin, 548
Madia, 755
Marcassa, 606
Merbane, 6
Mitcham, 86 Ochna, 839
Ochnace, 839
Ochsner, A. J., 183
Ochrolechia tartarea, f. 308
Ochroma Lagopus, 621
Ochrosia, 691
Moorei, 807
OCHROSPOREAE, 235
Ocimum Basilicum, 709
Thymus, 713
viride, 828
Ocotilla Wax, 627
Odors, Fishy, 93
From decomposition, 93
From poisonous flowers, 64, 0ily, 93 rioui poisonous flowers, 64, 134
Oily, 93
Pig-peu, 98
Oedema, 177, 866
of glottis, 78
of lungs, 78
malignant, 175
Oenanthe crocata, 72, 648, 649, 856
fistulosa, 4, 856
Lachenalii, 856
Pluellandrium, 648, 649, 857
Oenanthotoxin, 648
Oenothera, 644
Oesterlein, 642
Oesterlein, 642
Oesterlein, 642
Oesterlen, 503
O'Gara, P. J., 96, 107, 324, 432, 433, 550, 565, 593
Ohio Buckeye, 123, 617, f. 619
Oidiomycosis, 306 Mitcham, 86 Mustard, 7, 77, 114, 489, 575 Myrcia, 640 Myrth, 575 Niger, 755 Nutmeg, 445 Olive, 683 Orange, 86 Orange, 86
Palm, 370
Palmarosa, 344
Peanut, 521
Pennyroyal, 131
Peppermint, 86, 87, 93, 709, Feppermint, 86, 87, 93, 709, 866
Pine needles, 774
Pinhoen, 587
Poley, 78
Poley, 78
Rounculus, 459
Rhodium, 699
Rose, 503, 506
Rosemary, 86
Sassafras, 86
Savin, 149, 866
Sesame, 698
Sunflower, 756
Sweet Birch, 86
Tanacetim, 789
Tansy, 756, 789-790, 866
Tolene, 528
Turpentine, 329, 330, 608
Vervain, 860
Wintergreen, 404, 621, 665
Wormseed, 107, 428
Levant, 791
Wormwood, 793 866 Statistics of deaths from, 59
Toxicology of, 482
Turkey, 481
Openheim, C., 902
Onsonin, 165, 177, 178, 182
Opintia, 125, 635, 636, 637
Dillenii, 635
Englemanni, f. 636
Ficus-Indica, 635
Karwinskiana, 637
Larrevi, 635
Pereskia, 635
polycantha, 635
Rafinesquii, 125, 635
Striptacantha, 635
Tuna, 635
Tuna, 635
Tuna, 635
Tuna, 635
CPUNTIALES, 157, 634-637
Opwyrda, R. J., 902, 906
Orange, 582, f. 582, 866
Bitter, 582
False, 866
Mock, 500
Oil, 36
Osage, 106, 406, 414-415
Root, 108
Sour, 582
Orchard Grass, Powdery Mildew of, 272
Rust on, 279
Soot disease of, 282, f. 283
Orchid Family, 392-395 Oidiomycosis, 306 Oidium, 100, 154, 249, 303-306
albicans, 12, 100, 302, 303
Furfur, 298
granulomatogenes, 305
hominis, 303-305
lactis, f. 17, 299
lithogenes, 305
minimum, 298
moniliodes, 272
tonsurans, 301
ii 866 tonsurans, 301
Oil, 866
Almond, 6, 315, 608
Anise, 648
Anthemis, 789
Attar of Roses, 86
Avocado, 478
Bay-berry, 639
Bergamot, 86, 583
Bitter Almond, 6, 315, 503
Bone, 145
Boneset, 70
Buttertree, 680
Camphorin, 478
Carapa, 575
Caraway, 648 Wormwood, 793 Wormwood, 773 Okra, 623 OLACEAE, 53, 839 Old Man Cactus, 635 Oldenburgia arbuscula, 816 Oldenlandia senegalensis, Oldenlandia senega 849 umbellata, 849 Olea dioica, 840 curopaea, 683 glandulifera, 683 Caraway, 648 Cashew, 608, 864 Cassia, 86, 523 OLEACEAE, 683-684, 840

INDEX

| ORCHIDACEAE, 90, 105, 392- | grandis, 580 | Pancovia, 89 |
|---|--|--|
| ORCHIDACEAE, 90, 105, 392- 395, 417, 692, 840 Orchids, 524 Orchis, 90 coriothora, 840 | lasiandra, 580 | Pancratium illyric |
| Orchids, 524 | Pes-coprae, 840 purpurata, 840 | maritimum, 805 |
| Orchis, 90 | purpurata, 840 | zeylanicum, 80! |
| | Smithiana, 840 | PANDANALES, |
| maculata, 392 | tetraphylla, 580 | Pandanus, 332 |
| odoratissima, 840 purpurea, 840 Simia, 840 Orcutt, C. R., 902 Ordeal Bean, 528 Plant, Madagascar, 148 Poisons 804 866 | violacea, 120, 580 Oxaluria, 73 | /ragrans, 333 |
| Simia 840 | Oxy-benzyl-thiocyanate, 489 | odoratissimus, 8 |
| Orcutt, C. R., 902 | Oxycannabin, 106 | Thomensis, 841 |
| Ordeal Bean, 528 | Oxycanthin, 112, 472 | Pandorina, 92, 93 |
| Plant, Madagascar, 148 | Oxycannabin, 106 Oxycanthin, 112, 472 Oxycoccin, 666 Oxygen in alkaloids, 144 | 193 |
| Poisons, 804, 866 Tree, 692 Oregon Grape, 112, 472, f. | Oxygen in alkaloids, 144 | Moruni, f. 192 |
| 1 ree, 692 | Oxydendron arboreum, 821 | Paneolus papilion |
| Oregon Grape, 112, 472, 7. | Oxylobium parviflorum, 533, 832 | Pangium, 52 edule, 53, 54, 8 Panic Grass, Bla |
| Water Hemlock, 656-658, /- | Oxypolis, 650, 661 | Panic Grass, Bla- |
| 6.57 | Oxypolis, 650, 661 rigidior, 126, 661, 857 | 280 |
| 657 Orfila, M. J., 3, 5, 902 Organic acids, 150 | Oxyria digyna, 419 Oxytropis, 535, 564, 567-570 deflexa, 567 foetida, 832 Lamberti 30, 110, 564, 565 | Panicled Dogwood |
| Organic acids, 150 | Oxylropus, 535, 564, 567-570 | Panicum, 53, 89, junceum, 826 |
| Origanum noriounaum, 710 | loctida 932 | sanguinale, 826 |
| Marjorana, 710 Ormocarpum glabrum, 832 Ormosia coccinea, 832 | Lamberti 39 119 564 565 | Papa. 297 |
| Ormosia coccinea, 832 | Lamberti, 39, 119, 564, 565, 566, 567-570, 832 Distribution, 570 lapponica, 567, 832 | Papa, 297 Papain, 597, 627 Papaver, 480-483 |
| dasycarpa, 527 | Distribution, 570 | Papaver, 480-483 |
| Ormsby, 306 | lapponica, 567, 832 | aculeatum, 841 |
| OROBANCHACEAE, 698 | muumorus, 507 | andiani, 113 |
| Ornithin, 10 | spicatus, 569 | orientale 113 |
| muscari. 377 | sulphurea, 567, 832 Ozonium, 154 | dubium, 113 nudicaule, 481 orientale, 113 Rhoeas, 112, 42 |
| nutans, 835 | | somniferum, 4, 111, 112, 481-482, 84 |
| thyrsoides, 377, 835 | · P | 111, 112, |
| Ornithogalum, 835, 864, 866 muscari, 377 nutans, 835 thyrsoides, 377, 835 umbellatum, 377, 835 | D | 481-482, 84 |
| OROBANCHACEAE, 840 | Pabsch, H., 902 | Papaveraceae, 11 479-485, 48 Papaveramin, 147 |
| Orobanche minor, 698 | Pachira macrocarpa, 624 | Panaveramin 147 |
| ramosa, 698 Orobus, 52 | Pachistima, 614 Cambyi, 614 | Papaverin, 59, 11 |
| Oroxylin, 739 | Myrsinites, 614 | 481 |
| Oroxylon indicum, 739 | Pachygone, 52 | Group, 147 |
| Orpine Family, 114, 501-503 | ovata, 2, 838 | Papaw, 111, 476, |
| Orpine Family, 114, 501-503 Orr, S. S., 904 Orris Root, 389 | Pachyrhizus angulatus, 521, | American, f. 47 |
| Orris Root, 389 | 832 | Paper, 404 Bark, 642 |
| Osage Orange, 106, 406, 414- 415 | Inberosa, 832 | Mulharm 106 |
| Oscillaria, 93 | Paederia foetida, 849 Paeonia Moutan, 446 | PAPILIONACEAE, 5. |
| Oscillaria, 93 Oscillatoria, f. 92, 97, 184, | | Paprika africana, |
| 182-180 | Pacony, 446 Page, C. G., 902 Paige, J. B., 902 Painted Cup, 734 Painted Cup, 734 | Pappoose Root, 46 |
| OSCILLATORIACEAE, 97, 185- | Page, C. G., 902 | Papyrus, 369 |
| 186 Osier Red 664 | Painted Cun 734 | Paradisea Liliastr |
| Osier, Red, 664 Osmanthus frograns, 683 Osmohydrophora nocturna, | Paint-root, 386 | Paralysis, 74, 804 Rulbar, 174 Caused by Aroic |
| Osmohydrophora nocturna, | Paint-root, 386 Paitrier, 897, 902 Pale Dock, 419, 420 Mold, f. 261, 262 Spiked Lobelia, 136 | Caused by Aroie |
| 53, 810 | Pale Dock, 419, 420 | Parasitic Diseases |
| Otmorhien longistulie 648 | Mold, /. 261, 262 | Poisons, 10 |
| Osmunda, 322, f. 322 cinnamomea, 313, 322 | Spiked Lobelia, 136 | Yeast, 249 |
| Claytoniana, 101, 313, f. | Palicourea rigida, 849 Paliurus aculeatus, 847 | Paregoric, 6 PARIETALES, 1 |
| 231, 322 | australis, 621 | Parillin, 62 |
| 231, 322 regalis, f. 309, 313, 322 | Pallin, W., 902 | Paris, 89 |
| OSMUNDACEAE, 101, 313, 321- | Pallin, W., 902 Palm, 370, 523 | obovata, 835 quadrifolia, 4, 3 |
| Ostaitis 173 | Betel-nut, 370 | quadrifolia, 4, 3 |
| Osteitis, 173 | Cocoa-nut, 370 Corozo, 370 | Park, R., 903 |
| Osteonicles, 53 arbutifolia, 847 | Corypha, 370 | Park, Davis and (|
| Osteomyelitis, 167, 169, 173 | Date, 370 | Parker, G. H., 93, Parker, W. T., 9 |
| Ostertag, 26 Ostrich Fern, 321 | Date, 370 Oil, 370 | Parkia africana, 8 |
| Ostrich Fern, 321 | Royal, 370 | Paronychia argent |
| Oswego Tea, 614, 629 Othonopsis intermedia, 816 | Traveler's, 391 | honariensis. 827 |
| | Washingtonia, 370 Wine, 370 | honariensis, 827 capitata, 827 |
| media, 169, 177 | PALMACEAE, 840 | Parry's Aster, 776 |
| Otomycosis, 199, 262 | PALMAE, 90, 370, 417 | Pine, 128 |
| Ottis, 171 media, 169, 177 Otomycosis, 199, 262 Ott, Dr. J., 564, 902 Otto, K., 862, 902 Ougeinia dealbergioides, 832 Oxalates, 72, 150, 866 OXALIDACEAE, 120, 575, 577, 579-580, 840 | PALMACEAE, 840 PALMAE, 90, 370, 417 Palmarosa Oil, 344 | Parmena vuipina, |
| Otto, K., 862, 902 | Palmatin, 450 | Parsley, 126, 648, |
| Oralates, 72, 150 | Palmer C C 432 | Family, 126, 64 Fool's, 76, 126, |
| Oxalic acid. 6, 72, 150, 866 | Palmarosa Off, 344 Palmatin, 450 Palmer, Dr., 237 Palmer, C. C., 432 Palmer, Dr. Edward, 357 Palmitic acid 503 | Parsnip. 126, 65 |
| OXALIDACEAE, 120, 575, 577. | Palmitic acid, 593 | Parsnip, 126, 65 866 |
| 579-580, 840 | Palmitin, 594 | Cow, 126, 663 |
| 579-580, 840 Oxalis, 150, 579-580 | Pammel, L. H., 883, 898, 902 | Cow, 126, 663 Cultivated, 662 Water, 126, 659 |
| 71 CE 10 3 E 11 U, 0 TO | Palmitin, 594 Pammel, L. H., 883, 898, 902 Panaeolus papilionaceus, 244 Panax, 647, 808 ginseng, 647 quinquefolium, 647 Pancin, 51 | Water, 126, 659 |
| amara, 840 corniculata, 230, 580 | ginseng 647 | Creening f |
| crenata, 580 | quinquefolium, 647 | Wild, 49, 650. |
| crenata, 580 flava, 580 | Pancin, 531 | Cut-leaved, 76 Creeping, f. 6 Wild, 49, 650, 663, f. 662 |
| | | |

icum, 805 5 05 41 , 332 841 3, 98, 191, iaceus, 860 82, 747 ack-spot on, d, 664 826 479, 483, 841 4, 60, 73, f. 479, f. 481, 41 112-113, 144, 86, 841, 866 13, 146, 147, 477 76, 476-577 530 , 523 169, 484 rum, 835 ids, 371 s, 866 157, 627-634 377, 835 Co., 613 3, 903 903 832 itea, 436 861 5, 650, 651 647-664 5, 659, f. 660 651, 661-663. 59 76 . 650 . 0, 656, 661

Parthenium, 757
Hysterophorus, 816
integrifolium, 816
Parthenogenesis, 208
Partridge Berry, 742
Pea, 117, 529, f. 536, 536-Pea, 11 537 7 Paspalam, 339, 349
Scrobiculatum, 349, 826
Pasque Anemone, f. 454
Pasque-flower, 109, 446, 453.
455, f. 454, 456
Passiflora caerulea, 633, 841
edulis, 633
foerida, 747, 841
Herbertiana, 841
hispida, 841
incarnata, 633, 841
laurifolia, 841
quadrangularis, 841
rubra, 841
PASSIFLORACEAE, 53, 89, 628, 633, 841 633, 841 Passion Flower, Blue, 633 Fassion Flower, Ditte, 033 Family, 633 Pasteurellosis, 177 Pastinaca, 650, 661-663 satica, 126, 661-663, f. 662, 857 Patchouli, 710 Patek, J., 903 Pathogenic Poisons, 1 Paul, B. G., 878, 903 Paullinia, 89, 123 costata, 850 Cupana, 606, 850 curassavica, 850 macrophylla, 850
meliaefolia, 850
thalictrifolia, 850
trigonia, 850
Paulowilhelmia polysperma, Paulowillelmin polysperma, 804

Speciosa, 804

Pauwels, W. M. I. B., 52, 903

Pavesi, V., 903

Pavesi, V., 903

Pavin, 52

Pavin, 617

Pavonia zeylanica, 837

Pawpaw, 627

Payena latifolia, 53, 681, 851

Payer, G. T., 903

Pea, 531, 866

Chick, f. 110, 120, 521, f. 573, 573-574

Everlasting, 572-573

Field, 521 Field, 521 Field, 521 Fodder, 45 Garden, 1. 521, 521 Hoary, 558 Hog, 204 Idaho, 120 Jack, 521 Partridge, 117, 529, 536-537 117, 529, f. 536,

Peck, C. 11., 96, 234, 238, 240, 284, 803, 903 Peckolt, Th., 903 Pectenin, 637 Pectenin, 637
PEDALIACEAK, 698
Pediastrum, f. 02, 190
Pedicularis, 734, 738
bracteosa, 738
canadensis, 134, 734, 738
groenlandica, 134, 738
lanceolata, 134, 738
palustris, 738
racemosa, 738
sudetica, 738
sylvatica, 738, 852
Pedilanthus titlyymaloides,
823 823 Pedler, A., 903, 915 Peganum antidysintericum, 849 Harmala, 849, 858 Peinemann, Dr., 88 Pelargouinm, 5 peltatum, 825 South African, 577 zonale, f. 578 Peli, 747 zonale, f. 578

zonale, f. 578

Peli, 747

Peli, 747

Pellagra, 11, 77, 287, 34.

Pellagrocein, 11

Pellate, 637

Pelletier, Joseph, 3

Pellew, C. E., 888, 903

Pellicotti, 572

Pellicotty, 754

Root, 754

Pellizi, 290

Pellon, 11, 11., 903, 916

Pelosin, 473, 478

Pellotinia, 866

Peltandra virginica, 372

Peltigera horizontalis, 861

Penhallow, D. P., 903

Penicillium, 256-261, f. 258

glaucum, 24, 82, f. 25

zofo, 260, 505

minimum, 256-257 287, 343, 255, 256, 260, 505
minimum, 256-257
Penn, W. A., 903
Pennsylvania Persicaria, 421
Pennycress, Field, 495, f. 495
Pennyroyal, 709, 711
American, 711
Oil of, 131
Pennywort, Indian, 648
Pentactethra macrophylla,
531, 832
Pentstemon, 733 531, 832 Pentstemon, 733 grandiflorus, 733 Pentsia virgata, 816 Peperomia 396 Peper, 78, 396 Black, 77, 87, 396, f. 396 Cayenne, 77, 87, 133, 714, f. 725, 725,726 Guinea, 725 Red, 87, 133, 714, f. 725, 725,726 Strubby, 133 Shrubby, 133 Tree, 608 White, 87 Peppergrass, 486, 493 European, 486, 493 Large, 494 Small, 113, f. 494, 494 Peppermint, 709, 713, f. 713, 866 Japanese, 709 Oil of, 86, 87, 709 Pepsin, 373 Peraphyllum ramosissimum, 847 Percival, C. H., 903 Percy, S. R., 903

Pereira, Jonathan, 59, 482 Pereira Bark, Alkaloids of, Pereira, Jonathan, 57, 402 Pereira Bark, Alkaloids of 147 Pereirin, 147 Perezia oxylepis, 816 Pericampylos incarius, 838 Pericarditis, 169, 171 PERIDINALES, 153, 188 Pericarditis, 173 Periostetis, 173
Periostetis, 173
Periostetis, 171
Periploca graeca, 695, 809
vomitoria, 809
PERISPORIALES, 268-273
Perithecium of Aspergillus, f. Peritonitis, 167, 169, 171, 172, Periwinkle, 691 Perkins, J., 903 Pernossi, L., 883, 903 Peronospora alsinearum, f. calotheca, f. 205 leptosperma, f. 205 parasitica, 206 Schleideniana, 205 Parasinica, 206
Scilieideniana, 205
trifoliorum, 205, f. 206
Peronosporaceae, 204-208
Perredes, P. E. F., 903
Perret, A. H., 903
Perrot, Em., 903
Perssa gratissima, 478
Persian Cyclamen, f. 677
Persicaria Pennsylvanica, 421
Persimmon, 128, 681
Indian, 681
Japanese, 681
Peruvian Bark, See Bark
Petalostigma cordifolia, 823
Petasites officinalis, 816
Peters, A. T., 96, 214, 290, 291, 325, 346, 347, 869. 903, 911, 912
Peterson, F., 887, 904
Peterson, Maude G., 904
Petiveria alliacea, 841
tetandra, 841
Petrioria alliacea, 841
Petrioria alliacea, 841
Petrioria alliacea, 841 Petiveria unidea, tetandra, 841 Petroleum, Poisoning by, 6 Petroleum, hortense, 126, 648 648.
Petunia violacea, 716
Peucedanum ambiguum, 857
foeniculaceum, 857
graveoleus, 648
officinale, 648
Oreoselinum, 857 Oreosclinum, 648
Ostruthium, 857
Peucedone, 648
Peyre, A., 904
Peziza, 253
Pfaff, F., 613, 904
Phacelia, 703-704
circinata, 131
Hairy, f., 702, 704
Menziesii, 704
Rough, 131
RAUDIACEAE, 253 ROUGH, 152 PHACIDIACEAE, 253 Phacidium Medicaginis, 253 PHAEOSPOREAE, 194 PHAEOSPOREAE, 194 Phajus callosus, 840 Phalonopsis amabilis, 840 392, Lueddemanniana, 840 On, f. 27
Phalen, J. M., 901, 904
Phaliota radicosa, 53
PHALLACELE, 99, 245, 860
Phallin, 32, 35, 80, 240, 241, 242 Phalaris arundinacea, Ergot Phallogaster, 245 PHALLOIDEAE, 245

| Phallus duplicatus, 860 | epiphyllanthus, 823 | Pigweed, 427, 431-433, f. 433 |
|---|--|---|
| impudicus, 239, 245 Ravenelii, 860 Phares, Dr., 782 | Gastromei, 823 | Common, f. 426, 427 |
| Phares Dr. 782 | lacunarius, 823 Niruri, 823 | Green, 107 |
| Pharmacognosy, 866 | piscatorum, 823 | Prostrate, 431, f. 432 Slender, 432 |
| Pharmacographia, 866 | urinaria, 823 | Slender, 432 Thorny, 107 |
| Pharmacopoeia, 866 | Phyllocactus, 625 | Pilea pumila, 409 Pilocarpidin, 582 Pilocarpin, 77, 78, 80, 146, 148, 582, 731, 732 |
| American, 866 British, 866, 874 | Phyoscyamin, 248 Physalis Alkekengi, 854 | Pilocarpin 77 78 80 146 |
| French, 866 | foetens, 854 | 148, 582, 731, 732 |
| German, 866 | virginiana, 854 | ruocarpus, 849 |
| Italian, 866, 882 Swedish, 866 | Physica bulgarylanta 6 | officinalis, 849 |
| Pharyngitis, 170 | Physcia pulverulenta, f. 309 Physcocyanin, 184 | pennatifolius, 148, 582, 849 racemosus, 849 |
| Phaseum cuspidatum, f. 300 | Physic Nut, Cuban, 596 Physic Nut, Cuban, 596 Physocarpus apulifolius, 504 | spicatus, 849 |
| Phaseo-lunatin, 54 Phaseolus, 52, 535, 574, 866 | Physocarpus apulifolius, 504 | Pilocerein, 637 |
| acanitifolius, 832 | Physostigma, 866 | Pilocereus Sargentianus, 637 Pilalobus crystallinus, 195 |
| angularis, 526 lunatus, 53, 54, 75, 87, 119, 520, 574, 747, 832 | venenosum, 85, 148, 528, 531, 832 | Pilz-atropin, 238 |
| lunalus, 53, 54, 75, 87, | Physochlaina orientalis, 854 | Pilzmerkblatt, 904 |
| multiflarus, 119, 520, 574, | praealta, 854 Physostigmin, 146, 148, 528, | Pimelia trichostachya, 642 Piment, 639 |
| 832 | _ 531, 866 | Pimenta acris, 839 |
| Munga v. glaber, 520, 574 | Poisoning by, 77, 78, 81 | officinalis, 639 Pimpernel, 128, 676 Pimpinella Anisum, 648 |
| semierectus, 832 | Phytclephas macrocarpa, 370 | l'impernel, 128, 676 |
| trilabus, 520 vulgaris, 520, 574, 832 | Phytobezoars, 866 Caused by Common Oats, | Saxifraga, 648, 857 |
| Phellandrene, 649 | 102, 339 | Saxifraga, 648, 857 Pimpinellin, 648 |
| Phellodendron amurense, 581 | Crimson Clover, 68, 118, | Pin Oak, 403 |
| Phenol, 330 Philadelphus caronarius, 115, | 556, 567 Helichrysum, 755 | Pinaceae, 101 Pine, 101, 328, 329-330 |
| 500, 851 | Opuntia, 635-636 | Austrian, 329 |
| grandiflarus, 115, 500, 851 Lemainei, 500, 851 | Phytalacca, 434-435, 866 | Jack, 330 |
| Lewisii, 851 | abyssinica, 63, 434, 841 acinosa, 841 | Loblolly, 330 |
| micraphyllus, 115, 500, 857 | decandra, 107, 150, 433-435, | Lodgepole, 330 Long-leaved, 329 |
| Philippines, Poisonous plants | f. 434 dioica, 434, 841 | Norfolk, 327 Screw, 332 Sugar, 329 |
| of the, 867 Phillips, 904 | dioica, 434, 841 icosandra, 841 | Screw, 332 Sugar, 320 |
| Phillygenin, 683 | littoralis, 434 | |
| Phillyrea media, 840 | Tincture of 434 435 | White, 329, f. 329 |
| Phillyrin, 683 | PHYTOLACCACEAE, 63, 89, 107, | Pineapple, 373 |
| Philodendrou bipinnatifidum, 808 | 423, 424, 433-435, 841- 842 | White, 329, f. 329 Pineapple, 373 Family, 372 Wild, 373 Pinellia tuberifera, 808 Pinene, 330, 640, 710 d. Pinene, 648 |
| hederaceum, 808 | Phytolaccie acid 435 | Pinellia tuberifera, 808 |
| Imbe, 808 Simsii, 808 | Phytolaccin, 107, 435 | Pinene, 330, 640, 710 |
| Phlegmon, 171 | Phytolaecotoxin, 435 | d-Pinene, 648 "Piney" Resin, 627 |
| Phicum praiense, Ergot on, | Phytophthora infestaus, 205, 208 | Pingnicula vulgaris, 698, 833 |
| 2/6 | Phaseoli, 208 | Pinguin, 373 Pinhoen Oil, 587 |
| Phlorizin, 505 Phoebus, Philipp, 4, 803, | Phytotoxins, 11 | Pinhoen Oil, 587 |
| Phoebus, Philipp, 4, 803, | Picea canadensis, 328 Engelmanni, 327 | Pinicrin, 328 Pink, Deptford, f. 437 |
| Phoenix, 90 | excelsa, 328, 817 | Family, 435-444 |
| dactylifera, 370, 840 | mariana, 328 | Hardy 436 |
| Phalidia macnlata, 838 Pholiota caperata, 860 | sitchensis, 327 Pichi 716 | Indian, 129, 688 Moss, 423 Rose, 689 |
| radicasa, 860 | Pichi, 716 Pick, F. J., 295 Pickerel-weed, 374 | Rose, 689 |
| Phoradendron, 106, 415 | Pickerel-weed, 374 | reast, 250 |
| flavescens, 106, 415, f. 416, 836 | ricolin, 728 | Pink-root, 129, 688, 866 East Tennessee, 688 |
| Phormium tenax, 375 | Picraena excelsa, 853 Picrasma quassioides, 853 | Pinkham, 904 |
| Phosphorus, 72 | Pieric acid, 72 | Pinnularia viridis, f. 189 |
| Poisoning by, 6 | Picris hicraciaides, 816 | Pinns, 101, 329-330 |
| Phatinia, 53 Phragmidium incrassatum, | Picropodophyllin, 112, 470 | Banksiana, 330 contorta, 330 |
| 225, f. 225 Rubi, 221 | Picrotin, 149 | echinata, 330 |
| Kuoi, 221 | Picrotoxin, 1, 77, 80, 112, 149, 435, 473, 475, 534, | heterophylla, 330 |
| Phragmites, 217 | 607 | Lambertiana, 329 Laricio, 329, 817 palustris, 329, 330 pinaster, 330 |
| Ergot on, 276 | Picrotoxinin, 149 | balustris, 329, 330 |
| Communis, 219 | Pietet, A., 728, 878, 904, 917 | pinaster, 330 |
| Phycamyces, 195, 197 | Pictou Disease, 140, 795 Pie-plant, 150, 418 | ponaerosa, 330 |
| nitens, 2 | Pierce, H. C., 440 | panderosa v. scopulorum, 330 |
| nitens, 2 PHYCOMYCETES, 98, 154, | Pieris floribunda, 821 | Strobus, 329, f. 329 |
| 193-209, 281, 859 | formosa, 65 mariana, 821 | sylvestris, 330, 817 |
| Physician, 194 | mariana, 821 mitida, 821 | Taeda, 330 |
| Phyllachora, 279-280 graminis, 280, f. 280 Trifolii, 279 | ovalifalia, 65, 821 | Pinweed, 627 Piner, C. V., 526, 904 |
| Trifolii, 279 | Piffard, H. G., 904 | Piper, C. V., 526, 904 Piper, 52 |
| Phyllanthus, 52, 89 Conami, 823 | Pigeon Grass Smut, 98 | angustifotium, 390 |
| Emblica, 823 | Pigeon Pea, 521 Pignut, 402 | Betle, 396 Carpunya, 842 |
| , 5-0 | | ourpunya, 0.72 |

Chaba, 396 Plasmopara, 195 Halstedii, 205 viticola, 205 Podosphaera tridactyla, f. 269, Cubeba, 396
Cubeba, 396
dariense, 842
longnm, 396
methysticum, 396, ft. 396, 842
nigrnm, 77, 396, ft. 396, 842
peltatum, 396, 842
peltatum, 396, 842
PIPERACEAE, 52, 396, 842
PIPERACEAE, 52, 396, 842
PIPERALES, 156, 396
Piperin, 396
Piperovatin, ft. 754
Pipsissewa, 665
Pipewort, 372
Piptadena peregrina, 832
Pircunia, 89, 842
Pircunia, 89, 842
Pircunia, 89, 842
Pircunia, 89, 842
Piscidia, 1, 52
Erythrina, 74, 832
Pishck, A., 904
Pisidium montanum, 839 Cubeba, 396 269 Podostemon Aceae, 498 Pogostemon Heyneanns, 710 viticola, 205
PLATANACEAE, 498, 500, 842
Platanns, 84
acerifolia, 500
coneata, 500
occidentalis, 500, 842
orientalis, 500, 842
Plato, C. G., 905
Platopuntia, 636
Platte Cedar, 330, 331 Poet's Narcissus, 386 Pohl, J. E., 905 Poikilocytosis, 169 Polistocytosis, 169
Poinsettia, 588
Poison Box, 533
Berry, 378
Darnel, 361-364, f. 362, 363
Defined, 1
Distribution in plants, 82
Hemlock, f. 50, 51, 72, 75, 648, 651
Ivy, 77, 122, 123, 394, 600, 608, f. 609, 609-613, 676, 679, 862
California, 123, 609
Lore, 866
Oak, 609, f. 611, 612
Register, 905
Sago, 378
Sumac, 609, 610, f. 610
Toot, 868
Tree, 866
Use of to plant, 82
York-road, 533
Foisoners, Professional, 2
Poisoning by Aconite, 380
Cas, 5
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cryptogamic, 20-37
Cr Poinsettia, 588 Platte, Cedar, 330, 331

Platygyna urens, 823

Plaut, H. C., 13, 199, 265, 295, 302

PLECTASCINEAE, 252, 256.267 236-267
Plectospora minutula, f. 307
Plectronia dicocca, 53, 849
Plenck, J. J., 3, 905
Pleurisy-root, 130, 695
Pleurisis, 167, 169, 170, 171
Pleurococcus, 190
Pliny, 2, 64, 365, 601, 749
Ploss, 61
Plowright, C. B., 35, 905
Plugge, P. C., 65, 905
Plugge, P. C., 65, 905
Plum, 505, 514
Chickasaw, 505, 514
Ground, f. 563
Hog, 608
Leaco, 505
Japan, 505, 514
Native, 614
Northern Wild, 505 Pisidium montanum, 839 Pisonia obtusata, 839 tomentosa, 839 Pistachia Khinjuk, 805 Lentiscus, 608
Nut, 608
Terebinthus, 608
vera, 608
Pisum arvense, 521 Poisoners, Trofessional, 2
Oisoning by Aconite, 380
Gas, 5
Cryptogamic, 20-37
Deaths from in England, 6
United States, 5, 6
Wales, 6
for criminal purposes, 5
Mushroom, 237, 241
practiced in India, 2
Statistics of, 5-6
Stock in Montana, 6
Symptoms of, 73, 74, 75, 76, 77-79, 868
Treatment for, 73, 74, 75, 76, 77, 79-81
Poisonous plants, Bibliography, 862-918
Poisons, Action of in plants, 82
Ancient use of, 2,3 sativum, f. 521 Pitch, Burgundy, 328 Northern Wild, 505 Pocket, f. 252 Sapodilla, 679 Wild, 116, f. 506 Northern, 505 Canada, 327 Hemlock, 327 Hemlock, 327

Pittecolobium, 1, 89, 534

Pittecolobium, 832

dulce, 525
fasciculatum, 832

Minehasse, 832

montanun, 832

Saman, 527, 530, 832

Pittosporacae, 89, 498, 842

Pittosporacae, 89, 498, 842 PLUMBAGINACEAE, 128, 675, 842 Plumbago capensis, 842 europea, 842 Family, 128 rosea, 842 Pittosporum, 89 coriaceum, 842 cornifolium, 842 crassifolium, 842 eugenioides, 842 ramily, 128
rosea, 842
scandens, 842
toxicaria, 842
Phlox, 698, 843
divaricata, 698
Drummondii, 698
maculata, 698
Perrenial, 698
Plumiera rubra, 807
Pneumonia, 171, 176, 177
Acute infectious, 169
Epidemic from parrots, 176
Hypostatic, 37
Purulent, 170
Pneumonomycosis, 29, 199, 263, 264
Pneumotoxin, 169
Poa, 276 Ancient use of, 2,3 Antidotes to, 2, 868 Classified by Blyth, 72 Brandt and Ratzeburg, 6 ferrugineum, 842 Roribundum, 842 Moribindum, 842
Huttonianum, 842
Huttonianum, 842
Moorei, 842
Moorei, 842
phillyracoides, 842
rhombifolinm, 842
tobira, 842
undulatum, 842
Piturie, 147, 716
Piturin, 716
Pius III poisoned, 2
Planchon, Gustave, 904
Planchonia valida, 638 Buchner, 5 Dioscorides, 2 Proscorides, 2 Fodere, 5 Kobert, 72-73 Orfila, 5 Smith, 73-77 Sobernheim, 5 Detection of, 3, 866 History of, 2 Indian use of, 1 Kinds Pneumotoxin, 169
Poa, 276
annua, Ergot on, 276
Septoria on, 285
pratensis, 277, f. 339
Powdery Mildew on, f.
271, 272
Ergot on, 276
serotina, Powdery Mildew
on, 272
Wolfs Pawdery Mildew on Kinds Planchonia valida, 638 Abortives, 76 Abrus, 5 Plane Tree, 84 Plankton, 93 Aconite, 5 Acrid, 5, 803 Acrid narcotic, 5 Planococcus citrus, f. 161 Planococcus citrus, f. 161 PLANTAGINALES, 158, 739-740 Agaricus, 5 Alkaloidal, 803 Animal, 2, 5 Asthenic, 75, 804 Astringent, 5, 804 Blood, 5, 72 Boletus, 5 739-740

Plantago arenaria, 740
indica, 740
major, 740
ovata, 740
Rugelii, 740

Plantain Family, 739-740
Water, 102, 335, f. 335
Family, 335

Plasmodiophora Brassicae, f.
159, 160

PLASMODIOPHORALES,
160 on, 272
Wolfii, Powdery Mildew on, 272
PODALYRIEAE, 530 Boletus, 5 Bryonia, 5 Cardiac, 74, 803 Cicuta, 5 Colchicum, 5 Convulsive, 72, 74, 803 Deliriant, 72, 73-74, 803 Depressant, 74 Podocarpus, 327 Podophyllin, 88, 112, 470, 471 Podophyllotoxin, 76, 88, 112, Podophyllum, 469-471 Emodi, 88, 470, 810 peltatum, 75, 88, 1 470, 470-471, 810 112, f. 160

| Digitalia 5 | Baykini, 843 | Poor-man's Weather-glass |
|--|--|---|
| Digitalis, 5 | Cubaniania 043 | Poor-man's Weather-glass, |
| Dirca, 5 Drastic Resins, 5 | Cyparissias, 843 | 120, 070 |
| Drastic Resins, 5 | gianauiasa, 843 | 128, 676 Poplar, 397 Balsam, 105 |
| Emetic alkaloids, 5 | glandulasa, 843 javana, 843 | Balsam, 105 Papawia pisacarpa, 806 Poppy, 90, 480-483 Alkaloids of, 90, 147, 479- |
| Hot acrid, 5 | Senega, 63, 121, f. 585, 585- 586, 617, 843 venenasa, 586 | Papawia pisacarpa, 806 |
| Hydrocyanic, 5 Hypericum, 5 Inebriant, 74, 804 | 586, 617, 843 | Poppy, 90, 480-483 |
| Hypericum 5 | venenasa, 586 | Alkaloids of 90 147 479. |
| Inchainnt 74 904 | minainiana 586 | 101 |
| Inebriant, 74, 804 | virginiana, 586 | 481 |
| Inflammatory, 5 | POLYGALACEAE, 63, 89, 121, 417, 575, 577, 584-586, | California, 479, f. 48a Common, f. 111, 479 Corn, 112, 483 |
| Iris, 5 | 417, 575, 577, 584-586, | Common, f. III, 479 |
| Irritant, 5, 72, 76, 804 Irritant narcotic, 5 | 843 | Corn, 112, 483 |
| Irritant narcotic, 5 | Polygalic acid, 62, 586 | |
| I olium 5 | POLYGONACEAE, 106-107, 418- | Garden, f. 50, 73, 111, f. 481, 481-483 |
| Lolium, 5 | 422 042 | 10° 401 402 |
| Lycoperdon, 5 | 423, 843 POLYGONALES, 156, 417- | 401, 401-483 |
| Melica, 5 | POLYGONALES, 156, 417- | Long Smooth-truited, 113 |
| Mineral, 2, 5, 866 | 423 | Mallow, 624 |
| More volatile, 5 | Palyganatum, 89 | Oil, 479 |
| Narcissus 5 | biffarum, 835 | Opium, 112 |
| Narcissus, 5 Narcotic, 5, 72, 804 Narcotic acrid, 5 | | Daialda 112 402 |
| Narcotic, 5, 72, 804 | giganteum, 377 | Prickly, 113, 483 |
| Narcotic acrid, 5 | afficinale, 835 | Mexican, 113, 483 |
| Narcotic alkaloidal, 5 | verticillatum, 835 | Red. 479 |
| Nervous, 5 | POLYCONEAE 52 | Populin, 397 |
| Oleander 5 | Polygonic acid 107 | Populin, 397 Papulus balsamifera, 105, 397, |
| Oleander, 5 | Polygonic acid, 107 Palyganum, 52, 419, 421-423 acre, 106, 421, 843 aviculare, 421, 423 | 040 |
| Pepper, 5 Phallus, 5 | 1 alyganum, 52, 419, 421-423 | 849 |
| Phallus, 5 | acre, 106, 421, 843 | candicans, 397, f. 399 deltaides, f. 398 Porcher, F. P., 905 |
| Plant, 2, 5 | aviculare, 421, 423 | deltaides, f. 308 |
| Pokeweed, 5 | barbatum, 843 | Porcher, F. P., 905 |
| | Bistarta, 843 | Porcupine Grace 102 255.257 |
| Poppy, 5 | Causalanda 421 042 | Porcupine Grass, 102, 355-357, |
| Prunus, 5 | Canvalvulus, 421, 843 | f. 356 Smut, f. 216 |
| Rhus, 5 Ruta, 5 | flaccidum, 843 | Smut, f. 216 |
| Ruta, 5 | Hydrapiper, 107, 421, 422- | Parriga asbestinea, 16 |
| Septic, 5 | 423, f. 423, 843 | Porta, G., 3 |
| Smartweed, 5 | hydrapiperaides, 422, 843 | Porter F D 187 |
| Spartium F | Muchlanbargii 421 | Porter, E. D., 187 Porto Rico, Economic plants |
| Spartium, 5 | Muehlenbergii, 421 | Porto Rico, Economic plants |
| Stuperying, 5 | arientale, 421 | of, 864 |
| Stupefying, 5 Unknown, 5 Uraemic, 77, 804 | pennsylvanicum, 42 | Partulaco grandiflara, 423 |
| Uraemic, 77, 804 | Persicaria, 421, f. 422 | aleracea, 423 |
| Vegetable, 5, 866 | sachalinense, 421 tinctarum, 421 POLYPETALAE, 395 | Portulacaceae, 423, 844 Patamagetan, 93, 332, f. 333, |
| Volatile parentie 5 | tinctarum 421 | Patamagatan 02 222 6 222 |
| Volatile narcotic, 5 Sale of, 867 | DOLVDERALAR 105 | 1 atamagetan, 75, 552, 7, 333, |
| Sale 01, 807 | POLIFEIALAE, 393 | 334 |
| Poisonous Larkspur, 463 | POLYPODIACEAE, 100, 313, 315- | natans, f. 333 Potassic chlorate, 72 |
| Plants among Legumes, | <u>·</u> 321 | Potassic chlorate, 72 |
| 530-535 | Palypadium, 89, 90, 315-316 | cyanide, Poisoning by, 6, |
| Treatises upon 866 | aureum, 316 | 77 |
| Treatises upon, 866 Poke, See Pokeweed | incanum 316 | mitrata 85 |
| Deliahana 424 746 | incanum, 316 | nitrate, 85 Potato, 132, 867 Air, 374 |
| Pokeberry, 434, 746 Pokeweed, f. 434, 434, 435 Family, 433-435, 771 | laciniatum, 825 | Potato, 132, 867 |
| Pokeweed, f. 434, 434, 435 | lignlatum, 825 | Air, 374 |
| Family, 433-435, 771 | percussum, 825 | Common, 714, 718, f. 718, 724-725 |
| Root, 435 | Phymatades, 825 | 724.725 |
| Poland, Poisonous plants of, | scandens, 825 | |
| rotatio, rotsonous plants of, | | Japanese, 710 |
| 867 | suspensum, 825 | Sweet, 698 |
| Palanisia, 495, 497 gravealens, 114, 497 | vulgare, f. 309, 311, 316, 531, 825 Polypody, Common, 311, 315, | Sweet, 698 Wild, 701 |
| gravealens, 114, 497 | 531, 825 | Potato-rot Fungus, f. 205, |
| trachysperma, 114 | Polypody, Common 311 315. | 208 |
| | | |
| Delement Date | 316 | Patentilla darmrica 88 |
| Palemanium bareale, 843 | 316 | Patentilla davurica, 88 |
| Palemanium bareale, 843 caeruleum, 843 | 316 POLYPORACEAE, 99, 234-235, | Patentilla davurica, 88 fruticasa, 88 |
| Palemanium bareale, 843 caeruleum, 843 | POLYPORACEAE, 99, 234-235, 861 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 |
| Palemanium bareale, 843 caeruleum, 843 | POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 |
| Palemanium bareale, 843 caeruleum, 843 Family, 130 flavum, 843 | POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis. 807 |
| Palemanium bareale, 843 caeruleum, 843 Family, 130 flarum, 843 gracile, 843 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 linspidus, 861 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis. 807 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis. 807 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 fauciflarum, 843 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis. 807 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 reptans, 843 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 222 squamnasus, 861 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352.353 t 252 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 reptans, 843 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352.353 t 252 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 fauciflavum, 843 reptans, 843 Poley Oil, 78 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352.353 t 252 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 fauciflarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352.353 t 252 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 222 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 | Patentillo davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352.353 t 252 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 fauciflarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, | POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 357 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 fauciflarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 Palystictus versicalar, 234 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 233 serennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 Palysticma rubrum, 273 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 fauciflarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichus spinnlasum, 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrinchum, 279 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Power, F. W., 560 Prasg. L. van. 902, 906 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavium, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichus spinnlasum, 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrinchum, 279 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Power, F. W., 560 Prasg. L. van. 902, 906 |
| Palemanium bareale, 843 caeruleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 Palythrincium, 279 Palythrincium, 279 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prain, Col., 490 Prain, Col., 490 Prairie Larkspur, 460,461 |
| Palemanium bareale, 843 caeruleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum. 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 trifalii, 100, 861 Palytichum, 310 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prain, Col., 490 Prairie Larkspur, 460,461 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 faucidarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum. 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 trifalii, 100, 861 Palytichum, 310 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prain, Col., 490 Prairie Larkspur, 460,461 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 233 perennis, f. 232 squammasus, 861 Sulphur 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 Palystichum spinnlasum, 273 Palythrinchum, 273 Palythrinchum, 279 trifalii, 100, 861 Palytrichum, 310 cammene, f. 311 POMEAE, 501, 506 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prain, Col., 490 Prairie Larkspur, 460,461 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 faucidarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum. 825 Palystichus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 311 POMEAE, 501, 506 Pomegranate, 638 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prain, Col., 490 Prairie Larkspur, 460,461 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarfaea, 89, 813 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum. 825 Palystichus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 311 POMEAE, 501, 506 Pomegranate, 638 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prain, Col., 490 Prairie Larkspur, 460,461 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 faucidarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarpaea, 89, 813 Palycytliemia, 169 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrincinm, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 311 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prain, Col., 490 Prairie Larkspur, 460,461 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 faucidarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarpaea, 89, 813 Palycytliemia, 169 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum. 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 311 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 hinnata, 63 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis. 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prairic Larkspur, 460-461 Thermopsis, 539 Vetchling, 572 Wake-robin, 385 Prantl, K., 882 Praschil, W. W., 906 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarfaea, 89, 813 Palydesmus, 283 exitiasus, 20, 100, 283-284 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 Sulphureus, 234 Palyscias, 808 nadasa, 808 Palystictus versicalar, 234 Palystictus versicalar, 234 Palystictus versicalar, 234 Palystictus versicalar, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 311 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 pinnata, 63 Pomme Blanche, 558 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prain, Col., 490 Prain, Col., 490 Prairie Larkspur, 460-461 Thermopsis, 539 Vetchling, 572 Wake-robin, 385 Prantl, K., 882 Praschil, W. W., 906 Praid erecta, 812 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 fauciflarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarfaea, 89, 813 Palycythemia, 169 Palydesmus, 283 exitiasus, 20, 100, 283-284 Palygala, 89, 121, 585-586, | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 233 perennis, f. 232 squammasus, 861 Sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 317 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 pinnata, 63 Pomme Blanche, 558 Pomme de Prairie, 558 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prairie Larkspur, 460-461 Thermopsis, 539 Vetchling, 572 Wake-robin, 385 Prantl, K., 882 Praschil, W. W., 906 Pratia erecta, 812 Prasunitz, C., 906 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 fauciflarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarfaea, 89, 813 Palycythemia, 169 Palydesmus, 283 exitiasus, 20, 100, 283-284 Palygala, 89, 121, 585-586, | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 233 perennis, f. 232 squammasus, 861 Sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 317 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 pinnata, 63 Pomme Blanche, 558 Pomme de Prairie, 558 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prairie Larkspur, 460-461 Thermopsis, 539 Vetchling, 572 Wake-robin, 385 Prantl, K., 882 Praschil, W. W., 906 Pratia erecta, 812 Prasunitz, C., 906 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 pauciflarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarpaea, 89, 813 Palycythemia, 169 Palydesmus, 283 exitiasus, 20, 100, 283-284 Palygala, 89, 121, 585-586, 774 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum. 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrinchum, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 317 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 pinnata, 63 Pomme Blanche, 558 Pomme Blanche, 558 Pomme Ge Prairie, 558 Pomd Scum, f. 192 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis. 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 393 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prairie Larkspur, 460-461 Thermopsis, 539 Vetchling, 572 Wake-robin, 385 Prantl, K., 882 Praschil, W. W., 906 Pratia erecta, 812 Prausnitz, C., 906 Precipitates, Red, 6 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 faucidarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarpaea, 89, 813 Palycythemia, 169 Palydesmus, 283 exitiasus, 20, 100, 283-284 Palygala, 89, 121, 585-586, 774 alba, 843 | BOLYPORACEAE, 99, 234-235, 861 Palyparus anthelminticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 232 squammasus, 861 Sulphur, 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 Palystictus versicalar, 234 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 311 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 pinnata, 63 Pomme Blanche, 558 Pomme de Prairie, 558 Pond Scum, f. 192 Pondweed, 332, f. 333 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattisa cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 357 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prairie Larkspur, 460-461 Thermopsis, 539 Vetchling, 572 Wake-robin, 385 Prantl, K., 882 Praschil, W. W., 906 Pretia creeta, 812 Prausnitz, C., 906 Precipitates, Red, 6 White, 6 |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 panciflarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarpaea, 89, 813 Palycythemia, 169 Palydesmus, 283 exitiasus, 20, 100, 283-284 Palygala, 89, 121, 585-586, 774 alba, 843 amara, 843 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 233 perennis, f. 232 squammasus, 861 Sulphur 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum. 825 Palystichum spinnlasum. 825 Palystichum spinnlasum. 273 Palythrinchum, 273 Palythrinchum, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 311 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 pinnata, 63 Pomme Blanche, 558 Pond Scum, f. 192 Pondweed, 332, f. 233 Pangamia Piscida, 832 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 357 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prairie Larkspur, 460-461 Thermopsis, 539 Vetchling, 572 Wake-robin, 385 Prantl, K., 882 Praschil, W. W., 906 Pratia erecta, 812 Prausnitz, C., 906 Precipitates, Red, 6 White, 6 Preiss, 533 |
| Palemanium bareale, 843 caerulcum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarpaea, 89, 813 Palycythemia, 169 Palydesmus, 283 exitiasus, 20, 100, 283-284 Palygala, 89, 121, 585-586, 774 alba, 843 amara, 843 angulata, 843 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 233 perennis, f. 232 squammasus, 861 Sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum, 825 Palystictus versicalar, 234 Palystigma rubrum, 273 Palythrincium, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 317 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 pinnata, 63 Pomme Blanche, 558 Pond Scum, f. 192 Pondweed, 332, f. 333 Pangamia Piscida, 832 Pantederia cardata, 374 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis. 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 353 Powell, R. D., 879, 905 Power, F. R., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Prame, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prairie Larkspur, 460-461 Thermopsis, 539 Vetchling, 572 Wake-robin, 385 Prantl, K., 882 Praschil, W. W., 906 Pratia erecta, 812 Prausnitz, C., 906 Precipitates, Red, 6 White, 6 Preiss, 533 Prenanthes alba, 417, 692, |
| Palemanium bareale, 843 caernleum, 843 Family, 130 flavum, 843 gracile, 843 humile, 843 panciflarum, 843 reptans, 843 Poley Oil, 78 POLEMONIACEAE, 89, 130, 698, 843 Palianthes tuberasa, 64, 386, 835 Pollantin, 767 Pollen as cause of Hay-fever, 767 Polk, 184 Palyalthia argentea, 477 Palyanthus, 386 Palycarpaea, 89, 813 Palycythemia, 169 Palydesmus, 283 exitiasus, 20, 100, 283-284 Palygala, 89, 121, 585-586, 774 alba, 843 amara, 843 | 316 POLYPORACEAE, 99, 234-235, 861 Palyparus anthelminiticus, 861 hispidus, 861 ignarius, f. 233 perennis, f. 233 perennis, f. 232 squammasus, 861 Sulphur 234 sulphureus, 234 Palyscias, 808 nadasa, 808 Palystichum spinnlasum. 825 Palystichum spinnlasum. 825 Palystichum spinnlasum. 273 Palythrinchum, 273 Palythrinchum, 279 trifalii, 100, 861 Palytrichum, 310 cammune, f. 311 POMEAE, 501, 506 Pomegranate, 638 Pametia, 89 pinnata, 63 Pomme Blanche, 558 Pond Scum, f. 192 Pondweed, 332, f. 233 Pangamia Piscida, 832 | Patentilla davurica, 88 fruticasa, 88 Paterium canadense, 847 afficinale, 847 Pattsia cantanensis, 807 Pouchet, G., 895, 905 Poverty Grass, Long-awned, 352-353, f. 353 Short-awned, f. 353 Short-awned, f. 357 Powell, R. D., 879, 905 Power, F. B., 4, 54, 118, 217, 452, 468, 470, 533, 546, 565, 567, 575, 662, 684, 905, 906 Power, F. W., 560 Praag, L. van, 902, 906 Prain, Col., 490 Prairie Larkspur, 460-461 Thermopsis, 539 Vetchling, 572 Wake-robin, 385 Prantl, K., 882 Praschil, W. W., 906 Pratia erecta, 812 Prausnitz, C., 906 Precipitates, Red, 6 White, 6 Preiss, 533 |

altissima, 816
serpentaria, 692
Prentiss, A. N., 510, 906
Prentiss, D. W., 637, 906
Prestott, A. B., 568, 906
Preston, E., 435
Prestania taxifera, 807
Price, T. M., 291, 906
Price, T. W., 665, 896, 906
Price, W., 906
Prickly Ash, 581
Lettuce, 137, 760-761, f. altissima, 816 Furopean, 756
Pear, 635, 636, f. 636
Poppy, 113, 483
Mexican, 113, 483
Prien, O. L., 139, 776
Priestly, 3
Prillieux, E., 880, 906
Primrose, 676, 677
Family, 128, 675-679
Parry's, 128
Primula, 89, 676, f. 676, 677, 867
Auricula, 844 761 Auricula, 844 abcanica, 128, 676, 677, 679, 844 Parryi, 128, 676, 677, f. 678, 844 reticulata, 844 sinensis, 676, 679, 844 vulgaris, 676 PRIMULALES, 157, 157, 675-679
PRIMULACEAE, 52, 89, 128
675-679, 844
Primulin, 676
Prince's Feather, 421
PRINCIPES, 155, 639-370
Pringlea antiscorbutica, 486
Printhogalum, 867
Pritzel, G. A., 906
Privet, 128, 684
Prackia theaeformis, 855
Proctor, 906
Proccupin, G., 906 679 128, Proctor, 906
Prophetin, 749
Praserpinaca palustris, 640
Prosopis, 89
juliflora, 120, 527, 535, 832
ruscifalia, 832
Prostrate Pigweed, 431, f. Pratea synaroides, 844
PROTEACEAE, 415, 844
PROTEALES, 156, 415 PROTEACEAE, 415, 844
PROTEALES, 156, 415
Protein, 10
PROTOASCINEAE, 248-253
PROTOBASIDIOMYCETES, 220, 221-232
Protococus, 92
Protocosin, 505
Protocurin, 148, 687
Protomyces macrasparus, 247
PROTOMYCETACEAE, 247
PROTOMYCETACEAE, 247
Protopin, 90, 113, 481, 485
Protoveratridin, 381
Protoveratridin, 381
Protoveratrin, 381
Protoveratrin, 372
Prouty, 243
Prouty, 243
Prullaurocerasin, 503, 867
Prudden, T. M., 906
Prunes, Hydrocyanic acid in, 88 Prunes, I PRUNEAE, 506
Prunus, f. 66, 252, 506, 514519, 867
amara, 847
amara, 847 americana, 116, 505, f. 506, Amygdalus, 53, 505, 847 Amygdalus v. amara, 503

Amygdalus v. cammunis, 517 Amygdalus v. cammunis, 517
angustifolia, 505
Armeniaca, 505, 514
azium, 505, 514
Besseyi, 116, 505
Capollin, 847
Caraliniana, 116, 847
Cerasus, 253, 505, 514
demissa, 514
demissa, 514
demissa, 515, 517, 847
damcstica, 505, 514, 847
instititia, 506
javanica, 747
Laura-cerasus, 504, 517, 847
Mahaleb, 524, 847
maritima, 253
nana, 505
Padus, 252, 504, 506, f.
519, 519
paniculata, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 503
peundula, 505
516, f. 516
persica, 503, 514, 847
pnmila, 116, 505
serotina, 64, 88, 117, 503,
505, 506, f. 515, 515
516, 518, 515
serotina, 64, 88, 117, 503,
505, 506, f. 515, 515
stiloba, 514
trilloba, 518, 848 midulata, 848 virginiana, 116, 503, 514, 518, 848 Prussic acid. 2. 3, 53, 72, 346, 348, 505, 517, 518, 534, 581 See also Hydrocyanic acid. Psedera heterophylla, 620 quinquefalia, 124, 858 rricuspidata, 620 Pseudoaconitin, 109, 450 Pseudoconydrin, 651 Pseudocumarin, 524 Pseudohyssovanin, 148, 7 Pseudohyoscyamin, 148, 715, 726 Pseudojervin, 148, 381 Pseudomanas campestris, 163 pyocyanea, f. 161 syncyanea, f. 161 Pseudomorphin, 481 Pseudo-narcissin, 386 Pseudosmodingium pernicio-sum, 805 sum, 805 Pseudatsuga Douglašii, 327 Psidium Guajava, 639 montanum, 53, 839 Psaralea, 535, 557-558 argophylla, 118, 558 cuspidata, 558 esculenta, 118, 558 elandulosa, 629, 832 esculenta, 118, 558
glandulosa, 629, 832
hypogaea, 558
macrostachya, 535, 832
pentaphylla, 832
Silvery, 118
Silver-leaf, 558
Slender, 118
tenuiflara, 118, 535, 832
Psychatria emotica, 849
Ipecacuanha, 742, 849
Ptaeraxylon obliquum, 837
ntile, 608 Ptaeraxylon obliquum, 837 ntile, 608 Ptelea trifoliata, 581, 849 PTERIDOPHYTA, 100-101, 155, 312, 325 Pterigeron adscendens, 816 Pteris, 315, 317 aquilina, 100, 315, f. 317, 317-318, 825 candata, 825 servilata, f. 314 serrulata, f. 314

Pteracarpus Dalbergioides, 523 esculentus, 832 Marsupium, 523, 832 tinctorius, 523 pycnostachyum, Pteracaulan 816 Pteraspara andramedca, 838 Pteraspermum diversifolium, 855 855
Ptomains, 10, 73, 144, 149, 164, 867
Ptosis, 77, 174
Ptyalism, 25
Puccinia, 224-230, 776
caranata, 99, f. 223, 225, 228, 821
coronata I, 229
caranata II, 229
caranata II, 229
caranata detablidic, 229 caronata dactylidis, 229 coronifera, 229 dispersa, 229 arspersa, 229 emaculata, 223, f. 223 glumarum, 226, 227-229 graminis, 99, f. 223, 223-227, f. 225, 228, f. 228, 231, 861 1. 225, 228, 1. 228. 231, 861
rubiga-vera, 226, 227, f. 227, 228
Sarghi, 229, 230, f. 230
xylorrhizac, 138, 861
PUCCINIACEAE, 99, 222-230
Biology, 222-224
Puccoon, Common, 468
Pueraria Thunbergiana, 526
Puff-ball, 160, 220, f. 246
Giant, 99
Lead-color, f. 246
Puihn, J. G., 906
Pulcgium, 78
Pulicaria dysinterica, 816
Pulsatilla, 446, 455, 460
xulgaris, 4
Pulse Family, 117-120, 501, 564
Pumcia ccid, 679 Pulse Family, 117-120, 501, 564
Pumic acid, 679
Pumpkin, 750
Punica granatum, 638, 837
PUNICACEAE, 638
Purgative, 804
Purging Cassia, 529, f. 532
Purins, 143, 144, 416, 147
Purple Jimson Wecd, 133, 729, 732-733
Larkspur, 44, 84, f. 461, 462-463, 467
Western, 109
Loco Weed, 563, 567-570, f. 568
Magnolia, 474
Ragwort, 756
Thorn Apple, 729-733
Purple-stemmed Angelica, 126 126 Poison Hemlock, f. 665 Purshia tridentata, 848 Purshia tridentata, 848
Purshianin, 621
Pus Organism, f. 168, 171
Pussley, Garden, 423
Putranjiera Raxburghii, 823
Putrescin, 10
"Putrid sore throat," 21
Puy, C. E. de, 880
Pyemia, 167, 171
Pygeum africanum, 53, 848
latifalium, 54
barvillorum, 54, 848 parviflorum, 54, 848 Pyocyanin, 10 PYRENOMYCETES, 281 PYRENOMYCETINEAE, 266-281 Pyrethrin, 754

| Pyridin, 145, 146, 728 |
|--|
| Pyridin, 145, 140, 726 |
| Pyrola chlorantha, 842 |
| elliptica, 842 |
| minor, 842 |
| rotundifolia, 842 |
| Pyrophosphoric acid, 124 |
| Pyrogallic acid, 608 |
| Pyronema confluens, Fertili |
| zation of, 1. 247 |
| Pyrus, 53, 506, 512, 848 |
| 1 y/ 113, 33, 300, 312, 040 |
| americana, 504, 512 |
| angustifolia, 512 |
| Aria, 503 |
| Aucuparia, 115, 504, 506 |
| 512, 629, 848 |
| baccata, 512 |
| cammunis, 115, 504, 512 |
| covanaria, 504, 512 |
| Cydonia, 503, 504, 512 ioensis, 512, f. 513 japonica, 503, 504 |
| ioensis 512 f. str |
| intensica 503 504 |
| lanata, 848 |
| |
| Malus, 116, 504, 505, f |
| 512, 512 |
| pinnatifida, 503 |
| rivularis, 512 |
| sambucifolia, 512 |
| sinensis, 512 |
| sitchensis, 512 |
| Sorbus, 848 |
| PYTHIACEAE, 204 |
| Pythium DeBaryannm, 204 |
| proliferum, 204 |
| Protifer and, 204 |
| |

Q

Quack Grass, 103, 364-365, f.

365
Black-spot on, 280, f. 280
Ergot on, 276
Quain, E., 906
Quaking Grass, 90
Quamsa, 378
Quarter evil, 175
Quassia, 583
Quebrachia Balansae, 608
Lorentzii, 608
Quebrachin, 692
Quebrachin, 692
Quebrachin, 692
Quebrachin, 692
Quebrachin, 692
Queen's Delight, 604
Root, 604
Queensland Hemp, 624
Quelt, 34
Queratrin, 560
Quercitrin, 506, 510, 584, 666
Quercus, 105, 402, 629, 866, 867
alba, 403, 824
chrysolepis, 105
coccinea, 403
Ilex, 403
Ilex, 403
Ilex, 403
Ileximica, 403, 824
Mullenbergii, 403
ralustris, 403
Rabur, 403
rubra, 105, 403, f. 804
Taza, 824
virginiano, 403
Quillagic acid, Sce Quillaic
acid
Quillaic acid, 7, 62, 505, 586
Quillaja-sapotoxin, 62
Quinamin, 742
Quincke, 504
Seeds, 503
Quincke, H, 295
Quincie acid, 144

Quinolin, 145, 146, 147 Quinin, 77, 78, 87, 145, 147, 741, 742 Quinoa, 427 Quisqualis indica, 814

Rabak, Frank 88
Rabbit Septicemia, 177
Rabinowitch, Lydia, 249
Rabuteau, 904
Radicula, 490, 491
Armoracia, 113, 486, 490491, 818
Nosturtium-aquaticum, 486
Rabletic v. histoida, 490 palustris v. hispida, 490 Radish, 486 Radikofer, L., 52, 803, 804, Rafinesque, C. S., 449, 906 Ragged Robin, 439 Raggi, 198 Raggi, 198
Ragweed, 137, 755, 764-767,

f. 765
False, f. 764
Great, f. 136, 137, 765-766
Pollen as cause of Hay
Fever, 137, 767
Small, f. 766, 766-767
Tall, f. 766
Ragwort, 795
Purple, 756
Raillet, 15
Raleigh, Sir W., 714 Rarllet, 15
Raleigh, Sir W., 714
Rambusch, W. T., 907
Ramie-grass Cloth, 406
Ramsdell, William, 71
RANALES, 156, 444-479
Randall, Dr., 277
Rondio, 89 aculeata, 849 dumetorum, 849 Ransom, 62 Ransom, W. B., 907 RANUNCULACEAE, 53, 83, 89, 108-111, 417, 444, 446-468, 844-846 468, 844-846
Ronunculus, 81, 110-111, 447, 448, 457-461, 867.
obortivus, 111, 458-459, f. 459, 460
ocris, 4, 77, 78, 110, 457, 458, f. 459, 460, 846
alpestris, 4, 846
aquatilis, 846
aquatilis, v. capillaceus, 446
arvensis, 460, 846
osiaticus, 846
auriconus, 846
auriconus, 846 auricomus, 846 bulbosus, 4, 111, 457, 459, biloosus, 4, 111, 457, 459, 846
fascicularis, 110
Ficaria, 446, 846
Flaninula, 460, 846
luybridus, 846
lanuginosus, 846
lappaceus, 846
leptosepala, 108
lingua, 846
Oil of, 459
polyanthemas, 846
repens, 4, 457
scelerotus, 4, 77, 78, 111, 459-458, f. 458
septentrionalis, 110, 457, 846
Thora, 4, 846
Rape, 486, 867
Fungus, 283-284
Raphanus, 867
raphanistrum, 277, 818
sativus, 486, 818 846

Raphia Fiber, 370 Raphia Fiber, 370
pedunculata, 370
vinifera, 370
Raspberry, 507-508
Black, 505, 508
Black-cap, 508
Wild, f. 507, 508
Garden, 505
Red, Wild, 505, 507, f. 507 Rat Poison, Ratsbane, 575 Ratti, 907 Ratti, 907 Rattle-weed, 867 Rattlebox, 118, 543-546, f. Rattlebox, 118, 543-543, 564 Rattlesnake Fern, 692 Grass, 692 Master, 692 Plantain, 692 Plantain, 692 Rattlesnake-root, 692, 755 Rattlesnake-wced, 692, 755 Ratzeburg, J. T. C., 4, 803, 862, 874, 907 Raum, J., 249 Rauwerda, 90 Rauwolfia serpentina, 807 trifoliata, 807 verticillata, 807 Ravenalo, 391
Ravenel, M. P., 23, 30, 263
Ravenna, 747 Ray, 290
Rawton, Oliver de, 907
Read, Alex, 907
Red Baneberry, 108, 467-468
Buckeye, 123, 617, f. 619
Cedar, 101, 102, 330-331, f.
331, 332
Clover, 279, 525, 553, f.
555
Currant, 498
Flax, Garden, 580
Gum, 500, f. 501
Haw, 500, f. 501
Haw, 500, f. 514
Maple, 615
Oak, 403, f. 404
Osier, 664
Pepper, 87, 133, 714, f.
725, 725-726
Poppy, 479
Raspberry, Wild, 505, 507, f. Ray, 290 Raspberry, Wild, 505, 507, 507
Rust, 227
Seaweed, f. 194
Top, 338
Ergot on, 277
Vetch, 572
Red-berried Elder, 135
Red-bot, 104, 385, 432
Redwood, 102
Reed, II. S., 907
Rees, 302
Reese, J. J., 907
Reinder Lichens, 307
Reinders, 621
Remak, 13, 18, 295
Remijia pedunculata, 742
Remsen, Dr., 612
Renner, Dr., 575, 907
Renon, L., 263
Reproduction of Bacteria Reproduction of Bacteria, 161 Reseda, 846 luteola, 479, 846 adorata, 479 RESEDACEAE, 479, 486, 846 Resin, 319, 330 Botany Bay, 375 Chaia, 625 Piney, 627 Retamin, 531

Reynolds, C. F., 775 RIIAMNALES, 157, 620-621, RIIAMNEAE, 52 RHAMNACEAE, 53, 89, 620-621, 846-847 Eich poissons among 2, 621 Fish poisons among, 2, 621 Rhamnetin, 124, 620 Rhamnin, 124, 620 Rhamno-cathartin, 124 Rhamno-cathartin, Rhamnose, 612 Rhamnus Aloternus, 847 colifornica, 847 caroliniono, 847 cathartica, 123, 124, 229, cathartica, 123, 124, 229, 620, 847, 630, 53, 124, 229, 620, 847, 640, 620 infectoria, 620
lonceoloto, 124, 229
Purshiana, 621, 847
tinctoria, 620
Wightii, 847
Rhein, 418
Rheotannic acid, 419
Kheum Emodi, 843
hybridum, 843
officinole, 418, 843
palmatum, 843
Rhaponticum, 418, 843
Rhaponticum, 418, 843
Rhiacanthus, 90
communis, 692, 804
Rhinacanthus, 90
communis, 692, 805
Rhipsolis, 637, 811
Rhizobium leguminosarum, f.
162, f. 163
Rhizocarpon geographicum, f.
308
Rhizoids of Botrydium granulotum, f. 92
Rhizonucor, 202
Rhizophora Mangle, 638, 847
Rhizophora Mangle, 638, 847
Rhizopus, 198, f. 258, 867
nigricons, 195, 198
Rhode Island, Poisonous
plants of, 867
Rhodium, 501
Rhodium, 601
Rhodium, 601
Rhodium, 601 Rhodinol, 506 Rhodium, Oil of, 699 Rhodium, Oil of, 699
Rhododendrol, 666
Rhododendron, 52, 64, 222, 666, 667, 669, 867
arborescens, 821
Azolea, 667
borbotum, 55
California, 127, 667
californicum, 127, 667, 821
camponulatum, 821
cutawbiense, 83, 127, 667, 668, 821
coucosicum, 821 concosicum, 821 concostcum, 821 chrysonthum, 85, 666, 821 cinneborinum, 821 douricum, 821 ferruginenm, 821 fulgens, 65 grande, 65 hirsutum, 821 indicum, 821 inateum, 821 japonicum, 666 Large, 127 ledifolium, 821 maximum, 127, f. 666, 667, 668-669, 821 occidentale, 127, f. 668, 668-669, 821 bonticum, 64, 668, 821 ponticum, 64, 668, 821 punctatum, 821 sinense, 821

sublanceolotum, 821 Sublanceolotum, 821
RHODOM ELACEAE, 859
Rhodomenia palmota, 859
RHODOFHYCEAE, 154, 194
RHODOSPOREAE, 235
RHODEADALES, 157, 479-497
Rhoeadin, 481
Rhodotypos herricides, 848
Rhubarb, 418, 419
Rhus, 79, 409, 608-614, 805-806, 865, 867
caustica, 806 caustica, 806 caustica, 806 chinensis, 806 Coriaria, 608 Cotinus, 608, 805 diversilobo, 123, 609, f. 611, 806 foridano, 806 glabra, 608, 613-614, 805 Griffilhii, 806 javanica, 806 littoralis, 806 lucida, 806 Metopium, 806 Michauxii, 806 Michauxii, 806
pernicioso, 806
rodicons, 806
Rydbergi, 123, 806
semialota, 608
striota, 806
succedanea, 608, 806
sylvestris, 806
Toxicodendron, 4, f. 12, 77, 78, 122, 394, 608-609, f. 609
venevada, 610, 806 609
venenata, 610, 806
vernicifera, 608, 806
vernici, 122, 609, f. 610,
806
Wallichii, 806
Rhynchosia minima, 832
Ribes aureum, 53, 500, 851
cereum, 851
gracile, 500
grossularia, 500
macrobotrys, 851 grossularia, 500
mocrobotrys, 851
nigrum, 498, 851
prostrotum, 851
vulgare, 498
Ribhert, M. II. W., 907
Rice, W. S., 907
Rice, Wild, 276, 338
Rice-cut Grass, 339
Rich, F. A., 101, 334, 325, 907
Richardia africana, 371 Rich, F. A., 101, 334, 325, 907

*Richardia africana, 371
Richardson, 907
Richet, C., 907
Ricin, 6-7, 11, 55, 72, 121, 594, 626, 687, 867

*Ricinodendron africanus, 587
Ricinoleic acid glycerid, 594
Ricinolein, 594
Ricinous, 55, 590, 594-595, 862, 864, 867

*Communis, 53, f. 56, 75, 121, 594-595, 823

*Rickets, 325
Ricketts, H. T., 306
Ricord-Madianna, J. B., 907
Riddinger, 467
Riceke, V. A., 862, 872, 847, 907
Ringworm, 12, 14, 301-302 Ringworm, 12, 14, 301-302 of Body, 301 Dog, 12 Horse, 14 Also sec Tinea Scalp, 301 Ritchie, J., 305 Rivina humilis, 842 Rivolta, S., 263 Rivularia, 93

RIVULARIACEAE, 186-187 Robbins, E. J., 424 Robbins, E. L., 899, 907 Roberts, G. H., 260 Robin, Ch., 298, 302, 595, 907 Robinia, 534, 535, 558-561, hispida, 559 neo-mexicana, 118, 559 neo-mexicana, 118, 559
Nicou, 531
Psend-ococio, f. 57, 64, f.
66, 118-119, 527, f. 559, 559, 559, 559, 560, 561, 832
ricoso, 559, 560-561, 832
Robinin, 119, 560
Robinson, B. L., 18, 488, 863, 886, 907
Robiquet, 505
Roccella tinctoria, 307
Roccellacella, 307
Roccellacella, 307 ROCCELLACEAE, 307
Rochea coccinea, 501
Rochebrine, A. T. de, 907
Rochussen, F., 755, 907
Rock Elm, 408
Rose, 627
Rocket Larkspur, 460
Rocky Mountain Bee Plant, 496, f. 496
Bush Honeysuckle, 135
Columbine, 446
Rodigras. E.m., 908
Rodin, H., 908
Roemeria violocea, 841
Rolly, 908
Roman Chamomile, 754 Roman Chamomile, 754 Root, Indian Cucumber, 104 Pellitory, 754 Red, 104 Root-tubercle Organism, f. 162 162 Roots, Poisonous, 867 Roretz, A. von, 908 Roripa hispida, 490 Roso, 504, 506, 510-512 arkansano, 115 blanda, 510 conina, 504, 510, 629, 848 centifolia, 510 centifolia, 510
cinnamomea, 504
damasceno, 503
gallica, 504, 510, 848
pratincola, 510, f. 511
rubiginoso, 115, 504
rngosa, 504
Rosaceae, 53, 63, 89, 90, 115117, 144, 498, 501, 503,
519, 847-848, 867
ROSALES, 157, 498-574
Rose, 629 ROSALES, 157, 498-5.
Rose, 629
Apple, 639
Arkansas, 115, f. 511
Attar of, 699
Bay, 667 Bay, 667 Mountain, 127, 667-668 Cinnamon, 504 Cinnamon, 504
Dog, 504
Family, 503-519
Oil, 503, 506
Pink, 689
Prairie, 504, 510
Rock, 627
Say's, 511
Smooth, 510
Sweetbriar, 504
Wood's, 511, 512
Rosebay, 667
Mountain, 127, 667-668
Rosemal, 450
Rosemary, 709 Rosemary, 709 Bog, 127 Oil of, 86

| Wild, 127, 673 | Ruscus aculcatus, 835 | Sacred Bean, 444 |
|---|--|---|
| Wild, 127, 673 Rosenthal, D. A., 908 | Rush, 374 | Lily, 444 |
| Rosmarinus othernalis, 709 | Common, 367, f. 373 | Safflower, 756 |
| Roseol, 506 Rossi, P., 908 Rostafinski, 158 Rostrup, E., 290 | Common, 367, f. 373 Scouring, 333 | Safflower, 756 Safford, W. E., 325-326, 908 Saffron, f. 387, 389, 734, 756- Cape, 734 Mendow, 148 |
| Rossi, P., 908 | Russelia juncea, 734 | Saffron, f. 387, 389, 734, 756 |
| Rostafinski, 158 | Russell, Dr., 775 | Cape, 734 |
| Rostrup, E., 290 | Russell, W., 908 | Meadow, 148 |
| Rot, Cabbage, 163 | Russella funcea, 734 Russell, Dr., 775 Russell, W., 908 Russia, Poisonous plants of, | Meadow, 148 South American, 389 Sagaretia theezans, 629 |
| Rotenbiller, von. 908 | 867 | Sagaretia theezaus, 629 |
| Rothe I 908 | Russian Licorice, 527 | Sage, 710 |
| Rotenbiller, von, 908 Rothe, L., 908 Rotschky, 728 | Olenster 641 | Lance-leaved, 710 |
| Pottlerin 590 | Oleaster, 641 Thistle, 68, 150, 424, 430, | South American, 710 |
| Rottlerin, 589 Roub, J. F., 908 | | |
| D Dh 1: 121 | f. 431, 433 | White, 424 |
| Rough Phacelia, 131 | Turpentine, 330 Russula, 90, 243 | Sage brush, 790, 792, f. 791 |
| Round Cardamon, 391 | Russula, 90, 243 | Saghalen Knotweed, 421 |
| Round-leaved Dogwood, 664 | Barlea, 860 | Sagittaria Engelmanniana, |
| Green Brier, 104 | enietica, 237, 243, 860 | 335, f. 336 latifolia, 102 |
| Roup, 177 Roupala Pohlii, 844 | 10etens, 53, 860 | 101110110, 102 |
| Rongala Politit, 844 | enietica, 237, 243, 860 foetens, 53, 860 fragilis, 860 | Sago, 370 Poison, 378 |
| vervaineana, 844 | rubra, 860 Rusts, 20, 160, 220 | Poison, 378 |
| Rourea glabra, 817 Roux, G., 302, 897, 908 Row, 199 Royal Fern, f. 309, 313, 322 | Rusts, 20, 160, 220 | Wild, 378 |
| Roux, G., 302, 897, 908 | Black-stem, 227 Clover, 20, 99, 230-231, f. | Sagot, M. P., 908 |
| Row, 199 | Clover, 20, 99, 230-231, f. | St. Hilaire, 908 |
| Royal Fern, f. 309, 313, 322 | 231 | St. Ignatius's Bark, 688 |
| 1 alm, 370 | Corn. f. 230 | Folson, 378 Wild, 378 Sagot, M. P., 908 St. Hilaire, 908 St. Ignatius's Bark, 688 Bean, 74, 145 St. John's-bread, 1, 526, 527 St. John's-wort, 124, 386 |
| Roystonea borinquena, 370 | Crowned, f. 223, f. 225, f. | St. John's-bread, f. 526, 527 |
| Rozites gongylophora, 235 | 228, 228, 229 | St. John's-wort, 124, 386, |
| Rubber, Para, 588 | Goldenrod, 222, 775 | 629-631 |
| Plants, 587, 691, 696 | Crowned, f. 223, f. 225, f. 228, 228, 229 Goldenrod, 222, 775 Grass, f. 223, 224-227, f. | Common, 125, 629-630, f. |
| Rubber, Para, 588 Plants, 587, 691, 696 Tree, f. 406 | 7. 223, 224-221, 7. 225 Maize, 229-230 Oat, 99 Red 222 | 630 |
| Rubia tinctorum, 742 Rubiaceae, 53, 89, 90, 135, 144, 741-744, 848-849 RUBIALES, 158, 740-748 | Maize, 229-230 | European, 631 Family, 629-631 Great, 125, 629, 630-631, f. 631 Kalm's, 629 |
| RUBIACEAE, 53, 89, 90, 135. | Oat. 99 | Family, 629-631 |
| 144 741.744 848.849 | Red, 222 | Great, 125, 629, 630,631 |
| RUBIALES 158 740-748 | Reproduction, 222 | 1 621 |
| Rubijervin, 381 | | Kalm's 620 |
| | Strawberry, 281 | Shrubbar 620 |
| Rubus, 507-508 | Teosinte, 230 | Shrubby, 629 Spotted, 630 |
| chamaemorus, 848 cuneifolius, 505, 848 | Tickle Grass, f. 223 Wheat, 99, 227, f. 228 Covered, 227, f. 227, 228 | Spotted, 630 |
| cuneijoiius, 305, 848 | Wheat, 99, 227, 1. 228 | Sakarava, 86/ |
| idaens, 505 idaens v. aculeatissimus, 505, f. 507 occidentalis, 505, f. 507, 508 | Covered, 221, J. 227, 228 | Sakalava, 867 "Sake," 261 Salep, 392 |
| idaeus v. aculeatissimus, | W nite, 7. 204 | Salep, 392 |
| 505, 1. 507 | Ruta graveolens, 76, 120, 581, | SALICACEAE, 105, 396-397, 849 |
| occidentalis, 505, f. 507, 508 | 849 | SALICALIES, 156, 396-399 |
| 040/4143, 304 | montana, 849 | SALICACEAE, 105, 396-397, 849 SALICALES, 156, 396-399 Salicin, 397, 505 |
| parviflorus, 505 | Rutabaga, 486 RUTACEAE, 53, 89, 120, 148, 575, 577, 581-583, 849 | Salicornia, 150 |
| phoenicolasius, 505 | RUTACEAE, 53, 89, 120, 148, | Salicylic acid, 505 |
| Prickles and spines of, 507- | 575, 577, 581-583, 849 | Salikounda, 967 |
| 508 | Rutin, 495 | Salix, 629 |
| villosus, 508. f. 508, 848 | Rye, 338 | amygdaloides, f. 397 Salmon, D. E., 277 Salmon Berry, 505 |
| Rudbeckia, 757, 779 | Grass, 361-364 | Salmon, D. E., 277 |
| hirta, 779 | | Salmon Berry, 505 |
| laciniata, 139, 756, 779 | Common, 361 Italian, 361 | Salpingitis, 171 |
| occidentalis, 779 | Smut, f. 218 Wild, 27, 90, 149, 546 | Salsify 756 |
| Rüdel, C., 908 Rue, 78, 524 | Wild, 27, 90, 149, 546 | Salsify, 756 Salsola, 426, 430 Kali v. tenuifolia, 424, 430; |
| Rue, 78, 524 | Black-spot on, 280 | Kali v tavuitalia A2A A30 |
| Common, 581 | Ergot on, 276 | 1 425 014 424, 4305 |
| Family, 120, 581-583 | Rhynchospora, 369 | f. 431, 814 |
| Meadow, 111 | Rynning 662 | tamariscifolia, 814 Webbii, 424, 814 |
| Ruedi, Dr. Carl, 565 | Rynning, 662 Ryon, A. M., 908 | C-14 D -1 107 405 |
| Ruellia ciliosa, 688, 698 | Ryparosa caesia, 53 | Salt Bush, 107, 425 |
| patula, 804 | , | Australian, 424 |
| strehens 804 | · s | Grass, 90 |
| `tuberosa, 804 | | Groundsel, 138 |
| Rumex, 419, 420-421, 867 | Sabadilla officinalis, 377 | Salts, Caustic, 72 |
| abyssinicus, 843 | Sabadillin, 381 | Corrosive, 72 |
| Acetosa, 150, 846 | Sabadin, 379, 381 | Saltwort, 430 |
| Acetosella, 106, 420, f. 420, | Sabadinin, 379, 381 | Shrubby, 424 |
| 846 | | Salvia, 756 |
| altissimus, 419, 420-421 | SABALACEAE, 849 | amarissima, 828 |
| cristus, 106, 419, f 420 | Sabanijeff, N., 872, 908 | coccinea, 710 |
| crispus, 106, 419, f. 420, 421, 846 | Sabatia angularis, 689 | lanceolata, 710 |
| Ecklonianus, 846 | Sabina, 78 | officinalis, 710, 828 |
| hymenosepalus, 418, 846 | Sabinea florida, 833 | pratensis, 828 |
| | Sahli, 241 | splendens, 710 |
| obiusifolius, 846 | Sahourand, 13, 16, 301, 908 | Salvadora persica, 849 |
| orbiculatus, 421 | Sabourne, 300 Saccardo, P. A., 288 | SALVADORACEAE, 849 |
| Patientia, 419 scutatus, 419 | Saccardo, P. A., 288 | Salvinia, 315 |
| D | Saccharomyces, 100, 249 apiculatus, 250 cerevisiae, 250. f. 251 | Salviol, 710, 756 |
| Rumicin, 421 | apiculatus, 250 | |
| Runge, 145 | cerevisiae, 250. f. 251 | Samadera indica, 853 |
| Rusby, H. H., 96, 101, 105, | ellipsoideus, 250, f. 521 | Sambucin, 747 |
| 114, 121, 389, 448, 456, | glutinus, 250 | Sambucus, 745-747, 867 canadensis, 133, 744, 745- |
| 471, 484, 491, 560, 584, | kefyr, 250 | canaaensis, 133, /44, 745- |
| Runge, 145 Rusby, H. H., 96, 101, 105, 114, 121, 389, 448, 456, 471, 484, 491, 560, 584, 617, 633, 746, 793, 803, | mycoderma, f. 249, 250 SACCHAROMYCETES, 249- | 747, 812 Ebulus, 746, 747, 812 |
| 904 000 | | |
| 804, 908 | SACCHAROMYCETES, 249- | Edulus, 740, 747, 812 |
| Ruschhaupt, F. R., 908 | SACCHAROMYCETES, 249- 253 | mexicana, 812 |

nigra, 53, 746, 747, 813 racemosa, 135, 744, 747, 813 Sambunigrin, 747 Saprolegnia, 209, 859 monoica, 209 Thureti, f. 208 498, 503, 552, 570, 580, 581, 584, 593, 620, 632, 702, 710, 909 Samolus, 2 SAPROLEGNIACEAE, 204, 208-Schauenstein, 688
Scheele, 3
Schell, Dr., 661
Schenck, K., 909, 912
Schimper, A. F. W., 909, 912
Schimpfky, R., 96, 453, 642, 718, 721, 727, 909
Schinus Molle, 608
Schizaea pusilla, f. 309, 315
Schizanthus pinnatus, 715
SCHIZEACEAE, 315
SCHIZOMYCETES, 97, 153, 160, 161-184, 859
Schizonella, 210
SCHIZOPHYCEAE, 97, 153, 184188 Schauenstein, 688 Samolus, 2
SAMYDACEAE, 53, 850
Sand Cherry, 116, 505
Flower, 109, 454
Sand-bur, 67, f. 68, f. 69, f.
349, 352, 722
Smut on, f. 218
Sand-box Tree, f. 588
Sandalwood, 415
Sandarac Tree, 328
Gum, 328
Wood, 328
Sand Flower, 109, 454 Sapucaya Nut, 638 Sarcobatus, 426, 429-430 Maximiliani, f. 429, 430, 209 814 vermiculatus, 68, 107
Sarcolobus carinatus, 809
narcoticus, 809
Sarcopetalum Harveyanum, 838 Sarcoptes, 294 equi, 293 scabei, 293 Sarcostemma wood, 328 Sand Flower, 109, 454 Sandoricum indicum, 837 Sandwort, European, 436 Sanfelice, F., 249, 304 Sanguinaria, 480, 483-485, 936 australe, 696, 810 glaucum, 810
Sarepta Mustard, 490
Sargasso Weed, 194
Sargassum bacciferum, 194
Sargent, C. S., 863
Sarracenia, 497 188 Biology of, 184 SCIHIZOPHYTA, 160-188, 859 867 canadensis, 113, 479, f. 484, 484-485, 841 Sanguinarin, 113, 480, 484, 97, 153, Schlagdenhauffen, Fr., 909 Schlegel, 26
Schleichera, 850
trijuga, 53, 606
Schleisinger, 36, 242
Schlitzberger, S., 909
Schlotterbeck, J. D., 90, 113,
483, 909
Schmelz, J. D., 909
Schmidt, E., 909
Schmidt, E., 909
Schmidt, E., 601
Schneidt, George, 548,
909 485 Sarracenia, 497
flava, 851
laciniata, f. 498
purpurea, 114, 497, 851
Trap of, f. 499
variolaris, f. 498, 851
SARRACENIACEAE, 114, 497, Sanguinarina, 113 Sanguisorba canadensis, 116 Sanicula marilandica, 692, 857 Sausevieria thyrsiflora, 827 SANTALACEAE, 415, 850 SANTALALES, 156, SARRACENIACEAE, 114, 497, 851 SARRACENIALES, 157, 497-498 Sarracenin, 114, 497 Sarsa-saponin, 62 Sarsaparilla, 647 Indian 695 Santalum album, 415 Santonica, 754 Santonin, 140, 149, 754, 791, 867 Sarsa-saponin, 62
Sarsaparilla, 647
Indian, 695
Saponin, 62
Sarson, 490
Sassafras, 112, 478-479
officinale, 112
Oil of, 86
variifolium, f. 478, 479
Satureia hortensis, 710
Saunders, C. F., 908
SAURURACERE, 396, 851
Saunsage, poisoning by, 77, 78
Saussurea Lappa, 755
Savanna Flower, 867
Savin, 6, 49, 72, f. 331, 332
American, 102
Oil of, 149
Savory, 710
Savory, 710
Savory, 757
Sayes, L. E., 468, 564, 565, 567, 751, 803, 863, 909
Saxifraga, 115
Andrewsii, 852
cortusaefolia, 852
cuneifolia, 852
Sibthorpii, 852
Saxifragacaece, 53, 89, 90, 115, 498, 851-852 867
Poisoning by, 77, 81
Sap Green, 620
SAPINDACEAE, 52, 63, 64, 89, 123, 604, 606-607, 850
SAPINDALES, 157, 604-621
Sapindus, 52, 89, 123
abyssinicus, 850
arborescens, 850
emarginatus, 65 909 909
Schneider, 505
Schneider, A., 909
Schneider, Max, 909
Schniegans, M., 909
Schoenlein, J. L., 17, 295
Schoenocaulon officinale, 835
Schoenus apogon, 819
Schorlemmer, C., 910
Schorlemmer, C41
Schoren, 241
Schramek, 895, 910
Schreber, 721
Schrieber, 721
Schribaux, F., 910 emarginatus. 65 marginatus. 123, 606, 850 Mukorossi, 850 Saponaria, 63, 606, 850 sapotoxin, 62 trifoliatus, 63, 850 Schribaux, E., 910 Schroeter, 96, 325, 439, 448, Sapinm indicum, 823 insigne, 823 sebiferum, 587 Sapodilla, 681 Family, 128, 679-681 Plum, 679 Tree, 681 460 Schuchardt, 572 Schuetz, K. E., 910 Schulze, J. E. F., 910 Schumell, T. E., 910 Schütte, H. W., 910 Schutz, Josef, 910 Schwarz, A. 910 Scilla. 78, 377 Sapogenin, 108, 586
Saponaria, 89, 436, 441-442
officinalis, 62, 63, 77, 108,
f, 436, 441, 442, f, 442,
812 Scilla, 78, 377 bifolia, 835 festalis, 835 Scillain, 148, 377 rubra, 63
Vaccaria, 442, f. 443, 812
Saponin, 4, 62-63, 72, 89, 104,
107, 108, 110, 111, 115,
117, 120, 130, 133, 135,
149, 315, 371, 375, 434,
435, 436, 437, 439, 441,
442, 447, 470, 500, 503,
505, 535, 567, 586, 606,
617, 621, 624, 637, 683,
698, 750, 774, 804, 867
Distribution in plants, 89
Group, 62, 149
Saponin-senegin, 62
Saponin-senegin, 62
Saponin-senegin, 62
Saponin-senegin, 62
Saponin-senegin, 62
Saponin-senegin, 63
Sapotin, 681
Sapotin, 681
Sapotin, 7, 437, 441, 505, rubra, 63 SAXIFRAGCEAR, 53, 89, 90, 115, 498, 851-852
Saxifrage Family, 89
Saxony, Poisonous plants of, 867 Scillin, 377 Scillipicrin, 377 Scabiosa succisa, 65, 820 Scabious, 741 Scaevola, 825 Scillitin, 377 Scillotoxin, 377 Scindapsus officinalis, 808 699 Scammonin, 699 Scammony, 699 Scirpus, 369 lacustris, 369 SCITAMINEAE, 155, 390-Scarlet Fever, 867 Scarlet Fever, 867
Gaura, 645
Lychnis, 439
Oak, 403
Runner, 119, 520, 528
Vetch, 526
Scenedesmus, 190
Schaffner, J. H., 96, 101, 104, 108, 112, 115, 118, 120, 126, 130, 139, 439, 471, 392, 852 Scleranthus perennis, 436 Sclerocarya Caffra, 806 Scleroderma vulgare, 247, 860 Sclerospora graminicola, 205, Sclerotinia, 253 Sclerotium clavus, 275

INDEX

| Scolecotrichum graminis, 282, | Cockle, Corn, 439-441 | Jacobaca, 140, 795, 797, |
|--|--|--|
| f 282 | Cow, 108 | 816 fft |
| Scopalamin, 60, 77, 78, 148, 716, 732 | Colchicum, 867 Corozo Palm, 370 | latifolius, 795 plattensis, 140 |
| Scoraina airopoiaes, 4 | Corypha Palm, 370 | toluccanus, 140 |
| Scopola, 60, 716 Scopolia carnicola, 854 | Cotton, 867 Crab's-eye, 527 | vulgaris, 816 vulneraria, 816 |
| japonica, 854 lurida, 854 | Croton, 149 | SENECIONEAE, 757 SENECIONIDEAE, 758 |
| physaloides, 854 | Cummin, 648 Cynoglossum, 867 | Senegin. 149. 586 |
| Scopolin, 716 | Datura, 729 Entada, 525 | Senegin, 149, 586 Senf, E., 395 Senn, Nicholas, 183 |
| Scorzonera, 757 Scotch Broom, 118, 408, 551 | Entada, 525 Eriobotrya, 867 | Senna, 536 |
| Heather, 65, 665 | Fennel, 648 | Leaves, 528 |
| Scatium acuminatum, 860 | Fenugreek, 527 Flax, 120, 581 | Substitute for, 534 Wild, f. 527 |
| Scott, C. J., 556 Scott, D. H., 910 Scott, J. L., 775, 910 | Galega, 535 Garcia, 867 | Wild, f. 537 Sennacrol, 528 |
| Scott, J. L., 775, 910 Scouring Rush, 323 | Ginkgo, 395 | Sennapicrin, 528 Sensitive Fern, 320-321 |
| Scouring Rush, 323 "Scours," 130 | Hura, 97 | Sensitive Fern, 320-321 Septentrionalin, 450 |
| Screenings containing cockle, 108, 439, 440 | Hyoscyamus, 726 Iatropha 587 | Septicemia, 167, 171, 867 Generalized, 172, 177 |
| Screw Pinc, 332 Scribner, F. Lamson, 903 | Jatropha, 587 Jequirity, 55, 527, 534 | Goose, 182 |
| Scribner, F. Lamson, 903 Scrophularia aquatica, 852 | Kapok, 621 Larkspur, 867 | Hemorrhagic, 177 hemorrhagica, 343 |
| marilandica, 134 | Leguminosae, 867 | Mouse, 176 |
| nadosa, 852 | Linseed, 581 Lupine, 867 Lychnis, 867 | Rabbit, 177 Streptococcic, 171 |
| SCROPHULARIACEAE, 89, 134, 698, 733-738, 852-853 | Lychnis, 867 | Septoria, 284-286 Bromi, f. 285, 285-286 |
| Scrophularin, 134 SCROPHULARINEAE, 52 | Mucuna, 524 Musk, 624 | 777 Winder 201 206 |
| Scum, 93 | Nux vomica, 867 | rubi, 284 tritici, 286 Segucia semperatirens 102 |
| Scutellaria galericulata, 131, 828 | Nux vomica, 867 Ocimum, 710 | Sequoia sempervirens, 102 |
| lateriflora, 828 | Olive, 683 Omphaloa, 867 | Serjania, 52, 89, 606, 607 |
| Sea Holly, 648 | Oxytropis, 567 Paprika, 523 Peanut, 521 | acuminata, 850 |
| Kale, 486 Mosses, 91 | Peanut, 521 | curassavica, 606, 850 erecta, 850 |
| Mosses, 91 Weed, Red. f. 194 | Physastiema, 528, 531 | inebrians, 850 |
| Sea Island Cotton, 623 Seaside Heliotrope, 705 | Poppy, 727 Prosopis, 55 Psaralea, 535 Quince, 503 | ichthyactona, 850 lethalis, 606, 850 |
| Seasonal variation affecting | Psaralea, 535 | mexicana, 850 |
| toxicity of plants, 83 Sebastiana Palmeri, 823 | Rattlebox, 546 | nodasa, 606. 850 piscatoria, 850 |
| Pavaniana, 823 | Rattlebox, 546 Rape, 867 | polyphylla, 850 trifoliatus, 606 |
| Secale luxurians, 275 Secalin, 277, 278 | Ricinus, 867 Sabadilla, 867 | Scrpentaria, 688 |
| Secalin-toxin, 149 | Sinapis, 867 | Sertürner, F. W. A., 143 |
| Secalinotoxin, 277 Sechium edule, 750 | SIMARUBACEAE, 867 Stavesacre, 464 | Service Berry, 504 |
| Secor, Eugene, 654 | Strychnos, 683 | Sesame, 698 Sesamun indicum, 698 |
| Secotium erythrocephalum, f. | Tamarind, 529 Trigonella, 531 | Sesban, 1, 561, 561 |
| 246 Securidaca longepedunculata, | Ulex, 530 | Scsbania, 534, 535, 561 aculeata, 524 |
| 843 | Vetch, 120, 527 Vicia, 867 | platycarpa, 561, f. 651 vesicaria, 118, 833 |
| Securigera Coronilla, 833 | Water Hemlock, 651 | vesicaria, 118, 833 Sessile-flowered Wake-robin, |
| Securinega, 52 ramiflora, 823 | Wormwood, 867 Segura, J. C., 910 | 385 |
| Sedative, 804 | Segura, J. C., 910 Sehlen, von, 298 | Setaria, 339, 350 |
| Sedge, f. 71, 369 Family, 367-368, 369 | SELAGINNELLACEAE, 853 | germanica, 350 glauca, f. 339 |
| Sedum, 501-503 acre, 114, 501, 502, 503, 818 | Selby, A. D., 130, 286, 701, | glauca, f. 339 italica, 102, f. 349, 350 scandens, 826 |
| acre, 114, 501, 502, 503, 818 purpureum, 501, 502, f. 502 | Seligman, C. G., 910 Selmi, F. 4, 910 Semecarpus Anacardium, | verticillata, f. 339 |
| stenopetalum, f. 502 | Semecarpus Anacardium, | viriais, 7. 207, 350 |
| Seeds, 144, 862 Aconite, 83 Adzuki Bean, 520 | 608, 806 heterophylla, 806 | Seymour, 910 Sexton, 662 |
| Adzuki Bean. 520 | Sempervirum montanum, 818 | Shaddock, 582 |
| Aleurstes, 587 | Semple, R. H., 910 Senebiera pinnatifida, 818 | Shaggy-mane Mushroom, 235 Shallon, 665 |
| Anagyris, 530 Anise, 648, 651 | Seneca Snakeroot, 585-586. | Shallot, 375, 383 |
| Baptisia, 530 | f. 585 Senecio, 755, 757, 758, 795- 797 | Shallu, 345 |
| Bassia, 681 Beans, 867 | 797 | Sharp, G., 911 Shaw Dr., 560 |
| Calabar Bean, 528 | aureus, 140 | Shaw, Dr., 560 Sheep-pox, 179 |
| Caraway, 648 Cassia, 535 | Balsamitae, 796 Burchelli, 795 | Sneep Sorrel, 106, 150, 1. 420 |
| Castanospermum, 527 | Cineraria, 756 | Sheldon, J. L., 290 Shell-bark Hickory, 401 |
| Castor Oil, 149 Cerbera, 692 | Cineraria, 756 cruentus, 756 elegans, 756 | Shellac, 406 |
| Cherry, 503 | Porsters, 755 | Shenstone, J. C., 911 |
| Cerbera, 692 Cherry, 503 Coffee Astragalus, 527 Coral Tree, 527 | guadalensis, 140, 816 Grayanus, 816 | Shepherdia argentea, f. 639, 641 |
| | ., | |

Shepherd's Purse, f. 492, 493, 867 Weather-glass, 128 Shield Fern, 319-320, f. 320 Male, 149, f. 300, 315 Extract of, 77 Shiomoyana, J., 911 Shola, 524 Shorea, 627 Shorea, 627 Short-awned Poverty Grass, Showy Lady's Slipper, 393-395, f. 394 Mentzelia, 633, 634, f. 634 Milkweed, 130, 696-697, f. Milkweed, 130, 696-697, f.
697
Moccasin Flower, 105
Shreve, F., 911
Shrubby Cinque-foil, 88
Pepper, 133
St. John's-wort, 629
Saltwort, 424
Sialagogue, 804
Siam Benzoin, 683
Sickungia rubra, 849
Sicydium monospermum, 819
Sicyos angulatus, 135, 751, 867
Sida. 124 Sida, 124 Sida, 124
jamaicensis, 624, 837
paniculata, 124, 624, 837
retusa, 623
rliombifolia, 624
Stinging, 124
urens, 124, 837
Sidamgrotzky, 26
Side-saddle Flower, 114
Sideroxylon borbonicum, 851
dulcificum, 851
tomentosum, 681 dulcificum, 851
tomentosum, 681
toxiferum, 851
Siebenmann, 199
Siegel, A., 911
Sieva Bean, 574
Sikimin, 474
Silage, Poisoning from, 24-28, 867
Silene, 89, 436, 437-439
antirrhina, 108, 438, f. 438, 813 813 Armeria, 437 Cucubalis, 813 Griffithii, 813 latifolia, 437-438 macrosolen, 813 noctiflora, 108, f. 437, 437-438, f. 438 stellata, 437 virginica, 813 viscosa, 813 ilesia, Poisonous plants of, Silesia, Pe Silk-worm Disease, 163 Silky Sophora, 117, 542, f. Silky Sophora, 117, 542, f.

Silphium, 757
Silver Maple, 615
Nitrate, Poisoning hy, 6
Plant, 419
Silver-bell Tree, 682
Silver-leaf Psoralea, 558
Silvery Lupine, 550
Psoralea, 118
Silybum, 758, 802
Marianum, 802, 816
Simaba Cedron, 583
valdivia, 583
Waldinii, 853
Simaruba amara, 853
Bark, 583
SIMARUBACEAE, 89, 121, 575 SIMARUBACEAE, 89, 121, 575, 577, 583-584, 853, 867 Simhlum, 245 Simon, J. F., 3, 911

Simpson, W. T., 911 Simpson Honey Plant, 134 Sinalbin, 489 Sinapin-sulphate, 489 Sinapin-suipnate, 407
Sinapis, 868
Sinapism, 113, 114, 490, 494
Sinigrin, 489, 491
Sinkler, 911
Siphiocampylos giganteus, 812
SIPHONGAMA, 101, 325
Sieal 386, 261 Sisal, 386 Sisyrinchium angustifolium, Sisyrinchium angustifolium, 828
Sitanion elymoides, 103
Sium, 651, 659, 868
cicutaefolium, 651, 857
crectum, 857
latifolium, 857
Skimmia japonica, 849
Skullcap, Marsh, 131
Skunk Cabbage, 103, 371
Slade, II. B., 96, 338, 911
Sleepy Catchfly, 107, 438, 91
Sleepy Catchfly, 107, 438, f.
Grass, 102, 355, 357-358
Slender Gerardia, 134, 738
Nettle, 106
Pigweed, 432
Psoralea, 118
Wheat Grass, 364
Slime Mold, f. 158, 159, 167
Fresh-water Green, f. 191
Slippery Elm, 408, f. 409
Sloanea dentata, 622
Slobbering, Cause of, 557
"Slobbers," 603
Sluyter, Theodor, 263
Smatl Beggar-ticks, 139
Cranberry, 665
Horse-weed, f. 777 828 Cranberry, 665
Horse-weed, f. 777
Lady's Slipper, White, 105,
393 Yellow, 105, f. 393 Peppergrass, 113, f. 494, 494 Ragweed, f. 766, 766-767 Stickseed, 780 Stinging Nettle, 412-413, f. Small-flowered Crowfoot, 458, f. 459 Marsh Elder, 763 Smartweed, 78, 106, 107, 421, Smartweed, 78, 100, 107, 421, 422
Disease caused by, 12
Family, Weeds of, f. 420
Smut on, 98, 219
Water, 421-422
Smilax, 89
Cultivated, 375
herbacea, 64
pseudo-syphilitica, 835
rotundifolia, 104
Smith, Bernhard S., 73-77, 803, 804, 911
Smith, Clinton D., 213, 214, 344
Smith, E. F., 100, 287, 293, 889, 911
Smith, F., 261, 911
Smith, John, 441, 596, 694
Smith, J. B. 911
Smith, Theobald, 214
Smith, W. G., 272, 911, 914
Smith, W. G., 272, 911, 914
Smithsonian Institution, 911
Smoke Tree, 608 Smoke Tree, 608 Smoooth Lepiota, 240 Sumac, 608, 613-614 Tobacco, 133 Zygadenus, 378-379 Smut, Barley, 98, 217 Cheat, 218 Corn, 98, 210-215

Foxtail, 217, f. 218
Millet, 214, f. 218
Oat, 215-217
Loose, 98
Tall Meadow, f. 216, 218
Pigeon Grass, 98
Porcupine Grass, f. 216
Rye, 218
Sandbur f. 218 Rye, 216 Sandbur, f. 218 Smartweed, 98, 219 Sorghum, 212 Sprouting Grass, f. 214 Stinking, 219-220, f. 219 Timothy, 218 Stinking, 218
Timothy, 218
Wheat, 217
Stinking, 98
Smyrna Opium, 85
Snakebites, Antidotes against, 417 Snake Grass, 692 Snake Grass, 692
Poison, 7
Venom, 2
Snakeroot, 773
Black, 692
Button, 648, 692, 755, 77374
Seneca, 121, 585-586, f. 585
Virginia, 417
Western, 773-774
White, 771-772
Snap Dragon, 733
Sneezeweed, 139, f. 142, 756,
781-784, f. 782, 868
Fine-leaved, f. 784
Mountain, 140
Narrow-leaved, 783-784 Mountain, 140
Narrow-leaved, 783-784
Sneezewood, 608
Snowberry, 135, 744, 747-748
Snowdrop, 386, 868
Tree, 682
Snow-on-the-Mountain, 64, Snow-on-the-Mountain
121, 598, f. 599
Snuff, 381
Soap Bark, 505
Brush, 120
Fish, 593
Plant, 104
Tree, 123
Soaphery, 123 Soapberry, 123 Tree, 606 Tree, 606
Soap-root, 380
Soap-root, 380
Soapwort, f. 436, 441, 688
Sobernheim, J. F., 3, 55, 911
Socrates, 2, 651
Sohn, C. E., 442, 911
Soil affecting toxicity of plant, 83, 87-88
Soir, 869, 911
Soja Bean, f. 520
SOLANACEAE, 52, 60, 72, 89, 131-133, 144, 698, 713-733, 853-855, 868
Solandra grandiflora, 854 Solandra grandiflora, 854 Solanein, 718 Solanein, 718
Solanidin, 132, 148, 718
Solanin, 60, 83, 132, 133, 144, 148, 149, 718, 721, 722, 724, 804
Solanum, 89, 714, 717, 718-725, 868
aculeatissimum, 854 aculeatissinum, 854
aviculare, 854
carolinense, 60, 132, f. 723,
724, 854
citrullifolium, 723
Dulcamara, 60, 138, 715,
718-719, f. 710
elaeagnifolium, 724
ellipticum, 854
eremophilum, 724, 854
esuriale, 854
grandiflorum, 854
Jamesii, 714

INDEX

| mammosum, 854 |
|--|
| mammosum, 854 Melongena, 714, 718, 854 nigrum, 4, 60, f. 61, 74, 131-132, f. 132, 719-721, f. 720, 854 rostratum, 718, 722-723, f. |
| f. 720, 854 |
| 163 |
| saponaceum, 854 sodomeum, 854 Stramonifalium, 854 |
| Sturtianum, 854 tarvum, 854 |
| triflorum, 1, 32, f. 721, 722, |
| tuberosum, 132, 714, 718, f. 718, 721, 855 |
| tuberosum, 132, 714, 718, f. 718, 721, 855 villosum, 855 Xanti, 855 |
| Soldier's Herb 106 |
| Solenostemma Argel, 810 Solidago, 757, 774-776 conadensis, 138, 774-775 Rust on, 222 |
| conadensis, 138, 774-775 Rust on, 222 |
| |
| missouriensis, 774 adara, 755, 774 rigida, 775-776 seratina, Rust on, 222 speciasa, 774 |
| seratina, Rust on, 222 speciasa, 774 |
| Solomon's Seal, 377 |
| Sonchus, 757, 759-760 |
| aleraceus, 759 |
| speciasa, 774 Virgaurea, 816 Solomon's Seal, 377 Sommerfeld, T., 911 Sonchus, 757, 759-760 arrensis, 759-760 aleraceus, 759 Rust on, 222 Saphora, 535, 542-543, 868 alapecuroides, 833 |
| angustifolia, 530 |
| flavescens, 90 japonico, 833 mollis, 833 |
| secunainora, 90, 117, 534, |
| 542-543, 833 sericea, 90, 117, 542, f. 542, 564, 833 Silky, 117, 542, f. 542 speciosa, 530 tamentosa, 90, 833 SOPHOREAE, 530 Sophorin, 530, 543 Soporific, 804 Sorbite, 506 |
| Silky, 117, 542, f. 542 speciasa, 530 |
| tamentosa, 90, 833 SOPHOREAE, 530 |
| Sophorin, 530, 543 Soporific, 804 |
| Sorbite, 506 Sordariaceae, 280 |
| SORDARIACEAE, 280 Sarghum, 54, 102, 338, f. 345, 345-348, 503, 747, 868 |
| 868 Smut, <i>f. 42</i> |
| Smut, f. 42 vulgare, 53, 346, 826 Sarosporium, 210 Sorrel, 579-580 |
| French, 419 Jamaica, 623 |
| Mountain, 419 |
| Sheep, 106, 150, f. 420 Wood, 120 |
| Violet, 580 Soudan, Poisonous plants of, 866 |
| Sour Cherry 505, 514 Dock, 106, f. 420 Orange, 582 |
| Orange, 582 South American Saffron, 389 |
| Sage, 710 Southern Buckthorn 679 |
| Mistletoe, 415, f. 416 |
| Mistletoe, 415, f. 416 Sow Thistle, 759-760 Annual, 759 Field, 759-760 |
| Soy Bean, 520, 548 Spanish Chestnut, 403 Needle, 67, f. 780, 780-781 |
| Needle, 67, f. 780, 780-781 |

Wormseed, 424 Vetch, 572 Sparganium, 332, f. 334 Spartein, 147, 551 Spartina cynosuroides, f. 69, 338, 339 Spartium junceum, 826 scoparium, 147 SPATHIFLORAE, 155, 370-372 Spathiphyllum candicans, 808 Spatzier, 633 Spearmint, 709 Speedwell, 228, 229, 733 Purslane, 735-736 Spergula orvensis, 436 Spermacoce, 90
capitota, 849
semierecta, 849
SPERMATOPHYTA, 155,
325-802 Sphacelenic acid, 278, 279 Sphacelia, 275 Sphacelinic acid, 28 Sphacelotheca, 210 Sphacelotoxin, 277 Sphaerella Fragariae, 281 Sphaeriaceae, 267, f. 268, SPHAERIACEALES, 280-281 Sphaerotheca Castagnei, f. 269, 270 Sphaerozyga, 186 Sphedamnocarpus angolensis, 837 Spice-bush, 477 Spider Poison, 7 Spiderwort, 373 Blue, 373 Spielcr, A. J. T., 911 Spigelia, 698, 868 Anthelmia, 836 glabrata, 836 marilandica 76, 129, 688, 836 pedunculata, 836 pedunculata, 836 Spigelin, 688 Spiked Lobelia, 753 Pale, 136 Spikenard, 647 Spilanthes Acmella, 816 aleracea, 816 Spinach, 150, 424, 427, 435 Australian, 427 New Zealand, 423 Spinacea oleracea, 424 Spinacea oleracea, 424
Spinal cord, Poisons acting
on, 74 on, 74 Symptoms of, 74 Treatment, 74 Spiny Amaranth, 432, f. 432 Clothur, f. 137 Spiraca, 189, 504 Aruncus, 848 bella, 848 cauescens, 848 Douglasii, 504 Filipendula, 848 Humbaldtii, 848 Humbaldtii, 848 lyspericifolia, 848 japonico, 503, 504, 848 laczigata, 848 salicifolia, 504 Thunbergii, 504 tamentosa, 848 tamentosa, 84 Ulmaria, 492 Spiranthes, 392 SPIRILLACEAE, 165, 182 Spirillum, 150 camma, 161 rubrum, 161

Spirachaeta anscrina, 182 Obermeieri, 182 pallido, 182 Spirogyra, 92, 94, 95, 189, 190-Spleenwort, 318-320 Common, 318 Spondias dulcis, 608 luteo, 608 mangifera, 806 purpurea, 608 Sponge, Fresh-water, 93 Gourd, 750 Sponia virgado, 53 Sporogum, 160 Spirachaeta anscrina, 182 Sporongium, 160 Spores, 160 Sporodinia, 195 Sporotrichosis, 868 Sporotrichum, 13, 15, 297, 302 Furfur, 100, 297, f. 297 giganteum, 299 minutissimum, f. 298, 298-299 tonsurans, 100, f. 300, 300-303 Epidermis invaded by, f. Epidermis invaded by, f. 296
Spot disease of currant, 281
Orchard Grass, 282, f. 283:
Strawberry, 281
Spotted St. John's-wort, 630
Spurge, f. 597, 597-598
Large, 597, f. 602
Touch-me-not, 123, 620
Spratt, G., 911
Spreading Dogbane, 129, 692
Nightshade, f. 721
Sprekelia formosissimo, 386, 805
Sprouch, C, H. E., 912 Sprouch, C. H. E., 912 Sprouting Grass Smut, f. 214 Snruce, 868 Black, 328 Norwey 227 Snruce, 868
Black, 328
Norway, 337, 328
Tideland, 327
White, 328
Spurge, 597-603, 868
Caper, 121, f. 590, 602
Common, 602
Cypress, 121, 600, f. 601
Family, 89, 586-604
Flowering, 121, 122, 598, f. 598
Yellow, f. 601
Laurel, 125, 642, f. 643
Myrtle, 121, 599
Nettle, 122, 596, f. 596
Spotted, f. 607, 597-598
Large, 597, f. 602
Spurrey, 436
Squash, Bush, 750
Crookneck, 750
Hubbard, 751
Winter, 750
Squaw-weed, 138
Squill, 148, 375, 377
Squirrel-tail Grass, 66, 103, 366-367, f. 368
Powdery Mildew on, 272
Injuries from, 360
Squirting Cucumber, 749
Stachys arvensis, 131, 828
palustris, 828 Stachys arvensis, 131, 828 palustris, 828 Sieboldii, 710 Stachytarpheta indica, 858 Stadhlinger, 482 Staff-tree, 614-615 Family, 614-615 Stagger-bush, 128, 674, f. 675,-675 Staggers, 100, 388 Mad, 259 Stomach, 261 Staggerwort, 140

Stagliorn Sumac, 608
Stahl, A., 3
Stahagmitis Mangle, 827
Stalberg, S., 912
Stalker, M., 9, 118, 187, 277, 356, 544, 564, 912
Stange, C. H., 24-28
Stapelia, 64, 695, 810
Stapf, O., 912
Staphisagrin, 108, 464, 868
Staphylea trifolia, 604
STAPHYLEACEAE, 604 STAPHYLEACEAE, 604 Staphylolisin, 168 Leucocidin, 168 Stapliylacaccus pyogenes
aureus, 168
Star Anise, 149, 648, 863, 868
Apple, 679, 681
Cucumber, 751
-of-Bethlehem, 377 Tacca, 374
Yam, 374
Yam, 374
Statice brasiliensis, 675
pectinata, 128, 842
Statutes on Poisoning, Iowa Statutes on Poisoning, Iowa code, 6 Stavesacre, 446, 460, 463 Seed, 464 Stearic acid, 593 Stearin, 594 Stearns, Fred, 468 Stebler, 96, 325, 439, 449, Stebler, 96, 325, 439, 449, 560
Stellaria, 436
crassifolia, 813
media, 107, 442-444, f. 444, 868 868
Steironema, 676
Ianceolatum, 676
Stemless Loco Weed, 39, 119, 566, 567-570, f. 568
Stemonitis, 159, 160
Stephania, 89
aculcata, 838
discolor, 838
Sterculia alata, 855
STERCULIACEAE, 53, 621, 855
Stercocaulan ramulosum, f. 307 307 Sterigmata, 21 Sterigmata, 21 Sterigmatacytis, 258 Stevens, A. B., 912, 915 Stevens, F. L., 206 Stewart, F. C., 126, 210, 663 Stickseed, 131, 706-707 Large-flowered, 707 Large-flowered, 707

Stick-tights, 786

Stillingia, 590

lineata, 823

sylvatica, 604, 824

Stillmark 595

Stimulant, 804

Stinging Nettle, Small, f.

412, 412-413

Nut, Brazilian, 596

Sida, 124

Stinkborn, 99, 239, 245 Sida, 124
Stinkhorn, 99, 239, 245
Stinkhorn, 99, 239, 245
Stinking Clover, 114
Smut of Wheat, f. 219, 219-220
Willie, 797
Stipa, 339, 353, 578, 579
avenacca, 66, 354
capillata, 354, 926
comata, 66, 70, 102, f. 355, 355-357
hystricina, 53 355-357 hystricina, 53 inebrians, 354, 355, 826 lessingiana, 90 Neesiana, 354 palmata, 826 pinnata, 826

robusta, 90, 102, 357-358, 826 826 sibirica, 354 spartea, 70, 102, 210, 354, 355, f. 356 tenacissima, f. 354 Western, 355, f. 355 Stipe, 160 Stitchwort, 436 Stizolobium Lyoni, 526 Stock 486 Stizolobium 55 Stock, 486 Stomatitis, 179, 272, 290, 642, 649, 868 Catarrhal, 20 Mycotic, 256, f. 259, 260, 047, aug Catarrhal, 20 Mycotic, 256, f. 259, 260, 282, 283 Stockberger, W. W., 688, 912 Stokes, J. S., 912 Stone Clover, 553 Stone, G. F., 613 Stonecrop, 114, 221, 501 Mossy, 501, 502 Stonewort, 92, 194, 688 Storax, 500 Storesin, 500 Stork'sbill, 67, 120, 578-579 Hemlock, f. 579 Stowe, W., 912 Stramonium, 72, 718 Poisoning by, 6, 78 Purple, 729 Strangles, 170 Strasburger, E., 82, 893, 901, 912 Stratuard, R., 912 912 Strattuard, R., 912 Strawberry, 504-505, 629, 868 Blite, 474 Bush, 475 Chilean, 505 Cultivated, 115, 609 European, 115 Wood, 509 Garden, 505 Indian, 505, 509 Rust, 281 Indian, 505, Rust, 281 Tomato, 715 Tree, 665 Virginia, Wild, 115, 509 Streptococcic Septicemia, 171 Streptacoccus melanogenes, Streptacoccus melanogene 28, 868 equi, 170 erysipelatos, 171 pyogenes, 87, 171 Stricptocolysin, 171 Strickland, C. W., 912 Striga euphrasioides, 852 String Bean, 520 Strabilanthes, callasus, 69 Strobilanthes callosus, 698, 807 Strong, A. B., 912 Strong, A. B., 912 Strophanthus, 78, 868 Eminii, 807 hispidus, 148, 692, 807 Kombe, 692 Pierrei, 807 Strophantin, 148 Stropharia aeruginosa, 860 Strümpell, Dr., 572 Strychnia, Statutes regarding, 6 Strychnin, 1, 72, 73, 82, 129, 144, 145, 146, 147, 628, 686, 688, 728, 729, 736, Antidotes against, 528 Discovery of, 3 Nitrate, 688 Poisoning by, 6, 77 Sulphate, 688 Toxicity of, 7

Treatment for poisoning Treatment for noisoning by, 81
Strychnos, 52, 685, 868
angustifolia, 836
bracliata, 836
Castalnaei, 148, 836
cagens, 836
colubrina, 836
Crevauxiana, f. 685, 686, 836 gubleri, 836 Icaja, 836 Iguatii, 74, 1 inuocua, 836 144, 688 111110cua, 836
Melinoniana, 836
Mulinoniana, 836
11111x-vanica, 74, 85, 147,
685, 688, 836
paniculata, 836
patatorum, 685, 836
Pseuda-quina, 836
Sauthierana, 836 patatorum, 685, 836
Pseuda-quina, 836
Santhierana, 836
spinasa, 836
spinasa, 836
spinasa, 836
spinasa, 836
stryplinadendran Barbatiman, 833
polyphyllum, 833
Stryplinadendran Barbatiman, 833
stractia, 629
Stubbleberry, 719
Studor, 241
Stuhr, W. A., 17-19, 20-24, 28-29, 37, 39-40, 303
Sturdevant, I., B., 912
Stylapharum, 868
diphyllum, 113, 480, 841
Stylopin, 113, 480
Stypandra glanca, 835
Styptic, 396
Stypandra glanca, 835
Styptic, 396
Styraccexex, 679, 682-683
Styracin, 683
Styracin, 683
Styracin, 683
Styracin, 683
Styraca Benzoin, 682, 683
Family, 682
Suacada fruticosa, 424
Subjee, 411
Succory, 758-759
Sudworth, G. B., 332
Sugar Beet, 107, 424-427, 868
Cane, 338
Maple, 615, f. 616
Pine, 329
Suclivant's Honeysuckle, 744 Maple, 013, 1. 010
Pine, 329
Suicide by Poisoning, 2, 868
Sullivant's Honeysuckle, 744
Sulpho-cyanate of allyl, 489
Sulphur Polyporus, 234
Sulphur dioxid, 72
Sulphuric acid, Poisoning by, Sulphuric acid, Poisoning by,

Sumac, 122, 608-614, 868
Chinese, 583, f. 584
European, 608
Japanese, 608
Poison, 609, f. 610
Smooth, 608, 613-614
Stag-horn, 608
Summer Chrysanthemum, 756
Cypress, 107
Sundew, 77, 114, 497
Common, f. 500
Family, 89, 114, 497-498
Sunflower, 756
Great, 756
Maximilian's, f. 763
Powdery Mildew of, 269
Superbin, 377
Supple-jack, 620
Surgeon General of U. S.
Army, 892
Surinam, Poisonous plants of, 867
Susotoxin, 150, 171 Susotoxin, 150, 171 Susum anthelminticum, 825 Sutton, R. I., 912 Swain, R. E., 888, 912

| Swainsona, 534 |
|---|
| coronillaefolia, 533, 833 Greyana, 533, 833 Oliverii, 833 phacoides, 833 |
| Oliverii, 833 |
| phacoides, 833 |
| |
| Swamp Camas, 103 Hellebore, 103 Laurel, 672 |
| Leucothoe, f. 673, 673 |
| Leucothoe, f. 673, 673 Lousewort, 738 Milkweed, 130 |
| |
| 332 |
| Turnip, 486 Swartzia triphylla, 833 |
| Sweet Alyssum, 486 |
| Sweet Alyssum, 486 Balm, 397 Basil, 709 Bay, 474 |
| Bay, 474 Birch, Oil of, 86 Calomel 371 |
| Birch, Oil of, 86 Calomel, 371 Cassava, 54, 587 |
| Cassava, 54, 587 |
| Cherry, 514 Cicely, 648 |
| Clover, 552 |
| Calonie, 371 Cassava, 54, 587 Cherry, 514 Cicely, 648 Clover, 552 White, 118, f. 551, 552 Yellow, 118, 552 Fern, 399 |
| Fern, 399 |
| Fern, 399 Flag, f. 398 Gale, f. 398 Family, 397 Hay, 105, 389 |
| Family, 397 |
| Hay, 105, 389 |
| 1 0(4(0, 0)0 |
| William, 437 Sweet-scented Bedstraw, 744 |
| Shrub, 476 Sweethrian, 115 |
| Swertia chirata, 689 |
| Swietinia humilis, 837 Mahogoni 575 837 |
| Sweetbriar, 115 Sweetla chirata, 689 Swietinia humilis, 837 Mahogoni, 575, 837 Switzerland, Poisonous plants of, 867 |
| |
| Sycamore 500 |
| Sycosis parasitaria, 301 Sylvacal, 604 Syme, W. A., 612, 613, 869, 912 |
| Syme, W. A., 612, 613, 869, |
| 912 Symphoricarpos 157 745 |
| Symphoricarpos, 157, 745, 747, 748 mollis, 135, 813 occidentalis, 744, 747 orbiculatus, 135, 744, 748 racemosus, 135, 744, 748, 813 |
| mollis, 135, 813 |
| orbiculatus, 135, 744, 748 |
| racemosus, 135, 744, 748, 813 |
| Symphytum officinals 704 |
| Symplocarpus foetidus, 103, |
| Symplocarpus foetidus, 103, 371, 808 Symplocos Alstonio, 629 |
| lanceolata, 614 rocemoso, 681 |
| Symptoms of Poisoning, 72 |
| Synaktonin, 450 |
| Sundiesa ramulaea 4 207 |
| symphorea, f. 309 SYNANTHAE, 155 |
| Synaptase, 505 Syncephalis cornu, 196 intermedia, 196 |
| Syncephalis cornu, 196 intermedia 196 |
| Synchytrium, 204 decipiens, 204 |
| decipiens, 204 SYNPETALAE, 157 |
| Sunura 188 |
| Syphilis, 182 Syringa, 89, 115 |
| Syringa, 89, 115 bersica, 683 |
| persico, 683 rulgaris, 63, 683 |
| Syringin, 684 Syringopicrin, 128 Sisymbrium, 486, 487-488 Alliaria, 818 |
| Sisymbrium, 486, 487,488 |
| Alliaria, 818 |
| altissimum, 114, f. 487, 488 |

officinale, 114, 487, 818 Sophia, 818 toxophyllum, 818

Tabernaemontano Borbonica, Tabernaemontano Bon 807 citrifolia, 807 coronaria, 807 malaccensis, 867 mouritiono, 807 Tabernanthe Iboga, 8 Tacco pinnatifida, 374 Starch, 374 807 Tacco pinnatifida, 374
Starch, 374
TACCACEAR, 374
Tachio guianensis, 825
Tacsonia, 53, 633
Taenia serrata, 568
Taenifuge, 804
Tagetes erecta, 756
Takosis, 168
Talauma, 90 macrocarpa, 474, 837 ovata, 837 Talinum polyandrum, 844 Talisia stricta, 850 Talisia stricta, 850
Tall Blackberry, 665
Buttercup, 10, f. 459
Cone-flower, 779
Crowfoot, 458
Gypsophyl, 436
Larkspur, 108, 462, 463, f. 464, 465, 466, 467
Mcadow Oat Grass, f. 340
Tallow Tree, 587
TAMARICACEAE, 627
Tamarind, 529, f. 531
Tamarindus indica, 529, f. 531 Tamarix gallica, 627 mannifera, 627 Tampicin, 702 Tamus communis, 820 Tanus communis, 820
Tanacetin, 756
Tanacetum, 757, 758, 789-790
balsomita, 789
umbelliferum, 816
vulgare, 756, 789-790, 793,
816 Tanaccium crucigerum, 810
Tanausek, 364
Tangerine, 582
Tanghin, 868
Tanghin, 868
Tanghin venenifera, 148
Tangier Pea, 526
Tannic acid, 144, 403, 503,
608 608 Tanuin, 105, 319, 327, 403, 418, 608
Tanret, Ch., 277, 278
Tansy, 789-790 Common, 140 Oil, 756, 789-790 Tanweed, 421 Tape-worm, Presence in lo-coed sheep, 568 Taphrinae, 252, 253 Tapioca, 75, 587 Tapiura, 2, 621 guianensis, 819 Tar-weed, 138, 755 Taraktogenos Blumei, 53 Taraxacerin, 755 Taraxacin, 755 Taraxacum, 757
officinale, 755, 756, f. 759
Tarragon, 756 Tartaric acid, 529, 868 TAXACEAE, 101 Taxin, 101, 148, 328

Toxus, 52, 148, 328
baccata, 4, 328, 817, 868
brevifolia, 101, 328, 817
canadensis, 101, 328, 817
Taylor, Λ. S., 4, 363, 913
Taylor, J. L., 130
Taylor, J. L., 130
Taylor, M. E., 357
Taylor, Thomas, 913
Tea, 628, 868
Black, 628
Chinese, 614
Green, 628
Labrador, 127, 629, 666
Mexican, 107
New Jersey, 621, 629 Mexican, 107, 600
New Jersey, 621, 629
Oswego, 109, 629
Plant, 628, 629
Substitutes for, 436, 629
Wild, 501
Teak Tree, Indian, 708
Teasel, 7, 741
Tecoma, 52
cergmensis 810 ceramensis, 810
jasminoides, 739
leucoxylon, 810
obtusala, 810
radicans, 135, 739, 810, 862
speciosa, 810
stans, 810
toxophora, 810
Tectono grandis, 708
Teilleux, 573
Telfairia pedato, 751, 819
Temperature affecting toxicity
of poison, 868 ceramensis, 810 of poison, 868 Templetonio, 868 egena, 833 Templetonio, 868
egena, 833
purpurea, 833
retusa, 833
Tenbosch, J., 913
Tennessee, Poisonous plants of, 866
Teosinte, 230
Rust on, 230
Tephrosia, 1, 52, 535, 558
astragoloides, 833
candida, 833
cinerea, 833
coronillaefolia, 833
densiflora, 833
frutescens, 833
macropoda, 833
nitens, 833
periculosa, 833
purpurea, 534
tomentoso, 833
toxicario, 118, 533, 558, 833
virginiana, 118, 558, 833
Vogelii, 833
Tephrosin, 533, 558
Teramus labialis, 833
Terminalia Bellerico, 814
Catappa, 638, 640
Chebula, 640, 814
fagifolia, 640
tomentoso, 814
TERNERGEMIACEAE, 52, 89,
855 TERNSTROEMIACEAE, 52, 89, TERNSTROEMIACEAE, 52, 89, 855
Tesnier, T. J., 913
Testicularia, 210
Tetanin, 10, 150
Tetanoid spasms, a symptom of deliriant poisoning, 73 Tetanolysin, 174 Tetanospasmin, 174 Tetanotoxin, 150 Tetanus, 150, 174, 868 Bacillus, 174 Toxin of, 77, 174

Tetracera, 52
alnifolia, 819
Assa, 819
Tetradenia fruticosa, 828
Tetragonia expansa, 423, 824
Tetrapanax papyrifera, 647
Tetrapleura, 89
cacrulea, 833
Thominoii 833 carrilea, 833
Thonningii, 833
Teucrium canadense, 709
Chamaedrys, 828
Marum, 828
scordium, 828
Texas Croton, 121, 592, f.
592, 593
Lily, 690
Thacher, J., 913
Thalictrin, 446
Thalictrim, 868
aquilegifolium, 53
flavium, 846
macrocarpium, 446, 846
polycarpium, 111
revolutum, 111
Thalins, 275
THALLOPHYTA, 91
Thaliophytes, 188, 310
Thapsia garganica, 857
villosa, 857
villosa, 857
Thaumatococcus Daniellii, 858
Thayter, B. 167 Thonningii, 833 858 Thaxter, R., 167 Thea assamica, 63, 629 Thea assamica, 63, 629
Kissi, 629
sinensis, 628
sinensis bohea, 628
sinensis viridis, 628
Sassanqua, 63, 629
THEACEAE, 63, 629, 629
Thebain, 146, 147, 481
Thecaphora, 210
Thein, 146, 629, 742
Theobromia Cacao, 621, 623
Theobromin, 144, 146, 621, 628 628 Theophrasta americana, 839 Theophyllin, 146, 628 Thermopsis, 118, 534, 535, Theophyllin, 146, 628
Thermopsis, 118, 534, 535, 538-539
Alleghany, 538-539
caroliniana, 90
mollis, 538-539
montana, 90, f. 539, 539
Prairie 539
rhombifolia, 117, 539, 833 Therapeutics, Literature upon, 868
Thespesia grandiflora, 623
Theveresin, 692
Theretia, 52
Ahouai, 807, 849
ncreifolia, 148, 692, 807,
849
Thevetin, 148 sp., 551 ** 849
Theyetin, 148, 692
Thin, G., 913
Thistle, 797-802, f. 802
Bull, 141, 798, f. 800
Canada, 141, 798, f. 799
Family, 757-802
Lady's, 802
Milk, 802
Prairie, 801
Russian, 68, 150, 424, 430, f. 437, 432
Sow, 759-760
Annual, 759 Sow, 759-760 Annual, 759 Field, 759-760 Woollv. f. 798, 801 Thlaspi, 487, 494-495 arvense, f. 495 Thompson, C. J. S., 913 Thomsonia napalensis, 372

Thorn Apple, 60, 74, 715, 729-733
Purple, 729-733
White, 513
Thorns, Rose, 67
Thorny Clotbur, f. 768
Pigweed, 107
Thottea dependens, 809
Three-flowered Nightshade, f. 721, 722
Three-leaved Poison Ivy, 609-610
Three-lobed Kidney Bean, 520 733 520 Three-seeded Mercury, 83, Three-seeded Mercury, 83, 122, 603, 604
Thresh, 726
Thrush, 302-303, 868
Thuja, 78
occidentalis, 328, 756, 817
Thuiin 328 Thujin, 328, 756, 817
Thujin, 328
Thujone, 328, 710, 756, 790
Thunberg, C. P., 913
Thunbergia alata, 698
Thurber, Dr., 679
Thurber, Goo., 913
Thyme, Garden, 709
Thymelaca Tartonraira, 855
THYMELARCEAE, 52, 125, 640, 642-643, 855
Thymol, 89, 648, 709, 710
Thymus albus, 89
officinalis, 710
Serpyllus, 89
ovulgaris, 709
"Ti," 375
Tickseed, Small, 780 "Ti," 375
Tickseed, Small, 780
Tickseed, Small, 780
Tickle Grass, Rust on, f. 223
Tidy, C. II., 913
Tieute, 687
Tiger Lily, 375
Tiglinic acid, 593
Tilia americana, 621
cordata, 621
TILLACEAE, 53, 89, 621, 855
Fish-poisoning among, 2, 52
Tiliacera racemasa, 838 Tiliacora racemosa, 838
Tiliacora racemosa, 838
Tiliandsia usneoides, 372
Tilletia, 98, f. 218, 219-220, 861
hordei, 219
secalis, 219
Tritici, 220, 861
TILLETIACEAE, 98, 210, 219-220, 861
Tillie, T. B., 884
Timothy, 338
Ergot on, f. 27, 276
Tinea, 295
circinata, 301
circumscripta, 300 Circinata, 301
circumscripta, 300
disseminata, 300
favosa, 294-296
of Fowl, 14
Horse, 14
Sycosis, 300
fonsurans, 12, 14, 300
'Transmission of, 14
'Ting, Chung Yu, 237
Tireli, 290
Tirelli, V., 913
Tissue change, Poisonous products of, 73
Tizzioni, Guido, 913
Toad-flax, 134, f. 735, 735-736
Toadstool, 31, 160, 220, f. 232, 503
Lepiota, 243
See Mushrooms also
Tobacco, 72, 75, 86, 133, f. 714, 725-729, 868
Adulterants of, 773 circumscripta, 300

Alkaloids of, 147, 728
Common, 727-729
Flowering, 133, 715
Indian, 75, 136, 752
Nicotine in, 86, 87, 728
Smooth, 133
Wild, 729
Toccoc guiannessis, 640
Toddalea aculeata, 88
Toddy, Philippine, 370
Toffana of Naples, 2
Tofieldia calyculata, 835
Tokishige, 249, 913
Tolenc Oil, 528
Tolnifera, 90
Tolyposporium, 210
Tolyposporium, 210
Tomato, 132, 1-713, 714
Currant, 715
Strawberry, 715
Tree, 716
Tonka Bean, 523, 552, 868
Tonguin, 523
Tonsilitis, 171, 177
Toot Poison, 868
Torenia asiatica, 733
Torrey, John, 913
Torrey, John, 913
Torrey, John, 913
Torrey, John, 913
Torreya nucifera, 817
Touch-me-not, Spotted, 123,
Tovey, J. R., 882, 913 Touch-me-not, Spotted, 123, 620
Tovey, J. R., 882, 913
Towel Gourd, 750
Toxalbumin, 52, 55, 165, 804
Abrus, 55-56, 149
Black Locust, 56-58
Castor Oil Seed, 55, 149
Toisoning from, 55-58
Toxicodendrol, 394, 612, 620
Toxicodendrol, 394, 612, 620
Toxicodendron capense, 588
slobosum, 824 Toxicodendron capense, 58 globosium, 824
Toxicology, 1, 3, 868
Vegetable, 868
Veterinary, 868
Toxins, 10, 149, 164, 868
Animal, 149
Defined, 10, 164
Food, Classified, 10-11
Plant, 149 Treatment for poisoning by, 81 Toxiresin, 475
Trachoma, 265
Trachymene australis, 857
Tracy, S. M., 87, 913
Tradescantia, 89, 374
crassifolia, 814
elongata, 814
virepianum, 373 elongata, 814
virginianum, 373
Zebrina, 374
Traganthin, 530
Tragia, 590, 593-594
cordifolia, 824
involucrata, 824
nepetaefolia, 122, 594
urens, f. 593, 594
volubilis, 824
Tragopogon porrifolius Tragopogon porrifolius, 756
Trailing Arbutus, 127, 665
Wolf's-bane, 109
Trametes radiciperda, 234
Trapa natans, 840
Trapa natans, 840
Traveler's Palm, 391
Tree Fern, 313
Pepper, 608
Tomato, 716
Tree-of-Heaven, 121, 583, 584, f. 584
Tree-toot, 607
Trees, Malignant effect of, 868
Trelease, Wm., 68, 91, 186, Trelease, Wm., 68, 91, 186, 279, 635, 636, 806, 913 Trema aspera, 857

Turnera diffusa, 628 TURNERACEAE, 627

INDEX

| "Trembles," 771, 772 |
|---|
| "Trembles," 771, 772 Tremella lutescens, f. 220 TREMELLINEAE, f. 220 |
| i remelladan Pelalinusum, I. |
| Treub, M., 54, 82, 747, 913 Trevesia, 808 Trianthema, 89, 824 Portulacastrum, 824 |
| Trevesia, 808 |
| Portulacastrum, 824 |
| |
| maximus, 120, 858 |
| Tribulus, 89 cistaides, 858 marimus, 120, 858 terrestris, 120 Trichadenia zeylanica, 53 Tricalypsa Sonderiana, 849 Trichia decibieus, f. 150 |
| Tricalypsa Sonderiana, 849 |
| Trichia decipiens, f. 159 varia, f. 158 Trichilia, 89 |
| Trichilia, 89 |
| emetica, 575 |
| maschata, 837 trifolia, 837 |
| Trichaloma album, 860 saponaceum, 860 sulplurcum, 860 Trichomanes alatum, f. 309 radicans, 313 Trichophytic Fungi, 295, 300 Trichophyton, 12, 13, 14, 302, 868 atractabhara, 301 |
| sulphurcum, 860 |
| radicans, 313 |
| Trichophytic Fungi, 295, 300 |
| 868 |
| atractophoran, 301 cctothrix, 13, 301 cndo-ectothrix, 13 |
| cndo-ectothrix, 13 |
| endothrix, 13 equinum, 15 eretmorphoron, 301 |
| eretmorphoron, 301 |
| flavum, 15 mentagrophytes, 15 |
| minimunt, 200 |
| oidiophara, 301 fterygodes, 301 |
| toncuranc 301 |
| verrucasum asini, 15 verrucasum v. cqui, 15 Trichophytosis, 14-16 Remedy for, 16 Trichosanthes, 89 |
| Trichophytosis, 14-16 |
| Trichosanthes, 89 celebica, 819 |
| cclebica, 819 cucumerina, 751 |
| |
| globosa, 819 falmata, 135, 751 fubera, 819 |
| 1711/411414, 017 |
| wallichiana, 819 Trichasparum, 297 giganteum, 299 |
| giganteum, 299 |
| Trientalis, 89 |
| granieum, 299 ovoides, 299 Trientalis, 89 Trifolium, 535, 553, 557 agrarium, 553 alexandrinum, 525 |
| alexandrinum, 525 |
| arvense, 553 |
| alexanarium, 525 arvense, 553 clegans, 833 hybridum, 525, 553, 833 incarnatum, 68, 118, 556, f. 560, 557, 833 nigrescens, 833 pratense, 118, 525, 553, f. |
| incarnatum, 68, 118, 556, f. |
| nigrescens, 833 |
| |
| frocumbens, 553 repens, 118, 525, 553, 557, f. 557, 833 terrestris v. hispidissimus, |
| f. 557, 833 |
| terrestris v. hispidissimus, 67 |
| Triganella cretica, 833 Faenum-graecum, 527, 531 |
| Trigonallin 521 640 |
| Triguera ambrosiaca, 855 |
| Trilisa, 90, 524, 757, 772-773 |
| Triguera ambrosiaca, 855 Trilisa, 90. 524, 757, 772-773 Hairy, 773 adoralissima. 138, 772-773 paniculata, 773 |
| |
| Trillium, 89, 377, 385 erectum, 104, 385, 835 |
| erectuni, 104, 385, 835 |
| |

| grandiflorum, 104, 385, 835 |
|--|
| Lama Haward 104 |
| Large-flowered, 104 |
| nivale, 385 |
| recurvatum, 385 sessile, 385 Trimble, W. K., 751, 913 Trimen, Henry, 872, 913 Triodia irritans, 826 Triosteum, 745 majus, 813 perfoliatum, 135, 744, 745, |
| sessile, 385 |
| Trimble, W. K., 751, 913 |
| Trimen, Henry, 872, 913 |
| Triodia irritans, 826 |
| Trinsteum 745 |
| maine 013 |
| hanfaliatum 125 744 745 |
| perfoliatum, 135, 744, 745, |
| |
| Triphragmium Ulmariae, f. |
| 221 |
| Triple-awned Grass, 352 Trisctum, 226 Triticin, 103, 365 Triticum Spelta, 226 Tritiona, 389 Trollius asiaticus, 846 |
| Trisctum, 226 |
| Triticin 103 365 |
| Tuitianin Challe 226 |
| Trincum Spena, 220 |
| Tritonia, 389 |
| Trollius asiaticus, 846 |
| Chinensis, 447 |
| europaeus, 111, 846 |
| laxus, 111 |
| bunilus 447 846 |
| laxus, 111 pumilus, 447, 846 Tropa-cocain, 575 |
| Thomason again 120 400 ere |
| TRUPAEULACEAE, 120, 480, 3/3 |
| Iropaealum, 808 |
| majus, 855 |
| Tropa-cocain, 575 Tropaealum, 868 majus, 855 Tropia eaid, 716 Tropia eaid, 716 Trow, A. H., 209 Truax, Peter, 775 True, R. H., 87, 346, 565, 603, 688, 913 Truffle, 253, 4, 254 |
| Trow. A. H., 209 |
| Truay Peter 775 |
| Thur. D LF 97 246 565 603 |
| True, R. H., 67, 340, 303, 003, |
| 688, 913 |
| Truffle, 253. f. 254 |
| False, 247 |
| Trumpet Creeper, 135, 739 |
| Honeysuckle, 744 |
| Tenvillin 575 |
| Tanan H 012 |
| 1 ryon, ri., 913 |
| Trypsin, 3/3, 62/ |
| Truax, Peter, 775 True, R. H., 87, 346, 565, 603, 688, 913 Truffle, 253, f. 254 False, 247 Trumpet Creeper, 135, 739 Honeysuckle, 744 Truxillin, 575 Tryon, H., 913 Trypsin, 373, 627 Tschirsch, A., 94, 863, 901, 914 Tsuga canadensis, f. 327, 327, |
| 914 |
| Tsuga canadensis, f. 327, 327, |
| |
| |
| |
| |
| |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstinum, 253, f. 254 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstinum, 253, f. 254 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstinum, 253, f. 254 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstinum, 253, f. 254 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstinum, 253, f. 254 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstinum, 253, f. 254 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculin, 180 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculin, 180 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnalum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnalum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberous-rooted Corydalis, |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnalum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberous-rooted Corydalis, |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnalum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberous-rooted Corydalis, |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnalum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberous-rooted Corydalis, |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercele bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubociarin, 687 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercele bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubociarin, 687 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tubocinarin, 687 Tubocurarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tubocinarin, 687 Tubocurarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tubocinarin, 687 Tubocurarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tubocinarin, 687 Tubocurarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tuberose, 64, 586 Tubocinarin, 687 Tubocurarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocoirarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocoirarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocoirarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocoirarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocoirarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocoirarin, 148 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocurarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tubercus, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocurarin, 148 TUBULIFLORAE, 757, 762-802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tubercus, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocurarin, 148 TUBULIFLORAE, 757, 762-802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tubercele bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tuberose, 64, 786 Tuberous-rooted Corydalis, 480 Tuberous-rooted Corydalis, 480 Tuberous-rooted Corydalis, 7ebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocurarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tubercele bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tuberose, 64, 786 Tuberous-rooted Corydalis, 480 Tuberous-rooted Corydalis, 480 Tuberous-rooted Corydalis, 7ebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocurarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tubercele bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tuberose, 64, 786 Tuberous-rooted Corydalis, 480 Tuberous-rooted Corydalis, 480 Tuberous-rooted Corydalis, 7ebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocurarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tubercuse, 64, 386 Tuberose, 64, 386 Tuberose, 64, 186 Tuberose, 64, 186 Tuberose, 64, 186 Tuberose, 64, 186 Tuberose, 64, 186 Tuberose, 64, 186 Tuberose, 780 Tuberose, 687 Tubocurarin, 148 TUBIFLORAE, 158, 698-802 Tubocurarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocurarin, 148 TUBULIFLORAE, 757, 762-802 Tuzzek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tung, 587 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocurarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 Tung, 587 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 Tuberce bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocurarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 Tung, 587 |
| 817 heterophylla, 327 "Tuba," 370 Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocurarin, 148 TUBULIFLORAE, 757, 762-802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 Tung, 587 Tungsten, Salts of, 7 Tupa, 52 Tupelo Gum, 664 |
| 817 heterophylla, 327 "Tuba," 370 Tuba," 370 Tuber acstivum, 253, f. 254 magnatum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculin, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocurarin, 148 TUBULIFLORAE, 757, 762-802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 Tung, 587 Tungsten, Salts of, 7 Tupa, 52 Tupelo Gum, 664 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberose, 70, 386 Tuberose, 70, 386 Tuberose, 70, 70 Tuberose, 70 Tuberos |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercel bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocinarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 Tung, 587 Tungsten, Salts of, 7 Tupa, 52 Tupelo Gum, 664 Turboin, 533 Turkey Opium, 481 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercel bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocinarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 Tung, 587 Tungsten, Salts of, 7 Tupa, 52 Tupelo Gum, 664 Turboin, 533 Turkey Opium, 481 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercel bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocinarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 Tung, 587 Tungsten, Salts of, 7 Tupa, 52 Tupelo Gum, 664 Turboin, 533 Turkey Opium, 481 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercel bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocinarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 Tung, 587 Tungsten, Salts of, 7 Tupa, 52 Tupelo Gum, 664 Turboin, 533 Turkey Opium, 481 |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercle bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberose, 70, 386 Tuberose, 70, 386 Tuberose, 70, 70 Tuberose, 70 Tuberos |
| 817 heterophylla, 327 "Tuba," 370 Tuber acstivum, 253, f. 254 rubrum, f. 254 rubrum, f. 254 rubrum, f. 254 Tubercel bacillus, 170 Organisms, 180 Tuberculosis, 2, 8, 180 Tuberose, 64, 386 Tuberose, 64, 386 Tuberous-rooted Corydalis, 480 Tebeuf, K. F. v., 288, 914 TUBIFLORAE, 158, 698-802 Tubocinarin, 687 Tubocinarin, 148 TUBULIFLORAE, 757, 762- 802 Tuczek, E., 914 Tufted Buttercup, 110 Tulasne, L. R., 275 Tulip, 375, 377 Grass, 868 Tree, 474 Tulipa, 375 Gesneriana, 835 sylvestris, 835 Tumble-weed, 424, 431 Tumbling Mustard, 114, f. 487, 488 Tuna, 635, 868 Tung, 587 Tungsten, Salts of, 7 Tupa, 52 Tupelo Gum, 664 Turboin, 533 Turkey Opium, 481 |

Turnip, 486
Indian, 372, f. 372
Swedish, 486
Turnip-rooted Celery, f. 647, 648
Turpentine, 329, 330, 868
Austrian, 329
Chios, 608
French, 330
Russian, 330
Turpethin, 702
Tussilago, 757
Farfara, 755
Tuttlehead, 733
Tutin, 149, 607
Tutin, Frank, 575
Tutu Districts, Cattle in, 607
Plants, 607 Plants, 607 Twin-leaf, 88, 469 Tylophora asthmatica, 695, 810 fasciculata, 810 laevigata, 810 TYLOSTOMATACEAE, 245
Tympanites, 38, 403, 483, 490,
553, 557, 560, 570, 589,
804 Typha latifolia, 332, f. 334, 855 TYPHACEAE, 102, 855 Typhoid Fever, Cause of, 8 Typhotoxin, 10 Tyrotoxicon, 150 Ubelacker, Dr., 747 Ugenol, 683 Ulex, 118 eurapaeus, 90, 530, 551 Jussiae, 90 Ulexin, 530 ULMACEAE, 405, 856 Ulmus americana, f. 407, 408 campestris, 408 fulva, 408, f. 409 montana, 408 montana, 408
racemosa, 408
Ulva latissima, 190
UMBELLALES, 645-664
UMBELLIFERAE, 44, 125-127,
417, 646, 647-664, 856,
868 Poisoning by, 49-51 Umbelliferone, 648 UMBELLIFLORAE, 157 Umbellularia californica, 112, 478, 828 Umbra Tree, 434 Chinese, 121 Umncy, C., 881, 914 Uncaria, 742 Uncinula spiralis, 269 Unna, P. G., 296, 301 Unna, F. G., 296, 301

Ungnadia speciosa, 850

Upas Tree, 85, 148, 406, f.
408, 587, 868

United States, Flora of, 866

Medicinal plants of, 866

Pharmacopoeia, 866

Poisonous plants of, 866-867 Uraemia, 73 Uraemic poisoning, 804 Symptoms of, 77 Urceola, 691 Urechitea, 692 Urechites suberecta, 807 UREDINEAE, f. 221, 861 Urginea, 377 Scilla, 835

Uric acid, 146

Vallisneria spiralis, 332
Van de Warker, Ely, 790
Van der Velde, 168
Van Dresser, 11., 663
Van Es, L., 914, 915
Van Hasselt, 601
Vandellia crustacea, 852
minnta, 852
Vangueria spinosa, 849
Vanilla, 392, 773
Grass, f. 341
guianensis, 392 Urocystis, 218
agropyri, 219
acculta, f. 218, 219
Uroglena, 188
Uramyces, 230-231
appendiculatus, 230
Fabae, f. 221
pisi, 230
strictus, 230 striatus, 230 Trifolii, 99, 230-231, f. 231, 861 Trifolii, 99, 230-231, f. 231, 861
Ursone, 665
Urtica, 410, 411-413
dioica, 106, 413, f. 413, 857
pracilis, 106, 412, 857
holosericea, 413, 857
membranacea, 412
pilmlifera, 412, 867
spatula, 412
nrens, f. 412, 412-413, 857
URTICACEAE, 53, 106, 405, 409415, 857
Fish poisons in, 2
URTICALES, 156, 404-415
Urticaria, 12, 78, 412, 419
Urticarial poisoning, 804
Urticarial poisoning, 804
Urticarial phairs, Bugloss, f. Vanilla, 392, 773
Grass, f. 341
guianensis, 392
planifolia, 392
planifolia, 392
planifolia, 392
planifolia, 392
vanillin, 392, 683
Vanquelin, 734
Vapors, Poisoning by, 77
Variolaria amara, 861
Varnell, Prof., 290
Vasey, Geo., 357, 914
Vateria, 627
Vancheria, 191
Vaughan, V. C., 150, 5
572, 888, 897, 914
Vaulabelle, A. de, 914
Vegetable, Beefsteak, 234
Butter, 681
Calomel, 371
Poisons, 149
Vellelia paradoxa, 825 Nettle, f. 71 Sedge, f. 71 Grass, f. 71 USTILAGINACEAE, 98, 210-219, Velleia paradoxa, 825 Velvet-leaf, 124, 627 Venus Fly-trap, 498 Veratalbin, 379 861 USTILAGINEAE, 217, 364 Ustilago, 210, 868 avenae, 98, 215-217, f. 216, 861 Veratalbin, 379
Veratramarin, 381
Veratridin, 381
Veratridin, 381
Veratrism, 46, 77, 81, 82, 377, 381
Veratrism, 44, 46
Veratrism, 46, 52, 148, 376, 377, 379, 380-383, 835, 868
album, 4, 75, 148, 381, 835
alkaloids of, 148, 381
Arrow poisons, 2
califarnicum, 104, 381-383, 835 861
bromivora, 210, 215, f. 218
Cesatii, f. 218
Crameri, f. 214
esculenta, 210
Fischeri f. 214
Hardei, 98, f. 211, 217, 861
hypadites, f. 216, 217
maydis, 217
minima, 210
neglecta, 98, 211, f. 218,
861 861 Californicum, 104, 301-363, 835 lobelianum, 381 viride, f. 45, 75, 85, 103, 148, 381, f. 382, 835 Woodii, 103 Verbascum, 52, 734, 868 Blattaria, 734-735, 852 nuda, 98, 217, 861 panici-glauci 217 panici-glauci miliacea, f. 214, 218
perennans, 216, f. 218
seegetum, 217
Sorghi, f. 212
striaeformis, f. 218
Tritici, 98, 217, 861
ntriculasa, 98, 219, 861
zeae, 98, 210-215, f. 211, dubium, 852 arientale, 852 phlamaides, 852 pulverulemum, 852 pnlverulentum, 852 simplex, 852 sinnatum, 852 thapsaides, 852 Thapsus, 134, 733, 734-735, f. 735, 852 Verbena, 708, 868 Aubletia, 708 Family, 131, 707-708 lasstata, 708, 858 officinalis, 858 stricta, 708 urticaefolia, 708 trenosa, 858 Utricularia vulgaris, 698

857-858

VERNONIEAE, 757

864

Vaccaria vulgaris, 108 Vaccinium, 222, 629, 665 arboreum, 665 caespitosum, 665 canadense, 665 corymbosum, 665 v. pallidum, 665 macrocarpon, 665 membranaceum, 665 membranaceinn, 665 Myrtillus, 629 Oxycoccus, 665, 666 pennsylvonicum, 665 stamineum, 665 vacillans, 665 Vaginitis, 179 Vahlen, E., 914 Valertin, G. C., 914 Valeriana diaica, 857 officinalis, 857 VALEBLANGEAF 857 VALERIANACEAE, 857

Veronica, 734
Beccabunga, 853
officinalis, 853
peregrina, 735-736
Virginica, 134, 733, 736,
853 Veronal Poisoning, 868 Verschaffelt, E., 914 Verschaffelt, E., 914
VERTICII,LATAE, 156, 395
Verticillium, 24
graphi, 199
Vervain, Blue, 708
White, 708
Vesicant, 804
Vesicaria gnaphalades, 818 Vesticaria gnaphalades, 818 Vestia lycioides, 855 Vetch, 45, 120, 525, 526 American, 570 Black-purple, 526 Common, 120, 570-572, f. 571
European, 87
German, 572
Hairy, 527, 570
Milk, 562-567
Red, 572
Scarlet, 526
Spanish, 572
Wild, 87
Woolly-pod, 526
Vetch-Bread, 572
Vetchling, 572-573
Marsh, 572
Prairie, 572
Viburnum Lantana, 813
macraphyllum, 813
Opnlus, 813 571 macraphyllim, 813
Opulus, 813
Opulus, 813
Vicat, P. R., 914
Vicia, 570-572
americana, 570
atrapurpurens, 526-535
dasycarpa, 527
Ervilia, 833
Faba, 522, 523, 572
fulgens, 526
lirsnia, 527
sativa, 53, 120, 269, 525,
570, f. 571, 572
villosa, 525, 570
Vicin, 120, 572
Victoria regia, 445
Vigna Catjang, 520, f. 532
Villamilla pernviana, 434
Villaresia Maorei, 839
Villiers, A., 914
Villosa, 508
Vinca majar, 691, 807 Vinca majar, 691, 807 minar, 691, 807 pusilla, 807 Vincetaxicum, 695
nigrum, 810
afficinale, 696, 810
Vinegar, Acetic acid in, 163
Tarragon in, 756 Viala, 631-632, 858 iala, 631-632, 858
blanda, 631
cucullata, 125
Natallii, 631
odorata, 125, 631, 858
pedata, 631, 858
pedatifida, 631
pubescens, 631, f. 632
sepincola, 858
striata, 631
tricalar, 631, f. 632, 632-633 venosa, 858 Verbenaceae, 131, 707-708, Verdier, 874, 882, 914 Vermifuge, 804 Vermont, Economic plants of, 633 VIOLACEAE, 125, 417, 628, 631-633, 858 Violaquercitrin, 632 Violet, Bird-foot, 631 Blue, Common, 125 Canada, 631 Dog's-tooth, 104 Vernania, 757 anthelminticum, 816 nigritiana, 816 noveboracensis, 222

| European, 83 |
|--|
| Fragiant, 631 |
| Pansy, 632 |
| Suppl 125 632 |
| White-flowered, 631 |
| Wood Sorrel, 580 |
| Yellow, 631, f. 632 Violin, 632, 633 |
| Violin, 632, 633 |
| Viner's Bugloss, 131 |
| Virchow, R., 231, 263 |
| Virchow, R., 231, 263 Virginia Creeper, 124, 150, |
| 620 |
| Peppergrass, 72 |
| Snakeroot, 417 |
| Strawberry, Wild, 115, 509 |
| Strawberry, Wild, 115, 509 Virgin's Bower, 109, 446, 456, f. 457 |
| 456, f. 457 |
| Viscum olbum, 106, 836 |
| seedlings, 595 |
| Vismia viridiflora, 629 |
| VITACEAE, 124, 620, 858 |
| VITACEAE, 124, 620, 858 Vitex littorolis, 708 |
| Negundo, 708 |
| pteropoda, 858 |
| vestita, 858 |
| l'itis elongota, 858 |
| Minohau, 858 |
| Saponorio, 858 |
| sesselifolio, 858 |
| Vitriol, Oil of, 143 |
| l'oondesio subterroneo, 521 |
| Vogel, 3 |
| Volkens C 915 |
| Volkens, G., 915 Volvocaceae, 98, 191-193 |
| Volvorio bombycina, 243-244 |
| redriacea 860 |
| volvoceo, 860 Volvox, 93, 98, 191, 193 glebotor, 193 |
| globotor 193 |
| minor, 193 |
| Vuillemin, P., 298, 299 |
| Vuillet, Jean, 895 |
| · amer, Jean, ove |
| W |
| • |
| 111 1 102 110 611 615 |

Waahoo, 123, 148, 614, 615, f. 615
Wager, II., 206
Wainwright, J. W., 915
Wakerobin, f. 371, 385
Erect, 104
Large-flowered, 385
Prairie, 385
Sessile-flowered, 385
White-flowered, Large, 385
White-flowered, Large, 385
Walte-flowered, Large, 385
Walter, S., 695, 915
Wall-flower, 486
Waleff, S., 695, 915
Wallef, S., 695, 915
Waller's Farmer, 915
Waller's Farmer, 915
Waller's Farmer, 915
Wallorh, K. F. W., 13
Walnut, Black, f. 400, 401
California, 401
Iapanese, 401
Walseh, J. J., 295
Walsh, L. II., 915
Wolshia omorphello, 569
Wolshuro, 52, 89
piscidio, 837
Wandering Jew, 374
Warburton, C. W., 276, 346, 915
Warren, L. E., 805-806, 915
Warren, L. E., 805-806, 915
Washingtonia, 370
Palm, 370
Water, Algae in, 91-95
Avens, 505
Cherry-laurel, 503
Chestnut, 640
Chinquapin, 444
Cress, 486

Crowfoot, 10ek, Great, 420 10ek, Great, 420 10ropwort, 649 11emloek, 126, 648, 651-659 Crowfoot, 446 1 Propwort, 649
1 Icmloek, 126, 648, 651-65
Lily, 444-445
Blue-flowered, 445
Yellow, f. 445
Moulds, f. 208
Parsnip, Cut-leaved, 76
Creeping, f. 650
Pepper, 422-423
Wild, 422-423, f. 423
Plantain, 102, 335, f. 335
Family, 335
Pollution of, 91
Smartweed, 421-422
'ater as source of diseas Water as source of disease, 8-10
Water-bloom, 93, 186, 868
Waterleaf Family, 131, 703-704
Watermelon, 750, f. 748
Water-weed, 332, 445
Watkins, II. C., 909
Watkins, W. II., 915
Watson, E. B., 403
Watson, Miss C. M., 564
Watson, Sereno, 342, 564, 915
Wattles, Australian, 523 Way, 915 Weatherby, C. A., 915 Weather-glass, Poor-man's, 128 Shepherd's, 128 Webb, Paul C., 915 Webber, H. J., 287, 582, 625 Wechsberg, 168 Weeds, Poisonous, 868 Weems, J. B., 903 Weevers, 82 Weems, J. B., 903
Weevers, 82
Weihardt, 482, 767
Welleome Laboratory, 4
Wellman, F. C., 915
Weivitschia, 326
Wenzell, 277
Weschke, Carl, 915
West India Arrow-root, 77, 391
Cedar, 575
Gherkin, 750
Locust Tree, 527
Westbrook, F. E., 915
Western, Aconite, 450, 452, 453, f. 452
Cleome, 114
Cone-flower, 779
Dogwood, 664
Honeysuckle, 744
Larkspur, Purple, 109
Lupine, 118
Needle Grass, f. 66, 102, 355, f. 355
Nettle, 106
Snakeroot, 773-774
Yellow Pine, 330
Westgate, J. M., 915
Wettstein, R. von, 915
Wheat, 225-338
Bunt, f. 219, 219-220
Cow, 134, 734
Egyptian, 345
Grass, 276, 364
Western, 364, 365
India, 419
Rust, 99, 227, f. 228, 228 India, 419 Rust, 99, 227, f. 228, 228 Smut, 217 Stinking, 98, 219, 220, f. Whelpley, II. M., 915 Whipple, G. C., 93, 189, 915 White, E. A., 915 White, G. R., 916 White, J. C., 96, 104, 112,

134, 215, 217, 455, 456, 464, 471, 485, 490, 503, 583, 587, 600, 601, 608, 609-612, 634, 679, 739, 794, 803, 804, 916 White, R. B., 417 White, Amanita, 34 Ash, 683 Aster, Small, 776 Aster, Small, 776
Bancberry, 108
Birch, 404
Cedar, 327, 328
Clover, 118, 525, 553, 557, 1. 557
Sweet, 118, f. 551
Gentian, 690
Helicbore, 75, 103, 381
Indigo, Wild, 117, 541, f. Jasnine, Large Flowered, f. 684 Lady's Slipper, Small, 105, 393 Lupine, European, 547 Mulberry, 406 Mustard, 77, 486, 488, f. 489, 490 Oak, 403 Oak, 403 Pepper, 87 Pine, 329, f. 329 Rust, f. 204 Sage, 424 Snakeroot, 771 Thorn, 513 Vervain, 708 hite-flowered, W White-Mowered Wake-robin, Large, 385
White-weed, 138, 756, 778-779, f. 778
Widal, F., 263, 916
Wigand, Albert, 916 Wikstroemia Chomoedophne, 855
Wilcox, E. M., 100
Wilcox, E. V., 5, 44, 84, 96, 97, 109, 378, 379, 383, 462, 466, 548, 568, 626, 656, 762, 793, 795, 916
Wilcox, R. W., 903, 916
Wilcox, T. E., 916
Wild Apple, Balsam, 751
Apple, Crab, 512
Barely, 66, 67, 103, 338, 368, f. 368
Carrot, 663-664
Cherry, Black, 64, 117, 503, 505, f. 515, 515-516
Red, 116, f. 504, 505, 516-519, f. 576
Western, 515
Cinnamon, 627
Coffee, 745
Comfrey, 131, 706
Cueumber, 751
Ginger, f. 415, 416
Grape, Blue, 620
Heliotrope, 131
Hydrangea, 115, 500
Indigo, Large-bracted, 541
Large White, 117, 541, f. 541, f. 541, f. 541, f. 541, f. 541, f. 541, f. 541, f. 560 Wikstroemia Chomoedophne, Large White, 117, 541, 1, 541, Pineapple, 373

Strawberry, European, 509
Woods, Dr. A. F., 348
Woodville, Wm., 917
Woody, Aster, 138, 776
Woolly, Loco Weed, 39, 562, 567, 568
Thistle, f. 798, 801
Woolly-pod Vetch, 526
Worden Grape, 620
Wyoming Larkspur, 109
Wormseed, 107, 428, f. 428
Oil of, 107, 428
Levaut, Oil of, 791
Spanish, 424
Wormwood, 74, 754, 755, 756, 790, 791-794, f. 791
Biennial, 140, f. 792
Common, 140, 793
Wright, 168
Wright, J. S., 917
Wright, R., 917
Wright's Datura, 133, 732-733
Wyman, Dr. Walter, 343
Wyndham, R., 881 Plum, northern, 116, f. Potato, 701 Rice, 276, 338 Rosemary, 127, 673 Rye, 27, 90, 149, 276, 280, 546 Ryc, 27, 90, 149, 276, 280, 546
Sago, 378
Senna, f. 337
Tea, 501
Tobacco, 729
Vetch, 87
Water Parsnip, 49, 650, 656, 661-663, f. 662
Yam, 374, 688
Wildeman, F. de, 916
Willy, H. W., 916
Williams, Dr., 10
Williams, Dr., 10
Williams, N. J., 916
Williams, X. J., 916
Williams, X. J., 916
Williams, T. A., 348, 357, 916
Williams, W. T., 259, 916
Williams, W. T., 916
Williams, W. T., 916
Williams, W. T., 916
Williams, W. T., 916
Williams, W. T., 917
Willoughly, W. G., 917
Willoughly, W. G., 917
Willoughly, Bo-397
Peach-leaved, f. 397
Willow.herb, 629
Grant 644 Nanthin, 147
Bases, 146
Derivatives, 146
Di-methyl, 146
Methyl, 146
Xanthisma texana, 138, 816
Xanthinm, 757, 767-770
canadense, 67, f. 68, 137, 768-770, f. 760, 816
spinosum, 137, 767, f. 768, 816 Family, 396-397
Peach-leaved, f. 397
Willow-herb, 629
Great, 644
Wilmer, B., 917
Wilson, Dr., 572
Wilson, Dr., 572
Wilson, F. W., 25, 96, 130, 694, 917
Wilson, James, 878, 917
Wilson, J. C., 917
Wilson, P., 917
Wilson, P., 917
Wilson, P., 917
Wilson, P., 917
Wilson, P., 917
Wilson, P., 917
Wilson, P., 917
Wilson, P., 917
Wilson, P., 917
Wilson, P., 917 Zalachas, 729 Zamang, 527 816 strumarium, 137, 816 Xanthopuccin, 108, 468 Xanthostrumarin, 137, 769 Wilt, Corn, 163
Wind-flower, 109, f. 455, 455456
Wine Palm, 370
Wineberry, 505
Winegate, J. B., 917
Winkler, L., 917
Winnebago Lake, 93
Winslow, Kenelm, 55, 96,
109, 251, 278, 381, 450,
471, 482, 518, 575, 593,
628, 687, 689, 726-727,
728, 731, 736, 742, 917
Wintergreen, 621, 665
Oil of, 404, 621, 665
Winter's-bark, 474
Winton, A. L., 917
Winton Disease, 795
Wire Grass, 374
Vistaria, 64, 527, 534, 629
chinensis, 527, 534, 629, 833
speciosa, 527
Wisterin, 534
Witches Broom' on Birch,
253
on Cherry, 253
Witchers Broom' Xanthorrhoea, 358 hastilis, 375 Xenophon, 64 Xeranthemum annuum, Xeranthemum annuum, 53
Xerophylla Douglasii, 835
Ximenia americana, 53
Xylia dolabriformis, 833
Xylomelium pyriforme, 844
Xylopia odoratissima, 806
polycarpa, 806
salicifolia, 806
Xylorhiza Parryi, 776
Xyridaceae, 372, 858
Xyris communis, 858 Yagna, 370 Yani Bean, 521 Chinese, 699 Family, 374 Family, 374
Japanese, 374
Starch, 374
Wild, 374, 688
Yarrow, 149, 756, 785-787, f. 786, f. 787
Yampon, 123
Yeast, f. 249, 250-251
Chinese, 198
Common, 250-251, f. 250, f. on Cherry, 253
on Cherry, 253
on Cherry, 253
withania somnifera, 855
Withania somnifera, 855
Withaus, R. A., 872, 917
Wittstein, G. C., 917
Woehler, 505
Wokas, 444
Wolfberry, 744
Wolfferny, 744
Wolfferny, 744
Wolfferny, Dr. 199
Wolfgang, 482
Wolfs-bane, Trailing, 109
Wood, 276, 503
Wood, Alphonso, 917
Wood, Alphonso, 917
Wood, Anemone, 77, 109
Nettle, 77, 106, 413, 414
Sorrel, 120, 580
Family, 120 251
Parasitic, 249
Pink, 250
Yellow Birch, 404, f. 405
Cleome, 496
Clover, Hop, 553
Sweet, 118, 552
Flax, Large-flowered, 120, 581 Flowering Spurge, f. 601 Jasmine, 688-689 Lady's Slipper, Small, 105, f. 393

Pine, Western, 330
Puccoon, 446, 468
Violet, 631, f. 632
Water Lily, f. 445
Yellow-eyed Grass, 372
Yellow-flowered Alfalfa, 526
Bitter-weed, 539
Nightshade, 692
Yellow-leaf disease of barley, 281, f. 281
Yerba, 614
Maté, 614, 742
Santa, 703
Yew, 4, 72, 101, 148, 326, 328, 868
American, 101, 328
Furopean, 101, 328
Yopan, 742
York-road poison, 533
Yncca, 89, 375
angustifolia, 375
filamentosa, 375, 835
glauca, 63, 835
imperialis, 375
Yuki, 380
Yulan Tree, 474

Z Zalachas, 729
Zamang, 527
Zamia, 326
media, 819
Zanthorhiza, 840
Zanthorkiza, 840
Zanthorylum, 89
alatum, 120, 849
americanum, 581
caribaeum, 849
Clava-Herculis, 581
Naranjillo, 849
scandens, 120, 849
Zea amylacea, 341
amyleasaccharata, 3 amyleosaccharata, 341
canina, 342
everta, 341
indentata, 341
indurata, 341
indurata, 341
Mays, 102, f. 337, 826
History of, 340-342
saccharata, 341
tunicata, 341
Zedoary, 392
Zehl, I., 917
Zenobia speciosa, 821
Zephyranthes, 386-388
atamasco, 105, 387-388, f.
388, 805
Ziegler, Ernst, 304
Zinc, Sulphate of, 6
Zingiber officinale, 77, f. 390,
391, 858
ZINGIBERACEAE, 391, 858 amyleasaccharata, 341 Zingiber ofteniale, 77, 1. 390, 391, 858
Zinnia, 756
Zinnia, 756
Zizania, Ergot on, 276
latifolia, 210
Zizyphus, 2, 89, 621
Jourcira, 847
Jujuba, 621
Lotus, 621, 847
Zoethout, W., 9, 918
Zoidiogama, 154
Zoogloea, 161
Zoon, J., 917
Zopf, W., 918
Zwaluwenberg, A. V., 909, 918
Zygadenus, 377-379
angustifolium, 835
chloranthus, 378-379
elegans, 6, 103, 835
paniculatus, 378
smooth, 378-379 venenosus, 6, 103, f. 376, 377.378, 379, 835
Zygnema, f. 92, 189, 190, f. 191
TYGNEMATACEAR, 190

ZYGOMYCETES, 154, 195-204 ZYGOPHYLLACEAE, 89, 575, 858 Zygophyllum coccineum, 858 Fabago, 858

iodocarpum, 858 Zygorhyncus, 195 Zygospores, 13 Zylia, 89 Zymase, 250





This preservation copy
was printed and bound at
Bridgeport National Bindery, Inc.,
in compliance with U.S. copyright law.
The paper used meets the requirements
of ANSI/NISO Z39.48-1992
(Permanence of Paper).

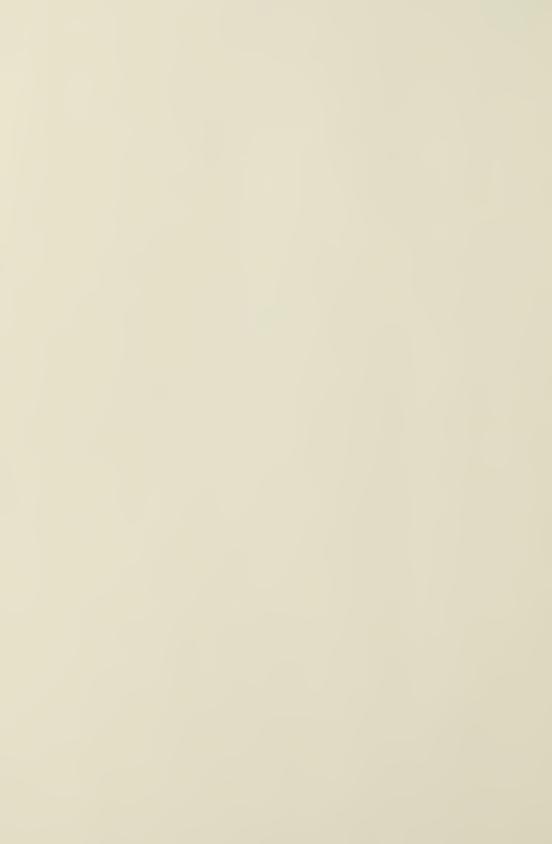
DPD



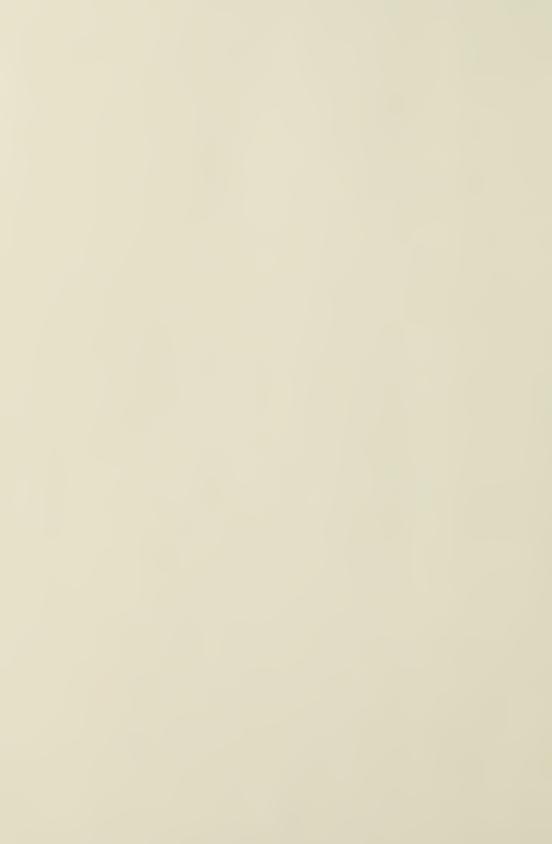
1997

















3 FEI-0 !

DATE DUE

| , | | |
|--------------|--|-------------------|
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |
| | | |
| GAYLORD | | PRINTED IN U.S.A. |



Science qQK 100 .U6 P32 1911
Pammel, L. H. 1862-1931.

A manual of poisonous plants

